1	Co-invasion of the ladybird Harmonia axyridis and its parasites Hesperomyces virescens fungus and
2	Parasitylenchus bifurcatus nematode to the Caucasus
3	
4	Marina J. Orlova-Bienkowskaja ^{1*} , Sergei E. Spiridonov ² , Natalia N. Butorina ² , Andrzej O. Bieńkowski ²
5	
6	¹ Vavilov Institute of General Genetics, Russian Academy of Sciences, Moscow, Russia
7	² A.N. Severtsov Institute of Ecology and Evolution, Russian Academy of Sciences, Moscow, Russia
8	* Corresponding author (MOB)
9	E-mail: marinaorlben@yandex.ru
10	
11	Short title: Co-invasion of Harmonia axyridis and its parasites to the Caucasus
12	

13 Abstract

14 Study of parasites in recently established populations of invasive species can shed lite on sources of 15 invasion and possible indirect interactions of the alien species with native ones. We studied parasites of 16 the global invader Harmonia axvridis (Coleoptera: Coccinellidae) in the Caucasus. In 2012 the first 17 established population of *H. axyridis* was recorded in the Caucasus in Sochi (south of European Russia, 18 Black sea coast). By 2018 the ladybird has spread to the vast territory: Armenia, Georgia and south 19 Russia: Adygea, Krasnodar territory, Stavropol territory, Dagestan, Kabardino-Balkaria and North 20 Ossetia. Examination of 213 adults collected in Sochi in 2018 have shown that 53% of them are infested 21 with Hesperomyces virescens fungi (Ascomycota: Laboulbeniales) and 8% with Parasitylenchus 22 bifurcatus nematodes (Nematoda: Tylenchida, Allantonematidae). Examined H. axyridis specimens were 23 free of parasitic mite *Coccipolipus hippodamiae*. An analysis of the phylogenetic relationships of 24 Parasitylenchus bifurcatus based on 18S rDNA confirmed the morphological identification of this 25 species. Hesperomyces virescens and Parasitylenchus bifurcatus are firstly recorded from the Caucasus

- and Russia, though widespread in Europe. It probably indicates that they appeared as a result of co-
- 27 invasion with their host. Harmonia axyridis was released in the region for pest control, but laboratory
- 28 cultures are always free of *H. virescens* and *P. bifurcatus*. Therefore, detection of *H. virescens* and *P.*
- 29 *bifurcatus* indicates that population of *H. axyridis* in the Caucasus cannot derive exclusively from
- 30 released specimens. We did not find *H. virescens* on 400 specimens of 31 other ladybird species collected
- 31 in the same localities with *H. axyridis* in the Caucasus. No reliable correlation between infestation by *H.*
- 32 virescens and P. bifurcatus has been found. Besides these two parasites an unidentified species of the
- 33 order Mermithida is recorded. It is the first documented case of *H. axyridis* infestation by a parasitic
- 34 nematode of this order in nature.
- 35 Key words Harmonia axyridis; harlequin ladybird; invader; parasites; Hesperomyces virescens;
- 36 Parasitylenchus bifurcatus; alien species; biological invasions; co-invasion

37 Introduction

Despite a large body of work on invasion ecology, interactions of invasive species with their natural enemies, in particular parasites, are poorly studied [1]. Study of parasites of alien species in young, recently established populations is of great importance for understanding of rotes of invasion [2]. Besides that, some parasites of alien species can affect native species [3]. Therefore, the study of parasites might reveal possible indirect interactions of an alien species with native ones and some reasons of its invasive success. The aim of our investigation was to determine what parasites affect the harlequin ladybird *Harmonia axyridis* (Pallas) (Coleoptera: Coccinellidae) in the Caucasus recently invaded by this species.

45 Harmonia axvridis native to East Asia has been introduced widely for biological control of 46 agricultural pests and established almost all over the world (see detailed description of the native range 47 [4] and overview of global invasion [5]). It established in Western Europe the late 1990s and then 48 expanded its range rapidly. Outbreak of H. axyridis in some regions caused a number of negative 49 ecological consequences including decline of native ladybird species [6]. Approximately by 2010 the 50 expansion of European range to the east reached Russia and adjacent regions [7]. The reasons of the great 51 invasive success of this species and decline of native ladybirds attract attention of hundreds of scientists 52 (see review by Roy et al. [5]). Recent studies have shown that some symbionts of this global invader 53 (parasites and microorganisms) have contributed to its success and sometimes even become a "biological 54 weapon" against its competitors - native ladybirds [1, 3, 8].

Harmonia axyridis was widely used for biological control of Aphidae and other pests in the Caucasus since 1927. In particular, in the 1980s more than 107,000 of specimens brought from the Far East were released in Georgia [9]. But in spite of these massive releases, *H. axyridis* did not establish before the 21st century. The first established population in the region was recorded in Sochi in 2012 [10, 11]. Then *H. axyridis* quickly spread and became common all over the Black Sea coast of the Caucasus and in adjacent regions. By 2018 it has been recorded in Armenia, Georgia including Abkhazia, and south

61	Russia: Adygea,	Krasnodar te	erritory, Stavrop	ol territory, Rost	ov region,	Dagestan,	Kabardino-Balkaria	a
	, ,		5 7 1	57	0,	0 ,		

- and North Ossetia (Fig 1) [12–14]. The releases of *H. axyridis* continued at least to 2010 [15]. So it was
- 63 unclear if the population in the Caucasus originated from released specimens or appeared a result of
- 64 expansion of European invasive range of the species [11, 14, 16].

- 66 Fig 1. Known localities of *Harmonia axyridis* and its parasites in the Caucasus.
- 67 1 localities of infestation of Harmonia axyridis with Hesperomyces virescens and Parasitylenchus
- 68 bifurcatus, 2 localities of infestation of Harmonia axyridis with Hesperomyces virescens, 3 other
- 69 localities of Harmonia axyridis. Regions of Russia: AD Adygea, DA Dagestan, KA Kabardino
- 70 Balkaria, KR Krasnodar region, NO North Ossetia, ST Stavropol region. Localities of detection of
- 71 the parasites: A Golovinka, B Razbityj Kotel, C Central District, D Agur, E Adler, F Veseloe.
- 72 Description of localities and sources of information are indicated in supporting information (S1 Table).
- The map is compiled with the help of DIVA GIS program using the free basic map: http://divagis.org/Data.
- 75 Establishment of *H. axyridis* in the Caucasus causes three main questions:
- 76 1. What are the sources of invasion of *H. axyridis* to the Caucasus?
- Why is *H. axyridis* currently established in the region, though previously failed to establish in
 spite of massive releases?
- 79 3. How will *H. axyridis* affect native ladybird species?
- 80 Though our study of parasites cannot directly answer these difficult questions, it could probably shed
 81 some light on them.
- 82 We restrict usage of the term "parasites" to those organisms living at the expense of a single

83

84 of the host (in contrast to parasitoids) [1]. Three species of parasites have been recorded to infest 85 Harmonia axvridis in the world: Hesperomyces virescens Thaxt. fungi, Coccipolipus hippodamiae

host, which are multicellular (in contrast to pathogenic microorganisms) and do not directly cause death

86 (McDaniel and Morrill) mites, and Parasitylenchus bifurcatus Poinar and Steenberg nematodes [1].

