

1 **Discrete or indiscrete? Redefining the colour polymorphism of the land snail**

2 ***Cepaea nemoralis***

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7 Running head: Redefining a snail colour polymorphism

8 Biologists have long tried to describe and name the different phenotypes that make
9 up the exuberant colour polymorphism of the land snail *Cepaea nemoralis*.
10 Traditionally, the view is that the ground colour is one of a few major colour classes,
11 either yellow, pink or brown, but in practise it is frequently difficult to distinguish the
12 colours, and consistently define different shades of the same colour. To understand
13 whether colour variation is continuous, and to investigate how the variation may be
14 perceived by an avian predator, we applied psychophysical models of colour vision
15 to shell reflectance measures. The main finding is that both achromatic and
16 chromatic variation are indiscrete, being continuously distributed over many
17 perceptual units, with the major axis of chromatic variation representing differences
18 in saturation, or purity of colour. Nonetheless, clustering analysis based on the
19 density of the distribution revealed three groups, roughly corresponding to human-
20 perceived yellow, pink and brown shells. There is also large-scale geographic
21 variation between these morphs across Europe, and some covariance between shell
22 colour and banding patterns. Although further studies are necessary to understand
23 the evolutionary origins and impact of natural selective upon this variation, the
24 observation of continuous variation in colour is intriguing, given that the underlying
25 supergene that determines colour should prevent phenotypes from “dissolving” into
26 continuous trait distributions.

27 Keywords: *Cepaea*, colour, polymorphism, snail, supergene

28 Throughout the past century, the study of animal colour has been critical in making
29 progress in understanding the principles of biology, especially with respect to
30 genetics and evolution (McKinnon and Pierotti 2010; McLean and Stuart-Fox 2014;
31 Cuthill et al. 2017; San-Jose and Roulin 2017). For instance, early studies on the
32 inheritance of colour traits were important in establishing an understanding of basic
33 Mendelian genetics (Wheldale 1907; Staples-Browne 1908). Subsequently, studies
34 of the distribution and predation of colour morphs have and continue to shape our
35 understanding of how natural and sexual selection operate in wild populations
36 (Hugall and Stuart-Fox 2012; Dale et al. 2015; Delhey et al. 2017). Most recently,
37 candidate gene and latterly genomic approaches have been used to identify the
38 underlying genes that determine the colour differences (references in Hoekstra 2006;
39 McLean and Stuart-Fox 2014; San-Jose and Roulin 2017).

40 For practical reasons, many of these prior studies have taken advantage of
41 traits that exhibit relatively simple, discrete variation and straightforward inheritance
42 patterns, but this risks missing the extraordinary variation of life forms and colour
43 traits. It is also likely that in nature discrete variation is the exception rather than the
44 rule – and this is becoming more evident as researchers increasingly use
45 instrumentation to measure colour (Montgomerie 2006), rather than being obliged to
46 bin types into human-defined categories.

47 Historically, two of the most important animals in studying colour
48 polymorphism have been the peppered moth *Biston betularia* and the grove snail
49 *Cepaea nemoralis* and its sister taxon, *C. hortensis* (collectively “*Cepaea*”, the
50 preferred common name), because individuals are relatively easy to collect and
51 study, and the colour morphs show straightforward inheritance. However, while
52 ongoing and long-term studies on these animals continue to provide compelling

53 evidence for the fundamental role of natural selection in promoting and maintaining
54 variation in natural populations, as well as the impact of modern-day habitat change
55 (Silvertown et al. 2011; Cook et al. 2012), progress in understanding the precise
56 mechanism of the polymorphisms has diverged in the two systems.

57 In the peppered moth, the precise mutation that determines the colour
58 differences was reported to be in a known patterning gene (van't Hof et al. 2016). In
59 contrast, an understanding of the 'exuberant' (Franks and Oxford 2009)
60 polymorphism of *Cepaea* – whether yellow, pink or brown shells and zero to five
61 bands – has stalled since the 1970s. In part, this may be a reaction to the question of
62 Jones *et al.* (1977) on whether the *Cepaea* polymorphism is “a problem with too
63 many solutions?” Actually, the intention of that work was to emphasise the perfect
64 case study provided by *Cepaea*; as simple explanations for phenotypic variation are
65 the exception, Jones *et al.* were making the point that it is important to study
66 organisms for which polymorphism may be explained by a variety of processes,
67 precisely because they are more realistic. Some 40 years later, this comment is still
68 relevant, given that genomic technologies and DNA sequence analyses should allow
69 us to uncover the relative contributions of each of these processes to contemporary
70 diversity – but it is nonetheless understandable that most study systems are still
71 selected because of their relative simplicity.

72 Despite a general lack of progress, the *Cepaea* polymorphism retains
73 excellent potential as a model system in evolutionary biology. Previous studies have
74 laid the foundations for future progress, including exploiting many long term studies
75 (Cook et al. 1999; Davison and Clarke 2000; Silvertown et al. 2011; Cameron and
76 Cook 2012; Özgo and Schilthuizen 2012; Cameron et al. 2013; Schilthuizen 2013;
77 Cook 2014; Ozgo et al. 2017), increasing understanding of the pigments and shell

78 proteome (Mann and Jackson 2014; Williams 2017) and especially, using new
79 genomic methods to identify the genes involved (Richards et al. 2013; Kerkvliet et al.
80 2017). Of particular interest here, we note (as do others, Surmacki et al. 2013) that
81 there is now a pressing need to quantify objectively the polymorphism of *Cepaea*
82 shells, and to understand how this is perceived by an avian predator, because only
83 then can we properly understand how the polymorphism is maintained. The specific
84 problem is that in the past *Cepaea* shell colours have usually been treated as one of
85 three or more distinct classes (e.g. Cain and Sheppard 1954) – yellow, pink or brown
86 – partly due to a lack of objective measures of colour, especially those that can be
87 used in the field and between different observers and contexts (Cain et al. 1960;
88 Cain et al. 1968; Jones et al. 1977). There has been also the significant issue that
89 human perception of colour is not necessarily objective or the same as that of an
90 avian predator (Surmacki et al. 2013).

91 Now that objective methods of measuring and analysing colour are widely
92 available (Endler 1990; Montgomerie 2006; Maia et al. 2013; Delhey et al. 2015) and
93 easy to use, at least in a controlled laboratory setting, we set out to measure
94 quantitatively the ground colour of *Cepaea* shells, and so to define the nature of the
95 polymorphism. Specifically, by measuring the shell colour of snails collected across
96 the breadth of the European distribution, we used psychophysical models of colour
97 vision to assess how chromatic variation is perceived by birds (but not categorised;
98 Caves et al. 2018).

99 Previously, Surmacki *et al.* (2013) used quantitative measures of colour on
100 relatively few individuals to assess how shells match to various backgrounds. Here,
101 we investigate the extent to which the distribution of snail shell colour is continuous
102 along the main axes of chromatic variation, using more than a 1000 individuals and

103 Gaussian finite mixture modelling (Scrucca et al. 2016) to test whether colours fall
104 into clusters in multivariate space. We also aimed to understand if quantitative
105 measures on a relatively small sample of shells can describe – rather than explain –
106 geographic patterns in colour morph frequency across Europe, as others have done
107 in much larger qualitative surveys.

108 The findings have significance for understanding the *Cepaea* polymorphism,
109 and the nature of the selection that acts upon it, as well as more generally
110 highlighting the need to measure colour objectively in other systems, before being
111 able to test for possible explanations.

112 *Methods*

113 Data collection

114 Individual *C. nemoralis* snails were mainly gathered opportunistically by volunteer-
115 led collection and field trips across Europe (Grindon and Davison 2013). Snails were
116 frozen upon arrival at the University of Nottingham, subsequently thawed and the
117 body extracted from the shell. The ground colour and banding of the shell was then
118 scored qualitatively by an experienced person (A.D.) and a student, as either yellow
119 (Y), pink (P) or brown (B), and unbanded (O), mid-banded (M), or all other banding
120 patterns (B, usually five-banded). Subsequent statistical analyses were carried out at
121 the level of the individual and the level of the population (sample site). So that we
122 could compare broad-scale patterns across Europe, larger groups were also used –
123 individual sample sites were therefore grouped into one of six larger groups (Table
124 S1; Figure 1).

125 An Ocean Optics spectrometer (model USB2000+UV-VIS-ES) and a light
126 source (DT-MINI-2-GS UV-VIS-NIR) were used to measure individual reflectance

127 spectra of shells, using a WS-1 diffuse white reflectance standard to set the baseline
128 light spectrum (Teasdale et al. 2013; Taylor et al. 2016), and complete darkness to
129 set the dark spectrum standard. Reflectance measurements were taken on the
130 underside of the dried shell, because it was usually the least damaged region, least
131 exposed to sunlight, and well away from any bands. Point samples were taken for
132 each shell at a 45° incident angle, 2 mm from the shell. Individual shells were
133 measured three times, non-consecutively, with the software recalibrated against light
134 standards every 2-5 measurements. Readings were collected using Ocean Optics
135 SpectraSuite v. 2.0.162 (software settings: integration time 750msec, boscar width 5,
136 scans to average 10); then the raw data smoothed and binned into 5 nm categories
137 using Pavo version 0.5-6 (Maia et al. 2013).

138 Analysing chromatic and achromatic variation

139 We used the framework provided by Delhey et al. (2015) to analyse the reflectance
140 spectra. In this framework, a psychophysical model of colour vision (Vorobyev and
141 Osorio 1998; Vorobyev et al. 1998) is used to assess whether chromatic differences
142 between reflectance spectra exceed a discrimination threshold, or 'just noticeable
143 difference' (JND), which can be perceived by a receiver, such as an avian predator.
144 The key to these models lies with the degree to which a particular combination of
145 reflectance and illuminant spectra stimulate each of the different photoreceptors in
146 the retina. In birds, these photoreceptors are the four single cones used for colour
147 vision, which are sensitive to long (L), medium (M), short (S), and very short (VS)
148 wavelengths of light (Cuthill 2006).

