Title: Standardized biogeographic grouping system for annotating populations in

pharmacogenetic research

Authors:

Rachel Huddart[1,8], Alison E. Fohner[1,2,8], Michelle Whirl-Carrillo[1,8], Genevieve L. Wojcik[1],

Christopher R. Gignoux[3], Alice B. Popejoy[1,4], Carlos D. Bustamante[1,5], Russ B. Altman[1,5,6,7] ,

Teri E. Klein[1,7]*

Affiliations:

[1] Department of Biomedical Data Science, Stanford University, Stanford, CA 94305, USA

[2] Department of Epidemiology, University of Washington, Seattle, WA 98195, USA

[3] Division of Bioinformatics and Personalized Medicine, and Department of Biostatistics,

University of Colorado, Anschutz Medical Campus, Aurora, CO 80045, USA

[4] Stanford Center for Integration of Research on Genetics and Ethics, Stanford, CA 94305, USA

[5] Department of Genetics, Stanford University, Stanford, CA 94305, USA

[6] Department of Biomedical Engineering, Stanford University, Stanford, CA 94305, USA

[7] Department of Medicine, Stanford University, Stanford, CA 94305, USA

[8] These authors contributed equally to this work

* **Correspondence:** Dr. Teri E. Klein, Shriram Center for BioE & ChemE, 443 Via Ortega,

Stanford, CA 94305-4125, Phone: (650) 736-0156, feedback@pharmgkb.org

Abstract:

The frequencies of pharmacogenetic alleles vary considerably between populations which has

important implications for the impact of these alleles in different populations. However, current

population grouping methods to communicate these patterns are insufficient as they are

inconsistent and fail to reflect the distribution of genetic variability around the world. To

facilitate and standardize the reporting of global variability in pharmacogenetic allele

frequencies, we present nine broad biogeographical groups, defined by global autosomal genetic

structure. These groups are based on data from large-scale initiatives, including the 1000

Genomes Project and the Human Genome Diversity Project, and reflect population genetic

history, genetic distances between populations, and geographical proximity, with borders falling

predominantly along national boundaries to simplify application of the grouping system. The

geographically-defined groups are American, Central/South Asian, East Asian, European, Near

Eastern, Oceanian, and Sub-Saharan African. Because of their distinct genetic patterns and

frequent inclusion in published data, we also present two admixed groups: African

American/Afro-Caribbean and Latino. Here, we characterize this population grouping system in

the context of broad genomic data and present its utility for annotating pharmacogenetic studies

and alleles. We recognize that broadly grouping global populations is an oversimplification of

human diversity, does not capture complex social and cultural identity, and uses arbitrary

geographic borders. However, this grouping method is consistent with robust population genetic

patterns and meets a need of the pharmacogenetics field by enabling consistent communication

of the scale of variability in global allele frequencies.

Report Text:

Interindividual variability in pharmacogenes has important consequences for drug

efficacy and toxicity.[1; 2] Unlike the low frequencies of alleles that are considered actionable with

respect to disease risk, pharmacogenetic variants with clinical relevance are common and, in fact,

both presence and absence of variants provide valuable dosing information.[3; 4] The frequencies

of many pharmacogenetic alleles vary greatly by global population, meaning that people with

different ancestries can have considerably different likelihoods of carrying an allele that is

associated with a particular drug response. For example, the CYP3A5*3 allele has been found at

a frequency of 98% in an Iranian population but at 11% in a Ngoni population from Malawi. [5; 6]

A single value for global allele frequency would fail to reflect this pattern. Presenting the

differences in frequencies of pharmacogenetic alleles is important for communicating the scale

of their expected impact on drug response and the degree of variation between populations. This

information is invaluable for furthering pharmacogenetic research and implementation.

Many pharmacogenetic studies present allelic data for very specific populations, such as

from a single country or ethnic group, which are difficult to incorporate into broader research or

implementation. Literature curation and gene summaries, such as those from the

Pharmacogenomics Knowledgebase (PharmGKB: www.pharmgkb.org), must group these

specific populations when annotating pharmacogenetic studies to allow users to easily compare

information from multiple studies. As such, tagging studies with population group identifiers is

an important component of knowledge extraction from curated literature. These population group

labels then are used in aggregating and evaluating overall evidence for gene-drug associations,

which eventually inform clinical implementation guidelines, such as those of the Clinical

Pharmacogenetics Implementation Consortium (CPIC: www.cpicpgx.org).

