1 Effects of planting density on the growth and photosynthetic characteristics of Alternanthera

2 *philoxeroides* under different nutrient conditions

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23 Abstract

24	Density and nutrient level are important factors that might affect the growth of invasive
25	plants. To reveal the effects of plant density on the performance of invasive plant Alternanthera
26	philoxeroides under different nutrient conditions, a greenhouse experiment was conducted in
27	which A. philoxeroides was planted at three densities (low, medium and high) under three
28	nutrient levels (low, medium and high). The results showed that both planting density and
29	nutrient levels had significant effects on the growth of the plant. The biomass of individual plant
30	and all plants in one pot under medium nutrient level were the highest while the photosynthetic
31	rate and total chlorophyll content were the highest at the high nutrient level. Under different
32	nutrient levels, the photosynthetic rate was the highest at medium planting density. The biomass
33	of single plant decreased with the increase of population density, while the total biomass in the
34	whole pot increased with the increase of density. These characteristics might contribute to the
35	invasion of <i>A. philoxeroides</i> and help the plant to form monodominant community.
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37	keywords: Alternanthera philoxeroides; Biomass; Nutrients; Population density; Photosynthesis.
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45 Introduction

46	Biological invasion has far reaching impacts on ecological functions and the ecological
47	diversity of native environments [1-5]. The invasive species have high fecundity, and once the
48	invasion succeeds, it is easy to form the predominant population. The success of biological
49	invasion is not only determined by the physiological and genetic characteristics of invasive
50	species, but also by the factors such as population density and nutrient levels [6-10].
51	The nutrient is one of the important factors for plant growth. Previous studies have found
52	that traits associated with the resource used determine the invasion success of several invasive
53	species [11, 12]. Invasive plants generally show stronger photosynthesis and growth when
54	invading habitats with richer nutrients [12-15]. Therefore, the nutrient level is also associated
55	with the invasion success [9]. The research on the influence of nutrient on invasive plants plays
56	an important role in understanding the successful plant invasion.
57	The competition among plants is also an important factor affecting the spatial distribution,
58	dynamics and species diversity, promoting the succession of communities [16]. Invasive plants
59	can replace local plants through interspecific competition, accomplishing the process of
60	successful invasion [17]. However, when invasive plants spread to a certain extent, there is a
61	degree of intraspecific competition. In order to adapt to this change, plants tend to change their
62	own characteristics, such as plant height, biomass of branches and physiological characteristics
63	of leaves, etc [18, 19]. And this phenotypic plasticity allows invaders to allocate more nutrients
64	than their native counterparts to increase biomass [20, 21]. Several studies found that the
65	physiological indexes of invasive plant under high density are less than those under low
66	population density [8, 22]. Plant density determines the competitiveness of aquatic clonal plants

67	in complex habitats [23]. Therefore, we can analyze the growth mechanism by studying the
68	morphological changes of plant. At the same time, the effect of population density on invasive
69	plants is one of the core problems in the study of invasion ecology at present [24].
70	Aquatic species might have a fast response to nutrient enrichment, increasing their biomass
71	rapidly, which is particularly true in the aquatic invasive species [9, 25-27]. Therefore, aquatic
72	invasive species affects the productivity and management of land and water resources
73	worldwide [28]. A. philoxeroides is one of these aquatic invasive plants. It is a clone weed that is
74	native to South America and it is a stoloniferous and rhizomatous perennial herbaceous plant
75	[28-30], which propagates clonally and expands rapidly in both aquatic and terrestrial habitats
76	[31-34]. A. philoxeroides has experienced an invasion history of more than 80 years in China and
77	become one of the most harmful invasive species [34]. At present, there are many researches
78	focused on the physiological characteristics of A. philoxeroides, its response to natural
79	environmental factors, and biological and chemical control [10, 35-37]. Many studies have shown
80	that if the plant has a high photosynthetic rate, and it usually grows and propagate rapidly [38,
81	39]. Moreover, the invasive capacity of invasive plants is closely related to the ability of
82	photosynthesis, they often enhance its invasive ability by increasing the photosynthesis of
83	ramets [40, 41]. Thus, studying the photosynthetic capacity of A. philoxeroides is essential for
84	understanding the potential of invasive species and to develop appropriate control strategies.
85	Our previous studies have compared the growth of <i>A. philoxeroides</i> with native plants [10].
86	So in the present study, we conducted a greenhouse experiment which combined nutrient levels
87	with planting density to explore their effects on the growth and photosynthetic characteristic of
88	A. philoxeroides. We asked the following questions: (1) Will the increase of planting density

- inhibit the growth of *A. philoxeroides* under various nutrient levels? (2) How do the nutrient level
- 90 and planting density affect the invasion of *A. philoxeroides*.