149 To analyse chromatic variation, the quantum catches for each cone type were
150 converted into three chromatic coordinates (x, y and z), where Euclidean distances
151 between points reflect perceptual differences, using the formulas of Cassey *et al.*

152 (2008). As there are no data for the song thrush, *Turdus philomelos*, which is the
153 main predator of *Cepaea*, we used sensitivity functions for the closest available
154 relative, the blackbird *Turdus merula* (Hart et al. 2000; Hart 2001), namely cone
155 proportions of VS: 0.528, S: 0.904, M: 1.128, and L: 1, sensitivity functions of 373,
156 461, 543 and 603, respectively. The analysis assumed that the L cone has a noise-
157 to-signal ratio of 0.05, so that the ratios for the other cones were VS: 0.0688, S:
158 0.0526 and M: 0.0471. The irradiance spectrum of 'standard daylight' (d65) was
159 used for the main analyses. However, analyses were also run for 'woodland shade'
160 to understand the influence of illuminant on avian perception of colour (Vorobyev et
161 al. 1998).

162 To identify the main axes of chromatic variation, we carried out a Principal
163 Components Analysis (PCA) on the chromatic coordinates (x, y and z), preserving
164 the perceptual distances (JNDs) by using a covariance matrix rather than a
165 correlation matrix (Delhey et al. 2015). To understand whether there are potential
166 clusters within the chromatic coordinate data, Gaussian mixture modelling was
167 carried out using Mclust 5.3 in R version 3.3.3 (Scrucca et al. 2016). A number of
168 models were compared, each of which assumed a different number of clusters (from
169 1 to 10), normally distributed in multivariate chromatic space. Several classes of
170 model were considered, each with a different assumption about the homogeneity of
171 variance and orientation among clusters. The best fitting model was then determined
172 as the one with highest Bayesian Information Criteria (BIC), with significant
173 differences determined using a bootstrap approach.

174 The methods of Delhey et al. (2015) were also used to assess achromatic
175 variation. In birds, sensitivity to achromatic cues is supposed to be mediated by
176 double cones which have the same pigment as L cones in birds but different oil

177 droplets, so have a wider sensitivity range. Values of achromatic contrast were
178 therefore estimated, again in JNDs, by computing achromatic contrast between each
179 reflectance spectrum and a reference (a very low value of double cone quantum
180 catch, 0.001), corresponding to a dark spectrum, and using the same noise-to-signal
181 ratio.

182 Analysis of morph frequencies

183 We investigated evidence for effects of location and banding on the likelihood that a
184 snail belonged to a particular morph, using generalised linear mixed effects models
185 (GLMMs) with binomial errors. Each morph was considered separately, with each
186 snail to be scored as belonging to the focal morph (1) or not (0). The three analyses
187 are not independent, since each snail can only belong to one morph. Banding
188 pattern was fitted as a fixed factor, whilst the effect of geographic location was
189 examined at three spatial scales. Variation in morph frequency at a local level was
190 modelled with random effect for site. Variation at a regional level was considered by
191 fitting a fixed effect of geographic region. Finally, continental scale variation was
192 modelled by looking for fixed linear and quadratic effects of latitude and longitude.
193 The fact that region and latitude/longitude are partially collinear was reflected in the
194 model-fitting procedure. We first fitted a saturated model with all main effects, except
195 for region, and their two-way interactions (excluding interactions involving quadratic
196 effects). Then, fixed terms were removed in a stepwise fashion, testing the effect of
197 deletion using likelihood ratio tests, until only significant terms remained. Effects of
198 latitude/longitude were then substituted with an effect of region and we compared the
199 Akaike Information Criterion (AIC) of the resulting models, to test if region was better
200 at capturing any large-scale geographic variation. Testing random effects in
201 generalised linear mixed models is problematic, so we compared the AIC of the

202 saturated GLMM with that of a generalised linear model without the random term for
203 site to provide an approximate test of the importance of site.

204 *Results*

205 Variation in colour

206 We measured the individual reflectance spectra of 1172 shells, mainly collected from
207 across Europe (Table S1; Figure 1) and then transformed them into visual space
208 coordinates, xyz. To visualise this chromatic variation, and the relationship with
209 human-scored and Mclust-defined colour categories (below), the xyz coordinates
210 were plotted in visual colour space (Figure 2). There were no obviously discrete
211 groups.

212 A PCA on the xyz coordinates showed a first axis which explained 87% of
213 chromatic variation. PC1 had a moderate positive loading for x (0.61), and a
214 moderate negative loading for y (-0.64) and z (-0.46). Two further axes explained
215 11% and 2% of the variation, the second having a positive loading on all axes (0.75,
216 0.28, 0.61, respectively), and the third a mixture (-0.26, -0.71, 0.65). The range of
217 observed variation on each axis was considerable: 41, 22 and 8 JNDs for x, y and z
218 respectively (Figure 2). Plotting the average normalized reflectance spectra for each
219 quartile of each principal component showed how the three PC axes correspond to
220 chromatic variation (Figure 3). Variation along PC1 represents relatively high
221 stimulation of L cones and lesser stimulation of S cones, relative to M cones.
222 Variation in PC2 showed relatively higher stimulation of L cones and lesser
223 stimulation of M cones. PC3 showed relatively high stimulation of the M cones
224 compared to lesser stimulation of the S and L cones. Only PC1 showed any
225 differences in the VS region but the shells barely reflected in the UV.

226 To investigate whether snail shells cluster in chromatic space, and whether
227 observed clusters correspond to human-scored qualitative colour morphs, Gaussian
228 finite mixture modelling was applied to the xyz visual space coordinates. The best
229 model (VVV, ellipsoidal, varying volume, shape, and orientation; BIC -15727.3; $P <$
230 0.001 compared 2nd best model) recovered three clusters, roughly corresponding to
231 human-scored yellow (46%, n=539), pink (44%, n=511) and brown (10%, n=122)
232 (Table 1). The next best fitting model also recovered three clusters (VEV; BIC -
233 15749.3; $P <$ 0.001 compared with 3rd best model) and the third recovered four
234 clusters (EEV; BIC -15755.5; the 4th cluster contained only 16 individuals).

235 Comparing human-scored (A.D.) and Mclust-defined groups, the overall
236 concordance was good at 76% (Table 1), with a similar error rate (75%) for the
237 student group. The highest proportion of discordant scores were human-scored
238 yellow shells that Mclust classed as pink (10% for A.D.; 12% for student group), with
239 the other major discrepancies being human-pink classed as Mclust-brown (8%), and
240 human-yellow classed as Mclust-pink (4%). The main difference was that human-
241 scoring reported relatively few brown shells (n=37), whereas the same group in
242 Mclust was larger (n=122). With misclassifications adjusted relative to the total
243 number of each Mclust shell type, 81% of the brown group were in a different
244 human-scored group (74% pink, 7% yellow), compared to 26% of the pinks (3%
245 brown 23% yellow) and just 8% of yellows (8% pink, 0% brown). Thus, while the
246 overall correspondence between human and Mclust scoring of shell colour was
247 good, the yellows were scored accurately (92%), pinks less so (74%) and brown
248 poorly (19%).

249 Plots of human-scored colours along the three PCs (Supplementary Movie 1)
250 and Mclust-categories were concordant with the above analyses (Figures 2, 4;

251 Supplementary Movie 2). Broadly, PC1 did not separate different human-perceived
252 colours or categories of shell, but instead mainly represents differences in saturation,
253 or purity of colour, between individuals (Figure 4). PC2 separated brown from yellow
254 and pink, and PC3 broadly separated all three colours, yellow, pink and brown.

255 The above analyses were repeated using woodland shade rather than
256 standard daylight. The main difference was that while Mclust again recovered three
257 groups, brown shells were more common (14%, n=168), with fewer pinks (40%,
258 n=474) and approximately the same number of yellows (45%, n=530).

259 Finally, achromatic variation was also considerable, varying over more than
260 100 JNDs, and without any obvious differences between Mclust-defined colour
261 morphs (Supplementary Figure 1).

262 Geographic variation between morphs

263 Large-scale geographic variables (latitude, longitude and region) had significant
264 effects on the probability that a snail was pink or yellow, but not the probability that a
265 snail was brown (Table 2). Pink morphs were significantly less common at mid-
266 latitudes (Figure 5). Snails with more than one band (B) and those which were mid-
267 banded (M) were more likely to be pink in the west, while unbanded (O) snails were
268 more likely to be pink in the east (Figure 5). In contrast, yellow snails were less
269 common at high latitudes, and were affected by an interaction between longitude and
270 banding which was the reciprocal of that seen in pink snails (Figure 6). Morph
271 frequencies also varied at a local level: a saturated mixed model including the
272 random effect of site was much better (Brown AIC = 443.7; Pink AIC = 868.9; Yellow
273 AIC = 852.8) than an equivalent model without the random effect (Brown AIC =
274 646.4; Pink AIC = 1407.9; Yellow AIC = 1416.2). Banding was associated with colour

275 morph in various ways. In addition to the interaction between banding and longitude
276 in pink and yellow snails mentioned above, unbanded snails (O) were generally more
277 likely to be brown (14% of all unbanded snails), than snails that were mid-banded
278 (M; 11.6%) or had several bands (B; 5.8%).

279 *Discussion*

280 By measuring the ground colour of *Cepaea nemoralis* shells collected across the
281 breadth of the European distribution, we used psychophysical models of avian vision
282 to understand how the shell colour may be perceived by birds, and to describe how
283 this varies in geographic space, and with respect to other characters such as
284 banding. The findings have significance for understanding the *Cepaea*
285 polymorphism, and the nature of the selection that acts upon it, as well as more
286 generally highlighting the need to objectively measure colour variation in other
287 systems before beginning to test for possible explanations.

288 Broadly, we found that both chromatic (Figure 2) and achromatic variation
289 (Supplementary Figure 1) is considerable, occurring over many perceptual units
290 (JNDs). If this variation, both within and among human-perceived colour morphs,
291 affects prey detection or identification by avian predators, then the presumption is
292 that the polymorphism must be impacted by natural selection. The current available
293 evidence suggests that animals in general use chromatic and achromatic signals for
294 separate tasks, for example, using achromatic signals to identify the location, shape
295 and motion of objects, while chromatic signals identify surface quality (Osorio and
296 Vorobyev 2005). However, while this is also likely the case for avian predators,
297 specific experimental evidence from birds is sparse (Osorio et al. 1999; Kang et al.
298 2015; White and Kemp 2016).

