

Research Article

Title: Visual modelling validates prey detection by means of diurnal active photolocation in a small cryptobenthic fish

Running Title: Modelling active photolocation

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1 **Abstract**

2 Active sensing has been well documented in animals that use echolocation and electrolocation. Active
3 photolocation, or active sensing using light, has received much less attention, and only in
4 bioluminescent nocturnal species. Recently, however, evidence has suggested the diurnal triplefin
5 *Tripterygion delaisi* uses controlled iris radiance, termed ocular sparks, for prey-detection. While this
6 form of diurnal active photolocation was behaviourally described, a study exploring the complete
7 physical and theoretical process would provide a more compelling case supporting this mechanism. In
8 this paper, we investigate the conditions under which diurnal active photolocation could assist *T. delaisi*
9 in detecting potential prey items. In the field, we sampled prey gammarids (Genus *Cheirocratus*) from
10 foraging substrates, and characterized the spectral properties of their body and eyes, which possess
11 strong reflectors between the ommatidia. In the laboratory, we quantified ocular spark sizes and the
12 angular dependence of their radiance. Together with environmental light measurements and the visual
13 properties of *T. delaisi*, we modeled diurnal active photolocation under various scenarios. Our results
14 corroborate that diurnal active photolocation can help *T. delaisi* detect gammarids at distances relevant
15 to foraging, 4.5 cm under favourable conditions and up to 2.5 cm under average conditions. Because
16 ocular sparks are widespread across many different fish species, diurnal active photolocation for micro-
17 prey may be a common predation strategy.

18 Active sensory systems have been well studied in several animals. For example, the echolocating
19 behavior of bats, by which the reflection of emitted sound waves contributes to navigation in the dark,
20 was detailed starting in 1938 (1, 2), and active electrolocation, by which the disruptions of weak
21 electrical fields are used to detect potential prey and predators, is well known from model organisms
22 such as *Apteronotus leptorhynchus* (3, 4). In contrast, active photolocation, the process by which
23 organisms emit light to survey their environment, seems limited to bioluminescent organisms; only
24 deep-sea dragonfish (Fam. Stomiidae), lanternfish (Fam. Myctidae), and nocturnal flashlight fish (Fam.
25 Anomalopidae) are assumed to use active photolocation (5-7). However, recent evidence suggests
26 active photolocation, by means of controlled light redirection, could also be used in diurnal fish to assist
27 in prey detection, and may be generally common across fish species (8).

28 Michiels et al. (8) described a mechanism that allows the triplefin *Tripterygion delaisi* to redirect
29 ambient light by taking advantage of its laterally protruding lenses and reflective irides, and discussed
30 how this may assist in the detection of camouflaged micro-prey. The central basis of the mechanism is
31 that downwelling light strikes the dorsal part of the eye, is focused by the protruding lens onto the iris
32 below the pupil, and is reflected in the horizontal plane of vision. The focussed light can be radiated by
33 the red fluorescent section of the iris producing a 'red ocular spark', reflected by a blue-white area
34 below the pupil generating a 'blue ocular spark' (**Fig. 1**), or turned on and off by rotating and tilting the
35 iris (see Fig. 2 in (8)). Because downwelling light in the aquatic environment is many times more intense
36 than sidewelling light (9, 10), blue ocular sparks appear much brighter than the background. Michiels et
37 al. (8) emphasized that ocular sparks are too weak to illuminate an entire scene, but suggested they may
38 be sufficiently radiant to reveal strong and/or directional reflectors in nearby target organisms.

39 Indeed, strongly reflecting structures are abundant in aquatic ecosystems, specifically in the
40 eyes of both vertebrates and invertebrates (11-14). For example, camera eyes that possess either a
41 *tapetum lucidum* or *stratum argenteum* are retroreflective, and produce the eyeshine observed when

42 illuminating nocturnal animals. This type of reflected eyeshine is only perceived if the illuminating
43 source is coaxial to the receiver's eye because most of the light is returned to the source in a narrow
44 angle. Furthermore, invertebrates such as stomatopod larvae also possess strong reflectors that can
45 function to camouflage their opaque retinas (12). Though not true retroreflectors, the reflectance of
46 marine invertebrate compound eyes is often stronger towards coaxial alignment (8, 12, 15). Strong
47 directional reflectors and coaxially generated illumination are key components of the mechanism
48 proposed by Michiels et al. (8) because the ocular sparks are generated on the irides, immediately
49 adjacent to the pupil. Thus, ocular sparks could make use of the reflectance of prey eyes to increase the
50 probability of detection, as has been suggested for nocturnal, bioluminescent species (15-17).

51 The experiment reported in Michiels et al. (8) was conducted in the laboratory and focused on
52 ocular spark modulation in response to prey presence and background hue. No studies have yet
53 explored the physical and theoretical basis of the complete process to describe the conditions under
54 which ocular sparks could assist triplefins in detecting prey under natural conditions. In this study, we
55 use simple mathematical expressions and visual modelling to determine the parameters that would
56 enable triplefins to benefit from blue ocular sparks for prey detection. In the field, we collected
57 measurements of ambient light fields and characterised the reflective properties of a background in
58 which gammarids (Crustacea: Amphipoda), important triplefin prey items (18, 19), are found. In the
59 laboratory, we measured the ocular spark properties of the triplefin and the optical properties of the
60 eyes and bodies of gammarids. Finally, we combined these data with triplefins' species-specific visual
61 system characteristics (20) to inform visual models.

62



63

64 **Figure 1.** *Tripterygion delaisi* with blue ocular spark. Photo credit: Nico K. Michiels

