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An interdisciplinary study around the reliquary of the late cardinal Jacques de Vitry

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31 **Abstract**

32 The reliquary of Jacques de Vitry, a prominent clergyman and theologian in the early
33 13th century, has experienced several transfers over the last centuries, which seriously
34 question the attribution of the remains to the late Cardinal. Uncertainty about the year of his
35 birth poses an additional question regarding his age at death in 1240. The reliquary, located in
36 the Saint Marie d'Oignes church, Belgium, was reopened in 2015 for an interdisciplinary
37 study around his relics as well as the Treasure of Oignes, a remarkable cultural heritage
38 notably built from Jacques de Vitry's donation. Anthropological, isotopic and genetic
39 analyses were performed independently on the remains found in the reliquary. Results of the
40 analyses provided evidence that the likelihood that these remains are those of Jacques de
41 Vitry is very high: the remains belong to the same human male individual and the historical
42 tradition about his age is confirmed. In addition, a separate relic (left tibia) was analysed and
43 found to match with the remains of the reliquary (right tibia). The unique Jacques de Vitry's
44 mitre, made of parchment, was sampled non-destructively and the extracted parchment
45 collagen was analysed by a proteomic method in order to determine the animal species. The
46 results showed that, surprisingly, not all parts of the mitre were made from the same species.
47 All together, these findings are expected to fertilize knowledge carried by historical tradition
48 around the relics of Jacques de Vitry and his related cultural heritage.

49 **Introduction**

50 Jacques de Vitry was a prominent clergyman and theologian, successively regular
51 canon, bishop and cardinal of the Roman Catholic Church, who was active in Europe and
52 Middle East during the first part of the thirteen-century (S1A Fig). His life and personality
53 are mainly known from his writings (e.g. *Historia Orientalis*), crusade preaches and sermons.
54 Unfortunately, no autobiography is available and facts about his youth are scarce. His date of
55 birth is uncertain with two hypotheses coexisting: 1165-1170 (anonymous source, ca. 1250
56 [1]) or 1175-1180 (contemporary scholar Jean Donnadiou [2]), the former source being
57 questionable (Jacques de Vitry neither studied theology in Paris in 1187 nor was the
58 confessor of the king of France, as reported by the anonymous source). The date and place of
59 his death, on the other hand, are known precisely as 1st May 1240 in Rome [3].

60 Having studied theology in Paris (between 1190 and 1208), he received episcopal
61 consecration around 1210. He left Paris around 1208 to join the priory Saint Nicolas
62 d'Oignes (funded in 1187) of the Diocese of Liège as an Augustinian canon regular. There,
63 he met Marie d'Oignes (S1B Fig) and became the confessor and the biographer of the
64 visionary, ecstatic beguine who died in 1213 at the age of 36 and was subject to popular
65 devotions [4, 5]. Between 1212 and 1216, he preached the crusade against the Albigenses.
66 Following his election as bishop of Saint John of Acre, he left the priory for the Holy Land in
67 1216. Following Damietta defeat in 1221, he decided to come back to Europe. A year after
68 his return, in 1225, he resumed his itinerant life as preacher of the sixth crusade. In 1229, he
69 was elevated to the College of Cardinals and settled in Rome where he died on May 1st 1240.
70 His body was buried in the Dominican headquarter convent in Rome [6].

71 According to his testimonial will to lay at rest near to Marie d'Oignies, his remains were
72 transferred to Oignies a year later and put in a marble monument, close to Marie's one in the
73 monastery church [7]. Several transfers of his relics took place in the next centuries. In 1636,
74 the reliquary was open for transfer of his relics to a new place. Two teeth were removed on
75 that occasion and given to clergymen for devotion [8]. On July 25th 1759, the reliquary was
76 open for transfer to another place inside the church [9]. During the demolition of the priory in
77 1808, the relics were transferred to the Saint Martin church (Aiseau, Belgium) where they
78 were put in a lead container, dated from 1844. After the collapse of the Saint Martin church
79 building in 1970, the reliquary was opened in 1971 for the transfer of the remains to the Saint
80 Marie d'Oignies church, Belgium, which was built in 1908 near the place of the ancient
81 priory. The reliquary (and its content) is still displayed in this church today.

82 Throughout his life but also by testimony, Jacques de Vitry enriched the priory of Oignies
83 with books, relics, tissues, vessels and other religious artworks [10]. He granted, among
84 others, an ivory cross, a portable altar and two mitres, one of which being a unique object as
85 it features miniatures on parchment. All these objects and others collected later constitute the
86 Treasure of Oignies, a unique cultural heritage ensemble, which was recognized as such by
87 the Belgian Federal State in June 2010 [11].

88 In 2015, the Archaeological Society of Namur (SAN), which has the scientific responsibility
89 of the Treasure of Oignies, set up a consortium in partnership with several Belgian
90 universities and research institutes in order to undertake an ambitious, interdisciplinary
91 scientific study around the reliquary of Jacques de Vitry (referred as the CROMIOSS
92 project). The study was motivated by the fact that the reliquary of the prominent clergyman
93 and theologian has experienced several transfers during the last centuries, which seriously
94 question the attribution of the remains to the late Cardinal. Uncertainty about the year of his
95 birth poses an additional question regarding his age at death. Given the patrimonial
96 importance of the Treasure of Oignies, the research consortium decided to englobe within its
97 inquiry a material study of one of the bishop's mitre (Fig 1), the only known example of a
98 mitre composed of parchment. In spite of this exceptional feature, this mitre has so far
99 received little attention from art historians.

100 Following the opening of the reliquary on September 8th 2015, human remains were found in
101 the lead container. The interdisciplinary study reported hereafter relies on a critical
102 confrontation between the historical tradition and the scientific results obtained from
103 anthropological, isotopic, genetic and proteomic analyses.

104 **Fig 1. Jacques de Vitry's mitre made of parchment.** Donation of Jacques de Vitry to the priory of Oignies
105 (collection of the Treasure of Oignies, Belgium). (A) Early photography (© Armand Dandoy, 1879). (B)
106 Contemporary photography (© Vedrin, Guy Focant) with sampling spots indicated for parchment proteomic
107 analyses (1, 2: cap; 3 cap border; 4, 5: left and right lappets).

