The landscape of coadaptation in *Vibrio parahaemolyticus*

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Abstract

Vibrio parahaemolyticus is a human gastrointestinal pathogen that thrives in warm brackish waters and is unusual amongst bacteria in having a population structure that approximates panmixia. We take advantage of this structure to perform a genome-wide screen for coadapted genetic elements. The great majority of interactions that we detect are between accessory genes, many involved in carbohydrate transport and metabolism, providing evidence for a plug and play-like architecture, in which elements have evolved to be functional immediately on arrival in a new genome. The most complex co-adaptations we identify include hundreds of core genome SNPs as well as accessory genome elements and involve genes encoding lateral flagella and cell wall biogenesis, implying that several genetically distinct strategies have evolved for colonizing surfaces. Our results provide evidence that, as in human relationships, coadaptation involves progressively increasing levels of commitment, with the most involved interactions becoming irreversible and presaging speciation.

Introduction

Biological life is made possible by the cooperative action of thousands of genes. Much insight in to the way that genes interact with each other to produce complex phenotypes has been gained by experiments such as systematic knock out screens in yeast (Costanzo et al. 2016), *Escherichia coli* (Babu et al. 2011) and other organisms and clonal evolution in controlled environments (Tenaillon et al. 2012, Jerison and Desai 2015), but it is challenging for any laboratory experiment to characterize the complexity of real environments or the fitness effects of natural polymorphisms. Genes in the same genome co-adapt to each other progressively, with the course of evolution depending on the variants that segregate in the population as well as the fitness landscape, which may itself change over time. Therefore, to understand the role interactions between genes play in the evolution of complex phenotypes, laboratory experiments need to be complemented by studies of natural variation.

One approach to identifying fitness interactions is to identify statistical associations between genetic variants segregating within natural populations (Campo et al. 2008, Kryazhimskiy et al. 2011, Cui et al. 2015, Figliuzzi et al. 2016, Skwark et al. 2017, Puranen et al. 2018). If variants are found together more frequently than expected, this might indicate that they function together to improve the fitness of the organism. There are three major challenges in its application. The first is obtaining a large enough sample to achieve statistical significance to detect associations, given the enormous number of potential interactions are possible between variants throughout the genome. The second is that it is difficult to separate historical contingency from adaptation. Are genetic variants found together in the same organisms because they are beneficial together or simply because they have been inherited together? The third is that most methods are based on detecting pairwise interactions, but selection on quantitative traits or other complex genetic interactions might also be common (Kuzmin et al. 2018).

Here we address the first two of these challenges by studying patterns of linkage

disequilibrium in a panmictic bacterial population, in which recombination has been

extensive enough to largely scramble genetic variation (Yang et al. 2018, manuscript in

preparation). *Vibrio parahaemolyticus* lives in coastal waters and causes gastroenteritis in

humans and economically devastating diseases in farmed shrimps and is capable of

replicating in less than 10 minutes in appropriate conditions (Makino et al. 2003), while the

doubling time in the wild of the related bacteria *Vibrio cholerae* has been estimated as

slightly over an hour (Gibson et al. 2018). Approximately 0.017% of the genome recombines

each year (Yang et al. 2018), while there are thousands of cell divisions per year, meaning

that for any given pair of interacting loci there are hundreds of thousands of generations of

selection, on average, between recombination events affecting either one of them. As a result,

even epistatic interactions that involve only small selective coefficients s (for example $s = 1.0$

$\times 10^{-4}$) can create an imprint on the genome in the form of strong linkage disequilibrium

(Arnold et al. 2018), that can potentially be detectable with relatively modest sample sizes.

We have taken a conservative approach to identifying statistical associations, rigorously

filtering the set of genomes used in our discovery dataset to eliminate any hint of population

structure. We performed more than 25 billion Fisher exact tests between variants in the core

and accessory genomes, using a cut-off of $P < 10^{-10}$ with the aim of assembling a

comprehensive list of common genetic variants that have strong linkage disequilibrium

between them due to fitness interactions. We find that the great majority of interactions

involve small numbers of accessory genome elements, with surprisingly little involvement of

the core genome. However, we also identify three complex multi-locus interactions, in which

core and accessory genomes have evolved in parallel to create coadapted gene complexes

with distinct strategies. Our approach and results provide a starting point for systematically

investigating fitness landscapes in natural populations, suggests that organisms progressively

modify their own landscape through coadaptation and highlights the fundamental importance

of lateral flagella variation to *V. parahaemolyticus* ecology.

Results

Detection and characterization of interaction groups

To avoid false positives due to population structure, we restricted our initial analysis to the

strains from the Asia population, VppAsia, within our global collection of 1,103 isolates and

iteratively removed isolates until there was no sign of clonal structure (Methods), leading to a

discovery dataset of 198 strains (Figure 1a, Figure S1). We performed a Fisher exact test of

associations between all pairwise combinations of 151,957 SNP variants within the core and

14,486 accessory genome elements. As has been observed previously (Cui et al. 2015), most

of strong associations occurred between sites within 3 kb on the chromosome (Figure 1b). In

order to exclude associations that arise only due to physical linkage, we excluded all sets of

associations that spanned less than 3 kb, including between accessory genome elements. This

left us with 452,849 interactions with $P < 10^{-10}$, which grouped into 90 networks of

associated elements, all of which involved at least one accessory-genome element, with 8

also including core genome SNPs and 38 of which included multiple genome regions. In total

these networks included 1,936 SNPs in 110 core genes and 1593 accessory genome elements.

