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New de novo assembly of the Atlantic bottlenose dolphin (*Tursiops truncatus*) improves genome completeness and provides haplotype phasing.

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Abstract

High quality genomes are essential to resolve challenges in breeding, comparative biology, medicine and conservation planning. New library preparation techniques along with better assembly algorithms result in continued improvements in assemblies for non-model organisms moving them toward reference quality genomes. We report on the latest genome assembly of the Atlantic bottlenose dolphin leveraging Illumina sequencing data coupled with a combination of several library preparation techniques. These include Linked-Reads (Chromium, 10x Genomics), mate pairs, long insert paired ends and standard paired ends. Data were assembled with the commercial DeNovoMAGIC™ assembly software resulting in two assemblies, a traditional "haploid" assembly (Tur_tru_Illumina_hap_v1) that is a mosaic of the two haplotypes and a phased assembly (Tur_tru_Illumina_dip_v1) where each scaffold has sequence from a single homologous chromosome. We show that Tur_tru_Illumina_hap_v1 is more complete and accurate compared to the current best reference based on the amount and composition of sequence, the consistency of the mate pair alignments to the assembled scaffolds, and on the analysis of conserved single-copy mammalian orthologs. The phased de novo assembly Tur_tru_Illumina_dip_v1 is of the highest quality available for this species and provides the community with novel and accurate ways to explore the heterozygous nature of the dolphin genome.

Keywords: de novo genome assembly, bottlenose dolphin, Tursiops truncatus, 10x Genomics, DeNovoMAGIC™, Illumina

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Introduction

Technical advances in the past decade have reduced sequencing costs and improved access to sequencing data. Subsequent improvements in DNA extraction, preparation, and assembly algorithms facilitate low cost accurate de novo genome assemblies. Such assemblies are essential for constructing haplotype diversity databases for breeding, comparative biology, medicine and conservation planning. Even highly complex genomes now benefit from higher contiguity and improved protein coding coverage [1-4]. Consortium efforts to catalogue biodiversity of pivotal species of comparative evolutionary significance will continue to drive novel low-cost approaches toward reference quality assemblies with chromosome level resolution [5-7]. Here we use a combination of methods to drive improvements in assembly structure for the Atlantic bottlenose dolphin (*Tursiops truncatus*). This genome assembly, like that of the Hawaiian Monk seal and African wild dog, is being published with the goal to facilitate research on comparative genomics, provide structure for cataloging biodiversity and ultimately support decisions around species conservation and management [8, 9].

The bottlenose dolphin is one of the most widely studied marine mammals, however the taxonomy of the *Tursiops* genus remains unresolved. Numerous species designations have been suggested but not adopted due to a lack of resolution afforded by available data [10]. Even with new molecular genetic markers, we have reached a limitation on resolution from genetic data available to delineate species, subspecies and populations [11]. To usher this species into the era of genomics, a high-quality reference genome is essential. It provides structure to catalogue diversity within and between species at the whole genome level. In addition, the parallel molecular trajectory between dolphin and other mammalian species [12] makes the bottlenose dolphin a useful model to understand aspects of human health such as metabolic processes/diabetes [13-15], proteomics [16, 17] and aging [18].

A preliminary dolphin genome was first submitted to NCBI (TurTru1.0; GCA_000151865.1) using low coverage (2.82X) Sanger sequencing for the purpose of cross-species comparison [12, 19, 20]. Subsequent improvements were achieved through the addition of 30X Illumina short read data and 3.5X 454 data (Ttru_1.4; GCA_000151865.3). A much more complete genome was submitted in 2016 leveraging improvements in library preparation and assembly methods (Meraculous v. 2.2.2.5 and HiRise v. 1.3.0-116-gf50c3ce; Dovetail, Inc) with 114X coverage of Illumina HiSeq data prepared with proximity ligation Hi-C protocol (Tur_tru v1; GCA_001922835.1; [16]).

With the collection of data from multiple sources including Linked-Reads (Chromium, 10x Genomics; [21]), mate pairs (MP), long insert paired ends and standard paired ends, we provide an improved haploid reference quality dolphin genome assembly as well as the first haplotype phased diploid assembly. We refer to our unphased assembly as Tur_tru_Illumina_hap_v1 and to the phased assembly as Tur_tru_Illumina_dip_v1. Using Tur_tru v1 for comparison, our assembly shows increased contiguity and completeness with high consistency to the MP data and orthologous mammalian protein alignments. Additionally, by aligning Tur_tru_Illumina_hap_v1 to the Human reference genome, we illustrate the synteny of the dolphin scaffolds to human chromosome 1 [22, 23].

