Distinct higher-order thalamic circuits channel parallel streams of visual information in mice

Corbett Bennett[1*], Samuel D. Gale[1*#], Marina E. Garrett[1,2], Melissa L. Newton[2], Edward M. Callaway[2], Gabe J. Murphy[1], Shawn R. Olsen[1]

1. Allen Institute for Brain Science

2. Salk Institute for Biological Studies

* contributed equally

corresponding author (samg@alleninstitute.org)

MLN's present affiliation is the University of California at Berkeley

Abstract

Higher-order thalamic nuclei, such as the visual pulvinar, play essential roles in shaping cortical dynamics and connecting functionally-related cortical and subcortical brain regions. A coherent framework describing pulvinar function remains elusive due to its anatomical complexity, involvement in diverse cognitive processes, and the limited experimental tools available in many species. We combined large-scale anatomical circuit mapping with high-density electrophysiological recordings to dissect a homolog of pulvinar in mice, the lateral posterior nucleus (LP). We define three LP subregions based on correspondence between connectivity and functional properties. These subregions form parallel corticothalamic loops and contain separate representations of visual space. Silencing visual cortex or the superior colliculus revealed that these input sources drive activity and shape visual tuning in separate LP subregions. By specifying the information carried by distinct circuits through LP and identifying their downstream targets, our data provide a roadmap for understanding pulvinar function in visual processing and behavior.

Introduction

Higher-order thalamus plays a critical role in cortical function, both as a route by which cortical areas communicate and a relay of subcortical input to cortex. The pulvinar, the higher-order thalamic nucleus in the visual system, is reciprocally connected with multiple visual and frontal cortical areas and therefore mediates an indirect pathway for cortico-cortical communication through cortico-thalamo-cortical ("transthalamic") circuits[1]. The pulvinar also receives a dense projection from superficial layers of the superior colliculus (SC), potentially forming a secondary route of information flow from retina to cortex that runs parallel to the retino-geniculate pathway.

Traditionally, the pulvinar was thought to have primarily modulatory roles in visual processing. However, recent studies have shown that inactivating the pulvinar severely attenuates visual responses in primary and higher-order visual cortex and can potently disrupt visual perception and behavior[2,3]. These findings suggest the pulvinar is a key component of the core visual processing hierarchy. In addition, the pulvinar is implicated in diverse sensory and cognitive functions including spatial attention[4,5], detection of threatening stimuli[6], and perceptual

binding[7,8]. The broad range of putative pulvinar functions likely reflects the complexity of its various subregions and interconnectivity with the cortex and other targets. Thus, many fundamental questions about pulvinar function remain unanswered: (1) what are the functionally relevant subdivisions of the pulvinar; (2) what are the relative contributions of cortical and SC input in shaping pulvinar visual responses; (3) what information is carried by the cortex-pulvinar-cortex and SC-pulvinar-cortex pathways?

The lateral posterior thalamic nucleus (LP) in rodents is homologous to the primate visual pulvinar[9]. Like the pulvinar, mouse LP is reciprocally connected with all visual cortical areas, receives a strong projection from SC, and likely consists of several distinct subregions[10–12]. These similarities, combined with the array of tools available in mice, suggest the potential for experiments in mouse LP to resolve outstanding questions about the role of higher-order thalamus in cortical processing. Previous studies in mice have shown that LP neurons encode a variety of sensory and motor signals and convey these to visual cortex[13–15]. However, these studies focused on limited portions of LP and did not relate functional differences to anatomical connectivity. Moreover, none of these studies tested the causal role of cortical or SC input in driving LP visual responses.

Here, we performed comprehensive anatomical input/output mapping and high-density electrophysiological recordings across the full extent of LP to elucidate the relationship between structural and functional organization in the mouse corticothalamic visual system. We quantitatively mapped parallel processing streams that connect specific higher-order visual cortical areas with distinct LP subregions. We identified a posterior subregion that receives driving input from SC and is reciprocally connected with “ventral stream” visual cortical areas, an anterior subregion that receives driving input from primary visual cortex (V1) and is reciprocally connected with “dorsal stream” visual cortical areas, and a medial subregion interconnected with frontal and associational cortices.

Results

Anatomical connectivity mapping divides LP into three broad subregions

LP activity is shaped by input from cortical and subcortical structures and influences diverse downstream targets via its output projections. Thus, functionally-relevant subregions of LP might be defined by distinct patterns of input and output connections. To generate maps of LP connectivity, we used viral tracer injections in source and target regions. We define LP input volumes by quantifying axon labelling in LP resulting from anterograde viral tracer injection into each source region including SC, eight cortical visual areas, and two frontal cortical areas (Fig. 1a, top row; all injections taken from Allen Mouse Brain Connectivity Database). We defined LP output volumes by measuring fluorescence from LP cell bodies labelled by retrograde virus injections into each target region (Fig. 1a, bottom row). Cortical visual areas were precisely targeted based on retinotopic mapping of area borders prior to viral injection. Input and output volumes were mapped to a standardized anatomical coordinate system for comparison (the Allen Institute Common Coordinate Framework, CCF; Fig. 1a).

To quantify the degree to which axons from different source regions overlap in LP, we computed the normalized dot product between each input volume (Fig. 1b). The resulting similarity matrix suggested that specific groups of input sources target distinct LP voxels. To test this idea, we performed hierarchical clustering of LP voxels based on their input. This analysis revealed three statistically significant clusters that differed in their predominant sources of input

(Fig. 1c,d): (1) posterior LP (pLP; cluster 1) receives input from SC and ventral-stream visual cortical areas (LI and POR); (2) anterior LP (aLP; cluster 2) receives input from V1 and dorsal-stream visual cortical areas (AL, RL, AM, PM); and (3) medial LP (mLP; cluster 3) receives input from frontal cortical areas (ACA and ORB). These clusters represent a first-order parcellation of LP into broad zones, which may contain additional fine-scale organization (see Discussion). Similar patterns of axonal labeling were observed in LP when we restricted fluorophore expression to layer 5 or 6 cortical neurons (in Rbp4-KL100 or Nstr1-GN220 Cre mice, respectively) or used a virus that preferentially labels pre-synaptic terminals (rather than fibers of passage; Fig. S1a,b).

To quantify the input/output connectivity of LP, we compared the overlap of anterogradely-labeled axons and retrogradely-labeled cell bodies from each input source and output target (Fig. 1e). This analysis revealed putative transthalamic pathways linking visual areas by indirect projections through LP. We found that most of these pathways are reciprocal: if an LP voxel receives input from a region, it tends to project back to that region (evident from high values along the diagonal of Fig. 1e). However, we also found evidence for relay pathways linking distinct areas (evident from values off the diagonal). For example, mLP voxels (cluster 3) receive input almost exclusively from frontal cortical regions (ACA and ORB) and project back to these regions as well as to dorsal-stream visual cortical areas (PM, AM, RL, AL, Fig. 1d,e). Similarly, aLP voxels (cluster 2) receive most of their input from dorsal-stream visual cortical areas but project both back to these regions and ventral-stream visual cortical areas (LI and POR). Finally, pLP voxels (cluster 1) project to the amygdala (LA) in addition to reciprocal connections with LI and POR, but do not project back to SC. A complementary analysis using maps of LP output defined by anterograde tracer injections in different portions of LP yielded similar results (Fig. S1c-e).

Cortical areas are connected directly through cortico-cortical projections and indirectly through higher-order thalamic regions such as LP (transthalamic pathways). To determine whether the same or different visual cortical areas are connected via cortico-cortical and transthalamic pathways (Fig. 1f, inset), we compared: (1) the density of axons from each cortical area to all the other cortical areas (cortico-cortical projections; Fig. 1g), with (2) the overlap of axons in LP from each cortical area (input volumes) and the cell bodies in LP (output volumes) projecting to each of those areas (transthalamic connections; Fig. 1e). There is a significant correlation between these measures of cortical connectivity (Fig. 1f), suggesting that transthalamic connections of visual cortical areas through LP largely mirror direct cortico-cortical pathways.

Figure 1

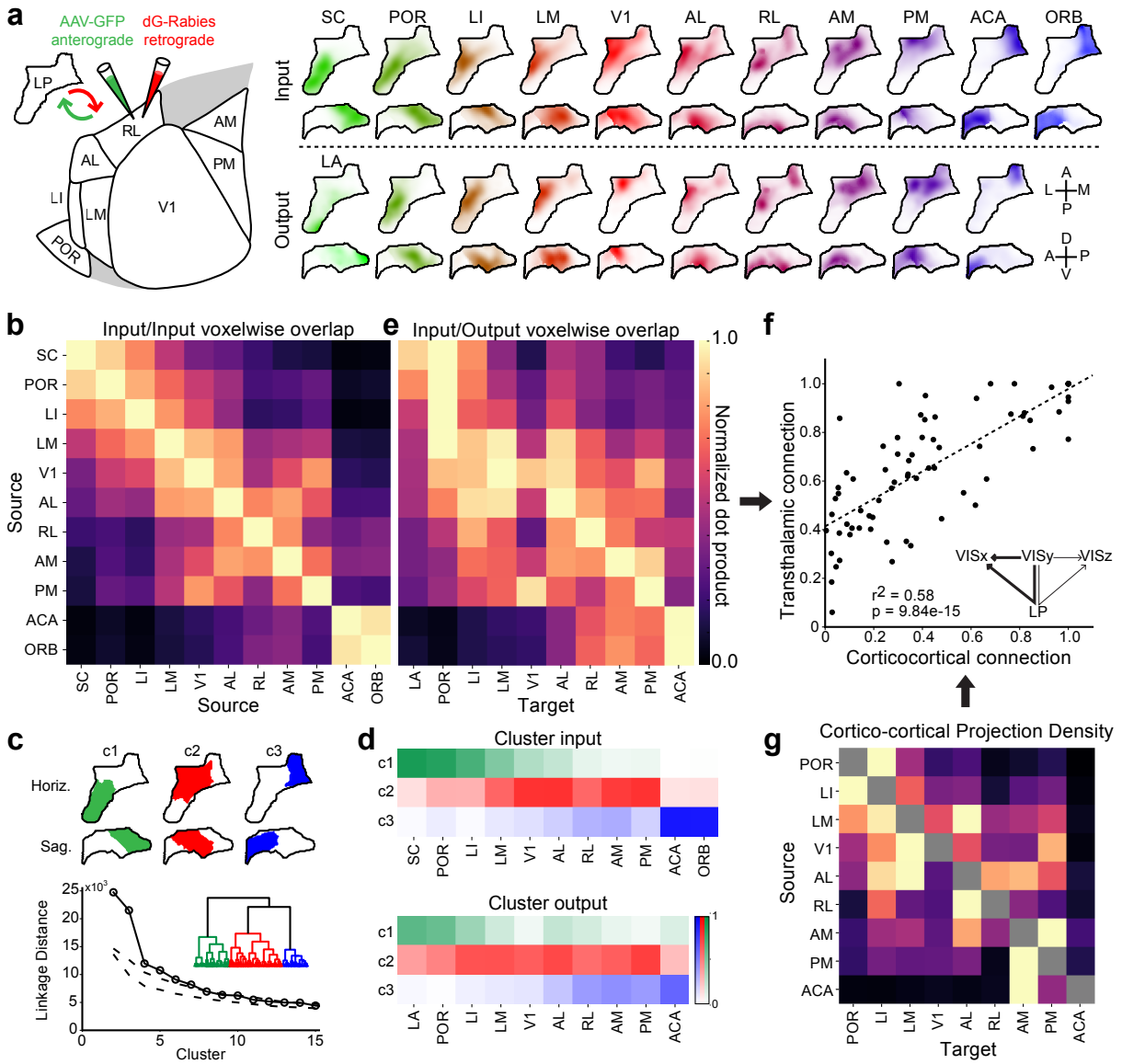

Figure 1. Input/output connectivity reveals three broad LP subregions. (a) Left: schematic of circuit mapping experiments. All regions shown in horizontal plane. Anterograde and retrograde virus injections were in separate mice. Right: LP inputs (defined by anterogradely labeled axons) and outputs (defined by retrogradely labeled somata) from/to various cortical and subcortical regions. The input/output volumes are shown as horizontal and sagittal projections and represent average fluorescence across multiple tracer injection experiments registered in the Allen Institute Common Coordinate Framework (see Methods). **(b)** Overlap of inputs to LP from different sources (normalized voxel-wise dot product). **(c)** Hierarchical clustering LP voxels based on anatomical input using Ward's linkage criterion. Top: horizontal (top) and sagittal (middle) projections of LP voxels belonging to the first three clusters. Bottom: Linkage distance of LP voxels based on pattern of input from 11 sources shown in (b). Dendrogram (inset) shows linkage distance and clusters. Linkage distance for the first 15 clusters are compared to clusters formed from random shuffling of data across voxels for each input source (dashed lines; 1-99% confidence interval). **(d)** Overlap of each cluster with input volumes (top) and output volumes (bottom) for each region. **(e)** Overlap in LP of input and output maps for each source-target area pair (row-

normalized). **(f, g)** Comparison of direct cortico-cortical and indirect cortico-pulvinar-cortical (transthalamic) connectivity. The density of axons directly connecting visual cortical areas (panel g; cortico-cortical connections, row-normalized) are compared to the overlap of input from and output to the same source-target pairs in LP (panel e; putative transthalamic connections). Abbreviations: superior colliculus (SC), lateral amygdala (LA), postrhinal area (POR), laterointermediate area (LI), lateromedial area (LM), primary visual cortex (V1), anterolateral area (AL) rostrolateral area (RL), anteromedial area (AM), posteromedial area (PM), anterior cingulate (ACA), orbital cortex (ORB).

LP contains at least two maps of visual space corresponding to posterior (SC-recipient) and anterior (V1-recipient) subregions

If the LP subregions we defined anatomically are functionally distinct, they may, like visual cortical areas, have separate maps of visual space. Indeed, SC and V1 contain full representations of visual space but project to largely non-overlapping parts of LP (Fig. 1a,b). We used the retinotopic organization of SC and V1 and the topography of their axons in LP to predict the representation of visual elevation and azimuth in LP (Fig 2a-f, S3). The predicted elevation map showed separate spatial gradients of visual elevation representation in LP that converge at the boundary of SC and V1 input to LP. Thus, anatomical connectivity predicts that LP contains at least two distinct representations of visual elevation.

To test this prediction, we used Neuropixels high density electrode probes[16] and a sparse noise stimulus to measure the spatial receptive field location of neurons over the full extent of LP in awake mice (Fig. 2g). The location of each probe in LP was marked with DiI, and every recorded cell was assigned a position in the CCF (Fig. 2h,i; 1579 neurons, Methods). The dispersion of receptive field locations of nearby neurons was substantially higher in LP than the dorsal lateral geniculate nucleus (dLGN; Fig. 2j). Nonetheless, averaging the receptive field locations of nearby LP neurons revealed a smooth map of elevation that, like the elevation map predicted by anatomy, reversed its gradient near the center of LP where SC and V1 axons converge (Fig. 2k). We confirmed our findings from population data by serially recording responses to sparse noise in multiple locations across LP in the same mouse (Fig. 2l). As we moved the probe to more anterior locations, the receptive field elevation gradient reversed (Fig. 2m). This data strongly suggests that aLP and pLP are functionally distinct subregions containing separate maps of visual space.

Similar methods were used to construct maps of azimuth representation in LP. Both the topography of axons and the measured receptive fields showed a gradient of azimuth representation in the region of LP receiving input from V1 but no clear azimuth map in the region of LP receiving SC input (Fig. S2).

Mapping receptive field organization in pLP is potentially complicated by the fact that this region contains adjacent zones receiving unilateral or bilateral SC input[11]. In other species, SC axons are topographically organized in the part of LP receiving unilateral input but more diffuse in the part receiving bilateral input[17–19]. We found that SC axons were topographically organized for elevation but not azimuth in both the unilateral and bilateral SC-recipient portions of LP, consistent with data from rats[20]. However, this organization was more precise in unilateral SC-recipient LP (Fig. S3).

