Effects of Ocean Climate on Spatiotemporal Variation in Sea Urchin Settlement and Recruitment

Daniel K. Okamoto[*1,2], Stephen Schroeter[†3], and Daniel C. Reed[‡3]

[1]Department of Biological Science, Florida State University, Tallahassee, Florida 32303
[2]Ecology, Evolution and Marine Biology, University of California Santa Barbara, Santa Barbara, CA 93106 USA
[3]Marine Science Institute, University of California Santa Barbara, Santa Barbara, CA 93106 USA

[*]corresponding author: dokamoto@bio.fsu.edu

[†]email: schroete@lifesci.ucsb.edu

[‡]email: dan.reed@lifesci.ucsb.edu

Abstract

Sea urchins are voracious herbivores that influence the ecological structure and function

of nearshore ecosystems throughout the world. Like many species that produce planktonic

larvae, their recruitment is thought to be particularly sensitive to climatic fluctuations in

temperature that directly or indirectly affect adult reproduction and larval transport and survival.

Yet how climate alters sea urchin populations in space and time by modifying larval recruitment

and year-class strength on the time-scales that regulate populations remains understudied. Using

a, spatially replicated weekly-biweekly dataset spanning 27 years and 1100 km of coastline, we

characterized seasonal, interannual, and spatial patterns of larval settlement of the purple sea

urchin (*Strongylocentrotus purpuratus*). We show that large spatial differences in temporal

patterns of larval settlement were associated with different responses to fluctuations in ocean

temperature and climate. Importantly, we found a strong correlation between larval settlement

and regional year class strength suggesting that such temporal and spatial variation in settlement

plays an important role in controlling population dynamics. These results provide strong

evidence over extensive temporal and spatial domains that climatic fluctuations shape broad-

scale patterns of larval settlement and subsequent population structure of an important marine

herbivore known to control the productivity, community state and provisioning services of

marine ecosystems.

Introduction

Large scale climate oscillations (e.g., El Niño/Southern Oscillation, North Atlantic Oscillation) lead to changes in ocean temperature, biogeochemistry and the severity and frequency of disruptive events that affect ocean circulation, upwelling and primary productivity (2014; Mantua et al., 1997). Such shifts impose wide-reaching ecological impacts, in part by altering animal recruitment and food web structure in space and time (Sydeman et al., 2015). Hence, understanding how climate variability alters the recruitment of marine species is particularly important for effective conservation and management of the ocean's resources. Climatic fluctuations give rise to shifts in numerous factors that shape both adult reproduction and larval supply, including primary productivity, temperature, and advection and transport. Given these multiple direct and indirect effects of climate on recruitment, significant challenges remain in achieving such understanding for benthic species with planktonic larvae due to the substantial effort needed to characterize spatial and temporal variation in larval settlement and the numerous sensitive vital rates that contribute to it.

For benthic species like sea urchins, understanding causes and consequences of recruitment variability has both ecological and economic implications. Sea urchin grazing can alter the structure of some of the world's most diverse and productive marine ecosystems, including coral reefs (Edmunds & Carpenter, 2001), seagrass meadows (reviewed by Valentine & Heck Jr, 1999) and kelp forests (reviewed by Filbee-Dexter & Scheibling, 2014). In addition, sea urchins form the basis of important nearshore fisheries in many regions of the world (e.g. Andrew et al., 2003; Kato & Schroeter, 1985). As a result, climate-driven changes in sea urchin populations have the potential to profoundly affect the ecological structure and functioning of

marine ecosystems and the economic value of the fisheries that they support. Much of the

research on controls of sea urchin population dynamics has focused on the roles of predation and

disease in controlling adult abundance and their cascading influence on community structure

(e.g. Burt et al., 2018; Estes & Duggins, 1995; Filbee-Dexter & Scheibling, 2014; Lafferty,

2004). Yet short-term empirical studies (months to a few years) suggest that environmentally

regulated larval supply is likely an important driver of adult urchin dynamics (Hernández et al.,

2010; Ling et al., 2009). Despite the widespread recognition of the importance of recruitment

variation in controlling population fluctuations in many marine species (Shelton & Mangel,

2011), relatively few studies have examined the biotic and abiotic processes controlling the

supply of sea urchin larvae in nature (but see Hernández et al., 2010; Ling et al., 2009), and the

degree to which they affect the abundance and dynamics of older life stages over time scales that

impact population dynamics.

Fluctuations in climate can affect spatial and temporal patterns of larval supply by

influencing the production of larvae by benthic adults, transport of larvae to adjacent habitats,

and the survival of larvae in the plankton. Increases in ocean temperature can impact larval

production by: (1) increasing adult mortality via the spread of water-borne pathogens (reviewed

by Feehan & Scheibling, 2014), and (2) reducing adult fecundity and inhibiting gametogenesis

by altering food quantity and quality (Basch & Tegner, 2007; Cochran & Engelmann, 1975;

Foster et al., 2015; Okamoto, 2014). Because sea urchins produce feeding larvae that spend

weeks to months in the plankton, increases in ocean temperature can also affect larval

development, growth and survival, either directly, or indirectly by altering the availability of

their phytoplankton food source (Bertram & Strathmann, 1998; Byrne et al., 2009; Hoegh-

Guldberg & Pearse, 1995; Strathmann, 1987). Finally, climate related changes in patterns of

ocean circulation can affect the transport of larvae from source to destination (but see Morgan,

2014; Siegel et al., 2008). Thus, the effects of climatic change on sea urchin recruitment

represent cumulative impacts on adult abundance and reproduction, complex current patterns that

transport larvae, behavioral responses of larvae, and larval development and survival. Because

patterns of ocean temperature, circulation and upwelling can vary dramatically in space, the

effects of climate oscillations on sea urchin recruitment potentially vary over large spatial scales.

A dearth of long-term, high frequency, spatially extensive data has prevented characterizing

temporal and spatial variability in larval settlement in sea urchins, the degree to which it is

explained by different sources of environmental variation, and the relative importance of these

drivers in accounting for fluctuations in population size.

Here we analyzed a 27-year weekly to biweekly time series of the recruitment of newly

metamorphosed larvae (hereafter referred to as larval settlement) at sites distributed across 1100

km of coast in California to investigate sources of spatial and temporal variability in larval

settlement of the purple sea urchin *Strongylocentrotus purpuratus*. Our objectives were to: (1)

quantify variation in larval settlement across different temporal and spatial scales; (2) evaluate

whether larval settlement on artificial substrates predicts year-class strength in natural

populations; and (3) determine the relative importance of adult abundance, larval and adult food

supply, sea surface temperature and broad-scale fluctuations in ocean climate in contributing to

the observed variation in larval settlement

Study system

Populations of the purple sea urchin (*Strongylocentrotus purpuratus*) occupy shallow

subtidal and intertidal rocky substrata from at least 27°N on the western coast of the Baja

Peninsula (Olivares-Bañuelos et al., 2008) to at least 59°N on the Kenai Peninsula in Alaska

(Field & Walker, 2003). Purple urchins are broadcast spawners and the seasonality of their

spawning is generally thought to be driven by spring photoperiod and temperature (Cochran &

Engelmann, 1975; Gonor, 1973; Pearse et al., 1986). Energy available for gonad production is

largely determined by macroalgal food availability; specifically, sea urchins in barrens devoid of

abundant macroalgae can show dramatic reductions in fecundity (>99%), and resumption of

feeding in emaciated urchins can lead to gonadal recovery within 2-3 months (Okamoto, 2014).