Current methods for grouping global populations in pharmacogenetics are based on

subjective, vague, and inconsistent geographical boundaries, or on populations that are

geographically straightforward to cluster and reflect little admixture.[7-11] As an example of the

issues with current grouping methods, some studies cluster participants of Egyptian descent with

African populations, while others cluster them with Middle Eastern populations.[12; 13] While this

discrepancy illustrates inconsistencies of geographic borders, the clustering of African-descent

populations of the Americas with populations from Africa, as seen in the 1000 Genomes African

(AFR) superpopulation, provides another example of the challenges of employing a small

number of categories. The genetic patterns seen in American populations with African ancestry

differs dramatically from populations in Africa due to admixture primarily with European and

American Indian populations. [14-16] While sharing common ancestry, the recent admixture

typically observed in the Americas can complicate average allele frequency estimation or, at a

minimum, make these combined groupings less homogeneous.[15] These insufficient grouping

systems, often ad-hoc and not fully representative of what we have learned through population

genomic studies, create a barrier to understanding and interpreting pharmacogenetic allele

frequencies in a globally representative fashion.

Up until July 2018, PharmGKB annotated studies using the five race categories defined

by the US Office of Management and Budget (OMB): White, Black or African American,

American Indian or Alaska Native, Asian, and Native Hawaiian or Pacific Islander, with an

additional ethnicity OMB category of Hispanic/Latino. While PharmGKB serves as a global

resource, these OMB groups are US-centric and lack the capacity to truly capture the scale of

global human diversity. We also investigated the utility of the biogeographic categories

employed by the Human Genome Diversity Panel - Centre d'Etude du Polymophisme Humain

(HGDP - CEPH), which groups its 52 populations into Africa, Europe, Middle East, South and

Central Asia, East Asia, Oceania and the Americas.[7; 17; 18] These population labels work well for

the populations included in the HGDP data set, which are not located in ambiguous regions

between group borders and which mostly contain populations with little admixture. However,

papers curated at PharmGKB can include populations located all over the world, including in the

transitional zones between HGDP geographical regions and admixed populations. This leads to

ambiguity in how such populations would be grouped using HGDP categories. In conclusion,

existing systems are insufficient for capturing the diversity of study populations in a replicable

manner that is consistent with patterns of human genetic variation.

Therefore, we sought to define a grouping system of global populations that could be

used consistently to annotate pharmacogenetic studies and relevant alleles, and could capture

global human population genetic patterns. Using population genetics data sources, including the

1000 Genomes Phase 3 data release and the HGDP, we propose a simple and robust grouping

pattern based on nine broad biogeographic regions that represent major geographic regions of the

world (**Figure 1**). It is important to note that classifying individuals and communities into a few

distinct groups with defined boundaries conflicts with our understanding of human variation,

history, and social/cultural identities. *As a result, we respectfully present these groups as a tool*

to represent broad differences in frequencies of genetic variation rather than as a classification

of human diversity.

We chose this geographic clustering pattern because geography has historically been the

greatest predictor of genetic variation between human populations, with genetic distance

increasing as geographic distance increases.[19] This geographic pattern aids consistency in

population groupings by setting boundaries along national borders. We have also included two

admixed groups due to their distinct genetic patterns compared to the geographically-defined

groups and because they are relatively frequently described in pharmacogenetic cohorts. Nine

groups were selected to be enough to illustrate the broad diversity in global allele frequencies,

yet small enough to apply easily and to be tractable in grouping specific populations.[20-23] We

then used autosomal genotype data to validate the ability of these groups to capture broad genetic

variation and to optimally reflect global patterns. The groups are given below with their

abbreviations.

Geographical populations:

To simplify utility, geographic boundaries between groupings are drawn predominantly

along country borders, with only Russia divided into east and west along the Ural Mountains

boundary due to the large size and genetic heterogeneity of the country. We intend these groups

to represent peoples with a predominance of ancestors who were in the region pre-Diaspora and

pre-colonization.

