

1 **Solitary bee larvae prioritize carbohydrate over protein in parentally provided pollen**

2

3 Alexander J. Austin^{1,2†} and James D. J. Gilbert¹

4

5 ¹Department of Biological and Marine Sciences, University of Hull, UK

6 ²Strategy & Environment, Ku-ring-gai Council, NSW, Australia

7 [†]To whom correspondence should be addressed. Email: alexander.j.austin17@gmail.com

8

9 **Acknowledgements**

10 The authors thank Victor Swetez, Emma Chapman, Toby Bagnall, Fiona Tainsh and Shannon
11 Woodmansey for logistical assistance; Stephen Simpson, Audrey Dussutour, Mathieu Lihoreau,
12 Michal Filipiak, Sheena Cotter, Lori Lawson Handley, Domino Joyce, Elizabeth Duncan, Francis
13 Gilbert and Lucy Browning for valuable discussions and comments on the manuscript. The study
14 was funded by the University of Hull; JDJG was also initially supported in this work by a
15 research grant from the Association for the Study of Animal Behaviour.

16

17 **Author Contributions**

18 JDJG and AA conceived and designed the study. AA gathered all data. Both authors analysed the
19 data, and wrote and edited the manuscript.

20

21 **Data Accessibility**

22 All data in this study will be made available upon acceptance via a digital repository such as
23 DRYAD, Zenodo, Open Science Framework etc.

24

25 Abstract

- 26 1. Most organisms must regulate their nutritional intake in an environment full of complex
27 food choices. While this process is well understood for self-sufficient organisms,
28 dependent offspring, such as bee larvae, in practice have limited food choices because
29 food is provided by parents. Nutrient balancing may therefore be achieved by offspring,
30 by parents on offspring's behalf, or by both, whether cooperatively or in conflict.
- 31 2. We used the Geometric Framework to investigate the capacity of dependent larval mason
32 bees (*Osmia bicornis*) to regulate their intake of protein and carbohydrate. Female *Osmia*
33 seal eggs individually inside cells they have provisioned with pollen, and have no contact
34 with developing offspring, allowing offspring choices to be studied in isolation.
35 Herbivorous insect larvae are typically expected to balance protein and carbohydrate to
36 maximise growth and reproduction.
- 37 3. Contrary to prediction, carbohydrate and not protein mediated both growth and survival
38 to pupation. Accordingly, larvae prioritised maintaining a constant intake of carbohydrate
39 and self-selected a relatively carbohydrate biased diet compared to other hymenopterans,
40 while tolerating wide excesses and deficiencies of protein, rendering them potentially
41 vulnerable to dietary change or manipulation. Reasons for prioritising carbohydrate may
42 include (1) the relative abundance of protein in their normal pollen diet, (2) the relative
43 paucity of nectar in parental provisions making carbohydrate a scarce resource, or (3) the
44 requirement for diapause for all *O. bicornis* larvae. Larvae were intolerant of moderate
45 dietary dilution, likely reflecting an evolutionary history of nutrient-dense food.
- 46 4. Our results demonstrate that dependent offspring can remain active participants in
47 balancing their own nutrients even when sedentary, and, moreover, even in mass
48 provisioning systems where parents and offspring have no physical contact. Research
49 should now focus on whether and how evolutionary interests of parent and dependent
50 offspring coincide or conflict with respect to food composition, and the implications for
51 species' resilience to changing environments.

52

53

54 Keywords: pollination, foraging ecology, agriculture, nutritional geometry, limiting nutrient,
55 diapause, ecological trap, environmental change, bee health

56

57 **Introduction**

58 Most animals manage their nutrient intake by combining nutritionally different foods
59 (Simpson & Raubenheimer 2012). However, the importance of this ability depends upon the
60 nutritional variability of the animals' typical food (Despland & Noseworthy 2006;
61 Raubenheimer, Simpson & Mayntz 2009). Extreme specialists, for example, can lose the
62 capacity to regulate nutrition (Warbrick-Smith *et al.* 2009; Poissonnier *et al.* 2018). One way in
63 which organisms can experience limited nutritional choice is if they are dependent upon others
64 for nutrition, or "alloregulation" (Lihoreau *et al.* 2014), such as dependent offspring of altricial
65 birds, human toddlers, and many larval insects. Under these circumstances, by what rules
66 offspring regulate their own consumption should depend upon provisioning rules of parents. On
67 the one hand, offspring tend to have different requirements from parents (Harper & Turner 2000;
68 Michaelsen *et al.* 2003) - particularly for protein, given their elevated rates of somatic growth
69 and development. Accordingly, parents often make different nutritional choices for their
70 offspring versus when foraging for themselves (Royama 1970; Dussutour & Simpson 2009; Burt
71 & Amin 2014). For example, granivorous birds usually provision young with insects, rather than
72 the seed diets of adults, to fulfil protein requirements (Wiens & Johnston 2012). If parents
73 alloregulate offspring nutrition tightly, then offspring should have no need for self regulation,
74 like extreme specialists (Poissonnier *et al.* 2018). On the other hand, parents may provide
75 suboptimal nutrition for offspring - either through inefficiency (e.g. Seidelmann 2006), or if
76 parents' and offspring's evolutionary interests do not coincide (Trivers 1974). Here, offspring
77 may be able to use nutritional regulation to mitigate costs arising from their parents' nutritional
78 choices. While there has been much research into evolutionary compromises involving offspring
79 solicitation and corresponding parental responses (e.g. Smiseth, Wright & Kölliker 2008), less is
80 known about whether or how offspring may exert control by discriminating among parental
81 provisions.

82 The Geometric Framework for Nutrition (GF) allows us to investigate foraging decisions
83 made by animals in multi-dimensional "nutrient space" (Simpson & Raubenheimer 1993). The
84 GF can be used to determine animals' nutritional choices relative to their "intake target" - the
85 optimal amount and balance of multiple macronutrients - as well as their "rule of compromise"
86 that governs their choices when restricted to suboptimal food (Raubenheimer & Simpson 1999b).
87 The GF has provided insights into the nutritional ecology of a broad range of taxa (reviewed in

88 Simpson & Raubenheimer 2012). Its application to dependent offspring, though, has typically
89 been as part of studies of social insect systems (e.g. Helm *et al.* 2017) and studies have often
90 inferred offspring requirements indirectly from patterns of alloparental feeding in studies more
91 broadly focused on adult foraging (see Dussutour & Simpson 2009; Cook *et al.* 2010; Vaudo *et*
92 *al.* 2016). In such systems, multiple adults normally contact offspring, progressively feeding and
93 adjusting nutrition in response to feedback (Field 2005; Schmickl & Karsai 2017), making the
94 responses of individual larvae difficult both to follow and interpret.

95 In solitary bees, by contrast, typically females provision offspring individually with a
96 pollen ball before sealing the cell and leaving. This behaviour makes solitary bees an ideal,
97 manipulable model for directly studying the nutrition of dependent larvae (Strohm *et al.* 2002)
98 independently of provisioning decisions made by parents. Larvae of bees, like most aculeate
99 hymenopterans, rely on parents or alloparents for nutrition (Field 2005). Nutritional requirements
100 for bee adults and offspring differ, often radically (Weeks *et al.* 2004; Filipiak 2019); adults
101 primarily feed on carbohydrate-rich nectar (although see Cane 2016) while larvae feed mostly on
102 protein-rich pollen (Filipiak 2019). Solitary bees, along with most other hymenopterans and
103 many other parental insects, typically have a simple one-to-one parent-offspring relationship
104 whereby parents “mass provision” their young, providing a finite, fixed-mass food provision, and
105 have no contact with their young during development (Costa 2006). Such systems are almost
106 unstudied in a rigorous nutritional context (but see e.g. Roulston & Cane 2002). In these species,
107 there is no opportunity for parents to adjust nutrition according to offspring feedback, and the
108 larva must therefore make the best of what it is given. It may be that offspring regulate their own
109 nutrition to compensate for variation, as in more independent insect larvae (Lee *et al.* 2002), or
110 possibly to mitigate costs imposed by parents. Alternatively, they may have lost this capacity,
111 like extreme specialists (Warbrick-Smith *et al.* 2009; Poissonnier *et al.* 2018). We know very
112 little about how bee larvae deal with variable nutrition (but see Helm *et al.* 2017) - a knowledge
113 gap with potentially important consequences, considering the proposed link between nutritional
114 stress and bee declines (Roulston & Goodell 2011; Goulson *et al.* 2015).