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92 Materials and methods

93 Ethics statement

94 We collected plant material for our study with the official permission of the Environmental

95 Protection Bureau of Weishan County, and the Management Committee of the Weishan Lake

96 Constructed Wetland Park. We did not collect endangered or protected species.

97

98 Plant material and experimental design

99 The A. philoxeroides seedlings were collected from Weishan Lake Wetland Park, Shandong

- 100 province, in July 2017. The seedlings were cultured a week in the greenhouse of Fanggan
- 101 Research Station of Shandong University (36°26'N, 117°27'E). The method of sand culture was

102 used in the experiment, and the river sand was washed thoroughly before planting. The length of

- the selected stem was about 11cm, and then was transplanted into a pot (h: 23.5cm; d: 22cm)
- 104 with 7 kg sand. Set up two factors in our experiment: the nutrient level of the sand and planting
- density of A. philoxeroides. The nutrient treatments consisted of three levels (low, medium and
- high; labeled as A, B and C) and three kinds of planting density (low, medium and high; 1, 2 and 4
- 107 seedlings per pot, respectively). In total, there were nine treatment combinations, each dealing
- 108 with five replicates. The nutrient levels and planting density settings in the experiment are
- shown in Table 1.
- 110 Table 1. Nutrient levels and planting density of *A. philoxeroides* at different treatment.

Nutrient levels	TN	Amount of chemical	planting density	planting number
	(g kg ⁻¹ sand)	fertilizer added		(plant pot ⁻¹)
		(g kg ⁻¹ sand)		
			1	1
А	0.5	2.5	2	2
			4	4
			1	1
В	0.7	3.5	2	2
			4	4
			1	1
С	2.1	10.5	2	2
			4	4

111 *A, B, and C: low, medium and high nutrient levels.

112 In the experiment, the compound fertilizer with the ratio of nitrogen and phosphorus (N: P = 4: 1) similar to Nansi Lake was used as the nutrient source. According to our previous 113 114 experiments, the medium nutrient gradient in this experiment is the most suitable for the 115 growth of A. philoxeroides [37]. Water 200 mL every day, in order to ensure the normal growth of A. philoxeroides, and would not lead to the loss of fertilizers in the pot. At the time of 116 fertilization, grounding the fertilizer into powder, then half dissolved in 200 mL of water each 117 118 time, added for two days, to prevent once adding cause damage to plants. At the time of experiment, the low nutrient level was added the fertilizer only in the first week; medium and 119 120 high nutrient levels were added every two weeks during the experiment. The time of the

experiment was from the July 24, 2017 to September 20, 2017.

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123 Determination of photosynthetic rate and light response

124 **CURVE**

125 The photosynthetic characteristics of each group were measured by a Portable 126 Photosynthesis System (*LI-COR 6800*, USA) using PAR of 1000 μ molm⁻²s⁻¹. Leaves were measured 127 under ambient CO₂ concentration [385 μ molmol⁻¹] [42]. The light response curves were 128 measured under the PAR of 1600, 1200, 1000, 800, 600, 400, 200, 100, and 0 μ molm⁻²s⁻¹ [42]. 129 The fitting of the light response curve adopts a non - right - angle hyperbolic model, and the 130 model formula is:

$$Pn = \frac{\alpha I + Pn_{\max} - \sqrt{(\alpha I + Pn_{\max})^2 - 4\theta \alpha I Pn_{\max}}}{2\theta} - Rd$$

131

132 In this formula, α is the apparent quantum rate; I is the light quantum flux density; Pn_{max} 133 is the maximum photosynthetic rate; Rd is the dark respiration rate; θ is the angle parameter 134 that reflects the degree of bending of the light response curve, and the range of values is $0 \leq$ 135 $\theta \leq 1$ [43]. According to the light response curve, we calculated the light compensation point 136 (LCP) and the light saturation point (LSP).

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Determination of chlorophyll content

139 In each treatment, using a volume fraction of 95% ethanol extract chlorophyll, then the 140 spectrophotometer was used to measure the absorbance values at 649nm and 665nm 141 wavelength [44]. The concentration of chlorophyll a, chlorophyll b and total chlorophyll were 142 calculated according to the formula:

 $Ca = 13.95A_{665} - 6.88A_{649}$ $Cb = 24.96A_{649} - 7.32A_{665}$ $Cr = Ca + Cb = 18.08A_{649} - 6.63A_{665}$

144 In the formula, Ca, Cb and Cr represent the concentrations of chlorophyll a, chlorophyll b and

total chlorophyll respectively, [mg cm⁻²]; A_{649} and A_{665} represent the absorbance at the

146 wavelengths of 649 nm and 665 nm respectively [45].

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148 Determination of specific leaf area (SLA) of *A. philoxeroides*

- 149 Before harvest, randomly selected 20 mature leaves of each treatment, wiped clean and
- 150 flatted on the scanner. The determination of leaf area using Photoshop software.
- Leaf area = the percentage of leaf pixels / background pixels · background paper area [46-48].
- 152 SLA = leaf area / leaf dry weight.
- 153

154 Determination of morphological indexes of A. philoxeroides

- 155 We measured the length of stolon and recorded the number of internode before harvest.
- 156 The internode length of *A. philoxeroides* under different treatments was calculated.

157

Determination of biomass index of *A. philoxeroides*

- 159 At the end of the experiment, the roots of *A. philoxeroides* were washed thoroughly, and
- 160 the leaves, stems and roots of each treatment were respectively put into the envelopes,
- numbered and then dried up to constant weight in an oven which the temperature was 80 $\,^\circ\mathbb{C}$,
- 162 recorded the dry weight of each part.