299 In our analysis, we found that chromatic variation in shells is continuously
300 distributed in visual space, meaning that there are no wholly discrete colours (Figure
301 2). Perhaps surprisingly, we found that the most variable chromatic axis (PC1; 87%)
302 that would be visible to a bird reflects the degree of saturation, or purity of colour.
303 Axes separating human-perceived colours showed less variation, PC2 (11%)
304 separating brown from yellow/pink, and PC3 (2%) broadly separating yellow, pink
305 and brown.

306 Despite the lack of discrete colours, density-based clustering recovered three
307 main shell types, which roughly correspond to human-perceived yellow, pink and
308 brown (Table 1; Figure 3). Brown shells were more common according to the
309 objective analysis than perceived by humans, with the frequency higher again when
310 using woodland shade as an illuminant. Therefore, prior studies that (necessarily)
311 used changes in frequencies of human-perceived colours to understand natural
312 selection on snail shells may have missed a significant part of the picture – not only
313 may birds use both achromatic and chromatic cues to differentiate morphs, but they
314 should also be able to perceive chromatic differences to a much finer precision than
315 a simple trivariate yellow, pink or brown categorisation that humans are obliged to
316 use in qualitative surveys. Of course, this does not mean that birds react to the many
317 morphs equally – it is possible that they categorically perceive a continuous variable
318 (Caves et al. 2018). Further investigations are needed, especially using a bird such
319 as the song thrush.

320 The effects of geographic location and banding pattern on variation in the
321 reflectance spectrum of snails were also examined, the initial aim being to develop
322 methods to *describe* variation, rather to *explain* it (e.g. by looking for correlations with
323 environmental variables, putative selective agents, etc., as others have done;

324 Silvertown et al. 2011). Generally, we found that geographic variables (latitude,
325 longitude and region) and banding are generally associated with different
326 frequencies of the three traditional colour morphs, with the main directional trend
327 being that yellow snails are most common at mid-latitudes, as was found in much
328 larger studies (Jones et al. 1977; Silvertown et al. 2011). Similarly, as previously
329 reported (Cain et al. 1960), epistasis meant that unbanded snails (O) were generally
330 more likely to be brown, and banded snails (B) were less likely to be brown.
331 Therefore, by establishing a method for quantitatively measuring colour, and
332 showing that a relatively small sample can be used to infer wide geographic patterns,
333 this work provides a baseline for further studies on the polymorphism.

334 *Discrete or indiscrete?*

335 Laboratory crosses in the past have revealed that the variation in the *Cepaea* shell
336 phenotype is predominantly controlled by a ‘supergene’, which in a recent definition
337 is a *genetic architecture involving multiple linked functional genetic elements that*
338 *allows switching between discrete, complex phenotypes maintained in a stable local*
339 *polymorphism* (Thompson and Jiggins 2014; Llaurens et al. 2017). This meaning fits
340 with the traditional view – and the classical ‘Fordian’ theory of polymorphism (Ford
341 1964) – that the ground colour of the shell is one of three more or less discrete
342 colour classes, either yellow, pink or brown, and indeed, is part of the reason that
343 *Cepaea* snails became a well-studied system. However, while scoring the shell
344 colour into different, discrete types is straightforward in offspring of individual crosses
345 in the lab, the acknowledged reality is that it is sometimes difficult to classify shells
346 consistently (e.g. see Table 1), especially when they are apparently intermediate in
347 form.

348 Now, in our study, we have shown definitively that the colour polymorphism of
349 *Cepaea nemoralis* is not discrete (Figures 2, 4). This finding emphasises the specific
350 practical problem for projects collecting and using shell polymorphism data,
351 especially those based entirely in the field and using citizen science (e.g. Evolution
352 Megalab; Silvertown et al. 2011). However, it also illustrates a more general
353 problem: if we do not have a precise definition of the *Cepaea* polymorphism and an
354 understanding of the underlying genetic control, then how can we claim to
355 understand the evolutionary and ecological factors that maintain colour variation?

356 Mathematical modelling is one method that can be used to explore the
357 evolution of polymorphism, and of most relevance to this work, the circumstances
358 that may or may not lead to discrete phenotypes. Historically, the argument of Ford
359 was that despite the fact that supergenes may appear as Mendelian loci, they were
360 actually rather complicated arrangements of several loci that are effectively
361 prevented from being broken up by recombination under most normal
362 circumstances. Thus, in both colour polymorphism in general (e.g. in side-blotched
363 lizards; Sinervo and Lively 1996) and specifically relating to supergenes (e.g. in
364 butterfly mimicry rings; Joron et al. 2011; Kunte et al. 2014), the distinctiveness of
365 the morphs is a central feature of the genetic control; the genetic architecture
366 specifically prevents phenotypes from “dissolving” into continuous trait distributions
367 (Ford 1964).

368 Much of the existing research has therefore begun from the premise of
369 understanding how evidently discrete types come about, and thus give insight into
370 the adaptive evolution of genome structure (Cuthill et al. 2017). In simulations it has
371 been shown that natural selection tends to favour lowered recombination when
372 intermediate genotypes are at a disadvantage; unlinked loci modify the phenotype to

373 adapt to local conditions (e.g. to a local Batesian model butterfly; Charlesworth and
374 Charlesworth 1975b, a; Llaurens et al. 2017). More recently, and perhaps most
375 directly relevant to understanding the *Cepaea* polymorphism, Kopp and Hermisson
376 (2006) devised a model to investigate the evolution of a quantitative trait under
377 frequency-dependent disruptive selection. Their finding was that over generations
378 most of the genetic variation tends to concentrate on a small number of loci.

379 The historic background is perhaps part of the reason that most of the recent
380 progress in understanding supergenes has mainly come from species or systems
381 that show simple, wholly discrete phenotypes, for example in butterfly mimicry rings
382 (Joron et al. 2011), or heterostylous plants (Li et al. 2016). However, in *Cepaea* there
383 are many colour morphs, such that colour variation is quantitative and due to a
384 supergene; in other species such as the guppy *Poecilia* and the cichlid
385 *Labeotropheus*, the inheritance of often considerable colour variation is due to
386 several loci, some sex-linked and others not (Tripathi et al. 2009; Thompson and
387 Jiggins 2014; Wellenreuther et al. 2014). Thus, developing theory on the impact of
388 negative frequency-dependent (apostatic) selection must be able to account for
389 these complexities, including those where supergenes are absent and variation is
390 quantitative, otherwise there is a risk that models will simply reaffirm what we already
391 know.

392 In one recent model, it was shown that crypsis and apostatic selection
393 together may act to maintain a large number of morphs within a population, and in
394 another apostatic selection was shown to maintain variation between similar species
395 (Franks and Oxford 2011, 2017). In another more recent study, a simulation was
396 used to explore the influence of predator perspective, selection, migration, and
397 genetic linkage on colour allele frequencies. The relative sizes of predator and prey

398 home ranges can result in large differences in morph composition between
399 neighbouring populations (Holmes et al. 2017). Finally, in an empirical study blue
400 jays *Cyanocitta cristata* searched for digital moths on mixtures of dark and light
401 patches at different scales of heterogeneity. It was found that complex backgrounds
402 with many moth-like features elicited a slow, serial search that depended heavily on
403 selective attention. The result was increased apostatic selection, producing a broad
404 range of moth phenotypes (Bond and Kamil 2006). All of these circumstances may
405 apply to the *Cepaea* colour polymorphism.

406 Overall, there is an open debate – but little empirical data – on how the
407 relative heterogeneity of the environment/substrate, density, distance or motion may
408 influence the selection for crypsis or negative frequency dependence (Cuthill et al.
409 2017; Barnett et al. 2018). As Surmacki *et al.* (2013) summarised, if heterogeneous
410 areas consist of large patches of diverse habitats then this may promote the
411 evolution of specialist morphs through selection for crypsis, producing a few distinct
412 or specialist morphs, each more or less well matched to the coloration of the
413 preferred habitat type (Endler 1978; Bond 2007). If instead there are a mixture of
414 small microhabitats, apostatic selection is more likely to result in multiple morphs that
415 may be equally cryptic in all “grains” of the habitat. This is because in such
416 circumstances, predators use search images of the most common morph, and this
417 can lead to frequency-dependent selection.

418 *Supergenes return*

419 In contrast to a relative paucity of field data, and a relatively lack of progress in
420 establishing baseline theory, advances in DNA sequencing technology have meant
421 that knowledge on the genetics of colour polymorphism is advancing rapidly. As
422 hypothesised, in the still relatively few supergenes that have been fully

423 characterised, the discrete phenotypes are maintained due to close physical
424 proximity of the gene(s) and/or tight linkage (Joron et al. 2011; Kunte et al. 2014;
425 Gautier et al. 2018).

426 In comparison, a few more general studies on colour polymorphism, rather
427 than on supergenes specifically, have begun to reveal the extent of phenotypic
428 variation, and whether discrete or indiscrete. For example, reflectance spectra have
429 been used to show that even though humans perceived the colour variation in the
430 eggs of African cuckoo finch *Anomalospiza imberbis* as falling into discrete
431 categories, the variation was actually continuous (Spottiswoode and Stevens 2010,
432 2011). Similarly, tawny dragon lizard *Ctenophorus descresii* does have discrete
433 colour morphs, but there is still considerable variation within each morph (Teasdale
434 et al. 2013). Further quantitative studies in other lizards in which colour
435 polymorphism has traditionally been treated as qualitative are also increasingly
436 showing that there are significant overlaps in colour (Cote et al. 2008; Vercken et al.
437 2008; Paterson and Blouin-Demers 2017).