115 In this study, we used a commercially important solitary bee species, *Osmia bicornis*, to
116 investigate how dependent larvae cope with varying nutrition, and whether they can regulate
117 their own intake. *O. bicornis* are pollen generalists (Falk 2015) and the solid, roughly spherical
118 pollen balls that parents provide to offspring are variable in species composition (Haider *et al.*

119 2014). Although *O. bicornis* larvae are sedentary, they are capable of limited movement, in
120 principle allowing them to preferentially consume specific parts of a fixed provision (note that
121 other invertebrates are also capable of extracting and consuming preferred nutrients from
122 nutritionally complex food items (Mayntz *et al.* 2005). The species is entirely solitary with no
123 known tendency for offspring to “help at the nest” as in other bees (Hogendoorn & Velthuis
124 1993; Rehan *et al.* 2014) so there is no reason to believe mothers would alter offspring nutrition
125 to force them to help, as in other systems (Lawson, Helmreich & Rehan 2017) and therefore no
126 obvious potential for parent-offspring conflict over offspring nutrition. Natural variation in
127 pollen ball nutrient content is largely unquantified (although see Budde & Lunau 2007), so there
128 is no prior expectation about the capacity of larvae to regulate their consumption. We used a
129 classic GF design (e.g. Lee *et al.* 2008), focusing on protein and carbohydrate, with two
130 experimental phases. In the first “no-choice” phase we raised *O. bicornis* larvae on fixed diets of
131 differing protein to carbohydrate ratios to determine their rules of compromise and the diet
132 composition that maximised fitness. In a second “choice” phase, we then provided larvae with
133 targeted choices between sets of two imbalanced diets that differed in their protein:carbohydrate
134 ratios to determine whether larvae defend an intake target. Given the central role of protein in
135 growth of insect larvae (Scriber & Slansky 1981; Behmer 2009), and following Hunt & Nalepa’s
136 (1994) exhortation to “follow the protein”, we predicted that (1) protein would be a key driver of
137 fitness in larval *O. bicornis*, (2) larvae would accordingly aim for a relatively protein-biased
138 intake target, and (3) larvae would prioritize regulating intake of protein over carbohydrate.

139

140 **Methods**

141

142 *Study organism*

143 *Osmia bicornis* is a common, cavity-nesting solitary bee (Falk 2015), and a commercially
144 important pollinator (Jauker *et al.* 2012). *O. bicornis* larvae were obtained as diapausing adults in
145 cocoons (Mauerbienen®). These were released at the nesting site at the University of Hull in
146 April 2017, and emerging adults allowed to breed. Early trials revealed that fresh eggs and
147 newly emerged larvae were too fragile for manipulation. Therefore, newly emerged larvae were
148 left alone for two days before we transferred them to a single-occupancy nest and assigned each

149 to an experimental treatment. Details of nesting apparatus and monitoring protocols are available
150 in the supplementary methods.

151

152 *Diet Formulation*

153 Existing artificial diet protocols for solitary bees have met with limited success in terms
154 of larval survival (Nelson, Roberts & Stephen 1972; Fichter, Stephen & Vandenberg 1981). We
155 used six diets, consisting of three different protein:carbohydrate (P:C) ratios (Diet A = 1:1.2,
156 Diet B = 1:2.3 & Diet C = 1:3.4) and two total macronutrient concentrations (concentration 1 =
157 90% nutrients, 10% diluent, or concentration 2 = 70% nutrients, 30% diluent; see table S1). Diet
158 ratios were chosen based on a combination of the nutrient ratios in honeybee-collected pollen
159 loads and published data for protein content of *O. bicornis* pollen balls (Budde & Lunau 2007).
160 Diets were diluted with sporopollenin, the primary constituent of the exine of pollen (Mackenzie
161 *et al.* 2015), an extraordinarily stable natural polymer. Sporopollenin is a novel dietary diluent
162 for bees; its suitability has been demonstrated in a separate study (Tainsh *et al.* 2020). For a more
163 detailed description of sporopollenin and its preparation, see supplementary methods.

164

165 *Experiment 1: No-choice phase*

166 Two-day-old larvae, randomized by parentage, were allocated to one of 6 treatments
167 corresponding to our 6 artificial diets (n = 20/treatment). Provisions were made to resemble the
168 size of natural provisions (mean initial artificial provision weight = 0.323g +/- 0.034g). Once
169 provisioned, larvae were placed in an incubation chamber (Gallenkamp, IH-270) at 23°C and
170 80% RH. Provisions were replaced weekly to avoid desiccation and mould formation, or when
171 fully consumed by larvae, ensuring the diet was always available in excess. Weight of provision
172 consumed was recorded upon provision replacement. A “water control” group, containing
173 artificial diets but no larvae, was used to track water loss from the diets, going through the same
174 weighing regime as above with weight loss recorded at each swap. Nests were checked daily for
175 mortality. Cocoon weight was recorded at the completion of pupation.

176

177 *Experiment 2: Choice phase*

178 In the choice experiment, 32 two-day-old larvae of mixed parentage were randomly
179 divided among four treatments. Treatments consisted of strategic pairwise combinations (see Fig

180 1; Table 1) of four possible diets: A1 (1P:1.2C, 90%), A2 (1P:1.2C, 70%), C1 (1P:3.4C, 90%),
181 C2 (1P:3.4C, 70%). Because *O. bicornis* larvae are sedentary and receive a single provision, it is
182 not biologically appropriate to present choices between two diets simultaneously. Therefore,
183 choices were offered temporally by swapping the provision every other day, presenting one diet
184 at a time. This required the larvae to differentially feed over time to compensate for temporal
185 imbalance, in order to converge on an intake target (see e.g. Raubenheimer & Jones 2006). All
186 larvae were kept on the same treatment from two days post-hatching up to pupation, whereupon
187 diet replenishment ceased. The diet that the larvae would be fed first was randomly assigned via
188 coin toss prior to the experiment.

189

190 *Statistical Analysis*

191 All analyses were conducted in R version 3.4.2 (R Core Team, 2017). For the no-choice
192 experiment, we calculated total nutrients consumed (protein and carbohydrate) from raw diet
193 consumption data for each swap, adjusted for water loss and dilution. Values were then summed
194 for each larva.

195 To investigate consumption rules, including rules of compromise, we first asked whether
196 diet ratio and concentration affected consumption of (a) the total provision, (b) protein, or (c)
197 carbohydrate, using models of each respective variable with “ratio” and “concentration” as
198 predictors. Rules of compromise can include nonlinear effects, particularly curves around the
199 intake target (Simpson & Raubenheimer 1993). To account for potentially curvilinear
200 relationships we also added ratio^2 as a predictor, as well as two-way interactions between all
201 predictors.

202 To assess fitness consequences of macronutrient consumption, we analysed cocoon
203 weight at pupation and survival to pupation. For both analyses, to analyse potentially nonlinear
204 effects of nutrient consumption upon fitness, we used polynomial regression, fitting both first-
205 and second-order polynomial terms for “protein consumed [P]” and “carbohydrate consumed
206 [C]”. We analysed cocoon weight using a linear model with “cocoon weight” as a response. The
207 full model contained linear (P and C) and quadratic effects for both nutrients (P^2 and C^2) and
208 their interaction ($P \times C$), as well as diet concentration (high or low), and two-way interactions
209 between concentration and nutrients ($\text{conc} \times P$, $\text{conc} \times P^2$, $\text{conc} \times C$, $\text{conc} \times C^2$). We used
210 standard diagnostics to check the fit of models, and used a reverse stepwise process to determine

211 the minimal model, at each step dropping the least significant term until the model contained
212 only significant terms. To analyse survival, we used parametric survival analysis in the *survival*
213 package in R and fitted the same full model as described above. We assessed model fit
214 graphically by inspecting the Kaplan-Meier estimates of the residuals against the assumed
215 Weibull distribution. Again we used reverse stepwise selection to determine the minimal model,
216 comparing models with likelihood ratio tests against a chi squared distribution. To visualise these
217 fitness effects, we calculated response surfaces for cocoon weight and survival, and visualised
218 them using non-parametric thin-plate splines.

