

1 **Title:**

2 **Static stability predicts the continuum of interleg coordination patterns in *Drosophila***

3

4 **Running title:**

5 **Stability predicts coordination patterns**

6

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17

18 **Keywords:**

19 motor control, locomotion, insect walking, stability, interleg coordination

20 **Summary statement:**

21 A simple stability-based modelling approach can explain why walking insects use different
22 leg coordination patterns in a speed-dependent way.

23

24 **Abstract**

25 During walking, insects must coordinate the movements of their six legs for efficient
26 locomotion. This interleg coordination is speed-dependent; fast walking in insects is
27 associated with tripod coordination patterns, while slow walking is associated with more
28 variable, tetrapod-like patterns. To date, however, there has been no comprehensive
29 explanation as to why these speed-dependent shifts in interleg coordination should occur in
30 insects. Tripod coordination would be sufficient at low walking speeds. The fact that insects
31 use a different interleg coordination pattern at lower speeds suggests that it is more optimal
32 or advantageous at these speeds. Furthermore, previous studies focused on discrete tripod
33 and tetrapod coordination patterns. Experimental data, however, suggest that changes
34 observed in interleg coordination are part of a speed-dependent spectrum. Here, we explore
35 these issues in relation to static stability as an important aspect of interleg coordination in
36 *Drosophila*. We created a model that uses basic experimentally measured parameters in fruit
37 flies to find the interleg phase relationships that maximize stability for a given walking speed.
38 Based on this measure, the model predicted a continuum of interleg coordination patterns
39 spanning the complete range of walking speeds. Furthermore, for low walking speeds the
40 model predicted tetrapod-like patterns to be most stable, while at high walking speeds tripod
41 coordination emerged as most optimal. Finally, we validated the basic assumption of a
42 continuum of interleg coordination patterns in a large set of experimental data from walking
43 fruit flies and compared these data with the model-based predictions.

44

45 Introduction

46 Legged locomotion (i.e., walking) is an important behavior for most terrestrial animals; in
47 many species, it is the primary mode of locomotion used in various contexts such as
48 foraging, migrating, finding mates, hunting, or escape. Because of its importance for these
49 behaviors, it can be assumed that walking has become highly optimized during evolution.
50 However, walking is not a fixed behavior and must be adaptable regarding basic parameters
51 like speed or direction. The most prominent of such adaptations is interleg coordination—the
52 temporal and spatial relationship between leg movements. In large vertebrates like dogs,
53 horses, and humans, changes in walking speed are accompanied by changes in interleg
54 coordination, termed *gait* transitions (Alexander, 1989). A gait can be defined as a distinct
55 mode of locomotion used within a particular speed range. For instance, a horse will first walk
56 at low speeds then transition to trot at an intermediate speed and, finally, switch to gallop at
57 high speeds (Orlovsky et al., 1999). The transition between two gaits occurs at a
58 characteristic locomotion speed and is discontinuous regarding at least one parameter
59 associated with walking behavior (Alexander, 1989). It is important to note that gaits are not
60 defined by a particular set of movement parameters but by a discontinuous, rather than
61 gradual, transition.

62 Interleg coordination during walking has also been studied extensively in arthropods, mainly
63 insects (for reviews see Ayali et al., 2015; Bidaye et al., 2017; Borgmann and Büschges,
64 2015; Cruse, 1990; Cruse et al., 2009). As in vertebrates, these animals adapt their interleg
65 coordination as they change walking speed (Graham, 1972; Wahl et al., 2015; Wendler,
66 1964; Wilson, 1966; Wosnitza et al., 2013). Several prototypical patterns have been
67 described in the literature; insects use wave gait coordination at low walking speeds
68 (Hughes, 1952), tetrapod coordination at intermediate speeds, and tripod coordination at
69 high speeds (Strauss and Heisenberg, 1990; Wosnitza et al., 2013). Each of these
70 locomotion modes corresponds to a particular interleg coordination pattern. During wave gait
71 coordination, at most one leg executes a swing phase at any given time, while metachronal
72 waves of protraction progress from the hind to the front leg on each side of the animal's
73 body. In tetrapod coordination, at most two legs are in swing phase at a particular time.
74 Finally, tripod coordination is characterized by concurrent swing phases of ipsilateral front
75 and hind legs and the contralateral middle leg.

76 Commonly, these interleg coordination patterns in insects are referred to as gaits in the
77 literature (Bender et al., 2011; Dürr et al., 2018; Nishii, 2000; Ramdya et al., 2017; Spirito
78 and Mushrush, 1979); however, to our knowledge, it has never been explicitly shown that the
79 different forms of locomotion found in insects actually fulfill the definition of gaits as

80 suggested by Alexander (1989)—namely, that these are discrete modes of locomotion and
81 not merely special cases along a continuum.

82 Based on data from the cockroach *Periplaneta americana* (Hughes, 1952) and the stick
83 insect *Carausius morosus* (Wendler, 1964), Wilson (Wilson, 1966) proposed a set of simple
84 rules for the generation of interleg coordination in six-legged insects. In contrast to the
85 common assumption of actual gaits in insects, these rules predicted that insects should use
86 a speed-dependent continuum of interleg coordination patterns. Wilson also pointed out that
87 these rules should result in the natural emergence of all known coordination patterns,
88 including wave gait-like, tetrapod, and tripod coordination, as part of this continuum.
89 Similarly, Spirito and Mushrush (1979) clearly showed a continuum of phase relationships
90 between legs in walking *P. americana*. Results from *Drosophila melanogaster* support the
91 notion of a continuum of coordination patterns; the tripod coordination strength calculated in
92 a study by Wosnitza et al. (2013) showed no clear discontinuities when analyzed over the
93 complete range of walking speeds.

94 These studies suggest that walking insects change interleg coordination in a speed-
95 dependent, continuous, and systematic manner and either imply, describe, or explain this
96 continuum. However, to our knowledge there has been no explicit attempt to explain why
97 these changes occur (i.e., what the adaptive value of these changes might be). Tripod
98 coordination, which is typically used at high walking speeds, would also be suitable for slow
99 walking; indeed, fruit flies can also use tripod coordination at lower speeds (Gowda et al.,
100 2018; Wosnitza et al., 2013). However, there is no directly evident reason why insects should
101 shift to different, more tetrapod-like interleg coordination patterns at low speeds. The fact that
102 a tendency for this shift nevertheless can be observed in most insects suggests that some
103 aspect of the shift to other interleg coordination pattern must be more optimal at lower
104 speeds as compared to tripod. Of course, exceptions are known: dung beetles (genus
105 *Pachysoma*), for instance, sometimes use a peculiar galloping gait (Smolka et al., 2013), and
106 *P. americana* can switch to quadrupedal and even bipedal running during high speed escape
107 (Full and Tu, 1990).

