

1 Full title:

2 Comparative evaluation of the Geenius™ HIV 1/2 Confirmatory Assay and the HIV-1 and HIV-
3 2 Western blots in the Japanese population

4 Short title:

5 Comparison between the Geenius HIV 1/2 Confirmatory Assay and the HIV-1 and HIV-2
6 Western blots

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27

28 **Abstract**

29 Accurate diagnosis of earlier HIV infection is essential for treatment and prevention.
30 Currently, confirmation tests of HIV infection in Japan are performed using Western blot (WB),
31 but WB has several limitations including low sensitivity and cross-reactivity between HIV-1 and
32 HIV-2 antibodies. To address these problems, a new HIV testing algorithm and a more reliable
33 confirmation and HIV-1/2 differentiation assay are required. The Bio-Rad Geenius™ HIV-1/2
34 Confirmatory Assay (Geenius) has recently been approved and recommended for use in the
35 revised guidelines for diagnosis of HIV infection by the Center for Disease Control and
36 Prevention (USA). We made comprehensive comparison of the performance of Geenius and the
37 Bio-Rad NEW LAV BLOT 1 and 2 (NLB 1 and 2) which are WB kits for HIV-1 and HIV-2,
38 respectively, to examine if Geenius is a suitable alternative to these WB assays which are now
39 being used in HIV testing in Japan. A total of 166 HIV-1 positive samples (146 from patients
40 with established HIV-1 infection and 20 from patients with acute infection), five HIV-1
41 seroconversion panels containing 21 samples and 30 HIV-2 positive samples were used. In
42 addition, a total of 140 HIV negative samples containing 10 false-positives on screening tests
43 were examined. The sensitivity of Geenius and NLB 1 for HIV-1 positive samples was 99.3%
44 and 98.6%, respectively. Geenius provided more positive results in the samples from acute
45 infections and detected positivity 0 to 32 days earlier in seroconversion panels than NLB 1. NLB
46 2 gave positive results in 12.3% of HIV-1 positive samples. The sensitivity of both Geenius and
47 NLB 2 for HIV-2 positive samples was 100%. The specificity of Geenius, NLB 1 and NLB 2
48 was 98.5%, 81.5% and 90.0%, respectively. Geenius is an attractive alternative to WB for
49 confirmation and differentiation of HIV-1 and HIV-2 infections. The adaptation of Geenius to
50 the HIV testing algorithm may be advantageous for rapid diagnosis and the reduction of testing
51 costs.

52

53 **Introduction**

54 The risk of HIV transmission during acute and early infection is much higher than that
55 during established infection [1]. Furthermore, early initiation of antiretroviral therapy (ART)
56 substantially reduces the risk of transmission to sexual partners [2] and improves clinical
57 outcomes, compared with delayed ART [3]. Accurate diagnosis of earlier HIV infection is
58 important for treatment and prevention strategies.

59 Currently, diagnosis of HIV infection in Japan is carried out mainly in two different
60 algorithms: (i) a sample tested positive on HIV-1/2 antigen/antibody assay is retested with HIV-1
61 Western blot (WB-1) and HIV-2 Western blot (WB-2) simultaneously, and then, if the results on
62 both assays are negative, applied to nucleic acid test (NAT) of HIV-1 plasma RNA; this
63 algorithm is recommended by the National Institute of Infectious Diseases (Japan) [4]; (ii) a
64 sample that tested positive on HIV-1/2 antigen/antibody assay is then retested with WB-1 and
65 NAT at the same time, and then, if the results on both assays are negative, applied to WB-2; this
66 is recommended by the Japanese Society for AIDS Research [5]. These algorithms, however,
67 have several limitations associated with Western blot that include false negative or indeterminate
68 results in the early phase, cross-reactivity between HIV-1 and HIV-2 [6], and a labor-intensive
69 and time-consuming protocol.

70 In 2014, the Center for Disease Control and Prevention (CDC) in the US published
71 revised guidelines for diagnosis of HIV infection in which the use of an HIV-1 and HIV-2
72 antibody differentiation assay is recommended after a repeatedly reactive HIV-1/2
73 antigen/antibody test [7]. The FDA-approved Multispot™ HIV-1/HIV-2 Rapid Test (Bio-Rad
74 Laboratories) was initially validated for this purpose. Thereafter, Bio-Rad developed a new
75 confirmatory and differentiation test, the Geenius™ HIV-1/2 Confirmatory Assay (hereafter
76 called Geenius). Geenius received a CE mark in February 2013 and clearance from the Food and
77 Drug Administration in October 2014. Although Geenius has been evaluated in many studies [8–
78 17], there have been few studies on comparison between Geenius and WB. Moon et al. compared
79 the performance of Geenius and WB-1 [16] but did not tested WB-2, and thus they did not
80 comparatively evaluate the HIV-1/2 differentiation ability of Geenius and WB-1/WB-2.