108 **Materials and Methods**

109 **Putative remains of Jacques de Vitry**

110 With the approval of the competent authorities, the reliquary (S2A Fig) was opened
111 on September 8th 2015, in the presence of the Bishop of the Diocese of Tournai, journalists
112 and scientists. Caution was taken in order to avoid contamination of the relics by human
113 DNA: restricted access, use of gloves and masks. Because the reliquary was previously open
114 and relics were transferred several times since the second burial, it was first necessary to
115 determine the human nature of the relics and to check whether or not they belonged to a
116 single individual. A primary inspection of the lead container content was performed during
117 the opening ceremony (S2B Fig). A wooden frame containing a tibia, which was supposed to
118 belong to Jacques de Vitry, was also exhibited on that occasion. In fact, the story of this tibia
119 is rather tumultuous. It was probably displayed in a private place and stolen at an unknown
120 date. Found by the police in the 20th century, it was first given back to the village of Oignies
121 erroneously (actually, Oignies-en-Thiérache located in the Province of Namur, about 70 km
122 from Oignies) before finally reaching the church Saint Marie of Oignies in the 20th century.
123 In the reliquary, the remains were wrapped in a textile, which also contained small fragments
124 of bone, plant remains, textile fibres, golden flakes and insect remains belonging to various
125 groups of beetles (xylophagous (*Anobium punctatum*), detritivorous (*Ptinus* and *Tenebrio*
126 *sp.*), necrophagous (*Trox scaber*) and granivorous (*Sitophilus granarius*)). The first three
127 groups are part of taxa that are occasionally found in a funerary context and the presence of
128 *Trox scaber* would suggest that organic material remnants, i.e. muscles, skin or hair, were
129 still present. The reliquary, the frame and their contents were then transferred to the
130 University of Namur (Laboratory of Anatomy) in order to take bone samples for further
131 genetic and isotopic analyses.

132 **Anthropological study**

133 A detailed inventory of the remains found in the reliquary was first established in
134 order to estimate the minimum number of individuals. The sex determination of the studied
135 individual was complicated by the absence of pelvic bones, which show the greatest sexual
136 dimorphism. For this reason, we applied a sex determination method using cranium
137 characteristics [12] and discriminant functions based on tibia measurements [13, p. 240-242].
138 In order to estimate the age at death, we used the dental wear [14], the closure of the cranial
139 sutures [13, p. 120-121] and cementum analysis [15]. The stature was estimated using
140 equations of Olivier et al., and computed from the length of long bones originating from a
141 French sample [16].

142 **Isotopic study**

143 Collagen extraction was performed following Longin's (1971) method. A 1% NaOH
144 wash step (15 minutes) was introduced between demineralization and hydrolization steps.
145 First, all the bone samples were demineralized in 10 ml 8% HCl for 20 minutes, and rinsed
146 with MilliQTM-water. After that, each sample was immersed for 15 minutes in 1% NaOH, and
147 again rinsed with MilliQTM-water. Then, after adding 1% HCl for neutralization, it was
148 washed with MilliQTM-water. For all the steps mentioned above, Ezee-filters were used.
149 Gelatinization of the extract was done in water (pH 3), at 90° C for 12 hours. The resulting

150 gelatin was filtered with a Millipore 7 micrometre glass filter, and freeze-dried. Age
151 determinations (^{14}C analyses) were carried out on the AMS instrument at the Royal Institute
152 for Cultural Heritage (KIRK-IRPA), Brussels (Lab code RICH-) [17]. CO_2 was released by
153 sample combustion in the presence of CuO and Ag . Graphitization was performed with H_2
154 over a Fe catalyst. Targets were prepared at KIK-IRPA [18]. ^{14}C calibrations were performed
155 using OxCal (version 3.1) [19] and the IntCal13 calibration curve date [20]. The C:N ratio,
156 $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ analyses were performed on a Thermo Flash EA/HT elemental analyser,
157 coupled to a Thermo Delta V Advantage Isotope Ratio Mass Spectrometer via ConFlo IV
158 interface (Thermo Fischer Scientific, Bremen, Germany). Standards used were IAEA-N1,
159 IAEA-C6, and internally calibrated acetanilide. Analytical precision was 0.25‰ for both $\delta^{13}\text{C}$
160 and $\delta^{15}\text{N}$ based on multiple measurements of the standard acetanilide.

161 **Genetic study**

162 DNA extraction and amplification were performed at UNamur and KULeuven in
163 dedicated laboratories for ancient DNA analysis. Standard precautions for contamination
164 were taken including separated areas for pre- and post-PCR procedures, restricted access to
165 pre-PCR areas, multiple negative controls in the DNA extraction and amplification reactions,
166 replication of DNA extractions and PCR reactions, contamination control with DNA profiles
167 of all laboratory staff.

168 At UNamur laboratory, bone nuclear DNA was extracted according the procedure described
169 by Mundorff and Davoren [21]. Briefly, the petrous bone and the tibiae were cleaned with
170 10% bleach and bone powder was withdrawn by drilling with a decontaminated 8-mm drill
171 under a previously decontaminated chemical hood. Nuclear DNA was then extracted from
172 ≈ 3 g of bone powder using the Set Buffer Trace Bone kit and Nucleospin DNA Trace from
173 forensic sample kit (Macherey-Nagel, Germany). Nine different microsatellite markers were
174 amplified on the ancient bone DNA and contemporary DNA (used as control DNA) using the
175 AmpFISTR Minifiler PCR Amplification kit (Applied Biosystems) and were further analysed
176 with GeneMapper on the Abi 3130xl Genetic Analyser (Applied Biosystems) at URBE
177 (UNamur). Standard PCRs of 5 Y chromosome short tandem repeats (DYS19, DYS389II,
178 DYS448, DYS456, DYS635) were performed using sets of primers designed by Kwon and
179 coworkers [22].

180 At KULeuven laboratory, the procedures described by Ottoni et al. [23] were used for
181 decontamination and grinding of the bone material and the two upper teeth samples, as well
182 as for DNA extraction through silica-based spin columns (QIAquick PCR Purification Kit,
183 Qiagen). One of the teeth was reused for genetic analysis after cement chrono-analysis. This
184 tooth was embedded in epoxy resin and cut into two so that only the dentin and the root
185 channel could be removed with a decontaminated dental drill. Multiplex DNA amplification
186 of autosomal short tandem repeats (STRs) and STRs from the Y chromosome was done
187 respectively according to Dognaux et al. and Larmuseau et al. [24–26] except for the number
188 of cycles, which was raised to 34. The autosomal multiplex of 9 STRs (fragment size
189 between 70 and 275 bp) includes also primers to amplify a sequence in an intron of the
190 Amelogenin gene present on the sex chromosomes (123 bp on the X and 129 bp on the Y)

191 [27]. Three multiplexes were used for the Y-chromosome STRs including in total 40 STRs
192 with a size range between 74 and 420 bp. Fragment analysis was performed on an Applied
193 Biosystems 3130XL Genetic Analyzer (Thermo Fisher Scientific) with data analysis using
194 GeneMapper ID v3.2 software (Thermo Fisher Scientific). Analysis of the first and the
195 second hypervariable segments (HV-I and HV-2) of the mtDNA control region was
196 accomplished by amplification of, respectively, five and two overlapping fragments ranging
197 in size from 109 to 166 bp, followed by direct sequence analysis according to Ottoni et al.
198 [23]. Forward and reverse sequencing was performed using the BigDye Terminator v3.1
199 Cycle Sequencing Kit (Thermo Fisher Scientific) according to the protocol of the
200 manufacturer. Sequence analysis was done on an Applied Biosystems 3130XL Genetic
201 Analyzer (Thermo Fisher Scientific) with data analysis using DNA Sequence Analysis
202 Software v5.2 (Thermo Fischer Scientific) and aligning of the sequences against the revised
203 Cambridge Reference Sequence using BioEdit v7.0.4 [28, 29].