87 Hesperomyces virescens (Ascomycota: Laboulbeniales) is an obligate ectoparasite that has been 88 reported to infect adults of over 30 ladybird species (Coleoptera: Coccinellidae) in all continents except 89 Australia and Antarctica [1]. The association *H. axyridis–H. virescens* has been reported from Austria, 90 Belgium, Czech Republic, Croatia, Germany, France, Hungary, the Netherlands, Poland, Slovakia, the 91 UK, USA, Canada, Argentina, Ecuador, South Africa, and China [1]. Hesperomyces virescens has not 92 been detected in Russia or in the Caucasus before [1]. Native range of this fungus is unknown. It was 93 assumed that it could originate from North America [17]. Its entire life cycle takes place on integument of 94 adult ladybird. The sticky spores have shorts life span, are exclusively spread by activities of the host, 95 and transmission takes place during direct contact of specimens (mating or overwintering in 96 aggregations) [18].

97 Coccipolipus hippodamiae (Acarina: Podapolipidae) is an ectoparasitic mite. All life stages live 98 on the underside of the elytra of an adult ladybird and feed on the haemolymph. It is reported to affect 99 five Coccinellidae species in North America, Africa and Europe. Transmission also takes place during 100 mating or overwintering in aggregations.

101 Parasitylenchus bifurcatus (Nematoda: Allantonematidae) is an obligate endoparasite. Its only 102 known host is Harmonia axyridis. Fertilized infective female enters an adult ladybird through spiracles or 103 thin parts of the integument. It develops into a female of the first parasitic generation, producing eggs, 104 from which the second parasitic generation females develop. The latter give birth to juvenile males and 105 females, maturing and mating inside the body of the host. Then new infective females appear and leave 106 the host. Mechanism of transmission is unknown [19]. Parasitylenchus bifurcatus originally described

- from Denmark [19], has been also reported in the Netherlands [20], Czech Republic, Poland [1], the USA
 and Slovenia [21]. There have been no documented records of infestation of *H. axyridis* by nematodes
 belonging to the order Mermithida in nature.
- 110 No parasites of *H. axyridis* in the Caucasus and European Russia have been recorded [5]. We

111 have found *Hesperomyces virescens*, *Parasitylenchus bifurcatus* and an unidentified species belonging to

112 the order Mermithida on *H. axyridis* in the region.

113 Materials and methods

114 Collection of Harmonia axyridis

115 Adult Harmonia axyridis (213 specimens) were collected in the city of Sochi (south of European 116 Russia, Krasnodar region, Black sea coast of the Caucasus) in April 17-May 15 2018. Localities of 117 collection: Veseloe (43.41, 39.98), Central District (43.58, 39.73), Adler (43.42, 39.94), valley of Agur 118 river (43.56, 39.83), Golovinka (43.79, 39.47), Razbityj Kotel (43.69, 39.73). The beetles were collected 119 by shaking of different trees and shrubs branches and sweep-netting of grasses and collected by hand. 120 Probably all these specimens have overwintered, since the larvae were firstly detected on May 1 and 121 pupae on May 11. No specimens of young generation with soft elytra were detected. All beetles were 122 placed in plastic containers and kept alive at a temperature of about +4°C.

123 Screening for parasites

In mid-June each specimen was examined under a stereomicroscope to detect if it was infested with parasites. First, the ladybird was examined externally from above and from below to determine the presence of ectoparasites. Then its elytra were removed to detect if tergites were infested with mites or other parasites. Then abdomen of each specimen was dissected to find endoparasites. This method allows to detect different parasites in one specimen. Dissection was performed in 0.9% NaCl solution. Collected nematodes were heat-killed (at 65°C) and fixed with TAF solution. Permanent slides of nematodes in anhydrous glycerin were prepared following the Seinhorst method [22]. Totally 60 permanent slides were

131 made. Permanent nematode slides and TAF-fixed nematodes are kept in the Helminthological Museum

132 of the Russian Academy of Sciences (Moscow). Morphometric analysis encompassing measurements of

133 common nematode body features was carried out on five fixed nematode specimens of each life stage,

- 134 employing Zeiss Jenaval microscope. Photographs were taken employing Leica 5500B microscope.
- 135 Identification

136 Primarily the identification of the parasites was based on morphological features. Identification of

137 *Parasitylenchus bifurcatus* was confirmed through nucleotide sequences analysis. The nematodes

138 recovered from the ladybird haemocoel were frozen individually in sterile 0.7 ml Eppendorf tubes for

139 DNA extraction, which was performed according to Holterman *et al.* [23]. The worm-lysis solution (950

140 μl of a mixture of 2 ml of 1M NaCl, 2 ml of 1M Tris-HCl, pH 8 plus 5.5 ml of deionised water plus 10 μl

141 of mercaptoethanol and 40 μ l of proteinase K, 20 mg ml⁻¹) was prepared directly before DNA extraction.

142 Aliquots of 25 µl of sterile water and 25 µl of worm-lysis solution were added to each tube with a

143 nematode and incubated at 65°C for 90 min. The tubes containing the homogenate were then incubated at

144 99°C for 5 min to deactivate proteinase K. About 1.0 μl of homogenate was used as PCR template.

PCR reactions were performed using Encyclo Plus PCR kit (Evrogen[®], Moscow, Russia) according to the manufacturer's protocol. Primer pairs Nem18S_F (5'-CGC GAA TRG CTC ATT A CA ACA GC-3') and 26R (5'-CAT TCT TGG CAA ATG CTT TCG-3') were used to obtain partial (about 900 bp long) sequence of 5' half of the mitochondrial 18S rDNA [24]. PCR cycling parameters included primary denaturation at 94°C for 5 min followed by 34 cycles 94°C for 45 s, 54°C for 60 s and 72°C for 1 min, followed by post-amplification extension at 72°C for 3 min.

A pair of primers D2A (5'-ACA AGT ACC GTG AGG GAA AGT TG -3') and D3B (5'-TCG GAA GGA ACC AGC TAC TA-3') was used to amplify approx. 800 bp long sequence of D2D3 expansion segment of 28S rDNA [25]. PCR cycling parameters included denaturation at 95°C for 3 min, followed by 35 cycles of 94°C for 30 s, 54°C for 35 s, and 72°C for 70 s and followed by postamplification extension at 72°C for 5 min.

156 PCR products were visualised in 1% agarose gel. Then bands containing all obtained PCR product 157 were excised from 0.8% agarose gel for DNA extraction with Wizard SV Gel and PCR Clean-Up System 158 (Promega, Madison, USA). Samples were directly sequenced using the same primers as used for primary 159 PCR. The sequences were combined and aligned using the Clustal X program after the addition of 160 sequences from the GenBank [26]. Similar sequences were searched for in NCBI GenBank with BLAST 161 algorithm [27]. Subsequently, the sequences were edited using the Genedoc 2.7 program [28], to prepare 162 a file for the analysis in MEGA7.0.14 [29]. Phylogenetic trees were obtained with different methods (MP 163 - maximum parsimony, NJ - neighbour joining and ML - maximum likelihood) and pairwise nucleotide 164 differences were calculated. Obtained sequences were analysed in with three methods: maximum 165 parsimony (MP), neighbor joining (NJ) and maximum likelihood (ML). Obtained sequences were 166 deposited in GenBank MH718837 for the 18S rDNA sequence and MH722215 for the 28S rDNA.