438 In *Cepaea nemoralis* snails, the colour and banding elements of the
439 supergene have been mapped (Richards et al. 2013) but we remain ignorant of the
440 underlying genetics and the precise nature of the selection that acts upon the
441 polymorphism. For instance, models of supergene evolution require that intermediate
442 phenotypes are disadvantaged – this makes sense with respect to Batesian mimics
443 or distylous flowers – but in snails a rare intermediate might be at an advantage, due
444 to apostatic selection. At the molecular level, one scenario is that the extreme and
445 effectively continuous colour variation of the shells is due to a corresponding high
446 number of colour alleles within the supergene. An alternative scenario is that there
447 actually relatively few colour alleles, with much of the chromatic variation due to

448 effects of other modifying loci (Charlesworth and Charlesworth 1975b). A final
449 consideration is that while colour variation might be continuous across a grand
450 geographic scale, if most local populations are founded by few individuals, then local
451 variation might be discrete, which is all that matters from a selective point of view.
452 This is more likely to be the case when both colour and banding are considered as
453 the visible phenotype, especially since they are frequently in linkage disequilibrium
454 (Cook 2017).

455 Overall, by establishing a method for quantitatively measuring colour, this
456 work provides a baseline for further studies on the polymorphism, both from the
457 perspective of understanding the nature of selection, and ultimately, also the genes
458 involved. To reconcile and test competing theories with the empirical observations,
459 the next steps must be to identify the component parts and evolutionary origins of
460 the supergene in *C. nemoralis*, develop a model of frequency-dependent selection,
461 and further understand how birds react to specific elements of the chromatic
462 variation. When all of these findings are brought together, only then can we begin to
463 understand the evolutionary and ecological factors that maintain this “problem with
464 too many solutions.” Whatever the final outcome, there is no risk that *Cepaea* snails
465 will be relegated to “other adaptive polymorphism” (Thompson and Jiggins 2014),
466 especially because, as Jones *et al.* (1977) suggested, it is important to study
467 organisms for which polymorphism may be explained by a variety of processes,
468 precisely because they are more realistic.

469 Supplementary material

470 Supplementary Movies 1, 2. Supplementary Figure 1. Supplementary Table 1.

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480 Data archiving

481 Raw reflectance data will be included with the manuscript or uploaded to Dryad upon
482 acceptance of the manuscript.

483 *Literature cited*

- 484 Barnett, J. B., C. Michalis, N. E. Scott-Samuel, and I. C. Cuthill. 2018. Distance-
485 dependent defensive coloration in the poison frog *Dendrobates tinctorius*,
486 Dendrobatidae. *Proc Natl Acad Sci USA* 115:6416-6421.
- 487 Bond, A. B. 2007. The evolution of color polymorphism: crypticity searching images,
488 and apostatic selection. *Annu Rev Ecol Evol Syst* 38:489-514.
- 489 Bond, A. B. and A. C. Kamil. 1998. Apostatic selection by blue jays produces
490 balanced polymorphism in virtual prey. *Nature* 395:594-596.
- 491 Bond, A. B. and A. C. Kamil. 2002. Visual predators select for crypticity and
492 polymorphism in virtual prey. *Nature* 415:609-613.
- 493 Bond, A. B. and A. C. Kamil. 2006. Spatial heterogeneity, predator cognition, and the
494 evolution of color polymorphism in virtual prey. *Proc Natl Acad Sci USA*
495 103:3214-3219.
- 496 Cain, A. J., J. M. B. King, and P. M. Sheppard. 1960. New data on the genetics of
497 polymorphism in the snail *Cepaea nemoralis* L. *Genetics* 45:393-411.
- 498 Cain, A. J. and P. M. Sheppard. 1954. Natural selection in *Cepaea*. *Genetics* 39:89-
499 116.

- 500 Cain, A. J., P. M. Sheppard, and J. M. B. King. 1968. Studies on *Cepaea*. I. Genetics
501 of some morphs and varieties of *Cepaea nemoralis* (L). Philosophical
502 Transactions of the Royal Society of London Series B-Biological Sciences
503 253:383-&.
- 504 Cameron, R. A. D. and L. M. Cook. 2012. Habitat and the shell polymorphism of
505 *Cepaea nemoralis* (L.): interrogating the Evolution Megalab database. J
506 Molluscan Stud 78:179-184.
- 507 Cameron, R. A. D., L. M. Cook, and J. J. D. Greenwood. 2013. Change and stability
508 in a steep morph-frequency cline in the snail *Cepaea nemoralis* (L.) over 43
509 years. Biol J Linn Soc 108:473-483.
- 510 Cassey, P., M. Honza, T. Grim, and M. E. Hauber. 2008. The modelling of avian
511 visual perception predicts behavioural rejection responses to foreign egg
512 colours. Biol Lett 4:515-517.
- 513 Caves, E. M., P. A. Green, M. N. Zippel, S. Peters, S. Johnsen, and S. Nowicki.
514 2018. Categorical perception of colour signals in a songbird. Nature.
- 515 Charlesworth, D. and B. Charlesworth. 1975a. Theoretical genetics of Batesian
516 mimicry I. Single-locus models. J Theor Biol 55:283-303.
- 517 Charlesworth, D. and B. Charlesworth. 1975b. Theoretical Genetics of Batesian
518 Mimicry II. Evolution of supergenes. J Theor Biol 55:305-324.
- 519 Cook, L. M. 2014. Morph frequency in British *Cepaea nemoralis*: what has changed
520 in half a century? J Molluscan Stud 80:43-46.
- 521 Cook, L. M. 2017. Reflections on molluscan shell polymorphisms. Biol J Linn Soc
522 121:717-730.
- 523 Cook, L. M., R. H. Cowie, and J. S. Jones. 1999. Change in morph frequency in the
524 snail *Cepaea nemoralis* on the Marlborough Downs. Heredity 82:336-342.
- 525 Cook, L. M., B. S. Grant, I. J. Saccheri, and J. Mallet. 2012. Selective bird predation
526 on the peppered moth: the last experiment of Michael Majerus. Biol Lett
527 8:609-612.
- 528 Cote, J., J. F. Le Galliard, J. M. Rossi, and P. S. Fitze. 2008. Environmentally
529 induced changes in carotenoid-based coloration of female lizards: a comment
530 on Vercken et al. J Evol Biol 21:1165-1172.
- 531 Cuthill, I. C. 2006. Color perception. Pp. 3-40 in G. E. Hill, and K. J. McGraw, eds.
532 Bird coloration. Cambridge (MA): Harvard University Press.
- 533 Cuthill, I. C., W. L. Allen, K. Arbuckle, B. Caspers, G. Chaplin, M. E. Hauber, G. E.
534 Hill, N. G. Jablonski, C. D. Jiggins, A. Kelber, J. Mappes, J. Marshall, R.
535 Merrill, D. Osorio, R. Prum, N. W. Roberts, A. Roulin, H. M. Rowland, T. N.
536 Sherratt, J. Skelhorn, M. P. Speed, M. Stevens, M. C. Stoddard, D. Stuart-
537 Fox, L. Talas, E. Tibbetts, and T. Caro. 2017. The biology of color. Science
538 357.

- 539 Dale, J., C. J. Dey, K. Delhey, B. Kempenaers, and M. Valcu. 2015. The effects of
540 life history and sexual selection on male and female plumage colouration.
541 Nature 527:367-370.
- 542 Davison, A. and B. Clarke. 2000. History or current selection? A molecular analysis
543 of 'area effects' in the land snail *Cepaea nemoralis*. Proc R Soc Lond B Biol
544 Sci 267:1399-1405.
- 545 Delhey, K., V. Delhey, B. Kempenaers, and A. Peters. 2015. A practical framework
546 to analyze variation in animal colors using visual models. Behav Ecol 26:367-
547 375.
- 548 Delhey, K., B. Szecsenyi, S. Nakagawa, and A. Peters. 2017. Conspicuous plumage
549 colours are highly variable. Proc R Soc Lond B Biol Sci 284.
- 550 Endler, J. 1978. A predator's view of animal color patterns. Pp. 319-364 in M. K.
551 Hecht, W. C. Steere, and B. Wallace, eds. Evolutionary biology. Plenum
552 Press, New York.
- 553 Endler, J. A. 1990. On the measurement and classification of color in studies of
554 animal color patterns. Biol J Linn Soc 41:315-352.
- 555 Ford, E. B. 1964. Ecological Genetics. Methuen, London.
- 556 Franks, D. W. and G. S. Oxford. 2009. The evolution of exuberant visible
557 polymorphisms. Evolution 63:2697-2706.
- 558 Franks, D. W. and G. S. Oxford. 2011. The interrelationship between crypsis and
559 colour polymorphism. Ecol Lett 14:295-300.
- 560 Franks, D. W. and G. S. Oxford. 2017. The co-evolution of anti-predator
561 polymorphisms in sympatric populations. Biol J Linn Soc 122:729-737.
- 562 Gautier, M., J. Yamaguchi, J. Foucaud, A. Loiseau, A. Ausset, B. Facon, B.
563 Gschloessl, J. Lagnel, E. Loire, H. Parrinello, D. Severac, C. Lopez-Roques,
564 C. Donnadieu, M. Manno, H. Berges, K. Gharbi, L. Lawson-Handley, L.-S.
565 Zang, H. Vogel, A. Estoup, and B. Prud'homme. 2018. The genomic basis of
566 colour pattern polymorphism in the harlequin ladybird. bioRxiv.
- 567 Grindon, A. J. and A. Davison. 2013. Irish *Cepaea nemoralis* land snails have a
568 cryptic Franco-Iberian origin that is most easily explained by the movements
569 of Mesolithic humans. PLoS One 8:e65792.
- 570 Hart, N. S. 2001. Variations in cone photoreceptor abundance and the visual ecology
571 of birds. Journal of comparative physiology. A, Sensory, neural, and
572 behavioral physiology 187:685-697.
- 573 Hart, N. S., J. C. Partridge, I. C. Cuthill, and A. T. Bennett. 2000. Visual pigments, oil
574 droplets, ocular media and cone photoreceptor distribution in two species of
575 passerine bird: the blue tit (*Parus caeruleus* L.) and the blackbird (*Turdus*
576 *merula* L.). J Comp Physiol A 186:375-387.