204 **Proteomic study**

205 Non-invasive collagen extraction and sample preparation were done following the
206 ZooMS method [30].

207 **Sampling procedure**

208 Sampling was performed in a clean area inside biological laboratory, with the mitre
209 laid down on a clean table. Sampling consisted in gentle rubbing of the surface of parchment
210 parts of the mitre (S3 Fig). For each sampling, a new piece of PVC erasers (Mars, Staedler)
211 and new nitrile gloves were used, and the table was cleaned with isopropanol. Samples
212 (eraser crumbs containing parchment collagen) were taken in duplicate at different locations
213 where the parchment was bare (non-decorated parts): on the cap as well as on the right and
214 left lappets (Fig 1B). Eraser crumbs were collected in a 1.5ml Eppendorf tube and were
215 stored at 4°C until collagen extraction.

216 **Collagen extraction and digestion**

217 50 µl of NH₄HCO₃ 50mM buffer was added to each sample. Eppendorf tubes were
218 spin down at maximum speed in a centrifuge. 200 ng of trypsin (Promega) was added to each
219 sample and incubated during 4 hours under light agitation. The samples were acidified with a
220 solution of trifluoroacetic acid (TFA) to a final concentration of 1% (vol./vol.). Eppendorf
221 tubes were centrifuged during 5 min at maximum speed and supernatant containing the
222 peptides were transferred to another Eppendorf vial.

223 **Peptides desalting and concentration**

224 ZipTip C18 (Millipore) pipettes were used. After washing and conditioning of the
225 ZipTip according the manufacturer's instructions, the peptides were loaded and desalted with
226 a solution of H₂O, 0,1% TFA (vol./vol.). Peptides elution was done with 10 µl of 80%
227 acetonitrile (ACN) / 0.1% TFA (vol./vol.). All samples were then vacuum dried (Heto) and
228 recovered with a solution of 2% ACN/ 0.1% TFA (vol./vol.).

229 **Mass spectrometry analysis**

230 All samples were analysed using liquid chromatography (UltiMate 3000, Thermo
231 Systems) coupled to electrospray tandem mass spectrometry (MaXis Impact UHR-TOF,
232 Bruker) (LC-MSMS). The digests were separated by reverse-phase liquid chromatography
233 using a 75 μm X 150-mm reverse phase column (Acclaim PepMap 100 C18). Mobile phase
234 A was 95 % H_2O /5 % ACN, 0.1 % formic acid. Mobile phase B was 80% ACN/20% H_2O ,
235 0.1 % formic acid. The digest was injected, and the organic content of the mobile phase was
236 increased linearly from 5 % B to 40 % B in 15 min and from 40 % B to 100 % B in 5min.
237 The column effluent was connected to a Captive Spray (Bruker). In survey scans, MS spectra
238 were acquired for 0.5 s in the m/z range between 50 and 2200. The 10 most intense peptides
239 2^+ or 3^+ ions were sequenced. The collision-induced dissociation (CID) energy was
240 automatically set according to mass-to-charge (m/z) ratio and charge state of the precursor
241 ion. MaXis and Thermo Systems instruments were piloted by Compass HyStar 3.2 (Bruker).
242 Peak lists were created using DataAnalysis 4.0 (Bruker) and saved as mgf file for use with
243 ProteinScape 3.1 (Bruker) with Mascot 2.4 as search engine (Matrix Science). Enzyme
244 specificity was set to trypsin, and the maximum number of missed cleavages per peptide was
245 set at one. Hydroxylation (KP) and oxidation (M) were allowed as variable modification.
246 Mass tolerance for monoisotopic peptide window was 7 ppm and MS/MS tolerance window
247 was set to 0.05Da. The peak lists were searched against a home-made collagen protein
248 database and a contamination protein database. Scaffold software (Proteome Software) was
249 used to validate protein and peptide identifications, and also to perform the search of species
250 marker peptides. Our species marker database contained specific peptides that allowed us to
251 differentiate *Capra hircus*, *Ovis aries* and *Bos taurus*.

252 **Results and discussion**

253 **Results**

254 **Anthropological analysis of the remains**

255 Inside the container, a cranium, 26 bones and 7 teeth were found, wrapped in a black
256 satin shroud sealed with staples and a red wax seal. Mandible, ribs, pectoral and pelvic girdle
257 bones were absent (Fig 2 A), precluding efficient identification of the sex. No lower teeth
258 were found and 9 upper teeth were missing: 7 were lost post mortem and 2 ante mortem.

259 **Fig 2. Remains found in the reliquary.** (A) Inventory of the remains. (B) Radiography of the left and right
260 tibias. (C) Left tibia (top) and right tibia (bottom). (D) Evidences of cuts on the bones (articulations).

261 Visual and metric inspection of the remains led us to conclude that they belong to the same
262 individual. Metric comparison between the right and left tibias (Fig 2C) revealed almost no
263 difference, except the discoloured aspect of the right tibia (the one put in the frame as a
264 distinct relic), which might be the result of cleaning treatment applied before framing.
265 Radiography of both tibias (Fig 2C) showed that the internal structures of both tibias exhibit

266 strong similarity (concordance of Harris lines), which confirms they belong to the same
267 individual.

268 Five cranial female characteristics (slightly delimited glabella, arched traces of nuchal lines
269 and delimited supraorbital ridge; smooth external occipital protuberance and vertical frontal
270 bone) and two male features (large mastoid process and quadrangular orbits) were identified.
271 Inspection of tibia morphologies produced male or female diagnostics, depending on the
272 morphological character considered.

273 Dental wear indicated an age ranging from 45 to 55. Cranial vault sutures indicated death
274 between 30 and 60. Analysis of tooth cementum gave an age estimate from 53 to 62. The
275 “consensus biological age” was around 55, which was supported by the fact that the specimen
276 presented a few signs of degenerative osteoarthritis. Stature was estimated, based on the left
277 tibia length. If the individual were a male (female), his (her) height would have been around
278 1.66 (1.62) meter.