65

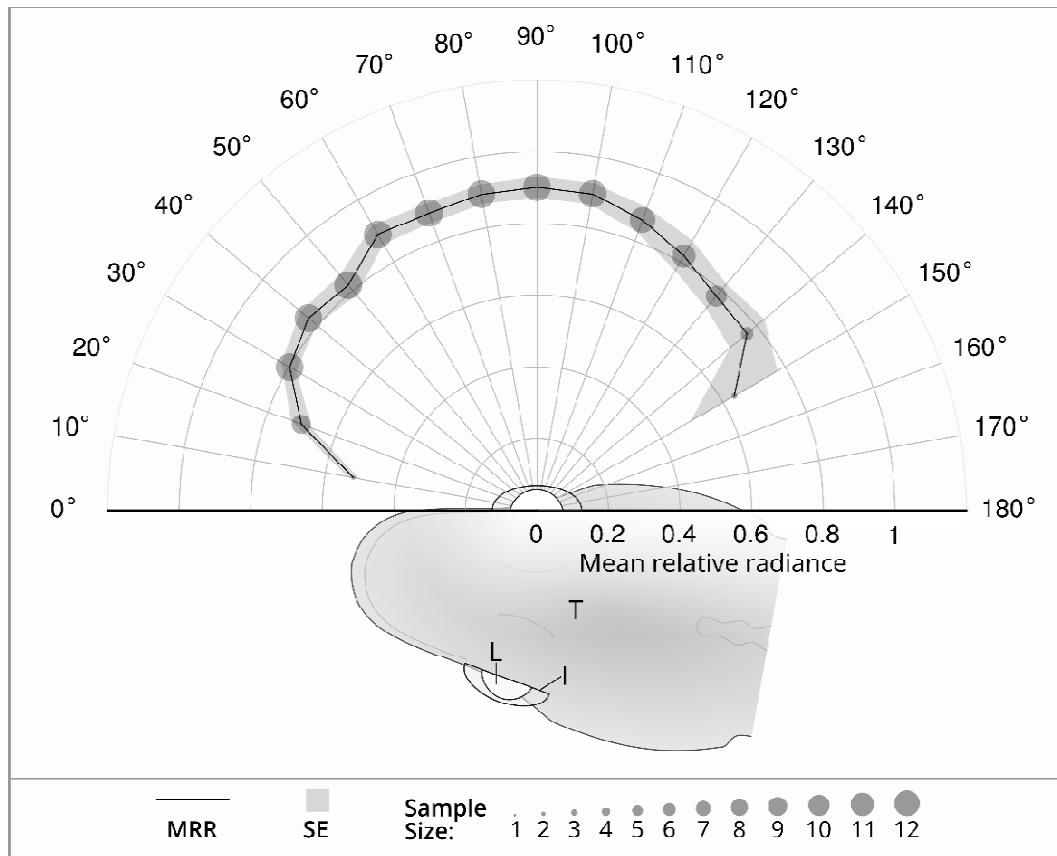
66 **Results and Discussion**

67 *Properties of ocular sparks*

68 Previous work showed that the relative radiance of the average ocular spark peaks around 472 nm at
69 2.15 times that of a diffuse white standard, and that the total area under the curve between 380 and
70 700 nm averaged 1.36 times that of a diffuse white standard (range 0.63 to 2.09, n = both eyes of 5 fish;
71 data from (8)). To further characterize the properties of the blue ocular spark we measured its size, and
72 its radiance at different angles in relation to a white diffuse standard under controlled light
73 environments. The radius equivalent (area as a circular disk) of the ocular sparks ranged from 0.10 mm
74 to 0.24 mm (mean = 0.16 mm, n = 10 fish). The relative radiance of the spark was similar across all
75 angles measured along the equatorial axis (**Fig. 2**). Combined, these results suggest that the
76 chromatophores composing the reflective patch are not strongly specular. Hence, reflectance values
77 higher than 1 were attributed to light being focused onto an area smaller than the lens catchment area.
78 While a narrow beam of energy increases the maximum distance of an active sensing signal, it limits the
79 active space from which animals can gather information, leaving them 'blind' in other directions (21).
80 Hence, directional emission would not be particularly advantageous in an active visual sensing system,

81 as the exact position of the reflector would have to be known. Under these circumstances, a broad
82 active sensing signal would be useful for scanning a large area of the visual environment for strong
83 directional reflectors.

84



86 **Figure 2.** The mean relative radiance (MRR) of the blue ocular spark does not vary along the equatorial axis. MRR is
87 represented by the solid line, the corresponding standard error (SE) by the light grey area, and the sample size of
88 each measured angle by the size of the discs. Fish is seen from a dorsal perspective and naming scheme for angles
89 in relation to the iris of *Tripterygion delaisi*: 90° = normal angle, 0° = angle parallel at the anterior start of the semi-
90 circle and 180° at the posterior end. T = *T. delaisi* body, I = Iris, L = Lens.

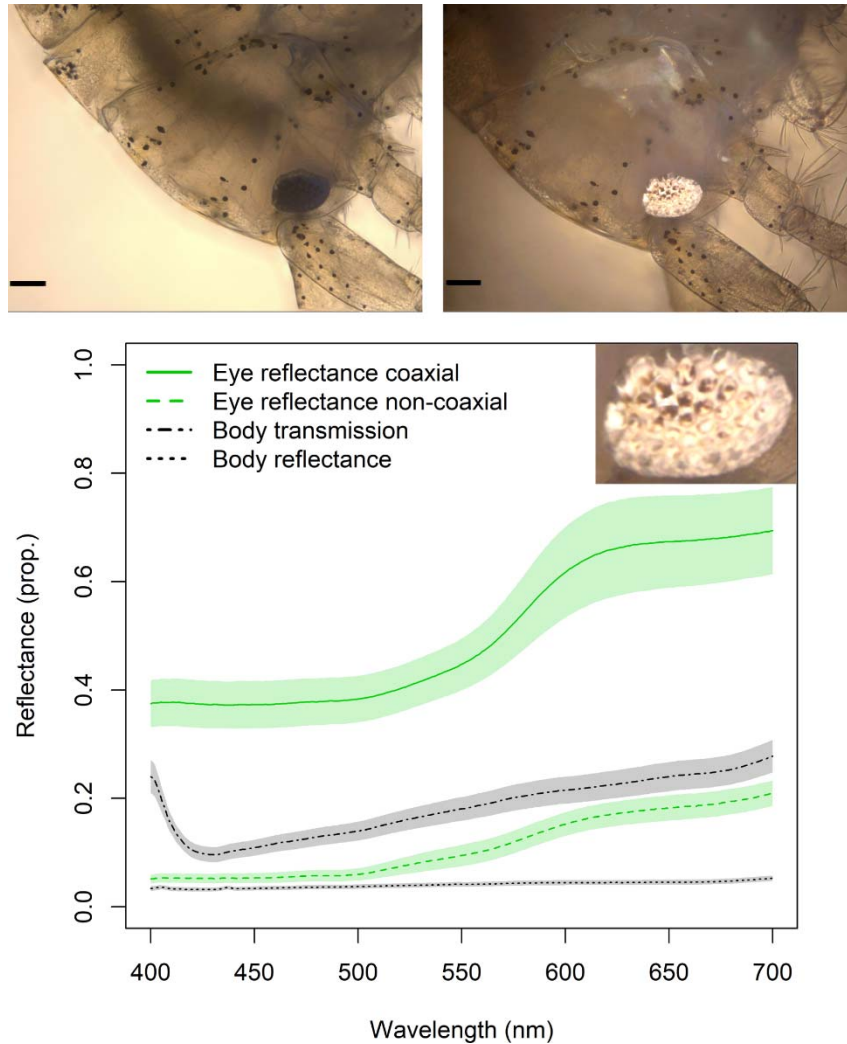
91 *Spectral properties of Cheirocratus gammarids*

92 Our measurements on gammarids eyes (*Cheirocratus* sp.) were collected from the entire compound eye,
93 which is generally reflective in decapod shrimps (15). Focus stacking images revealed the reflective units
94 of gammarid eyes are not found in the optical pathway of the eye (**Fig. 3**), but appear to be between
95 ommatidia akin to those described in *Pullosquilla thomassini*, *Pseudosquilla richeri*, and *Harpisquilla*
96 *sp.* (12). While these reflectors would not improve vision in dim light as other reflectors do (22), they
97 would help camouflage the gammarid eye (12). We measured the reflection and transmission spectra of
98 gammarid body and eyes using a spectroradiometer coupled to a compound microscope (see Methods
99 section for details) under 10×10 magnification ($n = 19$). On average, the body of the gammarids
100 transmitted more light than they reflected, which would make them well camouflaged against any
101 background (**Fig. 3**). Overall eye reflectance, within the 400 to 700 nm wavelength range and illuminated
102 with a coaxial light source (epi-illumination), was on average 4.09 times greater than when illuminated
103 with a light source set at 45° from normal (range = 2.68 to 9.87, $n = 18$; **Fig 3**). The close match between
104 body transmission and non-coaxial eye reflectance further suggests gammarids could hide their eyes
105 against any given background under most light environment scenarios.