American (AME): The American genetic ancestry group includes all Native, Indigenous, and

Aboriginal people from both North and South America, including American Indian, Alaska

Native, First Nations, Inuit, and Métis in Canada, and Indigenous peoples of Central and South

America.

Central/South Asian (SAS): The Central and South Asian genetic ancestry group includes

populations from Pakistan, Sri Lanka, Bangladesh, India, and ranges from Afghanistan to the

western border of China.

East Asian (EAS): The East Asian genetic ancestry group includes Japan, Korea, and China, and

stretches from mainland Southeast Asia through the islands of Southeast Asia. In addition, it

includes portions of central Asia and Russia east of the Ural Mountains.

European (EUR): The European genetic ancestry group includes populations of European

descent, including European Americans. We define the European region as extending west from

the Ural Mountains and south to the Turkish and Bulgarian border.

Near Eastern (NEA): The Near Eastern genetic ancestry group encompasses northern Africa, the

Middle East, and the Caucasus. It includes Turkey and African nations north of the Saharan

Desert.

Oceanian (OCE): The Oceanian genetic ancestry group includes the indigenous populations of

the Pacific Islands, including Hawaii, Australia, and Papua New Guinea.

Sub-Saharan African (SSA): The Sub-Saharan African genetic ancestry group includes

individuals from all nations of Sub-Saharan Africa, including Madagascar.[24]

Admixed populations:

We include two admixed groups representing populations with recent mixing of geographically-

based populations and therefore, have distinct genetic patterns which are not adequately reflected

by any single geographically-based group. While many populations reflect a degree of

admixture, we selected these two populations because they are frequently reported in

pharmacogenetic studies.

African American/Afro-Caribbean (AAC): Individuals in the African American/Afro-Caribbean

genetic ancestry group reflect the extensive admixture between African, European, and

Indigenous ancestries[25] and, as such, display a unique genetic profile compared to individuals

from each of those lineages alone. Examples within this cluster include the Coriell Institute's

African Caribbean in Barbados (ACB) population and the African Americans from the

Southwest US (ASW) population, [26] and individuals from Jamaica and the US Virgin Islands.

Latino (LAT): The Latino genetic ancestry group is not defined by an exclusive geographic

region, but includes individuals of Mestizo descent, individuals from Latin America, and self-

identified Latino individuals in the United States. Like the African American/Afro-Caribbean

group, the admixture in this population creates a unique genetic pattern compared to any of the

discrete geographic regions, with individuals reflecting mixed Native and Indigenous American,

European, and African ancestry.

The Central/South Asian, East Asian and European groups presented here are equivalent

to the 1000 Genomes South Asian (SAS), East Asian (EAS) and European (EUR) super

populations, respectively. As such, we have adopted the relevant 1000 Genomes super

population codes as abbreviations for each of these groups to maintain consistency. While the

1000 Genomes Ad Mixed American (AMR) super population shows complete overlap with the

Latino group, we have opted to use the abbreviation LAT for this group. This removes the

potential for confusion between the Latino group and the other admixed group of African

American/Afro-Caribbean.

Figure 1 illustrates the countries included in each group and removes any ambiguity of

the group boundaries. We intend this map to be used as a guide for grouping populations. Study

subjects of an ancestry that is not within the geographic cluster in which they currently live will

be included in the geographic cluster reflecting their ancestry. For example, South Africans of

Dutch descent would be included in the European cluster rather than the Sub-Saharan African

cluster. However, when lacking a clear description otherwise, the population will be included in

the group that includes its home country. This approach highlights the importance of

understanding and recording detailed self-identity and self-reported ethnicity in the context of

genetic studies.

To validate the genetic variability distinguished by these population groups, we

conducted Principal Components Analysis (PCA) of unrelated individuals from 1000 Genomes.