- 577 Hoekstra, H. E. 2006. Genetics, development and evolution of adaptive pigmentation
578 in vertebrates. *Heredity* 97:222-234.
- 579 Holmes, I. A., M. R. Grundler, and A. R. D. Rabosky. 2017. Predator perspective
580 drives geographic variation in frequency-dependent polymorphism. *Am Nat*
581 190:E78-E93.
- 582 Hugall, A. F. and D. Stuart-Fox. 2012. Accelerated speciation in colour-polymorphic
583 birds. *Nature* 485:631-634.
- 584 Jones, J. S., B. H. Leith, and P. Rawlings. 1977. Polymorphism in *Cepaea*: a
585 problem with too many solutions? *Annu Rev Ecol Syst* 8:109-143.
- 586 Joron, M., L. Frezal, R. T. Jones, N. L. Chamberlain, S. F. Lee, C. R. Haag, A.
587 Whibley, M. Becuwe, S. W. Baxter, L. Ferguson, P. A. Wilkinson, C. Salazar,
588 C. Davidson, R. Clark, M. A. Quail, H. Beasley, R. Glithero, C. Lloyd, S. Sims,
589 M. C. Jones, J. Rogers, C. D. Jiggins, and R. H. ffrench-Constant. 2011.
590 Chromosomal rearrangements maintain a polymorphic supergene controlling
591 butterfly mimicry. *Nature* 477:203-206.
- 592 Kang, C., M. Stevens, J. Y. Moon, S. I. Lee, and P. G. Jablonski. 2015. Camouflage
593 through behavior in moths: the role of background matching and disruptive
594 coloration. *Behav Ecol* 26:45-54.
- 595 Kerkvliet, J., T. de Boer, M. Schilthuizen, and K. Kraaijeveld. 2017. Candidate genes
596 for shell colour polymorphism in *Cepaea nemoralis*. *Peerj* 5:e3715.
- 597 Kopp, M. and J. Hermisson. 2006. The evolution of genetic architecture under
598 frequency-dependent disruptive selection. *Evolution* 60:1537-1550.
- 599 Kunte, K., W. Zhang, A. Tenger-Trolander, D. H. Palmer, A. Martin, R. D. Reed, S.
600 P. Mullen, and M. R. Kronforst. 2014. Doublesex is a mimicry supergene.
601 *Nature* 507:229–232.
- 602 Li, J. H., J. M. Cocker, J. Wright, M. A. Webster, M. McMullan, S. Dyer, D.
603 Swarbreck, M. Caccamo, C. van Oosterhout, and P. M. Gilmartin. 2016.
604 Genetic architecture and evolution of the S locus supergene in *Primula*
605 *vulgaris*. *Nat Plants* 2.
- 606 Llaurens, V., A. Whibley, and M. Joron. 2017. Genetic architecture and balancing
607 selection: the life and death of differentiated variants. *Mol Ecol* 26:2430-2448.
- 608 Maia, R., C. M. Eliason, P. P. Bitton, S. M. Doucet, and M. D. Shawkey. 2013. pavo:
609 an R package for the analysis, visualization and organization of spectral data.
610 *Methods Ecol Evol* 4:906-913.
- 611 Mann, K. and D. J. Jackson. 2014. Characterization of the pigmented shell-forming
612 proteome of the common grove snail *Cepaea nemoralis*. *BMC Genomics* 15.
- 613 McKinnon, J. S. and M. E. R. Pierotti. 2010. Colour polymorphism and correlated
614 characters: genetic mechanisms and evolution. *Mol Ecol* 19:5101-5125.

- 615 McLean, C. A. and D. Stuart-Fox. 2014. Geographic variation in animal colour
616 polymorphisms and its role in speciation. *Biol Rev (Camb)* 89:860-873.
- 617 Montgomerie, R. 2006. Analysing Colors. Pp. 90-147 in G. E. Hill, and K. J. McGraw,
618 eds. *Bird Coloration*. Cambridge (MA): Harvard University Press.
- 619 Osorio, D., A. Miklosi, and Z. Gonda. 1999. Visual ecology and perception of
620 coloration patterns by domestic chicks. *Evol Ecol* 13:673-689.
- 621 Osorio, D. and M. Vorobyev. 2005. Photoreceptor spectral sensitivities in terrestrial
622 animals: adaptations for luminance and colour vision. *Proceedings of the*
623 *Royal Society B-Biological Sciences* 272:1745-1752.
- 624 Ozgo, M., T. S. Liew, N. B. Webster, and M. Schilthuizen. 2017. Inferring
625 microevolution from museum collections and resampling: lessons learned
626 from *Cepaea*. *Peerj* 5.
- 627 Ožgo, M. and M. Schilthuizen. 2012. Evolutionary change in *Cepaea nemoralis* shell
628 colour over 43 years. *Global Change Biology* 18:74-81.
- 629 Paterson, J. E. and G. Blouin-Demers. 2017. Distinguishing discrete polymorphism
630 from continuous variation in throat colour of tree lizards, *Urosaurus ornatus*.
631 *Biol J Linn Soc* 121:72-81.
- 632 Richards, P. M., M. M. Liu, N. Lowe, J. W. Davey, M. L. Blaxter, and A. Davison.
633 2013. RAD-Seq derived markers flank the shell colour and banding loci of the
634 *Cepaea nemoralis* supergene. *Mol Ecol* 22:3077-3089.
- 635 San-Jose, L. M. and A. Roulin. 2017. Genomics of coloration in natural animal
636 populations. *Philos Trans R Soc Lond B Biol Sci* 372.
- 637 Schilthuizen, M. 2013. Rapid, habitat-related evolution of land snail colour morphs on
638 reclaimed land. *Heredity* 110:247-252.
- 639 Scrucca, L., M. Fop, T. B. Murphy, and A. E. Raftery. 2016. Mclust 5: clustering,
640 classification and density estimation using Gaussian finite mixture models.
641 *The R Journal* 8:205-233.
- 642 Silvertown, J., L. Cook, R. Cameron, M. Dodd, K. McConway, J. Worthington, P.
643 Skelton, C. Anton, O. Bossdorf, B. Baur, M. Schilthuizen, B. Fontaine, H.
644 Sattmann, G. Bertorelle, M. Correia, C. Oliveira, B. Pokryszko, M. Ozgo, A.
645 Stalazas, E. Gill, U. Rammul, P. Solymos, Z. Feher, and X. Juan. 2011.
646 Citizen science reveals unexpected continental scale evolutionary change in a
647 model organism. *Plos One* 6:e18927.
- 648 Sinervo, B. and C. M. Lively. 1996. The rock–paper–scissors game and the evolution
649 of alternative male strategies. *Nature* 380:240.
- 650 Spottiswoode, C. N. and M. Stevens. 2010. Visual modeling shows that avian host
651 parents use multiple visual cues in rejecting parasitic eggs. *Proc Natl Acad*
652 *Sci USA* 107:8672-8676.

- 653 Spottiswoode, C. N. and M. Stevens. 2011. How to evade a coevolving brood
654 parasite: egg discrimination versus egg variability as host defences. Proc R
655 Soc Biol Sci Ser B 278:3566-3573.
- 656 Staples-Browne, R. 1908. On the inheritance of colour in domestic pigeons, with
657 special reference to reversion. Proc Zool Soc Lond 78:67-104.
- 658 Surmacki, A., A. Ozarowska-Nowicka, and Z. M. Rosin. 2013. Color polymorphism in
659 a land snail *Cepaea nemoralis* (Pulmonata: Helicidae) as viewed by potential
660 avian predators. Naturwissenschaften 100:533-540.
- 661 Taylor, C. H., T. Reader, and F. Gilbert. 2016. Hoverflies are imperfect mimics of
662 wasp colouration. Evol Ecol 30:567-581.
- 663 Teasdale, L. C., M. Stevens, and D. Stuart-Fox. 2013. Discrete colour polymorphism
664 in the tawny dragon lizard (*Ctenophorus decresii*) and differences in signal
665 conspicuousness among morphs. J Evol Biol 26:1035-1046.
- 666 Thompson, M. J. and C. D. Jiggins. 2014. Supergenes and their role in evolution.
667 Heredity 113:1-8.
- 668 Tripathi, N., M. Hoffmann, E. M. Willing, C. Lanz, D. Weigel, and C. Dreyer. 2009.
669 Genetic linkage map of the guppy, *Poecilia reticulata*, and quantitative trait
670 loci analysis of male size and colour variation. Proc R Soc Biol Sci Ser B
671 276:2195-2208.
- 672 van't Hof, A. E., P. Campagne, D. J. Rigden, C. J. Yung, J. Lingley, M. A. Quail, N.
673 Hall, A. C. Darby, and I. J. Saccheri. 2016. The industrial melanism mutation
674 in British peppered moths is a transposable element. Nature 534:102-105.
- 675 Vercken, E., B. Sinervo, and J. Clobert. 2008. Colour variation in female common
676 lizards: why we should speak of morphs, a reply to Cote et al. J Evol Biol
677 21:1160-1164.
- 678 Vorobyev, M. and D. Osorio. 1998. Receptor noise as a determinant of colour
679 thresholds. Proc R Soc Lond B Biol Sci 265:351-358.
- 680 Vorobyev, M., D. Osorio, A. T. D. Bennett, N. J. Marshall, and I. C. Cuthill. 1998.
681 Tetrachromacy, oil droplets and bird plumage colours. J Comp Physiol A
682 183:621-633.
- 683 Wellenreuther, M., E. I. Svensson, and B. Hansson. 2014. Sexual selection and
684 genetic colour polymorphisms in animals. Mol Ecol 23:5398-5414.
- 685 Wheldale, M. 1907. The inheritance of flower colour in *Antirrhinum majus*. Proc R
686 Soc Biol Sci Ser B 79:288-305.
- 687 White, T. E. and D. J. Kemp. 2016. Color polymorphic lures target different visual
688 channels in prey. Evolution 70:1398-1408.
- 689 Williams, S. T. 2017. Molluscan shell colour. Biol Rev (Camb) 92:1039–1058.
690

Table 1. Comparison between human perceived colour categories and Mclust-defined groups. Shells that were scored the same are on the diagonal (in bold). Yellow and pink were most common, and so the absolute number of discordant scores was relatively low. Brown had by far the highest proportion of discordant scores.