219 In the choice experiment, the mean final consumption of each nutrient was investigated
220 using linear models with diet combination, dilution and their interaction as predictors, and using
221 Tukey's post hoc tests to compare individual treatments against each other. Under a null
222 expectation we would expect larvae to eat randomly from each diet (Fig 1). Thus, for each larva
223 we calculated the deviation from this null expectation. We then tested whether these values
224 systematically departed from zero for protein and carbohydrate, and whether these departures
225 from random consumption differed by treatment group. We used a linear model with "deviation
226 from random consumption" as the response variable and "treatment group" as a predictor.

227 Larvae that died pre-pupation were not used in the calculation of the mean protein and
228 carbohydrate consumption for diets in either experiment, or for cocoon weight, but were used in
229 analyses involving survival.

230

231

232 **Results**

233 *No-choice phase*

234 Dietary P:C ratio had a significant effect on the overall amount of provision consumed,
235 with larvae consuming more provision on high P:C ratio diets ($F_{1,78}=21.55$, $p<0.0001$). Total
236 consumption was also affected by diet concentration ($F_{1,78}=14.03$, $p<0.001$); larvae on less
237 concentrated diets consumed more provision, indicating compensatory feeding. The quadratic
238 term was not significant (Fig. 2a; Table S2a).

239 Dietary P:C ratio had a strong effect on the total amount of P eaten ($F_{1,79}=146.93$,
240 $p<0.0001$); more protein was eaten by larvae raised on the higher P:C diets (Fig. 3). Diet

241 concentration had no effect on the amount of P consumed; neither was there a ratio:concentration
242 interaction, nor a quadratic effect of ratio (Table S2b).

243 In contrast, larvae consumed similar amounts of C across all diets, with neither
244 concentration nor dietary P:C ratio (linear or quadratic) having an influence on the amount of C
245 consumed (Table S2c). A mean of 0.23 ± 0.01 g of C was consumed by (surviving) larvae across
246 all diet treatments (Fig. 3).

247 Cocoon weight varied differently with macronutrient intake depending on the overall
248 concentration of the diet (carbohydrate \times conc interaction, $F_{1,72}=6.50$, $p=0.01$; protein \times conc
249 interaction, $F_{1,72}=4.82$, $p=0.03$). At 90% nutrient density, cocoon weight was correlated
250 positively with the amount of carbohydrate consumed, and negatively with protein (Fig. 4a). For
251 our range of diets, the greatest weights were obtained by larvae that ate above approx. 0.3g C and
252 below 0.15g P. In contrast, at 70% nutrient density, cocoons were lower in weight than on the
253 90% diets, and were fairly uniform in weight irrespective of macronutrient intake (Fig 4b). No
254 quadratic effects were observed, nor interactions involving quadratic effects, meaning that we
255 did not identify an optimal amount of P or C that maximised cocoon weight within the range of
256 diets we used (Table S3a).

257 The relationship between survival and nutrition similarly depended upon dietary
258 concentration (carbohydrate \times conc interaction: $\chi_1=6.50$, $p=0.01$). Survival of larvae fed diets at
259 90% concentration depended primarily upon carbohydrate consumption (Fig 5a). Those larvae
260 that consumed high amounts of carbohydrate saw the highest survival irrespective of how much
261 protein was consumed. At lower levels of carbohydrate, interestingly, protein weakly mediated
262 survival (protein \times carbohydrate interaction: $\chi_1=-4.88$, $p=0.046$). Survival of larvae raised on the
263 more dilute diets was much lower, and was not substantially affected by intake of P or C (Fig
264 5b). Again, there were no significant quadratic terms, whether as main effects or as part of
265 interactions (Table S3b).

266

267 *Choice phase*

268 We found no evidence of larvae defending a common intake target *sensu stricto*
269 (*Raubenheimer & Simpson 1993, 1999b*), i.e. a common ratio *and* amount of nutrients
270 consumed, which would have been evident as all groups clustering at a common point in nutrient
271 space in Fig 6a. Nevertheless, consumption deviated from random so as to converge upon a

272 target P:C ratio (see e.g. Deans, Sword & Behmer 2019) represented by a common line, or
273 “nutritional rail”, of approx. 1:1.8 (Fig. 6a). The amount of protein consumed by larvae was
274 significantly affected by diet combination: more protein was consumed by individuals offered
275 diet combinations that were overall more concentrated ($F_{3,23}=7.43$, $p<0.01$, Fig 6a; Table S4a).
276 Similarly, carbohydrate consumption was significantly affected by diet combination ($F_{3,23}=4.58$,
277 $p=0.01$, Fig 6a; Table S4b). Unlike with protein, though, this pattern appeared to be driven by the
278 diets at the extreme; only the most concentrated diet pair (C1A1) differed from the least
279 concentrated pair (C2A2; Fig 6a); other pairwise comparisons were not significant (Table S4).

280 Despite the lack of a common intake target, larvae were not consuming diets at random
281 (Fig 6b, Table S4c, d). For both carbohydrate and protein we saw differences in consumption
282 from what would have been expected for each larva based on random consumption, and this
283 effect was dependent on the specific set of diet choices (protein, $F_{4,20}=19.67$, $p<0.001$;
284 carbohydrate, $F_{4,20}=51.65$, $p<0.001$). When visualised as the amounts of protein and
285 carbohydrate consumed during each 48h treatment period (Fig. 7), it is clear that larvae were
286 achieving a degree of homeostasis in carbohydrate consumption (Fig 7b) compared to what
287 would be expected under random consumption of each diet choice (Fig 7a), whereas their
288 consumption of protein (Fig 7d) aligned closely with what would be expected under random
289 consumption (Fig 7c).

290

291 Discussion

292 We found that carbohydrate was positively associated with both body size and survival in
293 *Osmia bicornis* larvae (Figs. 4a, 5a), although within our range of dietary ratios we did not
294 specifically identify an optimum (fitness-maximising peak) in intake for either carbohydrate or
295 protein. Accordingly, given a choice, larvae converged on a relatively carbohydrate-biased
296 protein:carbohydrate ratio of 1:1.8 (Fig. 6a). Moreover, larvae prioritised carbohydrate over
297 protein intake, showing tighter control over carbohydrate consumption than over protein
298 consumption (Fig 7), and they pupated after eating about 0.23 g carbohydrate irrespective of
299 protein and of dietary dilution (Fig 3). Yet this carbohydrate target fell short of the amount of
300 carbohydrate that maximised cocoon weight or survival to pupation (Fig 4a, 5a). Dietary dilution
301 imposed costs upon larvae regardless of nutritional intake, in the form of greater mortality and
302 lower cocoon weights (Figs 4b, 5b). Taken together, these results show that (1) larval *O. bicornis*

303 are at least partially responsible for their own nutritional regulation, and (2) their performance
304 and consumption rules suggest adaptation to a pattern of carbohydrate-limited growth and
305 survival. In what follows, we suggest how and why these patterns in *O. bicornis* depart from
306 expected results based on studies of related organisms, and more generally what these findings
307 suggest about nutritional cooperation and/or conflict between parents and offspring in (mass)
308 provisioning species.