81 In Japan, while Geenius has not been approved yet, there is a growing interest in the
82 CDC-recommended HIV diagnostic algorithm because it is expected to decrease the number of
83 indeterminate results, allow earlier identification of HIV infections, and reduce the number of
84 NAT to resolve the ambiguity of WB results.

85 The aims of this study are to compare the confirmation and differentiation performance
86 of Geenius and NEW LAV BLOT 1 and 2 (Bio-Rad Laboratories, Tokyo, Japan, hereafter called
87 NLB 1 and 2), which are WB-1 and WB-2 kits, respectively, and to examine if Geenius is a
88 suitable alternative to WB in the HIV testing algorithm in Japan.

89

90 **Material and methods**

91

92 **Samples and patients**

93 A total of 166 HIV-1 positive samples were used: 146 were obtained from patients with
94 established HIV-1 infection and 20 from patients with acute infection. Among the patients with
95 established infection, 73 were obtained from patients receiving ART at the Keio University
96 Hospital or Atsugi City Hospital and had been diagnosed with HIV-1 infection by either of
97 Dainascreen[®] HIV Combo (an HIV-1 p24 Ag/HIV-1/2 Ab immunochromatographic test, Alere
98 Medical, Tokyo, Japan) or the Architect[®] HIV Ag/Ab Combo Assay (an automated HIV-1 p24
99 Ag/HIV-1/2 Ab test, Abbott Japan, Chiba, Tokyo), followed by NLB 1 and 2 and, if necessary,
100 the Cobas AmpliPrep/Cobas TaqMan[®] HIV-1 Test (an automated qualitative HIV-1 RNA test,
101 Roche Diagnostics, Tokyo, Japan, hereafter called Cobas). The other 93 samples were obtained
102 from individuals seeking HIV testing in public health centers located in Kanagawa and Osaka:
103 85 were positive on Dainascreen HIV Combo and 8 were positive on the Architect[®] HIV Ag/Ab
104 Combo Assay. Their infections were confirmed by NLB 1 and 2 or Cobas. Established HIV-1
105 infection was defined by positive results on both NLB 1 and Cobas; acute HIV-1 infection was
106 defined by an indeterminate or negative result on NLB 1 but a positive Cobas result.

107 Five HIV-1 seroconversion panels comprised of seven, five, four, three and two
108 samples, respectively, were obtained from patients attending the Shirakaba Clinic in Tokyo,
109 Japan.

110 Thirty samples of two commercially obtained HIV-2 panels were used: five from HIV-2
111 Mixed Titer AccuSet™ Performance Panel (PRE301B, SeraCare Life Sciences, Millford, MA)
112 and 25 from Plasma-CPD-A Anti HIV-2 (HemaCare, Los Angeles, CA).

113 A total of 140 HIV negative samples were obtained from individuals seeking HIV
114 testing in the public health centers, which were tested as mentioned above; 10 of them were
115 false-positive on screening tests using Dainascreen HIV Combo, which were negative or
116 indeterminate on NLB 1 and 2, and negative on Cobas.

117 Comparative testing by Geenius and NLB 1 and 2 was conducted between May 2016
118 and April 2017 in Kanagawa Prefectural Institute of Public Health, Osaka Institute of Public
119 Health, and Keio University School of Medicine according to the manufacturer's instructions.

120

121 **Geenius**

122 Geenius is a single-use immunochromatographic test for the confirmation and
123 differentiation of individual antibodies to HIV-1 and HIV-2 in whole blood, serum or plasma
124 samples using HIV synthetic peptides or recombinant proteins for HIV-1 (p31 [POL], gp160
125 [ENV], p24 [GAG] and gp41 [ENV]) and HIV-2 (gp36 [ENV] and gp140[ENV]). Geenius is
126 aimed at confirming the presence of antibodies to HIV-1 and HIV-2 in samples reactive by
127 screening tests. Banding patterns and intensities on a Geenius cassette were read by an
128 automated reader connected to a personal computer and interpreted using the Geenius algorithm.
129 This cartridge assay allows rapid evaluation within 30 min. Interpretive results involve HIV
130 negative, HIV-2 indeterminate, HIV-1 indeterminate, HIV indeterminate, HIV-1 positive, HIV-2
131 positive, HIV-2 positive (with HIV-1 cross-reactivity), and HIV positive untypable.