106

107 *Triplefin and gammarid eye size*

108 From scaled pictures, we determined that triplefin pupil size averaged 0.78 mm (range 0.66 mm to 0.92
109 mm, $n = 35$ fish, one eye each) and the gammarid eye size averaged 0.0625 mm (range 0.0205 to 0.1020
110 mm, $n = 11$). Based on inter-photoreceptor distances measured from triplefin retinal mounts the visual
111 acuity at the fovea is conservatively set around 6 cycles per degree, meaning that triplefins should be
112 able to resolve the eye of the average gammarid eye from a distance of ~ 48 mm (23).



113

114 **Figure 3.** Top: Example gammarid for which measurements of body transmission and reflectance, as well as eye
115 reflectance were obtained. Top left: viewed under 10 × 10 magnification using transmission illumination, scale bar
116 is 100 μm; Top right: viewed with coaxial illumination, scale bar is 100 microns. Bottom: Reflectance and
117 transmittance of the body ($n = 19$ individuals) and eye ($n = 18$ for coaxial and $n = 10$ for non-coaxial reflectance);
118 lines indicate average of measurements, shaded area indicate standard error of the mean. Inset shows that the
119 highly reflective structures are between ommatidia. Photo credit: Pierre-Paul Bitton.

120

121 *Active photolocation of the gammarid eye*

122 We compared the radiance of gammarid eyes as perceived by triplefins with and without the
123 contribution of a blue ocular spark under various scenarios. Any change in radiance would allow the
124 triplefin to detect potential prey items, e.g. by switching the ocular spark on and off, or when a mobile

125 prey changed the relative position of its eye. To explore the conditions suitable for diurnal active
126 photolocation, we varied four parameters within observed ranges. These four were determined to affect
127 active photolocation most strongly (see Methods). (1) **Spark size**: Because the photon flux that reaches
128 the gammarid eye is directly related to the solid angle subtended by the ocular spark, we varied its
129 radius equivalent on a continuous scale from 0.09 to 0.25 mm. (2) **Spark radiance**: The photon flux
130 reaching the gammarid eye is also proportional to the relative radiance of the ocular spark so we varied
131 it on a continuous scale from a mean area under the curve of 0.63 to 2.09. (3) **Gammarid eye**
132 **reflectance**: The difference in gammarid eye radiance with and without the contribution of an ocular
133 spark depends on the relationship between the reflective properties of the gammarid ocular reflectors
134 under coaxial illumination and non-coaxial illumination. The non-coaxial component is used to estimate
135 how bright the eye is under the prevailing conditions, without the addition of an ocular spark; the
136 coaxial reflectance is used to calculate the contribution of the ocular spark to the total gammarid eye
137 radiance. We evaluated the impact of the relationship between the coaxial and non-coaxial reflectance
138 of gammarid ocular reflectors using three categories: large difference (non-coaxial reflectance is 9.87
139 times weaker than coaxial reflectance; maximum observed), average difference (4.09 times weaker),
140 and small difference (2.68 times weaker; minimum observed). (4) **Shading of prey**: Finally, redirecting
141 downwelling light into the horizontal plane would allow triplefins to generate greater contrasts with
142 greater shading of prey, while the triplefin remains exposed to the same downwelling light. We
143 investigated the influence of prey shading using four categories: no shade, weakly shaded, average
144 shade, and strongly shaded (see Methods).

145 For each set of conditions, we calculated the maximum prey detection distance by means of
146 active photolocation by calculating the chromatic and achromatic contrast between the gammarid eye
147 with and without the radiance induced by a blue ocular spark as perceived by the triplefin at different
148 distances (range 0.5 – 4.5 cm), and by comparing these values with specific chromatic and achromatic

149 contrast thresholds. The range of distances used is relevant to triplefin feeding behaviour, as it normally
150 targets prey items from one to three cm (personal observations). For chromatic contrast calculations we
151 used the receptor-noise limited model (24) parameterized using triplefin-specific visual characteristics
152 (20, 23), with a Weber fraction set at 0.05 (25, 26). For achromatic contrasts, we followed the
153 calculations in Siddiqi et al. (27) using a realistic Weber fraction of 0.02 (unpublished data; (28)), and a
154 conservative fraction of 0.05. To verify that the radiance from the ocular spark does not concurrently
155 change the luminance of the body of the gammarid, we also performed all calculations using body
156 transmittance and reflectance, assuming the gammarid was resting on the algal species *Halopteris*
157 *filicina*, a common triplefin foraging substrate (19).

158 Using a realistic Weber fraction of 0.02, the results from our models show that diurnal active
159 photolocation would assist with micro-prey foraging under wide ranging conditions (**Fig. 4**) by
160 generating perceivable achromatic contrasts in the eye of gammarids when modulating the ocular spark.
161 Chromatic contrast calculations did not yield values above six mm and are therefore considered
162 ineffective for gammarid detection using blue ocular sparks (results not shown). Neither achromatic nor
163 chromatic contrast calculations created perceivable contrasts on gammarid bodies (no detection
164 distance above five mm, results not shown). A conservative Weber fraction of 0.05 limited the
165 parameter space under which active photolocation based on achromatic contrasts would be beneficial
166 (**Fig. S1**), but demonstrated nonetheless the potential for ocular sparks to enhance prey detection at
167 distances relevant to triplefin foraging behaviour.