Results from the field and laboratory indicate that the thermal upper limit to completion of

gametogenesis is about 17°C (Basch & Tegner, 2007; Cochran & Engelmann, 1975). Fertilized

zygotes develop into planktonic echinoplutei larvae that feed exclusively on phytoplankton

(Strathmann, 1987). After spending several weeks to months in the plankton, individuals begin

final metamorphosis and settle to the benthos (Strathmann, 1978) at a size of ~500 μm in

diameter (Okamoto unpublished data). Larval settlement varies dramatically among locations at

both small and large spatial scales (Ebert, 2010). Once settled, fully competent individuals

occupy cobble and other complex substrata and between 12-24 months become visible in benthic

surveys at 1-2 cm diameter. For a full review see Rogers-Bennet & Okamoto (Rogers-Bennett &

Okamoto, 2020).

Collection of newly settled urchins along the coast of California, USA (1990-2016)

Settlement of newly metamorphosed purple sea urchins was sampled in three major

regions along the California coast from 1990 through 2016, with a total of 54,588 replicate

observations. Sampling regions (from south to north) included two sites in San Diego County

(Scripps Pier and Ocean Beach Pier), four sites in the Santa Barbara Channel (Anacapa Island,

Stearns Wharf, Ellwood Pier and Gaviota Pier) and one site at Point Cabrillo in Fort Bragg

(Figure 1). San Diego and the Santa Barbara Channel lie within the Southern California Bight

and Fort Bragg is in northern California. At each site, newly settled urchins were collected using

nylon-bristled scrub brushes (2.5 cm long bristles and a 6 x 9 cm wooden base) suspended 1 to 2

m from the benthos (Ebert et al., 1994). The majority of deployments included 4-8 replicate

brushes collected weekly at each site from 1990 to 2003, and biweekly thereafter through 2016.

Upon collection brushes were transported to the laboratory in plastic bags and rinsed through a

350 μm mesh sieve. Purple urchins were sorted from other species, counted and preserved. A

more detailed description of the sampling methods, geographic and taxonomic coverage and

links to the data and metadata are provided in Schroeter et al. (2019)

Methods

Spatio-temporal trends in larval settlement

We estimated seasonal, annual, and spatial patterns in larval settlement using an

integrated spatiotemporal Bayesian model that accounted for the spatially and temporally

intercorrelated and heterogeneous nature of the multivariate time series. We used a Bayesian

state-space formulation for the model for several reasons. First, episodic periods of low

replication, low observation numbers, missing data, or slight variation in the sampling interval

mean that the true number of settlers may not be reflected in the empirical mean value from

brush data (i.e. the true value is not always observable). Second, when brushes contained

hundreds or more individual urchins, species identification consisted of subsampling urchins to

estimate proportions of individual species (in this case *S. purpuratus* versus the rarer red urchin,

Mesocentrotus franciscanus). This subsampling routine requires accounting for the uncertainty

in the sampled species ratios in the estimation of settler abundance, which we did by

incorporating a Bayesian prior in the form of a Beta distribution with hyperparameters as the

observed ratio and total subsample count (the Jeffrey's prior for ratios from binomial counts).

Third, initial examination of the data indicated substantial seasonal and interannual variability

that was multiplicative among-samples within each period, as well as serial autocorrelation and

spatial components that needed to be simultaneously accounted for.

We used a Poisson observation likelihood to link the biweekly mean estimate from the

trend equation ($\hat{\mu}_{t,l}$ - see Table 1, Equations 3 & 4, and Table 2) to observed counts of larval

settlers (N) for each brush (b) within each site within each year. To account for the above

described nuances of the data we estimated biweekly settlement density ($\hat{\mu}_{t,l}$) as the sum of a

site-specific mean ($\beta_{0,l}$), log annual trend ($\text{Annual\_Trend}_{s,t}$), log seasonal trend

($\text{Seas\_Trend}_{t,l}$), and a spatially correlated lognormal process error ($\varepsilon_{t,l}$);

Eq. 1 $\ln \hat{\mu}_{t,l} = \beta_{0,l} + \ln \text{Annual\_Trend}_{t,l} + \ln \text{Seasonal\_Trend}_{t,l} + \varepsilon_{t,l}$ (Trend Equation)

We estimated trends on the log-scale so that process errors, and seasonal and interannual trends

were multiplicative and strictly positive on the original scale.

The annual trends were assumed to be correlated in both space and time, where spatial

and temporal covariances were assumed to be independent. The annual spatial covariance was

assumed to be unstructured (i.e. no formal distance decay structure) because we only had seven

sites. The annual temporal covariance was determined by a Matérn 3/2 kernel because it

provided sufficient flexibility to capture interannual trends. The seasonal trend (log-scale)

within each site was estimated using a seasonal periodic temporal kernel within the model to

capture the cyclical nature of seasonality. The process error ($\varepsilon_{t,l}$) was modeled with an

unstructured spatial covariance to account for correlations among sites in their deviations from

model expectations. We estimated the model posteriors using Stan (Carpenter et al., 2016) with

three 1000 iteration chains after a 1000 iteration burn-in. Stan model code is provided in the

supplement. For model equations and symbology see Tables 1 and 2.

Relationship between benthic year-class strength and larval settlement.

If year class strength of sea urchins is limited by larval supply, then we should see

increases (decreases) in the abundance of juveniles in years following anomalously high (low)

larval settlement. We therefore tested whether larval settlement in the Santa Barbara Channel

(mean of Anacapa, Stearns Wharf, Ellwood Pier and Gaviota Pier) was predictive of subsequent

juvenile recruitment (hereafter referred to as "year-class strength") on natural reefs in the region.

We calculated densities of juveniles (<2cm test diameter) from the Channel Islands Kelp Forest

Monitoring (KFM) Program (Kushner et al., 2013), including only the 9 sites with time series

extending from 1990 through 2016. We examined the relationship between larval settlement and

year-class strength using a generalized linear mixed effects model (GLMM) with a negative

binomial likelihood (a Poisson likelihood indicated overdispersion) and survey site as a random

effect. We tested the hypothesis of no correlation using a likelihood ratio test comparing the

models that do versus do not include settlement as a covariate. Models were estimated using

glmmTMB (Magnusson et al., 2017) in R (see Electronic Supplement for further details).

Relationships between larval settlement and biotic and abiotic conditions

We estimated the strength of relationships between biweekly larval settlement and

various hypothesized physical and biological drivers using an integrated regression model that

included a regularized regression within a Bayesian time-series modelling framework (Fig. 2).

We used a Poisson observation likelihood to link estimated latent trends $\hat{\mu}_{t,l}$ to the settlement

count observations (see Table 1 Equations 3 & 4, and Table 2). We estimated the log biweekly

density ($\ln \hat{\mu}_{t,l}$) as a function of the centered and standardized covariates (denoted by the vector

$\boldsymbol{x}_{t,l}$) while directly estimating and accounting for seasonal trends (Seasonal_Trend$_{t,l}$), and a

separable spatially and temporally correlated, multivariate lognormal process error ($\varepsilon_{t,l}$):

Eq. 2 $\ln \hat{\mu}_{t,l} = \beta_{0,l} + [\boldsymbol{x}_{t,l}]' \boldsymbol{\beta}_l + \ln \text{Seasonal\_Trend}_{t,l} + \varepsilon_{t,l}$ (Regression Equation)

We assumed the temporal correlation in the process error $\varepsilon_{t,l}$ followed a first order

autoregressive model.