309 Larvae grew and survived best on our highest carbohydrate (i.e. lowest
310 protein:carbohydrate) diets. Accordingly, across our range of diets, larvae maintained a constant
311 carbohydrate intake while tolerating excesses or deficiencies of protein (a “no-interaction” rule
312 of compromise; Raubenheimer & Simpson 1999b) - although it is conceivable that alternative
313 rules of compromise, such as the “equal distance rule” more typically seen in generalist
314 herbivores (Raubenheimer & Simpson 1999b; Behmer 2009), might have been evident over a
315 broader array of diets. Both these patterns are unusual because insect herbivores are generally
316 considered to be limited by protein (e.g. Bernays & Chapman 2007; although see Le Gall &
317 Behmer 2014). In the few existing studies involving larval bees, e.g. honeybees (Helm *et al.*
318 2017), bumblebees (Kraus *et al.* 2019) and sweat bees (Roulston & Cane 2002), protein and not
319 carbohydrate mediated larval growth and/or survival. More broadly, insect larvae often grow and
320 survive best on balanced or moderately high protein:carbohydrate ratios (Roeder & Behmer
321 2014; Rodrigues *et al.* 2015), although low protein:carbohydrate ratios are associated with
322 longevity in adults (e.g. Lee *et al.* 2008). Moreover, animals generally prioritise regulation of the
323 nutrient that is typically limiting in their normal diet, and tolerate variation in nutrients that are
324 abundant (Raubenheimer & Simpson 1999a). Tolerance of wide variation in protein is thus
325 usually seen in predators (e.g. Raubenheimer *et al.* 2007; Kohl, Coogan & Raubenheimer 2015).
326 In contrast, herbivores often regulate protein more tightly than carbohydrate (Lee *et al.* 2002; Le
327 Gall & Behmer 2014; VanOverbeke, Thompson & Redak 2017). It is worth noting that protein
328 *did* weakly mediate survival in our larvae to some extent, although only at low carbohydrate -
329 possibly as a result of switching to protein as an energy source.

330 During the choice phase, when allowed to self-select diets, *O. bicornis* larvae converged
331 on a protein:carbohydrate ratio of 1:1.8. This ratio is considerably more carbohydrate-biased than
332 that preferred by bumblebees foraging on behalf of microcolonies (1:0.25, Vaudo *et al.* 2018;
333 1:0.08, Kraus *et al.* 2019), and (to a lesser extent) than ants foraging for colonies with offspring

334 (1:1.5, Dussutour & Simpson 2009). It is also more carbohydrate-biased than that selected by
335 reproductive, solitary phytophages such as grasshopper adults and lepidopteran larvae (1:0.25 -
336 1:1.4, reviewed in Behmer 2009) and is closer to diets selected by *Drosophila* larvae (1:2,
337 Rodrigues *et al.* 2015). Notably, though, 1:1.8 was more protein-biased than the ratio that we
338 found maximised both cocoon weight and survival (1:3.4), suggesting *O. bicornis* larvae may
339 choose diets that favour other fitness-related quantities (such as reproduction and/or
340 developmental time) over body size/survival, as in *Drosophila* (Lee *et al.* 2008; Rodrigues *et al.*
341 2015). As a cautionary note, the specific source of nutrients may also affect the preferred ratio:
342 for example, adult honeybees exhibited different target P:C ratios when fed different protein
343 sources (Altaye *et al.* 2010). Whether larvae are similarly sensitive is still unknown.

344 Two main features of *O. bicornis*' ecology may help to explain their prioritization of
345 carbohydrate, and their relative preference for this macronutrient, compared to what we know of
346 related taxa. First, the relative paucity of carbohydrate in *O. bicornis*' larval diet may help to
347 explain these findings. Despite being herbivorous, *Osmia* larvae are unlikely to be protein-
348 limited, because pollen is among the most protein-rich of plant tissues (Mattson 1980).
349 Moreover, in *Osmia* specifically, nectar constitutes only a tiny fraction of the pollen ball, less
350 than 4% (Maddocks & Paulus 1987; see Radmacher & Strohm 2010), in contrast to many other
351 bees where nectar is a principal source of carbohydrate for larvae (e.g. Kraus *et al.* 2019). *O.*
352 *bicornis* larvae may therefore be limited more by the amount of digestible carbohydrate within
353 pollen than by dietary protein (see Roulston & Cane 2000). Second, *O. bicornis* is (to our
354 knowledge) the first truly solitary hymenopteran studied under the GF; other studies have
355 concerned individuals likely to become workers of social species. Unlike social hymenopterans,
356 *O. bicornis* offspring are all reproductive and undergo diapause (Fliszkiewicz *et al.* 2012) - both
357 activities dependent on the fat body, where carbohydrate-derived fat is stored (Kawooya & Law
358 1988; Ziegler & Van Antwerpen 2006; Hahn & Denlinger 2007; Wasielewski *et al.* 2013). Thus,
359 *O. bicornis* larvae may have additional requirements for carbohydrate over and above those of
360 developing nonreproductive, nondiapausing hymenopteran workers. These contrasting findings
361 reinforce the idea that bees' nutritional needs may be just as diverse as their ecologies.

362 Although larvae retained the ability to regulate carbohydrate by over- or under-eating
363 protein, they nevertheless coped very poorly with dietary dilution (Fig 4b, 5b), despite displaying
364 compensatory feeding behaviour (Fig 2) that suggests they both detected and responded to such

365 dilution. The dilution was not excessive (70% nutrient density) compared to similar studies
366 offering very highly dilute diets (14%, Raubenheimer & Simpson 1993; 16.8%, Lee,
367 Raubenheimer & Simpson 2004). The locusts and caterpillars in those studies, though, are
368 adapted for diets that vary greatly in nutrient density, beginning dilute and becoming even more
369 dilute over the season (Scriber & Slansky 1981). By contrast, pollen is among the most
370 consistently nutrient-rich parts of a plant (Roulston & Cane 2000) and does not broadly vary in
371 composition over a season (DeGrandi-Hoffman *et al.* 2018). With a normal diet of unadulterated
372 pollen and very little nectar, *Osmia* larvae may have had no need to evolve mechanisms to cope
373 with dilution. In comparison, caterpillars reared on an invariant diet for generations lost the
374 ability both to regulate intake and to cope with dilution (Warbrick-Smith *et al.* 2009). *Osmia*
375 larvae appear to have retained the former capacity, but lost the latter, suggesting a normal diet
376 that is dense in nutrients, but variable in composition.

377 In systems where parents gather food for offspring from the environment, both parents
378 and offspring can be active participants in nutritional regulation. The lack of protein regulation
379 shown by *O. bicornis* larvae highlights the importance of understanding (a) whether mother *O.*
380 *bicornis* adjust protein content of provisions in response to imbalances in the landscape, and (b)
381 whether larvae have physiological adaptations (e.g. post-ingestive processing) for tolerating
382 protein imbalance. Budde & Lunau (2007) found that *O. bicornis* provisions contained about
383 19% protein regardless of pollen species used, suggesting a degree of homeostasis by parents.
384 Yet human activity is reducing floral diversity and quality (Ziska *et al.* 2016; Papanikolaou *et al.*
385 2017). Evidence is mixed concerning whether, in practice, parent bees assess pollen nutrients at
386 the flower (reviewed by Nicholls & Hempel de Ibarra 2016). Both bumblebees and ants balance
387 nutrition on behalf of colonies (Dussutour & Simpson 2009; Vaudo *et al.* 2018), regulating more
388 tightly when foraging for offspring - protein in the case of both taxa (Dussutour & Simpson
389 2009; Kraus *et al.* 2019) and carbohydrate in ants (Dussutour & Simpson 2008; Cook *et al.*
390 2010). On the other hand, protein gathered by honeybees varies passively with landscape usage
391 (while maintaining carbohydrate and lipid; Donkersley *et al.* 2014). Which regulatory strategy
392 *Osmia* parents and larvae collectively pursue may have important implications for their
393 vulnerability to human-induced landscape change, and so should now be a focus for research.