We find evidence for three functionally and anatomically distinct LP subregions. What rules govern the representation of each higher visual cortical area (HVA) within these larger subregions? HVAs in the mouse often contain incomplete retinotopic maps biased to a particular region of visual space[21,22]. These biases might be reflected in the organization of HVA

connections in LP. We computed the elevation bias for the portion of LP receiving input from, or projecting to, each HVA by weighting the LP elevation map by the anatomical input and output volumes shown in Figure 1. We found a significant correlation between these elevations and the mean elevation measured for each HVA via intrinsic imaging (Fig. S4). This result indicates that the LP input and output maps associated with a given HVA are in part predicted by that HVA's retinotopic bias.

Figure 2

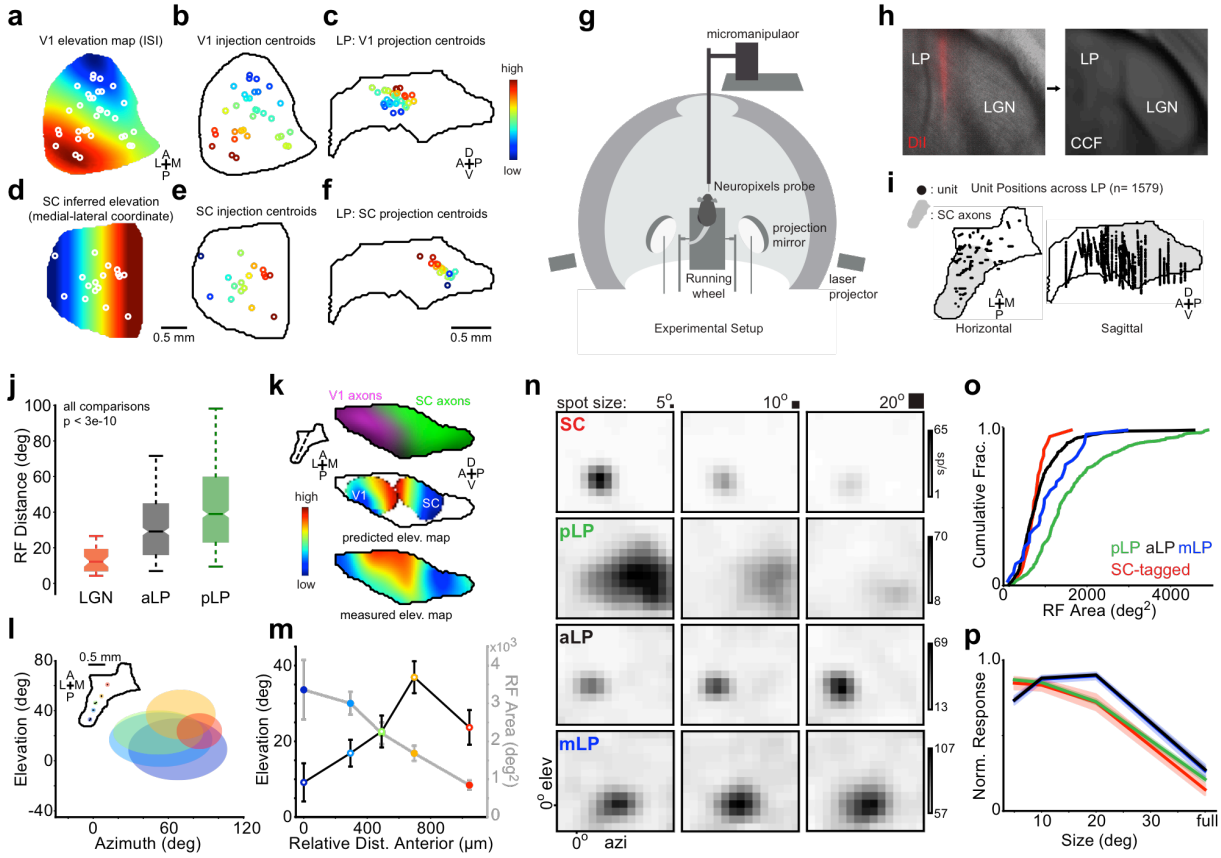

Figure 2. Posterior and anterior LP have separate maps of visual space and distinct receptive field properties. (a) Mean ISI elevation map for V1 with location of V1 injections from the Allen Connectivity Database superimposed (white circles indicate injection centroids). **(b)** V1 injections colored by assigned elevation according to map in (a). **(c)** V1 projection centroids in LP (sagittal plane) for injections in (b) colored by assigned elevation. These centroids were smoothed to create the V1 predicted elevation map in (k). **(d-f)** Same as (a-c) for SC injections. Elevation in SC is inferred from the medial-lateral coordinate (Methods). **(g)** Diagram of experimental setup for visual stimulation and neural recording. **(h)** DiI labeling of probe tract recovered from post-hoc histology and registered to the CCF. **(i)** Recording locations for all LP neurons displayed on horizontal and sagittal projections of LP. Gray region denotes SC-recipient LP. **(j)** Receptive field distance for pairs of cells separated by 20 microns or less in dLGN (orange), aLP (black) or pLP (green). Only cells from the same probe insertion were compared. Box edges indicate first and third quartiles. Notch indicates 95% CI for median (band). Whiskers denote 5th and 95th percentiles. **(k)** Top: LP slice showing SC (green) and V1 (magenta) input to LP. Plane of slice indicated by dotted line in inset. Middle: Predicted LP elevation map based on anatomical V1 input (left) or SC input (right). Bottom: Composite elevation map for all LP cells. **(l)** Data from experiment in which five insertions were made serially in one mouse. Recording locations for each insertion are shown in the inset. Ellipses are centered at the mean RF center for each insertion (color-coded to match inset). Ellipse shape reflects

mean RF shape for neurons at each location. **(m)** Mean receptive field area (closed circles) and elevation (open circles) for each recording location in (l). Colors as in (l). Error bars represent standard error. **(n)** Off receptive fields for example SC (optotagged), pLP, aLP and mLP neurons (rows). Receptive fields were mapped with sparse noise consisting of 5, 10, and 20 degree squares (columns). **(o)** Cumulative distribution of receptive field area and **(p)** mean size tuning for SC (red), pLP (green), aLP (black) and mLP (blue) neurons. Shaded regions in (p) denote standard error.

Receptive field size and size tuning differ across LP subregions

To determine whether the functional properties of LP neurons differ in the three anatomically-defined subregions, we compared visual responses in pLP (the subregion that receives input from SC), mLP (the subregion that receives input from ACA), and aLP (no input from either SC or ACA). In addition, we compared visual responses in LP to putative LP-projecting SC neurons recorded in Nstr1-GN209 Cre x Ai32 mice. In these mice, the SC cell type that projects to LP, but not other SC cell types, expresses channelrhodopsin-2 (ChR2) and responds with sustained spiking to long flashes of blue light[23]. Visual responses of optotagged SC neurons differed substantially from those of non-optotagged neurons recorded in the same mice (Fig. S5).

Reponses to sparse noise consisting of black and white squares of varying size revealed sub-region specific differences in the size and shape of spatial receptive fields and size tuning in LP. Receptive fields in pLP were horizontally elongated and significantly larger than aLP receptive fields (Fig. 2n,o; Table S3; median aspect ratio: 1.30 pLP, 1.15 aLP, 1.05 mLP, 1.07 SC). Receptive fields of mLP neurons were intermediate in size. Despite their large spatial receptive fields, pLP neurons, like optotagged SC neurons, responded most strongly on average to the smallest square size presented (5^0; Fig. 2n,p). These neurons thus respond best to small stimuli presented at any location within a relatively large region of space. In contrast, aLP and mLP neurons responded most strongly to the largest square size (20^0). These results, and others presented below, are consistent with functional differences, in addition to anatomical differences, between LP subregions.

Responses to object and background motion differ in LP subregions

LP-projecting cells in SC are responsive to small, slowly-moving objects in the visual field but not full-field background motion[23]. To determine whether object motion is differentially encoded in LP subregions, we recorded responses to a stimulus consisting of a moving random checkerboard background (full field) and a small (10^0), differentially-moving “patch” of random checkerboard pattern. The patch is only visible when its speed and/or direction of motion differs from that of the background. LP-projecting SC neurons respond most strongly to the small checkerboard patch moving over a stationary checkerboard background (background speed 0 ^{0}/s; Fig. 3a,e,f). In some cases, these neurons also respond moderately when the patch and background move in opposite directions and/or with a large difference in speed, but typically respond weakly or not all to background motion alone (patch speed 0^0/s). Like these SC neurons, pLP neurons responded more strongly to patch motion alone and less strongly to background motion alone or combinations of patch and background motion compared to aLP or mLP neurons (Fig. 3b-f; Table S3). This data further implies that pLP is functionally distinct from aLP and mLP, and, along with size tuning data (Fig. 2n,p), is consistent with the possibility that input from SC plays an important role in shaping visual responses of pLP neurons.

Responses to full-field drifting gratings revealed additional differences between neurons in LP subregions. Neurons in aLP and mLP typically preferred faster speeds and higher temporal frequencies than pLP neurons (Fig. S6). Direction-selective neurons in pLP tended to prefer motion towards the upper-temporal visual field similar to the bias found in SC[24–26] (Fig. S6g), while neurons in aLP and mLP tended to prefer motion towards the nasal visual field (Fig. S6h-j).

Posterior LP neurons respond more strongly to looming stimuli

The portion of LP that receives input from SC and projects to the amygdala has been implicated in mediating a freezing response to looming stimuli[27,28]. To characterize responses of LP neurons to looming stimuli, we presented expanding spots that increased in angular size at a rate that simulates an object approaching at constant speed; the rate of expansion is a nonlinear function of time-to-collision and the object's size/speed ratio[29] (Fig. 3g). SC and pLP neurons responded more strongly to looming stimuli than aLP or mLP neurons (Fig. 3i; S7a). In many SC and pLP neurons, the time of the peak response was linearly related to the object size/speed ratio such that the peak response occurred at a constant angular spot size (Fig. 3h). Similar responses to looming stimuli are observed in other animals and are referred to as η cell responses[29–32]. Other types of responses (e.g. ρ and τ cells) were not clearly observed in our experiments (Fig. S7b). We define η cells as those with correlations greater than 0.9 between the time of peak response and the object size/speed ratio. Cells defined as η cells were more common in pLP than other parts of LP (64% of LP-projecting SC, 25% of pLP, and 5% of aLP and mLP neurons; Fig. 3j). Within pLP, other properties of η cells also differed from non-η cells. In response to the checkerboard stimulus, pLP η cells exhibited a stronger preference for object (patch) motion compared to background motion than non-η cells (Fig. 3k). Thus, a population of pLP neurons may convey information about looming stimuli, and object motion in general, to the amygdala and ventral-stream visual cortical areas.

Clustering LP neurons based on visual response properties alone independently segregates posterior and anterior neurons

Our results demonstrate that distinct patterns of anatomical connections segregate LP into subregions that contain neurons with different functional properties. Is the inverse also true – do functional properties predict the spatial organization of neurons in LP? Hierarchical clustering of LP neurons based on visual response properties (size tuning, RF area, and patch-background index) produced two statistically significant clusters (Fig. 3l). Compared to neurons in cluster 1, neurons in cluster 2 had larger receptive fields, stronger surround suppression, and stronger preference for object motion compared to background motion (Fig. 3n-p). These properties are similar to those of neurons located in pLP. Indeed, 52% of pLP neurons were assigned to cluster 2, while only 7% of aLP neurons and 3% of mLP neurons belonged to this cluster (Fig. 3m). Thus, functional properties independently predict spatial segregation of neurons in LP.

Figure 3

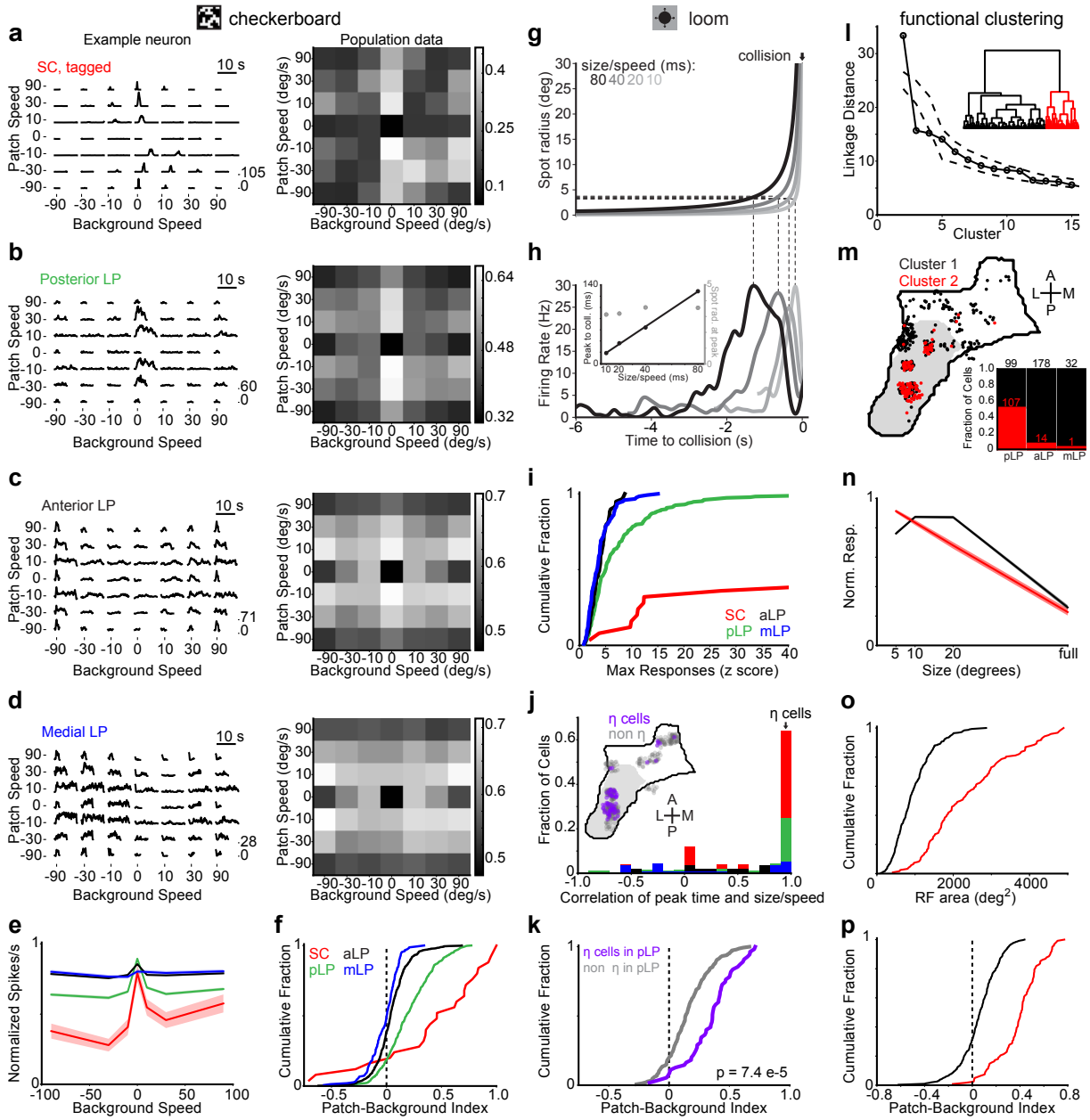

Figure 3. Visual response properties reveal functional-anatomical segregation in LP. (a) Spike density functions of an example optotagged SC neuron (left) and mean population response (right) to random checkerboard background (full field) and patches (10^0) moving relative to each other at various speeds (positive speeds are nasal to temporal). Checkerboard squares are 1^0. Patch speed 0^0/s trials are background motion only. Background 0^0/s trials consist of patches moving over a stationary random checkerboard background. Since the texture of patches and background are indistinguishable, patches moving with the same speed and direction as background are invisible (equivalent to patch speed 0^0/s trials). **(b,c,d)** Same as (a) for pLP, aLP, mLP neurons. **(e)** Population tuning curves for background speed (normalized max response down columns of checkerboard response matrix) for SC (red), pLP (green), aLP (black) and mLP (blue). **(f)** Cumulative distributions of patch-background index values for SC, pLP, aLP and mLP in response to the checkerboard stimulus. Patch-background index is the difference between the maximum responses to patch (background speed 0^0/s) and background (patch

speed 0^0/s) motion divided by their sum. **(g)** Trajectory of spot radius for looming stimuli at four size-to-speed ratios. **(h)** Firing rate of example neuron in posterior LP to looming stimuli depicted in (g). Dotted lines relate spot radius to time of peak firing rate. Inset: time of peak firing rate relative to collision plotted against size to speed ratio for example neuron in (h) (filled black circles; left axis) and spot radius at peak firing rate plotted against size to speed ratio for same neuron (open gray circles; right axis). **(i)** Cumulative distribution of max loom response across all conditions for neurons in SC (red), pLP (green) , aLP (black) and mLP (blue). Note that, due to their low spontaneous firing rates, many SC neurons had z-scores greater than 40. **(j)** Histogram of correlation between peak response time and size to speed ratio for cells in SC and LP subregions (colors as in (i)). Cells with a correlation value greater than 0.9 were classified as η-type. Inset: location η cells shown in a horizontal projection of LP. Gray region denotes SC-recipient LP. **(k)** Cumulative distribution of checkerboard patch-background index values for η (purple) and non-η (gray) neurons in posterior LP. **(l)** Inset: dendrogram representing hierarchical clustering (Ward's linkage criterion) of LP neurons based on visual response properties. Main: linkage distance for the first 15 clusters compared to clusters formed from the same data randomly shuffled across neurons for each visual response parameter (dashed lines; 1-99% confidence interval). **(m)** Horizontal projection of the location in LP of neurons from each cluster. Inset: stacked bar plot showing the fraction of cells in each cluster across LP subregions (numbers give total cell count in each bar). **(n,o,p)** Mean size tuning (n) and cumulative distributions of receptive field area (o) and patch-background index (p) for the two clusters.