		Colour (human)				
		yellow	pink	brown	total	% total misclassified
Colour (Mclust)	yellow	495	44	0	539	8.2
	pink	118	379	14	511	25.8
	brown	9	90	23	122	81.1
	total	622	513	37		

Table 2. Results of likelihood ratio tests of the terms in binomial GLMMs of the effects of geographic variables and banding phenotype on the probability that a snail belonged to each of the three colour morphs. Significant p-values are in bold. The effects of modelling large-scale geographic variation in two ways are illustrated by the AIC values for the best model in which linear and quadratic effects of latitude and longitude were included (AICa), and the best model in which geographic region was included (AICb). All models include a random effect for site.

	Brown			Pink			Yellow		
AICa (df)				865.2 (11)			850.2 (8)		
AICb (df)				875.4 (22)			872.09 (22)		
Term	df	χ^2	p	df	χ^2	p	df	χ^2	p
Latitude	1	1.747	0.186	1	0.218	0.641	1	2.62	0.106
Longitude	1	1.572	0.21	1	1.382	0.24	1	0.001	0.986
Latitude ²	1	2.019	0.155	1	1.832	< 0.001	1	4.732	0.03
Longitude ²	1	2.587	0.108	1	0.214	0.644	1	0.355	0.551
Banding	2	10.751	0.005	2	0.286	0.867	2	3.056	0.217
Latitude x longitude	1	0.239	0.625	1	0.0877	0.767	1	0.023	0.88
Latitude x banding	2	1.411	0.494	2	6.345	0.042	2	4.972	0.083
Longitude x banding	2	4.255	0.119	2	12.935	0.002	2	27.043	< 0.001
Region	6	10.586	0.102	6	7.481	0.279	6	5.924	0.432
Banding x region	12	12.824	0.382	12	34.564	0.001	12	31.448	0.002

Figure 1. Sample sites across Europe, grouped by geographically contiguous regions. A (England, n=397), B (Ireland, n=144), C (North Spain and Pyrenees, n=112), D (North France, Belgium, Germany, n= 178), E (Scandinavia, n=77), F (Poland, n=126) and all others (n=138).

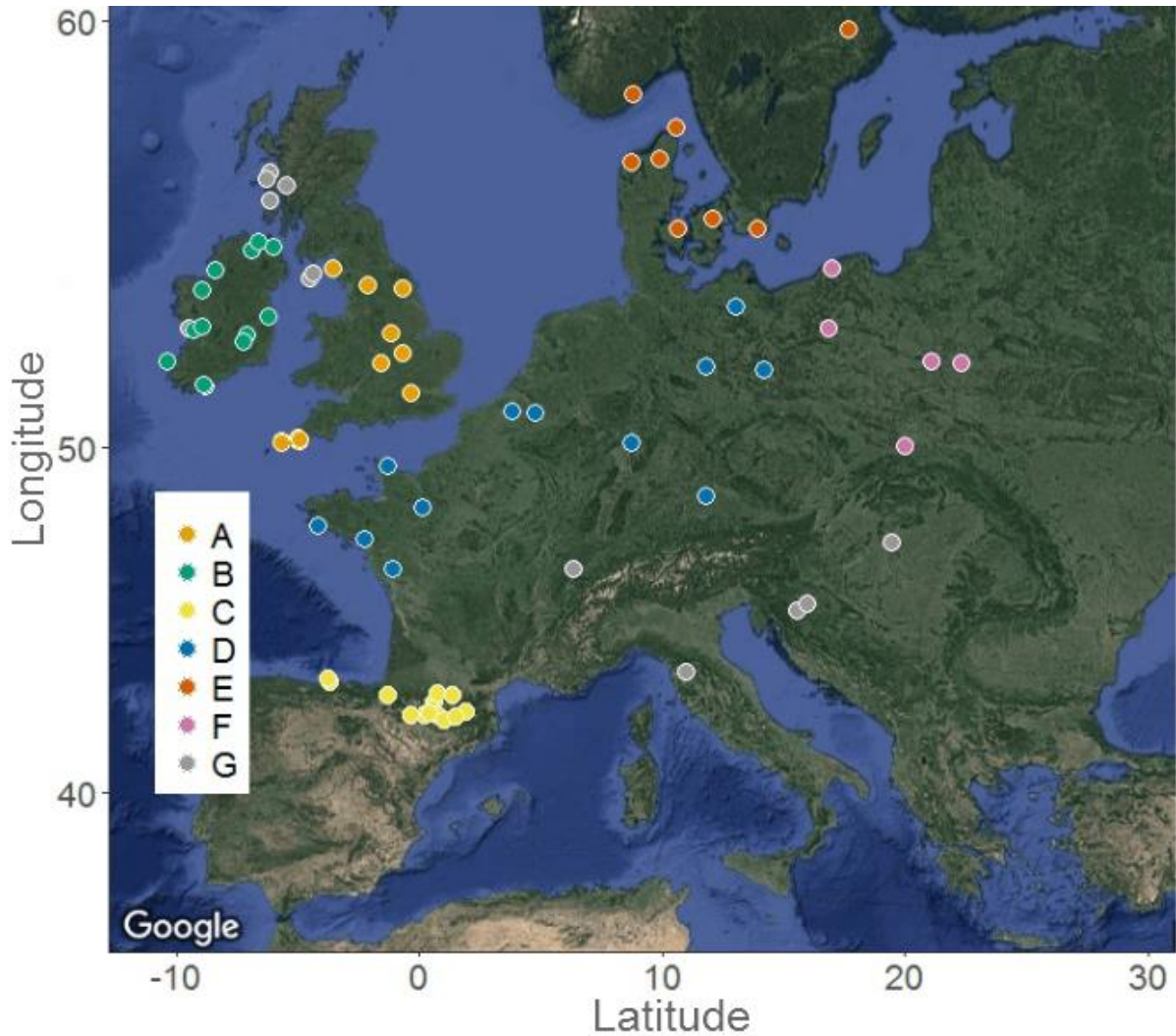


Figure 2. Axes of chromatic variation in the shell of *C. nemoralis*, using avian visual space, shown from two different perspectives (see also Supplementary Movie 2) . Units on x, y and z axes are in JNDs. The solid lines illustrate variation along the first three principal components; individual points are coloured according to Mclust classification of the shell, either yellow, pink or brown. Top: Variation along PC1 (87%) mainly represents differences in saturation between shells. PC2 (11%) shows relatively higher stimulation of L cones and lesser stimulation of M and S cones, and tends to separate brown from pink/yellow. Bottom: PC3 (2%) shows relatively high stimulation of the M cones compared to lesser stimulation of the S and L cones, and tends to separate yellow from pink and pink from brown.

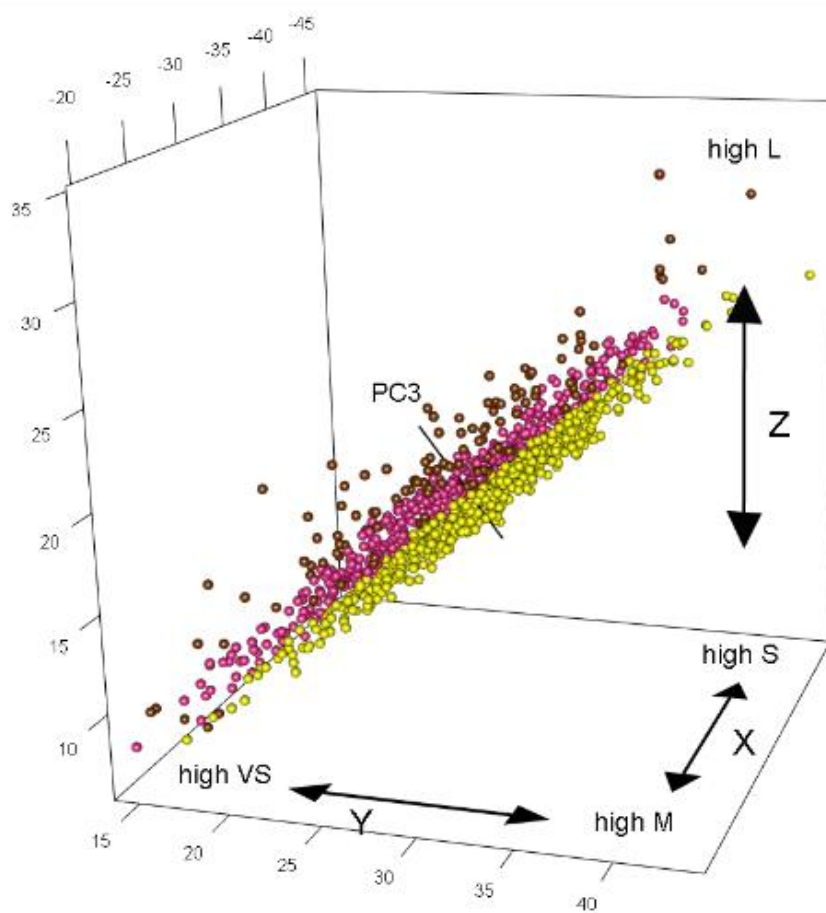
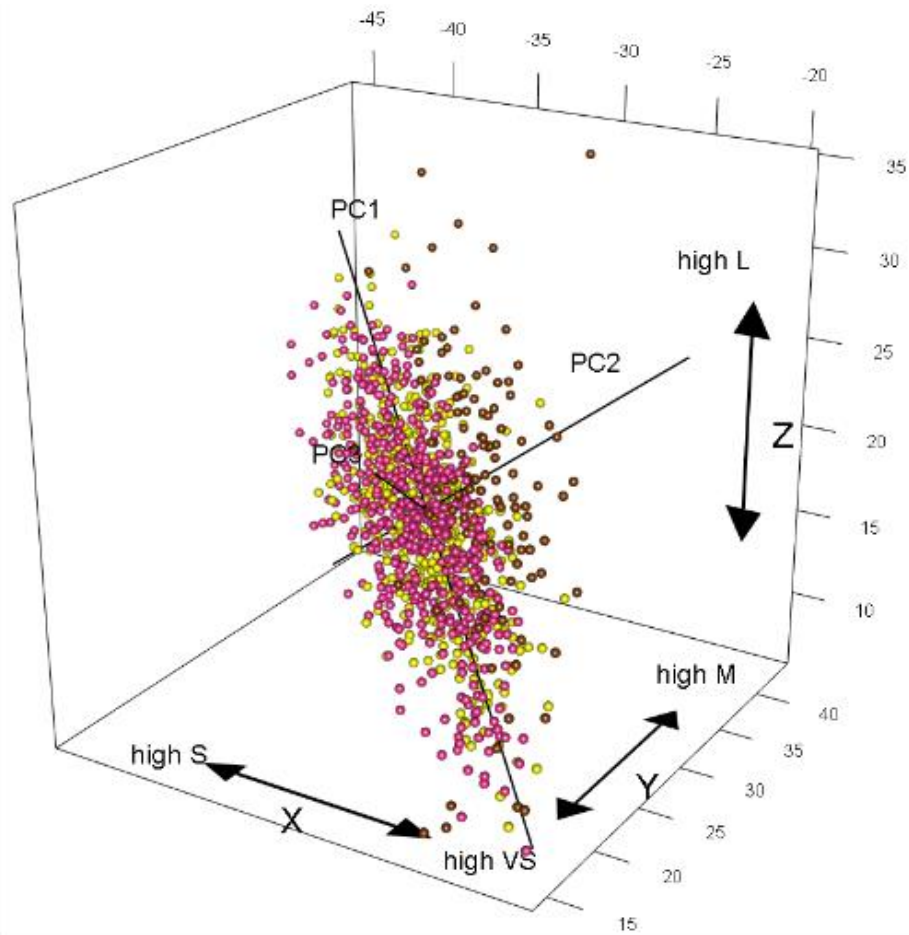