132

133 **NLB 1 and 2**

134 NLB 1 and 2 are the only WB kits approved by The Pharmaceuticals and Medical
135 Devices Agency (PMDA) of Japan for confirmation of HIV-1 and HIV-2 infection, respectively.
136 Bands were observed visually. Interpretation of banding patterns is performed as follows: for
137 HIV-1, the presence of at least two of three ENV bands (GP160, GP120 and GP41) is considered
138 positive, no HIV-1 specific band negative, and other patterns indeterminate; for HIV-2, the
139 presence of one ENV, one GAG and one POL band is considered positive, no HIV-2 specific
140 band negative, other patterns indeterminate.

141

142 **Statistics**

143 Sensitivity and specificity were determined by considering indeterminate results as not
144 positive and not negative, respectively, with 95% confidence interval [95% CI]. Cohen's kappa
145 (κ) was calculated to assess agreement between Geenius and NLB 1.

146

147 **Results**

148

149 **Samples in established HIV-1 infection**

150 Geenius, NLB 1, and NLB 2 results on 146 samples from patients with established HIV-
151 1 infection are compared in Table 1. Geenius provided 145 HIV-1 positive results including one
152 HIV positive untypable (sensitivity, 99.3% [95% CI, 96.2–99.8]) and one HIV-1 indeterminate
153 result. NLB 1 showed 144 positive result (sensitivity, 98.6% [95% CI, 95.1–99.6]) and two
154 indeterminate results: the indeterminate results were observed on samples from patients
155 receiving ART. It is notable that only four samples were negative by NLB 2, which may be due
156 to high cross-reactivity.

157

158 **Table 1. Comparison of Geenius with NLB 1 and 2 results for established and acute HIV-1**
159 **infection samples.**

| | WB | | Geenius | | | | |
|---|-------|---------------|----------------|---------------------|----------------|---------------------|------------------------|
| | | | HIV-1 positive | HIV-1 indeterminate | HIV-2 positive | HIV-2 indeterminate | HIV positive untypable |
| Established HIV-1 Infection ^a (<i>n</i> = 146) | NLB 1 | Positive | 143 | 0 | 0 | 0 | 1 |
| | | Indeterminate | 1 ^c | 1 ^c | 0 | 0 | 0 |
| | | Negative | 0 | 0 | 0 | 0 | 0 |
| | | Total | 144 | 1 | 0 | 0 | 1 |
| | NLB 2 | Positive | 18 | 0 | 0 | 0 | 0 |
| | | Indeterminate | 122 | 1 | 0 | 0 | 1 |
| | | Negative | 4 | 0 | 0 | 0 | 0 |
| | | Total | 144 | 1 | 0 | 0 | 1 |
| Acute HIV-1 infection ^b (<i>n</i> = 20) | NLB 1 | Positive | 0 | 0 | 0 | 0 | 0 |
| | | Indeterminate | 7 | 6 | 0 | 0 | 0 |
| | | Negative | 0 | 0 | 0 | 0 | 0 |
| | | Total | 7 | 6 | 0 | 0 | 0 |
| | NLB 2 | Positive | 0 | 0 | 0 | 0 | 0 |
| | | Indeterminate | 5 | 3 | 0 | 0 | 0 |
| | | Negative | 2 | 3 | 0 | 0 | 0 |
| | | Total | 7 | 6 | 0 | 0 | 0 |

160 ^aFourth-generation enzyme immunoassay positive, NLB 1 positive and NAT positive at the time
161 of initial diagnosis.

162 ^bFourth-generation enzyme immunoassay positive, NLB 1 negative and NAT positive at the time
163 of sample collection

164 ^cOn ART at the time of sample collection.

165

166 **Samples in acute HIV-1 infection**

167 Geenius, NLB 1, and NLB 2 results on 20 samples from patients with acute HIV-1
 168 infection are compared in Table 1. Geenius reclassified seven of the NLB 1 indeterminate
 169 samples as positive, showing that Geenius has a higher detection sensitivity than NLB 1.