279 Close visual inspection of the bones revealed peculiar, unexpected and interesting features:
280 cut marks (Fig 2D) in the periphery and within articulations of the right shoulder, the right
281 and left elbows, the right and left knees. These cuts were most likely made in order to remove
282 muscles and ligaments.

283 **Isotopic analysis of tibias and skull fragments**

284 Isotopic analyses ($\delta^{13}\text{C}$, $\delta^{15}\text{N}$ and ^{14}C) were carried out on two samples taken from the
285 left tibia, one sample from the right tibia and one sample from the skull. The results are
286 shown in Table 1.

287 **Table 1. Results of isotopic analyses: sample laboratory codes and names, ^{14}C ages (BP),**
288 **calibrated ages (2σ), stable isotope fractionations ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$), C:N ratio of collagen extracted**
289 **from different bone samples from the remains.**

Lab-code	Sample name	^{14}C age (BP)	Calibrated age (2σ)	$\delta^{13}\text{C}$ (‰)	$\delta^{15}\text{N}$ (‰)	Atomic C:N ratio
RICH-22318.1.1	Left Tibia 1	915±28	1030AD (95.4%) 1190AD	-19.9	10.8	3.3
RICH-22318.2.2	Left Tibia 2	903±28	1030AD (95.4%) 1210AD	-20.0	11.0	3.3
RICH-22319.2.1	Right Tibia	896±26	1040AD (95.4%) 1220AD	-19.8	11.2	3.2
RICH-23856	Skull	957±26	1020AD (95.4%) 1160AD	-19.0	11.0	2.9

290 BP, Before Present. σ , standard deviation.

291 The C:N ratio of the collagen of all samples fell within the C:N range proposed by De Niro
292 [31] for well-preserved collagen, namely between 2.9 and 3.6. The stable isotopes ($\delta^{13}\text{C}$,
293 $\delta^{15}\text{N}$) results suggested that both tibias and the skull might belong to the same individual. The

294 ¹⁴C results confirmed this hypothesis by passing positively the χ^2 -test. The average of the
 295 four ¹⁴C dates was calculated: 919±13BP (χ^2 -test: df=3, T=3.3(5% 7.8)), which led to a
 296 skeleton date between 1040 and 1170AD (95.4% probability) after calibration (Fig 3).

297 **Fig 3. Radiocarbon dating results from the remains.** (A) Calibrated ages (2 σ) of the left tibia, right tibia and
 298 skull. (B) Average age of the four ¹⁴C dates determined for these remains.

299 Genetic analysis of the remains

300 Analysis of nuclear DNA from the petrous bone and both tibias was first performed at
 301 UNamur using 9 human DNA microsatellite markers. However, the analysis of DNA extracts
 302 from the remains turned out to be not reproducible among the different samples and the
 303 presence of the Y chromosome could not be established (data not shown). A standard PCR
 304 was then used to amplify 5 short tandem repeats (STR) of the Y chromosome using the same
 305 DNA samples. The Y chromosome was found in the petrous bone DNA, whereas the tibias
 306 DNA did not give reproducible results (Fig 4). These results suggest that the genetic material
 307 is strongly damaged in the tibias and, to a lesser extent, in the petrous bone.

308 **Fig 4. PCR amplification of 5 STRs from the Y chromosome.** Left tibia (LT), right tibia (RT), petrous bone
 309 (S), negative control (-), positive control (+).

310 Genetic analysis of the remains was performed independently at KULeuven following
 311 standard ancient DNA (aDNA) protocols. Five samples were analysed: tibia powder, tooth
 312 embedded in epoxy resin (sample reused after cement chrono-analysis), a petrous bone
 313 fragment and two upper teeth. Analysis of short tandem repeats from the Y chromosome on
 314 the tibia DNA extract gave no results, indicating that the quantity and/or quality of the
 315 extracted nuclear DNA were insufficient or that the DNA originated from a female. Identical
 316 results were obtained for the resin embedded tooth and the petrous bone DNA extracts. The
 317 results obtained by UNamur and KULeuven for nuclear DNA are consistent with the
 318 treatment of the remains attributed to Jacques de Vitry (discussed below) which would have
 319 impacted the preservation of the DNA [32, 33]. In contrast, partial profiles for both
 320 autosomal (including sex determination with amelogenin gene) and Y-chromosome STRs
 321 were obtained with the two upper teeth DNA extracts (Table 2).

322 **Table 2. Genotyping results of the analysis of nine autosomal STRs, Amelogenin and 40 Y-**
 323 **chromosome STRs in DNA extracts from two upper teeth.**

Autosomal STRs	AMEL ¹	D2S441	D1S1656	D12S391	D10S1248	D21S11	D22S1045	D18S51	D1S1677	FGA
Amplicon length (bp)	123-129	72-112	115-168	169-227	78-142	150-217	68-119	139-219	74-118	119-278
Upper tooth 1	Y	11	16.3	-	14	-	-	14	13,15	18,(22) [20]
Upper tooth 2	Y	(11),14	-	-	14 [13]	32.2	-	14,(20) [19]	13,(15)	18,22
Y-STRs	DYS438	DYS392	DYS458	DYS385	DYS460	DYS481				
Amplicon length (bp)	87-140	90-130	130-160	237,316	95-123	105-168				
Upper tooth 1	9	11	14	12,13	12,13	22				
Upper tooth 2	-	-	14	-	10	-				

324 ¹ Amelogenin.
 325 [] PCR fragments attributed to stochastic effects of either high stutter or drop-in; () alleles with lower peak
 326 height (heterozygote imbalance).

327 The autosomal STR results were reproducible between the DNA extracts as well between
 328 different PCR reactions indicating that the two teeth originated from the same individual. The
 329 characteristics of these profiles (drop-out of alleles or loci, drop-in of one or two alleles, high
 330 stutter frequency, heterozygote imbalance) were consistent with ancient DNA, where DNA
 331 damage and fragmentation lead to non-amplification of alleles or loci, and where low
 332 amounts of DNA lead to stochastic effects (drop-out, high stutter effect and heterozygote
 333 imbalance) that will influence the results [34]. The fact that very partial profiles were
 334 obtained for the Y-chromosome STRs can be explained by the higher sensitivity of the
 335 autosomal STR multiplex for low amounts of degraded DNA, except for the Amelogenin
 336 amplicons where the X-allele could not be amplified in the DNA extracts of the two upper
 337 teeth.

338 All five DNA extracts were also subjected to the analysis of the non-coding region of human
 339 mtDNA. In contrast to the nuclear DNA results, all DNA extracts with the exception of the
 340 DNA extract from the tibia powder revealed a mitochondrial (mt) sequence (Table 3).