167 Collection and external examination of other ladybird species

168 Since Hesperomyces virescens fungi can develop not only on H. axyridis, but also on over 30 other

169 ladybirds [1], we decided to examine other ladybirds collected by the same methods and in the same

170 localities as *H. axyridis*. Four hundred specimens of ladybirds of 31 other species (both native and

171 introduced) have been collected and screened for ectoparasitic fungi. The list of these ladybird species:

172 Adalia bipunctata (Linnaeus), Anisosticta novemdecimpunctata (Linnaeus), Calvia decemguttata

173 (Linnaeus), Chilocorus bipustulatus (Linnaeus), Chilocorus renipustulatus (Scriba), Coccinella

174 quinquepunctata Linnaeus, Coccinella septempunctata Linnaeus, Coccinula quatuordecimpustulata

175 (Linnaeus), Cryptolaemus montrouzieri Mulsant, Exochomus quadripustulatus (Linnaeus), Halyzia

176 sedecimguttata (Linnaeus), Harmonia quadripunctata (Pontoppidan), Hippodamia variegata (Goeze),

177 Lindorus lophanthae (Blaisdell), Nephus bipunctatus (Kugelann), Parexochomus nigromaculatus

178 (Goeze), Propylea quatuordecimpunctata (Linnaeus), Psyllobora vigintiduopunctata (Linnaeus), Rodolia

179 cardinalis (Mulsant), Scymnus frontalis (Fabricius), Scymnus haemorrhoidalis Herbst, Scymnus

180 interruptus (Goeze), Scymnus subvillosus (Goeze), Scymnus suturalis Thunberg, Serangium montazerii

- 181 Fürsch, Stethorus pusillus (Herbst), Subcoccinella vigintiquatuorpunctata (Linnaeus), Tytthaspis
- 182 sedecimpunctata (Linnaeus), Vibidia duodecimguttata (Poda).
- 183 Results
- 184 We have found three parasitic species on examined *H. axyridis* adults: *Hesperomyces virescens*
- 185 (Ascomycota: Laboulbeniales, Laboulbeniaceae), Parasitylenchus bifurcatus (Nematoda: Tylenchida
- 186 Allantonematidae) and unknown species belonging to Mermithida (Nematoda). All examined H. axyridis
- 187 specimens were free of parasitic mite *Coccipolipus hippodamiae*.
- 188 *Hesperomyces virescens* Thaxt
- 189 Characteristic yellowish-greenish thalli of Hesperomyces virescens were detected on the host integument

190 (Fig 2). Their morphology corresponds to the detailed description by De Kesel [30]. Identification was

191 confirmed by mycologist E.Yu. Blagoveshchenskaya.

- 192
- Fig 2. *Hesperomyces virescens* on *Harmonia axyridis*. (a) thalli (bar 50 μm), (b) ladybird covered with
 thalli (bar 3 mm).

195

196 Specimens of *H. axyridis* infested with *H. virescens* were found in all six localities, where the beetles 197 were examined. The distance between the most western locality (Golovinka) and the most eastern 198 (Veseloe) is more than 60 km. The thalli were found in 112 (53%) adults of H. axyridis (Table 1) and 199 were situated on elvtra, pronotum, sternites, legs and mouthparts of the beetles. It seems that H. 200 virescens does not kill or significantly damage H. axyridis, since even the beetles covered with large 201 number of fungi thalli moved actively. Males were infested more often than females (62% and 48% 202 respectively). No signs of ectoparasitic fungi have been detected on 400 examined adults of other 203 ladybird species.

Table 1. Infestation of *H. axyridis* with *Hesperomyces virescens* and *Parasitylenchus bifurcatus*.

	Number	Number of	Number of	Number of	Total
	rumoer				10101
	of	specimens	specimens	specimens	
	specimens	infested with	infested with	infested with	
	without	only	only	P. bifurcatus &	
	parasites	Hesperomyces	Parasitylenchus	H. virescens	
		virescens	bifurcatus		
Males	22	37	1	1	61
Females	69	67	9	7	152
Total	91	104	10	8	213

205

206 Parasitylenchus bifurcatus Poinar, Steenberg

Parasitic nematodes of the family Allantonematidae were found in *H. axyridis* 18 specimens collected at three out of six sampling locations (see Fig 1). Three different nematode life cycle stages, including a subsequent generation parasitic female, vermiform (infective) female and male, were found in the sampled beetles. The number of females of the subsequent generation varied from 5 to 32 per a beetle. The number of vermiform nematode specimens varied considerably, sometimes being as high as about a couple of hundred per a beetle.

Morphometric analysis of nematodes by standard body features allowed to identify the nematodes extracted from ladybirds as *Parasitylenchus bifurcatus* Poinar, Steenberg, 2012 (Table 2). The characteristic features of *P. bifurcatus* nematodes were as follows: a straight stylet lacking basal thickenings, a forked tail tip in the vermiform females and juvenile males, spicules straight, wedgeshaped or triangular, with narrow bursa and gubernaculum (Fig 3). We found that subsequent generation of parasitic females also has forked tail tip.

- 219 Table 2. Morphometric characteristics of the nematode *P. bifurcatus* specimens, isolated from the
- 220 ladybird Harmonia axyridis collected in Sochi.
- 221

Character	Parasitic females of the	Vermiform (infective)	Males (n= 5)
	subsequent generation	females (n= 5)	
	(n= 5)		
Body length, µm	1118.0 (930.0–1660.0)	590.0 (530.0-670.0)	403.0 (396.0-480.0)
Body width, µm	124.0 (85.0–184.0)	12.6 (12.0–13.0)	15.0(14.0–16.0)
Stylet length, µm		11.5 (11.0–12.0)	9.0 (8.0–11.0)
Head to excretory	155.4 (141.0–165.0)	50.6 (42.0–57.0)	66.0(62.0-75.0)
pore distance, µm			
Vulva position, %	89.4 (88.0–93.0)	88.0 (87.0–90.0)	
Tail length, µm	39 (25.0-48.0)	33.2 (30.0–37.0)	34.4(25.5–40.0)
Spicule length, µm			12(11–13)

222

223

224

Fig 3. *Parasitylenchus bifurcatus*: (a) subsequent generation female of *P.bifurcatus* (bar 98 μm), (b)
subsequent generation female, tail (bar 28 μm), (c) vermiform (infective) female, (d) tail (bar 12 μm), (d)
male, tail (bar 12 μm).

228

Parasitic females of the subsequent generation, sampled in Sochi, were close in the size of their body to the specimens of *P. bifurcates* from Denmark [19]. Their body length of the former was 1118.0 μ m (930.0–1660.0) and the body width was 124 μ m (85.0–184.0), compared with 1300 μ m (920–1600)

232	and 195 μ m (158–271) respectively in Danish specimens. In the population of <i>P. bifurcates</i> , described in
233	Slovenia [21], females of the subsequent generation were smaller: 886.4 μ m (782.0–1098.0) in length and
234	72.3 μ m (59.0–81.0) in width, compared with females from the Sochi population.