164 Statistical analysis

165	The date of different variables, such as biomass, photosynthetic rate, SLA and total
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- 166 chlorophyll content was analyzed by two-way ANOVA with SPSS 21.0 software (Table 2). The
- 167 significance test in all tests was performed at a level of P <0.05. Use Origin 8.5 software to draw
- 168 charts.

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169 **Results**

170 Table 2. The two-way ANOVA of the effects of the independent variable nutrient level and

171 plant density, and their combination on studied parameters of species A. philoxeroides in

172 the experiment.

Indices	Source	df	F	р
Single leaf biomass	Density	2	56.564	<0.001
	Nutrient	2	114.140	<0.001
	Nutrient $ imes$ Density	4	7.131	<0.001
Single biomass	Density	2	201.859	<0.001
	Nutrient	2	97.273	<0.001
	Nutrient ×Density	4	17.055	<0.001
Total leaf biomass	Density	2	25.660	<0.001
	Nutrient	2	108.667	<0.001
	Nutrient ×Density	4	5.568	0.001
Total biomass	Density	2	28.790	<0.001
	Nutrient	2	68.308	<0.001
	Nutrient ×Density	4	3.485	0.017

Internode length	Density	2	77.514	<0.001
	Nutrient	2	4.017	0.027
	Nutrient ×Density	4	2.251	0.083
SLA	Density	2	49.876	<0.001
	Nutrient	2	195.844	<0.001
	Nutrient \times Density	4	1.751	0.160
Photosynthetic rate	Density	2	35.230	<0.001
	Nutrient	2	42.370	<0.001
	Nutrient ×Density	4	3.346	0.020
Total chlorophyll	Density	2	10.451	<0.001
content	Nutrient	2	156.668	<0.001
	Nutrient \times Density	4	0.502	0.735

173

174 Effects of density on Plant Biomass Index and Morphology

175 Index under different nutrient gradients

176 Results showed that the interaction between nutrient level and plant density significantly

- 177 affect the leaf biomass of per plant, single plant biomass, total leaf biomass and total biomass of
- a whole pot (Table 2).
- 179 Under three nutrient levels, the leaf biomass of single plant and the biomass of single plant all
- 180 decreased significantly with the increase of planting density (Table 3). Among various nutrient
- 181 levels, under three planting density, the reduction rate of leaf dry weight per plant was 47.9%,
- 182 53.9%, 62.5%; the reduction rate of biomass per plant was 57.2%, 64.4%, 63.4% (Table 3). At

183	different nutrient levels, the difference in the leaf biomass of per plant between low density (1)
184	and medium density (2) was slightly greater than the difference between medium density (2) and
185	high density (4) (Table 3). At every nutrient levels, there was a significant difference in the
186	biomass of single plant among all three planting densities (Table 3). In low nutrient level (A), the
187	difference between medium planting density (2) and high planting density (4) is slightly larger
188	than that between low planting density (1) and medium planting density (2) (Table 3). Under
189	medium nutrient (B) and high nutrient (C), the gap between low density (1) and medium planting
190	density (2) is slightly larger than medium planting density (2) and high planting density (4) (Table
191	3).
192	At the three nutrient levels, the total leaf biomass and the total biomass of a pot all
193	increased significantly with the increase of planting density (Table 3). To total leaf biomass of A.
194	philoxeroides, there was no significant difference between low planting density (1) and medium
195	planting density (2) under the treatment of low (A) and medium (B)nutrient levels and the
196	difference among the three planting densities was not significant under the high nutrient level (C)
197	treatment (Table 3). For total biomass of a pot, at low nutrient level (A), the difference between
198	medium planting density (2) and high planting density (3) is significantly smaller than that
199	between low planting density (1) and medium planting density (2), besides there is no significant
200	difference between low planting density (1) and medium planting density (2) at medium (B) and
201	high (C) nutrient levels (Table 3).
202	According to the two-way ANOVA analysis (Table 2), there were have obvious interaction
203	between nutrient level and plant density on internode length of A. philoxeroides.
204	With the increase of planting density, the average internode length of A. philoxeroides

- 205 decreased in varying degrees, and the internode length was the longest at the treatment of low
- 206 planting density (1) (Table 3). At the treatment of three nutrient levels, the average internode
- 207 length of medium nutrient level (B) was the highest, is 5.6 cm (Table 3).

208 Table 3. Different nutrient levels and different planting densities of *A. philoxeroides* biomass

and morphological index.