As seen in **Figure 2A**, the first two principal components (PCs) separate populations by

geographic region, especially along continental boundaries, and illustrate the increasing genetic

distance between populations of increasing geographic distance. As can be seen in the

overlapping PC distribution of individuals of different population groups, human genetic

diversity is a spectrum,[18] and therefore the geographic boundaries of these groups should be

understood as an obligatory divide to create relevant groupings, with the acknowledgement that

these borders are constrained by modern country borders and therefore are inherently arbitrary in

geographic space.[18] However, as shown in **Figure 2B**, only a few PCs are needed to accurately

predict these population clusters. Even with only 4 PCs, the minimum area under the curve

(AUC) for correct cluster prediction is 97.9% for most populations using multiple logistic

regression. The only outlier is the African American/Afro-Caribbean cluster, consistent with

ancestral similarity to the African cluster.[14; 27] Here still, with a larger number of PCs, the AUC

is above 93%, even with the observed ancestry outliers present in the 1000 Genomes African

Americans in the Southwest US (ASW) population.[28] While no categorization will result in

perfect prediction, given the spectrum of human diversity, the statistical validation of this

clustering from broad autosomal data makes these clusters both relevant and useful for

PharmGKB.

In **Figure 3**, we demonstrate that the groups we have selected are effective for

representing the diversity of global allele frequencies in pharmacogenes. We present here the

frequency of four single nucleotide polymorphisms (SNPs) with important pharmacogenetic

implications. The 'A' allele of rs1065852 is the defining SNP of the *cytochrome P450 2D6*

*(CYP2D6) *10* haplotype and is also found in combination with other variants in multiple

CYP2D6 haplotypes. Haplotypes containing this SNP are associated with decreased CYP2D6

activity, which has important implications for drugs that are CYP2D6 substrates, including

codeine, selective serotonin reuptake inhibitors, ondansetron, and tricyclic antidepressants.[29-31]

The *CYP2C9* alleles *2* (defined by rs1799853), *3* (defined by rs1057910), and *8* (defined by

rs7900194) are associated with reduced enzyme function and therefore are associated with

recommended changes to the dosing of warfarin and phenytoin, which are substrates of

CYP2C9.[32; 33] Using data from the 1000 Genomes, we show the frequency of the four SNPs in

these biogeographic groups. The range of frequencies globally illustrates the importance of

showing allele frequency by group in order to convey its impact on drug response globally.

The SNP rs1065852 shows stark continental patterns (**Figure 3A**). The 'A' allele is found

at high frequencies within East Asian populations, ranging from 66.2% in Vietnam (KHV) to

36.1% in Japan (JPT). This allele is less frequent in other continental populations, such as Africa

(3.5-16.5%), Europe (14.6-24.7%), and South Asia (10.4-25.6%). As can be seen from the range

of frequencies of the three *CYP2C9* alleles, the most common reduced function allele varies

globally, with the *8* allele much more common in African populations (1.8-7.6%) than the *2*

(<1%) or *3* (monomorphic in Africa) (**Figure 3B-D**). Conversely, the *8* allele is rare in

European populations (<1%), while *2* (8.1-15.2%) and *3* (5.6-8.4%) are more common.

Patterns such as this one can result in bias in the utility of dosing algorithms, such as the

International Warfarin Pharmacogenetics Consortium (IWPC) dosing algorithm for warfarin,

which adjusts dose based on the presence of the *2 and *3 alleles but does not include the *8

allele.[34]

While individual pharmacogenetic testing (either pre-emptive or at point-of-care) remains

the most effective and appropriate way to implement pharmacogenetic knowledge for the care of

an individual,[35; 36] we recognize the need in clinical and genetic research for a standardized

method to broadly group populations based on biogeographic region. For example, identifying

populations with high frequencies of certain pharmacogenetic alleles can help to direct targeted

screening when resources are constrained and inform priorities for future pharmacogenetic

research.[19] However, the groups we present are large and the summary information presented

should be understood as an approximation dependent on existing studies in that region, which

may be limited to a few locations. As such, these groups are not suitable for use in guiding

specific implementation programs; rather, they should be seen as a tool for research purposes.