394 Additionally, the ability to discriminate among nutrients provided by parents may be one
395 tool offspring can use to exert some control over their nutrition, even in the absence of contact

396 with parents. *Osmia* parents may provide suboptimal resources simply because of inefficiency in
397 gathering pollen: efficiency drops across the season (Seidelmann 2006) and is lower in smaller-
398 bodied parents (Seidelmann, Ulbrich & Mielenz 2010). Moreover, less efficient parents actively
399 switch to producing male offspring (Seidelmann *et al.* 2010), so male and female offspring may
400 experience different selection for regulation. This is well documented in other groups (e.g.
401 Maklakov *et al.* 2008) and sex differences in larval regulation should now be a focus for
402 research. But it is also well known that the evolutionary interests of parents and offspring
403 frequently differ over how resources should be allocated (Trivers 1974; Crespi & Semeniuk
404 2004; Kilner & Drummond 2007; Haig 2010). The potential for offspring to use nutritional
405 regulation to mitigate parentally imposed costs has been relatively overlooked, as most research
406 to date has focused on parent-offspring conflict over *amount* of parental provisions, despite clear
407 potential for conflict over composition (e.g. in discus fish, Buckley *et al.* 2010). Among
408 primitively social Hymenoptera, some parents actively stunt offspring by restricting provisions
409 (Lawson *et al.* 2017), securing their help by forcing them to become workers (Craig 1983). But
410 the composition of food provided by parents is also critical to offspring fitness (e.g. Roulston &
411 Cane 2002) and in extreme cases caste-determining (Anderson 1984). *O. bicornis* are solitary
412 and lack castes, but this does not preclude parent-offspring disagreement over the optimal
413 balance of offspring nutrition, as in e.g. *Drosophila* (Rodrigues *et al.* 2015).

414 More broadly, understanding the relative roles of offspring (intake regulation and post-
415 ingestive processing) versus parents in nutrient balancing, as well as their evolutionary interests,
416 will be key to understanding the nutritional ecology of species with parental provisioning. Such
417 species include not just bees and other Hymenoptera, but other important ecosystem service
418 providers such as dung beetles (Frank *et al.* 2017) and burying beetles (Hopwood, Moore &
419 Royle 2013), as well as altricial birds (Wiens & Johnston 2012) and even humans (Burt & Amin
420 2014). Allorregulation by parents is not a given; the relative roles and interests of parent and
421 offspring in these groups are likely to reflect species' ecologies. Recent studies have found
422 nutritional mismatches between oviposition sites selected by parents and the nutritional
423 requirements of the offspring that will develop in those sites (Rodrigues *et al.* 2015; Lihoreau *et*
424 *al.* 2016). Parent sweat bees (*Lasioglossum zephyrum*) appear not to regulate protein in larval
425 provisions, despite protein mediating offspring performance (Roulston & Cane 2002). In *O.*
426 *bicornis*, we have shown that offspring retain the ability to regulate their nutritional intake

427 despite all food selection being done by parents whom they never meet. Larvae appeared to pay
428 closest attention to regulating dietary carbohydrate, consistent with this nutrient mediating both
429 growth and survival. Yet protein remains a key requirement for development; key now is to (a)
430 establish the nutritional rules used by parents when provisioning offspring, and whether these
431 coincide with or depart from those employed by larvae, and (b) establish specifically how protein
432 balance is achieved, and whether parents or larvae carry that responsibility.

433

434

435

436 **References**

- 437 Altaye, S.Z., Pirk, C.W.W., Crewe, R.M. & Nicolson, S.W. (2010) Convergence of
438 carbohydrate-biased intake targets in caged worker honeybees fed different protein
439 sources. *The Journal of experimental biology*, **213**, 3311–3318.
- 440 Anderson, M. (1984) The evolution of eusociality. *Annual review of ecology and systematics*,
441 **15**, 165–189.
- 442 Behmer, S.T. (2009) Insect herbivore nutrient regulation. *Annual review of entomology*, **54**,
443 165–187.
- 444 Bernays, E.A. & Chapman, R.F. (2007) *Host-Plant Selection by Phytophagous Insects*.
445 Springer Science & Business Media.
- 446 Buckley, J., Maunder, R.J., Foey, A., Pearce, J., Val, A.L. & Sloman, K.A. (2010) Biparental
447 mucus feeding: a unique example of parental care in an Amazonian cichlid. *The Journal*
448 *of experimental biology*, **213**, 3787–3795.
- 449 Budde, J. & Lunau, K. (2007) Rezepte für ein Pollenbrot--heute: *Osmia rufa*. *Entomologie*
450 *heute*, **19**, 173–179.
- 451 Burt, N.M. & Amin, M. (2014) A mini me?: exploring early childhood diet with stable isotope
452 ratio analysis using primary teeth dentin. *Archives of oral biology*, **59**, 1226–1232.
- 453 Cane, J.H. (2016) Adult Pollen Diet Essential for Egg Maturation by a Solitary *Osmia* Bee.
454 *Journal of insect physiology*, **95**, 105–109.
- 455 Cook, S.C., Eubanks, M.D., Gold, R.E. & Behmer, S.T. (2010) Colony-level macronutrient
456 regulation in ants: mechanisms, hoarding and associated costs. *Animal behaviour*, **79**,
457 429–437.
- 458 Costa, J.T. (2006) *The Other Insect Societies*. Harvard University Press.

- 459 Craig, R. (1983) Subfertility and the evolution of eusociality by kin selection. *Journal of*
460 *theoretical biology*, **100**, 379–397.
- 461 Crespi, B. & Semeniuk, C. (2004) Parent-offspring conflict in the evolution of vertebrate
462 reproductive mode. *The American naturalist*, **163**, 635–653.
- 463 Deans, C., Sword, G.A. & Behmer, S.T. (2019) First evidence of protein-carbohydrate
464 regulation in a plant bug (*Lygus hesperus*). *Journal of insect physiology*, **116**, 118–124.
- 465 DeGrandi-Hoffman, G., Gage, S.L., Corby-Harris, V., Carroll, M., Chambers, M., Graham, H.,
466 Watkins deJong, E., Hidalgo, G., Calle, S., Azzouz-Olden, F., Meador, C., Snyder, L. &
467 Ziolkowski, N. (2018) Connecting the nutrient composition of seasonal pollens with
468 changing nutritional needs of honey bee (*Apis mellifera* L.) colonies. *Journal of insect*
469 *physiology*, **109**, 114–124.
- 470 Despland, E. & Noseworthy, M. (2006) How well do specialist feeders regulate nutrient
471 intake? Evidence from a gregarious tree-feeding caterpillar. *The Journal of*
472 *experimental biology*, **209**, 1301–1309.
- 473 Donkersley, P., Rhodes, G., Pickup, R.W., Jones, K.C. & Wilson, K. (2014) Honeybee nutrition
474 is linked to landscape composition. *Ecology and evolution*, **4**, 4195–4206.
- 475 Dussutour, A. & Simpson, S.J. (2008) Carbohydrate regulation in relation to colony growth
476 in ants. *The Journal of experimental biology*, **211**, 2224–2232.
- 477 Dussutour, A. & Simpson, S.J. (2009) Communal nutrition in ants. *Current biology: CB*, **19**,
478 740–744.
- 479 Falk, S.J. (2015) *Field Guide to the Bees of Great Britain and Ireland*. British Wildlife
480 Publishing.
- 481 Fichter, B.L., Stephen, W.P. & Vandenberg, J.D. (1981) An Aseptic Technique for Rearing
482 Larvae of the Leafcutting Bee *Megachile rotundata* (Hymenoptera, Megachilidae).
483 *Journal of apicultural research*, **20**, 184–188.
- 484 Field, J. (2005) The evolution of progressive provisioning. *Behavioral ecology*, **16** (4), 770-
485 778
- 486 Filipiak, M. (2019) Key pollen host plants provide balanced diets for wild bee larvae: A
487 lesson for planting flower strips and hedgerows (ed R Rader). *The Journal of applied*
488 *ecology*, **56** (6), 1410-1418
- 489 Fliszkiewicz, M., Giejdasz, K., Wasielewski, O. & Krishnan, N. (2012) Influence of winter
490 temperature and simulated climate change on body mass and fat body depletion
491 during diapause in adults of the solitary bee, *Osmia rufa* (Hymenoptera: Megachilidae).
492 *Environmental entomology*, **41**, 1621–1630.