341 **Table 3. Results of mtDNA Sanger sequencing for different bone and teeth samples from the**
 342 **remains.**

	HVI (16008-16366)								HVII (48-155)		
	16126	16145	16192	16224	16256	16261	16270	16304	73	150	152
rCRS¹	T	G	C	T	C	C	C	C	A	C	C
Resin embedded tooth	T	G	C/T	T/C	C/t	C	C/T	C	G/a	C/T	C
Petrous bone	T	G	C	T	C	C	C	C	A	C	C
Upper tooth 1	T/c	G/a	C/T	T	C/t	C/T	C/t	C	A/G	C	C
Upper tooth 2	T	G	C/T	T/C	C	C	C	C	A/G	C	C/T
Analyst	T	G	C	T	C	C	C	T	A	C	T

343 ¹ cCRS, revised Cambridge Reference Sequence [28].

344 Analyst is the person who performed the DNA extraction and sequence analysis.

345 The samples of the reused tooth and the two upper tooth samples showed at several positions
 346 the presence of two nucleotides, which is evidence for the presence of exogenous DNA in the
 347 DNA extracts. The origin of this DNA remains unknown (laboratory contamination can be
 348 excluded) and can probably be related to the handling of the remains during the different
 349 historical transfers. The fact that four out of five DNA extracts showed preservation of
 350 human mtDNA can be explained by the higher copy number of mtDNA and, to a lesser
 351 extent, additional protection against degradation by the double membrane of the
 352 mitochondrion which would increase the chance to extract preserved mtDNA from
 353 archaeological remains [35, 36]. The mtDNA sequence of the DNA extract from the petrous
 354 bone fragment revealed no evidence for exogenous DNA and was identical to the revised
 355 Cambridge Reference Sequence [28]. This mtDNA sequence, the most frequent (about 10%)
 356 in West Eurasian populations (<https://empop.online/>; v3/R11), could also be present in the
 357 other DNA extracts, which would be consistent with the hypothesis that all the skeletal
 358 remains belong to the same individual. This hypothesis would also imply that the origins of

359 the exogenous DNA in the other samples are different, which is supported by the different
360 mtDNA sequences observed for the two upper teeth where reproducible nuclear DNA
361 profiles were obtained.

362 **Proteomic analysis of collagen from parchment parts of the mitre**

363 Identification of the origin species of parchment parts of the mitre was carried out by
364 proteomic analysis of the collagen protein extracted using a non-invasive sampling method
365 (S3 Fig). Searches for species biomarkers gave unambiguous results on samples taken from
366 four different locations on the mitre: cap, lower part of cap, left and right lappets (Fig 1 B).
367 Origin species (sheep, calf or goat) were identified in parchment samples according to the
368 number of occurrences of species-specific peptide markers obtained during sequencing of
369 collagen I and III proteins (Table 4).

370 **Table 4. Results of species identification in parchment parts of the mitre.**

Sample	Location	Species-specific peptides (SSP) detected	Number of SSP validated and sequenced	Identified species
1	Cap	1	19	<i>Ovis aries</i> (sheep)
2	Cap	1	12	<i>Ovis aries</i> (sheep)
3	Lower part of cap	2	3	<i>Bos Taurus</i> (calf)
4	Left lappet	7	52	<i>Bos Taurus</i> (calf)
5	Right lappet	2	5	<i>Bos Taurus</i> (calf)

371

372 **Discussion**

373 **History of the remains**

374 In order to ease interpretation of the results of the scientific studies, we develop here
375 additional historical considerations apart from the biography of Jacques de Vitry given above.
376 Before Jacques de Vitry settled in Oignies, the relatively recent and poor priory did not attract
377 attention. No doubt the arrival of Jacques de Vitry changed the destiny of the priory,
378 favouring its enrichment by the acquisition of relics [37, 38]. The meeting with Marie
379 d'Oignies had important consequence for the destiny of the future bishop and cardinal since it
380 was at the origin of his decision to be buried in Oignies. Actually, it was a current practice in
381 the high society of that time to notify by testimony the will to be buried in homeland in case
382 of death occurred abroad. Repatriation of dead human bodies from long distances had
383 practical consequences in terms of treatments applied to the corpse in order to ensure
384 hygienic and safe transfer. This implied dismembering of the corpse and use of recipes to
385 treat the remains [39-41]. It is noteworthy that this practice was forbidden in 1299 by Pope
386 Boniface VIII as it was regarded as an abominable custom for Christians. About 50 years
387 before the ban, in due respect to his testimonial will, it is reasonable to assume that the dead

388 body of Jacques de Vitry has experienced dismembering and subsequent treatment of the
389 remains. It is also noteworthy that a left tibia put in a wooden frame was kept as a separate
390 relic (Fig 2 C). No clear explanation has been given so far about the origin of this relic,
391 except that it is attributed to Jacques de Vitry thanks to a label present in the frame and
392 bearing the following notice in French: “Ceci est un ossement du Vénérable Jacques de Vitry.
393 Cardinal et Evêque d’Acre en Palestine. Mort à Rome le 30 Avril 1240. Transporté à son
394 Monastère D’Oignies l’année suivante, d’où provident cette précieuse Relique” wich means:
395 “This is a bone from the Respectable Jacques de Vitry. Cardinal and Bishop of Acre in
396 Palestine. Died in Rome on May 1st, 1240. Transported to his Monastery of Oignies next
397 year, from where this invaluable Relic comes.” Therefore, a question naturally arises
398 regarding the belonging of this framed tibia to the remains found in the reliquary.

399 **Are the remains those of Jacques de Vitry?**

400 In the absence of historical portrait or description, traits of Jacques de Vitry remain
401 unknown. No information is available regarding his stature, physiognomy and health.
402 Moreover, we face the case of a secondary burial, not the original one. His age at death is
403 uncertain: between 60 and 75. Regarding the sex, it turned out that the morphology of the
404 (incomplete) skeleton did not allow us to determine it. Therefore, we decided to resort to
405 genetic analysis with the hope to be able to determine the sex from preserved DNA.
406 Regarding the age, anthropological results (dental wear, cranial vault and tooth cementum)
407 tend to support the contemporary hypothesis regarding the birth year of Jacques de Vitry,
408 with a slightly lower age estimate. The biological profile (age, sex, stature) obtained from the
409 anthropological study did not allow us to deliver a verdict about the attribution of the remains
410 to Jacques de Vitry. Cut marks on bones, on the other hand, are compatible with
411 contemporary medieval practices used for hygienic and safe repatriation of dead human
412 bodies. Evidences of cuts on several bones and around joints suggest dismembering of the
413 corpse [42-44] in order to facilitate post mortem transfer. This hypothesis is supported by
414 historical reports of contemporary dismembering practices used for remote burial and agrees
415 with the translation of Jacques de Vitry remains from Rome to Oignies, on a distance of more
416 than 1400 km. According to historical recipes, dismembering was followed by boiling of
417 bones in water, wine or vinegar [39-41]. Such treatments would have damaged seriously the
418 genetic material in the bones [45, 46].