It should be noted that this grouping system does have some limitations. Classifying

individuals into these population groups can be complicated by social and cultural identities,

especially for admixed individuals,[7; 9; 37-39] and membership of an individual within one of these

population groups is inherently an imperfect surrogate for predicting the likelihood that the

individual carries a particular genetic variant.[36; 40] As can be seen in the analysis of rs1065852

above, the frequency of the 'A' allele can vary by up to 30% between populations which are all

included in the East Asian group. Furthermore, while the grouping system is based on overall

genome-wide average patterns, which typically follow a clinal variation pattern correlated with

geographic proximity,[7; 22; 23; 41; 42] variation in individual genes or individual populations do not

always follow these gradual patterns.[8-11; 36]

Because aggregate annotations of pharmacogenetic research and summary allele

frequencies are based only on available studies, additional studies are needed that include a

greater diversity of populations to make pharmacogenetic research and allele frequency

summaries more representative.[43] For example, the Sub-Saharan Africa (SSA) grouping

represents a large swath of human genomic diversity, and increased representation of these

populations in pharmacogenetics studies may lead to the discovery of clinical differences within

the larger grouping. Furthermore, large, reference genetic studies with targeted allele

information, like that emerging from the Population Architecture using Genomics and

Epidemiology (PAGE) study (www.pagestudy.org), may provide compelling evidence to adjust

these group boundaries based on frequency patterns specific to pharmacogenetic alleles.

Despite these limitations, broad population groups are needed for illustrating global

diversity with respect to pharmacogenetic variation and the average predicted phenotypes in

populations. These nine proposed biogeographic groups provide a consistent way to present

these data based on a system that is grounded in robust data on population genetic patterns.

PharmGKB has begun to use these population groups in curation activities, and we recommend

that these groups and accompanying map be considered the standard grouping mechanism for

population pharmacogenetics. Ultimately, individual pharmacogenetic testing of all patients,

regardless of ancestry, is needed to deliver truly personalized medicine. However, the population

groups we present are useful for the standardized presentation of pharmacogenetic studies, global

allele frequency summaries in pharmacogenetic research and broad clinical screening.

Acknowledgments:

This work was funded by NIH/NIGMS R24 GM61374 and NIH/NHGRI U01 HG007419-04.

Conflicts of Interest:

274   CRG owns stock in 23andMe, Inc and is a founder of and advisor to Encompass Bioscience, Inc.

275   CDB is a member of the scientific advisory boards for Liberty Biosecurity, Personalis, 23andMe

276   Roots into the Future, Ancestry.com, IdentifyGenomics, and Etalon and is a founder of CDB

277   Consulting. RBA is a stockholder in Personalis Inc. and 23andMe, and a paid advisor for

278   Youscript. Remaining authors have no conflicts of interest.

References:

1. Roden, D.M. (2006). PHarmacogenomics: Challenges and Opportunities. Annals of Internal
Medicine 145.
2. Dunnenberger, H.M., Crews, K.R., Hoffman, J.M., Caudle, K.E., Broeckel, U., Howard, S.C.,
Hunkler, R.J., Klein, T.E., Evans, W.E., and Relling, M.V. (2015). Preemptive clinical
pharmacogenetics implementation: current programs in five US medical centers. Annu
Rev Pharmacol Toxicol 55, 89-106.
3. Tabor, H.K., Auer, P.L., Jamal, S.M., Chong, J.X., Yu, J.H., Gordon, A.S., Graubert, T.A.,
O'Donnell, C.J., Rich, S.S., Nickerson, D.A., et al. (2014). Pathogenic variants for
Mendelian and complex traits in exomes of 6,517 European and African Americans:
implications for the return of incidental results. Am J Hum Genet 95, 183-193.
4. Wright, G.E.B., Carleton, B., Hayden, M.R., and Ross, C.J.D. (2018). The global spectrum of
protein-coding pharmacogenomic diversity. The pharmacogenomics journal 18, 187-195.
5. Rahsaz, M., Azarpira, N., Nikeghbalian, S., Aghdaie, M.H., Geramizadeh, B., Moini, M.,
Banihashemi, M., Darai, M., Malekpour, Z., and Malekhosseini, S.A. (2012). Association
between tacrolimus concentration and genetic polymorphisms of CYP3A5 and ABCB1
during the early stage after liver transplant in an Iranian population. Experimental and
clinical transplantation : official journal of the Middle East Society for Organ
Transplantation 10, 24-29.
6. Bains, R.K., Kovacevic, M., Plaster, C.A., Tarekegn, A., Bekele, E., Bradman, N.N., and
Thomas, M.G. (2013). Molecular diversity and population structure at the Cytochrome
P450 3A5 gene in Africa. BMC genetics 14, 34.
7. Rosenberg, N.A., Pritchard, J.K., Weber, J.L., Cann, H.M., Kidd, K.K., Zhivotovsky, L.A., and
Feldman, M.W. (2002). Genetic structure of human populations. Science (New York,
NY) 298, 2381-2385.
8. Rajagopalan, R., and Fujimura, J.H. (2012). Will personalized medicine challenge or reify
categories of race and ethnicity? The virtual mentor : VM 14, 657-663.
9. Gannett, L. (2005). Group Categories in Pharmacogenetics Research. Philosophy of Science
72, 1232-1247.
10. Wilson, J.F., Weale, M.E., Smith, A.C., Gratrix, F., Fletcher, B., Thomas, M.G., Bradman, N.,
and Goldstein, D.B. (2001). Population genetic structure of variable drug response. Nat
Genet 29, 265-269.