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Results

Coverage. We generated sequence data for a total coverage of approximately 450X, the majority from PCR Free and Chromium 10X Genomics Linked-Read libraries (Table 1). Coverage was computed using 2.4Gbp estimated genome size. Genome assembly was conducted using DeNovoMAGIC™ software (NRGene, Ness-Ziona, Israel). More detail about the library preparation and the assembly process are found in the Methods section.

Haploid and diploid assemblies. We report on two assemblies in this manuscript, one traditional haploid consensus assembly Tur_tru_Illumina_hap_v1 that represents a mosaic of the maternal and paternal haplotypes, and the other haplotype-phased (ie, diploid) assembly where each scaffold represents sequence corresponding to a single haplotype, Tur_tru_Illumina_dip_v1. The quantitative statistics for both assemblies are listed in Table 2. The phased or diploid genome assembly was made possible using Illumina sequencing data by leveraging the combination of library prep methods including Linked-Reads, is a significant advance and will provide the community with a powerful genomic tool for the downstream analysis in the context of the true heterozygous dolphin genome.

Genome assembly comparison. Both assemblies were compared to the best available assembly Tur_tru v1 (NCBI accession GCA_001922835.1; [16]). We did not use the Ttru_1.4 assembly (NCBI accession GCA_000151865.3) because the contiguity statistics of the Ttru_1.4 are vastly inferior to the Tur_tru v1 with a contig N50 3 times smaller than Tur_tru v1 and scaffold N50 over 200 times smaller.

The statistics for the Tur_tru_Illumina_hap_v1 assembly show bigger scaffolds but slightly smaller contigs with about 13% more sequence in the scaffolds compared to Tur_tru v1 (Table 2). More sequence does not necessarily make for a better assembly considering that the extra sequence may be duplicated haplotypes or contaminants that do not belong to the original organism. To characterize the extra sequence, we first aligned the Tur_tru_Illumina_hap_v1 to the Tur_tru v1 assembly using the Nucmer aligner which is part of MUMmer4 package [24]. We used default settings for generating the alignments. We then analyzed the alignments using the dnadiff package included with MUMmer4. 87.5% of Tur_tru_Illumina_hap_v1 sequence aligned to 97.9% of Tur_tru v1. This shows that 12.5% of Tur_tru_Illumina_hap_v1 had no alignments to Tur_tru v1, while only 2.1% of Tur_tru v1 had no alignments to Tur_tru_Illumina_hap_v1. Therefore, there are 301Mbp of extra novel sequence in our new assembly Tur_tru_Illumina_hap_v1. We then used the BUSCO tool to show that the extra sequence is meaningful (Table 3). Tur_tru_Illumina_hap_v1 had 160 missing BUSCOs, compared to 270 missing in Tur_tru v1. The number of duplicated BUSCOs was higher by only 34 in our assembly compared to Tur_tru_v1. This suggests that most of the extra sequence in Tur_tru_Illumina_hap_v1 is not contamination or redundant sequence, and likely contains useful coding information. We examined the locations of the 110 BUSCOs that are only found in the Tur_tru_Illumina_hap_v1 and 97 of them fully or partially aligned to locations in the sequences that were missing in Tur_tru v1 assembly. The haplotype-resolved assembly is more fragmented and it is missing 393 BUSCOs. As expected most of the complete BUSCOs that were found (3371) are duplicated (2079), since they are found in different haplotypes.

Assembly validation through MP consistency. Since both Tur_tru v1 and Tur_tru_Illumina_hap_v1 reference the same species, we expect few rearrangements between the assemblies. To examine this, we compared the absolute and relative correctness of the scaffolds of Tur_tru_Illumina_hap_v1 assembly by aligning the Illumina data from the 5-7Kbp MP library to the scaffolds of Tur_tru_Illumina_hap_v1 and Tur_tru v1 assemblies using Nucmer tool [24]. We chose this library because it contained the largest number of valid 5-7Kbp mate pairs. We then chose the best alignment

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for each read by running "delta-filter -q" on the alignment file and classified the alignments of the MPs into the following categories (Table 4):