235 The BLAST-search of similar nucleotide sequences in NCBI GenBank was performed for all 236 three obtained sequences. These were the sequences of different clones and isolates of Parasitylenchus 237 bifurcatus, which were found as the closest to the obtained 18S rDNA sequence of nematodes from 238 Harmonia axyridis ladybirds from Russian Caucasus. All the similar sequences detected by BLAST-239 search were downloaded and used for comparison. Under all methods of analysis obtained sequence was 240 a member of strongly supported clade (100% bootstrap support) consisting of P. bifurcatus sequences 241 plus a sequence of unidentified Allantonematidae (Fig 4). Obtained sequence of H. axyridis parasite from 242 Russian Caucaus was 100% identical to some published sequences of P. bifurcatus (e.g. clones '314j', 243 '3j4i' and 'PaTyBif1'). Remarkably, an unidentified allantonematid nematode, the sequence of which is 244 deposited as JQ941710 was found to be identical to that obtained in the course of our study was found in 245 2010 in Germany in the haemocoel of H. axvridis (E.L. Rhule, unpublished). It seems, that these 246 nematodes from Germany also belong to the species P. bifurcatus. All other known 18 rDNA sequences 247 of P. bifurcatus differ from our sequence in one or two nucleotides. Several clades demonstrated close, 248 but not securely resolved relationships with the clade containing sequences of P. bifurcatus. Under all 249 methods of analysis the 18S rDNA sequence of *Howardula phyllotretae* Oldham, 1933 is in sister 250 relationships with P. bifurcatus clade (Fig 4). One such clade consists of the sequences of parasitic 251 tylenchids of fleas: Rubzovinema Slobodyanyuk, 1991, Spilotylenchus Launay, Deunff & Bain, 1983 and 252 Psyllotylenchus Poinar & Nelson, 1973 (Fig 4). Another related clade was represented by species of 253 Deladenus Thorne, 1941 and unidentified Tylenchomorpha gen.sp. Other sequences of Howardula Cobb, 1921 were most distant from P. bifurcatus (different in 59-60 bp) in our analysis and served as a root for 254 255 obtained cladograms.

256

The known and deposited in NCBI GenBank 28S rDNA sequences of entomoparasitic tylenchids

are less numerous than 18S rDNA data. Obtained cladogram demonstrates close relationships of studied

258 nematode with two other sequences obtained for unidentified species of *Parasitylenchus*: DQ328729 and

259 KM245038 (Fig 5). Both are related to the parasites of bark beetles in Russia and Czech Republic,

- 260 correspondingly. In the level of nucleotide differences these two Parasitylenchus sequences are the
- 261 closest to nematodes from *H. axyridis* (87 and 90 bp), when the nucleotide differences with all other
- studied entomoparasitic tylenchids exceed 100 bp. The sequence of Howardula phyllotretae together
- with that of Anguillonema amolensis Mobasseri, Pedram et Pourjam, 2017 are forming a clade which is
- 264 in sister position to *Parasitylenchus* Micoletzky, 1922 clade under all methods of analysis (Fig 5). As in

265 18S rDNA cladogram the sequences of flea parasites (Rubzovinema, Spylotylenchus, Psyllotylenchus) are

266 forming well supported clade (Fig 5). The relationships of this latter clade with Parasitylenchus

267 (*Howardula phyllotretae* + *Anguillonema amolense*) clade are strongly supported.

268

269 Fig 4. The relationships of Parasitylenchus bifurcatus from Sochi, Russia with other groups of insect-

associated tylenchids inferred from analysis of partial 18S rDNA. Bootstrap support is given near

271 corresponding nodes in the format MP/NJ/ML.

272

273 Fig 5. The relationships of Parasitylenchus bifurcatus from Sochi, Russia with other groups of insect-

associated tylenchids inferred from analysis of partial 28S rDNA. Bootstrap support is given near

corresponding nodes in the format MP/NJ/ML.

276 Parasitylenchus bifurcatus was found in 8% of all dissected adults of *H. axyridis*. Prevalence in females 277 and males was 10% and 3% respectively, but this difference is not reliable because of small number of 278 infested specimens. The incidence of the ladybird infestation by *Parasitylenchus bifurcatus* in Sochi is 279 lower, compared with the results of the former studies in the countries of Europe: the incidence of 280 infested ladybirds was as high as up to 35% in Denmark [19], and even up to 47% in Czech Republic [1]. 281 *Parasitylenchus bifurcatus* was found also in specimens infested with *Hesperomyces virescens* and free

of fungi, and no correlation between infestation of specimens with *Parasitylenchus bifurcatus* and

283 Hesperomyces virescens was observed (Fig 6).

284

Fig 6. Proportion of *Harmonia axyridis* adults infested with *Hesperomyces virescens* and *Parasitylenchus*

286 *bifurcatus*. Number of examined specimens is indicated in brackets.

287

288 The only one specimen of a nematode, belonging to the order Mermithida, was found in the only ladybird

289 specimen sampled in Veseloe. This was the first documented case of the ladybird H. axyridis infestation

290 by a parasitic nematode of this order in nature. Formerly von Linstow [31] reported *Mermis nigrescens*

291 Dujardin (Mermithida: Mermithidae) to be a parasite of the other ladybird species, *C. septempunctata*.

292 The infestation of Adonia variegata, C. septempunctata und Semiadalia undecimnotata with unidentified

293 mermithids in South-East of France was reported by G. Iperti [32]. H. Kaiser and W.R. Nickle [33]

294 described the Coccinella septempunctata infestation with Hexamermis sp. in Styria, Austria. An

295 overview of reports of coccinellid infestation with mermithid nematodes was presented by G.O. Poinar Jr

[34]. A study, specifically focused on the search of parasitic nematodes of this order in the Sochi area,

297 their identification to the species, and determination of the incidence of ladybird infestation is planned for

future.

299

Discussion

Haelewaters et al. [1] supposed that infection of young populations of *H. axyridis* in some newly invaded regions by *H. virescens* and *P. bifurcatus* (North America, Netherlands), occurred as a result of acquisition of native natural enemies. In the Caucasus co-invasion hypothesis is more relevant than acquisition hypothesis. First, *H. virescens* and *P. bifurcatus* were absent in the region and appeared at the same time with their host. Locality of the current records of the parasites (Sochi) is situated at more than 1400 km from the nearest known localities of both parasitic species. Second, other examined ladybird species are not infested with *H. virescens* in the Caucasus. Other ladybirds were not dissected to find

307 nematodes, but, since *H. axyridis* is the only known host of *P. bifurcatus*, there is no reason to suggest an 308 *H. axyridis* got infected from other ladybird species. Third, an analysis of the phylogenetic relationships 309 of *Parasitylenchus bifurcatus* based on 18S rDNA demonstrated complete identity of 18S rDNA 310 sequence of these nematodes from Russian Caucasus with some strains or clones found in Western and 311 Southern Europe. Cautiously, such identity can be considered as an indication on the possible transfer of 312 parasites together with insect hosts from Western part of Eurasia.

313 Since H. axyridis was released for biological control of pests in the Caucasus, it was unclear, how 314 the population of *H. axyridis* appeared: as a result of these releases or as a result of expansion of its 315 European range [11, 16]. The study of parasites has shed light on this question. Both *H. virescens* and *P.* 316 bifurcatus affect only adults, do not occur on other life stages and exclusively spread by activities of the 317 host [18]. Transmission takes place only from adult to adult, therefore the direct contact between different 318 generations of beetles is necessary for to maintain the life cycle of the parasites. Since different 319 generations are kept separately in laboratory culture [35], it is free of parasites. Therefore, detection of H. 320 virescens and P. bifurcatus indicates that population of H. axvridis in the Caucasus cannot derive 321 exclusively from specimens released from laboratory culture. At least part of ancestors of the Caucasian 322 population of Harmonia axyridis are from European invasive range. On the other hand, admixture of 323 released specimens is not excluded, since H. axyridis was released in Sochi for several decades. The 324 complex invasion scenaria are common place for alien insects in general [36] and for Harmonia axyridis 325 in particular [37]. This case and some other recent studies [2] confirm that parasitological analysis is a 326 promising approach of revealing of invasion rotes.

Roy et al [17] supposed that connection of *H. virescens* with *H. axyridis* coupled with the rapid expansion of *H. axyridis* globally suggests that this parasite will continue to spread throughout the rest of the world. Spread of *H. virescens* to the Caucasus confirms this suggestion.