11. Race, E., and Genetics Working Group. (2005). The Use of Racial, Ethnic, and Ancestral
Categories in Human Genetics Research. Am J Hum Genet 77, 519-532.
12. Relling, M.V., Gardner, E.E., Sandborn, W.J., Schmiegelow, K., Pui, C.H., Yee, S.W., Stein,
C.M., Carrillo, M., Evans, W.E., and Klein, T.E. (2011). Clinical Pharmacogenetics
Implementation Consortium guidelines for thiopurine methyltransferase genotype and
thiopurine dosing. Clin Pharmacol Ther 89, 387-391.
13. Scott, S.A., Sangkuhl, K., Stein, C.M., Hulot, J.S., Mega, J.L., Roden, D.M., Klein, T.E.,
Sabatine, M.S., Johnson, J.A., and Shuldiner, A.R. (2013). Clinical Pharmacogenetics
Implementation Consortium guidelines for CYP2C19 genotype and clopidogrel therapy:
2013 update. Clin Pharmacol Ther 94, 317-323.
14. Bryc, K., Auton, A., Nelson, M.R., Oksenberg, J.R., Hauser, S.L., Williams, S., Froment, A.,
Bodo, J.M., Wambebe, C., Tishkoff, S.A., et al. (2010). Genome-wide patterns of
population structure and admixture in West Africans and African Americans.
Proceedings of the National Academy of Sciences of the United States of America 107,
786-791.
15. Mathias, R.A., Taub, M.A., Gignoux, C.R., Fu, W., Musharoff, S., O'Connor, T.D., Vergara,
C., Torgerson, D.G., Pino-Yanes, M., Shringarpure, S.S., et al. (2016). A continuum of
admixture in the Western Hemisphere revealed by the African Diaspora genome. Nature
communications 7, 12522.
16. Martin, A.R., Gignoux, C.R., Walters, R.K., Wojcik, G.L., Neale, B.M., Gravel, S., Daly, M.J.,
Bustamante, C.D., and Kenny, E.E. (2017). Human Demographic History Impacts
Genetic Risk Prediction across Diverse Populations. Am J Hum Genet 100, 635-649.
17. Cann, H.M., de Toma, C., Cazes, L., Legrand, M.F., Morel, V., Piouffre, L., Bodmer, J.,
Bodmer, W.F., Bonne-Tamir, B., Cambon-Thomsen, A., et al. (2002). A human genome
diversity cell line panel. Science (New York, NY) 296, 261-262.
18. Rosenberg, N.A., Mahajan, S., Ramachandran, S., Zhao, C., Pritchard, J.K., and Feldman,
M.W. (2005). Clines, clusters, and the effect of study design on the inference of human
population structure. PLoS Genet 1, e70.
19. Burchard, E.G., Ziv, E., Coyle, N., Gomez, S.L., Tang, H., Karter, A.J., Mountain, J.L.,
Perez-Stable, E.J., Sheppard, D., and Risch, N. (2003). The importance of race and
ethnic background in biomedical research and clinical practice. The New England journal
of medicine 348, 1170-1175.
20. Auton, A., Brooks, L.D., Durbin, R.M., Garrison, E.P., Kang, H.M., Korbel, J.O., Marchini,
J.L., McCarthy, S., McVean, G.A., and Abecasis, G.R. (2015). A global reference for
human genetic variation. Nature 526, 68-74.
21. Elhaik, E., Tatarinova, T., Chebotarev, D., Piras, I.S., Maria Calo, C., De Montis, A., Atzori,
M., Marini, M., Tofanelli, S., Francalacci, P., et al. (2014). Geographic population
structure analysis of worldwide human populations infers their biogeographical origins.
Nature communications 5, 3513.
22. Jakobsson, M., Scholz, S.W., Scheet, P., Gibbs, J.R., VanLiere, J.M., Fung, H.C., Szpiech,
Z.A., Degnan, J.H., Wang, K., Guerreiro, R., et al. (2008). Genotype, haplotype and
copy-number variation in worldwide human populations. Nature 451, 998-1003.
23. Li, J.Z., Absher, D.M., Tang, H., Southwick, A.M., Casto, A.M., Ramachandran, S., Cann,
H.M., Barsh, G.S., Feldman, M., Cavalli-Sforza, L.L., et al. (2008). Worldwide human
relationships inferred from genome-wide patterns of variation. Science (New York, NY)
319, 1100-1104.
24. Hurles, M.E., Sykes, B.C., Jobling, M.A., and Forster, P. (2005). The dual origin of the
Malagasy in Island Southeast Asia and East Africa: evidence from maternal and paternal
lineages. Am J Hum Genet 76, 894-901.