1. **Same scaffold happy** – number of MPs where both mates aligned to the same scaffold in the correct orientation with mate separation within 3 standard deviations of the library mean
2. **Same scaffold short** -- number of MPs where both mates aligned to the same scaffold in the opposite orientation with mate separation of less than 1000bp; these MPs are not indicative of scaffolding misassemblies, they are simply a byproduct of the mate pair library preparation process as they are MPs that are missing the circularization junction site between the mates
3. **Same scaffold long** – number of MPs where both mates aligned to the same scaffold in the correct orientation, but the mate separation exceeded three standard deviations of the library mean
4. **Same scaffold misoriented** -- number of MPs where both mates aligned to the same scaffold in the opposite orientation with mate separation of more than 1000bp
5. **Mates aligned to different scaffolds** – number of MPs where the two mates aligned to different scaffolds
6. **Only one mate in the pair aligned** – number of MPs where only one read aligned to the assembly.

The comparison reveals that Tur_tru_Illumina_hap_v1 and Tur_tru v1 assemblies are similar in quality in terms of the MP alignments. The differences that stand out are the much larger (8 times more) number of mates that aligned to the same scaffold in the wrong orientation and the much larger number of the same scaffold long pairs in Tur_tru v1 compared to Tur_tru_Illumina_hap_v1 (Table 4). The difference in the number of same scaffold long pairs suggests that gap sizes are better estimated in the Tur_tru_Illumina_hap_v1 assembly. Both statistics point to a relatively higher number of mis-ordered or misoriented contigs in the scaffolds of Tur_tru v1 assembly. The likely cause of this difference is the scaffolding process used to create Tur_tru v1 assembly. The assembly was created with the HiRise assembler [25] using proximity ligation Hi-C data for scaffolding. The proximity ligation data provide MPs of all possible sizes, however, the MP distances and mate orientations are unknown. Since there are more shorter pairs than longer pairs due to the 3D structure of the DNA – it is much more likely to ligate parts of DNA that are closer to each other than the ones that are far apart. This property enables one to use these data for scaffolding. By mapping the pairs to the assembled scaffolds, one can measure how the distance between the mates in a pair varies with the number of pairs whose ends map to the same location in the assembly. However, the dependence is fairly weak on the short end, meaning that the number of pairs of about 10Kbp in length is not significantly different from the number of links of 12-13Kbp in length. This frequently results in mis-orientations and shuffling of scaffold positions for contigs that are smaller than 10-20Kbp in the scaffolding process. It is likely this type of misorientation and shuffling that we are observing in the Tur_tru v1 assembly and by this measure our Tur_tru_Illumina_hap_v1 assembly is much more consistent with the MP data, while maintaining good scaffold sizes.

Haplotype resolution. We tested the efficacy of the haplotype resolution in Tur_tru_Illumina_dip_v1 assembly by analyzing all pairs of large scaffolds (over 1Mbp) that aligned to the same location in the haploid assembly, implying that the two scaffolds in the pair represent two haplotypes. For example, we analyzed scaffold32860 (3,643,790bp) and scaffold2584 (5,424,824bp), that are the apparent haplotypes of the scaffold10 (12,182,427bp) in the Tur_tru_Illumina_hap_v1 assembly. We aligned all MPs from the 5-7Kb library to the Tur_tru_Illumina_dip_v1 assembly using Nucmer and then filtered the alignments allowing for a single best match for each read. We then examined the MPs that had best

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matches to scaffold32860 and scaffold2584. MPs whose mates had their best alignments not in the same scaffold could point to haplotype misassembly where the haplotype scaffolds "switched" haplotypes. We found no MPs where two mates had their best matches to the scaffolds corresponding to different haplotypes, suggesting that each of the two homologous scaffolds represents unique haplotypes and there is no "switching" of haplotypes present. We found 3,376 MPs where both mates had their best alignments to the same scaffold with correct distance and orientation, 9 in the same scaffold and wrong orientation, 3 stretched mates and 1,835 compressed (short) mates. The compressed mates do not indicate misassemblies, but rather they are the non-junction pairs that are the artefact of the circularization protocol used to create MPs: these pairs do not contain junction site between the mates. There are no apparent errors in any pairs of scaffolds that we examined leading to the conclusion that haplotype resolution was accurate.