Hesperomyces virescens is found to be widespread and common on *H. axyridis* at the Black sea coast of the Caucasus. But in spite of high prevalence of *H. virescens* on *Harmonia axyridis* we have not found it on other potential hosts in the Caucasus. No signs of Laboulbeniales ectoparasites have been

detected on other ladybird species, in spite seven of them were indicated by Ceryngier and Twardowska 333 334 [38] as hosts of *Hesperomyces virescens* in other regions: *Adalia bipunctata, Chilocorus bipustulatus,* 335 renipustulatus, Coccinula quatuordecimpustulata, Propylea Chilocorus quatuordecimpunctata, 336 Psyllobora vigintiduopunctata and Tytthaspis sedecimpunctata. The same situation was previously 337 observed by A. De Kesel in Europe in spite some potential hosts overwintered at the same sited with 338 Harmonia axyridis [1]. Also Cottrell and Riddick [18] found reduced interspecific transmission of H. 339 virescens (under laboratory conditions) and hence suggested the existence of hostadapted isolates or 340 strains of H. virescens.

341 Hesperomyces virescens does not seriously damage its hosts [17], but Parasitylenchus bifurcatus 342 capable of causing significant harm [19]. Co-infection of H. axyridis with H. virescens and P. bifurcatus 343 was recorded in Netherlands and positive association between these parasites was detected that correlated 344 with a reduced number of live beetles [20]. No correlation between infestation by *H. virescens* and *P.* 345 *bifurcatus* has been yet recorded in the Caucasus. But the number of collected specimens was small, so 346 possibility of such correlation cannot be ruled out. The study of co-infection of *H. axyridis* by these two 347 parasites is an intriguing subject for future studies, since co-infections might result in lower survival 348 rates.

349 The obtained nucleotide sequences are significant as the confirmation of primary parasitic 350 nematode identification based on morphological features. Several sequences for 18S rDNA of P. 351 *bifurcatus* are deposited in NCBI GenBank, and newly obtained ones for the specimens from Sochi are 352 identicval with some of these. Unlike 18S rDNA data, those for large ribosomal subunit (28S rDNA) of 353 Parasitylenchus nematodes are quite scarce and can only prove the sufficient informative value of this 354 locus for phylogenetic studies of entomoparastic tylenchids. These data securely demonstrated the 355 clustering of P. bifurcatus sequence with two known sequences of Parasitylenchus nematodes. The 356 results of both 18S and 28S analyses revealed some incongruence in contemporary taxonomy of these 357 nematodes. Thus, it is obvious in the obtained phylogenetic trees that some genera are polyphyletic: we

358 can find the species of Howardula in three clades of 18S rDNA cladogram, Deladenus sequences in two

359 clades of 18S and 28S rDNA cladograms.

360 Conclusions

- 361 1. The population of *H. axyridis* in the Caucasus recently invaded by this species is infested with
- 362 two parasite species, which are recorded for the Caucasus and Russia for the first time:
- 363 *Hesperomyces vinirescens* and *Parasitylenchus bifurcatus*. Probably these parasites have

364 appeared in the region as a result of co-invasion with *H. axyridis*.

- 365 2. Population of *H. axyridis* in the Caucasus appeared as a result of expansions of European range,
- 366 as 18S rDNA sequences of Caucasian *Parasitylenchus bifurcatus* and those from Western Europe
- 367 are 100% identical. It cannot derive exclusively from specimens released for biological control of
- 368 pests, because laboratory cultures are free of these parasites.
- 369 3. Though *Hesperomyces vinirescens* develops on many ladybird species in other regions, its only
 370 known host in the Caucasus is *Harmonia axyridis*.
- 4. An unidentified species of the order Mermithida is recorded on *Harmonia axyridis* in the
- 372 Caucasus. It is the first documented case of the ladybird *H. axyridis* infestation by a parasitic
- 373 nematode of this order in nature.

374 Acknowledgements

375 We are grateful to E.Yu. Blagoveshchenskaya (Department of Mycology and Algology of Moscow State

376 University) for identification of the parasitic fungi and taking the photo of thalli, to P. Ceryngier (Faculty

- 377 of Biology and Environmental Sciences, Cardinal Stefan Wyszyński University) for valuable
- information and to T.A. Mogilevich for the help in screening of the ladybirds for parasites.

379

380 **References**

381 1. Haelewaters D, Zhao SY, Clusella-Trullas S, Cottrell TE, De Kesel A, Fiedler L, et al. Parasites

- of *Harmonia axyridis*: current research and perspectives. Biocontrol (Dordr). 2017;62: 355–371.
 doi: 10.1007/s10526-016-9766-8.
- Reshetnikov AN, Sokolov SG, Protasova EN. Detection of a neglected introduction event of the
 invasive fish *Perccottus glenii* using parasitological analysis. Hydrobiologia. 2017;788(1): 65–73.
- doi: 10.1007/s10750-016-2987-0.
- Vilcinskas A, Stoecker K, Schmidtberg H, Röhrich CR, Vogel H. Invasive harlequin ladybird
 carries biological weapons against native competitors. Science. 2013;340(6134): 862–863. doi:
 10.1126/science.1234032.
- Orlova-Bienkowskaja MJ, Ukrainsky AS, Brown PMJ. *Harmonia axyridis* (Coleoptera:
 Coccinellidae) in Asia: a re-examination of the native range and invasion to southeastern
 Kazakhstan and Kyrgyzstan. Biol Invasions. 2015;17(7): 1941–1948. doi: 10.1007/s10530-015 0848-9.
- 394 5. Roy HE, Brown PMJ, Adriaens T, Berkvens N, Borges I, Clusella-Trullas S, et al. The harlequin
 395 ladybird, *Harmonia axyridis*: global perspectives on invasion history and ecology. Biol Invasions.
 396 2016;18(4): 997–1044. doi: 10.1007/s10530-016-1077-6.
- Roy HE, Adriaens T, Isaac NJ, Kenis M, Onkelinx T, Martin GS, et al. Invasive alien predator
 causes rapid declines of native European ladybirds. Divers Distrib. 2012;18(7): 717–725. doi:
 10.1111/j.1472-4642.2012.00883.x.
- 400 7. Ukrainsky AS, Orlova-Bienkowskaja MJ Expansion of *Harmonia axyridis* Pallas (Coleoptera:
 401 Coccinellidae) to European Russia and adjacent regions. Biol Invasions. 2014;16(5): 1003–1008
- 402 doi: 10.1007/s10530-013-0571-3.
- 403 8. Goryacheva I, Blekhman A, Andrianov B, Romanov D, Zakharov I. *Spiroplasma* infection in
- 404 *Harmonia axyridis*-Diversity and multiple infection. PLoS One. 2018;13(5): e0198190. doi:
- 405 10.1371/journal.pone.0198190.
- 406 9. Kuznetsov VN. Far Eastern coccinellids in the Transcaucasia. Zashchita rasteniy. 1988;5: 19. [In
 407 Russian]