Interacting SNPs were substantially enriched for non-synonymous variants (Table 1), which

is unsurprising given the role of natural selection in generating associations.

The largest network (network 1) accounted for the majority of interacting SNPs as well as a

significant fraction of accessory genome elements (Table 1). Hierarchical clustering of the

interacting elements in this network based on *P* values (Figure 1c) revealed a complex pattern

of interactions but with differentiated sub-networks and we defined three large interaction

groups (IGs), IG1, IG2 and IG3 within it, placing remaining interactions within IG4. IG4-93

are displayed in Figure 2 and 3 and Figure S2, while IG1-3 are shown separately in Figures 4

and 5.

We compared our results for core genome interactions with those obtained by SuperDCA, a

method that uses Direct Coupling Analysis to identify causal interactions (Puranen et al.

2018), using the default settings for the algorithm. The most important discrepancy is that

although IG1 involves perfect associations, with P values as low as 1.4×10^{-30}, the coupling

strengths are lower than for the other groups we identified and none appear amongst the top

5,000 couplings. This discrepancy is due the large number of SNPs involved with similar

association patterns, which means that coupling values are distributed between them (Figure

1d, Figure S3). Excluding IG1 SNPs, there is a strong correlation between SuperDCA

coupling strengths and Fisher exact test P values (Figure 1e). And at a stringent cutoff of 10^{-}

$^{2.2}$, SuperDCA identifies the same multilocus interactions as Fisher exact test does at $P <$

10^{-10}, with a few SNPs excluded (Figure 1f). The significance thresholds for both the Fisher

exact test and SuperDCA could be relaxed to identify a substantially larger number of true-

positive hits, at the likely expense of some false ones, but we do not investigate these

associations further here.

For IG1-4, COG classes M and N, cell wall biosynthesis and cell motility, are substantially

overrepresented, relative to their overall frequency in the genome (Figure 2c, Fisher exact

test, $P < 0.01$). Amongst the other interaction groups, class G, encoding carbohydrate

transport and metabolism are substantially overrepresented (Fisher exact test, $P < 0.01$),

particularly amongst groups involving incompatibilities. There are also differences in GC

content between accessory genes in IGs and others (Figure 2c), with IGs having higher mean

values, especially in compatibility IGs.

Structure of variation within interaction groups

For each interaction group, we investigated how genetic variation was structured within the

V. parahaemolyticus population using a larger "non-redundancy" set of 469 strains, which

includes isolates from all four of the populations identified in (Yang et al. 2018), but excludes

closely related isolates, differing at less than 2,000 SNPs. We performed hierarchical

clustering of the strains based on the interaction group variants (Figure 3, 4, 5, S2).

The most common type of interaction group is a single accessory genome region of between

3 kb (IG53) and 57 kb (IG41), of which there are 52. For example, IG5 consists of 10 genes

in a single block. 9 code for various functions related to carbohydrate metabolism and

transport while the 10th is a transcriptional regulator (Figure 3 and Table S2). Four strains

have 9 out of the 10 genes but otherwise the genes are either all present or all absent in every

strain. Interestingly, there appears to be a difference in frequency between VppAsia isolates

and others, with the island present in 52% of VppAsia strains and 90% of others.

Genome islands are often associated with transmission mechanisms such as phage and

plasmids (Dobrindt et al. 2004). In our data, one example is IG17 which contains 4 genes

annotated as being phage related and a further 19 hypothetical proteins (Figure 3). In the

reference strain, 16 of the genes occur in a single block VP1563-VP1586, while 7 genes are

present elsewhere in the genome. 56% of the 469 had none of the genes, while 36% had more

than 12 of them and 8% had between 1 and 5, which presumably represent remnants of an old

phage infection. Only one gene (VP1563) in IG17 was found in appreciable frequency in

strains that had none of the other genes. This gene might represent cargo of the phage

infection that is able to persist for extended periods in the absence of infection due to a useful

biological function of its own.

Another common interaction is incompatibility between different accessory genome

elements, which is found in total 27 of the interaction groups (Figure 2). For example, all of

the 469 strains either have the gene yniC, which is annotated as being a phosphorylated

carbohydrates phosphatase, or at least 4 out of a set of 5 genes VP0363-VP0367 that includes

a phosphotransferase and a dehydrogenase (IG6, Figure 3) in the same genome location. Only

one strain has both sets of genes. This interaction also involves a core genome SNP, in the

adjacent gene VP0368, which is annotated as being a mannitol repressor protein. Another

pattern is found in IG45, where there are three genes that are mutually incompatible in our

data, and 12% strains have none of the three genes. Only one of the three genes is annotated.