419 The results of isotopic analyses of the remains (Table 1) show that the left tibia, the right tibia
420 and the skull are from the same individual. Taking into account the standard deviations on the
421 ^{14}C ages, there are no significant differences between the dates. The upper bound of average
422 ^{14}C dates (1170AD) is anterior (70 years offset) to the year of death of Jacques de Vitry
423 (1240AD). On the other hand, the stable isotopes analyses indicate a mainly terrestrial diet. If
424 this happened to be the case, the skeleton could not be attributed to Jacques de Vitry.
425 However, the $\delta^{15}\text{N}$ value is quite enriched, which may indicate a small amount of fish
426 consumption since a carnivore has a $\delta^{15}\text{N}$ value of +8‰ (S1 Table) [47]. Based on the
427 depleted $\delta^{13}\text{C}$ value, it can be assumed this comes from freshwater fish and not marine fish,
428 which has a more enriched $\delta^{13}\text{C}$ value (compare Table 1 and S1 Table). Freshwater fish can

429 have reservoir ages ranging between several hundreds of years and two thousand years, as
430 demonstrated in [48]. A minimal consumption of freshwater fish with a large reservoir age
431 can explain the offset between the year of death of Jacques de Vitry and the obtained
432 radiocarbon ages. If this reservoir effect is present, then the remains are likely to belong to
433 Jacques de Vitry.

434 The objective of the genetic analysis was to determine the sex of the individual whose
435 remains were found in the reliquary. Based on the DNA results for the petrous bone and two
436 upper teeth (sex determination with amelogenin gene and analysis of Y-chromosomal STRs),
437 we are able to conclude that they originate from a male individual.

438 **Jacques de Vitry's mitre**

439 Jacques de Vitry bequeathed several personal objects, which constituted, among
440 others, the Treasure of Oignies. Among these, the mitre having miniatures on parchment is
441 not only unique but also intriguing: questions about his usage, origin and fabrication have not
442 been addressed so far. The results of proteomic analyses showed that parchment of the cap
443 was made from sheep, except the lower part where calf was identified. On the other hand,
444 parchment of both lappets was made of calf. This difference of origin species between
445 different parchment parts of the mitre is puzzling and has not been reported before. Was this
446 choice of different animal species intentional or not? If yes, was it motivated by differences
447 in parchment quality, price or availability according to the species? All these questions
448 certainly deserve further investigations from the point of view of the history of art.

449 **Conclusion**

450 An interdisciplinary study was carried out around the reliquary of the late cardinal
451 Jacques de Vitry, a prominent clergyman and theologian who was active in Europe and
452 Middle East during the first part of the thirteen century. Results of anthropological, isotopic
453 and genetic analyses provided evidence that the likelihood that the remains found in the
454 reliquary are those of Jacques de Vitry is very high. Parchment parts of a mitre having
455 belonged to Bishop Jacques de Vitry were analysed non-invasively by proteomic techniques
456 and found to be made of different animal species. These findings are expected to fertilize
457 knowledge carried by historical tradition around the relics of Jacques de Vitry and his related
458 cultural heritage.

459

460

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481 **Author contributions**

482 Conceived the study and requested authorizations for carrying analyses on Jacques de Vitry's
483 reliquary and mitre: FL AC. Genetic analyses: RD FT JYM. Anthropological analyses: CP.
484 Isotopic analyses: MB. Proteomic analyses: MD. Mitre sampling: CC. Coordination of the
485 writing of manuscript: OD. All authors discussed the results and approved the manuscript.

486

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619 **Supporting information**

620 **S1 Fig. Jacques de Vitry and Marie d'Oignies.** (A) Cardinal Jacques de Vitry on his deathbed (engraving).
621 (B) Saint Marie d'Oignies (engraving).

622 **S2 Fig. Reliquary of Jacques de Vitry.** (A) The reliquary (© Vedrin, Guy Focant). (B) The remains found in
623 the reliquary after opening on 8th September 2015 (© Vedrin, Guy Focant).

624 **S3 Fig. Non-invasive sampling of parchment parts of Jacques de Vitry's mitre.** Gentle rubbing of the
625 parchment surface with a PVC eraser for proteomic analyses (© Vedrin, Guy Focant).

626 **S1 Table. Isotopic fractionations of carbon ($\delta^{13}\text{C}$) and nitrogen ($\delta^{15}\text{N}$) according to diet.**