Figure 3. Interquartile ranges of the average normalised reflectance spectra for the principal component axes shown in Figure 2. These plots confirm that variation on PC1 mainly represents differences in saturation between shells; PC2 represents relatively higher stimulation of L cones and lesser stimulation of M and S cones; PC3 represents relatively high stimulation of the M cones compared to lesser stimulation of the S and L cones.

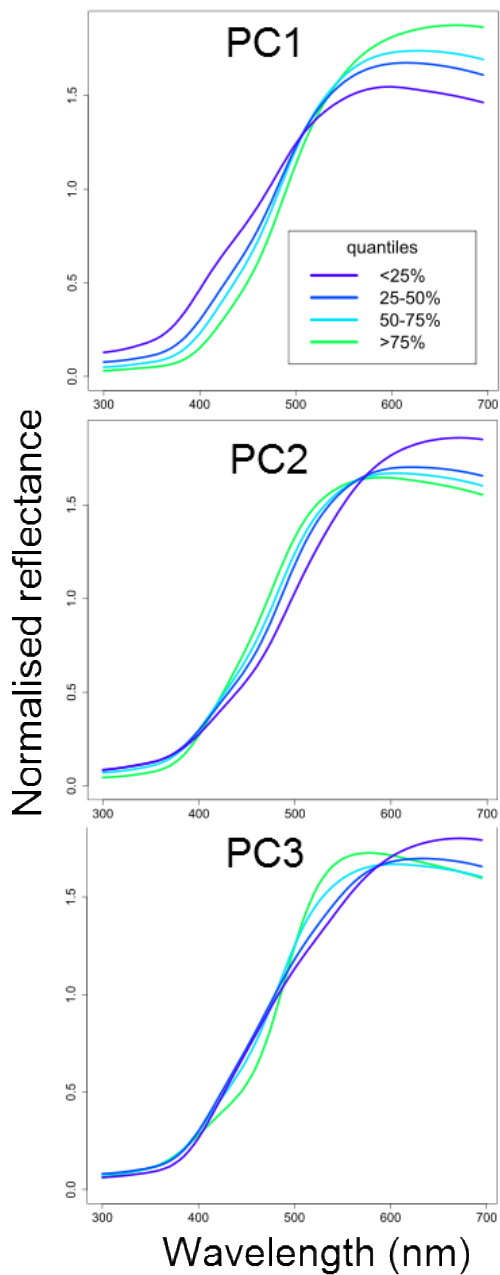


Figure 4. Scatterplot and associated density plot, showing variation along three principal component axes. Units are in JNDs. Points are coloured according to Mclust classification of the shell, either yellow, pink or brown. Top: PC1 versus PC2. Bottom: PC2 versus PC3.

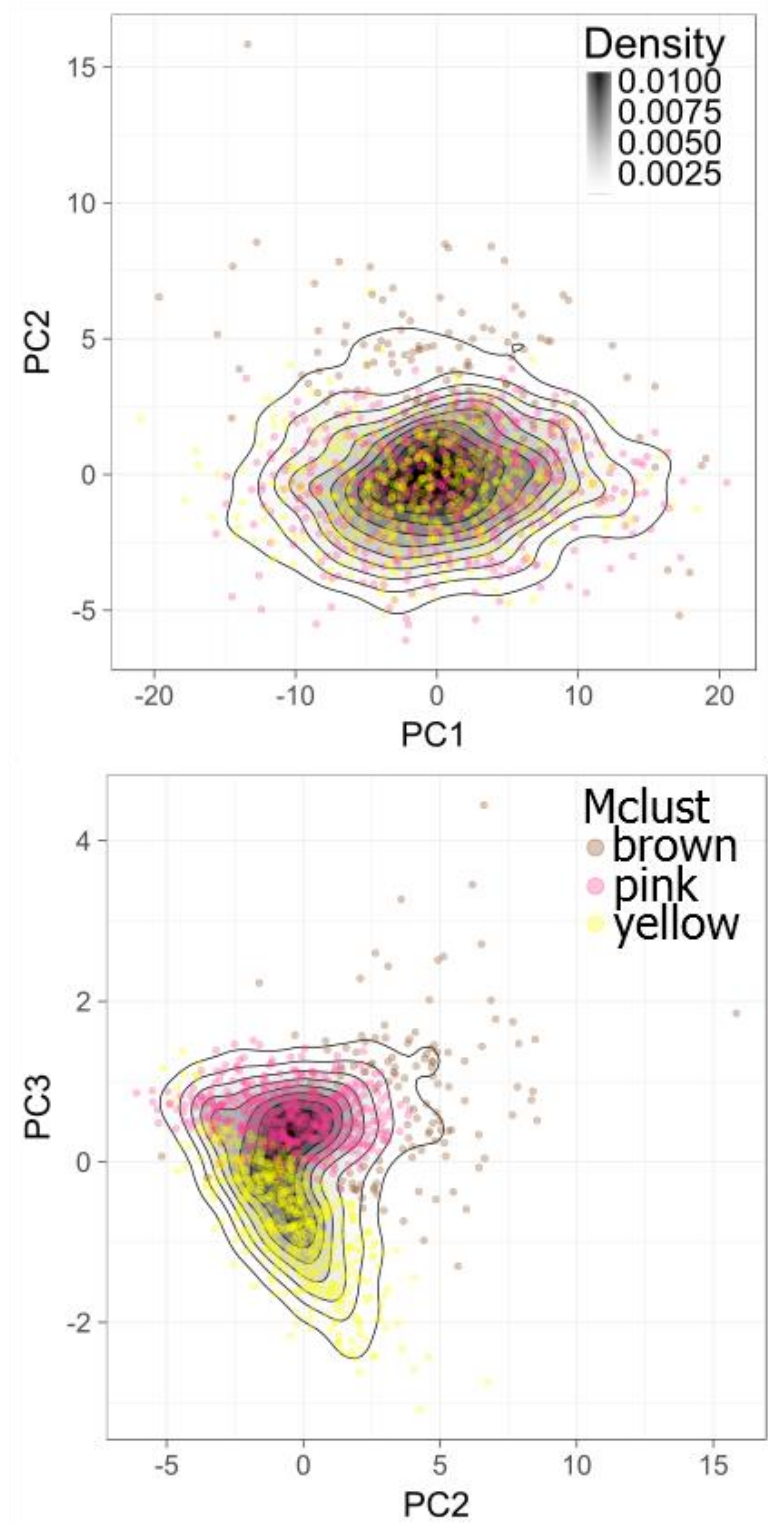


Figure 5. Scaled effects of latitude on the proportion of pink shells (top), and longitude on banding and proportion of pink shells.