Treatments	Single leaf	Total leaf biomass	Single biomass	Total biomass	Internode length
	biomass	(g)	(g)	(g)	(cm)
	(g)				
A1	2.90±0.56d	2.90±0.56e	22.83±3.35b	22.83±3.35d	5.66±0.28ab
A2	$1.97 \pm 0.43e$	3.94±0.88e	19.32±0.91c	38.64±1.83b	5.65±0.35b
A4	$1.51 \pm 0.33e$	5.80±1.30d	9.76±2.06de	39.03±8.25b	4.29±0.20d
B1	11.61±1.05a	11.61±1.05bc	40.52±3.03a	40.52±3.03b	6.60±0.61a
B2	7.21±2.13bc	14.42±4.25b	22.08±3.18b	44.16±6.36b	5.56±0.19c
B4	5.35±0.92c	21.38±3.66a	14.41±2.14cd	57.65±8.56a	4.65±0.50cd
C1	7.49±1.53b	7.49±1.53cd	21.38±2.60b	21.38±2.60d	6.05 ± 0.40 ab
C2	$5.06 \pm 0.63c$	$10.11 \pm 1.26c$	11.91±2.13d	23.83±4.26d	5.57±0.27c
C4	2.81 ± 0.77 de	11.25 ± 3.07 bc	7.83±0.91e	$31.30 \pm 3.64c$	4.55±0.06d

*A, B, and C: low, medium and high nutrient levels; 1, 2, and 4: the number of seedlings per pot.

211 Values are presented as means ±SD. Different letters indicate significant differences (p<0.05)

212

213 Effects of plant density on SLA of A. philoxeroides under

214 different nutrient levels

- According to the two-way ANOVA analysis (Table 2), there is no interaction between effects of
- 216 nutrient level and plant density on SLA of *A. philoxeroides*.
- 217 The SLA of *A. philoxeroides* increased significantly with the increase of nutrient level (Fig 1).
- 218 Different planting density had certain effects on the SLA of A. philoxeroides, which was the
- highest under medium planting density (2) and the lowest under high planting density (4), and
- both have significant differences (Fig 1).

Fig 1. The SLA of A. philoxeroides at different nutrient level and plant density. A, B, and

- 222 C: low, medium and high nutrient levels; 1, 2, and 4: the seedlings per pot. Values are
- presented as means ±SD. Different letters indicate significant differences (p<0.05).
- 224

Effects of density on photosynthetic rate and total chlorophyll

content of *A. philoxeroides* under different nutrient levels.

- 227 We found that nutrient levels and planting densities on the photosynthetic rate of *A*.
- 228 philoxeroides has a obvious interaction, and for the total chlorophyll content, they had no
- significant interaction to it (Table 2).

230 The analysis of photosynthetic rate and total chlorophyll content of A. philoxeroides under

- 231 different treatments showed that the change trend of the two indexes are basically the same,
- and both of them increased with the increasing nutrient level (Fig 2). The photosynthetic rate of
- 233 medium planting density (2) of the three nutrient levels were the highest and had significant
- differences, and they were the lowest under the treatment of high density (3) (Fig 2-A). The total
- chlorophyll content of the medium nutrient (B) and high nutrient (C) had obvious differences (Fig
- 236 2-B). The total chlorophyll content of *A. philoxeroides* reached the maximum under high nutrient

237 level (C), and for the planting density, when the planting density was medium (2), the total

chlorophyll content of *A. philoxeroides* reached the maximum (Fig 2-B).

Fig 2. The photosynthetic rate and chlorophyll content of *A. philoxeroides* at different nutrient levels and plant densities. A, B, and C: low, medium and high nutrient levels; 1, 2, and 4: the number of seedlings per pot. Values are presented as means ±SD. Different letters indicate significant differences (p<0.05).

243

244 Effects of density on light response curve of *A. philoxeroides*

245 under different nutrient levels.

246 The data were analyzed and fitted by SPSS software. The fitting coefficients (R²) of the light

- response curve of each group were all greater than 0.9, which showed that the curve fitting
- 248 degree was better and the photosynthetic characteristic of *A. philoxeroides* could be more
- 249 accurately reflected.
- 250 With the increase of the nutrient level, the maximum photosynthetic rate (Pn_{max}) of *A*.
- 251 *philoxeroides* increased in different degrees, so the Pn_{max} at the high nutrient level (C) were the
- 252 largest (Table 4). Among different planting density the Pn_{max} of *A. philoxeroides* were the largest
- at the treatments of medium planting density (2), and the Pn_{max} of A. philoxeroides was minimal
- under the high planting density (4) (Table 4). Besides, under the same nutrient level treatment,
- there were significant differences among the three different planting densities on the Pn_{max} of A.
- 256 *philoxeroides* (Table 4).
- 257 The light compensation point (LCP) and light saturation point (LSP) of *A. philoxeroides* under
- 258 different nutrient treatments were the biggest when planting density was medium (2), and when
- the planting density was high level (4), the value of LCP and LSP were the smallest (Table 4). At
- 260 the same planting density, both LCP and LSP increased with increasing nutrient levels, among
- them, there was a larger gap between medium nutrient (B) and high nutrient (C) (Table 4). Under

262 the same nutrient level, at the difference of the value of LCP and LSP between medium planting

263 density (2) and high planting density (4) was larger than that between medium planting density

264 (2) and low planting density (1) (Table 4).

265 Table 4. Light response curve parameters of *A. philoxeroides* at different nutrient levels and

266 planting densities.