168 Under the most favourable conditions, the ocular spark could generate detectable achromatic
169 contrasts at 45 mm distance, the maximum modelled. This distance represents almost a full body length
170 of an average sized triplefin (29) and is much longer than the average striking distance (personal
171 observations: NKM, P-PB, MS, UKH). Under unfavourable parameter combinations, diurnal active
172 photolocation would generate perceivable achromatic contrasts at less than 10 mm, limiting its

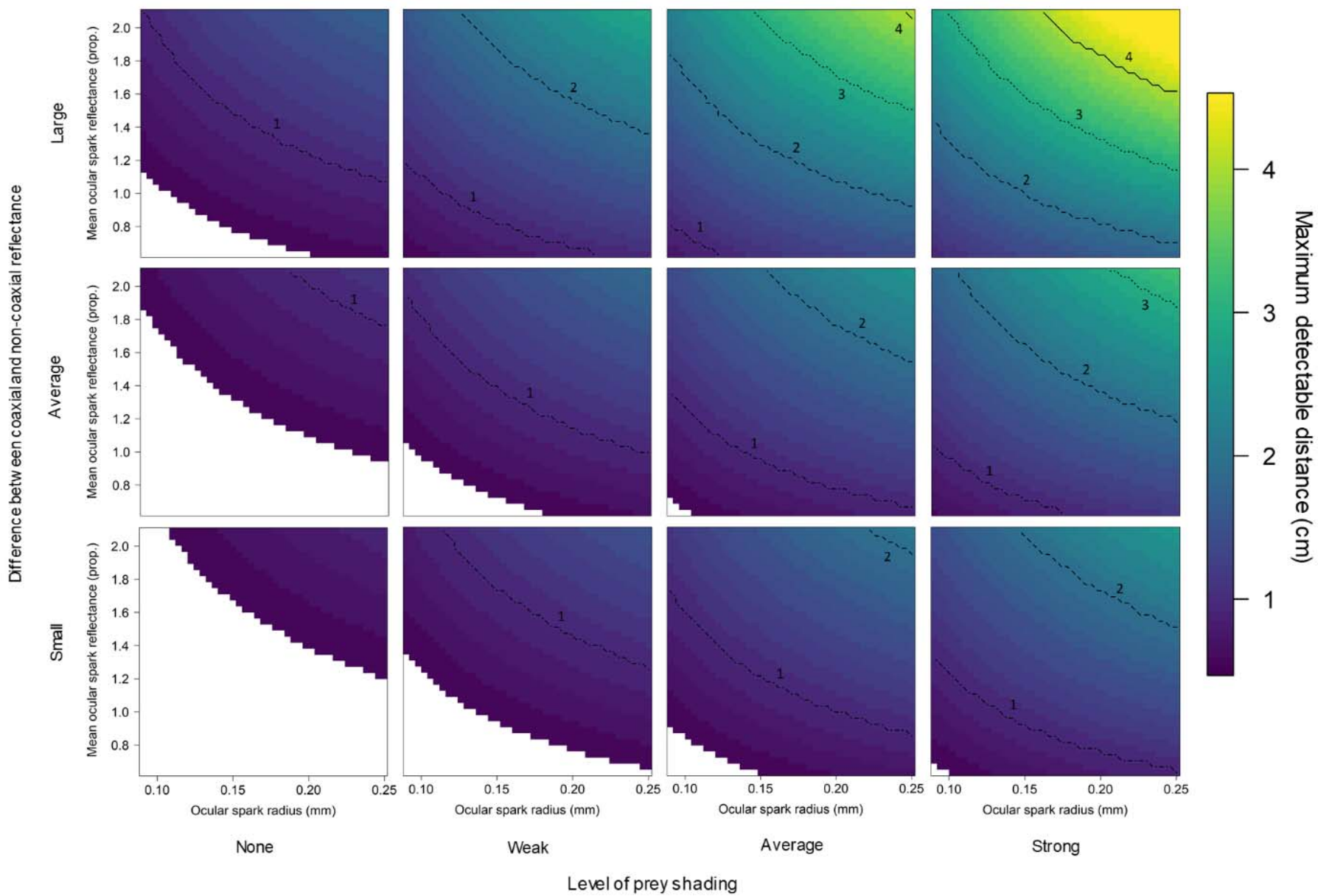
173 potential to increase prey detection. In general, diurnal active photolocation would not be beneficial
174 when triplefins forage on unshaded substrates (**Fig. 4** No shade). Under this scenario, only large and
175 bright ocular sparks, and strong coaxial reflectance of gammarid eyes, would generate perceivable
176 achromatic contrasts at distances > 10 mm. Even in poorly shaded areas, however, the ocular spark
177 would generate perceivable contrast in the eye of gammarids at > 10 mm. When foraging on average or
178 heavily-shaded substrate (**Fig. 4** third and fourth column), the distance at which active photolocation
179 would be beneficial would greatly depend on the relationship between the coaxial and non-coaxial
180 reflectance properties of the gammarid eyes. Under these shaded conditions, maximum detectable
181 distances of over 15 mm would be common, suggesting diurnal active photolocation is effective in many
182 situations. These results are in agreement with observations of triplefin foraging behaviour; they are
183 often found feeding at small micro-habitat structures (e.g., complex algal growth, encrusting epi-growth,
184 etc.).

185 The size of the ocular spark had a large effect on the model, simply because the amount of light
186 striking the gammarid eye is strongly dependent on the perceived size of the spark from the gammarids'
187 perspective. However, producing larger sparks may not be possible or beneficial. The evolution of the
188 size of the reflecting chromatophore patch on which the spark is focused, and therefore the photon
189 radiance available for active photolocation, is probably constrained by two factors. First, the maximum
190 amount of light that can be directed towards the chromatophore is limited by the catchment area of the
191 lens, which depends on the size, position and degree of protrusion through the pupil. This positioning is
192 likely to be driven much more by regular vision than active photolocation. Second, *T. delaisi* is a crypto-
193 benthic species which has evolved colour patterns particularly well suited for camouflage. Generating a
194 large, highly visible spark could become a disadvantage if it attracted potential predators. Indeed, larger
195 piscivorous fish are known to be attracted to brighter lures (30) and several such species are common in
196 the same habitat (e.g. Fam. Serranidae).

197 The relationship between the coaxial and non-coaxial reflectance properties of the gammarid eye
198 also had much influence on the maximum detection distance. The reflectors, as those of stomatopod
199 larvae (12), are of unknown origin but are likely quarter-stack multilayers of dielectric material with
200 different refractive indices (11). As in other photonic crystals, such as 2D crystals (31, 32) and thin-films
201 (33), these particular reflectors usually have much stronger reflection at normal incidence. The
202 relationship between coaxial and non-coaxial reflectance could then be a function of the number of
203 layers, their spacing, and regularity.

204 Overall, our results describe how active photolocation through blue ocular sparks in the diurnal
205 triplefin *Tripterygion delaisi* could assist in the detection of prey items at relevant foraging distances.
206 Measurements of the red ocular spark (8) show that these are overall weaker, but may perhaps
207 generate a chromatic contrast in blue rich light environments, such as those found at the greater depths
208 of *T. delaisi*'s ecological range. We conclude that diurnal active photolocation by means of ocular sparks
209 can supplement regular vision by making the highly reflective eye of potential prey targets shine under
210 nearly-coaxial illumination. Given the high number of fish species that have both protruding lenses and
211 highly reflective irides, active photolocation could be widespread among fish, and an important, yet
212 previously disregarded, vision enhancement mechanism.

213



215 **Figure 4.** Maximum detection distances of the eye of gammarids by means of blue ocular spark reflectance under
216 varying scenarios with Weber fraction set at 0.02. Top, middle, and bottom rows were obtained by varying the
217 relationship between the reflectance of gammarid eyes with coaxial epi-illumination and at 45° from normal.
218 Columns represent four scenarios of shading in which the prey item is located (See Material and Methods).
219 Conditions in which active photolocation would not assist in gammarid detection are in white.