Synteny between human and dolphin. Dolphin is a mammal, and currently the best mammalian reference genome is the human genome. To understand similarities between dolphin and human on the DNA level, we aligned the Tur_tru_Illumina_hap_v1 assembly to the primary chromosomes of the current haploid human reference genome GRCh38 [26]. Since human and dolphin are fairly distant species, we did not expect to find long DNA sequence alignments but instead we were looking for synteny where relatively short DNA fragments of scaffolds align in the same order and orientation between the two assemblies. We used MUMmer4 package for producing the alignments using the default settings. The alignment mummerplot (Figure 1) shows a striking synteny between the dolphin assembled scaffolds and human chromosome 1, visible even on the large-scale chromosome plot (Figure 1a). No large-scale synteny to the other human chromosomes can be readily observed. The synteny observation is possible due to large scaffold sizes in Tur_tru_Illumina_hap_v1. In Figure 1b, we show 22 scaffolds that have 50% or more of their sequence in syntenic alignments. The syntenic alignments of these 22 scaffolds span nearly the entire human chromosome 1 sequence. The synteny is not a new finding, it was first identified by Bielec et al (1998) [22] and was later extended to many other placental mammals [23]. The Tur_tru_Illumina_hap_v1 assembly clearly illustrates and confirms the expected synteny.

Methods

Sample Collection and DNA Extraction: The sample for this study came from a female Atlantic bottlenose dolphin (Sample ID 04329), captive born at SeaWorld of Orlando, Orlando, Florida from wild male and female Atlantic bottlenose dolphins. The animal was 36 years old at blood collection with a healthy medical history. Blood was collected using Qiagen- PAXgene™ Blood DNA Tubes (Qiagen). High molecular weight genomic DNA was isolated using the Illumina- MasterPure™ DNA Purification Kit and subsequently quantified and qualified using Quant-iT™ dsDNA Kit and E-Gel™ EX Agarose Gel (ThermoFisher).

Collecting Sequence Data

Paired End (PE) libraries. We generated the 450bp and 800bp PE libraries using the Illumina TruSeq® PCR-free DNA Sample Prep kit. The protocol was slightly modified at fragmentation and double-size selection steps by adjusting the Covaris DNA shearing protocols and by empirically titrating the ratios of SPRI magnetic beads over DNA to obtain insert sizes around 450bp and 800bp. We then evaluated the libraries for insert size and yield using Agilent Bioanalyzer and real-time qPCR assay, using Illumina DNA Standards and primer master mix qPCR kit (KAPA Biosystems, Roche), then normalized to 2nM prior to clustering and sequencing. Both the 450bp and

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800 bp libraries were then denatured and diluted to 8pM and 12pM respectively. The 800bp PE library was clustered and sequenced on the HiSeq 2000, using the Illumina HiSeq Cluster and SBS v4 kits for PE 2x160bp reads. The 450bp PE library was clustered and sequenced on the HiSeq 2500 v2 Rapid Run mode using the HiSeq Rapid Cluster and SBS v2 kits for PE 2x250bp reads.

Mate Pair (MP) libraries. To maximize sequence diversity and genome coverage, three separate MP libraries were constructed corresponding to 2-5Kb, 5-7Kb and 7-10Kb insert sizes using the Nextera® MP Library Preparation Kit according to the manufacturer's instructions (Illumina). All three libraries were generated from a single input of 4ug of genomic DNA size-selected on a 0.8% E-gel (Invitrogen). Proper sizing of gel-extracted products was confirmed using the Bioanalyzer High Sensitivity chip (Agilent) and 600ng was subsequently used as input for circularization. Following library preparation, the Bioanalyzer was used to confirm library quality. Each of the three libraries were quantified by qPCR (KAPA Biosystems Library Quantification Kit, Roche), denatured and diluted to 200pM after size-adjustment according to Bioanalyzer results, and clustered on the cBot (Illumina) according to the manufacturer's instructions. 2x150bp of Illumina paired-end sequencing was performed on the HiSeq 4000 using the HiSeq 3000/4000 Cluster and SBS kits.