170

171 **Seroconversion panels**

172 Five HIV-1 seroconversion panels were used to compare the detection ability of
 173 identifying positive samples during the early phase of infection between Geenius and NLB 1
 174 (Table 2). Geenius gave positive results 0 to 32 days earlier than NLB 1. Cross-reactive p26
 175 bands appeared in NLB 2 as the specific HIV-1 antibody titer increased, while no HIV-2-related
 176 band was observed in Geenius.

177

178 **Table 2. Comparison of Geenius with NLB 1 and 2 results for HIV-1 seroconversion**
 179 **panels^a.**

| Patient | Sample | Days ^b | Geenius | | NLB 1 ^d |
|---------|--------|-------------------|-----------------------------|-------|--|
| | | | HIV-1 ^c | HIV-2 | |
| A | 1 | 0 | Neg | Neg | Neg |
| | 2 | 9 | Pos (gp160, p24, gp41) | Neg | Ind (gp160, p68, p55, p24) |
| | 3 | 16 | Pos (gp160, p24, gp41) | Neg | Ind (gp160, p68, p55, p40, p31, p24, p |
| | 4 | 36 | Pos (gp160, p24, gp41) | Neg | Pos |
| | 5 | 42 | Pos (gp160, p24, gp41) | Neg | Pos |
| | 6 | 65 | Pos (p31, gp160, p24, gp41) | Neg | Pos |
| | 7 | 107 | Pos (p31, gp160, p24, gp41) | Neg | Pos |
| B | 1 | 0 | Neg | Neg | Neg |
| | 2 | 7 | Neg | Neg | Neg |
| | 3 | 40 | Pos (gp160, p24, gp41) | Neg | Pos |

| | | | | | |
|---|---|-----|-----------------------------|-----|--------------------------|
| | 4 | 47 | Pos (p31, gp160, p24, gp41) | Neg | Pos |
| | 5 | 85 | Pos (p31, gP160, p24, gp41) | Neg | Pos |
| C | 1 | 0 | Neg | Neg | Neg |
| | 2 | 7 | Pos (gp160, gp41) | Neg | Ind (gp160, p24) |
| | 3 | 39 | Pos (gp160, p24, gp41) | Neg | Pos |
| | 4 | 126 | Pos (gp160, p24, gp41) | Neg | Pos |
| D | 1 | 0 | Neg | Neg | Neg |
| | 2 | 7 | Ind (gp41) | Neg | Ind (p52, p40, p24, p18) |
| | 3 | 33 | Pos (gp160, p24, gp41) | Neg | Pos |
| E | 1 | 0 | Pos (gp160, p24, gp41) | Neg | Pos |
| | 2 | 8 | Pos (gp160, p24, gp41) | Neg | Pos |

180 ^aResults are shown by Pos (positive), Ind (indeterminate) or Neg (negative).

181 ^bTime from collecting the first sample.

182 ^cWhen a result was positive, reactive antigens are shown in parenthesis.

183 ^dWhen a result was indeterminate, reactive antigens are shown in parenthesis.

184

185 HIV-2 panels

186 Thirty samples of two commercial HIV-2 panels were used to compare Geenius, NLB 1,
 187 and NLB 2 (Table 3). All samples were positive with NLB 2 (sensitivity, 100% [95% CI, 88.4–
 188 99.5]); two samples were positive and 28 were indeterminate with NLB 1 (false-positive rate,
 189 6.7% [95% CI, 2.1–12.1]). Geenius gave 28 HIV-2 positive and two HIV positive untypable
 190 results (sensitivity, 100% [95% CI, 88.4–99.5]).

191

192 **Table 3. Comparison of Geenius with NLB 1 and 2 results for HIV-2 panel samples.**

| | | Geenius | | | |
|--|--|---------|-------|---------------------------|--------------|
| | | HIV-1 | HIV-2 | HIV-2 positive with HIV-1 | HIV positive |
| | | | | | |

| | | positive | positive | cross-reactivity | untypable | ne |
|-------|---------------|----------|----------|------------------|-----------|----|
| NLB 1 | Positive | 0 | 0 | 2 | 0 | |
| | Indeterminate | 0 | 10 | 16 | 2 | |
| | Negative | 0 | 0 | 0 | 0 | |
| | Total | 0 | 10 | 18 | 2 | |
| NLB 2 | Positive | 0 | 10 | 18 | 2 | |
| | Indeterminate | 0 | 0 | 0 | 0 | |
| | Negative | 0 | 0 | 0 | 0 | |
| | Total | 0 | 10 | 18 | 2 | |

193

194 Seronegative samples

195 A total of 130 screening negative samples were used to determine the specificity of
 196 three assays (Table 4). Concordant negative results between Geenius and NLB 1 were obtained
 197 for 104 samples; those between Geenius and NLB 2 for 116 samples. The specificity of Geenius,
 198 NLB 1, and NLB 2 were 98.5% (128/130) [95% CI, 94.6–99.5], 81.5% (106/130) [95% CI,
 199 73.8–87.2] and 90.0% (117/130) [95% CI, 83.5–94.0], respectively.