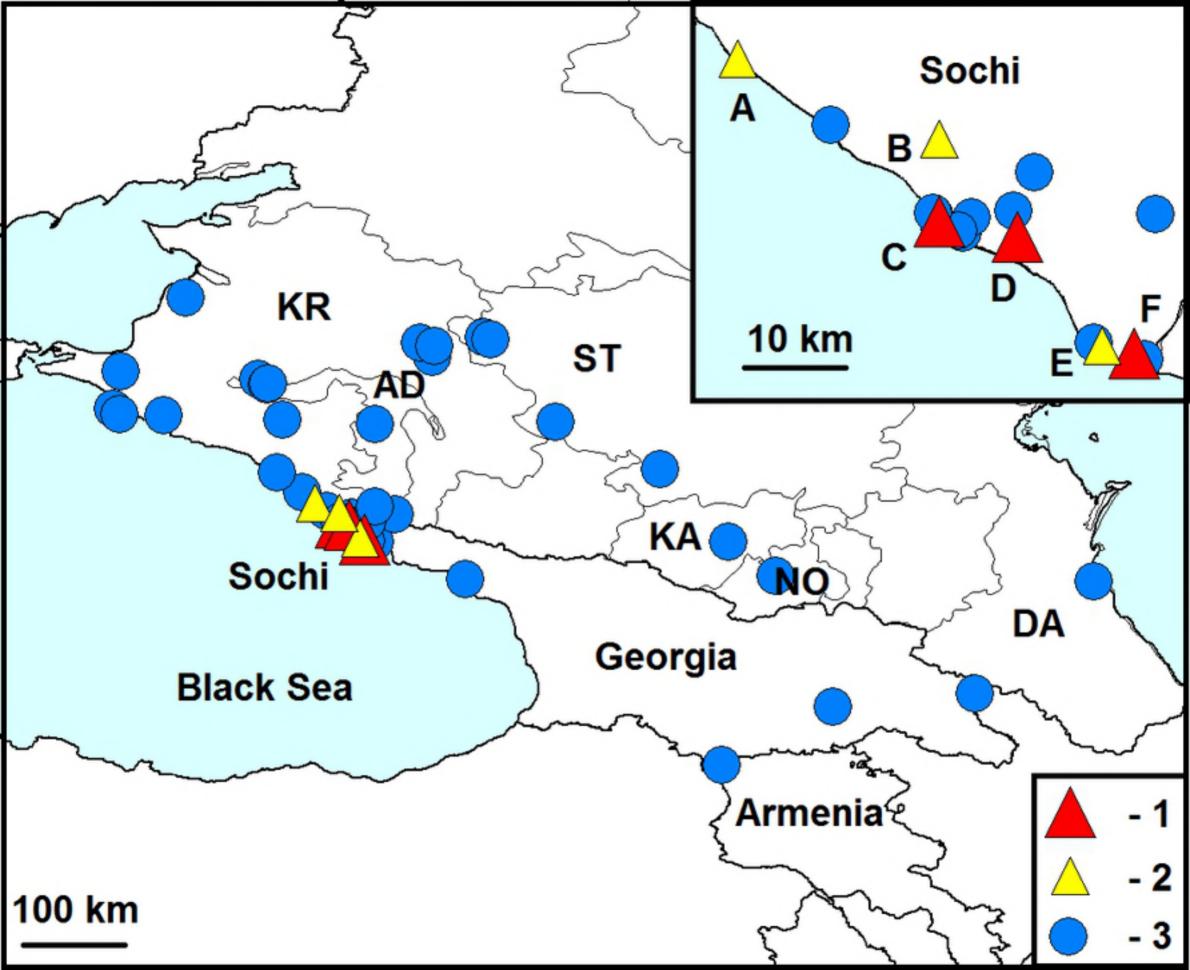
408	10. Mogilevich TA. My experiments with the ladybird Harmonia axyridis [cited 6 August 2018]. In:
409	Beetles and coleopterologists 2012. Available from:
410	https://www.zin.ru/aNIMAliA/Coleoptera/rus/mogilev1.htm. [in Russian]
411	11. Belyakova NA, Reznik SY. First record of the harlequin ladybird, Harmonia axyridis
412	(Coleoptera: Coccinellidae) in the Caucasus. Eur J Entomol. 2013;110(4): 699-702.
413	12. Orlova-Bienkowskaja MJ, Mogilevich TA. The first record of Asian ladybird Harmonia axyridis
414	(Pallas, 1773) (Coleoptera: Coccinellidae) in Kabardino- Balkaria and the history of the expansion
415	of this alien species in the Caucasus and south of European Russia in 2002–2015. Caucasian
416	Entomological Bulletin. 2016;12(1): 93–98. [In Russian]
417	13. Alekseev SK, Butaeva FG. Ladybirds (Coleoptera: Coccinellidae) of the vicinity of Alagir City
418	(Republic of North Ossetia-Alania). In: Actual problems of chemistry, biology and biotechnology:
419	Proceedings of the 10th All-Russian Scientific Conference (11-13 May 2016). North Ossetia
420	State University. Vladikavkaz: Publishing house of North Ossetia State Uiversity; 2016. pp. 71-
421	75. [in Russian]
422	14. Kalashian MY, Ghrejyan TL, Karagyan GH. Harlequin ladybird Harmonia axyridis Pall.
423	(Coleoptera, Coccinellidae) in Armenia. Russ J Biol Invasions. 2017;8(4): 313-315. doi:
424	10.1134/S207511171704004X.
425	15. Bugaeva LN, Ignat'eva TN, Novikov YuP., Kashutina EV. Problem of protection of vegetables at
426	organic food agriculture. Newsletter of the East Palaearctic Regional Section of the
427	Internationalorganizations for the biological control of harmful animals and plants, 2011;42: 32-
428	35. [in Russian]
429	16. Orlova-Bienkowskaja MJ. The outbreak of harlequin ladybird Harmonia axyridis (Pallas, 1773)
430	(Coleoptera, Coccinellidae) in the Caucasus and Possible Sources of Invasion. Russ J Biol
431	Invasions. 2014;5(4): 275–281. doi: 10.1134/S2075111714040055.