Modulation of LP neurons by motor activity

A subset of LP neurons that project to V1 are modulated by motor activity, in particular running and eye movements[13], in addition to visual stimuli. To test whether neurons across the three LP subregions were modulated by running, we compared visual responses to checkerboard and gratings stimuli during trials when mice were stationary or running (Fig. S8a-d). We found a weak but significant facilitation of visual responses during running in all LP subregions, with pLP being the least modulated. Cells that were modulated during the checkerboard stimulus tended to be similarly modulated during drifting gratings (Fig. S8d).

We also tested whether LP neurons were sensitive to eye movements by recording neural activity and eye position in the dark. Aligning the activity of LP neurons to the initiation of horizontal saccades revealed cells that were either excited or suppressed by eye movements (Fig. S8e-n). The fraction of cells that were significantly modulated during saccades was similar across LP subregions (22-24% excited, 6-10% inhibited; n=363 pLP, 472 aLP, and 49 mLP neurons from mice in which we recorded at least 10 saccades in the dark). In SC, 2/11 optotagged cells were excited and none were inhibited during saccades.

V1 and SC silencing differentially modulate activity in LP subregions

The visual response properties of pLP neurons are similar in several characteristics to those of SC neurons that project to pLP. However, pLP also receives direct input from ventral-stream visual cortical areas (Fig. 1a,b), and all visual cortical areas could potentially influence pLP indirectly via projections to SC[33]. To determine the relative impact of cortical and SC input to LP, we recorded spontaneous activity and visual responses of LP neurons while suppressing activity in visual cortex or SC.

To suppress activity in visual cortex, we directed blue light over V1 in VGAT-ChR2 mice (Fig. 4a; this manipulation activates inhibitory neurons which potently suppresses the local cortical network[34]). Cortical suppression strongly reduced both spontaneous activity and responses to the checkerboard stimulus in aLP neurons (Fig. 4b-f). To a lesser extent,

spontaneous activity and visual responses were also reduced in pLP neurons (Fig. 4c,d,g,h). Unlike aLP neurons, the strength of suppression of visual responses in pLP neurons depended on the stimulus condition ($p = 0.019$ for pLP, $p = 0.83$ for aLP, Kruskal-Wallis test). In pLP neurons, responses to the checkerboard patch moving on a stationary background were weakly suppressed compared to responses when the background was moving (Fig. 4h). Consequently, when cortex was inactivated, pLP neurons more strongly preferred object motion compared to background motion, becoming even more like SC neurons that project to pLP.

The effect of cortical suppression on activity of pLP neurons could partially reflect reduced cortical input to SC neurons that project to pLP[35] (Fig. 4a). Thus, we also measured the impact of cortical suppression on SC neurons (note that use of VGAT-ChR2 mice precluded optotagging of LP-projecting SC cells). Cortical suppression moderately reduced visual responses of neurons in the optic fiber layer of SC (SCop), where the somas of LP-projecting neurons are located (Fig. S9). Although the suppression of SCop neurons was similar in magnitude to the effect on pLP neurons, it did not significantly depend on checkerboard background speed ($p = 0.86$, Kruskal-Wallis test). Thus, suppression of SC activity does not fully explain the effects of cortical inactivation on pLP neurons, suggesting that the activity of pLP neurons is most likely shaped by convergent input from both SC and cortex.

To directly assay the contribution of SC to LP visual responses, we inactivated SC with TTX while recording in LP. The relative effects of SC inactivation on aLP and pLP neurons were opposite of those observed during cortical inactivation (Fig. 4k,q,r). Spontaneous activity and visual responses were more strongly suppressed in pLP than aLP (Fig. 4k,l). Moreover, pLP neurons lost their preference for object motion – responses to the checkerboard patch moving over a stationary background were more strongly suppressed than responses when the background was moving (Fig. 4m-s; $p = 3.7e-9$ for pLP, $p = 0.65$ for aLP, Kruskal-Wallis test). Thus, the functional properties of pLP neurons depend critically on receptive field-defining input from SC.

Both functional clustering and responses to looming stimuli (Fig. 3k-n) revealed diversity among pLP neurons, suggesting some pLP neurons may depend more strongly than others on SC input. Consistent with this possibility, the pLP neurons that were most suppressed by SC inactivation showed a greater preference for object motion during control trials than less suppressed pLP neurons (Fig. 4t). Thus, functional diversity in pLP is at least in part explained by strength of input from SC.

Figure 4

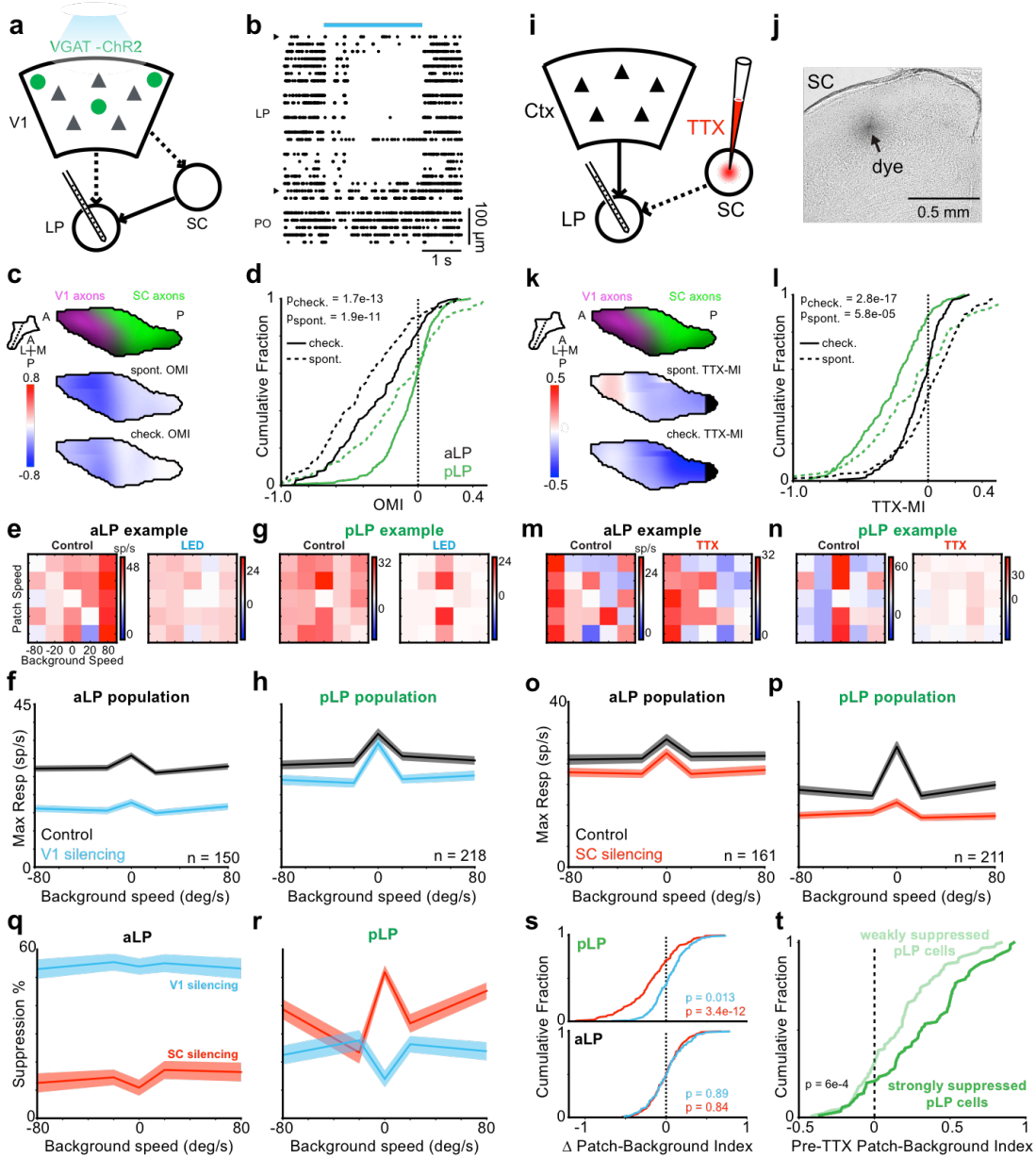

Figure 4. V1 and SC silencing have divergent effects on activity in anterior and posterior LP. (a) V1 silencing was accomplished by transcranial illumination of the cortex with blue light in VGAT-ChR2 mice. Recordings were simultaneously performed in LP. **(b)** Raster plot showing spontaneous activity of all thalamic units for one silencing trial. Blue bar indicates light delivery. Units are ordered by dorsal/ventral position. Carets demarcate LP boundaries. Units ventral of LP are in the posterior thalamic nucleus (PO). **(c)** Top: LP slice showing SC (green) and V1 (magenta) input to LP. Plane of slice indicated by dotted line in inset. Middle: Optogenetic modulation index (OMI) for spontaneous activity averaged across all units in LP. OMI is defined as (optogenetic firing rate - control firing rate)/(optogenetic firing rate + control firing rate). Bottom: OMI for checkerboard response. **(d)** Cumulative distribution of OMI for neurons in pLP (green) and aLP (black) during spontaneous activity (dotted lines) and the checkerboard stimulus (solid lines). **(e)** Example patch-checkerboard matrix for an aLP neuron during control (left) and V1 silencing (right) trials. **(f)** Mean background speed tuning during control (black) and V1 silencing (blue) for aLP population. Values are maximum projections along the columns of the checkerboard response matrix

averaged across cells. Shaded regions denote the SEM. **(g)**, **(h)** same as (e), (f) but for pLP. **(i)** SC silencing was accomplished by injecting TTX into the SC while recording in LP. **(j)** Bright-field image confirming the deposition of dye in sSC after a TTX injection. **(k-p)** As in (c-h) for SC silencing. **(q)** Suppression as a function of checkerboard background speed for aLP population during cortical (blue) and SC (red) silencing. **(r)** As in (q) for pLP. **(s)** Change in the patch-background index during cortical and SC silencing for pLP (top) and aLP (bottom). A negative shift indicates a reduction in patch preference. **(t)** Distribution of patch-background index values (before TTX injection in SC) of pLP neurons that were strongly (TTX-MI<-0.33) or weakly (TTX-MI>-0.33) inhibited by SC inactivation.

Discussion

We combined comprehensive and quantitative anatomical circuit tracing with systematic activity mapping across the entire LP complex in the mouse. By registering data from multiple modalities to a common anatomical coordinate system, we made quantitative comparisons between anatomy, functional properties, and perturbational effects. Together these data provide the most complete picture of the mesoscale circuit organization of higher-order visual thalamus to date. Moreover, we go beyond correlating anatomy and function by silencing the two main LP input pathways and showing divergent effects across LP. Neurons in pLP are driven by SC input and respond to looming stimuli and small, moving objects. Conversely, neurons in aLP are driven by V1 input and respond to large stimuli and full-field motion. A third subregion, mLP, may provide a transthalamic route for information flow from frontal/associational cortex to visual cortical regions (Fig. 5).

Input/output connectivity defines functionally relevant thalamic regions

The close correspondence between anatomically and functionally defined LP subregions supports an emerging view that the fundamental units of corticothalamic computation are not individual thalamic nuclei but more precise thalamic circuits linking functionally related cortical areas[36,37]. A potential criticism is that the three LP subregions we define might arbitrarily lie along a continuum of many input/output microcircuits. However, the reversal of the elevation map, and its close correspondence to the merging of axons from SC and V1, suggests that pLP and aLP are functionally distinct zones. Moreover, mLP is defined largely by a unique projection from frontal cortex, which categorically separates it from the other LP subregions. Thus, we believe our parcellation captures three broad but functionally relevant domains of LP. Future fine-grained investigation could reveal even more precise microcircuits within each subregion.

Network analysis of cortical projections suggests the mouse visual system can be divided into ventral and dorsal streams similar to those described in other species[38]. Our analysis of LP input/output connectivity indicate these cortical pathways are mirrored by cortico-thalamo-cortical projection patterns, with ventral stream areas reciprocally connected to pLP and dorsal stream areas reciprocally connected with aLP. The association of ventral stream areas with SC-recipient LP in the mouse is a reversal to that observed in the primate[39,40], potentially reflecting different functional roles of the ventral and dorsal streams in primates and mice.

The connectivity of mLP with frontal and associational cortices resembles the connectivity of medial and dorsomedial pulvinar in primates[41]. Although most mLP neurons were visually responsive, they were less likely to have organized receptive fields, and many did not respond to sparse noise (Table S2). These data suggest that mLP neurons are involved in higher-order visual processing and do not encode simple visual features. In the future, it will be

interesting to test whether these cells are particularly sensitive to behavioral context, similar to neurons in dorsomedial pulvinar[42].

Our anatomical data suggest that indirect transthalamic pathways connecting cortical areas through LP largely mirror direct corticocortical connectivity (Fig. 1g), as has been suggested in primates[37]. This parallel connectivity scheme could enable the pulvinar/LP to coordinate corticocortical communication by synchronizing activity across cortical areas[43]. How the thalamus modulates or transforms information carried in transthalamic pathways, and how these signals differ in content or impact from those of direct corticocortical pathways, is a crucial question for understanding cortical computation.

Figure 5

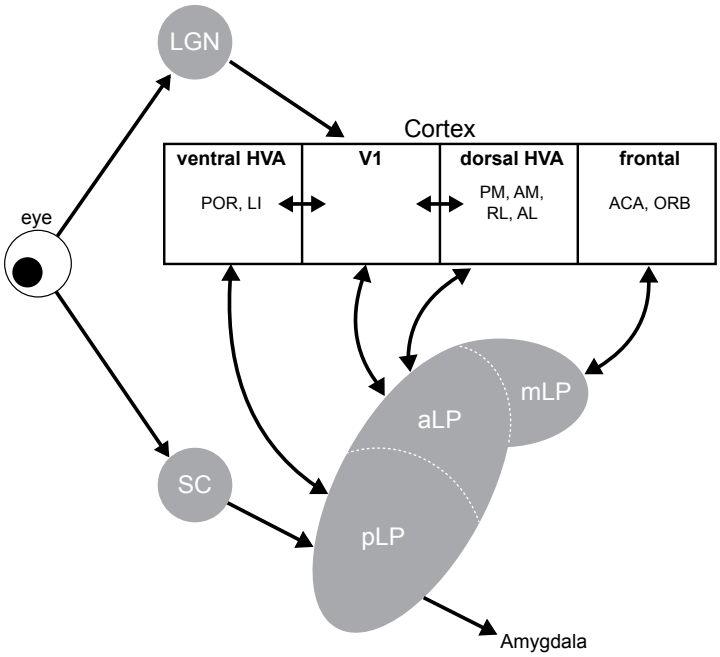

Figure 5. LP subregions channel parallel streams of visual information. Summary diagram depicting the involvement of the three LP subregions in retino-geniculate and retino-collicular visual pathways.