200

201 **Table 4. Comparison of Geenius with NLB 1 and 2 results for negative samples by fourth-**
 202 **generation immunoassay ($n = 130$).**

| | | Geenius | | | | | |
|-------|---------------|-------------------|------------------------|-------------------|------------------------|---------------------------|--|
| | | HIV-1 positive | HIV-1 indeterminate | HIV-2 positive | HIV-2 indeterminate | HIV positive untypable | |
| NLB 1 | Positive | 0 | 0 | 0 | 0 | 0 | |
| | Indeterminate | 0 | 0 | 0 | 0 | 0 | |
| | Negative | 0 | 1 | 0 | 1 | 0 | |
| | Total | 0 | 1 | 0 | 1 | 0 | |

| | | | | | | | |
|-------|---------------|---|---|---|---|---|--|
| NLB 2 | Positive | 0 | 0 | 0 | 0 | 0 | |
| | Indeterminate | 0 | 0 | 0 | 1 | 0 | |
| | Negative | 0 | 1 | 0 | 0 | 0 | |
| | Total | 0 | 1 | 0 | 1 | 0 | |

203

204 **False-positive samples**

205 It is important for a confirmatory assay to discriminate between acute HIV-1 infections
 206 and false positive screening results. Ten Dainascreen HIV Combo positive but Cobas negative
 207 samples were tested with the three assays (Table 5): eight were negative and two were
 208 indeterminate (positive p31 bands) with Geenius; six were negative and four were indeterminate
 209 with NLB 1; five were negative and five were indeterminate with NLB 2, suggesting Geenius is
 210 the most specific for HIV-1 false-positive screening samples among the three kits.

211

212 **Table 5. Comparison of Geenius with NLB 1 and 2 results for HIV-1 Combo positive but**
 213 **NAT negative samples ($n = 10$)^a.**

| Sample. | Geenius ^a | | NLB 1 | |
|---------|----------------------|----------|--------------------------|---------|
| | HIV-1 | HIV-2 | | |
| 1 | Negative | Negative | Indeterminate (p18) | |
| 2 | Negative | Negative | Indeterminate (p18) | Indeter |
| 3 | Negative | Negative | Negative | |
| 4 | Negative | Negative | Negative | |
| 5 | Negative | Negative | Indeterminate (p24, p18) | |
| 6 | Indeterminate (p31) | Negative | Indeterminate (p18) | Inde |
| 7 | Negative | Negative | Negative | |
| 8 | Negative | Negative | Negative | Inde |
| 9 | Indeterminate (p31) | Negative | Negative | Inde |

| | | | | |
|----|----------|----------|----------|------|
| 10 | Negative | Negative | Negative | Inde |
|----|----------|----------|----------|------|

214

215 **Concordance**

216 The overall concordance (κ) between Geenius and NLB 1 was 0.78 if positive,
217 indeterminate, and negative results were considered separately, and 0.95 if indeterminate results
218 were considered as negative.

219

220 **Discussion**

221 Japan is a country with low-level HIV epidemics. The cumulative reported incidence of
222 HIV infection through the end of 2016 was 27,443 [18]. Among them, the number of persons
223 with HIV-2 infection was six [19–22], and there has been no report of HIV-1 and HIV-2 dual
224 infection. According to PMDA, the confirmation of HIV-1 and HIV-2 infections should be
225 performed using WB-1 and WB-2, respectively. However, discrimination between HIV-1 and
226 HIV-2 infections is sometimes very difficult due to cross-reactivity of antibodies against the two
227 viruses. In such cases, it is recommended that the samples are retested from a screening test after
228 several weeks or tested with SERODIA[®]-HIV-1/2 (a particle agglutination assay to detect
229 antibodies to HIV-1 and/or HIV-2, Fujirebio, Tokyo, Japan) or Pepti-LAV 1/2 Assay (an enzyme
230 immunoassay for differentiation of HIV-1 and HIV-2 antibodies, Bio-Rad, Tokyo, Japan) to
231 distinguish HIV-1 and HIV-2 infections, while these differentiation assays also have a high
232 cross-reactivity. These additional tests are, however, laborious, time-consuming, and costly, and
233 cause a large burden in countries such as Japan where the prevalence of HIV-2 infection is
234 extremely low. In this study, we aimed to assess whether a new rapid test Geenius is an effective
235 alternative to WB-1 and WB-2 for confirmation and discrimination of HIV-1 and HIV-2
236 infections.