More detailed characterization of IG1-3

For IG1, the clustering revealed that strains fall into two cleanly differentiated groups (Figure

4). Amongst the 469 non-redundancy set, there are 43 strains from VppAsia and 1 strain from

VppUS2 in the minor group that we call eco-group 1.2, or EG1.2, with the remaining 425

belong to EG1.1. Chromosome painting (Figure 4b) shows evidence for sharp peaks of

differentiation of EG1.2 strains around the IG1 loci but with little evidence for differentiation

elsewhere (see the expected value revealed by vertical dot line). This pattern is entirely

different either for that observed between geographically differentiated populations or for

clonally related strains, which show higher-than-expected copying throughout the genome

but without sharp peaks (Figure S4). The composition of accessory genes also differs

markedly between eco-groups, in particular because EG1.2 strains has a particularly complex

polymorphic region (Ref2-09, 82 kb, Figure 4b) with substantial variation in gene content

between strains, which in total constitutes for a large fraction of the gene content (57 genes)

not shared with EG1.1 strains. Based on the annotation of these genes (Table S2), this region

might encode a type II secretion system.

Based on the clustering we split the genetic variants themselves into three tiers, shown in red,

blue and green in Figure 4. Tier 1 variants distinguish cleanly, or nearly cleanly between

EG1.1 and EG1.2 strains. Tier 2 variants are similar but with more discrepancies, while Tier

3 variants are typically fixed or nearly fixed within one eco-group and polymorphic within

the other. Each of the tiers includes multiple core and accessory genome regions. Some

regions are represented in only one tier, while others contain two or more. For example, the

lateral flagellar gene cluster (Ref1-23, VPA1538-VPA1557, Figure 4c) contains 383 Tier 1

SNPs, 24 from Tier 2 and 47 from Tier 3, however the extent of the region spanned by Tier 1

SNPs is smaller, so that for example flhA and flhB, at the center of the gene cluster only

contain Tier 3 SNPs, while lafK and lafA to the left and filS to the right contain multiple

fixed differences between eco-groups. In other regions of this genome cluster, Tier 1 SNPs

are flanked by those from other tiers.

IG2 includes 548 SNPs and 130 accessory genes which located in 26 regions (Figure 5a),

including a LuxR family transcriptional regulators, T6SS (type VI secretion system), and

cellulose synthesis related genes that had been identified in our previous research (Cui et al.

2015). Here with increased statistical power with more genome sequences, we found

additional interacting loci. Tier 1 contains the T6SS and the cellulose synthesis related genes,

reflecting their strong incompatibility, with nearly all EG2.1 strains containing a T6SS and all

those in EG2.2 containing genes annotated as part of the biosynthesis cluster. Tier 2 contains

mostly SNPs in core genome, which located at LuxR family transcriptional regulator and

uridine phosphorylase encoding genes. Tier 3 contains 85% of all variants in IG2, which

encoded genes that functional related with biogenesis of elements in cell membrane,

carbohydrate transport and metabolism, and transcriptional regulators. Furthermore, the

overall strength of the associations in the IG2 Tiers we have defined are much weaker than

those for IG1: while there are a handful of core genome regions where EG2.1 regions are

differentiated (Figure 5a), none of these is as clear-cut as the differentiated regions of IG1.

The loci of IG3 only covered eight genome regions, which contained 75 SNPs and 65

accessory genes, most of which are found in a single genome regions which includes the core

gene TonB3 (Tanabe et al. 2011)(Figure 5b). Clustering based on variants of IG3 revealed a

stair-like structure of variation. Tier 1 variants differentiate EG3.1 from other ecogroups and

include multiple SNPs in TonB3 and in the lateral flagellar gene cluster. Tiers 2 and 3

variants differentiate between EG3.2, EG3.3 and EG3.4, which are progressively more

different from EG3.1, both in terms of accessory genome complement and core genome

SNPs. Many of the accessory genes specific to EG3.4 are annotated to have functions

associated with cell wall biogenesis.

Phenotypic differences between EG1.1 and EG1.2

We performed a preliminary analysis of the phenotypic differences underlying EG1 by

determining the motility, growth rate, and biofilm formation ability (Figure 6) of in 7 EG1.1

and 4 EG1.2 strains on laboratory media. We failed to observe differences in swimming or

swarming capability under the conditions tested but EG1.2 strains revealed significantly

higher biofilm formation ability and faster growth rate than EG1.1 strains (Figure 6b and 6c),

and they revealed rough colony morphology, also a indication of increased biofilm formation,

under low salinity (1% NaCl) culture condition (Figure 6c).

There are in total 60 EG1.2 strains in the global collection of 1103 *V. parahaemolyticus*

strains. All but one, a VppUS2 isolate, is from the VppAsia population, with the majority (n=

48, 80%) in this study coming from routine surveillance on food related environmental

samples, including fish, shell fish, and water used for aquaculture. The strains revealed no

clear geographical clustering pattern in China, as they can be isolated from all six provinces

that under surveillance. Notably, only 4 EG1.2 strains were isolated from clinical samples,

including wound and stool, representing a lower proportion than for EG1.1 isolates

(453/1043), including if the two pandemic lineages are removed (268/798), suggesting this

eco-group has low virulence potential in humans.