108 In the present study, we explored the question of why walking insects change interleg
109 coordination in a speed-dependent manner. In large animals, energy optimality is typically
110 assumed to be the crucial factor responsible for the emergence of true gaits (e.g., Hoyt and
111 Taylor, 1981). Here, we consider static stability during walking as a potentially important
112 parameter. To investigate the influence of stability on coordination, we devised a compact
113 model that incorporates several kinematic parameters that are known from walking fruit flies
114 (*D. melanogaster*), such as swing duration, stance amplitude, and stance trajectory. Fruit
115 flies spontaneously walk at various speeds, so data from these animals is well suited to

116 explore a large range of walking speeds (Mendes et al., 2013; Strauss and Heisenberg,
117 1990; Wosnitza et al., 2013). The model was used to exhaustively test all theoretically
118 possible coordination patterns (defined herein as phase relationships between ipsilaterally or
119 contralaterally adjacent legs) for all experimentally observed walking speeds in *Drosophila*.
120 The predicted phase relationships between legs were then compared with a large body of
121 corresponding data from walking flies.

122 The results herein suggest that static stability plays a role in the selection of interleg phase
123 relationships. At high reference walking speeds, our model predicts that tripod-like
124 coordination is the optimal coordination pattern. This changes when the reference speed is
125 lowered to speeds that, in the fruit fly, are found in the intermediate or slow range; here, legs
126 are less tightly coupled, and the animal takes advantage of more stable coordination
127 patterns. The patterns predicted by the model resemble tetrapod-like and wave gait-like
128 coordination. Importantly, the model predicts a continuum of coordination patterns that
129 smoothly vary with walking speed. Experimental data confirm that walking flies shift their
130 coordination in a similar way; their motor output seems to also reflect not only theoretically
131 attainable stability but also how robustly such stability can be realized in the presence of
132 locomotor variability.

133

134 **Materials and Methods**

135 **Stability model**

136 Based on previous experimental findings (Wosnitza et al., 2013) we created a model that
137 incorporates several key aspects of walking in *D. melanogaster* and explicitly addresses the
138 speed-dependent nature of inter-leg coordination. The model makes the following
139 assumptions:

- 140 1. The duration of stance phase depends on walking speed.
- 141 2. Each leg's stepping frequency depends on walking speed.
- 142 3. The duration of swing phase does *not* depend on walking speed.
- 143 4. The stance amplitude does *not* depend on walking speed.
- 144 5. The phase relationships between pairs of ipsilateral legs are identical.
- 145 6. The phase relationships between pairs of contralateral legs are identical.

146 Data from a previous study (Wosnitza et al., 2013) validate these assumptions and are
147 presented in Figure 1. Least-squares fitting reveals that swing phase duration and step
148 amplitude are only weakly correlated with walking speed. In contrast, stance phase duration
149 and step frequency are strongly correlated with walking speed. Importantly, both stepping
150 frequency and stance duration can be accurately predicted assuming that swing duration and
151 step amplitude are constant (Fig. 1C and D).

152 The model presented here used these relations and a desired walking speed as a set point
153 to calculate the corresponding stepping frequency and stance duration. These two
154 parameters were then used in conjunction with experimentally measured average stance
155 trajectories (Fig. 2C; data from Wosnitza et al., 2013) to construct one complete step cycle
156 for each leg. All stance trajectories were defined in relation to the center of mass (COM) of
157 the fly. The COM's position was estimated by individual weight measurements of heads,
158 thoraces, abdomina, sets of six legs, and the wings ($n = 30$). These measurements showed
159 that the head contributed 12.5% of the fly's total weight, the thorax contributed 31%, and the
160 abdomen 45%. The combined weight of the legs (11%) and the wings (0.5%) were neglected
161 for the calculation of the center of mass. The head, thorax, and abdomen were then modeled
162 as conjoined ellipses that had the same dimensions and relative positions as their
163 counterparts. Using the individual weights and the positions and dimensions of the modeled
164 body parts, we calculated the position of the COM.

165 During virtual swing movement, a leg's tarsus was lifted off at the posterior extreme position
166 (PEP) and moved to the anterior extreme position (AEP). During the virtual stance
167 movement, the tarsus touched down at the AEP and moved with a uniform speed (i.e., the

168 set walking speed) to the PEP, where it was lifted off again. Two parameters, ϕ_I and ϕ_C ,
169 determined the phase relationships between the legs in this model (Fig. 2D); they can adopt
170 values between 0 and 1. ϕ_I defined ipsilateral phase relationships of step cycles between
171 hind and middle legs and between middle and front legs. Each set of three ipsilateral legs
172 was then treated as a unit (gray outline in Fig. 2D), and the phase relationship between these
173 contralateral units was determined by ϕ_C . Thus, for a particular walking speed and a set of
174 phase relationships, a particular leg's position and whether it was in stance could be
175 determined at a given time. The tarsal positions of the legs simultaneously in stance at a
176 given time were used to determine a support polygon; the minimum distance between the
177 COM and an edge of this polygon was defined as stability (Fig. 2E). Stability was positive
178 when the COM was within the support polygon and 0 when it was outside. When there were
179 fewer than three legs on the ground, stability was undefined and set to 0.

180 For a set walking speed, a stepping frequency and stance duration were uniquely defined,
181 and the average stance trajectories were assumed to be constant. Consequently, there were
182 two adjustable parameters in this model: ϕ_I and ϕ_C . To determine stability for different sets
183 of ϕ_I and ϕ_C , each of the two phases was varied systematically from 0 to 1 in steps of 0.02.
184 For each possible combination of phase relationships, we simulated one complete step cycle
185 and calculated its minimum stability. Thus, coordination patterns for which the COM always
186 remained within the support polygon returned positive values. Those with larger values keep
187 the COM towards the center of the polygon at all times, increasing the margin of stability.

188 **Flies and animal husbandry**

189 Fruit flies (*Drosophila melanogaster*) were raised at a temperature of 25 °C and 65%
190 humidity on a 12-h/12-h light/dark cycle. They were raised on a medium based on a recipe
191 by Backhaus et al. (1984). Experimental data were based on three different fly strains for the
192 experiments described herein: the wild-type strains *Berlin* and *CantonS* (WT, data from this
193 study and Wosnitza et al., 2013) and the mutant strain w^{1118} (data from Wosnitza et al.,
194 2013). These mutant flies have been reported to walk more slowly than wild-type strains, but
195 show no other apparent impairments (Wosnitza et al., 2013). Flies used during experiments
196 were between three and eight days old. Fly data presented in the manuscript were either
197 obtained during free-walking or tethered walking.