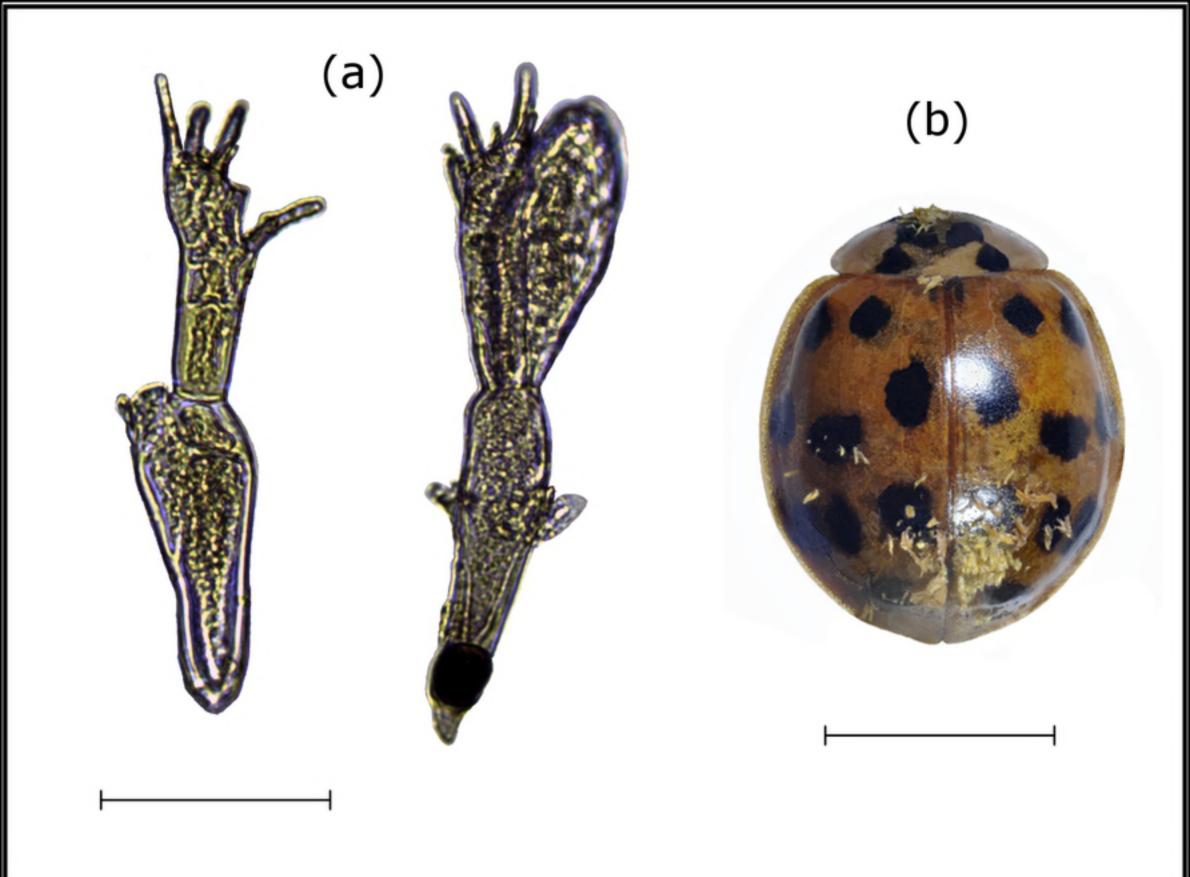
- 432 17. Roy HE, Rhule E, Harding S, Handley LJL, Poland RL, Riddick EW, Steenberg T. Living with
- 433 the enemy: parasites and pathogens of the ladybird *Harmonia*
- 434 *xyridis*. Biocontrol (Dordr). 2011;56(4): 663–679. doi: 10.1007/s10526-011-9387-1.
- 435 18. Cottrell TE, Riddick EW. Limited transmission of the ectoparasitic fungus *Hesperomyces*436 *virescens* between lady beetles. Psyche. 2012;814378. doi: 10.1155/2012/814378.
- 437 19. Poinar GO, Steenberg T. *Parasitylenchus bifurcatus* n. sp.(Tylenchida: Allantonematidae)
- 438 parasitizing *Harmonia axyridis* (Coleoptera: Coccinellidae). Parasit Vectors. 2012;5(1): 218. doi:
 439 10.1186/1756-3305-5-218.
- 440 20. Raak-van den Berg CL, van Wielink PS, de Jong PW, Gort G, Haelewaters D, Helder J, Karssen
- 441 G, van Lenteren JC. Invasive alien species under attack: natural enemies of *Harmonia axyridis* in
- 442 the Netherlands. Biocontrol (Dordr). 2014;59: 229–240. doi: 10.1007/s10526-014-9561-3.
- 443 21. Gerič Stare B, Širca S, Urek G. First report of nematodes *Parasitylenchus bifurcatus* Poinar &
 444 Steenberg, 2012 parasitizing multicolored Asian lady beetle *Harmonia axyridis* (Pallas, 1773) in
- 445 Slovenia. Acta Agric Slov. 2017;109(2): 457–463. doi: 10.14720/aas.2017.109.2.28.
- 446 22. Seinhorst JW. A rapid method for the transfer of nematodes from fixative to anhydrous glycerin.
 447 Nematologica. 1959;4(1): 67–69.
- 448 23. Holterman M, van der Wurff A, van den Elsen S, van Megen H, Bongers T, Holovachov O, et al.
 449 Phylum-wide analysis of SSU rDNA reveals deep phylogenetic relationships among nematodes
 450 and accelerated evolution towards crown clades. Mol Biol Evol. 2006;23: 1792–1800. doi:
 451 10.1093/molbev/msl044.
- 452 24. Floyd RM, Rogers AD, Lambshead PJD, Smith CR. Nematode-specific PCR primers for the 18S
 453 small subunit rRNA gene. Mol Ecol Notes. 2005;5: 611–612. doi: 10.1111/j.1471454 8286.2005.01009.x.
- 455 25. Nadler SA, Bolotin E, Stock SP Phylogenetic relationships of *Steinernema* Travassos, 1927
 456 (Nematoda: Cephalobina: Steinernematidae) based on nuclear, mitochondrial and morphological
 457 data. Systematic Parasitology. 2006;63: 161–181.

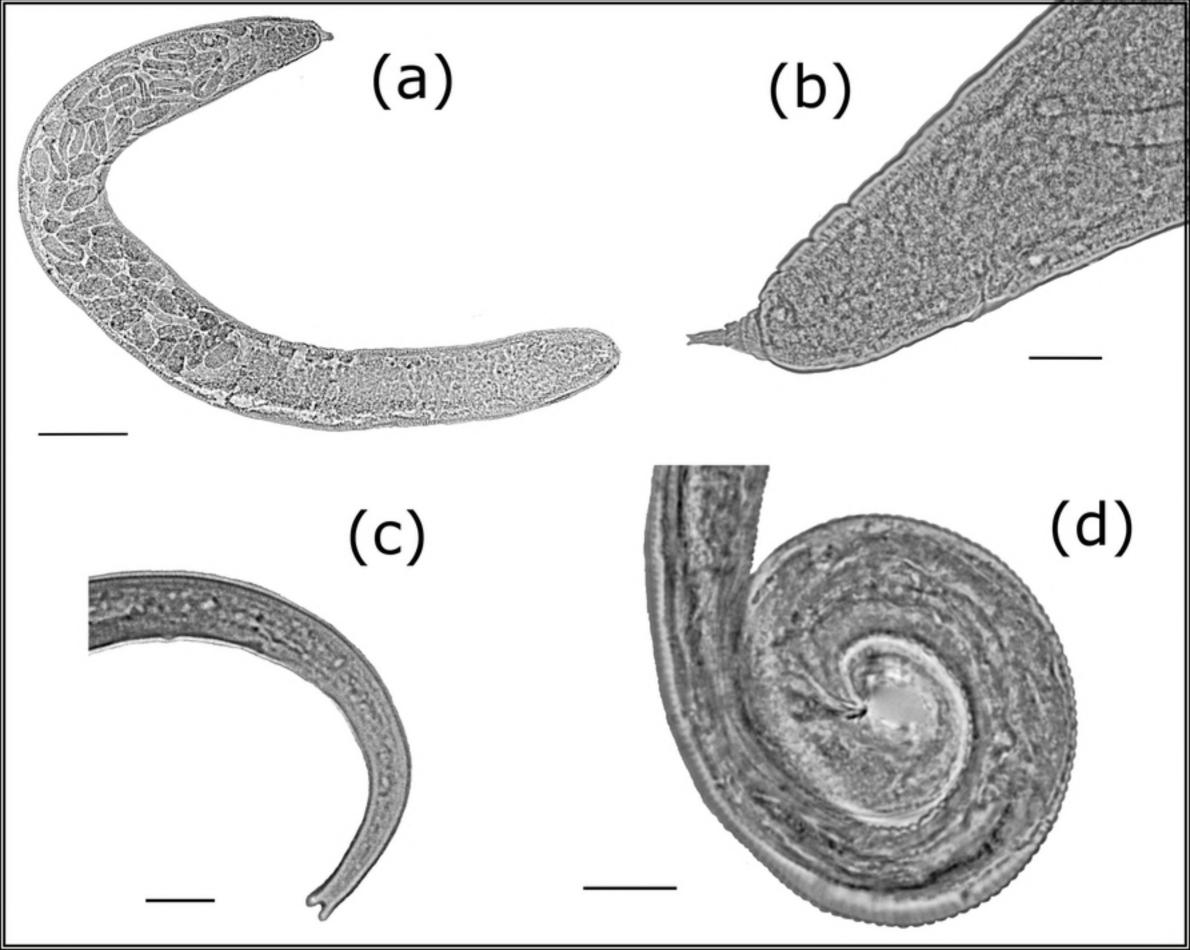
- 458 26. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins, DG. The Clustal X windows
- 459 interface: flexible strategies for multiple sequence alignment aided by quality analysis tools.
- 460 Nucleic Acids Res. 1997;24: 4876–4882. doi: 10.1093/nar/25.24.4876.
- 461 27. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. J Mol
 462 Biol. 1990;215: 403–410.
- 463 28. Nicholas KB, Nicholas HB Jr., Deerfield DW. Multiple Sequence Alignment Editor and Shading
- 464 Utility, Version 2.7.000: 1997. [cited 6 August 2018] Available from:
 465 http://www.psc.edu/biomed/genedoc.
- 466 29. Kumar S, Stecher G, Tamura K. MEGA7: Molecular evolutionary genetics analysis. Version 7.0
 467 for bigger datasets. Mol Biol Evol. 2016;33: 1870–1874. doi: 10.1093/molbev/msw054.
- 468 30. De Kesel A. *Hesperomyces* (Laboulbeniales) and coccinellid hosts. Sterbeeckia. 2011;30: 32–37.
- 469 31. Linstow OFP von. Das Genus Mermis. Archiv für mikroskopische Anatomie und
- 470 Entwicklungsmechanik 1898;53: 149–168.
- 471 32. Iperti G. Les parasites des Coccinelles aphidiphages dans les Alpes-Maritimes et les Basses472 Alpes. Entomophaga. 1964;9: 153–180.
- 473 33. Kaiser H, Nickle WR. Mermithiden (Mermithidae, Nematoda) parasitieren Marienkafer
- 474 (*Coccinella septempunctata* L.) in der Steiermark. Mitteilungen Naturwissenschaftlicher Verein
 475 für Steiermark. 1985: 115–118.
- 476 34. Poinar GO Nematodes for biological control of insects. Boca Raton: CRC Press; 1979.
- 477 35. Izhevsky SS. Introduction and application of entomophages. Moscow: Agropromizdat; 1990. [In
 478 Russian]
- 479 36. Garnas JR, Auger-Rozenberg MA, Roques A, Bertelsmeier C, Wingfield MJ, Saccaggi DL, Roy
- 480 HE, Slippers B. Complex patterns of global spread in invasive insects: eco-evolutionary and
- 481 management consequences. Biol Invasions. 2016;18(4): 935–952. doi: 10.1007/s10530-016-1082482 9.
- 483 37. Lombaert E, Guillemaud T, Lundgren J, Koch R, Facon B, Grez, A, et al. Complementarity of