Discussion

Bacterial traits such as pathogenicity, host-specificity and antimicrobial resistance naturally attract human attention but less obvious or even cryptic traits might be more important in determining the underlying structure of microbial populations. Studies of coadaptation based on genome sequencing of thousands of isolates have the potential to provide new insight into the ecological forces shaping natural diversity, and how that variation is assembled by individual strains to overcome the manifold challenges involved in colonizing specific niches, such as the human gastrointestinal tract. In other words, these studies provide a unique opportunity to see the world from the point of view of a bacterium.

We performed a genome wide scan for coadaptation in *V. parahaemolyticus*, performing pairwise tests for interactions amongst genetic variants and then clustered the significant pairwise interactions into 93 interaction groups. Our results highlight the central role of lateral motility in structuring ecologically significant variation within the species. We also find evidence that interactions move through progressive stages, analogous to differing degrees of commitment within human relationships, namely casual, going steady, getting married and moving out together (Figure 7).

Most interactions between core and accessory genomes are casual

A recent debate about whether the accessory genome evolves neutrally (Vos and Eyre-Walker 2017, Andreani, Hesse, and Vos 2017, Shapiro 2017) highlighted how little we know about the functional importance of much of the DNA in bacterial chromosomes. Using the same statistical threshold to assess significance, our interaction screen identifies many more examples of coadaptation between different accessory genome elements than of interactions

between the core and accessory genomes or within the core genome, implying that natural

selection has a central role in determining accessory genome composition.

Unsurprisingly, given the extensive literature highlighting the importance of genomic islands

to functional diversity of bacteria (Dobrindt et al. 2004), the most common form of

adaptation detected by our screen is the coinheritance of accessory genome elements located

in the same region of the genome. Based on a minimum size threshold of 3 kb, we find 52

(56%) such interactions, the largest of which is 57 kb (Table S3).

Previous approaches to detecting genome islands emphasize traits associated with horizontal

transmission, for example based on differences in GC content with the core genome, the

presence of phage-related genes or other markers of frequent horizontal transfer (Langille,

Hsiao, and Brinkman 2010). Our approach, based simply on co-occurrence identifies a wider

range of coinherited units and suggests that many islands have functions related to

carbohydrate metabolism.

Amongst interactions not involving physical linkage, the most common is incompatibility of

different accessory genome elements, representing 29% (27/93) of our IGs. For example, two

different versions of phosphorous pathway, one involving one gene, the other involving 5

(IG6, Figure 3).

We propose that the rarity of interactions between core and accessory genome in our scan

reflects the evolution of "plug and play"-like architecture for frequently transferred genetic

elements. Accessory genome elements are more likely to establish themselves in new host

genomes if they are functional immediately on arrival in many genetic backgrounds, while

from the point of view of the host bacteria, acquisition of essential functions in new

environments is more likely if a large fraction of accessory genome elements are immediately

functional without requiring adaptation of either the element itself or the rest of the bacterial

genome.

Sometimes it makes sense to go steady

When an accessory genome element with an important protein coding function arrives in a

new genome, it is likely that some optimization of gene regulation will be possible,

coordinating the expression of the gene with others in the genome. We found 11 different

interaction groups involving core and accessory genome regions. One simple example

included a regulatory gene VP0368 and an accessory genome element (IG6, Figure 3). In this

example, it is feasible for the core genome SNP and the associated accessory element to be

transferred together between strains in a single recombination event but coadaptation

involving two or more separate genome regions is likely to slow down the rate at which

strains gain and lose the accessory genome elements involved, which in turn makes further

coadaptation at additional genes more likely.

IG2, is an example of a complex coadaptation involving multiple core and accessory genome

regions. A large majority of strains in our dataset (439/469) either carry a cluster of genes

encoding a T6SS, or a cluster encoding cellulose biosynthesis genes, but few strains have

genes from both clusters (Figure 5a). Cells uses the T6SS to inject toxins into nearby bacteria

(Salomon et al. 2013) and cellulose production to coat themselves in a protective layer

(Tischler and Camilli 2004). Incompatibility might have a functional basis, for example

because cellulose production prevents the T6SS functioning efficiently, or an ecological one,

for example because cells that attack others do not need to defend themselves.

Marriage changes everything

IG1 differs from the other interaction groups in our scan in both the number of associated

regions and the strength of the associations. The interaction group include 454 SNPs in the 19

gene 18 kb lateral flagellar gene cluster (VPA1538-1557, Figure 4c), a further 917 core

genome SNPs in 62 genes and 152 accessory genes in 35 clusters. Strains cluster cleanly into

two groups, the more common group being designated EG1.1 and the rarer EG1.2.

Comparison with closely related species shows that the polymorphisms distinguishing EG1.1

and EG1.2 have evolved *de novo* within *V. parahaemolyticus*, with the EG1.2 variant

undergoing faster evolution (Figure S5).