220

221 **Materials and Methods**

222 To determine whether ocular sparks produced by *Tripterygion delaisi* could generate a perceivable
223 contrast in the eyes of gammarids, common triplefin prey items (18), we modelled interactions between
224 prey and predator while varying influential parameters. The parameter space under which diurnal active
225 photolocation may function was explored by (1) quantifying the downwelling and sidewelling light fields at
226 various locations within *T. delaisi* ecological ranges, (2) measuring the reflective properties of the most
227 common foraging substrate (*Halopteris filicina*), (3) measuring the size, and angle dependence of ocular
228 sparks, and (4) measuring the optical properties of gammarid eyes and bodies. We combined these data
229 to known properties of *T. delaisi*'s visual system (20) to calculate chromatic and achromatic contrast
230 between the radiance of gammarid eyes as perceived by a *T. delaisi* individual with and without the
231 radiance of its own blue ocular spark.

232

233 *Ocular spark radiance*

234 Radiance of the ocular spark was measured in live fish as described in Michiels et al. (8). To determine if
235 the radiance of ocular sparks is equal in all directions, we collected angle-resolved measurements by
236 securing whole triplefins, previously sacrificed by severing the spinal cord, in the center of a platform in
237 a stainless steel hemisphere of 15 cm diameter placed on a PVC ring holder inside a 7 l Plexiglass®
238 cylinder filled with fresh marine Ringer-solution. The reflective chromatophore patch responsible for
239 generating the sparks was positioned at the exact center of the hemisphere, which was also the exact
240 centre of cylinder, allowing measurements normal to the cylinder wall at all angles. Sparks were

241 generated by means of a stage lamp (ARRI® 650 Plus) mounted ~1.5 m above the fish. To avoid ambient
242 light effects, the room was otherwise kept dark. For each of the 12 fish, the radiance of the ocular spark
243 was measured with a Spectrascan PR-740 (PhotoResearch Inc., Syracuse, USA) fitted with a MLH-10X
244 lens (Computar®) at each 10° between 10° (anteriorly) and 150° (posteriorly) in relation to the frontal-
245 caudal axis of the fish's body (**Fig. 2**). The PR-740, uses Pritchard optics to collect measurements of
246 absolute radiance of a specific solid angle, which is visualized as a small black circular area in the
247 viewfinder. These values were expressed relative to the radiances of a polytetrafluorethylen (PTFE)
248 diffuse white standard (Berghof Fluoroplastic Technology GmbH, Eningen unter Achalm, Germany)
249 measured at the same angles and position immediately after each fish. The range of angles was not
250 covered for all fish explaining why the sample size varied between angles (**Fig. 2**). Because the lens'
251 resting state following death is slightly retracted, these measurements could only be used for comparing
252 relative radiance at various angles as they underestimate spark intensity in relation to the illuminant.
253 The size of the ocular spark was determined in 10 fish from scaled images analysed using ImageJ (34).

254

255 *Gammarid spectroradiometry*

256 Gammarids were isolated from *Halopteris filicina* algae collected between 5 and 10 m depth at STARESO
257 (Calvi, Corsica), and immobilized but kept alive using a 0.6 M MgCl₂ solution. Spectral measurements
258 were obtained with a PR-740 spectroradiometer mounted onto a Leica DM5000 B compound
259 microscope (Leica Microsystems, Wetzlar, Germany) under 10 × 10 magnification. For reflectance
260 measurements, we used an external halogen light source (KL2500 LCD, Schott AG, Mainz, Germany),
261 either incident through the microscope's housing (epi-illumination) or at 45° to the sample using an
262 external LLG 380 liquid light guide (Lumatec GMBH, Germany). For each gammarid we collected five
263 body and eye reflectance measurements coaxially illuminated, and five measurements of eyes
264 illuminated at 45°. We did not collect body reflectance at 45° because there was no evidence of coaxial

265 specularly or iridescence. The gammarid eye measurements were obtained from areas that covered
266 almost the entire eye under 10 x 10 magnification. A submerged PTFE standard was also measured five
267 times both with epi-illumination and with the light source at 45°. In all cases, the sample was
268 repositioned and refocused before each measurement. Averages of 5 measurements of the body and
269 eyes were expressed in relation to their relative standard. For transmission measurements we used the
270 12 V 100 W halogen lamp provided with the microscope in the transmitted light axis. For each
271 gammarid, we took five radiance measurements of the transmitted light as seen through haphazardly
272 selected locations on the body (plus Petri dish and MgCl₂ solution) and five measurements of the
273 transmitted light without the gammarid (but including the Petri dish and MgCl₂ solution). Transmittance
274 was then determined as the mean of the five measurements of the body divided by the reference.
275 Scaled images of the gammarids were also obtained at this time and the size of the eyes subsequently
276 estimated using ImageJ (34).

277

278 *Field light environments and background reflectance*

279 We measured the reflective properties of *Halopterus filicina*, a common foraging substrate for *T. delaisi*,
280 and the downwelling light, unshaded sidewelling light, and shaded sidewelling light of triplefin habitat at
281 the Station de Recherches Sous-marines et Océanographiques (STARESO) in Calvi (Corsica, France) in
282 June-July 2014 and 2017. Details of *Halopterus filicina* data collection protocol can be found in Harant et
283 al. (19). In short, substrate data were collected while scuba diving at a shallow site (5 m) characterized
284 by rocky slopes, steep walls and granite boulders. Measurements were obtained at various locations in
285 conjunction with a PTFE diffuse white standard (DWS) tilted at 45° to the surface as a combined
286 measure of downwelling and sidewelling light. The relative radiance between the substrate
287 measurements and light field is considered as the reflective property of *Halopterus filicina*. Light field
288 measurements were instead obtained between 2 and 30 m depth on substrates facing south. At each

289 depth (2, 4, 6, 8, 10, 14, 18, 24, and 30 m) we measured from a 45° angle the radiance of an exposed
290 PTFE standard set at normal incidence to the water surface (= angle of incidence 0°) for an
291 approximation of downwelling light, and from a 90° a PTFE standard set at 90° to normal for measuring
292 sidewelling light, and a PTFE standard set at 90° to normal and shaded by a 4 cm opaque black cover as a
293 measure of shaded sidewelling light environment. Three measurements were obtained for every
294 standard at every depth and averages used in analyses. All measurements were obtained using a PR-740
295 fixed at a focal distance of 50 cm in a custom built underwater housing (BS Kinetics, Achern, Germany).
296 The PR-740 was equipped with a colour correction filter (#287 double CT orange, LEE Filters, Andover,
297 England) which suppresses, but does not block the dominant blue-green spectral range. This increases
298 exposure time, allowing the instrument to obtain better readings in the weak, long-wavelength part of
299 the spectrum at depth. Radiance measurements were corrected for the transmission profile of the filter
300 and port of the housing before being used in the calculations.