Comparison with previous work

Rodent LP was previously divided into three subregions – caudomedial (LPcm), lateral (LPl), and rostromedial (LPrm) – based on cytoarchitecture and immunocytochemical markers[11,20,44]. These regions are similar to the LP subregions we identified based on anatomical connectivity. However, whereas we include all tecto-recipient LP in pLP, previous schemes separated LP subdivisions that receive bilateral or unilateral SC input (LPcm and the posterior half of LPl, respectively). In the mouse, the portion of LP that receives unilateral SC input is relatively small (Fig. S3), and individual LP neurons extend their dendrites across the border between LPcm and LPl[11]. For these reasons, we did not attempt to assign LP units to one or the other SC-recipient subdivision. Future studies may reveal functional differences between pLP neurons that receive bilateral or unilateral SC input.

The cortical projection to LP originates in both layer 5 (L5) and layer 6 (L6) pyramidal neurons. We found that L5 and L6 neurons from a given cortical area target similar regions in LP (Fig. S1a). It has been proposed that L5 provides a strong "driver" input to LP mediating feedforward transthalamic pathways, while L6 acts as a "modulator" in feedback pathways[1]. Our data suggest that both L5 and L6 participate in reciprocal corticothalamic feedback loops, similar to what has been described for frontal cortex connections with thalamus[45].

SC provides driving input to pLP

Dual retinofugal pathways through LGN and SC are an ancient, highly conserved feature of the vertebrate visual system[46]. In primates, the "secondary" visual pathway running through SC and pulvinar was traditionally thought to play a modulatory role in visual processing, perhaps relaying saccade-modulated visual signals[47,48]. Previous studies disagreed as to whether SC input could drive receptive field properties in pulvinar neurons[49,50]. Our data show that pLP neurons are tuned similarly to their input from SC. Moreover, silencing SC dramatically suppresses visual responses of pLP neurons and abolishes their preference for object motion (Fig. 4). Together these results demonstrate a major role for SC in driving visual responses in pLP and establish that the secondary visual pathway is an important parallel stream running alongside the geniculo-cortical pathway (Fig. 5). Our data further suggest that this pathway is particularly important for carrying information about small moving objects.

Differences between pLP and LP-projecting SC neurons suggest that pLP is not a simple relay of SC input. In particular, receptive fields of pLP neurons are substantially larger and horizontally elongated relative to those of LP-projecting SC neurons, suggesting that multiple SC neurons with horizontally displaced receptive fields might converge onto single pLP neurons. Such convergence is consistent with the topographical organization of SC axons for elevation but not azimuth in LP, and may explain the lack of an organized map of azimuth representation in pLP.

Cortical and subcortical inputs converge in pLP

Both SC and visual cortex project to pLP, suggesting individual pLP neurons could integrate input from cortical and subcortical sources. Since a large majority of pLP cells were suppressed during both SC and V1 inactivation experiments, many pLP neurons likely combine input from these two inputs. Interestingly, SC and cortical inactivation had differential effects on pLP excitability: cortical inactivation more strongly suppressed spontaneous than visually-evoked activity and reduced responses to background motion, while SC inactivation more strongly suppressed visually-evoked activity and abolished tuning for object motion (Fig. 4). Thus, cortical input to pLP may provide tonic modulatory drive as well as information about visual context which can be integrated with object motion signals from SC.

The relative strength of cortical and SC input could underlie the functional diversity of pLP neurons. Indeed, we found visual tuning differences between pLP neurons that were strongly or weakly suppressed by SC inactivation (Fig. 4t). Moreover, neurons in pLP were split between two functionally defined clusters, only one of which closely resembles SC input (Fig. 3l-p). Responses to looming stimuli also revealed functional diversity in pLP (Fig. 3k). The degree to which driving inputs from one or more sources (e.g. SC and/or cortical areas) converge on individual neurons is a critical question for LP and higher-order thalamic nuclei in general[51,52].

Recent studies have shown that higher order thalamus plays a fundamental role in shaping and maintaining cortical activity[2,3,53,54]. Our comprehensive characterization of the

structural and functional organization of LP provides a roadmap to decipher circuit-specific mechanisms by which higher-order thalamus contributes to visual processing and behavior.

Methods

Mice

Rabies injections in visual cortical areas were performed at the Salk Institute and approved by the Salk Institute's Institutional Animal Care and Use Committee (IACUC). All other experiments were performed at the Allen Institute and approved by the Allen Institute's IACUC. Mice of either sex were used and were >70 days old on the day of electrophysiological recordings. Transgenic mice were maintained on a C57BL/6J background. The number of mice used for each experiment is indicated in Supplementary Tables 1 and 2.

Anatomy

To generate LP input maps (Fig. 1), we utilized publicly available anterograde tracing data from the Allen Institute Mouse Brain Connectivity Database[10] (Table S1). For cortical areas, we used injections in C57BL/6J, Emx1 Cre, Rbp4-KL100 Cre, or Ntsr1-GN220 Cre mice. For SC, we used injections in C57BL/6J mice with viral expression in the superficial layers (experiments 112827164, 114754390, 126646502, 128001349, and 146078721). Experiments with injection volumes less than 0.05 mm^3 were excluded. For injections in visual cortical areas, experiments with injection volumes greater than 0.25 mm^3 were also excluded. For each source of input to LP, we averaged "projection energy" volumes aligned to the Allen Institute Common Coordinate Framework (CCF; 25 μm^3 voxels) after normalizing by the brightest voxel in LP. Voxels in LP were then smoothed in three dimensions with a Gaussian kernel with standard deviation of 75 µm. Maximum-intensity projections of smoothed LP voxels in the horizontal plane are displayed in Fig. 1a. For all analyses (e.g. comparing overlap of axons from different sources or clustering) the entire volume of voxels in LP were used.

To generate LP output maps, we injected G-deleted rabies encoding green fluorescent protein or the red fluorophore mCherry into each of the LP targets shown in Fig. 1. For injections in visual cortical areas, injections (35-150 nL, 350 µm deep) were targeted to the center of visual areas identified by intrinsic signal imaging[22]. Anterior cingulate (ACA) injections were 0.5 mm anterior and 0.25 mm lateral from bregma and 0.5 mm deep from the pia (100-150 nL). Lateral amygdala (LA) injections were 1.3-1.8 mm posterior and 3.1-3.3 mm lateral from bregma and 3.4-4.2 mm deep from the pia (50 nL at each of three depths). Mice were perfused seven days after rabies injection. Images of 50-100 µm thick coronal sections were aligned to the CCF in three steps. (1) Images were downsampled to 25 µm pixels and the first and last sections were manually aligned to the CCF template brain. Intermediate sections were then aligned via interpolation. (2) A global affine transformation (scale, rotation, shear) was applied to each section. The transformation matrix was obtained by comparing binarized versions of the image and corresponding template section using the Open Source Computer Vision Library function findTransformECC. (3) Local warping was applied using manually defined key points on the image and corresponding template section. Delauney triangles were defined from the key points, and affine transformations were applied to warp each triangle in the image to the location of the corresponding triangle in the template section. CCF-aligned rabies-injected brains were averaged and smoothed as described for anterograde tracing data to generate the images and analysis shown in Fig. 1.

To complement LP output volumes defined by retrograde rabies labelling, we analyzed anterograde tracing data from LP injections in the Allen Connectivity Database (Fig. S1). Injections were excluded if the injection volume was larger than 0.25 mm^3 or smaller than 0.01 mm^3, resulting in 13 injections. For each injection, the projection to each visual area was normalized by the total projection density to all visual areas. Output volumes were then computed for each target region by weighting the LP injection volumes by their normalized projection density to that region and smoothing with a 100 µm standard deviation Gaussian kernel in three dimensions. Due to potential viral labelling of nearby neurons in the dLGN, output volumes were not calculated for V1 or lateromedial visual cortex (LM), which are known to receive strong projections from dLGN.

To analyze cortico-cortical connectivity (Fig. 1g), we combined injections from the Allen Connectivity Database for each cortical area. Injections were excluded if they were larger than 0.2 mm^3 or smaller than 0.05 mm^3 or resulted in a total density of less than 0.005 mm^3 across all cortical targets. Injections into transgenic lines labelling inhibitory cells or with sparse cortical expression were also excluded. Each remaining injection was normalized by its total projection density across the targets listed in Fig. 1g. The strength of the projection from area X to area Y was defined as the median normalized projection density in area Y across all injections into area X. The resulting connectivity matrix was then row normalized to make Fig. 1g.

To analyze the topography of SC and V1 input to LP (Fig. 2, S4), we primarily used anterograde tracing data from the Allen Connectivity Database. For V1, we used injections in C56BL/6J or Emx1 Cre mice with injection volumes between 0.05 and 0.25 mm^3. To supplement the SC injection dataset (Table S1), we injected AAV-2.1-CAG-Flex-GFP and AAV-2.1-CAG-Flex-tdTomato into the SC of Ntsr1-GN209 Cre mice[55]. In these mice, Cre expression in superficial SC is restricted to cells that project to LP[24,56]. AAV was injected iontophoretically from a pipette with tip diameter of 20 µm using 3 nA current applied 7 seconds on, 7 seconds off for 3 minutes. Both viruses were injected into each mouse, offset along the medial-lateral or anterior-posterior axis (Fig. S3). One of the injections failed in 4 of 8 mice, resulting in 12 injections suitable for analysis. Images from these mice were aligned to the CCF as described for LP output mapping.

For each SC (n=17) or V1 (n=33) injection, we calculated injection and projection centroids in CCF coordinates using injection or projection density data (for injections from the Allen Connectivity Database) or raw fluorescence normalized to the brightest pixel in SC or LP (for our SC injections). Injection centroids were defined as the average CCF coordinate of voxels in SC or V1 with density or normalized intensity greater than 0.25, weighted by their density/intensity. Projection centroids were similarly calculated using voxels in LP. V1 injection centroids were assigned elevation and azimuth values based on ISI maps of elevation and azimuth aligned to the CCF and averaged across mice (Fig. 2, S2). SC injection centroids were assigned elevation and azimuth values based on their position along the medial-lateral or anterior-posterior axis, respectively, which approximates the measured receptive field maps of SC neurons[57] and the topography of V1 projection centroids in SC (Fig. S2d,k). Projection centroids were assigned the same elevation/azimuth as their corresponding injection centroids. To generate smoothed elevation and azimuth maps, projection centroid elevation or azimuth values were smoothed in three dimensions with a Gaussian kernel with standard deviation of 100 µm (SC and V1 data were smoothed separately and then combined in a single volume).

To compare LP input and output volumes to the retinotopic bias of each cortical visual area (Fig. S4), we utilized a mean elevation map for visual cortex based on ISI imaging from 14 mice (dataset from Garrett et al. 2014). For each visual area, we assigned a “measured”

elevation value corresponding to the mean of the ISI elevation map over that area. We then computed two "predicted" elevations for each area by weighting the LP elevation map by either its rabies-based output volume or anterograde input volume (from Fig. 1).

Electrophysiological recordings

Electrophysiological recordings from SC, LP, and dLGN neurons were made with Phase 2 Neuropixels probes[16] (128 channels arranged in two columns, with 20 µm between each recording site). Data were acquired at 30 kHz using the Open Epys acquisition board and GUI[58] and high-pass filtered (300 Hz). Mice were habituated to the recording rig for at least two weeks. On the rig, mice were head-fixed and allowed to run on a styrofoam cyclinder covered with rubber matting. On the day of recording, mice were anesthetized with isoflurane and a small craniotomy was made above the target brain region in the left hemisphere and covered with Qwikcast (World Precision Instruments). Mice recovered for at least two hours before recordings. For most mice, this procedure was repeated the following day for a new recording location (Table S2).

Optotagging SC neurons

The SC cell type that projects to LP was identified ("optotagged") by channelrhodopsin-2 (ChR2) activation in Ntsr1-GN209 Cre x Ai32 mice[23,59]. During these recordings, a 50 µm core diameter optical fiber was inserted in the brain near the Neuropixels probe to a depth just above the SC (~1 mm). Blue light was used to activate ChR2-expressing cells (2 s pulses, 0.5-10 mW measured from the fiber tip). Cells were considered optotagged if their mean firing rate during the last 1 s of the light pulses was greater than 5 standard deviations above the mean spontaneous rate. Spontaneous rate was calculated from the 1 s bin preceding each light pulse.

Visual Stimuli

The mouse's head was fixed at the center of a 24-inch diameter spherical dome (Fig. 2g). Visual stimuli were projected on the inner dome surface from two laser projectors (one on each side of the mouse) pointed at spherical mirrors placed below the running wheel. Four different visual stimuli were presented. (1) Sparse noise consisted of dark (0.6 cd/m^2) and light (5.8 cd/m^2) squares (5, 10, or 20^0 across) presented one at a time for 100 ms on a gray background (3.2 cd/m^2). The stimulus center position for each trial was chosen pseudo-randomly from a grid of 10^0 spacing and ranging from -20 to 120^0 in azimuth (negative is left of straight in front of the mouse) and -30 to 90^0 in elevation (negative is below the eye). All of the stimulus positions, sizes, and contrasts were sampled once per loop in random order before initiating a new loop. (2) Two sets of moving gratings stimuli were presented. For both, the gratings filled the entire right side of the dome, drifted for 2 s, and were followed by a 1 s gray screen period before the next trial. The first set of gratings included two orientations (vertical gratings moving in the nasal-to-temporal direction and horizontal gratings moving downward), six spatial frequencies (0.01, 0.02, 0.04, 0.08, 0.16, and 0.32 cycles/^{0}) and five temporal frequencies (0.5, 1, 2, 4, and 8 cycles/s), for a total of 60 trial types. Each trial type was presented once in random order before beginning a new loop. The second set of gratings included eight directions of motion, two spatial frequencies (0.02 and 0.16 cycles/^{0}), and two temporal frequencies (1 and 4 cycles/s). (3) A checkerboard stimulus consisted of random patterns of dark and light 1^0 squares that filled both sides of the dome. The background checkerboard pattern moved such that the right and left halves converged (temporal-to-nasal motion) or diverged (nasal-to-temporal motion) at the point directly in front of the mouse. Simultaneously, a 10^0 patch, also consisting of a random pattern of dark and light 1^0 squares, moved horizontally with direction and speed independent of that of the background

checkerboard pattern. Patch and background velocities were -90, -30, -10, 0, 10, 30, and 90 ^{0}/s (positive velocities are the nasal-to-temporal direction). The patch is invisible when the patch and background velocities are the same. The patch was presented at 0 and 40^0 elevation for LP recordings or targeted to the multi-unit receptive field elevation for SC recordings. (4) Looming stimuli were light or dark spots that expanded to simulate an object approaching at constant speed[29]. The expansion rate is a nonlinear function of time-to-collision and the object's size/speed ratio, which was 10, 20, 40, or 80 ms. The initial spot radius was 0.5^0 and the final spot radius was 80^0, which was held for 0.5 s before a 2 s gray screen inter-trial interval.

Cortical inactivation

Cortical inactivation experiments were conducted in VGAT-ChR2 transgenic mice[34]. Before each experiment, two optical fibers (200 µm core diameter) were positioned over the skull over visual cortex by stereotaxic coordinates (relative to lambda, the two fibers were 0 and 1 mm anterior and 2.8 and 2.6 mm lateral). Each fiber was coupled to a blue laser or LED calibrated to produce 2.5 mW measured at the fiber tip. During the sparse noise stimulus, control and cortical silencing conditions were interleaved in 25 trial (2.5 s) blocks. For all other stimulus protocols, control and cortical silencing trials were interleaved. Light delivery began one second before each silencing trial and ended 100 ms after the trial. Power was linearly ramped (100 ms) on and off. To allow cortex to recover after cortical silencing, four seconds were added between silencing and control trials/blocks. For all perturbation experiments, checkerboard stimulus conditions were reduced (patch and background velocities: -80, -20, 0, 20, 80 ^{0}/s) to increase trial repetitions. In separate mice not used for SC or LP recordings, we measured the lateral spread of cortical silencing using one fiber at 2.5 mW. Consistent with other studies[60], we found significant silencing at 1 mm and near complete silencing at 0.5 mm lateral from the fiber tip.