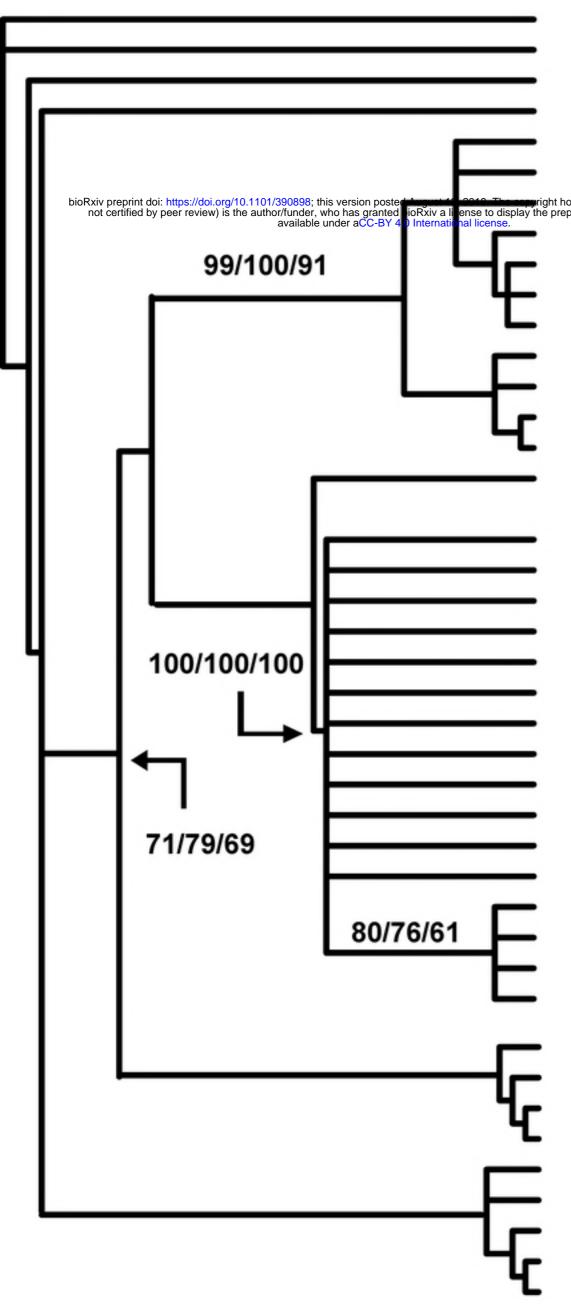
484		statistical treatments to reconstruct worldwide routes of invasion: the case of the Asian ladybird
485		Harmonia axyridis. Mol Ecol. 2014;23(24): 5979-5997. doi: 10.1111/mec.12989.
486	38.	Ceryngier P, Twardowska K. Harmonia axyridis (Coleoptera: Coccinellidae) as a host of the
487		parasitic fungus Hesperomyces virescens (Ascomycota: Laboulbeniales, Laboulbeniaceae): A case
488		report and short review. Eur J Entomol. 2013;110(4): 549-557.
489		
490		Supporting information captions
491		S1 Table. Localities of Harmonia axyridis and its parasites Hesperomyces virescens and

492 *Parasitylenchus bifurcatus* in the Caucasus









Howardula sp. AF519233 Hexatylus sp. KJ636357 Deladenus durus JQ957898 Neotylenchidae gen.sp. KY907662 Rubzovinema sp. KF155281 Rubzovinema sp. KF155283 ense to display the preprint in perpetuity. It is made Rubzovinema sp. KY119823 Panagrolaimus sp. KY119850 Rubzovinema sp. KY119814 Rubzovinema sp. KY119847 Rubzovinema sp. KF373731 Psyllotylenchus sp. KF373733 Spilotylenchus sp. KF373735 Rubzovinema sp. KY119815 Howardula phyllotretae JX291137

> Parasitylenchus bifurcatus LT547722 Parasitylenchus bifurcatus LT547723 Parasitylenchus bifurcatus Sochi, Russia Parasitylenchus bifurcatus KC875398 Allantonematidae JQ941710 Parasitylenchus bifurcatus LT547721 Parasitylenchus bifurcatus LT547720 Parasitylenchus bifurcatus LT547719 Parasitylenchus bifurcatus KC875401 Parasitylenchus bifurcatus KC875400 Parasitylenchus bifurcatus KC875397 Parasitylenchus bifurcatus KC875399 Parasitylenchus bifurcatus LT629307 Parasitylenchus bifurcatus LT629306 Parasitylenchus bifurcatus LT547725 Parasitylenchus bifurcatusLT547726

Deladenus posteroporosus KY098774 Deladenus siricidicola FJ004889 Tylenchomorpha gen.sp. LC147025 Tylenchomorpha gen.sp. LC147027 Fergusobia sp. AY589295 Howardula sp. AF519231 Howardula neocosmicus AF519226 Howardula aoronymphium AF519224 Howardula aoronymphium AY589304