25. Maples, B.K., Gravel, S., Kenny, E.E., and Bustamante, C.D. (2013). RFMix: a
discriminative modeling approach for rapid and robust local-ancestry inference. Am J
Hum Genet 93, 278-288.
26. Genomes Project, C., Abecasis, G.R., Auton, A., Brooks, L.D., DePristo, M.A., Durbin, R.M.,
Handsaker, R.E., Kang, H.M., Marth, G.T., and McVean, G.A. (2012). An integrated map
of genetic variation from 1,092 human genomes. Nature 491, 56-65.
27. Baharian, S., Barakatt, M., Gignoux, C.R., Shringarpure, S., Errington, J., Blot, W.J.,
Bustamante, C.D., Kenny, E.E., Williams, S.M., Aldrich, M.C., et al. (2016). The Great
Migration and African-American Genomic Diversity. PLoS Genet 12, e1006059.
28. Mimno, D., Blei, D.M., and Engelhardt, B.E. (2015). Posterior predictive checks to quantify
lack-of-fit in admixture models of latent population structure. Proceedings of the National
Academy of Sciences of the United States of America 112, E3441-3450.
29. Bell, G.C., Caudle, K.E., Whirl-Carrillo, M., Gordon, R.J., Hikino, H., Prows, C.A., Gaedigk,
A., Agundez, J., Sadhasivam, S., Klein, T.E., et al. (2016). Clinical Pharmacogenetics
Implementation Consortium (CPIC) guideline for CYP2D6 genotype and use of
ondansetron and tropisetron. Clin Pharmacol Ther.
30. Hicks, J.K., Sangkuhl, K., Swen, J.J., Ellingrod, V.L., Muller, D.J., Shimoda, K., Bishop, J.R.,
Kharasch, E.D., Skaar, T.C., Gaedigk, A., et al. (2016). Clinical pharmacogenetics
implementation consortium guideline (CPIC) for CYP2D6 and CYP2C19 genotypes and
dosing of tricyclic antidepressants: 2016 update. Clin Pharmacol Ther.
31. Hicks, J.K., Bishop, J.R., Sangkuhl, K., Muller, D.J., Ji, Y., Leckband, S.G., Leeder, J.S.,
Graham, R.L., Chiulli, D.L., A, L.L., et al. (2015). Clinical Pharmacogenetics
Implementation Consortium (CPIC) Guideline for CYP2D6 and CYP2C19 Genotypes
and Dosing of Selective Serotonin Reuptake Inhibitors. Clin Pharmacol Ther 98, 127-
134.
32. Caudle, K.E., Rettie, A.E., Whirl-Carrillo, M., Smith, L.H., Mintzer, S., Lee, M.T., Klein, T.E.,
Callaghan, J.T., and Clinical Pharmacogenetics Implementation, C. (2014). Clinical
pharmacogenetics implementation consortium guidelines for CYP2C9 and HLA-B
genotypes and phenytoin dosing. Clin Pharmacol Ther 96, 542-548.
33. Johnson, J.A., Caudle, K.E., Gong, L., Whirl-Carrillo, M., Stein, C.M., Scott, S.A., Lee, M.T.,
Gage, B.F., Kimmel, S.E., Perera, M.A., et al. (2017). Clinical Pharmacogenetics
Implementation Consortium (CPIC) Guideline for Pharmacogenetics-Guided Warfarin
Dosing: 2017 Update. Clin Pharmacol Ther.
34. International Warfarin Pharmacogenetics, C., Klein, T.E., Altman, R.B., Eriksson, N., Gage,
B.F., Kimmel, S.E., Lee, M.T., Limdi, N.A., Page, D., Roden, D.M., et al. (2009).
Estimation of the warfarin dose with clinical and pharmacogenetic data. The New
England journal of medicine 360, 753-764.
35. Foster, M.W., Sharp, R.R., and Mulvihill, J.J. (2001). Pharmacogenetics, Race, and
Ethnicity: Social Identities and Individualized Medical Care. Therapeutic Drug Monitoring
23, 232-238.
36. Yen-Revollo, J.L., Auman, J.T., and McLeod, H.L. (2008). Race does not explain genetic
heterogeneity in pharmacogenomic pathways. Pharmacogenomics 9, 1639-1645.
37. Braun, L., Fausto-Sterling, A., Fullwiley, D., Hammonds, E.M., Nelson, A., Quivers, W.,
Reverby, S.M., and Shields, A.E. (2007). Racial categories in medical practice: how
useful are they? PLoS medicine 4, e271.
38. Ortega, V.E., and Meyers, D.A. (2014). Pharmacogenetics: implications of race and ethnicity
on defining genetic profiles for personalized medicine. J Allergy Clin Immunol 133, 16-
26.
39. Bamshad, M., Wooding, S., Salisbury, B.A., and Stephens, J.C. (2004). Deconstructing the
relationship between genetics and race. Nat Rev Genet 5, 598-609.