237 Although the sensitivity of Geenius and NLB 1 was not significantly different (99.3% vs
238 98.6%) for samples from established HIV-1 infections, Geenius gave seven positive results in 20

239 NLB 1 negative or indeterminate samples from acute HIV-1 infections and provided positive
240 results earlier than NLB 1 in two of five seroconversion panels, showing that Geenius is more
241 sensitive than NLB 1. For 140 HIV-1 negative samples including 10 false-positive samples,
242 Geenius gave 136 negative and NLB 1 gave 112 negative results, showing that Geenius is more
243 specific than NLB 1.

244 Cross-reactivity of HIV-1 and HIV-2 antibodies between NLB 1 and NLB 2 was
245 remarkable compared with Geenius. When HIV-1 positive samples were examined, 18 of 144
246 NLB 1 positive samples were also positive with NLB 2. Geenius, however, resolved all of these
247 double-positive samples as HIV-1 positive. An overall discrimination rate of Geenius was 97.7%
248 (172/176) [95% CI, 94.3–99.1] and that of a combinational use of NLB 1 and NLB 2 was 87.5%
249 (154/176) [95% CI, 81.7–91.5], showing that Geenius has a higher discrimination ability than
250 NLB 1/NLB 2. Geenius still gave three HIV positive untypable results: one in 146 HIV-1
251 positive samples and two in 20 HIV-2 positive samples. It is practically impossible to determine
252 if these results reflect HIV-1/2 dual infection or cross-reactivity at present because the
253 application of HIV-2 NAT for confirmation of HIV-2 infection has not yet been established.

254 According to the HIV diagnostic algorithm recommended by CDC, samples that are
255 positive on screening tests but negative or indeterminate on HIV-1/HIV-2 antibody
256 differentiation immunoassay should be tested with an HIV-1 NAT [7]. Because Geenius gave
257 fewer negative or indeterminate results than NLB 1/NLB 2 in HIV-1 positive and HIV-1 false-
258 positive samples (Tables 1, 2, and 5), the use of Geenius will decrease the number of required
259 HIV-1 NAT compared to NLB 1/NLB 2, which may lead to the reduction of testing costs.

260 Geenius is characterized by the cassette involving immunochromatographic components
261 to detect HIV-1/2 antibodies and the automated reader using the proprietary interpretive
262 software. These devices make Geenius have several advantages over WB, including a simple,
263 easy and rapid procedure (within 30 min) and objective interpretation of banding patterns. It is
264 well known that technical skills and interpretation experience are required to perform WB. The

265 rapidity of Geenius may allow HIV testing in public health centers or outreach services to be
266 completed on the same day.

267 WB is frequently used for estimating the stage of early HIV-1 infections [23], based on
268 the study by Fiebid et al. [24], in which positive WB without p31 band is stage V and positive
269 WB with p31 band is stage VI. In this study, Geenius was shown to confirm HIV-1 seropositivity
270 earlier than WB, and thereafter detect p31 bands in panels A and B (Table 2). Keating et al. [25]
271 demonstrated that additional interpretive analysis of band intensities help estimation of recent
272 infections. Development of such algorithms may contribute to epidemiological studies on HIV
273 infections.

274

275 **Conclusions**

276 Geenius is an attractive alternative to WB for confirmation and differentiation of HIV-1 and
277 HIV-2 infections. The adaptation of Geenius to the HIV testing algorithm may lead to a more
278 rapid diagnosis and cost reduction.

279

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283

284 **References**

- 285 1. Miller WC, Rosenberg NE, Rutstein SE, Powers KA. Role of acute and early HIV infection
286 in the sexual transmission of HIV. *Curr Opin HIV AIDS*. 2010;5: 277–282. doi:
287 10.1097/COH.0b013e32833a0d3a PMID: 20543601.
- 288 2. Cohen MS, Chen YQ, McCauley M, Gamble T, Hosseinipour MC, Kumarasamy N, et al.
289 Antiretroviral therapy for the prevention of HIV-1 transmission. *N Engl J Med*. 2016;375:
290 830–839. doi: 10.1056/NEJMoa1600693 PMID: 27424812.