Many of the accessory-genes and SNPs are in perfect or near perfect disequilibrium with

SNPs in the flagellar gene cluster. These include loci encoding flagellar genes, T2SS and

other membrane transport elements. There are also 285 loci (27%, Tier 3) in weaker

disequilibrium, typically because they are polymorphic in one of the ecogroups. Many of

variants are likely to represent more recently evolved coadaptation. Some of these genes are

also associated with flagella or the T2SS related function but also encompass a broader range

of functional categories, including cell division and amino acid transport and metabolism

(Table S2).

Our laboratory phenotype experiments (Figure 6) suggest that biofilm formation is likely to

be a key trait underlying the different ecological strategies of EG1.1 and EG1.2 but the

variation in phenotypic response at different salinity levels and the absence of measurable

difference in swarming behavior despite the large genetic difference within the lateral flagella

genes highlight some of the manifold difficulties of interpreting natural variation using

phenotypes measured under laboratory conditions.

Despite the extensive differences that have accumulated between EG1.1 and EG1.2, there is

no evidence of restricted gene flow in most of the genome (Figure 4b), and even within the

flagellar gene cluster strongly differentiated regions are separated by a weakly differentiated

one (Figure 4c), implying that the coadaptation is being maintained by selection in the face of

frequent recombination. Initial divergence in flagellar function is likely to have led to

ecological differentiation, which led to bacteria having different nutritional inputs or

requirements and a broadening of the functional categories undergoing divergent selection.

How can the difference between IG1 and the other interaction groups in both the number of

associations and their strength be explained? *V. parahaemolyticus* is ubiquitous in shellfish in

warm coastal waters, within which it occurs at densities of around 1,000 cells per gram, so a

back of the envelope calculation suggests there are likely to be substantially more than 10^{15}

bacteria in the VppAsia population. The species also has a high estimated effective

population size (Cui et al. 2015, manuscript in preparation) and has strong codon bias, which

is often argued to be evidence that even tiny selective coefficients can drive adaptation (Sharp

et al. 2005). Furthermore, recombination only breaks up linkage disequilibrium between loci

slowly. Therefore, weaker and more variable patterns of association within IG2 and IG3 than

in IG1 is unlikely to be a simple consequence of the ineffectiveness of selection and is instead

likely to reflect complexity in the fitness landscape.

Strains gain flexibility by being able to switch between or modulate genomically encoded

strategies by homologous recombination. For example, expression of the T6SS might be

essential for survival in crowded habitats but detrimental in sparsely populated ones.

Recombinants between EG2.1 and EG2.2 at IG2 loci might represent transient adaptation of

strains to their immediate environment or long-term adaptation of intermediate strategies.

Furthermore, the phenotypic consequences of IG2 variants can depend on other loci in the

genome, such as IG1 variants, which is likely to reduce the strength of associations within

IG2. Crucially, the evolution of promiscuity is self-reinforcing because the presence of strains

using multiple strategies in the population also favors the presence of accessory genes and

core gene haplotypes that are fit on multiple backgrounds.

On the other hand, an absence of intermediate genotypes in the population can favor the

evolution of fastidiousness, with particular accessory genes and haplotypes becoming

essential components of some genetic backgrounds but deleterious on others. A likely

scenario is that differences between IG1.1 and IG1.2 at a lateral flagellar gene made

recombinants between the two versions of the gene inviable and also created divergent

selection at a handful of other loci that was largely independent of the external environment

or of interactions with other genes. The evolution of fastidiousness, like the evolution of

promiscuity, can be self-reinforcing, and might have led to progressive increase in the

differentiation of EG1.2 strains from EG1.1 until the coadaptation of IG1.2 variants to each

other and of IG1.1 variants to each other became more-or-less irreversible, like marriage in

England prior to the reign of King Henry VIII.

Coadapted gene complexes as speciation triggers

Running the tape forward, it is easy to envisage the number of coadapted regions of the

genome within IG1 undergoing progressive enlargement, until the entire genome becomes

differentiated. As coadapted regions become more numerous, the proportion of

recombination events between ecogroups that are maladaptive will increase, which might

prompt the evolution of mechanistic barriers to genetic exchange between them.

Mechanisms by which new bacterial species arise are frequently discussed in the literature

(Shapiro et al. 2012, Fraser et al. 2009, Falush et al. 2006) but there is currently little data on

how the process unfolds. IG1 is of interest both as an example of an intermediate stage of

divergence, prior to speciation, and because it suggests that substantial adaptive divergence

between gene pools can precede any barriers to genetic exchange, other than natural selection

at the loci involved. This – unique to our knowledge – example is exciting because the

distinct signature of selection should make it possible to dissect the genetic basis of

coadaptation in unprecedented detail. Broadly similar patterns of differentiation including

"genomic islands of speciation" have been observed for example between ecomorphs of

cichlid fishes (Malinsky et al. 2015), but the evolution of ecomorphs has been facilitated by

fish preferring to mate with similar individuals, which will have also inevitably lead to some

level of differentiation at neutral loci throughout the genome.

Conclusions

In *V. parahaemolyticus*, it has been possible to distinguish clearly between adaptive

processes, reflecting fitness interactions between genes and neutral ones, reflecting clonal and

population structure. This has allowed us to provide a description of the landscape of

coadaptation, involving multiple simple interactions and a small number of complex ones.