- 493 Frank, K., Brückner, A., Hilpert, A., Heethoff, M. & Blüthgen, N. (2017) Nutrient quality of
494 vertebrate dung as a diet for dung beetles. *Scientific reports*, **7**, 12141.
- 495 Goulson, D., Nicholls, E., Botías, C. & Rotheray, E.L. (2015) Bee declines driven by combined
496 stress from parasites, pesticides, and lack of flowers. *Science*, **347**, 1255957.
- 497 Hahn, D.A. & Denlinger, D.L. (2007) Meeting the energetic demands of insect diapause:
498 nutrient storage and utilization. *Journal of insect physiology*, **53**, 760–773.
- 499 Haider, M., Dorn, S., Sedivy, C. & Muller, A. (2014) Phylogeny and floral hosts of a
500 predominantly pollen generalist group of mason bees (Megachilidae: Osmiini).
501 *Biological journal of the Linnean Society. Linnean Society of London*, **111**, 78–91.
- 502 Haig, D. (2010) Colloquium papers: Transfers and transitions: parent-offspring conflict,
503 genomic imprinting, and the evolution of human life history. *Proceedings of the
504 National Academy of Sciences of the United States of America*, **107 Suppl 1**, 1731–1735.
- 505 Harper, E.J. & Turner, C.L. (2000) Nutrition and energetics of the canary (*Serinus canarius*).
506 *Comparative biochemistry and physiology. Part B, Biochemistry & molecular biology*,
507 **126**, 271–281.
- 508 Helm, B.R., Slater, G.P., Rajamohan, A., Yocum, G.D., Greenlee, K.J. & Bowsher, J.H. (2017)
509 The geometric framework for nutrition reveals interactions between protein and
510 carbohydrate during larval growth in honey bees. *Biology open*, **6**, 872–880.
- 511 Hogendoorn, K. & Velthuis, H.H.W. (1993) The sociality of *Xylocopa pubescens*: does a
512 helper really help? *Behavioral ecology and sociobiology*, **32**, 247–257.
- 513 Hopwood, P.E., Moore, A.J. & Royle, N.J. (2013) Nutrition during sexual maturation affects
514 competitive ability but not reproductive productivity in burying beetles (ed W
515 Blanckenhorn). *Functional ecology*, **27**, 1350–1357.
- 516 Hunt, J.H. & Nalepa, C.A. (1994) *Nourishment and Evolution in Insect Societies*. Westview
517 Press.
- 518 Jauker, F., Bondarenko, B., Becker, H.C. & Steffan-Dewenter, I. (2012) Pollination efficiency
519 of wild bees and hoverflies provided to oilseed rape. *Agricultural and forest
520 entomology*, **14**, 81–87.
- 521 Kawooya, J.K. & Law, J.H. (1988) Role of lipophorin in lipid transport to the insect egg. *The
522 Journal of biological chemistry*, **263**, 8748–8753.
- 523 Kilner, R.M. & Drummond, H. (2007) Parent--offspring conflict in avian families. *Journal of
524 ornithology*, **148**, 241–246.
- 525 Kohl, K.D., Coogan, S.C.P. & Raubenheimer, D. (2015) Do wild carnivores forage for prey or
526 for nutrients? *BioEssays: news and reviews in molecular, cellular and developmental*

- 527 *biology*, **37**, 701–709.
- 528 Kraus, S., Gómez-Moracho, T., Pasquaretta, C., Latil, G., Dussutour, A. & Lihoreau, M. (2019)
529 Bumblebees adjust protein and lipid collection rules to the presence of brood. *Current*
530 *zoology*, **65**, 437–446.
- 531 Lawson, S.P., Helmreich, S.L. & Rehan, S.M. (2017) Effects of nutritional deprivation on
532 development and behavior in the subsocial bee *Ceratina calcarata* (Hymenoptera:
533 Xylocopinae). *The Journal of experimental biology*, **220**, 4456–4462.
- 534 Lee, K.P., Behmer, S.T., Simpson, S.J. & Raubenheimer, D. (2002) A geometric analysis of
535 nutrient regulation in the generalist caterpillar *Spodoptera littoralis* (Boisduval).
536 *Journal of insect physiology*, **48**, 655–665.
- 537 Lee, K.P., Raubenheimer, D. & Simpson, S.J. (2004) The effects of nutritional imbalance on
538 compensatory feeding for cellulose-mediated dietary dilution in a generalist
539 caterpillar. *Physiological entomology*, **29**, 108–117.
- 540 Lee, K.P., Simpson, S.J., Clissold, F.J., Brooks, R., Ballard, J.W.O., Taylor, P.W., Soran, N. &
541 Raubenheimer, D. (2008) Lifespan and reproduction in *Drosophila*: New insights from
542 nutritional geometry. *Proceedings of the National Academy of Sciences of the United*
543 *States of America*, **105**, 2498–2503.
- 544 Le Gall, M. & Behmer, S.T. (2014) Effects of protein and carbohydrate on an insect
545 herbivore: the vista from a fitness landscape. *Integrative and comparative biology*, **54**,
546 942–954.
- 547 Lihoreau, M., Buhl, J., Charleston, M.A., Sword, G.A., Raubenheimer, D. & Simpson, S.J. (2014)
548 Modelling nutrition across organizational levels: from individuals to superorganisms.
549 *Journal of insect physiology*, **69**, 2–11.
- 550 Lihoreau, M., Poissonnier, L.-A., Isabel, G. & Dussutour, A. (2016) *Drosophila* females trade
551 off good nutrition with high-quality oviposition sites when choosing foods. *The Journal*
552 *of experimental biology*, **219**, 2514–2524.
- 553 Mackenzie, G., Boa, A.N., Diego-Taboada, A., Atkin, S.L. & Sathyapalan, T. (2015)
554 Sporopollenin, The Least Known Yet Toughest Natural Biopolymer. *Frontiers of*
555 *materials science*, **2**, 129.
- 556 Maddocks, R. & Paulus, H.F. (1987) Quantitative Aspekte der Brut-biologie von *Osmia rufa*
557 *L.* und *Osmia cornuta* Latr.(Hymenoptera, Megachilidae): Eine vergleichende
558 Untersuchung zu Mechanismen der Konkurrenzmindert zweier nahverwandter
559 Bienenarten. *Zoologische Jahrbücher. Abteilung für Systematik, Ökologie und*
560 *Geographie der Tiere*, **114**, 15–44.
- 561 Maklakov, A.A., Simpson, S.J., Zajitschek, F., Hall, M.D., Dessmann, J., Clissold, F.,
562 Raubenheimer, D., Bonduriansky, R. & Brooks, R.C. (2008) Sex-specific fitness effects