SC inactivation

For SC inactivation experiments, we positioned a glass pipette (15-20 µm tip diameter) filled with 25 µM tetrodotoxin (TTX) into SC (0.5-0.7 lateral and 0-0.2 anterior from lambda and 1.2-1.5 deep from brain surface). During pipette insertion, black-white alternating flashes were played in the right hemifield and the visually-evoked potential (VEP) was monitored on an oscilloscope. The VEP became noticeably larger and more consistent at ~1 mm depth, consistent with entry into the SC. The control dataset was collected after pipette insertion but before TTX injection. TTX was then injected by a picospritzer (2-10 PSI). During injection, the TTX pipette meniscus was video monitored to verify an injection volume of ~50 nL. We allowed 5 minutes after injection for TTX to diffuse before collecting the TTX dataset. The checkerboard stimulus was ongoing throughout the control, injection, diffusion, and TTX epochs. At the end of most experiments we moved the Neuropixels probe from LP to SC to verify the SC silencing. No spontaneous or visual-driven spikes were observed at locations 500 µm from the injection pipette tip. Activity appeared normal 1 mm from the injection pipette tip. Evan's blue (0.005%) was included in the pipette solution, and injection location was verified by post-hoc histology.

Eye Tracking

Images of the right eye were acquired at 60 Hz in the dark with an infrared (IR) camera. IR LEDs were placed around the camera lens. The pupil is large in the dark and its edges are partially occluded. Rather than tracking the pupil center, we determined the horizontal position of the lateral edge of the pupil relative to a corneal reflection. Saccade times were detected automatically using a velocity threshold and manually verified.

Data Analysis

Spike sorting was done in a two-step process. First, spikes were automatically detected and clustered using Kilosort[61]. Kilosort output clusters were then manually curated in phy[62]. All units passing this manual step were included for further analysis.

To register units to the CCF, brains were fixed overnight after the last experimental day and sliced on a vibratome (100 µm coronal sections). The first (posterior) and last (anterior) slices containing LP were aligned to corresponding CCF sections, and the remaining slices were aligned by linear interpolation. The probe track was then registered to the CCF by manually annotating the probe tip and the point at which it entered LP (based on DiI labeling). Individual units were then assigned positions along this track according to the distance from the tip to the channel with the maximum waveform amplitude. A similar method was used for SC recordings. Units were considered to be in LP if they fell within 100 µm of the LP border after CCF alignment (with the exception of dLGN units, which were readily identified by the sharp boundary between dLGN and LP in histological slices).

For analysis of responses to sparse noise, checkerboard and loom stimuli, a mean spike density function (SDF) was computed for each stimulus condition by convolving the raw spike train with a Gaussian kernel (10 ms standard deviation for sparse noise or 100 ms for checkerboard and loom) and averaging across trials. The response to a given stimulus condition was taken as the peak of the mean SDF over the stimulus window (50-150 ms after stimulus onset for sparse noise, 250 ms to end of trial for checkerboard, or trial onset to collision time for loom). To quantify a cell's responsiveness to a given stimulus, its peak response was compared to spontaneous activity. A cell was included in the analysis for a given protocol if its peak response across all conditions was greater than 5 standard deviations above the mean spontaneous firing rate. For sparse noise, the spontaneous firing rate distribution was estimated by randomly selecting n trials (where n was the number of repeats of the full stimulus set) and calculating the peak of the mean SDF from stimulus onset to 50 ms (a window that excludes the visual response). This process was repeated 200 times and the resulting 200 mean SDF peaks were taken as the spontaneous firing rate distribution. This same process was used for loom (with 100 repetitions), but the subsampled trials were restricted to those with a size to speed ratio of 80 ms (the longest trials), and the analyzed window ranged from stimulus onset to 1 second after onset. For checkerboard, the spontaneous rate was computed from trials in which the patch and background speed were 0^0/s (a static random checkerboard stimulus; the middle square of the checkerboard response matrix).

For gratings stimuli, the response to a given trial condition was defined as the mean number of spikes elicited during that condition in the window from 250 ms after stimulus onset to stimulus offset. The spontaneous rate was taken from randomly interspersed trials for which the gratings stimulus was omitted (isoluminant gray screen).

Responses to sparse noise were fit to a two-dimensional Gaussian to define receptive field location and area. Cells that did not respond to sparse noise (as defined above) or for which the fitting algorithm failed were not used for receptive field analysis. Receptive area is pi times the product of the major and minor radii of the fit at one standard deviation. To generate the measured LP elevation map, CCF voxels were assigned an elevation based on the mean elevation for cells assigned to that voxel (for the vast majority of voxels this was one or zero units). The resulting map was then smoothed with a Gaussian kernel (standard deviation 100 µm), linearly interpolating values for voxels without data. The same procedure was used for azimuth.

We used two measures to quantify the topographic organization of projection centroids or receptive field locations in LP. The first was the r^2 value of a three dimensional linear fit of the elevation or azimuth assigned to the CCF coordinate of each projection centroid or receptive field. The second was Moran's I, a measure of spatial autocorrelation, for these same values[63]. Randomly dispersed elevations or azimuths result in Moran's I near zero, perfect dispersion (i.e. a "checkerboard" of high and low values) results in a Moran's I near -1, and segregated azimuths or elevations (i.e. high values in one portion of LP and low values in another) results in a positive Moran's I. Standard error of the linear fit r^2 and Moran's I were calculated by randomly sampling (10000 repetitions with replacement) the data and taking the standard deviation of the 10000 resulting values.

To quantify the modulation of visual responses by motor activity in LP, we classified trials as running (mean speed > 5 cm/s) or stationary (mean speed < 1 cm/s). Because animals often spent the majority of the time in one behavioral state (usually running), we matched trial number and stimulus condition by randomly subsampling trials from the behavioral state with more trials. Cells were only included in the running analysis (Fig. S8) if they were deemed to have a significant response to the visual stimulus before separating trials by motor activity. The running modulation index was defined as (run + stationary)/(run – stationary), where run and stationary refer to the mean visual response across all stimulus conditions during running and stationary epochs respectively.

To quantify the effects of silencing cortex or SC on LP visual responses, we defined optogenetic and TTX modulation indices (OMI and TTX-MI) as (perturbation–control)/(perturbation+control), where "perturbation" and "control" were the peak visual response for the checkerboard stimulus averaged over the perturbation and control trials respectively. For spontaneous activity, the central square of the checkboard response matrix was used (static background stimulus). Suppression tuning curves were calculated for the best response at each background speed (maxima along the columns of the checkerboard response matrix). Only cells with significant responses to the checkerboard stimulus during control trials were analyzed.

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Author contributions

CB and SDG conceived of the project, performed experiments, analyzed data and wrote the manuscript. MEG and MLN performed rabies tracing experiments under EMC's supervision. SRO supervised CB and SDG and helped write the manuscript. GJM helped conceive the project. All authors contributed to editing the manuscript.

Competing interests

The authors declare no competing interests.

References

1. Sherman, S. M. Thalamus plays a central role in ongoing cortical functioning. *Nat. Neurosci.* **16,** 533–41 (2016).

2. Purushothaman, G., Marion, R., Li, K. & Casagrande, V. A. Gating and control of primary visual cortex by pulvinar. *Nat. Neurosci.* **15,** 905–912 (2012).

3. Zhou, H., Schafer, R. J. & Desimone, R. Pulvinar-Cortex Interactions in Vision and Attention. *Neuron* **89,** 209–220 (2016).

4. Petersen, S. E., Robinson, D. L. & Morris, J. D. Contributions of the pulvinar to visual spatial attention. *Neuropsychologia* **25,** 97–105 (1987).

5. Snow, J. C., Allen, H. A., Rafal, R. D. & Humphreys, G. W. Impaired attentional selection following lesions to human pulvinar: evidence for homology between human and monkey. *Proc. Natl. Acad. Sci. U. S. A.* **106,** 4054–9 (2009).

6. Van Le, Q. *et al.* Pulvinar neurons reveal neurobiological evidence of past selection for rapid detection of snakes. *Proc. Natl. Acad. Sci.* **110,** 19000–19005 (2013).

7. Ward, R., Danziger, S., Owen, V. & Rafal, R. Deficits in spatial coding and feature binding following damage to spatiotopic maps in the human pulvinar. *Nat. Neurosci.* **5,** 99–100 (2002).

8. Saalmann, Y. B. & Kastner, S. Cognitive and Perceptual Functions of the Visual Thalamus. *Neuron* **71,** 209–223 (2011).

9. Baldwin, M. K. L., Balaram, P. & Kaas, J. H. The evolution and functions of nuclei of the visual pulvinar in primates. *J. Comp. Neurol.* **525,** 3207–3226 (2017).

10. Oh, S. W. *et al.* A mesoscale connectome of the mouse brain. *Nature* **508,** 207–214 (2014).

11. ZHOU, N., MAIRE, P. S., MASTERSON, S. P. & BICKFORD, M. E. The mouse pulvinar nucleus: Organization of the tectorecipient zones. *Vis. Neurosci.* **34,** E011 (2017).

12. Tohmi, M., Meguro, R., Tsukano, H., Hishida, R. & Shibuki, K. The extrageniculate visual pathway generates distinct response properties in the higher visual areas of mice. *Curr. Biol.* **24,** 587–597 (2014).

13. Roth, M. M. *et al.* Thalamic nuclei convey diverse contextual information to layer 1 of visual cortex. *Nat. Neurosci.* **19,** 299–307 (2016).

14. Allen, A. E., Procyk, C. A., Howarth, M., Walmsley, L. & Brown, T. M. Visual input to the mouse lateral posterior and posterior thalamic nuclei: photoreceptive origins and retinotopic order. *J. Physiol.* **594,** 1911–1929 (2016).

15. Durand, S. *et al.* A Comparison of Visual Response Properties in the Lateral Geniculate Nucleus and Primary Visual Cortex of Awake and Anesthetized Mice. *J. Neurosci.* **36,** 12144–12156 (2016).

16. Jun, J. J. *et al.* Fully integrated silicon probes for high-density recording of neural activity. *Nature* **551,** 232–236 (2017).

17. Baldwin, M. K. L., Wong, P., Reed, J. L. & Kaas, J. H. Superior colliculus connections with visual thalamus in gray squirrels (Sciurus carolinensis): evidence for four subdivisions within the pulvinar complex. *J. Comp. Neurol.* **519,** 1071–94 (2011).

18. Baldwin, M. K. L., Balaram, P. & Kaas, J. H. Projections of the superior colliculus to the pulvinar in prosimian galagos (Otolemur garnettii) and VGLUT2 staining of the visual pulvinar. *J. Comp. Neurol.* **521,** 1664–82 (2013).

19. Chomsung, R. D., Petry, H. M. & Bickford, M. E. Ultrastructural examination of diffuse and specific tectopulvinar projections in the tree shrew. *J. Comp. Neurol.* **510,** 24–46 (2008).

20. Takahashi, T. The organization of the lateral thalamus of the hooded rat. *J. Comp. Neurol.* **231,** 281–309 (1985).

21. Zhuang, J. *et al.* An extended retinotopic map of mouse cortex. *Elife* **6,** (2017).

22. Garrett, M. E., Nauhaus, I., Marshel, J. H. & Callaway, E. M. Topography and Areal Organization of Mouse Visual Cortex. *J. Neurosci.* **34,** 12587–12600 (2014).

23. Gale, S. D. & Murphy, G. J. Active Dendritic Properties and Local Inhibitory Input Enable Selectivity for Object Motion in Mouse Superior Colliculus Neurons. *J. Neurosci.* **36,** 9111–9123 (2016).

24. Gale, S. D. & Murphy, G. J. Distinct Representation and Distribution of Visual Information by Specific Cell Types in Mouse Superficial Superior Colliculus. *J. Neurosci.* **34,** 13458–13471 (2014).

25. Drager, U. C. & Hubel, D. H. Responses to visual stimulation and relationship between visual, auditory, and somatosensory inputs in mouse superior colliculus. *J. Neurophysiol.* **38,** 690–713 (1975).

26. Ahmadlou, M. & Heimel, J. A. Preference for concentric orientations in the mouse superior colliculus. *Nat. Commun.* **6,** 6773 (2015).

27. Wei, P. *et al.* Processing of visually evoked innate fear by a non-canonical thalamic pathway. *Nat. Commun.* **6,** 6756 (2015).

28. Shang, C. *et al.* Divergent midbrain circuits orchestrate escape and freezing responses to looming stimuli in mice. *Nat. Commun.* **9,** 1232 (2018).

29. Gabbiani, F., Krapp, H. G. & Laurent, G. Computation of object approach by a wide-field, motion-sensitive neuron. *J. Neurosci.* **19,** 1122–41 (1999).

30. Sun, H. & Frost, B. J. Computation of different optical variables of looming objects in pigeon nucleus rotundus neurons. *Nat. Neurosci.* **1,** 296–303 (1998).

31. Liu, Y.-J., Wang, Q. & Li, B. Neuronal Responses to Looming Objects in the Superior Colliculus of the Cat. *Brain. Behav. Evol.* **77,** 193–205 (2011).

32. de Vries, S. E. J. & Clandinin, T. R. Loom-Sensitive Neurons Link Computation to Action in the Drosophila Visual System. *Curr. Biol.* **22,** 353–362 (2012).

33. Wang, Q. & Burkhalter, A. Stream-Related Preferences of Inputs to the Superior

Colliculus from Areas of Dorsal and Ventral Streams of Mouse Visual Cortex. *J. Neurosci.* **33,** 1696–1705 (2013).

34. Zhao, S. *et al.* Cell type–specific channelrhodopsin-2 transgenic mice for optogenetic dissection of neural circuitry function. *Nat. Methods* **8,** 745–52 (2011).

35. Zhao, X., Liu, M. & Cang, J. Visual Cortex Modulates the Magnitude but Not the Selectivity of Looming-Evoked Responses in the Superior Colliculus of Awake Mice. *Neuron* **84,** 202–213 (2014).

36. Halassa, M. M. & Kastner, S. Thalamic functions in distributed cognitive control. *Nat. Neurosci.* **20,** 1669–1679 (2017).

37. Shipp, S. The functional logic of cortico-pulvinar connections. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* **358,** 1605–24 (2003).

38. Wang, Q., Sporns, O. & Burkhalter, A. Network Analysis of Corticocortical Connections Reveals Ventral and Dorsal Processing Streams in Mouse Visual Cortex. *J. Neurosci.* **32,** 4386–4399 (2012).

39. Lyon, D. C., Nassi, J. J. & Callaway, E. M. A disynaptic relay from superior colliculus to dorsal stream visual cortex in macaque monkey. *Neuron* **65,** 270–9 (2010).

40. Berman, R. A. & Wurtz, R. H. Functional Identification of a Pulvinar Path from Superior Colliculus to Cortical Area MT. *J. Neurosci.* **30,** 6342–6354 (2010).

41. Kaas, J. H. & Lyon, D. C. Pulvinar contributions to the dorsal and ventral streams of visual processing in primates. *Brain Res. Rev.* **55,** 285–296 (2007).

42. Petersen, S. E., Robinson, D. L. & Keys, W. Pulvinar nuclei of the behaving rhesus monkey: visual responses and their modulation. *J. Neurophysiol.* **54,** 867–886 (1985).

43. Saalmann, Y. B., Pinsk, M. A., Wang, L., Li, X. & Kastner, S. The Pulvinar Regulates Information Transmission Between Cortical Areas Based on Attention Demands. *Science (80-.).* **337,** 753–756 (2012).

44. Nakamura, H., Hioki, H., Furuta, T. & Kaneko, T. Different cortical projections from three subdivisions of the rat lateral posterior thalamic nucleus: A single-neuron tracing study with viral vectors. *Eur. J. Neurosci.* **41,** 1294–1310 (2015).

45. Collins, D. P., Anastasiades, P. G., Marlin, J. J. & Carter, A. G. Reciprocal Circuits Linking the Prefrontal Cortex with Dorsal and Ventral Thalamic Nuclei. *Neuron* **98,** 366–379.e4 (2018).

46. Butler, A. B. Evolution of the thalamus: a morphological and functional review. *Thalamus Relat. Syst.* **4,** 35–58 (2008).