We have focused on interactions that generate strong linkage disequilibrium, but weaker and

more complex polygenic ones also have the potential to provide biological insight.

Most bacteria have population structure that deviates more markedly from panmixia

(manuscript in preparation). In some species this is likely due to smaller effective population

sizes, lower recombination rates or mechanistic barriers to genetic exchange between strains.

However, coadaptation can itself generate genome-wide linkage disequilibrium that might be

difficult to distinguish from clonal or population structure. Because the linkage

disequilibrium associated with IG1 is highly localized within the genome, it can, on careful

inspection be clearly be attributed to selection, but in other bacteria patterns are likely to be

less straightforward, making it challenging to understand to whether adaptive processes drive

population structure, or vice versa. Natural selection is the jewel of evolution but

distinguishing it from other processes requires in depth understanding of the relevant biology

in addition to suitable data and statistical methods.

Materials and Methods

Genomes used in this work

Totally 1,103 global *V. parahaemolyticus* genomes were used in this work (Table S1), which

also were analyzed in our another study (Yang et al. 2018). To reduce clonal signals, we

firstly made a "non-redundancy" dataset of 469 strains, in which no sequence differed by less

than 2,000 SNPs in the core genome. They were attributed to 4 populations, VppAsia (383

strains), VppX (43), VppUS1 (18) and VppUS2 (21) based on fineSTRUCTURE result

(Lawson et al. 2012). We then focused on VppAsia which has more strains, to generate a

genome dataset in which strains represent a freely recombining population. We selected 386

genomes from 469 non-redundancy genome dataset, including all the 383 VppAsia genomes

and 3 outgroup genomes which were randomly selected from VppX, VppUS1 and VppUS2

population, respectively. These 386 genomes were used in Chromosome painting and

fineSTRUCTURE analysis (Lawson et al. 2012) as previously described (Cui et al. 2015).

Initial fineSTRUCTURE result revealed multiple clonal signals still exist, thus we selected

one representative genome from each clone, combined them with the remaining genomes and

repeated the process. After 14 iterations, we got a final dataset of 201 genomes with no trace

of clonal signals, involving 198 VppAsia genomes that were used in further analysis.

The copy probability value of each strain at each SNP were generated by Chromosome

painting with "-b" option, and the average copying probability value of a given strain group

(e.g. EG1.2) at each SNP was used in Figure 4, 5 and Figure S4.

Variation detection, annotation and phylogeny

We re-called SNPs from 198 VppAsia genomes by aligning the assembly against reference

genome (RIMD 2210633) using Mummer (Delcher, Salzberg, and Phillippy 2003) as

previous described (Cui et al. 2015). Totally 565,466 bi-allelic SNPs were identified and

151,957 bi-allelic SNPs with minor allele frequency > 2% were used in coadaptation

detection. We re-annotated all the assemblies using Prokka (Seemann 2014), and the

annotated results were used in Roray (Page et al. 2015) to identify the pan-genome and gene

presence/absence, totally 41,053 pan-genes were found and 14,486 accessory genes (present

in > 2% and < 98% strains) were used in coadaptation detection. The pan-gene protein

sequences of Roary were used to blast (blastp) against COG and KEGG database to get

further annotation.

The Neighbour-joining trees were built by using the TreeBest software

(http://treesoft.sourceforge.net/treebest.shtml) based on sequences of concatenated SNPs, and

were visualized by using online tool iTOL (Letunic and Bork 2016).

Detection of coadapted loci

Totally 151,957 bi-allelic SNPs and 14,486 accessory genes identified from 198 independent

VppAsia genomes were used in coadaptation detection by two methods. Firstly we used

Fisher exact test to detect the linkage disequilibrium of each SNP-SNP, SNP-accessory gene,

and accessory gene-gene pair. Presence or absence of an accessory gene was considered as its

two alleles. Each variant locus (SNP or accessory gene) has 2 alleles, major and minor, of

which major represents the allele shared by majority of isolates. For each pair of loci X and

Y, the number of combinations between X_{major}-Y_{major}, X_{major}-Y_{minor}, X_{minor}-Y_{major}, X_{minor}-Y_{minor}

were separately counted and used in the contingency table to calculate the Fisher exact test P

value. It took 3 days to finish all the coadaptation detection in a computer cluster using 21

cores and 2 Gb memory. We also used SuperDCA (Puranen et al. 2018) to detect the

coadaptation between SNPs for comparison. It is based on direct coupling analysis (DCA)

model and has a much faster calculation speed compared with previous DCA methods.

However, it still took 25 days to finish the detection by using 32 cores and 86 Gb memory.

We removed coadaptation pairs with distance less than 3 kb to minimize the influence of

physical linkage. All identified SNPs in this study were located in the core genome, therefore

the physical distance between SNP pairs can be calculated according to their position in the

reference genome. To defined the distance between accessory genes, and between SNP and

accessory gene, we mapped the sequence of accessory genes against available 19 complete

maps of the *V. parahaemolyticus* genomes to acquire their corresponding position, and then

the gene that failed to be found in complete reference genomes were then mapped to the draft

genomes. If the accessory genes pair or SNP- accessory gene pair was found located in a

same chromosome or same contig of a draft genome, then the distance between paired

variants could be counted according to their position in the chromosome or contig. The

distance between paired variants that located in different chromosome or contig was counted

as larger than 3 kb and such pairs were kept in further analysis. Circos (Krzywinski and

Schein 2009) was used to visualize the networks of coadaptation SNPs in Figure 1f and

Figure S3.