- 291 3. Grinztejn B, Hosseinipour MC, Ribaud HJ, Swindells S, Eron J, Chen YQ, et al. Effects of
292 early versus delayed initiation of antiretroviral treatment on clinical outcomes of HIV-1
293 infection: results from the phase 3 HPTN 052 randomised controlled trial. *Lancet Infect Dis.*
294 2014;14: 281–290. doi: 10.1016/S1473-3099(13)70692-3 PMID: 24602844.
- 295 4. The National Institute of Infectious Diseases, Japan. Voluntary counseling and testing for
296 the diagnosis of HIV infection: update 2012. Available from:
297 <https://www.niid.go.jp/niid/ja/labo-manual.html#class5>. Cited 27 February 2018.
- 298 5. The Japanese Society for AIDS Research. Laboratory testing for the diagnosis of HIV
299 infection: approved guideline 2008. *The Journal of AIDS Research.* 2009;11: 70–72.
300 Available from: http://jaids.umin.ac.jp/journal/journal_vol11_no01_j.html. Cited 27
301 February 2018.
- 302 6. Guan M. Frequency, causes, and new challenges of indeterminate results in Western blot
303 confirmatory testing for antibodies to human immunodeficiency virus. *Clin Vaccine*
304 *Immunol.* 2007;14: 649–659. doi: 10.1128/CVI.00393-06 PMID: 17409223.
- 305 7. Centers for Disease Control and Prevention and Association of Public Health Laboratories.
306 Laboratory testing for the diagnosis of HIV infection: updated recommendations. 2014.
307 Available from: <http://stacks.cdc.gov/view/cdc/23447>. Cited 27 February 2018.
- 308 8. Malloch L, Kadivar K, Putz J, Levett PN, Tang J, Hatchette TF, et al. Comparative
309 evaluation of the Bio-Rad Geenius HIV-1/2 Confirmatory Assay and the Bio-Rad Multispot
310 HIV-1/2 Rapid Test as an alternative differentiation assay for CLSI M53 algorithm-I. *J Clin*
311 *Virol.* 2016;58 Suppl 1: e85–91. doi: 10.1016/j.jcv.2013.08.008 PMID: 24342484.
- 312 9. Abbate I, Pergola C, Pisciotta M, Sciamanna R, Sias C, Orchi N, et al. Evaluation in a
313 clinical setting of the performances of a new rapid confirmatory assay for HIV1/2
314 serodiagnosis. *J Clin Virol.* 2014;61: 166–169. doi: 10.1016/j.jcv.2014.06.015 PMID:
315 25037532.
- 316 10. Hawthorne Hallen A, Samuelson A, Nordin M, Albert J, Bogdanovic G. Evaluation of Bio-
317 Rad Geenius HIV-1 and -2 assay as a confirmatory assay for detection of HIV-1 and -2