Treatments	Pn _{max}	LCP	LSP
A1	4.79 ± 0.30 g	51.54±4.60fg	250.21±14.90e
A2	6.08±0.28f	56.39±4.46f	283.08±9.79d
A4	$3.77\pm0.25h$	40.68±3.92h	205.30±16.82f
B1	7.55±0.34e	65.48±3.67e	307.15±8.56c
В2	8.34 ± 0.50 d	72.2±2.951d	341.17±13.12b
B4	6.25±0.18f	45.86±1.42g	238.56±4.46e
C1	11.31±0.43b	91.21±4.79b	377.10 ± 30.20 ab
C2	13.40±0.56a	117.73±2.47a	392.55±6.47a
C4	10.00±0.82c	84.09±4.33c	355.47±21.62b

*A, B, and C: low, medium and high nutrient levels; 1, 2, and 4: the number of seedlings per

268 pot. Values are presented as means ±SD. Different letters indicate significant differences (p<0.05)

269

270 **Discussion**

271 In our experiment, nutrient and planting density had significant interaction effects on the

biomass accumulation of *A. philoxeroides*. Under the same planting density, the biomass

accumulation of *A. philoxeroides* could be promoted by increasing nutrients [49-51]. And our

274	previous studies concluded that compared with native plants, the A. philoxeroides had better
275	environmental adaptability and higher biomass and ratio of leaf area under different nutrient
276	conditions [10, 37]. At the treatment of medium nutrient level, the whole pot biomass of A.
277	philoxeroides among three different planting densities was the highest, indicating that too high
278	nutrient levels could be harmful for A. philoxeroides at the initial stage. At medium nutrient level,
279	the increase of planting density had the most obvious effects on the single plant biomass of A.
280	philoxeroides and the effects will be weakened at low nutrient. Therefore, we conclude that
281	under appropriate nutrient level, the effects of planting density on the growth of A. philoxeroides
282	were nutrient dependent. Some studies have shown that at same nutrient condition, the
283	biomass of invasive plants increased more than that of native plants [52]. So in our experiments,
284	under the same nutrient level, although the biomass of single plant was reduced with the
285	increase of planting density, the total biomass of the plant in whole pot was increased, especially
286	at low nutrient. That would enhance the invasive ability of <i>A. philoxeroides</i> .
287	In addition, according to the leaf biomass of A. philoxeroides, we analyzed the SLA of A.
288	philoxeroides. In our study, with the increase of nutrient level, the SLA of A. philoxeroides
289	increased, which is consistent with the studies on SLA of other plants [53-55]. Similarly, the
290	increase of planting density has different influences on the SLA of <i>A. philoxeroides</i> . In this study,
291	the SLA of <i>A. philoxeroides</i> was the highest under medium planting density treatment. SLA is one
292	of the important plant leaf traits and closely related to plant growth and survival strategy. Its
293	value can reflect the ability of plant leaves to intercept light and self-protection in bright light
294	[56]. It is closely related to the photosynthesis and respiration of plants [57]. We therefore
295	concluded that at the medium planting density, the leaf of <i>A. philoxeroides</i> had a higher net

296 photosynthetic rate. So we then analyzed the photosynthetic rate of rate of each treatments to

297

confirm our conclusion.

298	In this study, photosynthetic rate had increase with the increasing nutrient levels that is
299	consistent with previous studies [58-60]. Among the three kinds of plant density treatment, the
300	photosynthetic rate in medium planting density was the highest, and the difference with high
301	planting density was obvious. Chlorophyll is the main pigment of photosynthesis in plants, which
302	reflects the size of photosynthesis in plants [44]. By this experiment, we found that the content
303	of the total chlorophyll of A. philoxeroides had the same trend, indicating that under medium
304	planting density, plants can capture resources better [61].
305	In order to better understand the effects of planting density on the photosynthetic
306	characteristics of A. philoxeroides under different nutrient conditions, we studied the light
307	response curve parameters of A. philoxeroides. Photosynthetic parameters, such as maximum
308	photosynthetic rate (Pn _{max}), light compensation point (LCP) and light saturation point (LSP), are
309	important scientific basis for rapid growth of plants [62-64]. Light saturation point (LSP) can
310	reflected the adaptability of plants to strong light; the lower the light compensation point (LCP) is,
311	the better the normal photosynthesis is under the weak light and the maximum net
312	photosynthetic rate (Pn _{max}) reflects the utilization ability of <i>A. philoxeroides</i> to strong light under
313	different treatments. In our study, among the three kinds of nutrient, the Pn_{max} , LCP and LSP of
314	the A. philoxeroides seedlings in medium planting density were the largest, indicating that under
315	this treatment, A. philoxeroides had the strongest utilization ability and adaptability to glare, and
316	under high planting density, A. philoxeroides has better capability to utilize weak light. Moreover,
317	the photosynthetic parameters of the plant increased with the increasing nutrient levels. That is