Lateral flagellar gene cluster region in *Vibrio* genus

To identify the homologous sequences of *V. parahaemoluticus* lateral flagellar gene cluster

(VPA1538-1557) in the *Vibrio* genus, we downloaded all available *Vibrio* genome sequences

in NCBI, then aligned the nucleotide sequence of lateral flagellar gene cluster of *V.*

parahaemolyticus (NC_004605 1639906-1657888) against *Vibrio* genome dataset

(excluding *V. parahaemolyticus*) by using blastn. Totally 46 *Vibrio* genomes revealed above

than 60% coverage on lateral flagella region in *V. parahaemolyticus* genome and was used in

phylogeny rebuilding. We also included three randomly selected strains from EG1.1 and

EG1.2 respectively for comparison. In total 3,000 SNPs were identified in this region and

were used for NJ tree construction.

Determination of phenotypes

Bacteria strains. In the phenotype experiments, totally 11 strains were randomly selected

respectively from two EGs that defined by IG1 variants, including 7 EG1.1 strains (B1_10,

B1_3, B2_10, B4_8, C1_5, C5_2, C6_5) and 4 EG1.2 strains (B1_1, B3_1, B5_2, C3_10).

The strains stored at -80℃ were inoculated in the thiosulfate citrate bile salts sucrose agar

(TCBS) plates by streak plate method. Five clones for each strains were inoculated again in

another TCBS plate and then cultured overnight at 30℃ in 3% NaCl-LB broth overnight and

used for the following assays.

Motility assays. Five clones for each strain were cultured overnight at 30 ℃ and then

inoculated in the swimming plate (LB media containing 0.3% agar) and swarming plate (LB

agar with 3% NaCl). The swimming ability was recorded by measuring the diameter of

colony after 24 hours at 30 ℃. And the swarming ability was recorded after 72 hours at

24 ℃.

Growth curve. *V. parahaemolyticus* strains in 96-well plate were cultured overnight at 30°C

in 3% NaCl-LB broth. The optical density of each culture was adjust to an OD_{600} of 0.6.Then

1 ml of each culture was inoculated 100 ml of 3% NaCl-LB broth in a 96-well plate and

cultured at 30 °C. The growth of each culture were measured every 1 hour at the optical

density of 600 nm using Multiskan Spectrum.

Biofilm formation. *V. parahaemolyticus strains* were cultured overnight at 30°C in 3%

NaCl-LB broth. 2 µl of each overnight culture was inoculated to 100 µl of 3% NaCl-LB broth

in a 96-well plate and cultured at 30°C for 24 h statically. The supernatant was discarded and

each well was washed once with sterile phosphate-buffered saline (PBS). 0.1% Crystal violet

(wt/vol) was added to each well and incubated at room temperature for 30 min. The crystal

violet was decanted, and each well was washed once with sterile PBS. Crystal violet that

stained biofilm was solubilized with dimethylsulfoxide (DMSO), and then measured at the

optical density of 595 nm using Multiskan Spectrum (Thermo Scientific).

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Author Contributions

D. F., Y. C., and R. Y. designed the study and coordinated the project; Y. C., C. Y., and D. F.

analyzed the data; H. Q. and H. W. performed phenotype experiments; D. F. and Y. C. wrote

the manuscript. All authors approved the final version of the manuscript.

Competing Financial Interests statement

No

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Figure 1. Detection of coadaptation loci in *V. parahaemolyticus*. (a) NJ tree of 198 VppAsia strains based on 151,957 bi-allelic SNPs (minor allele frequency > 2%) (b) Q-Q plot of Fisher exact test *P* values between genetic variants. The vertical dotted line shows the threshold $P = 10^{-10}$. (c) Hierarchical clustering of interacting variants (column and row) in the largest network based on Fisher exact test *P* value. IG2-1 and IG2-2 were integrated into IG2. (d) Distribution of SuperDCA coupling strength value.(e) Correlation between Fisher exact test *P* value and SuperDCA coupling strength. (f) Overlap of strong linked SNP sites detected by Fisher exact test ($P < 10^{-10}$, excluding IG1 variants) and SuperDCA (coupling strength > $10^{-2.2}$). Red for interacted SNP pairs detected by both methods, blue for SNP pairs detected only by Fisher exact test. Only SNP/accessory gene pairs with distance > 3 kb were included in (c,e and f).