627

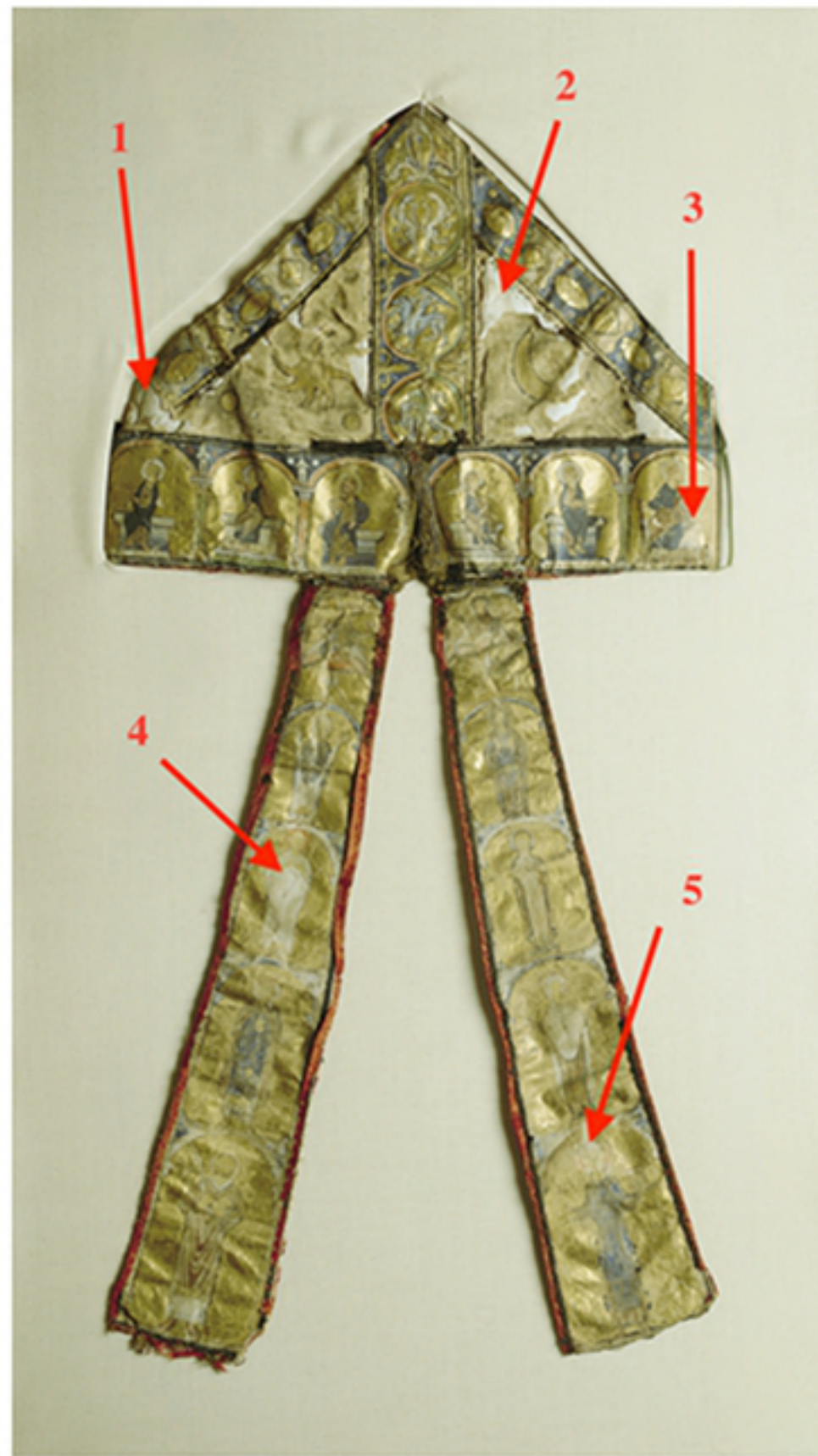
Bone collagen from animals having a 100% diet of	$\delta^{13}\text{C}$ (‰)	$\delta^{15}\text{N}$ (‰)
C3-plants	-21	+5
Meat C3-herbivores	-18	+8
C-4 plants	-7	+5
Marine food	-13	+18
River fish	-24	+16
Lake fish	-20	+16

628 Data reproduced from Lanting and van der Plicht (1996, 1998).

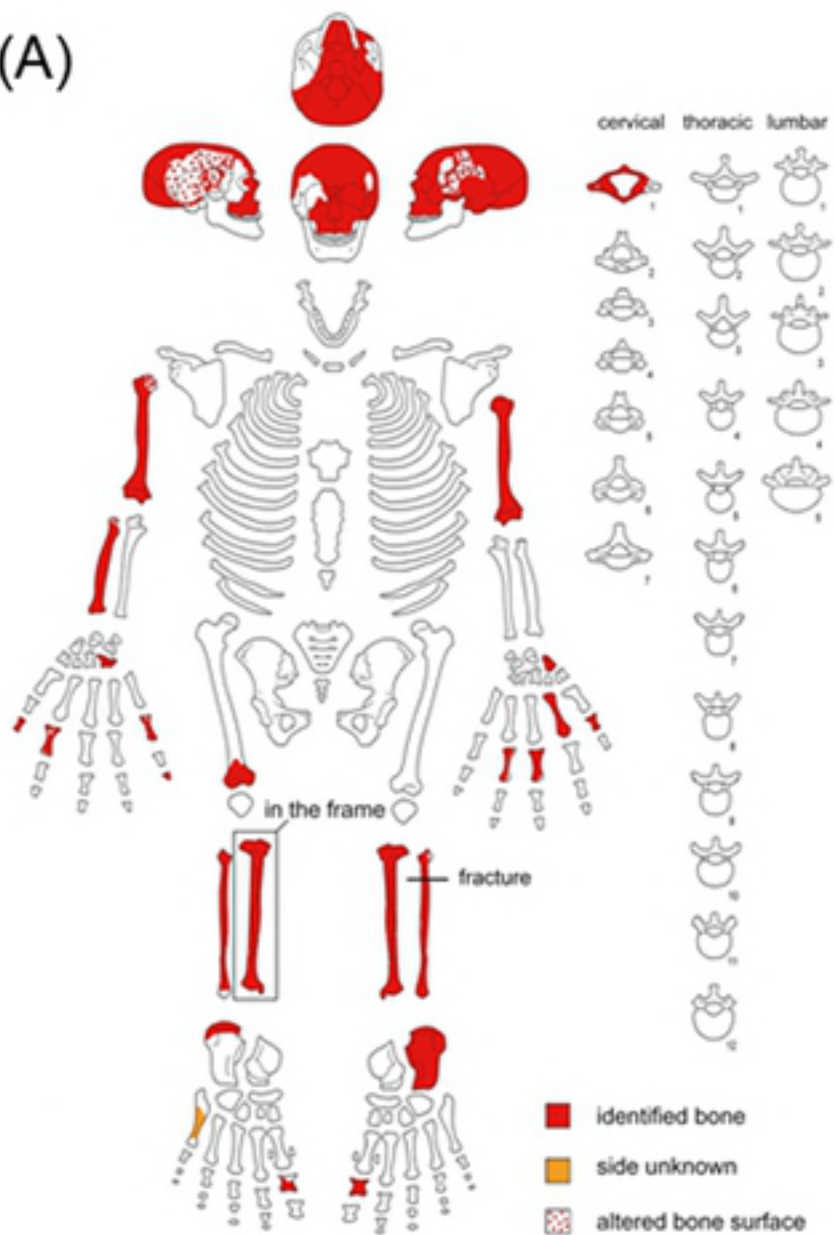
(A)



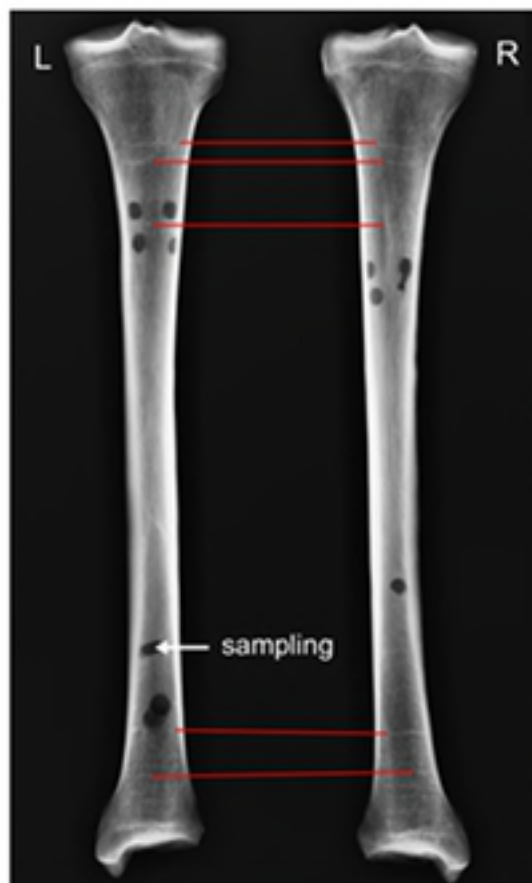
(B)