Because these analyses were correlative, we focused on unbiased parameter estimation

rather than hypothesis testing or model comparison per se. We estimated two separate models:

one included all locally measured (i.e. for a given site) environmental variables and the other

included a common set of composite indices of oceanographic climate ("global covariates" –

ENSO Index, Pacific Decadal Oscillation, North Pacific Gyre). Because inclusion of numerous

covariates can cause overfitting that leads to bias and uncertainty in the explanatory power of the

covariates, we erred on the side of sparsity and assigned the vector of regression coefficients

($\boldsymbol{\beta}_l$) a regularized horseshoe prior (i.e. the Finnish Horseshoe). Sparsity in this case assumed

that only a few of the covariates were meaningful without a priori knowledge of which

covariates were relevant and which were not. A sparse regression encoded this assumption to

allow the data to inform which covariates were relevant and how they were correlated with the

response variable. The Finnish horseshoe prior is a Bayesian version of the lasso (Carvalho et al.,

2010; Piironen & Vehtari, 2017) that produced a data-driven reduction in the influence of weaker

covariates by regularization of those coefficients given the data. For full details and model

equations, see Tables S.1 and S.2.

Below are the covariates used in the integrated regression model:

(i) Oceanographic climate indices (monthly, 1990-2016, all sites): We used three major

global indices of oceanographic climate. The multivariate El Niño Southern Oscillation Index

(MEI), the Pacific Decadal Oscillation (PDO - Mantua & Hare, 2002) and the North Pacific Gyre

Oscillation (NPGO - Di Lorenzo et al., 2008). The MEI provides a metric of the intensity of El

Niño Southern Oscillation (ENSO) fluctuations, which persist for 6-18 months and explains

much of the oceanographic variability in the tropics (Di Lorenzo et al., 2013; Wolter & Timlin,

1993; Wolter & Timlin, 1998). In contrast, the PDO and NPGO exhibit longer-period

fluctuations, explain much of the oceanographic variability in higher latitudes, and the low

frequency variability of these metrics are shaped through ENSO forcing of the Aleutian Low and

the North Pacific Oscillation, respectively (Di Lorenzo et al., 2013).

(ii) Coastal upwelling index (monthly, 1997-2016, all sites): The Bakun index (Bakun,

1973) provides an index of large-scale coastal upwelling and specifically describes the volume of

water that is transported offshore from Ekman transport (http://www.pfel.noaa.gov/ - sites =

33N-119W and 39N-125W. This index has been used as a large-scale proxy for processes that

may favor larval retention and advection from shore in addition to its value as a predictor of

223 coastal productivity (Menge et al., 2011; Menge & Menge, 2013; Shkedy & Roughgarden, 1997;

224 Wing et al., 2003). We emphasize that this index is not a location specific metric nor does it

225 consider fine-scale nearshore hydrodynamic processes that are also likely to affect larval

226 retention (Fisher et al., 2014; Morgan et al., 2009; Morgan et al., 2016; Morgan et al., 2018;

227 Shanks & Shearman, 2009; Shanks & Eckert, 2005; Shanks & Morgan, 2018; Shanks et al.,

228 2017) which remain outside the scope of this study, but worthy of investigation as drivers of

229 settlement trends. Rather we include the Bakun index as an indicator of broader scale coastal

230 upwelling that may affect regional trends in larval supply (e.g. Roughgarden et al., 1988).

232   *(iii) Sea surface chlorophyll (monthly, 1997-2016, all sites)*: Satellite imagery of sea

233 surface chlorophyll a provides a spatially and temporally resolved estimate of phytoplankton

234 biomass (mg m^{-3}) that is not available from in situ sampling. We used version 3.1 of the OC-CCI

235 merged ocean color time series (Sathyendranath et al., 2018) that combines SeaWIFs, MERIS,

236 MODIS and VIIRS to provide the temporal and spatial coverage required for this study. For

237 each larval settlement collection site, we aggregated data into a 30-day moving average (30 days

238 prior to brush collection) for an area 10 km from shore that stretched 150 km alongshore (the

239 average Lagrangian estimate for dispersal distances for species with a planktonic larval duration

240 (PLD) of 30-days (but see Shanks, 2009; Siegel et al., 2003)). For the site at Anacapa Island, we

241 included any point within a 150 km radius and within 10 km of any coastline. We used 30-day

242 moving averages for chlorophyll and temperature (below) because larvae were exposed to these

243 conditions for at least 30 days prior to settlement, and because averaging over 30 days minimized

244 the effects of serial autocorrelation in the data.

(*iv*) *Sea surface temperature (monthly, 1997-2016, all sites):* We used the 30-day moving

average of sea surface temperature, derived from Pathfinder AVHRR (Reynolds et al., 2007)

(advanced very high resolution radiometer) that was optimally interpolated at daily and 0.25

degree latitude/longitude resolution. We spatially aggregated data in the same way as sea

surface chlorophyll.

(*v*) *Fall kelp canopy biomass (annual, 1996-2016, Santa Barbara Channel & San Diego*

only): Giant kelp (*Macrocystis pyrifera*) is a preferred food and a major constituent of *S.*

purpuratus diets in southern California (Foster et al., 2015). The regional biomass of giant kelp

Macrocystis pyrifera can fluctuate dramatically from year to year (Bell et al., 2015) and cause

orders of magnitude variations in *S. purpuratus* fecundity (Okamoto, 2014). We estimated intra

and interannual variability in the biomass of giant kelp from Landsat Thematic Mapper satellite

imagery (Bell, 2017). Data were aggregated for the Santa Barbara Channel (including islands

and mainland from Point Conception to Santa Monica Bay through Ventura County) or the San

Diego region (mainland coast from the US-Mexico through Orange County including Santa

Catalina and San Clemente Islands). We used the 3-month running mean of kelp canopy

biomass during the period leading into the spawning season (Sept-Nov) because marked declines

in reproductive capacity require several months of consistently low food supply (Okamoto,

2014).

(*vi*) *Adult sea urchin density in the Channel Islands (annual, 1997-2016, Santa Barbara*

Channel only): The abundance of adult sea urchins can also fluctuate over time, which can affect

larval production and supply. We used the spatial geometric mean of adult biomass density (kg

m^{-2}) of purple sea urchins (calculated from surveys of density and size structure from the

Channel Islands Kelp Forest Monitoring program (Kushner et al. 2013) and a size-biomass

model from the Santa Barbara Coastal LTER (Reed, 2018) as an index of the spawning biomass

of purple sea urchins in the Santa Barbara Channel. We used the geometric mean (exponentiated

log-scale mean) as it accounts for spatial differences in overall mean abundance among sites.

We use a biomass index that incorporates size structure and density instead of just density

because individual maximum fecundity increases with size. For details on calculation of

biomass from the KFM data see the Electronic Supplement.

We allowed regression coefficients to vary among major regions (Fort Bragg, Santa

Barbara, and San Diego sites). We used this level of inference because the covariates are

aggregated over spatial regions that encompass all sites within a region due to the long

planktonic larval duration of sea urchins. The regression analyses included either all global

covariates (ENSO, PDO, NPGO) or all local covariates (temperature, chlorophyll, adult

densities, giant kelp biomass, and the Bakun upwelling index). We conducted separate analyses

for these two groups of covariates because global indices were directly correlated with local

covariates, or in the case of temperature were partially derived from them. For the analysis with

local covariates we isolated data to 1997 and onward to accommodate the range of chlorophyll

data availability, while global analysis included all data. Thus, our analyses provided non-

mechanistic explanatory variables for global covariates and mechanistic explanatory variables

for local covariates.