40. Urban, T.J. (2010). Race, ethnicity, ancestry, and pharmacogenetics. Mt Sinai J Med 77,
133-139.
41. Risch, N., Burchard, E., Ziv, E., and Tang, H. (2002). Categorization of humans in
biomedical research: genes, race and disease. Genome Biology 3.
42. Bamshad, M.J., Wooding, S., Watkins, W.S., Ostler, C.T., Batzer, M.A., and Jorde, L.B.
(2003). Human population genetic structure and inference of group membership. Am J
Hum Genet 72, 578-589.
43. Bustamante, C.D., Burchard, E.G., and De la Vega, F.M. (2011). Genomics for the world.
Nature 475, 163-165.
420
421
422

Figures:

Figure 1: Map of geographical boundaries included in each genetic ancestry group. The

biogeographical groups are color-coded and the group boundaries fall predominantly along

national boundaries to aid the assignment of group membership.

Figure 2: Principal component analysis comparing genetic distances of populations with

close geographic proximity using 1000 Genomes participants. (A) The genetic gradient

between populations is illustrated along PCs 1 vs 2 and PCs 3 vs 4, showing that, while

completely discrete population boundaries are challenging, the groupings proposed here provide

a statistically robust grouping. (B) AUCs of logistic regression to predict cluster membership,

showing high degree of population structure.   Note that, because none of the 1000 Genomes

populations fall into the American (AME) group, no reference data were available to include this

group in the analysis.

Figure 3: **Maps illustrating how the proposed biogeographical grouping system can be used to illustrate the variability in**

global allele frequencies.