198 **Free-walking assay**

199 A schematic of the free-walking setup is shown in Figure 3A. It consisted of an inverted glass
200 petri dish that we used as a transparent arena (diameter 80 mm) held by a circular frame
201 with a cutout below the dish. The cutout provided an unobstructed bottom view of the arena.

202 A surface mirror was placed below the arena at a 45° angle; this allowed for video recordings
203 at approximately the same height as the setup. In conjunction with the mirror, we used an
204 infrared (IR)-sensitive high-speed camera (VC-2MC-M340; Vieworks, Anyang, Republic of
205 Korea) to capture a bottom view of a central rectangular area on the surface of the arena of
206 approximately 30 x 36 mm, with a resolution of 1000 x 1200 pixels, 200-Hz frame rate, and a
207 shutter time of 200 μ s. Illumination was provided by a ring of IR light-emitting diodes (LEDs)
208 arranged concentrically around the arena and emitting their light mainly parallel to the
209 arena's surface. This resulted in a strong contrast between background and fly (see Fig. 3B).
210 The LEDs' activity was synchronized to frame acquisition of the camera. To prevent escape,
211 the arena was covered with a watch glass that established a dome-shaped enclosure, similar
212 to an inverted FlyBowl (Simon and Dickinson, 2010). To keep flies on the horizontal petri
213 dish, we covered the inside of the watch glass with SigmaCote (Sigma-Aldrich, St. Louis,
214 MO). Prior to an experiment, a single fly was extracted with a suction tube from its vial and
215 placed onto the arena, which was then immediately covered with the watch glass. Flies were
216 allowed to explore the arena for approximately 15 minutes, after which video acquisition was
217 started.

218 Flies were spontaneously active in the arena and frequently crossed the capture area. Video
219 data of this area was continuously recorded into a frame buffer of five-to-ten-second
220 durations. During an experiment, custom-written software functions evaluated the recorded
221 frames online and determined if a fly was present and if it had produced a continuous
222 walking track that was at least 10 body lengths (BL) long. Once the fly had produced such a
223 track and either stopped or left the capture area, the contents of the frame buffer were
224 committed to storage as a trial for further evaluation. Video acquisition and online evaluation
225 during acquisition were implemented in MATLAB (2016b; The MathWorks, Natick, MA).

226 **Tethered-walking assay**

227 A schematic of the tethered walking setup is shown in Figure 3C. It is a modified version of a
228 setup described previously (Berendes et al., 2016; Seelig et al., 2010). The setup consisted
229 of an air-supported polypropylene ball (diameter 6 mm) onto which a tethered fly can be
230 placed. Flies placed atop the ball in this manner will show spontaneous walking behavior and
231 use the ball as an omnidirectional treadmill. Ball movements were measured by two optical
232 sensors (ADNS-9500; Broadcom, Inc., San Jose, CA) with an acquisition speed of 50 Hz.
233 Each of these sensors provided information about 2D optic flow at the equator of the ball;
234 combining these data allowed for the reconstruction of the ball's rotational movement around
235 its three axes of rotation. Based on these movements, we reconstructed the fly's
236 instantaneous speed and the curvature of the virtual track during walking. Concurrently, and

237 synchronized to the acquisition of these data, we recorded high-speed video with a resolution
238 of 1200 x 500 pixels from a top view (other parameters and camera model same as above
239 references). Illumination was provided by an IR LED ring positioned around the camera's
240 lens (96 LEDs) and focused onto the fly. Low-level control of the optical sensors and
241 synchronization to the camera was implemented with custom-made hardware (Electronics
242 Workshop, Zoological Institute, University of Cologne), while high-level control and video
243 data acquisition were implemented in MATLAB. To improve visibility of the fly's legs, we
244 placed two surface mirrors on a gantry above the fly. The surface of the mirrors formed an
245 angle of 25° with the optical axis of the camera and, thus, provided two additional virtual
246 camera views (see Fig. 3D). Annotation of leg kinematics was done in these side views.

247 Prior to tethered-walking experiments, flies were cold-anesthetized and transferred into a fly-
248 sized groove in a cooled aluminum block (~4 °C), which held them in place for tethering.
249 Using a dissecting microscope, we then glued a copper wire (diameter 150 µm) to the fly's
250 thorax. For this, we used dental composite (Sinfony™; 3M ESPE AG, Seefeld, Germany) that
251 was cured within a few seconds with a laser light source (wavelength 470 nm). The wire was
252 inserted into a blade holder which, in turn, was attached to a 3D micromanipulator used for
253 exact positioning of the fly atop the ball. Similar to the free-walking condition, flies were given
254 approximately 15 min to familiarize themselves with the ball and the setup, as well as to
255 recover from anesthesia. Kinematic data from the ball and video data from the camera were
256 captured into separate ring buffers. Flies were spontaneously active; here, however, trial
257 acquisition was done manually.

258 **Data annotation and analysis**

259 The position of the fly throughout a trial in the free-walking paradigm was determined
260 automatically. In brief, each video frame was converted into a binary image, in which the fly
261 was detected as the largest area. This area was fitted with an ellipse; its major axis and
262 centroid were defined as the fly's orientation and center, respectively. Walking speed and
263 rotational velocity were calculated as changes of the center and rotation over time. In each
264 trial, the times and positions of all AEPs and PEPs of each leg were determined manually.
265 These positions were then transformed into a body-centered coordinate system based on the
266 fly's center and orientation. In the tethered-walking assay, walking speed and rotational
267 velocity were provided directly by the ball's motion sensors. All positional data (speed and
268 distance) were normalized to BL and subsequent analyses were carried out on these body-
269 centered and BL-normalized data.

270 An individual step was defined as the movement of a leg between two subsequent PEPs.
271 Swing movement was defined as the movement between a PEP and the subsequent AEP;

272 stance movement was defined as the movement between an AEP and the subsequent PEP.
273 The walking speed associated with one step was defined as the average walking speed
274 throughout the step. The instantaneous phase of a step was defined as a value between 0
275 and 1, which progressed linearly over time between the beginning and the end of the step.
276 The phase relationship between a pair of legs was calculated based on the difference
277 between the instantaneous phases of the two legs at the time of the PEP of one of the legs
278 (i.e., the reference leg). All annotations and calculations were carried out with custom-written
279 functions in MATLAB.