Results

Spatio-temporal trends in larval settlement

Substantial interannual variability in larval settlement of *S. purpuratus* was observed among years (Figure 3, Appendix Figure S1). Fluctuations in larval settlement were highly synchronous among sites within each of the two regions in the Southern California Bight (r = 0.73 and 0.85 for sites within the Santa Barbara Channel and San Diego, respectively; Figure 3b, c). Within the Santa Barbara Channel, pairwise correlations in interannual trends involving Gaviota, Ellwood and Stearns Wharf were higher (r = 0.86 to 0.90) than those involving Anacapa (r = 0.46 to 0.71). Anacapa began to decline in 2012, while the declines at the other sites did not begin until 2014 (Figure 3b, Appendix Figure S.1 b-e). While all of the Santa Barbara Channel sites except Anacapa exhibited eight continuous years of above average settlement following the low in 2005, San Diego sites remained mostly below average after 2003(Figure 3c, Appendix Figure S.1 f-g). This trend produced modest synchrony among San Diego and Santa Barbara sites from 1991 through 2007 (mean r = 0.68), but none thereafter (mean r = -0.06). In contrast, interannual trends in larval settlement at Fort Bragg were largely uncorrelated throughout the time series with sites in the Santa Barbara Channel (Figure 3a vs. 3b, mean r = -0.23) and sites in San Diego (Figure 3a vs. 3c; mean r = 0.12).

Larval settlement in southern California (San Diego and the Santa Barbara Channel) was highly seasonal, with similar patterns among sites (Figure 4a, Appendix Figure S2). On average 90% of recruitment occurred from March to July with a single peak in late April/early May (Figure 4a, Appendix Figure S2), and by July settlement, on average, was an order of magnitude lower than during the April-May peak, and by September is two orders of magnitude lower. By contrast, recruitment at Fort Bragg in northern California extended over a longer period of time

(90% occurred, between January and September) and typically included two peaks per year (a

large peak around July and a smaller peak in February and March; Figure 4a). The seasonal

peaks in settlement in southern California coincided with peaks in sea surface chlorophyll

(Figure 4a vs. 4b.) and troughs in sea surface temperature (Figure 4a vs. 4c). At Fort Bragg in

northern California the primary peak occurred slightly after the peak in chlorophyll a (Figure 4a.

vs. 4b). These peaks in settlement were largely consistent among years, albeit with large

variability in magnitude (Shown by red lines in Appendix Figure S1) and some interannual

variability within sites (see Appendix Figure S3 for a comparison in seasonal trend among

years).

Relationship between benthic year-class strength and larval settlement.

Recruitment of juvenile purple urchins at shallow subtidal reefs in the Santa Barbara

Channel exhibited a significant, positive correlation with larval settlement to brushes two years

prior (Figure 5, negative binomial GLMM - $\chi^2_{df=1}= 9.48$, p = 0.002). These results are robust to

whether collection sites included only Anacapa (the sole site in the Channel Islands) or

incorporated serial correlation using a site-specific first-order autoregressive model in the error

covariance matrix. Years with the highest larval settlement corresponded with a nearly three-

fold increase in the mean density of juvenile urchins two summers later (Figure 5). Over the

time series, average settlement densities in the Santa Barbara Channel varied by more than three

orders of magnitude among years. This interannual variation in settlement (averaged across the

sites in the Santa Barbara Channel) corresponded to more than a three-fold increase in the

density of benthic juveniles on average.

Relationships between larval settlement and biotic and abiotic conditions

Larval settlement of purple urchins showed strong correlations with local sea surface

temperature (SST, Figure 6 a-g, Figure 7 a) as well as the major climate indices (e.g. ENSO - Fig

6 h-n, Figure 7f). For temperature and ENSO, the sign of the correlation at Fort Bragg was

opposite that at sites in the Santa Barbara Channel and San Diego. The correlation between

settlement and temperature varied from strongly negative at sites in San Diego and Santa Barbara

to positive at the Fort Bragg site. Correlations between larval settlement and adult urchin

density, upwelling, chlorophyll and kelp biomass were either not different from zero or negative,

which was opposite of that expected at all sites (Figure 7 b-e). Larval settlement and sea surface

chlorophyll were uncorrelated at Fort Bragg and in the Santa Barbara Channel and negatively

correlated in San Diego – the opposite of the hypothesized relationship that more food in the

plankton would lead to higher larval settlement (Figure 7 d). The network of correlations among

these variables are depicted in (Appendix Figure S.3).

Larval settlement in southern California was orders of magnitude lower during warm, El

Niño conditions and during the negative phase of the NPGO with the more southern sites in San

Diego responding more strongly to temperature than those further north in the Santa Barbara

Channel (Figure 7 a). In contrast, Fort Bragg, the northern most site, responded positively to

warmer El Niño conditions, negatively to the NPGO, and positively to the PDO. Importantly,

relationships observed between larval settlement and ENSO and temperature in southern

California were opposite of those observed at Fort Bragg (Figure 7 f, g). The correlations

between larval settlement and SST and ENSO occurred on roughly 3-5 year cycles, while the

correlation with NGPO and PDO occurred on decadal scales.

Discussion

363   Climate variability can significantly impact marine ecosystems by affecting recruitment,

364 which in turn influences the dynamics of populations and communities. Yet determining which

365 species and locations will respond to shifting climate remains a difficult task. Using a multi-

366 decadal, high frequency, and spatially extensive time series we show how large-scale climatic

367 variation can give rise to spatially different responses in the settlement of larval sea urchins. We

368 found that settlement patterns were synchronous within Santa Barbara and San Diego and

369 synchronous between these two regions until the late 2000s. Importantly, these sites responded

370 negatively to elevated temperature and ENSO events. This contrasted sharply with settlement at

371 our northern California site (Fort Bragg), which was positively correlated to temperature and

372 ENSO. This difference was most obvious during the two strongest ENSO events in 1998 and

373 2014, when settlement responded dramatically in opposite directions.

375   While many recruitment-environment correlations often break down over time (Myers,

376 1998), the correlations between oceanographic factors and larval settlement in our study were

377 evident over a 27-year period that included multiple major ENSO events and several minor ones.

378 We observed repeatable patterns of diminished larval settlement in response to temperature and

379 ENSO events in southern California with opposite responses at Fort Bragg in northern

380 California. Others have found larval settlement in sea urchins to be positively or negatively

381 related to temperature depending on the species (Ebert, 1983; Hernández et al., 2010;

382 Himmelman, 1986). Our study is among the first to show that the relationship between larval

383 settlement and temperature can vary dramatically within a species depending on location.

Although sea urchin larval settlement data at the fine temporal scale of our study are not

available prior to 1990, there is a history of evidence for ENSO-related recruitment failures in

sea urchin populations of southern California. Between 1969 and the early 1980's, recruitment

of juvenile sea urchins was anomalously low during El Niño years (Ebert, 1983; Tegner &

Dayton, 1991). How patterns of larval settlement change in the future will undoubtedly depend

on the processes underlying their associations with ENSO events.

Climate related fluctuations in the supply of larvae have been hypothesized to affect

coastal species around the world (Caley et al., 1996; Grosberg & Levitan, 1992; Underwood &

Fairweather, 1989). In the case of sea urchins, larval supply has been implicated for dramatic

changes in the state of benthic communities (Estes & Duggins, 1995; Hernández et al., 2010;

Ling et al., 2009). Yet empirically demonstrating such links has historically proved challenging

because recruitment can be attenuated by high mortality of newly settled larvae (Connell, 1985;

Rowley, 1989), increasing the need for high frequency data of larval settlement spanning

multiple years. Data such as ours that meet these criteria are rare.