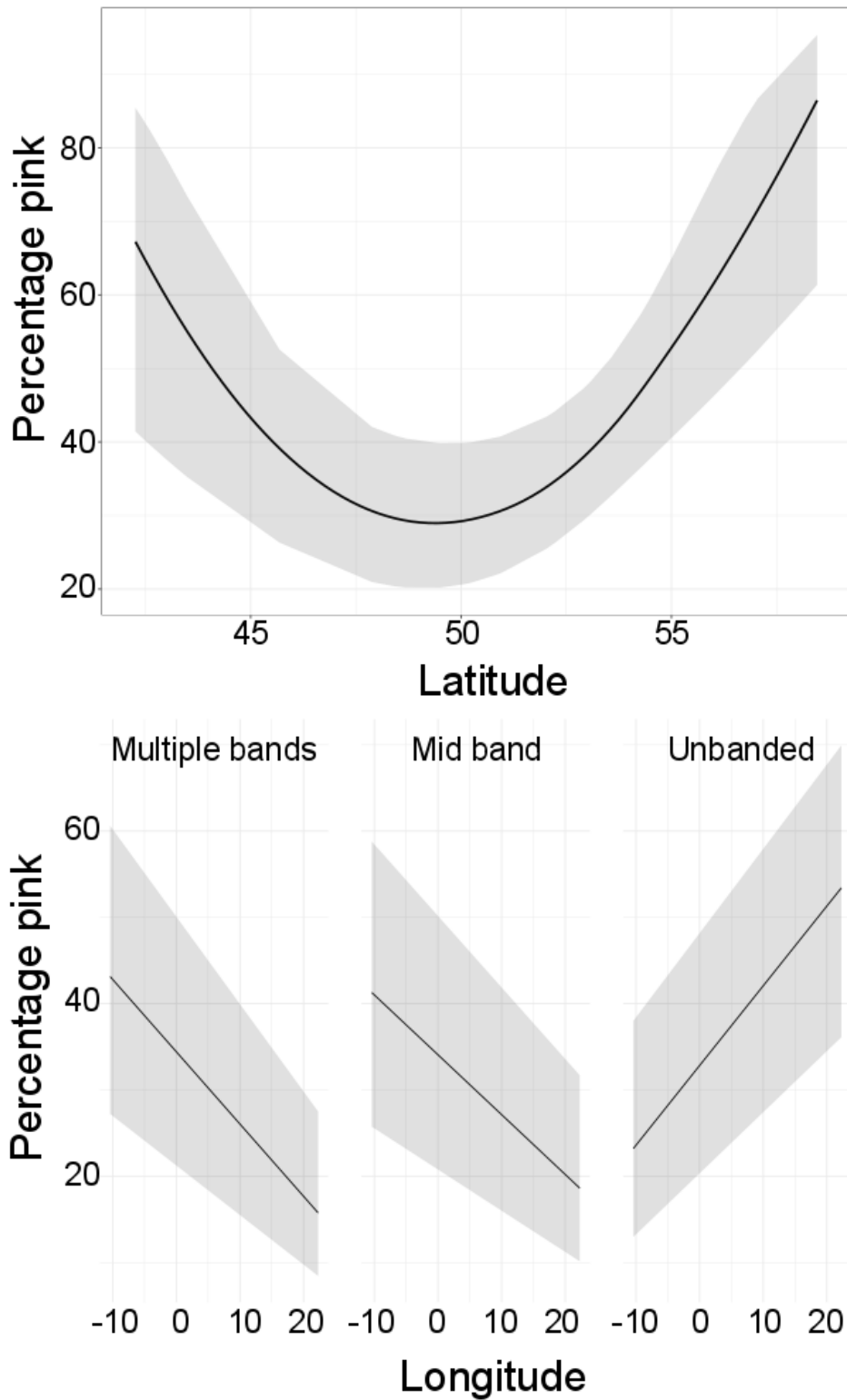
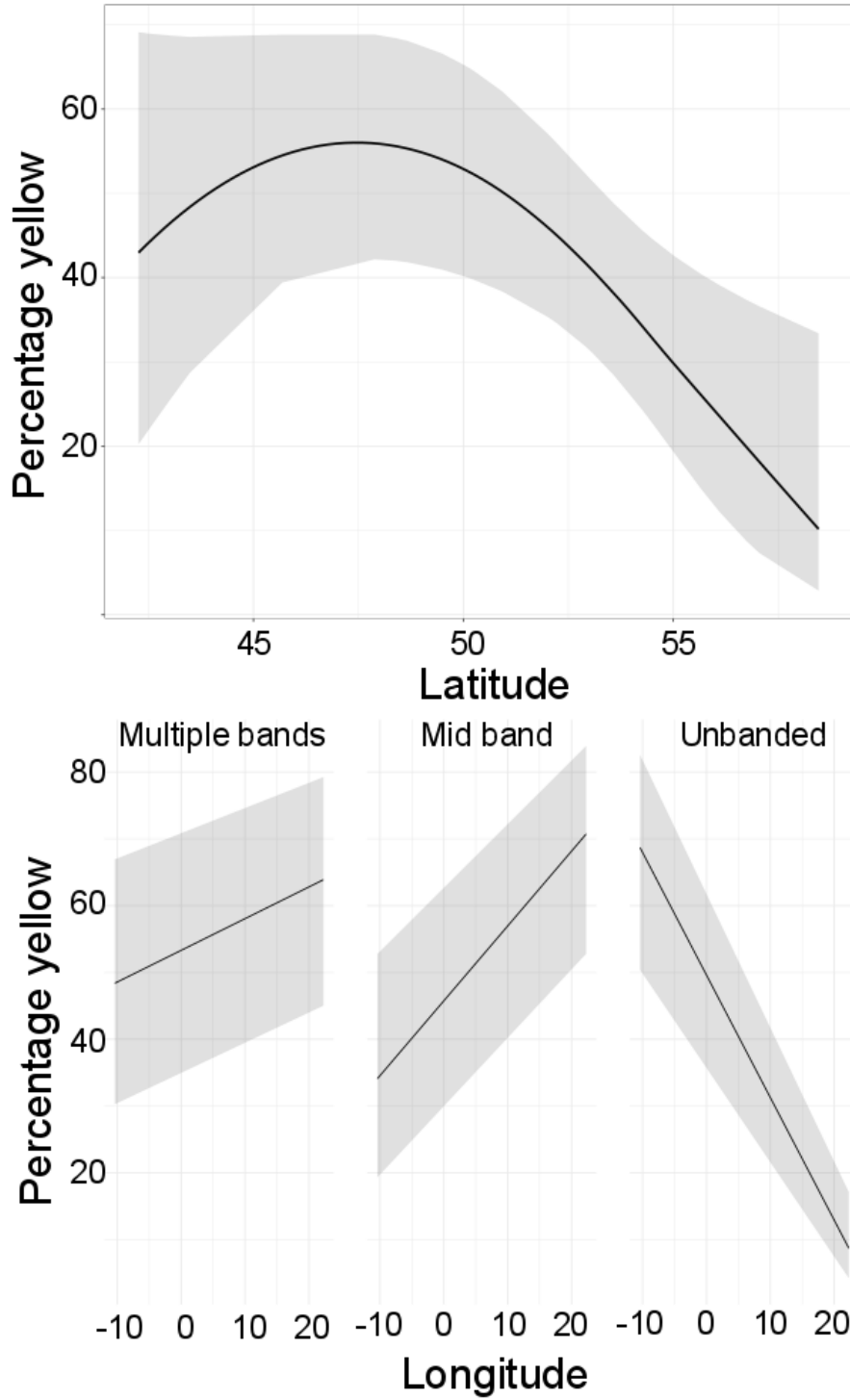
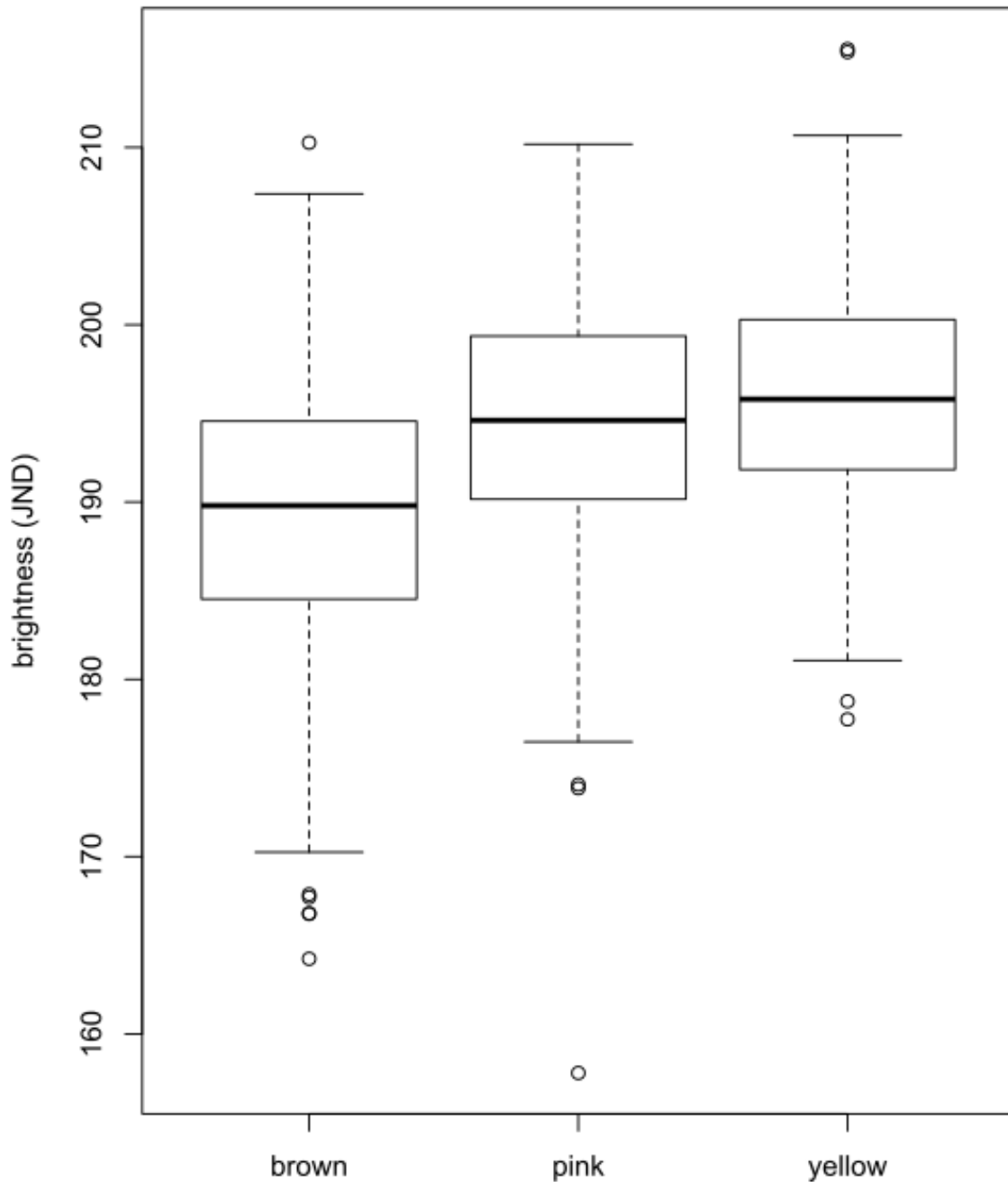


Figure 6. Scaled effects of latitude on the proportion of yellow shells (top), and longitude on banding and proportion of yellow shells.



Supplementary Figure 1. Boxplot showing extent of achromatic variation in Mclust-defined colour morphs of *Cepaea nemoralis*.



Supplementary Movie 1. Animation showing axes of chromatic variation in the shell of *C. nemoralis*, using avian visual space. Units on x, y and z axes are in JNDs. The solid lines illustrate variation along the first three principal components; individual points are coloured according to human-scoring of the shell, either yellow, pink or brown.

Supplementary Movie 2. Animation showing axes of chromatic variation in the shell of *C. nemoralis*, using avian visual space. Units on x, y and z axes are in JNDs. The solid lines illustrate variation along the first three principal components; individual points are coloured according to Mclust classification of the shell, either yellow, pink or brown.