(A)



(B)



(C)

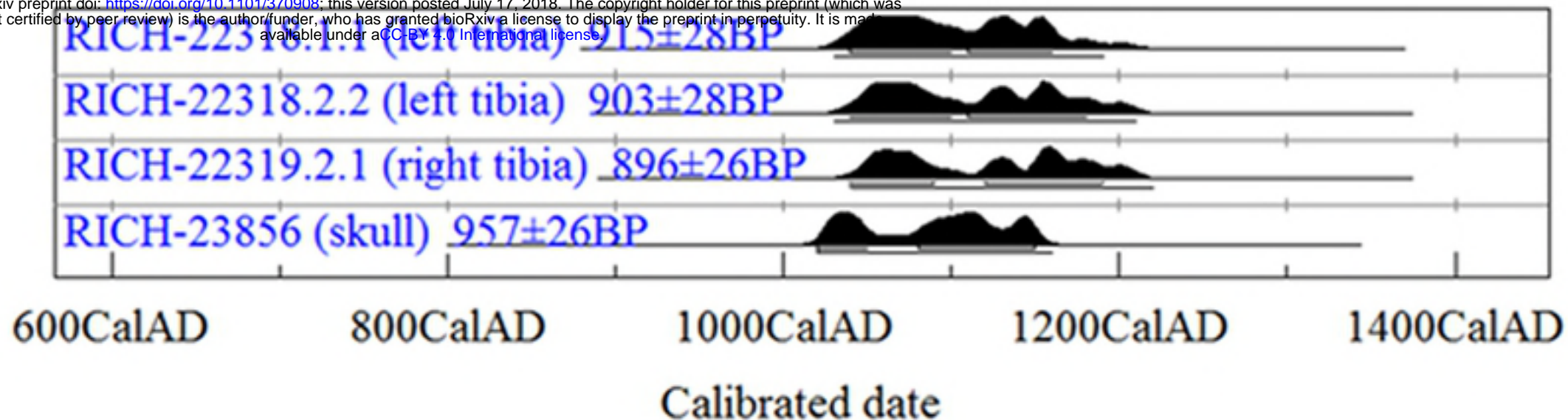


(D)

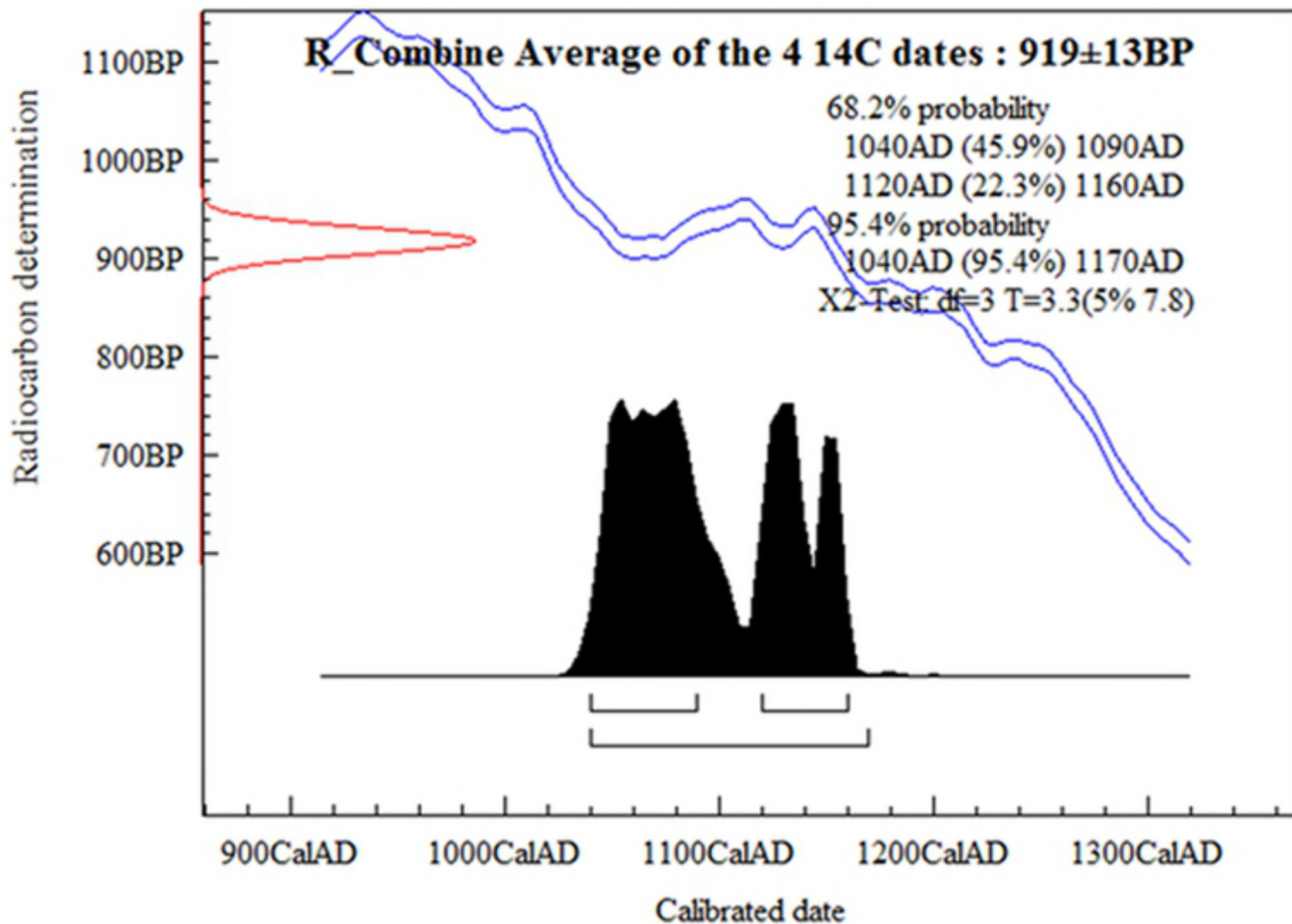


(A)

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(B)



DYS19

DYS389II

DYS448

DYS456

DYS635

- LT RT S +

LT RT S +

LT RT S +

LT RT S +

LT RT S +