The potential factors affecting larval supply that we examined other than temperature and

climate showed no meaningful correlations with larval settlement. Of particular note was the lack

of a correlation with ocean chlorophyll revealed by our analyses. This finding differed from that

observed for the tropical sea urchin *Diadema aff. antillarum* (Hernández *et al.* 2010) and other

echinoderms (e.g. Fabricius et al., 2010; Lamare & Barker, 1999) and highlights a need for

exploring mechanistically how spatial and temporal variability in ocean circulation and

phytoplankton productivity affect patterns of larval settlement. These outcomes do not suggest

that food is not important for urchin larvae, only that interannual variation therein did not explain

the interannual trends in settlement. Indeed, the mean seasonal trends in both southern and

northern California (which we show are synchronous in southern California) corresponded to

peaks in primary productivity, and both are regions of high productivity (Kahru et al., 2012).

Similar to other studies (Shanks & Shearman, 2009; Shanks & Morgan, 2018) we found

no relationship between large-scale fluctuations in indices of upwelling and larval settlement,

which runs counter to hypotheses that large-scale coastal upwelling shapes patterns of larval

supply by broadly altering productivity and retention (Menge & Menge, 2013; Roughgarden et

al., 1991). The major ocean currents in our study system (the Southern California Counter

Current and the California Current) are characterized by stochastic and spatially variable eddies,

fronts, filaments and bores (e.g. Bassin et al., 2005; Davis et al., 2008; DiGiacomo & Holt,

2001), whereas flows closer to shore tend to promote larval retention along wave exposed shores

(Morgan et al., 2016; Shanks et al., 2017). As a result, offshore transport and retention off the

coast of California is spatially nuanced and temporally inconsistent (Shanks & Eckert, 2005).

Such heterogeneity not only influences larval transport, but also when and where larvae

encounter productive food environments. Consequently, if larval food limitation plays a role in

shaping the dynamics of urchin settlement at our sites, then it likely arises out of more complex

dynamics than our spatially aggregated composite metrics were able to detect. For example,

processes that shape stratification, front formation and spatially isolated phytoplankton blooms in

combination with larval behavior may play a more important role than predicted by

simplifications of the higher or lower spatially averaged phytoplankton biomass or greater or

lesser coastal upwelling.

The oceanographic processes that affect spatial distribution and survival of larvae may

also affect predation driven mortality by altering the densities of predators and their larval prey.

While predation on echinoplutei certainly occurs, its relative importance in shaping larval supply

to the benthos remains difficult to quantify (Rumrill, 1990; Vaughn & Allen, 2010). For *S.*

purpuratus plutei, predation may be lower than for other taxa because they contain chemical and

structural defenses and display predator avoidance behavior (Cowden et al., 1984), particularly at

later stages of development, which are far less susceptible to predation (Rumrill, 1985). This is

supported by field studies that have shown predator mortality to be comparatively low for

echinoids (Allen & McAlister, 2007; Johnson & Shanks, 2003), which in part may be due to the

fact that accounting for natural densities of echinoplutei can dramatically reduce estimates of

laboratory predation rates (Johnson & Shanks, 1997). Integrating mechanistic models of ocean

circulation and mortality was beyond the scope of our study, yet such approaches are needed to

develop a more thorough understanding of processes that control settlement dynamics across

large gradients in oceanographic settings as they can control both population dynamics and range

limits (Gaylord & Gaines, 2000).

Although climate related changes can alter larval supply via their effects on the

production of larvae by adults, we found no relationship between larval settlement and regional

adult abundance or adult food (kelp). While fisheries research relies heavily on stock-recruit

dynamics, there is continual debate about whether adult dynamics actually control recruitment

patterns (Gilbert, 1997; Szuwalski et al., 2015). The lack of a positive correlation between adults

and larval settlement that we found for purple sea urchins adds to this debate by showing that

high abundances of adult urchins and kelp averaged across large spatial scales did not translate

into high larval settlement. We note that spatial structure in the dynamics of kelp and adult

urchins may have affected this outcome. For example, modest but uniform densities of adult sea

urchins in food rich kelp forests can provide very different outcomes for the production of

embryos compared to mosaics of dense adults in barren patches interspersed with forested

patches having few adults (Okamoto, 2014). Indeed, previous work has demonstrated that

assessing population productivity in spatially structured populations requires a spatially

structured analysis to account for processes of population regulation and competition (Chesson,

1996, 1998; Chesson et al., 2005; Thorson et al., 2015). That said, we found no evidence that

synchronous fluctuations in settlement arose from broad-scale collapses and proliferations in

populations of adult sea urchins or kelp at the appropriate lags and time-scales.

Temperature-related effects on larval production and survival and ENSO-related changes

in currents or oceanographic features that transport larvae represent two possible explanations for

the spatially opposing correlations between larval settlement and temperature and ENSO that we

observed. Many species have upper temperature thresholds beyond which reproduction is

impaired that are well below the lethal limits for adults. Temperatures in southern California

kelp forests routinely exceed 17°C during ENSO events (Reed et al. 2016), which may

adversely affect larval supply of purple sea urchins by limiting gamete production (Basch &

Tegner, 2007; Cochran & Engelmann, 1975), fertilization (Schroeder & Battaglia, 1985) larval

development and gene expression (Padilla-Gamiño et al., 2013; Runcie et al., 2012; Wong et al.,

2018), and larval survival (Azad et al., 2012). Such effects may in part explain why larval

settlement at Fort Bragg, where sea surface temperatures rarely exceed 16°C was not negatively

correlated with temperature. Unfortunately, there are no long-term datasets on reproductive

activity or planktonic larval densities and distributions to evaluate if and how the interannual and

seasonal trends observed here correspond to variation in reproductive activity and transport of

larvae.

Regional circulation is considered to be an important driver of larval supply for many

species, including those in the Southern California Bight (Blanchette et al., 2006; Broitman et al.,

2005; Hereu et al., 2004; McManus & Woodson, 2012; Woodson et al., 2012). ENSO events

can cause major changes in patterns of ocean circulation in the Southern California Bight and

elsewhere along the US west coast (Lynn & Bograd, 2002; Mitarai et al., 2009; Pineda & Reyns,

2017).   Spatial variability in such patterns could explain the opposing climatic responses in

larval settlement between Fort Bragg and our sites in southern California. El Niño events in

southern California are associated with lower stratification and reduced internal waves that

promote the onshore transport of larvae (Pineda, 1994; Pineda et al., 2018; Shanks, 1983). In

contrast, El Nino events in northern California are often associated with relaxed upwelling,

downwelling Kelvin waves and increased stratification (Chavez et al., 2002). Although we

cannot definitively partition out the effects of climate associated changes in temperature and

circulation on recruitment, our observations of larval settlement and subsequent year-class

strength point strongly to broader-scale ocean climate effects.

Our finding that larval settlement was a good predictor of regional year class strength in

natural populations indicates that large fluctuations in larval settlement of this prominent

herbivore could have far reaching ecological impacts that resonate throughout marine

ecosystems across a broad geographic region (Pearse, 2006). For example, the high settlement

that we observed at Fort Bragg in 2013-2015 coupled with a period of anomalously warm water

may explain the marked increase in the abundance of purple urchins and coincident loss of

canopy forming kelps reported for northern California beginning in 2014 (Rogers-Bennett &

Catton, 2019). Although regional processes appear to drive broad-scale patterns of larval

settlement in sea urchins, local hydrodynamics and other factors contribute to considerable fine-

scale spatial variation in larval supply and settlement in sea urchins (Ebert et al., 1994; Farina et

al., 2018; Hereu et al., 2004; Schroeter et al., 1996; Schroeter et al., 2009). For example, pelagic

larval abundance and settlement of *Paracentrotus lividus* varies substantially in space (Hereu et

al., 2004; Prado et al., 2012) as does post-settlement survival and abundance due to localized and

habitat dependent effects (Boada et al., 2018; Prado et al., 2012). Our inferences focused on

temporal trends and comparison of regional-scale patterns, rather than trying to infer causes and

consequences of variation in average settlement in space. Despite these caveats, our analyses

show remarkable synchrony in settlement within southern California and strong association with

regional benthic recruitment in the Santa Barbara Channel. Collectively, our results suggest that

predicted changes in ocean climate that lead to more frequent or severe marine heatwaves

(Frölicher and Laufkötter 2018) will reduce purple urchin recruitment in southern California, but

increase it in northern California with potentially cascading effects on benthic ecosystems similar

to those recently observed in northern California.