10x Chromium library. Genomic DNA quality was assessed by pulsed-field gel electrophoresis to determine suitability for 10X Chromium library preparation (10X Genomics). 1.125ng of input was used for library preparation according to the manufacturer's instructions without size-selection. Final library concentration was determined by qPCR (KAPA Library Quantification Kit, Roche) and size-adjusted according to Bioanalyzer DNA 100 chip (Agilent) results. 2x150bp of Illumina paired-end sequencing with an 8-base index read was performed on the HiSeq 4000 using the HiSeq 3000/4000 Cluster and SBS kits.

Genome Assembly

Genome assembly was conducted using the DeNovoMAGIC™ software platform (NRGene, Nes Ziona, Israel). This is a proprietary DeBruijn-graph-based assembler that was used to produce assemblies of several challenging plant genomes such as corn [1] and ancestral wheat Aegilops tauschii [3]. The following outlines design of the assembler, and steps of the assembly process.

Reads pre-processing. In the pre-processing step we first removed PCR duplicate reads, and trimmed Illumina adaptor AGATCGGAAGAGC and Nextera® linker (for MP library) sequences. We then merged the PE 450bp 2×250bp overlapping reads with minimal required overlap of 10bp to create stitched reads (SRs) using the approach similar to the one implemented in the Flash software [27].

Error correction. We scanned through all merged reads to detect and filter out reads with apparent sequencing errors by examining k-mers (k=24) in the reads and looking for low abundance k-mers. We have high coverage data (~450x), with each read yielding 127 (150-24+1) to 227 (250-24+1) K-mers. Thus average 24-mer coverage is at least 300x. 24-mers that only appear less than 10 times in the set of reads likely contain errors. We did not use the reads that contain these low abundance k-mers for building initial contigs.

Contig assembly. The first step of the assembly consists of building a De Bruijn graph (kmer=127 bp) of contigs from all filtered reads. Next, paired end and MP reads are used to find reliable paths in the graph between contigs for repeat resolving and contigs extension. 10x barcoded reads were mapped to contigs to ensure that adjacent contigs were connected only when there is evidence that those contigs originate from a single stretch of genomic sequence (reads from the same two or more barcodes were mapped to the same contigs).

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Split phased/un-phased assembly processes. Two parallel assemblies take place to complete the phased and un-phased assembly result. The phased assembly process utilizes the complete set of contigs. In the un-phased assembly process, the homologous contigs are identified and one of the homologs is filtered out, leaving a subset of the homozygous and one of the homologous contigs in heterozygous regions. The linking information of both homologous contigs is kept through the assembly process of the un-phased assembly, usually enabling longer un-phased scaffolds.

Scaffolding. All the following steps are done in parallel for both the phased and un-phased assemblies. Contigs were linked into scaffolds with PE and MP information, estimating gaps between the contigs according to the distance of PE and MP links. In addition, for the phased assembly, 10x data were used to validate and support correct phasing during scaffolding.

Gap filling. A final gap fill step used PE and MP links and De Bruijn graph information to locally construct a unique path through the graph connecting the gap edges. The path was used to close the gap if it was unique and its length was consistent with the gap size estimate.

Scaffold split/merge. We used 10x barcoded reads to refine and merge scaffolds. All barcoded 10x reads were mapped to the assembled scaffolds. Clusters of reads with the same barcode mapped to adjacent contigs in the scaffolds were identified to be part of a single long molecule. Next, each scaffold was scanned with a 20kb length window to ensure that the number of distinct clusters that cover the entire window (indicating a support for this 20kb connection by several long molecules) is statistically significant with respect to the number of clusters that span the left and the right edge of the window. If there was a statistically significant disagreement in the coverage by the clusters over the window, we broke the scaffold at the two edges of the window. Finally, the barcodes that were mapped to the scaffold edges (first and last 20kb sequences) were compared to generate a graph of scaffolds. The scaffolds are nodes and the edges are links connecting nodes with more than two common barcodes on the ends. We broke the links to the nodes that had more than two links and output the resulting linear paths in the scaffold graph as final scaffolds.

Summary

We show that Tur_tru_Illumina_hap_v1 is more complete and accurate compared to the current best reference Tur_tru v1, based on the amount and composition of sequence, the consistency of the MP alignments to the assembled scaffolds, and on the analysis of conserved single-copy mammalian orthologs. The additional 12.5% of sequence data identified and assembled here was found to contain 97 additional BUSCO alignments as compared to the latest published assembly Tur_tru v1. The large scaffolds represented by Tur_tru_Illumina_hap_v1 enabled and confirmed expected synteny to human chromsome 1. The phased de novo assembly Tur_tru_Illumina_dip_v1 is of the highest quality available and provides the community with novel and accurate ways to explore the heterozygous nature of the dolphin genome. These findings illustrate the impact of improved sample preparation and improved de novo assembly methods on progress toward more complete and accurate reference quality genomes. Better quality assemblies will improve our understanding of gene structure, function and evolution in mammalian species.