280 Results

281 Our model compactly represents possible interleg coordination patterns (ICP). Figure 4A
282 shows a plot of the ipsilateral phase angle, ϕ_I , against the contralateral phase angle, ϕ_C ,
283 which we call a $\phi_V\phi$ plot. Each (ϕ_C, ϕ_I) ordered pair represents one ICP. Once a walking
284 speed is set, the full stepping pattern can be determined based on the invariant features we
285 introduced into our model. Figure 4B-E shows several exemplary ICPs corresponding to
286 particular points in the $\phi_V\phi$ plot; walking speed was set to 5 BL s⁻¹. These examples are
287 meant to give the reader an intuitive understanding of the $\phi_V\phi$ plot. For example, when ϕ_I is
288 1/3, tetrapod-like ICPs emerge (Figure 4B-D). Figure 4B and C illustrate ICPs that have been
289 described in the literature as (ideal) tetrapod patterns. In those, two legs always execute their
290 swing movement at the same time; which legs swing together depends on ϕ_C (either 1/3 or
291 2/3). As we will show, these ideal tetrapod ICPs are not commonly observed in experimental
292 data, where animals typically use ICPs like the one shown in Figure 4D. The $\phi_V\phi$ plot can
293 also describe a tripod ICP (Figure 4E) commonly found in fast-walking insects.

294 The $\phi_V\phi$ plots reveal which ICPs are predicted to be the most stable at each walking speed.
295 Figure 5 shows the stabilities of all ICPs at various speeds (Fig. 5Ai-Hi) and the ICPs that
296 correspond to the most stable values of ϕ_I and ϕ_C (Fig. 5 Aii-Hii). Generally, the area
297 showing non-zero stability decreases as walking speed increases. This indicates that, at low
298 walking speeds, more combinations of ϕ_I and ϕ_C result in stable walking. However, unique
299 maxima (i.e., optimal combinations of ϕ_I and ϕ_C) can be found for each walking speed.
300 These phase values of highest stability (red dots in Fig. 5Ai-Hi) indicate that, at low walking
301 speeds, ϕ_I is approximately 0.2 (Fig. 5Ai) and increases continuously towards values of
302 approximately 0.4 (Fig. 5Hi). ϕ_I will, in fact, converge to 0.5 at even higher walking speeds
303 (data not shown). At the same time, the optimal value for ϕ_C remains 0.5 over the complete
304 speed range. The footfall patterns associated with the optimal ϕ_I and ϕ_C values in Figure
305 5Aii-Hii resemble ICPs found in the literature.

306 As walking speed increases, the stance phase duration becomes shorter, reducing the
307 general size of the stable region in each plot. The model predicts that the variance of both ϕ_I
308 and ϕ_C should decrease as walking speed increases, showing an increasingly smaller range
309 of ϕ_I and ϕ_C during the transitions towards tripod. This decrease in variability has been
310 described in the literature and is also apparent in the experimental data presented here (see
311 Fig. 6).

312 The most stable phase relationships predicted by the model have an anteriorly directed
313 swing phase sequence. This sequence, in which swing phase initiation progresses from the
314 hind leg to the middle leg and ends in the front leg during a complete ipsilateral step cycle,

315 has been described in many studies on six-legged walking in animals, both explicitly and
316 implicitly. The model has not been tuned to adhere to this particular progression; this
317 sequence emerges naturally. Furthermore, as the stability distribution suggests (Fig. 5Ai-Hi),
318 a posteriorly directed sequence, corresponding to ϕ_I values between 0.5 and 1, would be
319 noticeably less stable. This prediction implies a crucial role of the anteriorly directed swing
320 phase progression in walking. It is also noteworthy that the model does not predict the
321 existence of the idealized tetrapod ICP, in which two defined legs simultaneously execute
322 their swing movements. Instead, the model predicts a value of 0.5 for ϕ_C at all walking
323 speeds. The resulting ICPs resemble a tetrapod pattern where, at most, two legs are in
324 swing phase at the same time, but these legs do not enter swing phase simultaneously.

325 The most stable ICP predicted by the model always lies along the line $\phi_C = 0.5$, and its value
326 of ϕ_I depends continuously on the walking speed. To test the model's predictive ability with
327 regard to these values, we analyzed a pooled dataset (collected in this study and Wosnitza
328 et al., 2013) of 9552 steps (average of 1592 per leg). For 4372 contralateral comparisons
329 and 5849 ipsilateral comparisons ϕ_I and ϕ_C were well defined; in total, we analyzed 106
330 trials in 31 individuals. We limited our comparison with the model to steps that were
331 produced at walking speeds between 3 and 10 BL s⁻¹ during straight walking.

332 Figure 6 compares stability-optimal values from the model (red lines) with experimental data.
333 Average contralateral phase relationships cluster around 0.5 (green lines, Fig. 6C-D) over
334 the whole speed range, while average ipsilateral phase relationships increase smoothly from
335 values of approximately 0.35 to 0.5 (green lines, Fig. 6A-B, 6F-G). The predicted
336 contralateral phases are very similar to average experimental data (Fig. 6C-D, red and green
337 lines). In addition, the experimental data's variability decreases towards higher walking
338 speed, which might reflect the reduction in the range of values with non-zero stability (see
339 Fig. 5Ai to Hi). The predicted ipsilateral phases differ noticeably from average experimental
340 data; predicted phase values for ϕ_I are lower than the experimental data. There is, however,
341 a clear tendency towards lower phase values at lower walking speeds. Interestingly, the
342 experimental data seem to be constrained by the optimal phase values predicted by the
343 model at lower speeds, with almost no values below this lower boundary. Figure 5 reveals
344 that the most stable ϕ_I are very close to values associated with low stability or even
345 instability (white arrows). Intuitively, these values correspond to swing movement overlap in
346 ipsilateral neighboring legs (i.e. between hind and middle, or middle and front leg,
347 respectively); any perturbation in the ipsilateral phase relationship that shifts ϕ_I to this lower
348 value will therefore drastically reduce stability. As a consequence, the most stable ipsilateral
349 phase is also the least robust; a small reduction in the ipsilateral phase would destabilize the
350 animal's posture noticeably. Therefore, the animal appears to prefer more robust ICPs to the

351 most stable ICP. This, in turn, is also evident in the contralateral phase angle data, in which
352 the most stable ICP is also the most robust and the animal can realize this exactly.

353 One should also note that the model does not predict the existence of the idealized tetrapod
354 ICP, in which two predetermined legs simultaneously execute their swing movement.
355 Instead, the model predicts a value of 0.5 for ϕ_c at all walking speeds. The resulting ICPs
356 resemble a tetrapod pattern (i.e. at most two legs are in swing phase), but these legs do not
357 enter swing phase simultaneously. Discrete changes in gait, like those observed in walking
358 vertebrates, would be apparent as discontinuities in the experimental phase relationships;
359 none are obvious, though, indicating continuous transitions between ICPs.