Oceans are experiencing simultaneous shifts in temperature, water chemistry,

productivity and circulation, which have profound consequences for the ecological structure and

function of marine systems and the services that they provide (Wong et al., 2014). Our findings

provide valuable insights into the ecological consequences of climate related effects on patterns

of larval settlement of an important reef herbivore whose distribution spans most of the Pacific

coast of North America. Future investigations aimed at determining the specific biotic and

abiotic processes that regulate larval settlement in ecological important species such as *S.*

purpuratus will undoubtedly improve our understanding of how climate fluctuations affect

regional population dynamics to alter the structure and functions of marine ecosystems.

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Figure 5: The relationship between the summer density of juvenile purple sea urchins on reefs in

the Santa Barbara Channel and the density of recruits on larval collectors during March- July of

the previous year (i.e., 12-20 months prior). Points and error bars represent annual means (+/-

SE) averaged over all sites. The line with the band represents the estimated relationship and

95% uncertainty interval.

Figure 6: Standardized coefficients for the relationships between larval settlement of purple sea

urchins at each site and: (a-g) sea surface temperature, (h-n) the multivariate ENSO index, and

(m-u) the North Pacific Gyre Oscillation (NPGO). Small grey points represent biweekly means

and black points and lines represent annual means.

Figure 7: Standardized Bayesian multiple regression coefficients for relationships between *S.*

purpuratus settlement and local covariates (a-e) or global climatic indices (f-h). Models with

global and local variables were fit separately. Coefficients were estimated using a multiple

regression nested within a Bayesian multivariate time series model. Coefficients are *a priori*

biased towards zero using a regularized horseshoe prior for variable selection. For details see

Methods, Figure 2 and appendix for details.

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Figure 1: Map of larval settlement collection sites in California. Colors represent the average spatial gradient of sea surface temperature from 1991-2017. The hatched white lines show the shoreline buffer used to constrain sea surface temperature and sea surface chlorophyll index to local larval settlement observations.

Figure 2: Diagram of the model structure using either a Matérn temporal covariance (trend estimation model) or a horseshoe regression (correlation estimation model) to estimate seasonal anomalies. For details see Tables 1 and 2. *Bayesian Regression Model Only. † Trend Estimation Model Only

Figure 3: Standardized annual scale trends (log-scale, standardized to one SD) of larval settlement S. purpuratus from 1990 through 2016 for: (a) Fort Bragg, (b) sites in the Santa Barbara Channel, and (c) sites in San Diego. Colors within represent individual sites associated with the legend. Interannual trends are estimated using a 3/2 Matérn Gaussian process covariance within a Bayesian multivariate state-space model accounting for spatiotemporal correlations.

Figure 4: Seasonal trends at each site for: (a) standardized larval settlement of S. purpuratus, (b) biweekly mean sea surface chlorophyll a (chl a) and (c) biweekly mean sea surface temperature (SST). The seasonal trend in larval settlement at each site was standardized by the mean maximum seasonal value. Chl a and SST were measured using satellite imagery within 25 km of the coastline and 150 alongshore km of the location where settlement was sampled (hatched areas in Figure 1). See Methods for details.

Figure 5: The relationship between the summer density of juvenile purple sea urchins on reefs in the Santa Barbara Channel and the density of recruits on larval collectors during March- July of the previous year (i.e., 12-20 months prior). Points and error bars represent annual means (+/- SE) averaged over all sites. The line with the band represents the estimated relationship and 95% uncertainty interval.

Figure 6: Relationships between larval settlement of purple sea urchins at each site and: (a-g) sea surface temperature, (h-n) the multivariate ENSO index, and (m-u) the North Pacific Gyre Oscillation (NPGO). Small grey points represent biweekly means and black points and lines represent annual means.

Figure 7: Standardized Bayesian multiple regression coefficients for relationships between *S. purpuratus* settlement and local covariates (a-e) or global climatic indices (f-h). Models with global and local variables were fit separately. Coefficients were estimated using a multiple regression nested within a Bayesian multivariate time series model. Coefficients are a priori biased towards zero using a regularized horseshoe prior for variable selection. For details see Methods, Figure 2 and appendix for details.

Table 1: Equations for the multivariate spatiotemporal models. Variables, parameters and priors are described in Table 2. Note that there are two primary model forms: a trend estimation model, and a Bayesian regression model. Equations apply to both model forms except where specified.

Eq.	Description	Equation
3	total count likelihood	$\begin{cases} Tot_{t,s,b} \sim \text{Poisson}(\hat{\mu}_{t,s} \times D_{t,s,b}/p_{t,s,b}) & \text{when subsampled} \\ N_{t,s,b} \sim \text{Poisson}(\hat{\mu}_{t,s} \times D_{t,s,b}) & \text{otherwise} \end{cases}$
4	subsampling prior	$p_{t,s,b} \sim \text{beta}(N_{t,s,b} + 0.5, NO_{t,s,b} + 0.5)$ when subsampled
5	prior seasonal temporal correlation function	$\bar{r}_{<t_i,t_j>} = \exp\left[-\frac{2}{1.5^2}(0.5 - 0.5\cos(\pi|t_i - t_j|))\right]$ Periodic Kernel (MacKay 1998)
6[†]	prior annual temporal correlation function	$\bar{\bar{r}}_{<t_i,t_j>} = (1 + 3^2|t_i - t_j|)e^{-3^2|t_i-t_j|}$ Matérn 3/2 Kernel (Rasmussen & Williams 2006)
7	prior seasonal spatiotemporal covariance function (separable)	$\bar{\boldsymbol{\Sigma}} = \bar{\boldsymbol{\Omega}} \otimes [\bar{\boldsymbol{\sigma}}\bar{\boldsymbol{R}}]$
8[†]	prior annual spatiotemporal covariance function (separable)	$\bar{\bar{\boldsymbol{\Sigma}}} = \bar{\bar{\boldsymbol{\Omega}}} \otimes [\bar{\bar{\boldsymbol{\sigma}}}\bar{\bar{\boldsymbol{R}}}]$
9	matrix of seasonal trends	$\textbf{Seasonal} \sim \text{MVN}(0, \bar{\boldsymbol{\Sigma}})$
10[†]	matrix of annual trends	$\textbf{Annual} \sim \text{MVN}(0, \bar{\bar{\boldsymbol{\Sigma}}})$
11a[*]	first order autoregressive model	$\boldsymbol{\epsilon}_t = \boldsymbol{\phi}\boldsymbol{\epsilon}_{t-1} + \boldsymbol{e}_t\sqrt{(1 - \boldsymbol{\phi}^2)}$
11b	spatially correlated process error	$\boldsymbol{e}_t \sim \text{MVN}(0, \boldsymbol{\sigma}\boldsymbol{\Omega})$

∗ Bayesian Regression Model Only
† Trend Estimation Model Only

Table 2: Definition of parameters, variables, and priors for equations described in Table 1