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Table 1. Summary of the sequencing data collected to create Tur_tru_Illumina_hap_v1 and Tur_tru_Illumina_dip_v1.

Library type	Read Length	Insert Size	Genomic Coverage
PCR-free	2x250bp	450bp	101x
PCR-free	2x160bp	800bp	123x
Mate-Pair	2x150bp	2-4Kbp (peak 4.2Kbp)	37x
Mate-Pair	2x150bp	5-7Kbp (peak 6.0Kbp)	61x
Mate-Pair	2x150bp	8-10Kbp (peak 9.9Kbp)	58x
10X Chromium	2x150bp	-	70x

Table 2. Comparison of quantitative statistics for different assemblies of the bottlenose dolphin. We used genome size of 2,407,574,691bp equal to the total size of the scaffolds of the bigger haploid assembly, for comparison of the N50 contig and scaffold sizes between the two assemblies. The Tur_tru_Illumina_hap_v1 and Tur_tru v1 assemblies have comparable scaffold N50 sizes, and Tur_tru v1 has bigger contigs. The Tur_tru_Illumina_hap_v1 assembly has more sequence and our BUSCO analysis (Table 3) shows that it is likely more complete. The N50 comparisons to the haplotype-resolved Tur_tru_Illumina_dip_v1 assembly are shown for completeness, computed with 2x genome size (4,815,149,382bp).

	Tur_tru_v1	Tur_tru_Illumina_hap_v1	Tur_tru_Illumina_dip_v1
Total sequence	2,120,283,832	2,386,142,562	5,475,240,330
# of scaffolds	2647	485	644,368
Longest scaffold	96,299,184	83,924,496	10,429,594
Scaffold N50	23,564,561	26,997,441	774,534
Scaffold L50	26	30	1,529
# of contigs	116,650	139,921	902,141
Longest contig	403,070	320,783	298,006
Contig N50	37,749	30,679	27,325
Contig L50	17,321	23,581	52,793
GC content	40.85	41.26	41.95

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Table 3. Comparison of BUSCO 3.0.2 Mammalia single copy ortholologs among the three Dolphin assemblies. The table shows that the Tur_tru_Illumina_hap_v1 assembly is more complete, with 110 fewer missing single-copy orthologs compared to the Tru_tru v1 assembly. The Tur_tru_Illumina_hap_v1 assembly has 43 extra duplicated orthologs, which possibly points to incomplete filtering of redundant haplotypes. While Tur_tru v1 assembly has bigger contigs, the Tur_tru_Illumina_hap_v1 assembly has many fewer fragmented BUSCOs. The haplotype-resolved Tur_tru_Illumina_dip_v1 assembly is less contiguous and less complete. As expected, more than half of the complete BUSCOs are duplicated, corresponding to the two resolved haplotypes.

BUSCOs	**Tur_tru_v1 (NIST)**	**Tur_tru_Illumina_hap_v1**	**Tur_tru_Illumina_dip_v1**
Complete	3,647	3,837	3,371
Complete single-copy	3,614	3,760	1,292
Complete duplicated	33	77	2,079
Fragmented	187	107	340
Missing	270	160	393
Total	4,104	4,104	4,104

Table 4. Comparison of the alignments of 5kb MPs from 5-7Kbp library to Tur_tru_Illumina_hap_v1 and Tur_tru v1 assemblies. Same scaffold means that both mates mapped to the same scaffold; happy mates aligned in the correct orientation with mate distance within 3 standard deviations from the mean; misoriented mates aligned in wrong orientation; long mates aligned with the distance between the mates exceeding 3 standard deviations; short mates aligned with the distance of less than 1000bp.