301

302 *Gammarid eye photon flux with and without ocular sparks*

303 We modeled a three-dimensional interaction between triplefins and gammarids, both on the same
304 horizontal plane, and assuming their eyes were positioned at normal incidence. We calculated the
305 photon flux of the reflective eye of the gammarid, as perceived by the triplefin, with and without the
306 contribution of the ocular spark, by describing the interaction in simple equations (see complete
307 calculation details in SI). In short, the photon flux of the gammarid eye without the contribution of the
308 spark reaching the triplefin retina was determined by the sidewelling light reflected by the ocular
309 reflectors (non-coaxial), the solid angle subtended by the gammarid eye (in steradians) as perceived by
310 the triplefin as a function of the distance between the two eyes, and the area of the triplefin pupil as the
311 ultimate receptor area (measured in ImageJ from scaled images (34)). The photon flux due to the ocular
312 spark returned to the triplefin was further determined by the radiance of the ocular spark, the solid

313 angle of the ocular spark (in steradians) from the perspective of the gammarid eye, and the coaxial
314 reflective properties of the gammarid's ocular reflectors. All solid angles were calculated using Monte
315 Carlo simulations (35).

316 In the model, we used fixed mean values for parameters that had little influence on the results,
317 based on preliminary sensitivity analyses. We set the gammarid eye radius at 0.0625 mm, the triplefin
318 pupil radius at 0.78 mm, used the downwelling light profile measured at 10 m, the mean background
319 substrate reflectance (*Halopteryx filicina*), and the mean reflectance and transmittance of the gammarid
320 body. We explored the parameter space of the possible prey-predator interactions by varying the
321 factors that were determined to have the most influence on the contrast generated by the ocular spark
322 in the gammarid eye (parameter range and calculation details in Supplemental Information). We varied
323 the ocular spark radius measured in the laboratory (continuous range from 0.09 to 0.25 mm in intervals
324 of 0.004 mm), the ocular spark relative radiance (continuous from an area under the curve of 0.63 to
325 2.09 in relation to a white standard), the relationship between the coaxial and non-coaxial reflectance of
326 gammarid eyes (three categories: low difference based on the minimum observed, average difference
327 based on the measured mean, and large differences based on the maximum observed), and the
328 relationship between the downwelling and sidewelling light field (four categories: no shade, weakly
329 shaded, average shade, and strongly shaded). The 'no shade' was calculated as the average of the non-
330 shaded sidewelling light measurements divided by the average downwelling light, and the three shaded
331 categories were calculated as the minimum, average, and maximum observed shaded sidewelling light
332 measurements divided by the average downwelling light.

333 The spatial resolution of *T. delaisi* is conservatively estimated at 6 cycles/degree (23) which
334 means that the average gammarid eye diameter (0.125 mm) becomes a point source at ~48 mm. To
335 avoid modelling situations in which the gammarid eye is smaller than the smallest detectable point in
336 space by a triplefin, we limited the distance between the triplefin and the gammarid to a maximum of

337 45 mm. The minimum distance modelled relied on estimates of the distance of nearest focus (~5 mm;
338 based on calculations in (23)).

339

340 *Calculation of chromatic and achromatic contrasts*

341 Using retinal quantum catch estimations based on calculated photon flux, we calculated the differences
342 in chromatic and achromatic contrasts between the radiance of the gammarid eye with and without the
343 contribution of the ocular spark radiance. For the chromatic contrast we used the receptor-noise model
344 (Vorobyev and Osorio 1998), informed using the visual system characteristics of *T. delaisi* presented in
345 (20). In short, we used species specific ocular media transmission values, photoreceptor sensitivity
346 curves based on the single cone (peak at 468 nm), and the double cone (peaks at 517 and 530 nm)
347 following a vertebrate photoreceptor-template (36), and a relative photoreceptor density of single to
348 double cones set at 1:4 as found in the triplefin fovea (23). Since the Weber fraction (ω) for colour
349 contrast is not known for fish, we used a value of 0.05 as in previous studies from other groups (25, 26).

350 We calculated the achromatic contrast as $\log(Q_1/Q_2)/\omega$, where Q_1 and Q_2 are the quantum catches of
351 the two members of the double cones which are associated with the achromatic channel, under photon
352 flux₁ and photon flux₂ respectively (27). We used two different Weber fractions for our calculations: a
353 conservative value for fish ($\omega = 0.05$) according to work conducted on *Carassius auratus* (Hester 1968),
354 *Scardinius erythrophthalmus* (37), *Gadus morhua* (38), and *Lepomis macrochirus* (39), and the mean
355 value of these studies ($\omega = 0.02$ (28)). Recent work by our unit confirmed that triplefins have contrast
356 sensitivity at or below $\omega = 0.02$ (in prep.). Since this parameter is extremely influential in the calculation
357 of contrasts (40, 41), and one of the Weber fractions selected is conservative, our achromatic contrast
358 values also include conservative estimates. Both calculations of chromatic and achromatic contrast
359 result in measures of just-noticeable differences (JNDs), where values above one are considered to
360 represent the minimum discernable differences between the quantum catches. To ensure that the

361 contrast generated by the ocular spark was only influencing the radiance of the gammarid eye and not
362 the background, we performed the same calculations but for the gammarid body.

363

364 *Calculation of maximum discernable distance*

365 For each set of model conditions defined in the section *Gammarid eye photon flux with and without*
366 *ocular sparks*, we determined the maximum discernable distance of the ocular spark radiance returned
367 by the gammarid eye. This was achieved by calculating the chromatic and achromatic contrast at each
368 millimeter between 5 and 45 mm per set of conditions, and extracting the first value at which the
369 contrast was equal to or exceeding 1.0 JND (**Fig. S2**).

370

371 *Animal care and permits*

372 Fish were caught at STARESO between 5 and 20 m depth using hand nets while scuba diving. During
373 dives, fish were transported in 50 ml perforated Falcon™ tubes (Corning Inc, NY, USA) to permit water
374 exchange. At the field station the fish were held in a 50 L flow-through tank at ambient water
375 temperature, until transferred to facilities at the University of Tübingen, Germany. In these facilities,
376 individuals were kept separately in 15 L flow-through tanks (18°C, salinity 34‰, pH 8.2, 12 L: 12 D light
377 cycle) and fed once per day. The fish were sacrificed under permit 'Mitteilung 29.10.2014' from the
378 Regierungspräsidium (Referat 35, Konrad-Adenauer-Str. 20, 72072 Tübingen) under the supervision of
379 the animal welfare officer. We captured triplefins in STARESO under the station's general scientific
380 permit.

381

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389

390 *Data Archival*

391 All data and R scripts used in the analyses and preparation of figures will be made available on Dryad
392 upon acceptance, and are available to the editors and reviewers during the evaluation process.