318	similar to previous studies [65, 66]. It shows that the increase of nutrient will enhance the
319	utilization ability of A. philoxeroides to strong light. This suggests that at higher nutrient levels,
320	higher light intensities are required to produce more biomass, may be that is why the A.
321	philoxeroides has higher photosynthetic rate but the biomass is lower than the medium nutrient
322	level. Under the same nutrient level, the increase of planting density resulted in the decrease of
323	Pn _{max} , LCP and LSP and there were significant differences among the three planting densities. The
324	results suggested that the increase of planting density decreased the Pn _{max} of A. philoxeroides
325	and its ability to use strong light and its adaptability. Under the low nutrient level, there was no
326	obvious difference in the LCP between the medium and low planting density, which indicated
327	that under the low nutrient level, the high planting density A. philoxeroides had more obvious
328	photosynthetic ability at low light. But at the medium and high nutrient levels, the planting
329	density had more obvious influence on the LCP of A. philoxeroides, and the effects became more
330	obvious with the increase of nutrient level.
331	In conclusion, our study showed that at the three nutrient levels, the SLA, photosynthetic
332	rate and total chlorophyll content of A. philoxeroides at medium planting density were the
333	highest. What's more, although the biomass of single plant, SLA, photosynthetic rate and the
334	content of Chlorophyll reduced with the increase of planting density, the biomass of whole pot
335	tended to increase. These attributes may increase the competitive dominance of A. philoxeroides
336	and could help the A. philoxeroides population develop into a monodominant community.
337	

338 **References**

Pyšek P, Richardson DM. Invasive species, environmental change and management, and health.
Annual Review of Environment & Resources. 2010;35(1).

341 Vilà M, Weiner J. Are invasive plant species better competitors than native plant species? 2. 342 – evidence from pair - wise experiments. Oikos. 2004;105(2):229-38. Vicente JR, Pereira HM, Randin CF, Gonçalves J, Lomba A, Alves P, et al. Environment and 343 3. 344 dispersal paths override life strategies and residence time in determining regional patterns of invasion 345 by alien plants. Perspectives in Plant Ecology Evolution & Systematics. 2014;16(1):1-10. 346 Oduor AMO, Leimu R, Kleunen M. Invasive plant species are locally adapted just as frequently 4. 347 and at least as strongly as native plant species. Journal of Ecology. 2016;104(4):957-68. 348 5. Seebens H, Blackburn TM, Dyer EE, Genovesi P, Hulme PE, Jeschke JM, et al. No saturation in the 349 accumulation of alien species worldwide. Nature Communications. 2017;8:14435. 350 6. Crawley MJ. The Structure of Plant Communities: Blackwell Publishing Ltd.; 2003. 475-531 p. 351 7. Forsman A. Effects of genotypic and phenotypic variation on establishment are important for 352 conservation, invasion, and infection biology. Proceedings of the National Academy of Sciences of the 353 United States of America. 2013;111(1):302. 354 Li SL, Vasemägi A, Ramula S. Genetic variation facilitates seedling establishment but not 8. 355 population growth rate of a perennial invader. Annals of Botany. 2016;117(1). Sardans J, Bartrons M, Margalef O, Gargallo-Garriga A, Janssens IA, Ciais P, et al. Plant invasion is 356 9. 357 associated with higher plant-soil nutrient concentrations in nutrient-poor environments. Glob Chang 358 Biol. 2017;23(3):1282-91. 359 10. Zhang H, Chang R, Guo X, Liang X, Wang R, Liu J. Shifts in growth and competitive dominance of the invasive plant Alternanthera philoxeroides under different nitrogen and phosphorus supply. 360 361 Environmental & Experimental Botany. 2016;135:118-25. 362 11. Daehler CC. Performance Comparisons of Co-Occurring Native and Alien Invasive Plants: 363 Implications for Conservation and Restoration. Annual Review of Ecology Evolution & Systematics. 364 2003;34(1):183-211. 365 12. González AL, Kominoski JS, Danger M, Ishida S, Iwai N, Rubach A. Can ecological stoichiometry help explain patterns of biological invasions? Oikos. 2010;119(5):779-90. 366 367 13. Davis MA, Grime JP, Thompson K. Fluctuating Resources in Plant Communities: A General Theory of Invasibility. Journal of Ecology. 2000;88(3):528-34. 368 369 14. Eva S, Christoph K, Peterj E, HansjöRg D. Influence of light and nutrient conditions on seedling 370 growth of native and invasive trees in the Seychelles. Biological Invasions. 2009;11(8):1941-54. 371 15. Quan G, Mao D, Zhang J, Xie J. Effects of nutrient level on plant growth and biomass allocation of 372 invasive Chromolaena odorata. Ecological Science. 2015. 373 16. Xiang X, Ganlin WU, Duan R, Yan Y, Zhang X. Intraspecific and interspecific competition of Pinus 374 dabeshanesis. Acta Ecologica Sinica. 2015;35(2). 375 17. Chittka L, Schürkens S. Successful invasion of a floral market. Nature. 2001;411(6838):653. 376 18. Schooler S, Baron Z, Julien M. Effect of simulated and actual herbivory on alligator weed, 377 Alternanthera philoxeroides, growth and reproduction. Biological Control. 2006;36(1):74-9. 378 19. Liu LM, Song, H., Liu, H. The effect of water, light and density on the growth of the plant. 379 Environmental Protection Science. 2014;40(4):29-35. 380 20. Drenovsky RE, Martin CE, Falasco MR, James JJ. Variation in resource acquisition and utilization 381 traits between native and invasive perennial forbs. American Journal of Botany. 2008;95(6):681-7. 21. Funk JL. Differences in Plasticity between Invasive and Native Plants from a Low Resource 382 383 Environment. Journal of Ecology. 2008;96(6):1162-73. 384 22. S.Y. Q, Chang EZ, Dong JJ, Guo TT. Competitive effect between invasive plant Galinsoga parviflora