- 318 antibodies. *Clin Vaccine Immunol.* 2014;21: 1192–1194. doi: 10.1128/CVI.00153-14 PMID:
319 24943380.
- 320 11. Herssens N, Beelaert G, Fransen K. Discriminatory capacity between HIV-1 and HIV-2 of
321 the new rapid confirmation assay Geenius. *J Virol Methods.* 2014;208: 11–15. doi:
322 10.1016/j.jviromet.2014.07.025 PMID: 25075934.
- 323 12. Montesinos I, Eykmans J, Delforge ML. Evaluation of the Bio-Rad Geenius HIV-1/2 test as
324 a confirmatory assay. *J Clin Virol.* 2014;60: 399–401. doi: 10.1016/j.jcv.2014.04.025 PMID:
325 24932737.
- 326 13. Mor O, Mileguir F, Michaeli M, Levy I, Mendelson E. Evaluation of the Bio-Rad Geenius
327 HIV 1/2 assay as an alternative to the INNO-LIA HIV 1/2 assay for confirmation of HIV
328 infection. *J Clin Microbiol.* 2014;52: 2677–2679. doi: 10.1128/JCM.01184-14 PMID:
329 2478918.
- 330 14. Tinguely C, Schild-Spycher T, Bahador Z, Gowland P, Stolz M, Niederhauser C.
331 Comparison of a conventional HIV 1/2 line immunoassay with a rapid confirmatory HIV 1/2
332 assay. *J Virol Methods.* 2014;206: 1–4. doi: 10.1016/j.jviromet.2014.05.010 PMID:
333 24877900.
- 334 15. Friedrichs I, Buus C, Berger A, Keppler OT, Rabenau HF. Evaluation of two HIV antibody
335 confirmatory assays: Geenius™ HIV1/2 Confirmatory Assay and the recomLine HIV-1 &
336 HIV-2 IgG Line Immunoassay. *J Virol Methods.* 2015;224: 91–94. doi:
337 10.1016/j.jviromet.2015.08.015 PMID: 26315319.
- 338 16. Moon HW, Huh HJ, Oh GY, Lee SG, Lee A, Yun YM, et al. Evaluation of the Bio-Rad
339 Geenius HIV 1/2 confirmation assay as an alternative to Western blot in the Korean
340 population: a multi-center study. *PLoS One* 2015;10: e0139169. doi:
341 10.1371/journal.pone.0139169 PMID: 26422281.
- 342 17. Fordan S, Bennett B, Lee M, Crowe S. Comparative performance of the Geenius™ HIV-
343 1/HI-2 supplemental test in Florida’s public health testing population. *J Clin Virol.* 2017;91:
344 79–83. doi: 10.1016/j.jcv.2017.04.005 PMID: 28434810.

- 345 18. The committee of AIDS surveillance, the Ministry of Health, Labour and Welfare, Japan.
346 The annual AIDS surveillance report 2016, Japan. Available from: [http://api-](http://api-net.jfap.or.jp/status/2016/16nenpo/16nenpo_menu.html)
347 [net.jfap.or.jp/status/2016/16nenpo/16nenpo_menu.html](http://api-net.jfap.or.jp/status/2016/16nenpo/16nenpo_menu.html). Cited 5 March 2018.
- 348 19. Kusagawa S, Imamura Y, Yasuoka A, Hoshino H, Oka S, Takebe Y. Identification of HIV
349 type 2 subtype B transmission in East Asia. *AIDS Res Hum Retroviruses*. 2003;19: 1045–
350 1049. doi: 10.1089/088922203322588413 PMID: 14686325.
- 351 20. Kawahata T, Kojima Y, Mori H, Otake T, Koh KR, Hino M. First identification of a foreign
352 resident with HIV-2 infection in Japan. *Infectious Agents Surveillance Report*. 2004;25:
353 335.
- 354 21. Utsumi T, Nagakawa H, Uenishi R, Kusakawa S, Takebe Y. An HIV-2-infected Japanese
355 man who was a long-term nonprogressor for 36 years. *AIDS*. 2007;21: 1834–1835. doi:
356 10.1097/QAD.0b013e32827b1477 PMID: 17690592.
- 357 22. Ibe S, Yokomaku Y, Shiino T, Tanaka R, Hattori J, Fujisaki S, et al. HIV-2 CRF01_AB:
358 first circulating recombinant form of HIV-2. *J Acquir Immune Def Syndr*. 2010;54: 241–
359 247. doi: 10.1097/QAI.0b013e3181dc98c1 PMID: 20502347.
- 360 23. Ananworanich J, Phanuphak N, de Souza M, Paris R, Arroyo M, Trichavaroi R, et al.
361 Incidence and characterization of acute HIV-1 infection in a high-risk Thai population. *J*
362 *Acquir Immune Defic Syndr*. 2008;49: 151–155. doi: 10.1097/QAI.0b013e318183a96d
363 PMID: 18769355.
- 364 24. Fiebig EW, Wright DJ, Rawal BD, Garrett PE, Schumacher RT, Peddada L, et al. Dynamics
365 of HIV viremia and antibody seroconversion in plasma donors: implications for diagnosis
366 and staging of primary HIV infection. *AIDS*. 2003;5: 1871–1879. doi:
367 10.1097/01.aids.0000076308.76477.b8 PMID: 12960819.
- 368 25. Keating SM, Kassanjee R, Lebedeva M, Facente SN, MacArthur JC, Grebe E, et al.
369 Performance of the Bio-Rad Geenius HIV1/2 Supplemental Assay in detecting “recent” HIV
370 infection an calculating population incidence. *J Acquir Immune Defic Syndr*. 2016;73: 581–
371 588. doi: 10.1097/QAI.0000000000001146 PMID: 27509247.