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487 Supplemental Information

488

489 **Table S1.** Symbols used in the equations to calculate the photon flux of the gammarid eye reaching the
490 triplefin, with and without the contribution of the ocular spark

Symbol	Definition
L	Photon radiance (photons $s^{-1} sr^{-1} m^{-2}$)
S	Blue ocular spark relative radiance (proportion of a PTFE white standard)
d	Distance between triplefin and gammarid eyes (m)
r_t	Radius of triplefin pupil (m)
R_{ca}	Reflectance of gammarid eye (<u>co</u> axial) (proportion of a PTFE white standard)
R_{nca}	Reflectance of gammarid eye (<u>non-co</u> axial) (proportion of a PTFE white standard)
Φ	Photon flux coming from the gammarid eye reaching the triplefin pupil (photons s^{-1})
Ω	Solid angle of target as perceived by receiver (sr)

491

492

493 Photon flux calculations

494 We calculated the photon flux of the gammarid eye reaching the triplefin pupil with and without the
495 radiance of the ocular spark, assuming that the center of the triplefin pupil was at normal incidence to
496 the center of the eye of the gammarid, i.e. the full area of the pupil of the triplefin is visible to the
497 gammarid and vice versa. We also assume the effect of absorbance and scattering of the water to be
498 negligible since all energy transfers occur over distances shorter than 5 cm.

499

500 *Photon flux without ocular spark*

501 The base photon radiance of the gammarid eye (L_0) is a function of the sidewelling light field (L_{sw}) and
502 the reflectance of the gammarid eye with non-coaxial illumination:

503
$$L_0 = L_{sw} \times R_{nca} \quad (1)$$

504

505 The photon flux reaching the retina of the triplefin without the ocular spark (Φ_{ns}) is the proportion the
506 gammarid radiance multiplied by the solid angle of the gammarid eye (Ω_{gam}) and the area of the
507 triplefin pupil (πr_t^2):

508
$$\Phi_{ns} = L_0 \times \Omega_{gam} \times \pi r_t^2 \quad (2)$$

509

510 *Photon flux produced by ocular spark*

511 The photon radiance of the ocular spark (L_{os}) is determined by the downwelling light field (L_{dw}), the
512 catchment area of the lens, and the reflective properties of the iris chromatophores on which the light is
513 focused. The effect of the lens and reflective properties of the chromatophores have only been
514 measured together and are treated as a relative radiance value (S).

$$515 \quad L_{os} = L_{dw} \times S \quad (3)$$

516

517 The radiance of the gammarid eye (L_{gam}) caused by the reflection of the ocular spark is estimated by
518 multiplying the radiance of the ocular spark reaching the gammarid (L_{os}) with the solid angle of the
519 ocular spark (Ω_{os}) and the reflectance of the gammarid eye with illumination coaxial to the receiver
520 (R_{ca}). Because the properties of the gammarid eye are measured in relation to a diffuse white standard,
521 the photon exitance from the gammarid eye is converted to photon radiance by dividing by π
522 steradians:

$$523 \quad L_{gam} = L_{os} \times \Omega_{os} \times R_{ca} \times \pi^{-1} \quad (4)$$

524

525 The photon flux generated by the ocular spark which reaches the triplefin retina (Φ_{os}) is determined as
526 the proportion of the ocular spark generated gammarid eye radiance (Eq. 4) multiplied by the perceived
527 size of the gammarid eye, in steradians, and the area of the triplefin pupil:

$$528 \quad \Phi_{os} = L_{os} \times \Omega_{os} \times R_{ca} \times \pi^{-1} \times \Omega_{gam} \times \pi r_t^2 \quad (5)$$

529

530 The total photon flux reaching the retina of the triplefin with the ocular spark is then the sum of
531 equations (2) and (5).

532 A similar calculation was used for the effect of the ocular spark on the illumination of the
533 gammarid body. In these calculations we estimated the photon flux reaching the retina of the triplefin
534 with and without the contribution of the ocular spark, using the same solid angles. In contrast to
535 calculations with the gammarid eye, we used the same body reflectance values for the coaxially and
536 non-coaxially illuminated scenarios. The photon exitance from the body, both with and without the
537 contribution of the ocular spark was determined as the proportion of light that was reflected by the
538 body and the proportion of light that was transmitted through the body, reflected by the substrate, and
539 transmitted again through the body.

540

541 For all calculations, the solid angle of the gammarid eye from the perspective of the triplefin
542 pupil (Ω_{gam}), and the solid angle of the ocular spark from the perspective of the gammarid eye (Ω_{os}),
543 in steradians, were estimated by Monte Carlo simulation (35). The triplefin pupil, gammarid eye, and
544 ocular spark were treated as disks of zero thickness. The pupil and gammarid eye were always
545 positioned centered and at normal incidence to one another, and the ocular spark positioned at the
546 edge of the iris (displacement = 1.09 mm) in the same plane and normal vector as the triplefin pupil.
547 Because we estimate that the triplefin can focus on objects minimally at 7 mm and that average
548 gammarid eye becomes a point source beyond ~48 mm, we determined the solid angles for distances
549 between 5 mm and 45 mm. The calculations were based on 1E09 photon packets emitted from the
550 source; these generated solid angle estimates with 99.9% confidence intervals with errors ranging from
551 1.2 % of the solid angle value at 5 mm to 10.6 % at 45 mm.

552

553 Exploration of parameter space

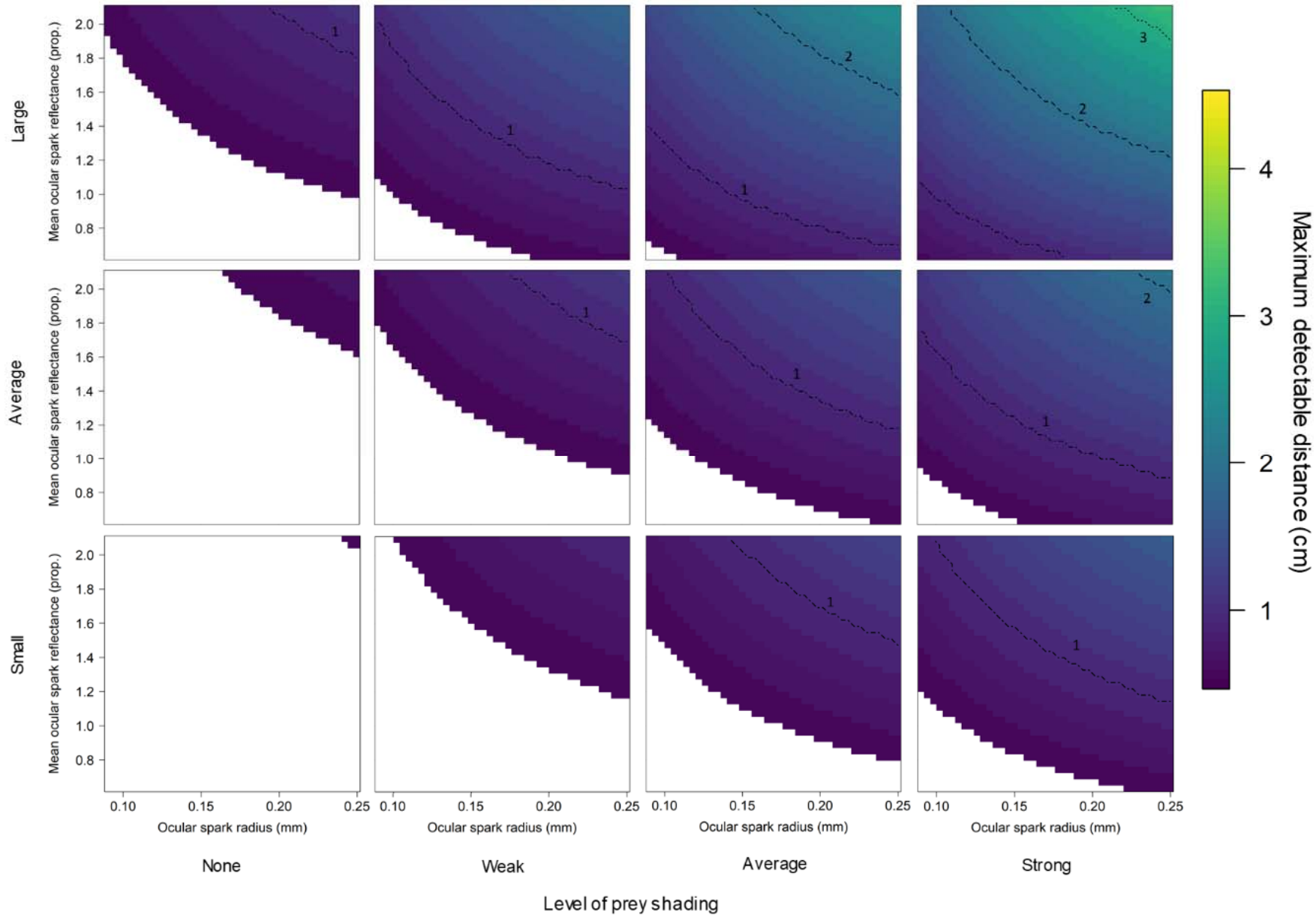
554 To explore the parameter space of our interaction between triplefins and gammarids, we varied the
555 parameters known to have the most influence on the calculated contrasts. To allow comparison and
556 visualization of the results, we chose to model two continuous parameters: ocular spark radius, and the
557 ocular spark relative radiance, and two categorical parameters: the relationship between the coaxial and
558 non-coaxial reflectance of the gammarid eyes, and the relationship between the downwelling and
559 sidewelling light field.