360

361 **Discussion**

362 A large body of data shows that walking at high speeds is associated with tripod coordination
363 in insects, while tetrapod-like and wave gait-like coordination patterns are more frequent at
364 lower speeds. The present work questions why insects change their interleg coordination
365 during walking in such a speed-dependent manner. To address this, we created a stability-
366 based model (Fig. 2) for predicting ICPs during walking in six-legged insects. The model
367 takes into account basic kinematic parameters (Fig. 1 and Fig. 2C) found in walking fruit flies
368 and explicitly accommodates walking speed as an important aspect. Using this model, we
369 exhaustively explored ipsilateral and contralateral interleg phase relationships over the
370 complete range of walking speeds and analyzed the influence of these phases on static
371 stability (Fig. 5). Furthermore, we compared the predicted optimal phase relationships to a
372 large body of experimental data measured in the present as well as a previous study
373 (Wosnitza et al., 2013). The results suggest that stability plays an important role in the
374 selection of an ICP at a particular speed. The model predicts several experimentally
375 observed aspects of insect walking. First, ICPs form a continuum spanning the complete
376 range of walking speeds. Furthermore, it predicts constant contralateral phase relationships
377 of 0.5 and a speed-dependence of ipsilateral phase relationships. The model also provides a
378 potential explanation for the experimentally observed reduction in phase variability at high
379 walking speeds, namely the reduced range of phase values that provide non-zero stability.
380 Finally, an anteriorly directed progression of swing phases in ipsilateral legs emerges in the
381 model.

382 **ICPs change continuously with walking speed**

383 The model predicts an animal's preferred ICP at each speed, albeit with some systematic
384 deviation. Furthermore, the speed-invariant contralateral phase angle is predicted to be 0.5,
385 which is also observed in experimental data. The model's prediction of the ipsilateral phase
386 angle represents one boundary in the experimental data and a sharp edge of stability for the
387 model. This suggests that the animal does not use the most stable ICP, but instead prefers
388 slightly less stable but more robust ICP at a given speed. Regardless, the animal does prefer
389 ICPs that are more stable than tripod, demonstrating that the thoracic ganglia do not function
390 as a centralized tripod generator. Instead, it is likely that a combination of central neural
391 mechanisms and mechanical influences contribute to the animal's variable, adaptive
392 locomotion.

393 Our model predicts continuous transitions between ICPs as the walking speed changes,
394 suggesting that fruit flies, and by extension other insects, may not exhibit true gaits like those
395 observed in vertebrates; gait transitions would manifest as discontinuities in such a speed-

396 dependent analysis. Indeed, the experimental data that we collected also showed no
397 evidence of discontinuities indicative of gait transitions. We believe that these data, and
398 those from previous studies in *Drosophila* (Berendes et al., 2016; Wosnitza et al., 2013),
399 support abandoning the term *gait* when referring to insect ICPs, because insect interleg
400 coordination does not fall into discrete coordination patterns. Instead, insect ICPs may be
401 thought of as a continuum of stance durations (Dürr et al., 2018). Based on these findings,
402 we would like to emphasize that walking speed has a strong influence on the parameters
403 measured here (phase relationships and footfall patterns). Studies investigating walking in
404 insects should, therefore, explicitly take into account and measure walking speed to avoid
405 conflating true changes in walking parameters between experimental conditions with mere
406 changes in walking speed.

407 **Idealized tetrapod ICPs are not preferred**

408 Both our model and the data we collected suggest that *D. melanogaster* does not utilize the
409 idealized tetrapod ICP, in which three pairs of legs sequentially enter swing phase together.
410 While our model suggests that the idealized tetrapod with $(\phi_C, \phi_I) = (1/3, 1/3)$ should be a
411 stable ICP (see Fig. 4B and C, as well as Fig. 5), it would be less robust than the observed
412 ICP where $(\phi_C, \phi_I) = (1/2, 1/3)$. This is because small changes to either ϕ_I or ϕ_C from
413 $(\phi_C, \phi_I) = (1/3, 1/3)$ would destabilize the animal, whereas ϕ_C would have to change
414 substantially from $(\phi_C, \phi_I) = (1/2, 1/3)$ to destabilize the animal. Previous studies of walking
415 in *D. melanogaster* have also reported that contralateral legs remain in antiphase at all
416 walking speeds, never giving rise to the idealized tetrapod gait (Strauss and Heisenberg,
417 1990). Keeping contralateral legs in antiphase at all speeds is also consistent with behavioral
418 descriptions of arthropod interleg coordination (Cruse, 1990) and could potentially simplify
419 interleg control.

420 Insect interleg coordination is likely determined by more than just the static stability over the
421 course of one step cycle, because the model predicted more extreme speed-dependent
422 changes in ICP (Fig. 5). This discrepancy might be explained by considering the robustness
423 of the coordination pattern—that is, how much error in the interleg phasing can be tolerated
424 before destabilizing the body. By this measure, our model would predict that the animal uses
425 tripod coordination at all speeds, because the stability surfaces in Figure 5 are centered on
426 $(\phi_C, \phi_I) = (1/2, 1/2)$. The data suggest that the animal instead utilizes a compromise between
427 the most stable and most robust ICP at a given walking speed, showing variation in the ICP
428 but avoiding potentially unstable ICPs. In fact, the mean (ϕ_I, ϕ_C) of the animal data always
429 lies near the 80th percentile of stable ICPs (data not shown). This means that 20% of other
430 available ICPs would be more stable. However, the animal uses them less frequently,

431 presumably because they are closer to unstable phase relationships. In our comparison
432 between model and experimental data (Fig. 6), the predicted most stable ipsilateral phases
433 (red line) seem to constitute a lower bound for the experimental data; this observation is
434 compatible with the hypothesis that the motor output reflects the expected variability.

435 **Extensions of the model**

436 Although our model successfully captured the experimental data collected for this study,
437 there are different locomotion scenarios that could be used to test this model in the future.
438 These fall into two main categories: support polygon variant and gravity vector variant.
439 Support polygon variant scenarios include animals with amputated legs and curved walking.
440 In this study, we restricted analysis to intact animals, walking with a very low curvature.
441 However, removing legs drastically affects the support polygon and leads to noticeable
442 changes in ICP in both fruit flies (Wosnitza et al., 2013) and cockroaches (Delcomyn, 1971;
443 Hughes, 1957). In addition, the stance trajectories of fruit fly walking along a curved path are
444 markedly different than during straight walking (Szczecinski et al., 2018). This also changes
445 the associated support polygon and, as a consequence, stability.

446 Gravity vector variant scenarios include animals walking on inclined, vertical, or inverted
447 substrates. In such cases, the animal is not trying to prevent falling directly toward the
448 substrate as in level locomotion, but at some angle to it, along it, or away from it,
449 respectively. Maintaining stability in such cases would benefit from or require adhesive forces
450 between the animal's foot and the substrate. In fact, larger insects, such as stick insects,
451 appear to use such mechanisms to improve stability even when walking on flat substrates
452 (Gorb, 1998; Paskarheit et al., 2016). Studies of insect-inspired climbing robots have shown
453 that the stability of climbing can be analyzed in a very similar way to how we analyzed the
454 stability of walking here, but with the addition of a force tangential to the substrate, provided
455 by the "uphill" leg (Daltorio et al., 2009). In the future, we will expand our model and test its
456 ability to predict ICPs of climbing fruit flies.