385 and Trifolium repens. Guangdong Agricultural Sciences. 2014;41(1):141-5. 386 23. Cipollini DF, Bergelson J. Plant density and nutrient availability constrain constitutive and 387 wound-induced expression of trypsin inhibitors in Brassica napus. Journal of Chemical Ecology. 388 2001;27(3):593-610. 389 24. Fraver S, D'Amato AW, Bradford JB, Jonsson BG, Jönsson M, Esseen PA. Tree growth and competition in an old - growth Picea abies forest of boreal Sweden: influence of tree spatial 390 391 patterning. Journal of Vegetation Science. 2014;25(2):374-85. 392 25. Butzler JM. THE ROLE OF NUTRIENT VARIABILITY IN AQUATIC ECOSYSTEMS. 2002. 393 26. Smith SDP. The roles of nitrogen and phosphorus in regulating the dominance of floating and 394 submerged aquatic plants in a field mesocosm experiment. Aquatic Botany. 2014;112:1-9. 395 27. Zhao H, Yang W, Xia L, Qiao Y, Xiao Y, Cheng X, et al. Nitrogen-Enriched Eutrophication Promotes 396 the Invasion of Spartina alterniflorain Coastal China. CLEAN - Soil, Air, Water. 2015;43(2):244-50. 397 28. Wang B, Li W, Wang J. Genetic diversity of Alternanthera philoxeroides in China. Aquatic Botany. 398 2005;81(3):277-83. 399 29. Li J, Ye WH. Genetic diversity of alligator weed ecotypes is not the reason for their different 400 responses to biological control. Aquatic Botany. 2006;85(2):155-8. 401 30. LianJin G, Tao W. Impact of invasion of exotic plant Alternanthera philoxeroides on interspecies 402 association and stability of native plant community. Chinese Journal of Eco-Agriculture. 403 2009;17(5):851-6. 404 31. Kolar, Cynthia S, Lodge, David M. Progress in invasion biology: predicting invaders. Trends in 405 Ecology & Evolution. 2001;16(4):199-204. 406 32. Zhang B, Jin, Y.G., Huai, H.Y., Shi, H.Y. Anatomical structure of hollow lotus leaf blade in two 407 habitats. Journal of weeds. 2001(4):6-7. 408 33. Liu J, Dong M, Miao SL, Li ZY, Song MH, Wang RQ. Invasive alien plants in China: role of clonality 409 and geographical origin. Biological Invasions. 2006;8(7):1461-70. 410 34. Pan X, Geng Y, Sosa A, Zhang W, Li B, Chen J. Invasive Alternanthera philoxeroides: biology, 411 ecology and management. Acta Phytotaxonomica Sinica. 2007;45(6):884-900. 412 35. Weng BQ, Lin S, Wang YX. Discussion on adaptability and invasion mechanisms of Alternanthera 413 philoxeroides in China. Acta Ecologica Sinica. 2006;26(7):2373-81. 414 36. Cao YS, Xiao YA, Zhou B, Wen-Jie HU. A THE PHENOTYPIC PLASTICITY OF ALTERNANTHERA 415 PHILOXEROIDES TO DIFFERENT WATER HABITATS. Journal of Jinggangshan University. 2012. 37. Chang RY, Wang RQ, Zhang YR, Liu J. Effects of N:P Ratio and Nutrient Level on the Competition 416 417 between Invasive Alternanthera philoxeroides and Native Oenanthe javanica. Advanced Materials 418 Research. 2012;534:337-42. 419 38. Mcdowell SCL. Photosynthetic Characteristics of Invasive and Noninvasive Species of Rubus 420 (Rosaceae). American Journal of Botany. 2002;89(9):1431-8. 421 39. Penuelas J, Sardans J, Llusià J, Owen SM, Carnicer J, Giambelluca TW, et al. Faster returns on 'leaf 422 economics' and different biogeochemical niche in invasive compared with native plant species. Global 423 Change Biology. 2010;16(8):2171-85. 424 40. Pearcy RW, Tumosa N, Williams K. Relationships between growth, photosynthesis and 425 competitive interactions for a C 3 and C 4 plant. Oecologia. 1981;48(3):371. 426 41. Liu J, He WM, Zhang SM, Liu FH, Dong M, Wang RQ. Effects of clonal integration on 427 photosynthesis of the invasive clonal plant Alternanthera philoxeroides. Photosynthetica. 428 2008;46(2):299.