47. Berman, R. A. & Wurtz, R. H. Signals Conveyed in the Pulvinar Pathway from Superior Colliculus to Cortical Area MT. *J. Neurosci.* **31,** 373–384 (2011).

48. Berman, R. A., Cavanaugh, J., McAlonan, K. & Wurtz, R. H. A circuit for saccadic suppression in the primate brain. *J. Neurophysiol.* **117,** 1720–1735 (2017).

49. Bender, D. B. Visual activation of neurons in the primate pulvinar depends on cortex but

not colliculus. *Brain Res.* **279,** 258–261 (1983).

50. Casanova, C. & Molotchnikoff, S. Influence of the superior colliculus on visual responses of cells in the rabbit's lateral posterior nucleus. *Exp. brain Res.* **80,** 387–96 (1990).

51. Groh, A. *et al.* Convergence of Cortical and Sensory Driver Inputs on Single Thalamocortical Cells. *Cereb. Cortex* **24,** 3167–3179 (2014).

52. Mease, R. A., Sumser, A., Sakmann, B. & Groh, A. Cortical Dependence of Whisker Responses in Posterior Medial Thalamus In Vivo. *Cereb. Cortex* **26,** 3534–3543 (2016).

53. Guo, Z. V. *et al.* Maintenance of persistent activity in a frontal thalamocortical loop. *Nature* **545,** 181–186 (2017).

54. Schmitt, L. I. *et al.* Thalamic amplification of cortical connectivity sustains attentional control. *Nature* **545,** 219–223 (2017).

55. Gerfen, C. R., Paletzki, R. & Heintz, N. GENSAT BAC Cre-Recombinase Driver Lines to Study the Functional Organization of Cerebral Cortical and Basal Ganglia Circuits. *Neuron* **80,** 1368–1383 (2013).

56. Gale, S. D. & Murphy, G. J. Distinct cell types in the superficial superior colliculus project to the dorsal lateral geniculate and lateral posterior thalamic nuclei. *J. Neurophysiol.* jn.00248.2018 (2018). doi:10.1152/jn.00248.2018

57. Dräger, U. C. & Hubel, D. H. Topography of visual and somatosensory projections to mouse superior colliculus. *J. Neurophysiol.* **39,** 91–101 (1976).

58. Siegle, J. H. *et al.* Open Ephys: an open-source, plugin-based platform for multichannel electrophysiology. *J. Neural Eng.* **14,** 045003 (2017).

59. Madisen, L. *et al.* A toolbox of Cre-dependent optogenetic transgenic mice for light-induced activation and silencing. *Nat. Neurosci.* **15,** 793–802 (2012).

60. Guo, Z. V. *et al.* Flow of Cortical Activity Underlying a Tactile Decision in Mice. *Neuron* **81,** 179–194 (2014).

61. Pachitariu, M., Steinmetz, N. A., Kadir, S. N., Carandini, M. & Harris, K. D. Fast and accurate spike sorting of high-channel count probes with KiloSort. *Adv. Neural Inf. Process. Syst.* **29,** 4448–4456 (2016).

62. Rossant, C. *et al.* Spike sorting for large, dense electrode arrays. *Nat. Neurosci.* **19,** 634–641 (2016).

63. Moran, P. A. P. Notes on Continuous Stochastic Phenomena. *Biometrika* **37,** 17 (1950).

Supplementary Figures and Tables

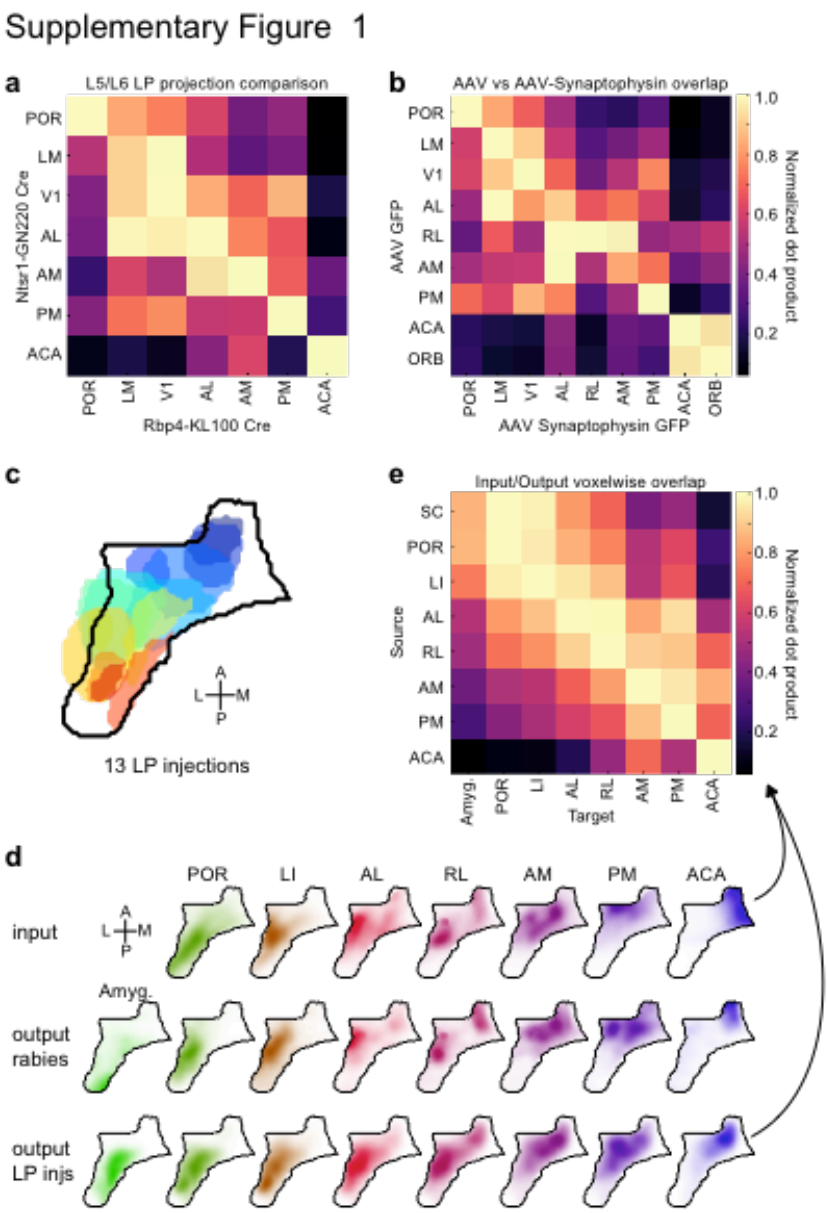

Figure S1. LP output volumes based on anterograde tracer injections in LP resemble LP output volumes based on rabies injection in cortical areas. (a) LP projection overlap matrix for injections in Rbp4-KL100 or Ntsr1-GN220 Cre mice (labeling layer 5 or 6 cortical neurons, respectively). The Allen Connectivity Database does not include injections in these Cre mice for all of the cortical areas shown in Fig. 1 (Table S1). **(b)** LP projection overlap matrix for injections of virus encoding synapse-localized GFP (AAV-Synaptophysin-GFP) or cytosolic GFP (AAV-GFP). **(c)** Injection volumes for 13 LP injections in the Allen Connectivity Database. **(d)** Horizontal projections of input volumes (top), rabies-based output volumes (middle, as in Fig. 1), and output volumes based on LP injections in (c). To make each injection-based output volume, the injection volumes in (c) were weighted by their projection density in the target structure of interest (Methods). V1 and LM were excluded due to possible contamination from dLGN

labelling. **(e)** Input/output overlap matrix (as in Fig 1b) comparing input volumes to LP injection based output volumes. Matrices in (a), (b), and (e) were row normalized.

Supplementary Figure 2

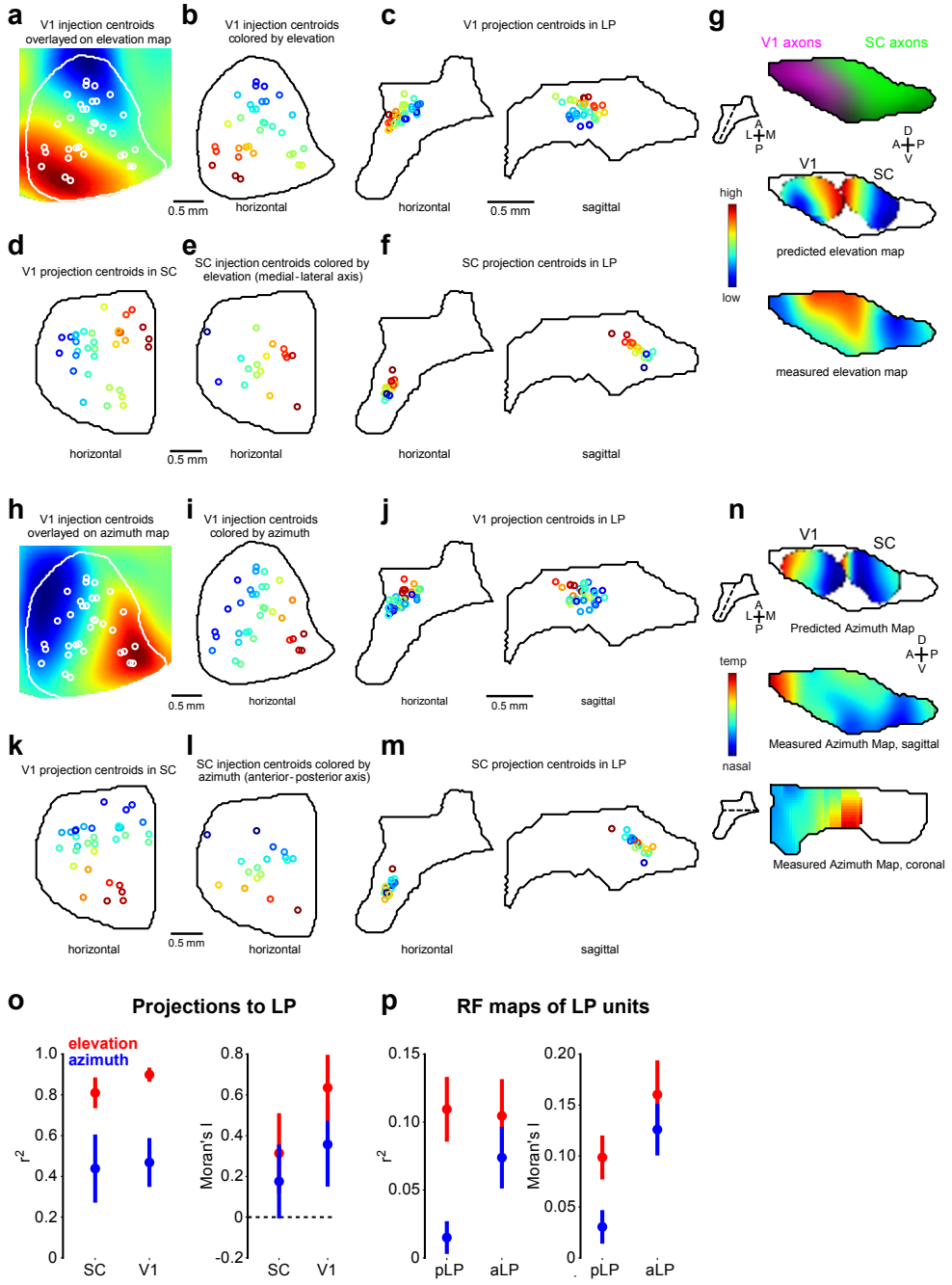

Figure S2. Prediction of LP elevation and azimuth maps by topography of V1 and SC input. (a) Mean elevation map of V1 (white outline) with location of V1 injections from the Allen Connectivity Database superimposed (white circles indicate injection centroids). **(b)** V1 injections colored by assigned elevation according to map in (a). **(c)** V1 projection centroids in LP for injections in (b) colored by

assigned elevation. These centroids were smoothed to create the V1 predicted elevation map in (g). **(d)** V1 projection centroids in SC to demonstrate that the SC elevation gradient is organized along the medial-lateral axis. **(e)** Location of SC injections colored by elevation inferred from medial-lateral coordinate. **(f)** SC projection centroids in LP. **(g)** Top: LP slice showing SC (green) and V1 (magenta) input to LP. Plane of slice indicated by dotted line in inset. Middle: Predicted LP elevation map based on anatomical V1 input (left, from (b, c)) or SC input (right, from (e, f)). Bottom: Composite elevation map for all LP cells. **(h-n)** as in (a-g) but for azimuth. **(o)** Left: r^2 values for linear fit of LP elevation (red) and azimuth (blue) projection centroids. Right: Moran's I (see Methods) calculated for LP projection centroids. **(p)** As in (o) for measured receptive field elevations and azimuths in pLP and aLP. Error bars represent standard error estimated from random sampling.

Supplementary Figure 3

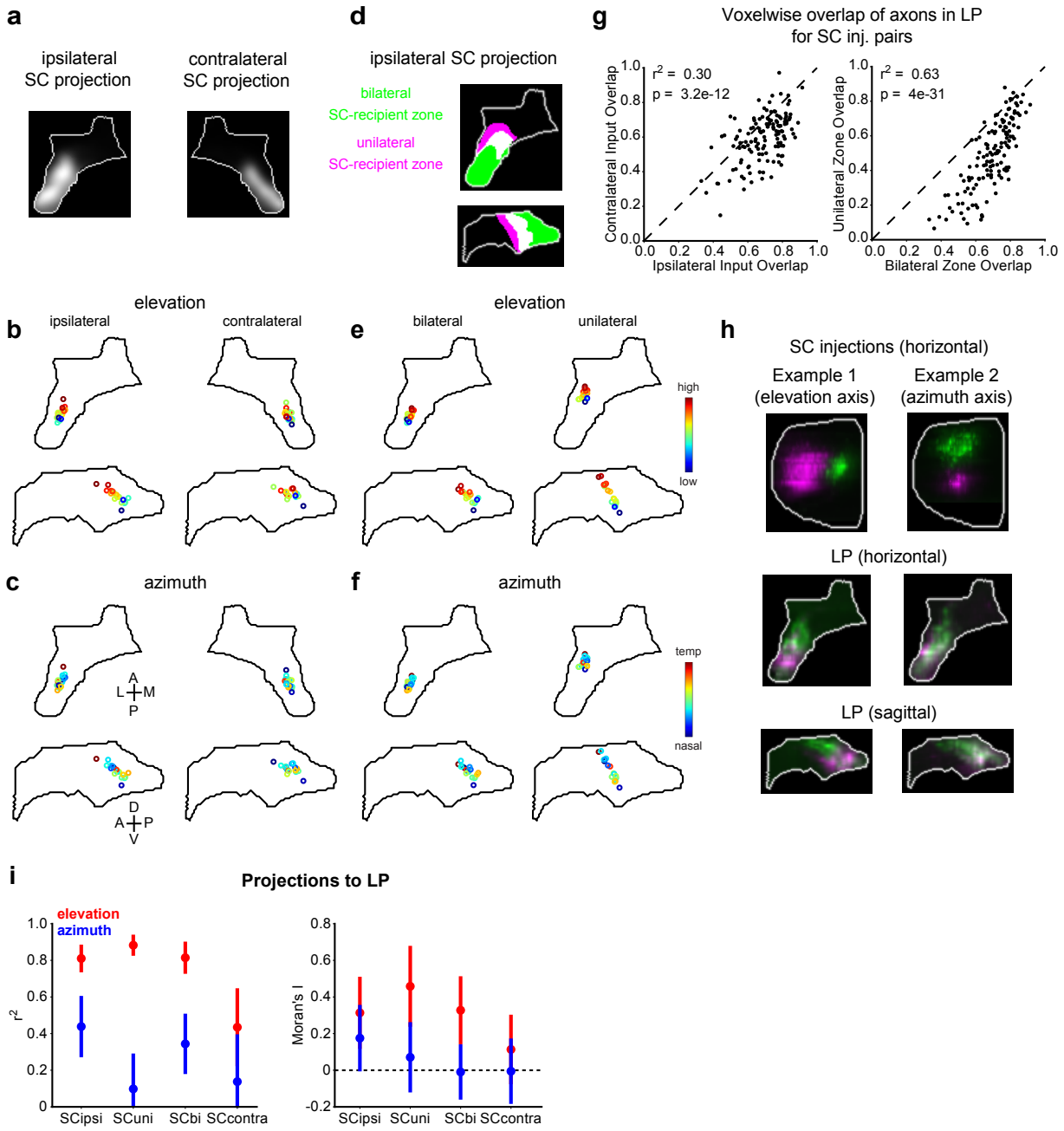

Figure S3. Topography of SC axons in unilateral and bilateral SC-recipient portions of LP. (a) Horizontal projections of average fluorescent intensity of ipsilateral and contralateral SC input to LP (as in Fig. 1a). **(b)** Ipsilateral and contralateral LP projection centroids for all SC injections colored by the relative elevation represented at the location of the injection centroid. **(c)** Same as (b) for azimuth. **(d)** Binarized versions of the data shown in (a) were used to define the portions of LP receiving bilateral (green) or unilateral (magenta) SC input. These subdivisions are mutually exclusive in the 3D volume but overlap (white pixels) in parts of the horizontal and sagittal projections of LP. **(e)** Projection centroids, colored by assigned elevation as in (b), of ipsilateral SC axons in the bilateral or unilateral SC-recipient portions of LP. **(f)** Same as (e) for azimuth. **(g)** Left: voxelwise overlap (normalized dot product) of SC axons in the bilateral SC-recipient portion of ipsilateral or contralateral LP for all pairs of SC injections.