457 **Possible mechanisms in the animal**

458 The goal of this work was not to explain how the animal generates different ICPs, but why.
459 However, it is worth considering which mechanisms may give rise to the phenomena
460 measured herein. Behavioral rules that describe interleg coordination in arthropods have
461 long been known (Cruse, 1990; Dallmann et al., 2017; Dürr et al., 2004). Several of these
462 behavioral rules explicitly address the temporal coordination between onsets of the swing
463 phases in adjacent legs (Rules 1-3, see Dürr et al., 2004). As a consequence, they ensure
464 that the probability of two adjacent legs executing their swing movements simultaneously is

465 low. Recent work with stick insects has shown that the onset of swing phase in a middle leg
466 correlates very tightly with the onset of stance phase in the ipsilateral hind leg (Dallmann et
467 al., 2017). The authors suggest that this is due to the middle leg measuring a decrease in the
468 load being supported, causing the leg to enter swing phase. Indeed, campaniform sensilla,
469 which sense cuticular strain induced by load changes, have been found to be sensitive to
470 unloading in the cockroach (Zill et al., 2009). Such a mechanism could be seen as an indirect
471 measurement of the animal's stability affecting their ICP. Whether this plays a role in *D.*
472 *melanogaster*, a particularly light animal, in which gravitational forces might not play a very
473 large role, remains to be investigated.

474 There is also evidence that walking in insects is more determined by centrally generated
475 motor output at high walking speeds while the influence of leg sensory structures is reduced
476 (Bender et al., 2011; Cruse et al., 2007). This is further supported by recent studies on *C.*
477 *morosus* (Mantziaris et al., 2017), *C. gregaria* (Knebel et al., 2017), and *D. melanogaster*
478 (Berendes et al., 2016). These studies have shown that neighboring legs have preferred
479 phases of oscillation, even when local sensory feedback is absent. This reduced sensory
480 influence at high walking speeds could, in turn, make the motor output less variable, thus
481 facilitating the convergence to the narrow range of stable ICPs. Ultimately, interleg
482 coordination likely arises through a combination of mechanisms that are mediated both
483 mechanically and neurally.

484 Assuming that static stability plays a role for the speed-dependent selection of an ICP, an
485 important question is whether stability, or some related proxy, is measured acutely and
486 continuously during walking or if the evolutionary pressure to remain upright that resulted in
487 interleg coordination rules that keep the body upright. Our experimental data from tethered
488 animals whose bodies were supported during walking did not noticeably vary from those from
489 freely walking individuals. In principle, these animals cannot fall over and acute
490 measurement of stability would result in different ICPs. These observations suggest that
491 walking flies do not measure stability as directly as mammals do, for example, utilizing
492 vestibular input (Buschmann et al., 2015).

493 The consequences of falling are less severe for a fruit fly than for larger animals (Hooper,
494 2012); if they do misstep and fail to support their body, their large damping to mass ratio
495 should slow down their fall more than for larger animals, such as humans. Nevertheless, fruit
496 flies still need to stay upright during walking. Falling impedes the animal's progress and
497 wastes energy and time, suggesting that it would benefit the animal to remain upright. This
498 might be even more critical during behaviors like courtship, during which males chase
499 females in close pursuit (Hall, 1994); falling over in this situation might reduce the chances of
500 mating. A similar line of argument can be made for escape from predators, in which precise

501 and smooth stepping is required (Parigi et al., 2014). Stability and the need to remain upright
502 have likely influenced the evolution of the observed ICPs in insects.

503 **List of symbols and abbreviations**

504 ϕ_I : Ipsilateral phase relationship

505 ϕ_C : Contralateral phase relationship

506 AEP: Anterior extreme position

507 BL: Body length

508 COM: Center of mass

509 ICP: Interleg coordination pattern

510 IR: Infrared

511 LED: Light-emitting diode

512 PEP: Posterior extreme position

513 PP: Polypropylene

514 w^{1118} : *D. melanogaster white* mutant strain

515 WT: *D. melanogaster* wildtype strains *Berlin* and *CantonS*

516

517 **Acknowledgements**

518 The authors would like to thank Michael Dübbert and Jan Sydow for excellent technical
519 support, Corinna Rosch for animal husbandry, and Sima Seyed-Nejadi, Sherylane Seeliger,
520 and Hans-Peter Bollhagen for lab support.

521 **Competing interests**

522 The authors declare no competing or financial interests.

523 **Author contributions**

524 N.S.S., T.B., and A.B. conceived the study, N.S.S., T.B., and A.S.C. carried out the
525 experiments, N.S.S. and T.B. created the model, N.S.S., T.B., and A.S.C. analyzed
526 experimental data, N.S.S., T.B., A.S.C., and A.B. wrote the manuscript. All authors read and
527 approved the final manuscript.

528 **Funding**

529 This study was supported by the German Research Foundation (DFG-Grant GRK 1960,
530 Research Training Group *Neural Circuit Analysis*, to A.B.), the Graduate School for Biological
531 Sciences at the University of Cologne (to A.S.C.), and the National Science Foundation
532 (Grant 1704436, to N.S.S.).

533

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534 **Figure legends**

535 **Figure 1:** Basic kinematic and temporal parameters of walking *D. melanogaster*. All
536 parameters are expressed as a function of walking speed. Points correspond to individual
537 steps. Left column corresponds to front legs (left and right, L1 and R1), middle column to
538 middle legs (L2 and R2), right column to hind legs (L3 and R3). (A) Step amplitude is only
539 weakly correlated with walking speed (regression line in red). (B) Swing duration is constant
540 over the observed speed range (regression line in red). (C and D) Both stance duration and
541 step frequency are strongly correlated with walking speed. Assuming that step amplitude and
542 swing duration are constant, stance duration and step frequency can be predicted with high
543 accuracy (green lines and corresponding coefficients of determination in C and D). This
544 figure was created with experimental data from Wosnitza et al. (2013).