- 563 of nutrient intake on reproduction and lifespan. *Current biology*, **18**, 1062–1066.
- 564 Mattson, W.J. (1980) Herbivory in Relation to Plant Nitrogen Content. *Annual review of*
565 *ecology and systematics*, **11**, 119–161.
- 566 Mayntz, D., Raubenheimer, D., Salomon, M., Toft, S. & Simpson, S.J. (2005) Nutrient-specific
567 foraging in invertebrate predators. *Science*, **307**, 111–113.
- 568 Michaelsen, K.F., Weaver, L., Branca, F. & Robertson, A. (2003) *Feeding and Nutrition of*
569 *Infants and Young Children*. WHO Regional Publications, European Series.
- 570 Nelson, E.V., Roberts, R.B. & Stephen, W.P. (1972) Rearing Larvae of the Leaf-Cutter Bee
571 *Megachile Rotundata* on Artificial Diets. *Journal of apicultural research*, **11**, 153–156.
- 572 Nicholls, E. & Hempel de Ibarra, N. (2016) Assessment of pollen rewards by foraging bees.
573 *Functional ecology*, **31**, 76–87.
- 574 Papanikolaou, A.D., Kühn, I., Frenzel, M., Kuhlmann, M., Poschlod, P., Potts, S.G., Roberts,
575 S.P.M. & Schweiger, O. (2017) Wild bee and floral diversity co-vary in response to the
576 direct and indirect impacts of land use. *Ecosphere*, **8** (11), e02008.
- 577 Poissonnier, L.-A., Arganda, S., Simpson, S.J., Dussutour, A. & Buhl, J. (2018) Nutrition in
578 extreme food specialists: An illustration using termites. *Functional ecology*, **32**, 2531–
579 2541.
- 580 Radmacher, S. & Strohm, E. (2010) Factors affecting offspring body size in the solitary bee
581 *Osmia bicornis* (Hymenoptera, Megachilidae). *Apidologie*, **41**, 169–177.
- 582 Raubenheimer, D. & Jones, S.A. (2006) Nutritional imbalance in an extreme generalist
583 omnivore: tolerance and recovery through complementary food selection. *Animal*
584 *behaviour*, **71**, 1253–1262.
- 585 Raubenheimer, D., Mayntz, D., Simpson, S.J. & Tøft, S. (2007) Nutrient-specific
586 compensation following diapause in a predator: implications for intraguild predation.
587 *Ecology*, **88** (10), 2598–2608.
- 588 Raubenheimer, D. & Simpson, S.J. (1993) The geometry of compensatory feeding in the
589 locust. *Animal behaviour*, **45**, 953–964.
- 590 Raubenheimer, D. & Simpson, S.J. (1999a) Integrating nutrition: a geometrical approach.
591 *Proceedings of the 10th International Symposium on Insect-Plant Relationships*, Series
592 Entomologica, pp. 67–82. Springer Netherlands.
- 593 Raubenheimer, D. & Simpson, S.J. (1999b) Integrating nutrition: a geometrical approach.
594 *Entomologia experimentalis et applicata*, **91**, 67–82.
- 595 Raubenheimer, D., Simpson, S.J. & Mayntz, D. (2009) Nutrition, ecology and nutritional
596 ecology: toward an integrated framework. *Functional ecology*, **23**, 4–16.

- 597 Rehan, S.M., Richards, M.H., Adams, M. & Schwarz, M.P. (2014) The costs and benefits of
598 sociality in a facultatively social bee. *Animal behaviour*, **97**, 77–85.
- 599 Rodrigues, M.A., Martins, N.E., Balancé, L.F., Broom, L.N., Dias, A.J.S., Fernandes, A.S.D.,
600 Rodrigues, F., Sucena, É. & Mirth, C.K. (2015) *Drosophila melanogaster* larvae make
601 nutritional choices that minimize developmental time. *Journal of insect physiology*, **81**,
602 69–80.
- 603 Roeder, K.A. & Behmer, S.T. (2014) Lifetime consequences of food protein-carbohydrate
604 content for an insect herbivore (ed G Davidowitz). *Functional ecology*, **28**, 1135–1143.
- 605 Roulston, T.H. & Cane, J.H. (2000) Pollen nutritional content and digestibility for animals.
606 *Plant systematics and evolution = Entwicklungsgeschichte und Systematik der Pflanzen*,
607 **222**, 187–209.
- 608 Roulston, T.H. & Cane, J.H. (2002) The effect of pollen protein concentration on body size in
609 the sweat bee *Lasioglossum zephyrum* (Hymenoptera: Apiformes). *Evolutionary*
610 *ecology*, **16**, 49–65.
- 611 Roulston, T.H. & Goodell, K. (2011) The role of resources and risks in regulating wild bee
612 populations. *Annual review of entomology*, **56**, 293–312.
- 613 Royama, T. (1970) Factors governing the hunting behaviour and selection of food by the
614 great tit (*Parus major*). *The Journal of animal ecology*, **39**, 619–668.
- 615 Schmickl, T. & Karsai, I. (2017) Resilience of honeybee colonies via common stomach: A
616 model of self-regulation of foraging. *PloS one*, **12**, e0188004.
- 617 Scriber, J.M. & Slansky, F., Jr. (1981) The Nutritional Ecology of Immature Insects. *Annual*
618 *review of entomology*, **26**, 183–211.
- 619 Seidelmann, K. (2006) Open-cell parasitism shapes maternal investment patterns in the
620 Red Mason bee *Osmia rufa*. *Behavioral ecology: official journal of the International*
621 *Society for Behavioral Ecology*, **17**, 839–848.
- 622 Seidelmann, K., Ulbrich, K. & Mielenz, N. (2010) Conditional sex allocation in the Red Mason
623 bee, *Osmia rufa*. *Behavioral ecology and sociobiology*, **64**, 337–347.
- 624 Simpson, S.J. & Raubenheimer, D. (1993) A Multi-Level Analysis of Feeding Behaviour: The
625 Geometry of Nutritional Decisions. *Philosophical transactions of the Royal Society of*
626 *London. Series B, Biological sciences*, **342**, 381–402.
- 627 Simpson, S.J. & Raubenheimer, D. (2012) The nature of nutrition: a unifying framework.
628 *Australian journal of zoology*, **59**, 350–368.
- 629 Smiseth, P.T., Wright, J. & Kölliker, M. (2008) Parent-offspring conflict and co-adaptation:
630 behavioural ecology meets quantitative genetics. *Proceedings. Biological sciences / The*

- 631 *Royal Society*, **275**, 1823–1830.
- 632 Strohm, E., Daniels, H., Warmers, C. & Stoll, C. (2002) Nest provisioning and a possible cost
633 of reproduction in the megachilid bee *Osmia rufa* studied by a new observation
634 method. *Ethology Ecology & Evolution*, **14**, 255–268.
- 635 Tainsh, F., Woodmansey, S.R., Austin, A.J., Bagnall, T.E. & Gilbert, J.D.J. (2020) Sporopollenin
636 as a dilution agent in artificial diets for solitary bees. *Apidologie*,
637 <https://doi.org/10.1007/s13592-020-00801-1>
- 638 Trivers, R.L. (1974) Parent-Offspring Conflict. *Integrative and comparative biology*, **14**,
639 249–264.
- 640 VanOverbeke, D.R., Thompson, S.N. & Redak, R.A. (2017) Dietary self-selection and rules of
641 compromise by fifth-instar *Vanessa cardui*. *Entomologia experimentalis et applicata*,
642 **163**, 209–219.
- 643 Vaudo, A.D., Farrell, L.M., Patch, H.M., Grozinger, C.M. & Tooker, J.F. (2018) Consistent
644 pollen nutritional intake drives bumble bee (*Bombus impatiens*) colony growth and
645 reproduction across different habitats. *Ecology and evolution*, **8** (11), 5765–5776
- 646 Vaudo, A.D., Patch, H.M., Mortensen, D.A., Tooker, J.F. & Grozinger, C.M. (2016)
647 Macronutrient ratios in pollen shape bumble bee (*Bombus impatiens*) foraging
648 strategies and floral preferences. *Proceedings of the National Academy of Sciences of the*
649 *United States of America*, **113**, E4035–42.
- 650 Warbrick-Smith, J., Raubenheimer, D., Simpson, S.J. & Behmer, S.T. (2009) Three hundred
651 and fifty generations of extreme food specialisation: testing predictions of nutritional
652 ecology. *Entomologia experimentalis et applicata*, **132**, 65–75.
- 653 Wasielewski, O., Wojciechowicz, T., Giejdasz, K. & Krishnan, N. (2013) Overwintering
654 strategies in the red mason solitary bee—physiological correlates of midgut metabolic
655 activity and turnover of nutrient reserves in females of *Osmia bicornis*. *Apidologie*, **44**,
656 642–656.
- 657 Weeks, R.D., Wilson, L.T., Vinson, S.B. & James, W.D. (2004) Flow of Carbohydrates, Lipids,
658 and Protein Among Colonies of Polygyne Red Imported Fire Ants, *Solenopsis invicta*
659 (Hymenoptera: Formicidae). *Annals of the Entomological Society of America*, **97**, 105–
660 110.
- 661 Wiens, J. & Johnston, R. (2012) Adaptive correlates of granivory in birds. *Granivorous Birds*
662 *in Ecosystems: Their Evolution, Populations, Energetics, Adaptations, Impact and*
663 *Control*, (Eds J. Pinowski and SC Kendeigh.) pp, 301–340.
- 664 Ziegler, R. & Van Antwerpen, R. (2006) Lipid uptake by insect oocytes. *Insect biochemistry*
665 *and molecular biology*, **36**, 264–272.