Right: overlap of ipsilateral SC axons in the bilateral or unilateral SC-recipient portion of LP. **(h)** Examples of SC injections separated on the axis along which receptive field elevation (example 1; medial-lateral) or azimuth (example 2; anterior-posterior) varies. For each example, two injections were made in the same brain using virus coding for different fluorophores. The top images show the injection sites and the images below show horizontal and sagittal projections of axons in LP. **(i)** Left: r^2 values for linear fit of SC elevation (red) and azimuth (blue) projection centroids in LP. Right: Moran's I for SC projection centroids in LP. Data is shown for all of ipsilateral LP (SCipsi), the unilateral (SCuni) and bilateral (SCbi) SC-recipient portions of ipsilateral LP, and contralateral LP (SCcontra).

Supplementary Figure 4

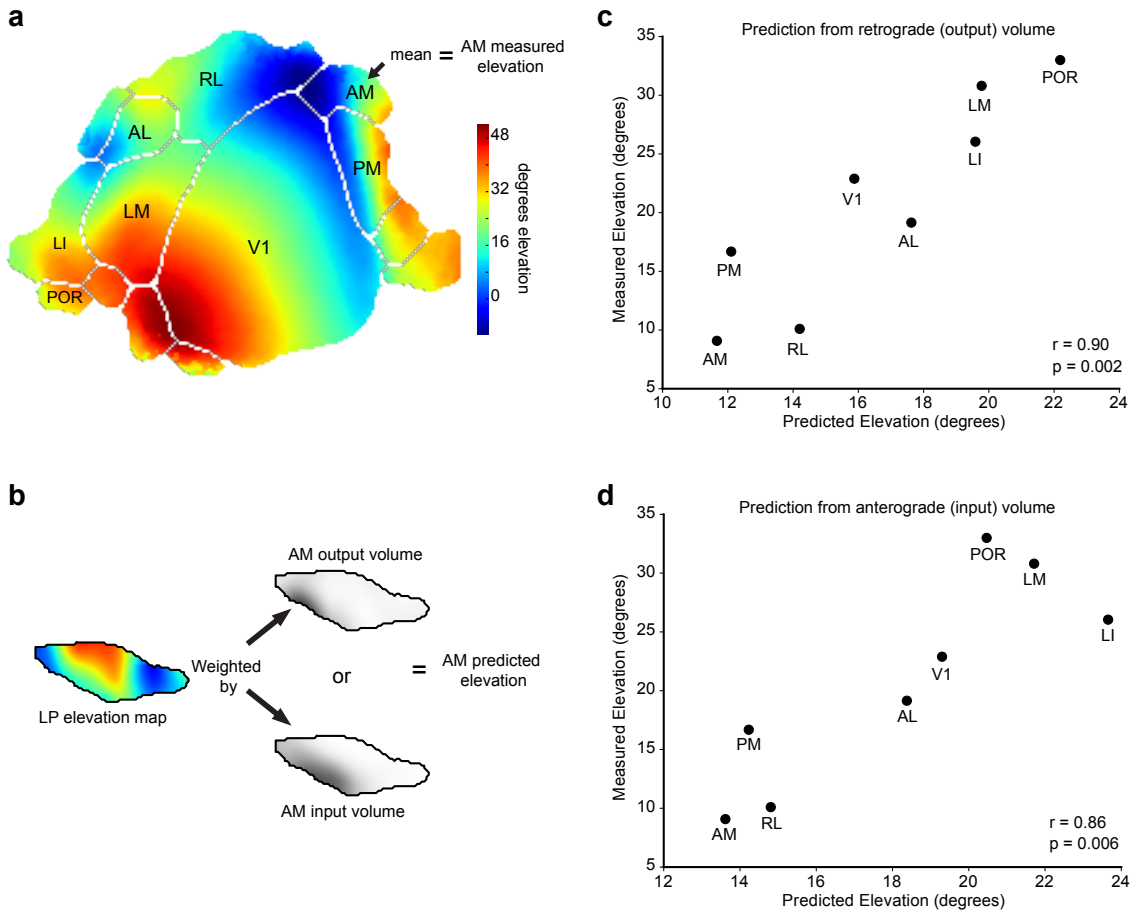

Figure S4. LP input/output projection volumes for eight visual cortical areas reflect biases in retinotopic coverage. (a) Average elevation map determined by ISI imaging across 14 animals[22]. The mean elevation across each area was assigned as the "measured" elevation in (c) and (d). **(b)** For each area (AM used here as example), a "predicted" elevation was assigned by weighting the LP elevation map by its retrograde output projection volume (used in (c)) or anterograde input projection volume (used in (d)). **(c,d)** Scatterplots comparing measured to predicted elevations based on retrograde volumes (c) or anterograde volumes (d).

Supplementary Figure 5

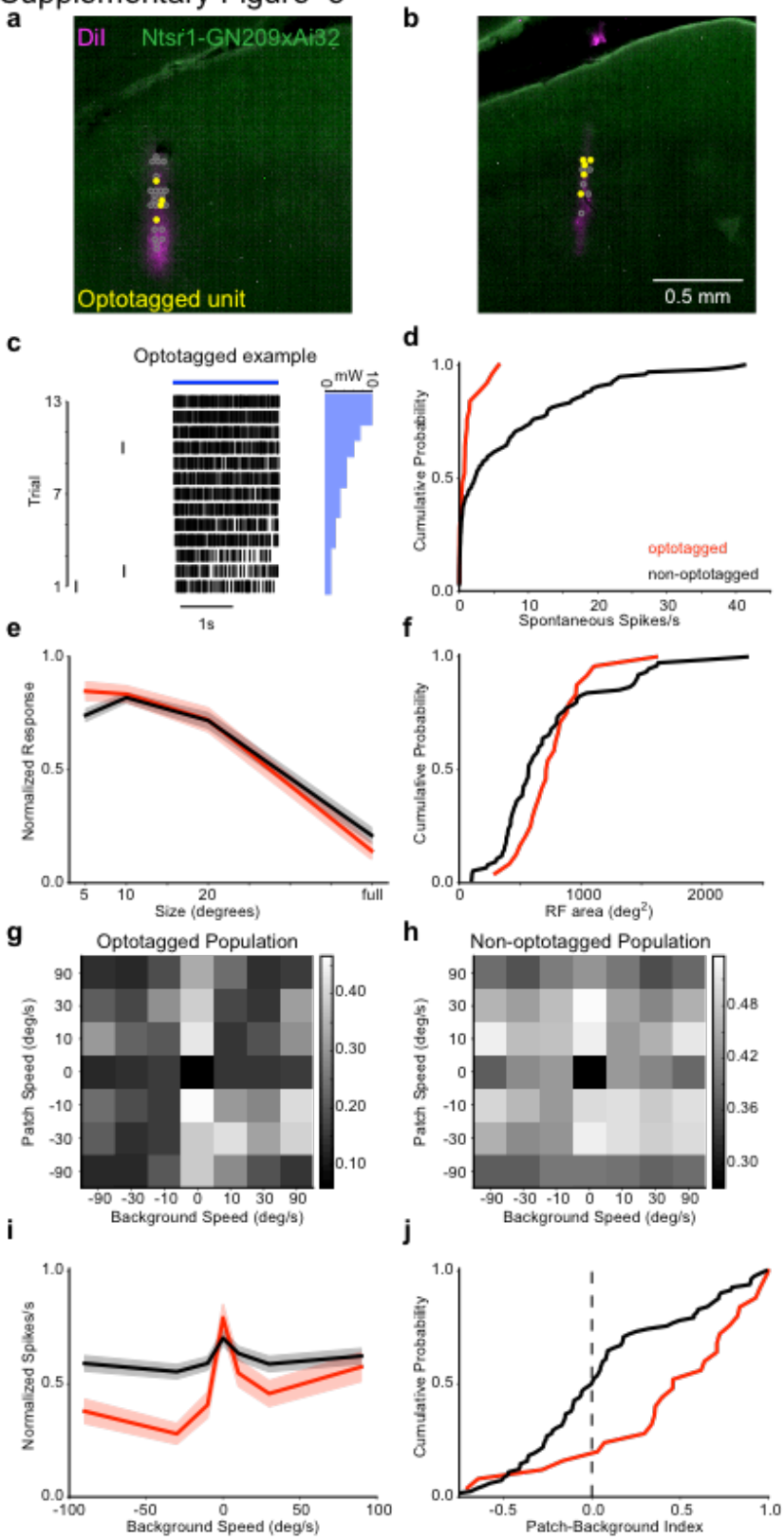

Figure S5. Optotagging identifies LP-projecting cells in superficial SC. (a, b) Coronal SC sections from two optotagging experiments showing distribution of optotagged (filled yellow circles) and non-optotagged (open gray circles) units in SC. **(c)** Raster showing response of example optotagged cell over 13 laser stimulation trials. Bar plot on right indicates laser power for each trial. **(d, e, f)** Cumulative distributions of spontaneous activity, population size tuning curves, and cumulative distributions of receptive field area for optotagged (red) and non-optotagged cells (black). **(g)** Population normalized checkerboard response matrix for optotagged SC cells. **(h)** Same as (g) for non-optotagged SC cells. **(i,j)** Population checkerboard background speed tuning curves (column maxima) and cumulative distributions for patch-background index for optotagged (red) and non-optotagged cells (black).

Supplementary Figure 6

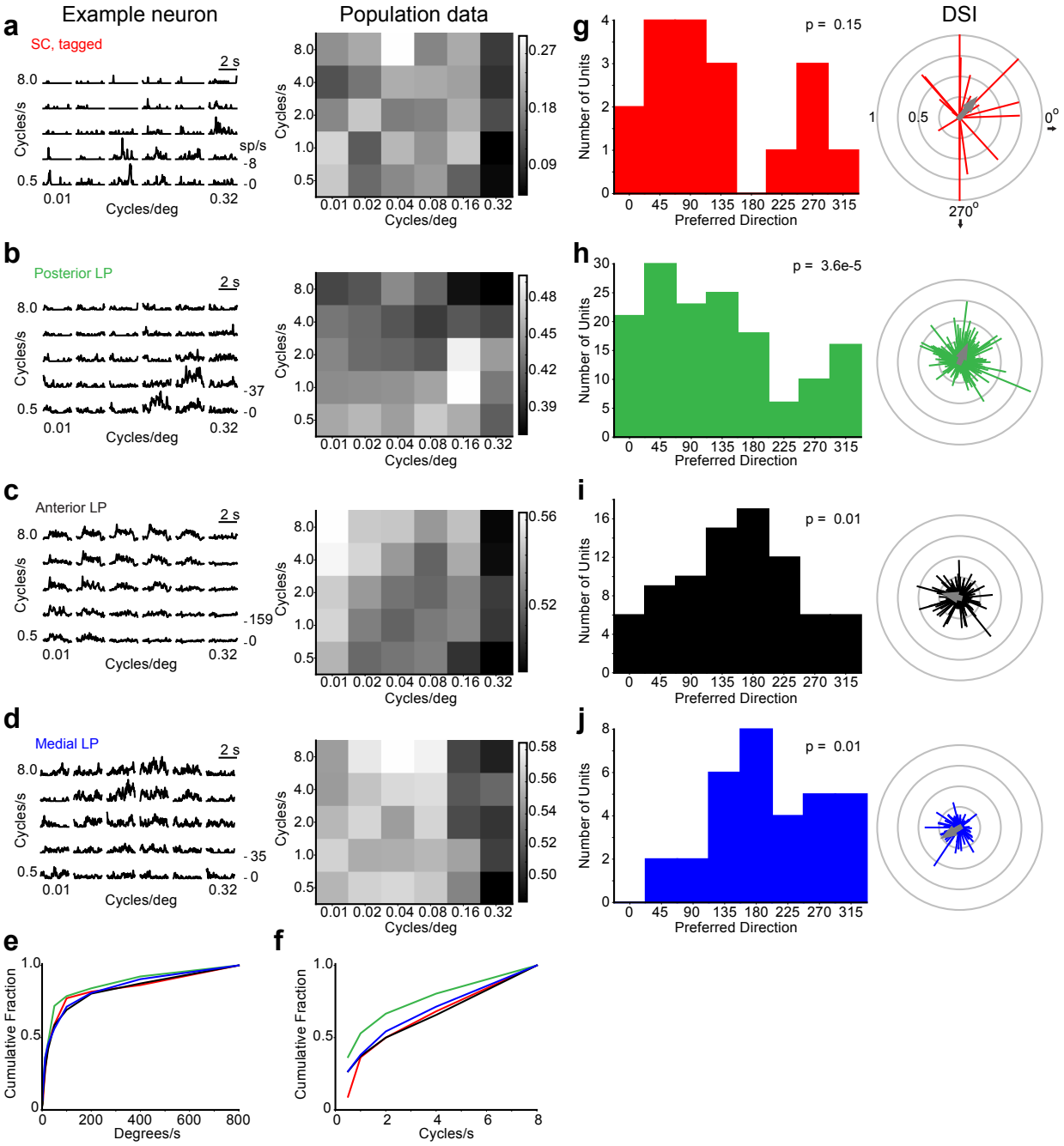

Figure S6. Comparison of spatio-temporal frequency and direction tuning in SC and LP subregions. (a) Spike density functions of an example optotagged SC neuron (left) and mean population responses (right) to drifting gratings of varying spatial and temporal frequency. **(b,c,d)** Same as (a) for pLP, aLP and mLP neurons. **(e,f)** Cumulative distributions of preferred speed and temporal frequency for SC and LP subregions. **(g)** Left: histogram of preferred directions for optotagged SC cells with direction selectivity (DSI, 1-circular variance) greater than 0.15. Right: polar plot representation of direction preference for SC cells. Each cell contributes one vector. Direction of vector indicates preferred direction and length indicates DSI. Gray arrow represents vector sum of population. **(h-j)** Same as (g) for LP subregions. P values in (g-j) calculated by Rayleigh's test for circular uniformity.

Supplementary Figure 7

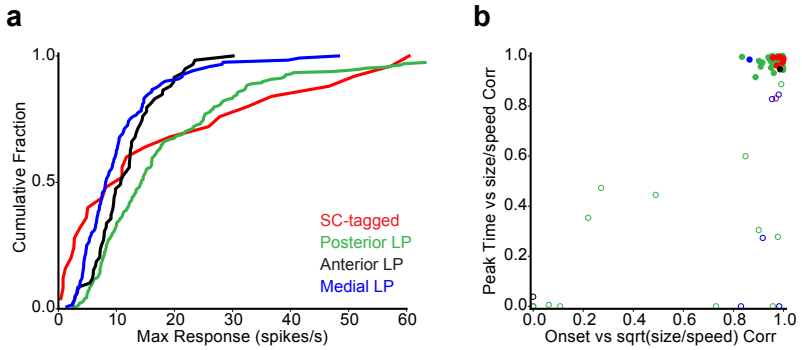

Figure S7. Classification of loom response types. (a) Cumulative distribution of maximum firing rate response to looming stimuli for cells in SC and LP subregions. **(b)** Scatterplot of correlation between peak response time and size to speed ratio (y axis) and correlation between response onset and the square root of the size to speed ratio (x axis). Cells were included only if they had a response greater than 3 standard deviations above baseline) for each size to speed ratio. Filled circles are cells that were classified as η-type. This data suggests that the majority of cells with a significant loom response display η behavior (upper right quadrant), and not ρ (lower right) or τ (lower left)—two other common loom response patterns[30].