545 **Figure 2:** Kinematic model and static stability. (A and B) The predicted relationships between
546 walking speed and stance duration (A) and stepping frequency (B; see Fig. 1C and D),
547 respectively, determines the frequency and stance duration for a particular speed. Thus, a
548 temporal sequence of swing and stance movements can be calculated for each leg. (C)
549 Average stance trajectories measured during experiments are combined with this temporal
550 sequence of swing and stance movements. A stance movement begins at the AEP (anterior-
551 most point in a stance trajectory), progresses with uniform speed (i.e., the set walking speed)
552 to the PEP (posterior-most point in a trajectory), is interrupted by the swing movement, and
553 then starts again from the AEP. Stance trajectories are described in body-centered
554 coordinates. Ellipses around AEPs and PEPs indicate one standard deviation of positional
555 variability measured in experiments (however, only average stance trajectories were used
556 here and variability was not taken into account). (D) Two values, ϕ_I and ϕ_C , describe the
557 phase relationships between ipsilateral legs and contralateral body sides, respectively. Since
558 all other parameters are either constant (stance trajectories) or uniquely defined by a
559 particular walking speed, these two values are the only free parameters in the model. (E)
560 Thus, for a given set of ϕ_I and ϕ_C and at a particular time within one complete step cycle, it
561 can be determined which legs are currently in stance and what their positions with regard to
562 the center of mass (blue dot) are. The legs currently in stance form a convex hull; the
563 minimal distance between the center of mass and the convex hull defines static stability
564 (green line). Its minimum value over one complete step cycle defines the stability for a
565 particular set of ϕ_I and ϕ_C .

566 **Figure 3:** Experimental setups. (A) Free-walking setup. Flies walked on top of a glass petri
567 dish covered with a watch glass (not shown for clarity). A concentric ring of IR LEDs provided
568 illumination (ring only shown partially). A high-speed camera captured a rectangular area of
569 the petri dish (dashed rectangle) via a surface mirror. As soon as the fly walked through the

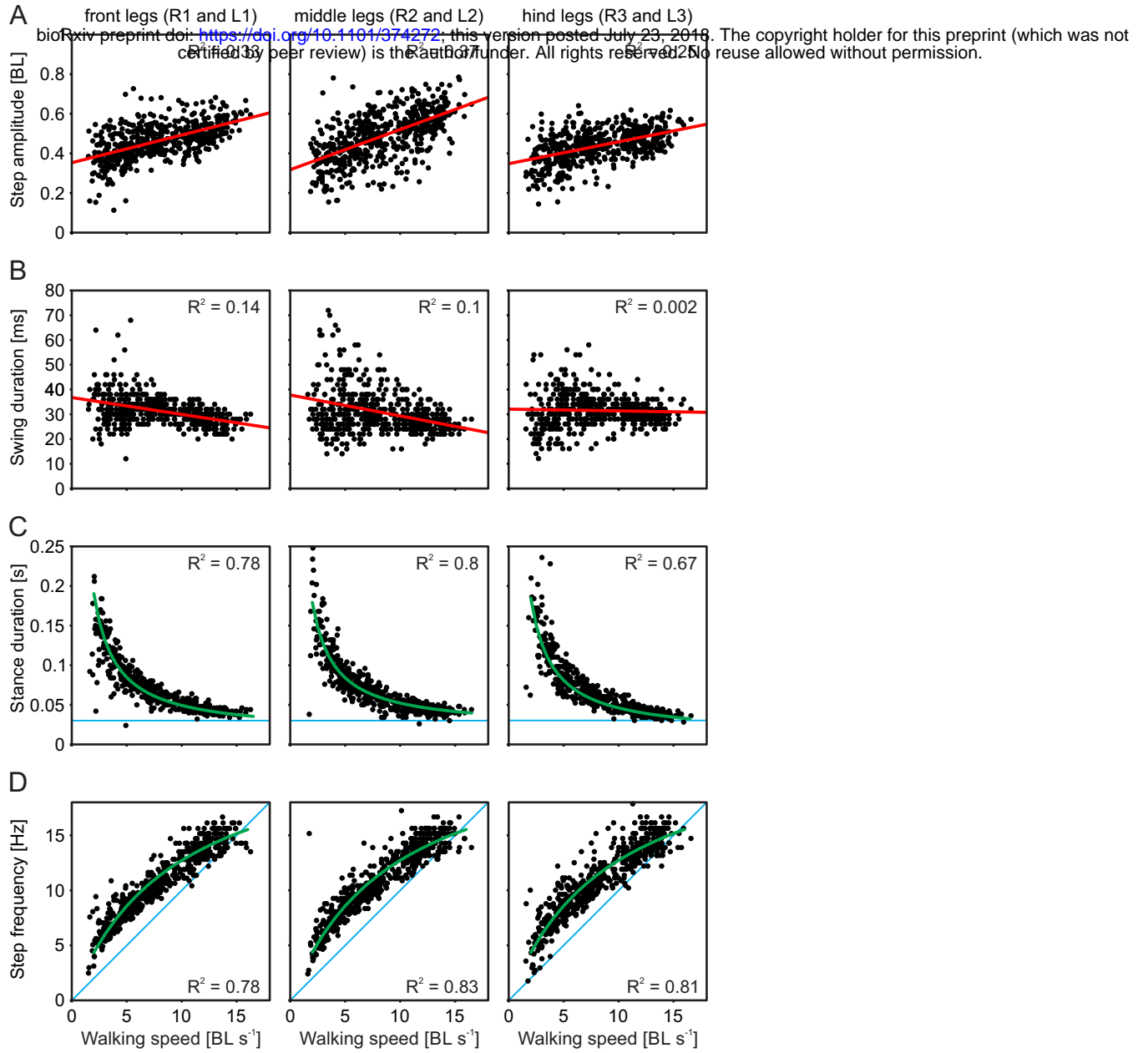
570 capture area, the recorded video was committed to storage for post-processing. (B) Example
571 cutout from a video frame captured in the free-walking setup. Leg tips are clearly visible and
572 have been manually annotated (for labels see Fig. 1). (C) Tethered-walking setup. Flies
573 walked on an air-supported PP ball whose rotational movements were captured by two
574 motion sensors. Illumination for the sensors was provided by IR lasers. The top of the ball
575 and the two mirrors were captured with a high-speed camera mounted above the setup;
576 illumination for the camera was provided by an LED ring around the camera lens. (D) Two
577 surface mirrors provided side views of the walking fly. Legs are clearly visible in these views;
578 leg tips have been annotated manually.