	Tur_tru_Illumina_hap_v1	**Tur_tru_v1**
Same scaffold happy	133,958,734	136,869,395
Same scaffold misoriented	485,207	3,294,244
Same scaffold long	184,893	871,303
Same scaffold short	82,085,819	139,116,178
Mates aligned to different scaffolds	11,160,889	25,545,033
Only one mate in the pair aligned	108,536,854	192,649,250

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Availability of data

The dolphin assembly Tur_tru_Illumina_hap_v1 has been deposited at NCBI under BioProject PRJNA476133. The dolphin assembly Tur_tru_Illumina_dip_v1 has been deposited at NCBI under BioProject PRJNA478376. Both assemblies are also available on the public FTP site ftp://ftp.ccb.jhu.edu/pub/alekseyz/Tur_tru_Illumina_v1/.

Competing interest

KV and CTL were both full-time employees of Illumina at the time this work was completed. Illumina is the company responsible for low cost high accuracy DNA sequencing. GBZ, KB, and TB are employees of NRGene, a company providing software analysis tools for de novo assembly.

Author contributions

KVM, CTL and MMV designed the project. KVM, CTL and AZ wrote the manuscript. GBZ, TB and KB generated genome assemblies. AZ conducted validation, MP consistency analysis, human chromosome 1 and BUSCO analyses; JSL provided the blood sample; JL, AN, MR, MG, EJ, and BS processed samples, generated sequencing and completed quality checks on sequence data; and all authors contributed to writing and editing the manuscript.

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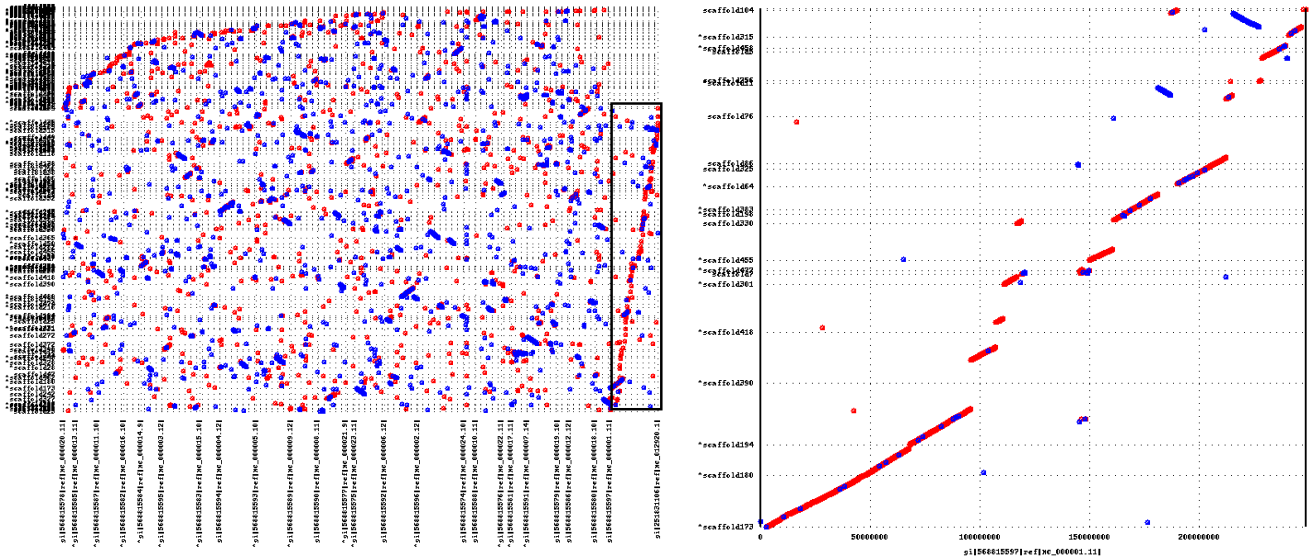
Figure1. This figure shows the alignment of the Tur_tru_Illumina_hap_v1 assembly to the human GRCH38 reference (primary chromosomes only). Each dot represents an alignment with red indicating forward direction and blue indicating reverse direction. Human reference coordinates are on the X axis and Tur_tru_Illumina_hap_v1 assembly alignment coordinates are on the Y axis. Panel (a) shows alignment of the entire assembly to the human reference with alignments to human chromosome 1 highlighted by the black box. One can clearly see the synteny that is present between the dolphin scaffolds and human chromosome 1. No other human chromosome shows clear synteny. Dolphin scaffolds with syntenic alignments spanning over 50% of the scaffold were extracted. Alignments only to human chromosome 1 are presented in panel (b).



(a) (b)

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