Figure 2. Landscape of coadaptation (excluding IG1-3). (a) Gene maps of different IGs. Arrows indicate genes and red for detected coadaptation core genes, blue for accessory genes and orange for genes with no coadaptation signal. Black vertical lines indicate SNPs in the interaction group. The colors of the bar on the left indicates average linkage strength of the loci in each IG. Vertical dotted lines were used to split compatible genes with physical distance larger than 3 kb, or genes located in different contigs, chromosomes and strains. Dotted rectangles indicate incompatible genes. IGs with genome block length larger than 60 kb are broken by double slash and shown in (b) after zooming out. COG classification labels are shown above the genes. (c) COG classification and GC content of all the genes used in detection (top) and of different types of coadaptation genes. Red for core genes and blue for accessory genes. The first number in brackets is the number of genes with COG annotation and the second is the total number of genes in the category.

Figure 3. Representative interaction groups. Hierarchical clustering of 469 non-redundancy strains (columns) based on coadaptation loci (rows) of 4 representative IGs. Colors of the heatmap indicate the status of genetic variants, with light orange/orange for two alleles of a SNP, light yellow/brown for absence and presence of the accessory genes. Bar colors below the tree on the top indicate the populations of strains according to the legend. Function summary of involved genes is shown on the top of each heatmap.

Figure 4. Interaction group 1.

Figure 4. Interaction group 1. (a) Hierarchical clustering of 469 non-redundancy strains (columns) based on coadaptated loci (rows) of IG1. Color scheme of the heatmap is the same as in Figure 3. Branch colors of the tree on the top indicate the populations of strains according to the legend. Branch colors of the tree on the left indicate the tiers of coadaptation loci, with red, blue and green for tier 1, 2 and 3, respectively. The same colors are used for tiers of coadaptation in panel (b) and (c). Colors of the bar on the right indicate the genome position of coadaptation loci, which is corresponding to the bar colors in panel (b). (b) Gene map of coadaptation genome blocks of IG1. Two reference genomes were used to show coadaptation variants in IG1. The left curve indicates the value of average copy probability of EG1.2 strains that copied from themselves throughout the genome of Ref1, with vertical dotted line indicates the expect value (number of EG1.2 strains /number of all strains). The bars indicate the reference genomes and different chromosomes are separated by a horizontal short bar. The labels of coadaptation genome blocks are shown above them and are corresponding to information in Table S2. Arrows and vertical lines separately indicate genes and SNPs, which were colored according to different coadaptation tiers, and core genes were colored grey and genes with no coadaptation signal were light orange. COG classification labels are shown above the genes. (c) The distribution of coadaptation SNPs in the lateral flagellar gene cluster region (VPA1538-1557). The top indicates the gene organization of lateral flagellar gene cluster. Light orange rectangles show the accessory genome region. The histograms indicate the distribution of SNPs along the gene cluster, with colors of bars indicate coadaptation tiers.

Figure 5. Interaction group 2 (a) and 3 (b). Layout and colors are the same as in Figure 4. EG1.2 and EG2.1 strains are shown below the tree on the top for comparison (black bars). The left curves indicate the average copy probability value of EG2.1 (a) and EG3.1 (b) strains copy from themselves throughout the reference genome, respectively. Twenty randomly selected SNPs separately from IG2-1 and IG2-2 were used in hierarchical clustering to minimize the influence of number of variants.

Figure 6. Phenotypes of 7 EG1.1 and 4 EG1.2 strains. (a) Swimming and swarming ability. (b) Growth curve. (c) Biofilm formation and colony morphology. Strains used in experiments were randomly selected and were marked in Figure 4a.

Figure 7. Overview of four stages of coadaptation. Circles indicate bacterial strains within a population. Stars indicate SNPs, with red and green indicating the two alleles. Blue rectangles indicate accessory genes or genome islands. Arrows indicate the transitions between stages. Casual interactions involve genes and SNPs coming and going on all genetic backgrounds. Steady interactions involve particular genes that are associated with each other but with frequent exceptions due to ongoing genetic flux and coadaptation with other loci. Married interactions involve a core of fastidiously associated loci with other loci that lead to further co-adaptation in multiple genome regions. The horizontal line in the fourth stage indicates a barrier to gene flow entailing speciation.

Table1. Summary of interactions detected in coadaptation screen.

| | Total[*] | Network 1 | | | | | IG5-93 | Frequency[***] |
		IG1	IG2	IG3	IG4	Frequency[**]		
SNP-SNP pair	2.3×10^{10}	283632	4398	1150	6	0.00%	22751	0.00%
SNP-Accessory gene pair	2.2×10^{9}	104096	7319	2446	112	0.01%	1188	0.00%
Accessory gene-gene pair	2.1×10^{8}	7999	2823	973	1692	0.01%	12264	0.01%
SNP number	151957	917	548	75	63	1.05%	333	0.22%
Synonymous (Syn)	117541	626	409	49	38	0.95%	226	0.19%
Nonsynonymous (NonSyn)	23673	236	122	21	21	1.69%	107	0.45%
NonSyn/Syn	0.2	0.38	0.30	0.43	0.55		0.47	
Core gene	3936	62	20	6	4	2.34%	18	0.56%
Accessory gene	14486	152	130	65	124	3.25%	1122	7.75%

  * All of the variations used in coadaptation screen.

 ** Ratio of the number of variations in Network 1 to the total number of variations.

*** Ratio of the number of variations in IG5-93 to the total number of variations.