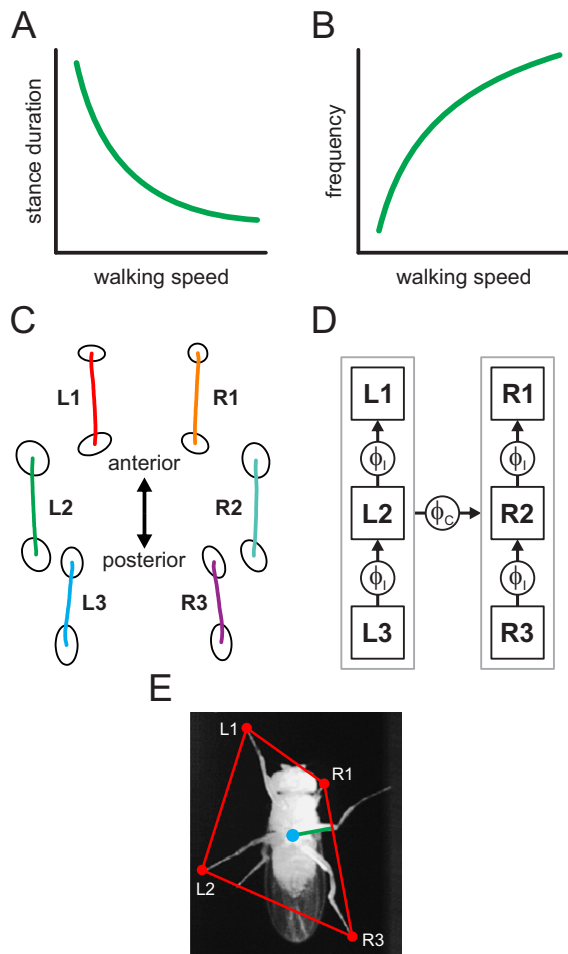
579 **Figure 4:** Representative inter-leg coordination patterns (ICPs). Based on the model
580 presented in Fig. 2, each combination of ϕ_I and ϕ_C is associated with a particular ICP. (A)
581 $\phi_V\phi$ plot with the position of four exemplary ICPs; each indicated point (B, C, D, E)
582 corresponds to an ICP in panels B to E. (B and C) Idealized tetrapod ICPs commonly
583 referred to in the literature. These correspond to $\phi_I = 1/3$ and $\phi_C = 1/3$ or $2/3$. (D) Tetrapod-
584 like ICP for which $\phi_I = 1/3$ and $\phi_C = 1/2$. This pattern can be found in walking fruit flies and is
585 also predicted as more stable than the ideal tetrapod ICP (see Results). (E) Tripod ICP
586 corresponding to $\phi_I = 1/2$ and $\phi_C = 1/2$. This ICP has frequently been reported in the
587 literature and occurs in fast-walking insects. Walking speed for all exemplary ICPs has been
588 set to 5 BL s^{-1} to facilitate comparison. Note that the tripod ICP in E will not occur naturally at
589 these speeds.

590 **Figure 5:** Model-derived stability plots and corresponding ICPs. (Ai to Hi) Each combination
591 of ϕ_I and ϕ_C is associated with a particular stability at a particular walking speed (here 3 to
592 10 BL s^{-1}). High stability is associated with yellow hues, low or zero stability is associated
593 with blue hues. The region of non-zero stability decreases with increasing walking speed and
594 contracts to an area around $\phi_I = 1/2$ and $\phi_C = 1/2$. In each $\phi_V\phi$ plot, the point of maximum
595 stability is indicated by a red dot. Note that at speeds of 4 BL s^{-1} and higher, the points of
596 maximum stability are very close to regions of zero stability (white arrows). (Aii to Hii) ICPs
597 that correspond to ϕ_I and ϕ_C of maximum stability in Ai to Hi. ICPs continuously change from
598 wave gait-like coordination at low speeds to almost tripod coordination at high speeds.
599 Increasing speed to even higher values will, in fact, result in tripod ICPs (not shown).

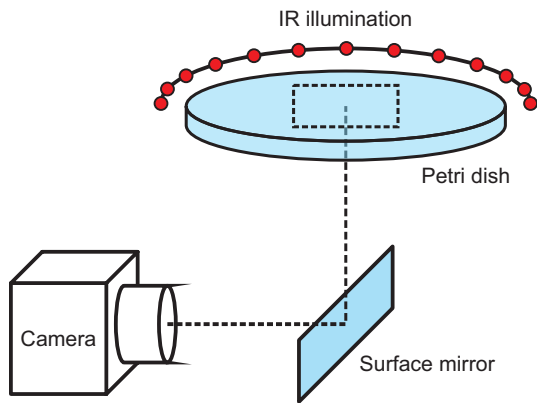
600 **Figure 6:** Phase relationships measured during experiments and predicted phases as a
601 function of walking speed. Here, we extended the data set from Wosnitza et al. (2013) with
602 data measured for the present study. Dots correspond to the phase relationship of individual
603 steps, and phase is measured between an observed leg and a reference leg (e.g., L3>L2
604 refers to the reference leg L3 and the observed leg L2). (A, B, F, G) Phase relationships
605 between ipsilateral middle and front legs (A and F) and hind legs and middle legs (B and G).

606 (C, D, E) Phase relationships between contralateral front legs (C), middle legs (D), and hind
607 legs (E). Green lines indicate running averages of experimentally measured phases;
608 magenta lines indicate model predictions for stability-optimal values of ϕ_l (A, B, F, G) and ϕ_c
609 (C, D, E).

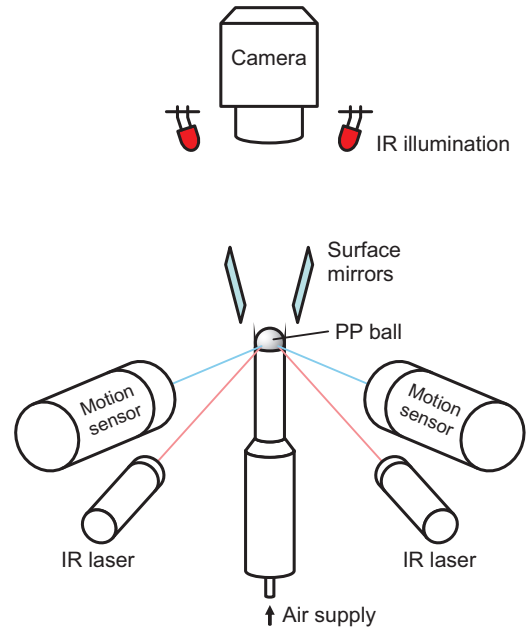




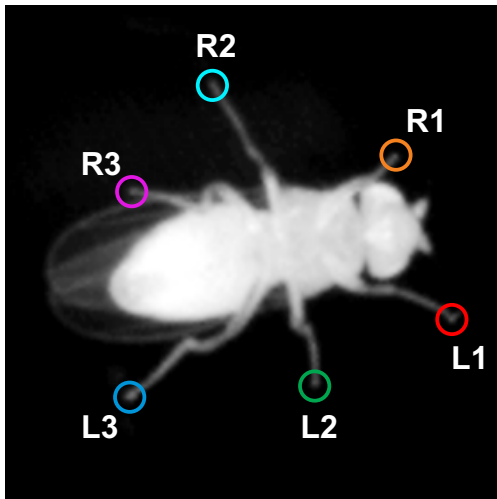
A



C



B



D