429 42. Zhu JW. Physiological and biochemical characteristics of hollow lotus seed grass under high 430 manganese stress and the study of glyphosate tolerance: Zheijang University: 2008. 43. Liang WB, Nie DL, Si-Zheng WU, Bai WF, Shen SZ. Photosynthetic light response curves of 431 432 Macropanax rosthornii and their model fitting. Nonwood Forest Research. 2014. 433 44. Chen QZ, Tang N, Zhang BJ, Wang LK, Yang P. Chromium-induced Photosynthetic Physiological 434 Parameters in Alternanthera philoxeroides. Hubei Agricultural Sciences. 2015. 435 45. Guo C, Wei X, Yun-Feng LI, Zhao FM. Physiological Characteristics of Osmotic Adjustment and 436 Content of Chlorophyll of H8 and H10 Carrying DNA Segments of Alternanthera philoxeroides. 437 Southwest China Journal of Agricultural Sciences. 2014;27(2):573-7. 438 46. Xiao Q, Ye, W.J., Zhu, Z., Chen, Y., Zheng, H.L. A simple method for measuring leaf area using 439 digital camera and Photoshop software. Chinese Journal of Ecology. 2005;24(6):711-4. 440 47. Chen WX, Huang, J.J. Comparative study on the method of measuring leaf area of two kinds of 441 plants. Jilin agricultural. 2010(10):50-1. 442 48. Cui SG, Qin, J.H. Image processing method for the determination of leaf area of rape. Hubei 443 Agricultural Sciences. 2017;56(14):2756-7. 444 49. Maron JL, Connors PG. A native nitrogen-fixing shrub facilitates weed invasion. Oecologia. 445 1996;105(3):302-12. 446 50. Burns JH. A comparison of invasive and non - invasive dayflowers (Commelinaceae) across 447 experimental nutrient and water gradients. Diversity & Distributions. 2004;10(5 - 6):387-97. 448 51. Maestre FT, Reynolds JF. Amount or Pattern? Grassland Responses to the Heterogeneity and 449 Availability of Two Key Resources. Ecology. 2007;88(2):501-11. 450 52. Lapointe BE, Bedford BJ. Stormwater nutrient inputs favor growth of non-native macroalgae 451 (Rhodophyta) on O'ahu, Hawaiian Islands. Harmful Algae. 2011;10(3):310-8. 452 53. Arendonk JJCMV, Niemann GJ, Boon JJ, Lambers H. Effects of nitrogen supply on the anatomy 453 and chemical composition of leaves of four grass species belonging to the genus Poa, as determined 454 by image - processing analysis and pyrolysis - mass spectrometry. Plant Cell & Environment. 455 1997;20(7):881-97. 456 54. Meziane D, Shipley B. Interacting determinants of specific leaf area in 22 herbaceous species: 457 effects of irradiance and nutrient availability. Plant Cell & Environment. 1999;22(5):447-59. 458 55. Niinemets UKK. Leaf structure vs. nutrient relationships vary with soil conditions in temperate 459 shrubs and trees. Acta Oecologica. 2003;24(4):209-19. 56. Zhang L, Luo T. Advances in ecological studies on leaf lifespan and associated leaf traits. Acta 460 461 Phytoecologica Sinica. 2004;28(6):844-52. 462 57. BenomarLahcen, DesRochersAnnie, Larocqueguy R. Changes in specific leaf area and 463 photosynthetic nitrogen-use efficiency associated with physiological acclimation of two hybrid poplar 464 clones to intraclonal competition. Canadian Journal of Forest Research. 2011;41(7):1465-76. 465 58. Van KM, Weber E, Fischer M. A meta-analysis of trait differences between invasive and 466 non-invasive plant species. Ecology Letters. 2010;13(2):235-45. 467 59. Feng YL, Auge H, Ebeling SK. Invasive Buddleja davidii Allocates More Nitrogen to Its 468 Photosynthetic Machinery than Five Native Woody Species. Oecologia. 2007;153(3):501-10. 469 60. Zhang W, Xiao H, Yin Z, Zeng X, Huang M, Feng Y, et al. Effects of simulated nitrogen deposition 470 on photosynthetic characteristics of the invasive plant Mikania micrantha. Ecology & Environmental 471 Sciences. 2013;22(12):1859-66. 472 61. Han LH, Liu, C., Wang, J.J. Effect of nitrogen fertilizer on the content of pigment in purple stem of

473 different population. Hubei Agricultural Sciences. 2012;51(3):475-7.

474 62. Sharp RE, Matthews MA, Boyer JS. Kok Effect and the Quantum Yield of Photosynthesis: Light

475 Partially Inhibits Dark Respiration. Plant Physiology. 1984;75(1):95-101.

- 476 63. Awada T, Radoglou K, Fotelli MN, Constantinidou HI. Ecophysiology of seedlings of three
- 477 Mediterranean pine species in contrasting light regimes. Tree Physiology. 2003;23(1):33-41.
- 478 64. Hui Z. Photosynthetic characteristics comparison between an invasive plant, Lantana camara
- 479 L., and associated species. Acta Ecologica Sinica. 2009;29(5):2701-9.
- 480 65. Jiao JY, Yin CY, Chen K. Effects of soil water and nitrogen supply on the photosynthetic
- 481 characteristics of Jatropha curcas seedlings. Chinese Journal of Plant Ecology. 2011;35(1):91-9.
- 482 66. Wang S, Han, X.R., Zhan, X.M., Yang, J.F., Liu, Y.F., Wang, Y., Li, N. The comparative study on
- fitting light response curve model of photosynthesis of maize under different nitrogen fertilizer levels.
- 484 Journal of Plant Nutrition & Fertilizer. 2014;20(6):1403-12.