Symbol	Description	Type	Prior/Input
$\hat{\mu}_{t,s}$	expected settlement density (per brush, per day) of purple urchins, defined in Eqs. 1 or 2 in the main text for each model.	estimated	Eqs. 1, 2
$N_{t,s,b}$	number of purple urchins counted at time t, site s and brush b. When urchin species within brushes are subsampled, data produced are a total urchin count ($Tot_{t,s,b}$), a total subsample count ($NO_{t,s,b}$, and a purple urchin subsample count ($N_{t,s,b}$).	input	data
$Tot_{t,s,b}$	total number of urchins counted at time t, site s and brush b	input	data
$p_{t,s,b}$	estimated proportion of purple urchins in the counts given the subsample (when urchin counts in brushes are subsampled).	input	data
$D_{t,s,b}$	exact number of days of the brush deployment	input	data
$\mathbf{X}$	covariate design matrix - for estimation model represents only site indices, for regression model includes covariates	input	data
$\boldsymbol{\beta}$	vector of site specific scale parameters and covariate coefficients	estimated	**Scale parameters**: Student-t: scale=2; **Coefficients**: regularized horseshoe prior (see methods).
$\boldsymbol{\phi}$	vector of site specific first order autoregressive parameters & estimated (regression model only)	estimated	Uniform -0.95 to 0.95
$\boldsymbol{\Omega}, \bar{\boldsymbol{\Omega}}, \bar{\bar{\boldsymbol{\Omega}}}$	spatial correlation matrices for process error, annual trends, and seasonal trends (estimated through the cholesky factor)	estimated	LKJ prior:scale= 2 (Lewandowski *et al.* 2009)
$\bar{\boldsymbol{R}}, \bar{\bar{\boldsymbol{R}}}$	temporal correlation matrices for annual and seasonal gaussian processes	input	eqs 5 & 6
$\boldsymbol{\sigma}, \bar{\boldsymbol{\sigma}}, \bar{\bar{\boldsymbol{\sigma}}}$	site specific standard deviation for process error, annual trends, and seasonal trends	estimated	half-Cauchy: scale = 2.5 (Polson & Scott 2012)

Supplementary Information - Appendices for
"Effects of Ocean Climate on Spatiotemporal Variation in Sea
Urchin Settlement and Recruitment"

Daniel K. Okamoto[*1,2], Stephen Schroeter[†3], and Daniel C. Reed[‡3]

[1]Department of Biological Science, Florida State University, Tallahassee, Florida 32303
[2]Ecology, Evolution and Marine Biology, University of California Santa Barbara, Santa Barbara, CA 93106 USA
[3]Marine Science Institute, University of California Santa Barbara, Santa Barbara, CA 93106 USA

[*]corresponding author: dokamoto@bio.fsu.edu
[†]email: schroete@lifesci.ucsb.edu
[‡]email: dan.reed@lifesci.ucsb.edu
[*]corresponding author: dokamoto@bio.fsu.edu
[†]email: schroete@lifesci.ucsb.edu
[‡]email: dan.reed@lifesci.ucsb.edu

Estimation of juvenile urchin densities

The KFM program surveys purple sea urchins at the Santa Barbara Channel Islands by counting the total number of individuals in defined areas and measuring approximately 100 individuals at each site. Thus, to model density we used the number of juveniles measured divided by the total number of individuals measured, multiplied by the total number of individuals counted, divided by the area surveyed:

$$Juvenile \ \ Density_{i,t} = \frac{Juveniles \ \ Measured_{i,t}}{Total \ \ Measured_{i,t}} \frac{Total \ \ Count_{i,t}}{Total \ \ Area_{i,t}}$$

We modeled density in the GLM using the number of measured juvenile urchins (2.5 cm in test diameter- the approximate cutoff size for reproduction - Kenner & Lares 1991). Let represent expected juvenile density at site i in year t. We constructed the regression as:

$$\ln \mu_{i,t} = \alpha_i + \beta \ \ \ln Larval \ \ recruitment_t$$

We allowed the intercept (α_i) to vary by site nested within each island because of overall differences in mean juvenile density among sites and islands. We used a negative binomial (NB) with the using direct mean and variance parameterization form:

$$Juveniles \ \ Measured_{i,t} \sim \text{NB}(\ln \mu_{i,t} - \ln[\frac{1}{TotalMeasured_{i,t}} \frac{TotalCounted_{i,t}}{TotalArea_{i,t}}], \sigma^2)$$

We constructed the model in this format (i.e. with the density denominator in the left hand side of the equation) to maintain the sample size and integer nature of the data while modeling the mean density.

Figure S.1: Observed and estimated purple urchin (*S. purpuratus*) settlement trends from 1990-2016 for each site. Large points represent empirical biweekly means, lines on the axis represent biweekly mean values equal to zero, small points represent counts on individual brushes (only positive values shown for clarity, although the models include zero values), red lines represent biweekly estimates, lines with blue 95% uncertainty intervals represent model estimates of the interannual trends. The model estimates the seasonal trend and the interannual trend at each site simultaneously. For reference, interannual trends for *S. purpuratus* shown here are the unstandardized versions of those shown in Figure 2.

Figure S.2: Seasonal settlement trends from 1990-2016 with the mean estimated seasonal trend for each site. Large points represent empirical biweekly means lines on the axis represent biweekly mean values equal to zero, small points represent counts on individual brushes (only positive values shown for clarity, although the models include zero values), black lines represent the posterior mean and bands represent the 95% uncertainty interval. The model estimates the seasonal trend and the interannual trend at each site simultaneously. For reference, seasonal trends for *S. purpuratus* shown here are the unstandardized versions of those shown in Figure 3.

Figure S.3: Cumulative annual mean settlement from the estimated biweekly means for each year. Thick black lines represent the overall mean (i.e. the empirical cumulative mean from Figure 3) and thin, colored lines represent the cumulative annual values of the red lines in Figure S1. Blue bands represent the 10 and 90% quantiles for each biweekly period.

Variable Correlations

To illustrate the intercorrelated nature of the covariates and responses, we estimated a network model for Gaviota Pier that includes the independent model estimates of *S. purpuratus* settlement and its seasonality. To do so, we first constrained network structure by excluding all nonsensical interactions (i.e. chlorophyll does not cause ENSO events and is thus eliminated a priori) and including all known directional interactions (i.e. estimated seasonality affecting settlement for is forced into the network). Learning of the network skeleton is achieved via the Hill Climbing algorithm using the bnlearn (Scutari 2010) package in R. Note that strong collinearity can result in the weaker correlation being ignored (e.g. ENSO over SST with Larval Recruitment).

Figure S.4: Bayesian network model structure inferred for the Santa Barbara Channel. Numbers represent the partial correlation coefficients accounting for all other variables pointing to that node. All correlations shown are significant at alpha= 0.05

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```
\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\
\\ Stan Code Required to Estimate Trends      \\
\\ Author: D.K. Okamoto                       \\
\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\

data {
  // input information
  int NO; // number of observations
  int NS; // number of sites
  int NM; // number of months
  int NYM; // number of months
  int NSUB; // number of observations with subsamples
  // input data
  //  Beta priors for uncertainty in subsamples
  vector[NSUB] P1; // SP observations (on the original scale)+0.5
  vector[NSUB] P2; // SP observations - subsample count (on the original scale) +
      0.5
  int SUB[NSUB];

  int N[NO]; // total count (on the original scale)
  int OBS[NO]; // continuous month-site observation index
  int PRED_MONTH[NM]; // month-site observation index associated with MA
  vector[NO] D; // number of days
  cholesky_factor_cov[NYM] D_seas;   // seasonal cholesky covariance factor
  cholesky_factor_cov[NM] D_ann;   // annual cholesky covariance factor
}

parameters {
  // mean volatility
  vector<lower=1e-7>[NS] sigma_S_mu;