Supplementary Figure 8

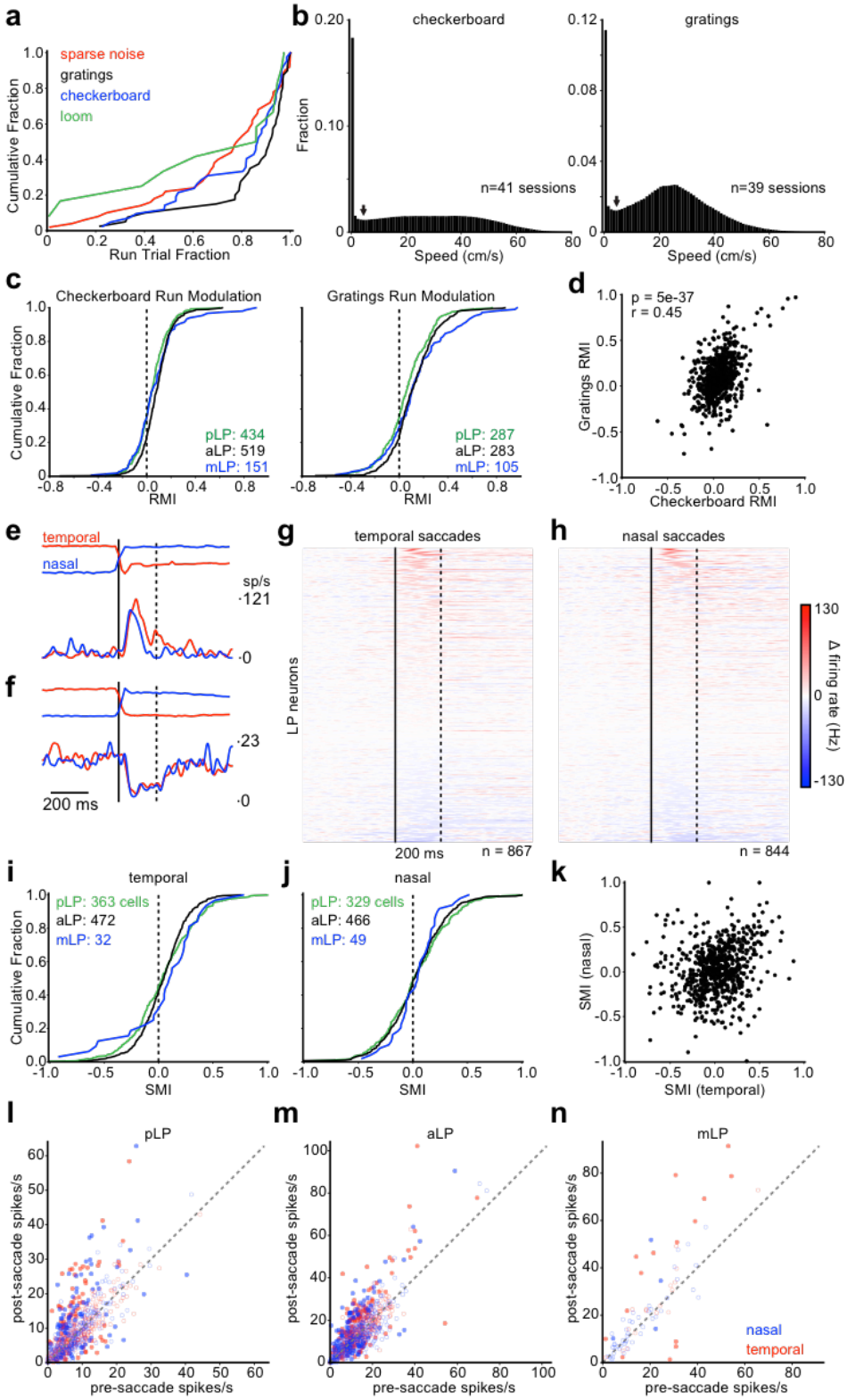

Figure S8. Motor activity weakly modulates cells across LP. (a) Cumulative distribution of run trial fraction across all recording sessions for four visual stimulus protocols. **(b)** Histograms of running speed across all sessions and mice for checkerboard and gratings protocols. The first bin defines "stationary" epochs (< 1 cm/s). Arrows indicate run threshold (5 cm/s). **(c)** Cumulative distributions of the run-modulation index (RMI)—defined as (run – stat)/(run + stat) where run and stat are the mean visual response during running and stationary trials respectively—during the checkerboard stimulus and drifting gratings. **(d)** Scatterplot comparing RMI during gratings and checkerboard for all LP cells with a significant response to both protocols (Methods). **(e)** Example unit excited by eye movements. Top traces are average eye trajectories for nasal (blue) and temporal (red) saccades. Bottom traces are mean SDFs triggered on saccade onset (solid line). The analysis period for (g-n) is bracketed by solid and dotted lines. **(f)** Same as (e) for example unit suppressed by eye movements. **(g)** Heat plot showing change in firing rate aligned to saccade onset (temporal direction) for LP cells. Only cells for which there were at least 10 saccades were included (n=867). Solid and dotted lines bracket analysis window as in (e, f). **(h)** Same as (g) for nasal saccades (n=844). **(i)** Cumulative distribution of saccade modulation index (SMI; difference between firing rates before and after the saccade divided by the sum of these rates) for cells in each LP subregion (colors as in (c)), calculated for eye movements towards the temporal visual field. **(j)** Same as (i), but for nasal saccades. **(k)** Scatterplot comparing SMI for temporal and nasal saccades. Every point represents one cell. **(l, m, n)** Scatterplots of firing rate before and after saccade onset for cells in pLP, aLP and mLP. Every cell contributes two points to a given plot corresponding to nasal (blue) or temporal (red) saccades. Filled circles denote cells with statistically significant modulation (Wilcoxon signed-rank test, $p<0.01$).

Supplementary Figure 9

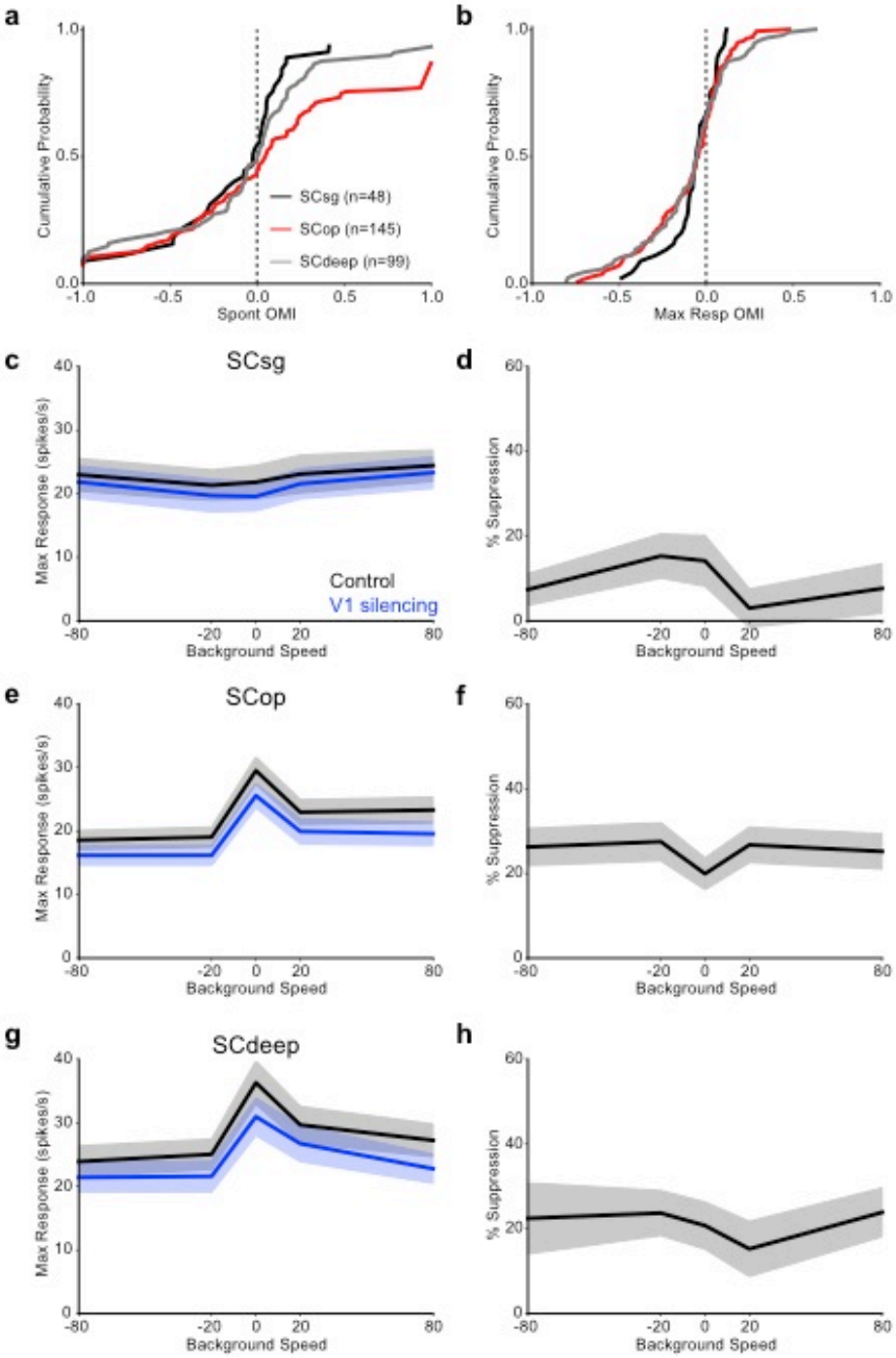

Figure S9. Cortical silencing moderately suppresses visual responses of SC neurons. (a) Cumulative distribution of the optogenetic modulation index (OMI) during spontaneous activity for SC cells in the superficial gray (SCsg), optic fiber layer (SCop), and deeper layers (SCdeep). **(b)** As in (a) but for the maximum response to the checkerboard stimulus. **(c)** Background speed tuning for the checkerboard stimulus (max response for each column of the checkerboard response matrix) during

control and V1 silencing trials for cells in the SCsg. **(d)** Suppression during cortical silencing across checkerboard background speeds. **(e,f,g,h)** Same as (c,d) but for cells in SCop and SCdeep.

Injection Target	SC	LA	POR	LI	LM	V1	AL	RL	AM	PM	ACA	ORB
From Allen Institute Mouse Brain Connectivity Database:												
All	5		8	2	11	51	5	4	8	7	16	13
Ntsr1-GN220 Cre			3		1	8	1		2	1	1	
Rbp4-KL100 Cre			4		2	10	2	2	2	3	5	5
AAV Synaptophysin			1		3	14	1	1	1	4	3	1
Other:												
AAV	12											
Rabies		2	2	3	3	2	5	2	5	2	4	

Supplementary Table 1. **Number of injections used for anatomy experiments**. In most cases, one injection was made in a mouse. For our AAV injections in SC, 8 of the 12 injections were dual injections of two different AAVs in the same hemisphere of 4 mice (see Methods). For Allen Institute Mouse Brain Connectivity Database experiments, “all” refers to experiments in wildtype, Emx1 Cre, Nstr-GN220 Cre, or Rbp4-KL100 Cre mice using AAV-GFP or AAV-Synaptophysin-GFP. These data were combined for LP input mapping shown in Fig. 1. Data from specific subsets of experiments (Nstr1-GN220 Cre, Rbp4-KL100 Cre, AAV-Synaptophysin-GFP) are shown in Fig. S1.

	SC		pLP	aLP	mLP	dLGN
	optotagged	non-tagged				
Sparse Noise	24/25 (96%)	75/97 (77%)	364/633 (58%)	306/751 (41%)	36/195 (18%)	42/64 (66%)
Gratings	22/25 (88%)	72/97 (74%)	353/494 (71%)	312/598 (52%)	105/195 (54%)	24/46 (52%)
Checkerboard	25/25 (100%)	82/97 (85%)	440/499 (88%)	519/640 (81%)	151/195 (77%)	60/64 (94%)
Loom	23/25 (92%)	59/97 (61%)	116/227 (56%)	15/59 (25%)	28/120 (23%)	
V1 Inactivation (checkerboard)		268/292 (92%)	218/266 (82%)	150/236 (64%)		
SC Inactivation (checkerboard)			211/243 (87%)	161/217 (74%)		

Supplementary Table 2. Number of cells recorded during electrophysiology experiments. Values indicate the fraction of cells that responded (see Methods) to each of the visual stimulus types shown in the left column, or to the checkerboard stimulus for V1 and SC inactivation experiments. For the sparse noise stimulus, a fitable spatial receptive field was also required to

include a cell in the numerator. Only a subset of the stimuli were presented during some experiments. SC optotagging experiments were in Ntsr1-GN209 Cre x Ai32 mice. V1 inactivation experiments were in VGAT-ChR2 mice. The number of mice used for control experiments were 5 (SC), 30 (LP) and 3 (dLGN); for cortical inactivation 5 (SC) and 6 (LP); and for SC inactivation 8 (LP). In some cases we recorded twice from the same mouse on consecutive days. The number of recording sessions for control experiments were 8 (SC), 50 (LP), and 3 (dLGN); for cortical inactivation 9 (SC) and 11 (LP); and for SC inactivation 10 (LP).

	SC vs			pLP vs		aLP vs	SC opto vs
Measurement	**pLP**	**aLP**	**mLP**	**aLP**	**mLP**	**mLP**	**SC non-opto**
Spontaneous Rate	**1.0E-14**	**1.0E-15**	**4.7E-15**	**4.6E-20**	**1.1E-06**	9.6E-02	**5.5E-03**
RF Area	**3.1E-08**	8.5E-01	6.8E-02	**7.0E-36**	**6.8E-04**	4.2E-02	8.3E-02
RF Aspect	**1.0E-03**	4.2E-02	7.5E-01	**8.3E-05**	**6.0E-04**	7.6E-02	9.9E-01
Size Tuning, 5 deg	9.4E-01	**4.5E-03**	2.0E-02	**3.3E-15**	**1.4E-05**	9.1E-01	6.6E-02
Size Tuning, 10 deg	6.2E-01	3.3E-01	5.4E-01	3.1E-01	7.9E-01	8.0E-01	8.3E-01
Size Tuning, 20 deg	9.6E-01	**3.0E-04**	**4.8E-03**	**2.9E-17**	**2.9E-05**	8.6E-01	6.4E-01
Size Tuning, 40 deg	2.5E-01	1.0E-02	3.2E-02	**3.4E-03**	1.2E-01	9.5E-01	8.0E-01
Preferred SF	6.6E-01	7.2E-01	9.9E-01	7.9E-01	3.3E-01	4.4E-01	4.3E-01
Preferred TF	2.7E-02	6.7E-01	4.6E-01	**1.4E-05**	1.2E-02	5.1E-01	3.6E-01
Preferred Speed	3.3E-01	9.4E-01	7.3E-01	**6.2E-03**	2.9E-01	4.1E-01	3.3E-01
Patch Background Index	**2.8E-04**	**4.3E-07**	**3.2E-07**	**1.3E-31**	**5.1E-28**	**4.4E-05**	**1.0E-03**
Max Loom Resp	1.1E-01	7.7E-01	6.3E-01	**1.1E-03**	**3.8E-11**	1.7E-02	8.2E-01
Max Loom (z-score)	**1.8E-10**	**1.2E-10**	**2.2E-12**	**5.2E-04**	**5.0E-08**	2.5E-01	**2.9E-07**

p < alpha
p >= alpha

Supplementary Table 3: Statistical comparisons for neurons in different recording regions. Dark squares indicate statistically significant comparisons (threshold for significance was 0.05/6 for first six columns and 0.05 for final column). All tests were non-parametric (Wilcoxon rank-sum).