560 The parameter 'ocular spark radius' ranged from 0.09 mm to 0.25 mm (based on actual
561 measurements ranging from 0.10 mm to 0.24 mm) in 41 intervals of equal increments (0.004 mm). The
562 range values for the parameter 'ocular spark relative radiance' was produced by first taking the mean
563 value of all measurements at each wavelength (binned in 1 nm interval) and varying the area under the
564 curve between the measured range of 63 % to 209 %. To produce square matrices of results, the value
565 range was also divided in 41 intervals of equal increments.

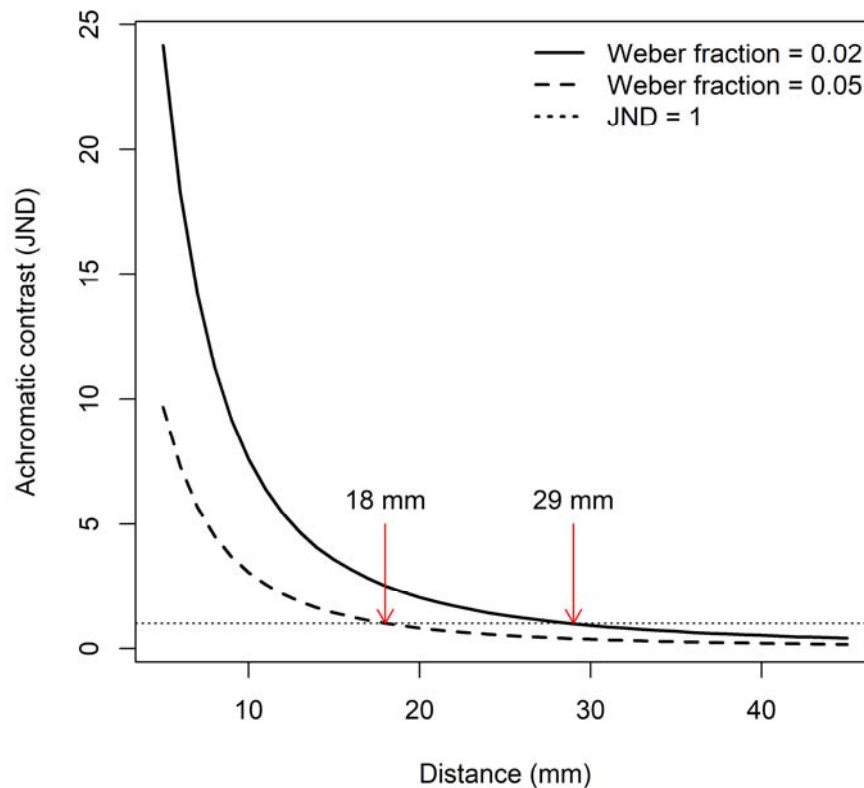
566 The relationship between the coaxial and non-coaxial reflectance of gammarid eyes was not
567 correlated in the samples measured. To explore the influence of this parameter we calculated the
568 average difference between the coaxial and non-coaxial eye reflectance measurement obtained from
569 each gammarid, calculated at each wavelength (binned in 1 nm interval), and varied the area under the
570 curve to represent the minimum value observed (10.1 %), the average value (24.4%), and the maximum
571 value observed (37.25 %).

572 We included four measures of the relationship between the downwelling and sidewelling light
573 fields: no shade, weakly shaded, average shade, and strongly shaded. The relationship between the
574 downwelling and unshaded sidewelling spectral light profile was obtained by taking their ratio at several
575 measured locations. The three categories of shaded sidewelling light were obtained by calculating the
576 average difference between the downwelling and shaded sidewelling light fields at each measurement
577 station, and varying the area under the curve to represent the minimum value observed (ratio DW/SW =
578 8.65), the average value (ratio = 16.62), and the maximum value observed (ratio = 26.63). These
579 conversion vectors were then applied to the downwelling light field obtained at 10 m depth.
580

Difference between coaxial and non-coaxial reflectance



582 **Figure S1.** Maximum detectable distances of ocular spark reflectance from the eye of gammarids under varying
583 scenarios (Weber fraction = 0.05). Top, middle, and bottom row were obtained by varying the relationship
584 between the reflectance of gammarid eyes with coaxial epi-illumination and at 45° from normal. Vertical rows
585 were obtained by varying the amount of shade on which prey items rests. Conditions in which active photolocation
586 would not assist in gammarid detection are in white.
587



588
589 **Figure S2.** Example extrapolation of the maximum distance at which reflections in the gammarid eye caused by
590 ocular spark radiance are discernable. The achromatic contrast is the perceived difference in photon flux from the
591 gammarid eye with and without ocular spark contribution. The maximum discernable distance is defined as the
592 distance at which the contrast is equal to one just-noticeable-difference (JND). All calculations were repeated
593 twice, once with the Weber fraction set at 0.05, once with the Weber fraction set at 0.02 (See Material and
594 Methods).
595