  // spatial volatility correlation matrix
  cholesky_factor_corr[NS] L_Omega_S; //
  cholesky_factor_corr[NS] L_Omega_Spatial;

  // seasonal and annual variance
  vector<lower = 0>[NS] sigma;
  vector<lower = 0>[NS] sigma2;

  // mean site settlement
  row_vector<lower= -10, upper= 10>[NS] mu_S;

  // iid errors for seasonality
  matrix<lower= -10, upper= 10>[NYM,NS] z_s;

  // iid errors for annual trends
  matrix<lower= -10, upper= 10>[NS,NM] w_z;

  // iid errors for residuals
  matrix<lower= -4, upper= 4>[NS,NM] e_z;

  // proportion of samples that are purps
  vector<lower= 0,upper= 1>[NSUB] theta;
}

transformed parameters {
  matrix[NYM,NS] LS; // log scale expected sp per brush, per day for every month

  matrix[NM,NS] S; // estimated abundance
```

```
  vector[NO] S_exp; // normal scale estimated sp for each observation

  vector[NO] theta2; // fraction that is SP

  matrix[NM,NS] w;

  // correlated errors
  matrix[NM,NS] e;

  LS  = D_seas*z_s;
  w = D_ann*transpose(L_Omega_Spatial*w_z);

  theta2 = rep_vector(1.0,NO);
  theta2[SUB] = theta;
  // put it all together
  e = transpose(diag_pre_multiply(sigma_S_mu,L_Omega_S)*e_z);
  S= rep_matrix(mu_S,NM)+diag_post_multiply(LS[PRED_MONTH,],sigma2)+
      diag_post_multiply(w,sigma)+e;
  S_exp = exp(to_vector(S)[OBS]) .*D ./theta2;
}

model {
  sigma ~ cauchy(0, 0.5);
  sigma2 ~ cauchy(0, 0.5);

  L_Omega_S ~ lkj_corr_cholesky(2);
  L_Omega_Spatial ~ lkj_corr_cholesky(2);

  for(i in 1:NS){
    z_s[i]~ normal(0,1);
    mu_S[i] ~ normal(0,5);
  }

  sigma_S_mu ~ cauchy(0,2.5);

  // uncertainty on proportions of purples in the sample
  // jeffrey's prior given subsampling
  theta~beta(P1, P2);

  // observation likelihood
  N~poisson(S_exp);
}
```

```
\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\
\\ Stan Code Required to Estimate Bayesian Regression \\
\\ Author: D.K. Okamoto                               \\
\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\
data {
  // input information
  int NO; // number of observations
  int NS; // number of sites
  int NM; // number of months
  int NYM; // number of months
  int NSUB; // number of observations with subsamples
  int NSP; // number of subsample counts
  int NP; // number of variables
  // input data
  //  Beta priors for uncertainty in subsamples
  vector[NSP] P1; // SP observations (on the original scale)+0.5
  vector[NSP] P2; // NON SP observations (on the original scale) + 0.5
  int SUB[NSUB];
  int ID[NO];
  int OBS[NO]; // continuous month-site observation index
  // input data
  int N[NO]; // total count (on the original scale)
  int OBS_MONTH[NO]; // continuous month-site observation index associated with MA
  int OBS_SITE[NO]; // month-site observation index associated with MA
  int PRED_MONTH[NM]; // month-site observation index associated with MA
  matrix[NM*NS,NP] X; // covariate matrix
  int xind[NM,NS]; // covariate matrix indices
  vector[NO] D; // number of days
  cholesky_factor_cov[NYM] D_seas;   // seasonal cholesky covariance factor
  real<lower= 0> m0;
  real<lower= 0> sigma_scale;
  real<lower= 0> slab_scale;
}

parameters {
  // mean volatility
  vector<lower=0>[NS] sigma_S_mu;

  // spatial volatility correlation matrix
  cholesky_factor_corr[NS] L_Omega_S; //
  cholesky_factor_corr[NS] L_Omega_Spatial_seas;

  // iid errors for seasonality
  matrix[NYM,NS] z_s;

  // seasonal and annual variance
  real<lower = 0> sigma[NS];

   // mean site settlement
  row_vector[NS] mu_S;

  // iid errors for residuals
  matrix<lower= -4,upper=4>[NS,NM] e_z;

  // AR1 parameters
  vector<lower= -0.9,upper= 0.9>[NS] phi;

  // proportion of samples that are purps
  vector<lower= 0,upper= 1>[NSP] theta;
```

```
  // iid errors for seasonal gaussian process
  vector[NP] beta_tilde;
  vector<lower=0>[NP] lambda;
  real<lower=0> c2_tilde;
  real<lower=0> tau_tilde;
  real alpha;
}

transformed parameters {
  matrix[NYM,NS] LS; // log scale expected sp per brush, per day for every month
  matrix[NM,NS] S; // estimated abundance
  vector[NO] S_exp; // normal scale estimated sp for each observation

  vector[NO] N_mu; // expected total count
  vector[NO] q_mu; // expected fraction of N that is SP
  vector[NS] sigma_star;

  vector[NM*NS] s_hat;

  // process variables
  vector[NO] theta2; // fraction that is SP

  // correlated errors
  matrix[NM,NS] e;
  matrix[NM,NS] e_hat;

  vector[NP] beta;
  vector[NP] lambda_tilde;

  real tau0 = (m0/(NP-m0))*(mean(sigma_S_mu)*sqrt(1.0*NM*NS));
  real slab_scale2 = square(slab_scale);
  real half_slab_df = 0.5 * 25;

  real tau = tau0 * tau_tilde;
  real c2 = slab_scale2 * c2_tilde;
  lambda_tilde  = sqrt( c2 * square(lambda) ./ (c2 + square(tau) * square(lambda))
      );
  beta = tau * lambda_tilde .* beta_tilde;

  // regression model
  s_hat = X*beta;

  // seasonal gaussian process model
  LS  = D_seas*z_s;

  // process error
  for(i in 1:NS){
    sigma_star[i] = sigma_S_mu[i]*sqrt(1.0-phi[i]^2.0);
  }
  e = transpose(diag_pre_multiply(sigma_star,L_Omega_S)*e_z);
  // put it all together

  for(j in 1:NS){
    e_hat[1,j] = e[1,j];
    S[1,j]=mu_S[j]+LS[PRED_MONTH[1],j]*sigma[j]+s_hat[xind[1,j]]+e_hat[1,j]-5;
    for(i in 2:NM){
      e_hat[i,j] =  phi[j]*e_hat[i-1,j]+e[i,j];
      S[i,j]=mu_S[j]+LS[PRED_MONTH[i],j]*sigma[j]+s_hat[xind[i,j]]+e_hat[i,j]-5;
```

```
    }
  }

  theta2 = rep_vector(1.0,NO);
  theta2[SUB] = theta[ID[SUB]];

  S_exp = exp(to_vector(S)[OBS]) .*D ./theta2;
}

model {
  sigma ~ normal(0.0, 2);

  L_Omega_S ~ lkj_corr_cholesky(2.0);

  for(i in 1:NS){
    z_s[i]~ normal(0.0,1.0);
    e_z[i]~ normal(0.0,1.0);
    mu_S[i] ~ cauchy(0.0,10);
  }

  sigma_S_mu ~ normal(0.0,sigma_scale);

  beta_tilde ~ normal(0, 1);
  lambda ~ cauchy(0, 1);
  tau_tilde ~ cauchy(0, 1);
  c2_tilde ~ inv_gamma(half_slab_df, half_slab_df);

  // jeffrey's prior given subsampling
  theta~beta(P1, P2);

  // observation likelihood
  N~poisson(S_exp);
}
```