666 Ziska, L.H., Pettis, J.S., Edwards, J., Hancock, J.E., Tomecek, M.B., Clark, A., Dukes, J.S., Loladze,
667 I. & Polley, H.W. (2016) Rising atmospheric CO₂ is reducing the protein concentration
668 of a floral pollen source essential for North American bees. *Proceedings. Biological*
669 *sciences / The Royal Society*, **283** (1828), 20160414
670
671
672

673 **Tables & Figures**

674

675 **Table 1. Sample sizes for each diet combination used for choice phase (allocated by random**
676 **coin toss). “Order” refers to diet order - e.g. for A1C1, Order 1 would receive A1 first**
677 **whereas Order 2 would receive C1 first, determined by coin toss. Surviving larvae are in**
678 **parentheses.**

	Order 1	Order 2	Total
A1C1	1 (1)	6 (5)	7 (6)
A1C2	5 (5)	4 (3)	9 (8)
A2C1	3 (2)	5 (5)	8 (7)
A2C2	5 (2)	3 (1)	8 (3)

679

680

681

682 **Figure captions**

683

684 **Figure 1. (a)** Expected protein and carbohydrate consumption if larvae ate indiscriminately
685 between two diets. Diet choices are pairwise combinations of diets A1, A2, C1 and C2, which
686 each contain protein and carbohydrate at different ratios and concentrations. Solid lines represent
687 P:C ratios; black points represent actual nutrient content of each diet, which depends upon
688 dilution as well as P:C ratio. Red points represent expected consumption if larvae eat randomly
689 (i.e. equally) from each of a choice of two diets (choices denoted by the red point labels). **(b)**
690 schematic describing how larvae were assigned to each diet grouping. Coloured arrows show the
691 period in days that each larva was fed a particular diet.

692

693 **Figure 2.** Amount of provision in grams consumed by larvae raised on the 3 different P:C ratio
694 artificial diets at the 2 different macronutrient concentrations (90% and 70% macronutrient
695 content).

696

697 **Figure 3.** Mean total (+/- 1 SE) amount of P and C consumed in grams by larvae on each
698 diet before pupation. Solid lines and letters represent three P:C ratios (A = 1:1.2, B = 1:2.3,
699 C = 1:3.4). Numbers following letters denote diet concentration (1 = 90%, 2 = 70%). Dotted
700 lines show global mean consumption of each nutrient.

701

702 **Figure 4.** Effects of P and C consumption upon cocoon weight (g) in larvae fed diets at (a)
703 90% and (b) 70% nutrient density. Transition from blue to red indicates heavier cocoons.
704 For context, mean total consumption of P and C for each diet is plotted (white points; data
705 as in Fig. 2) alongside raw data (grey points). Solid lines and letters represent three P:C
706 ratios (A = 1:1.2, B = 1:2.3, C = 1:3.4).

707

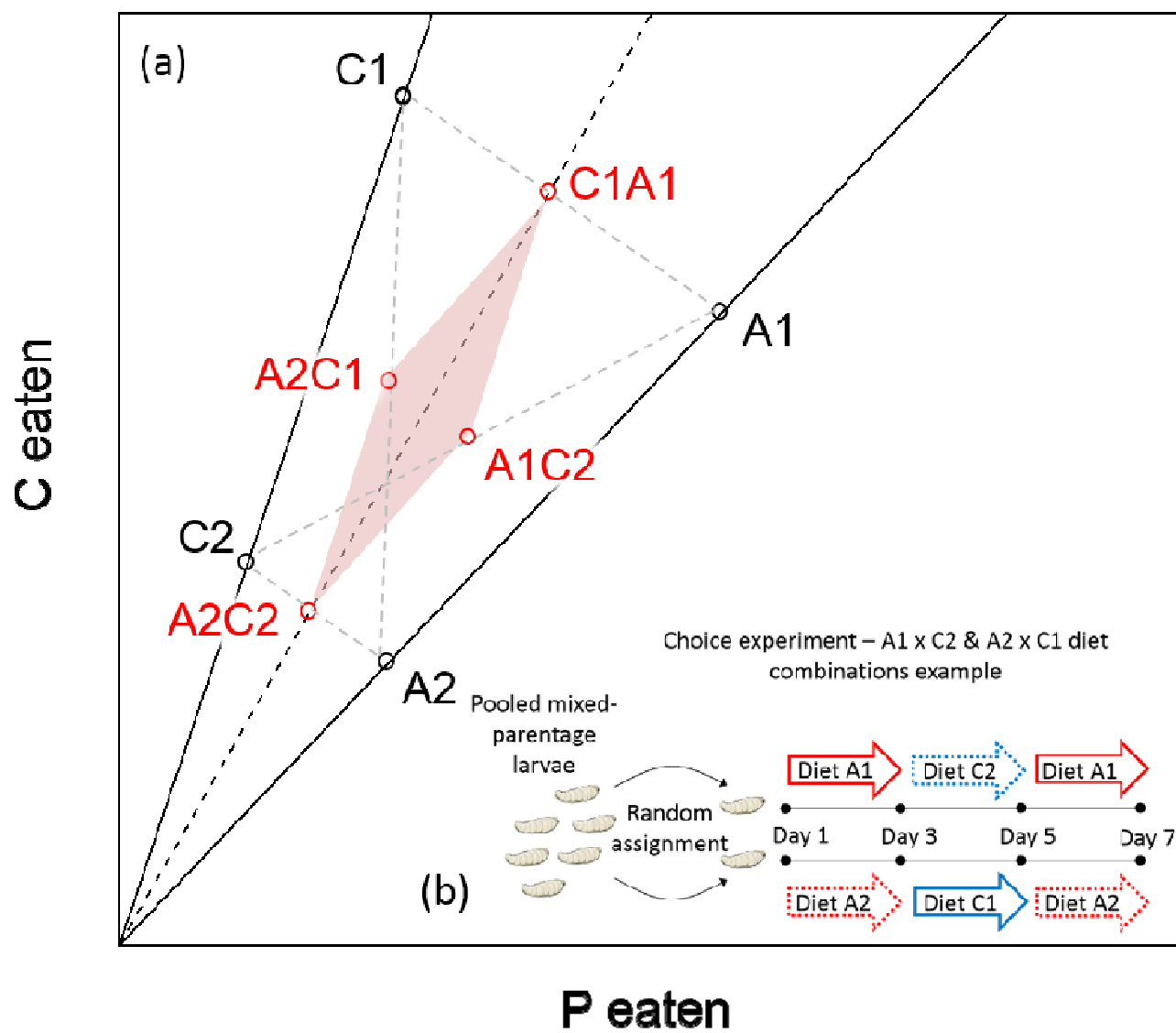
708 **Figure 5.** Effects of P and C consumption upon estimated survival time (colour) in larvae
709 fed diets at (a) 90% and (b) 70% nutrient density. Transition from blue to red indicates
710 longer survival. Black points, dead larvae; white points, larvae surviving to pupation. For
711 context, mean total consumption of P and C for each diet is plotted (large white points; data
712 as in Fig. 2). Solid lines and letters represent three P:C ratios (A = 1:1.2, B = 1:2.3, C = 1:3.4).

713

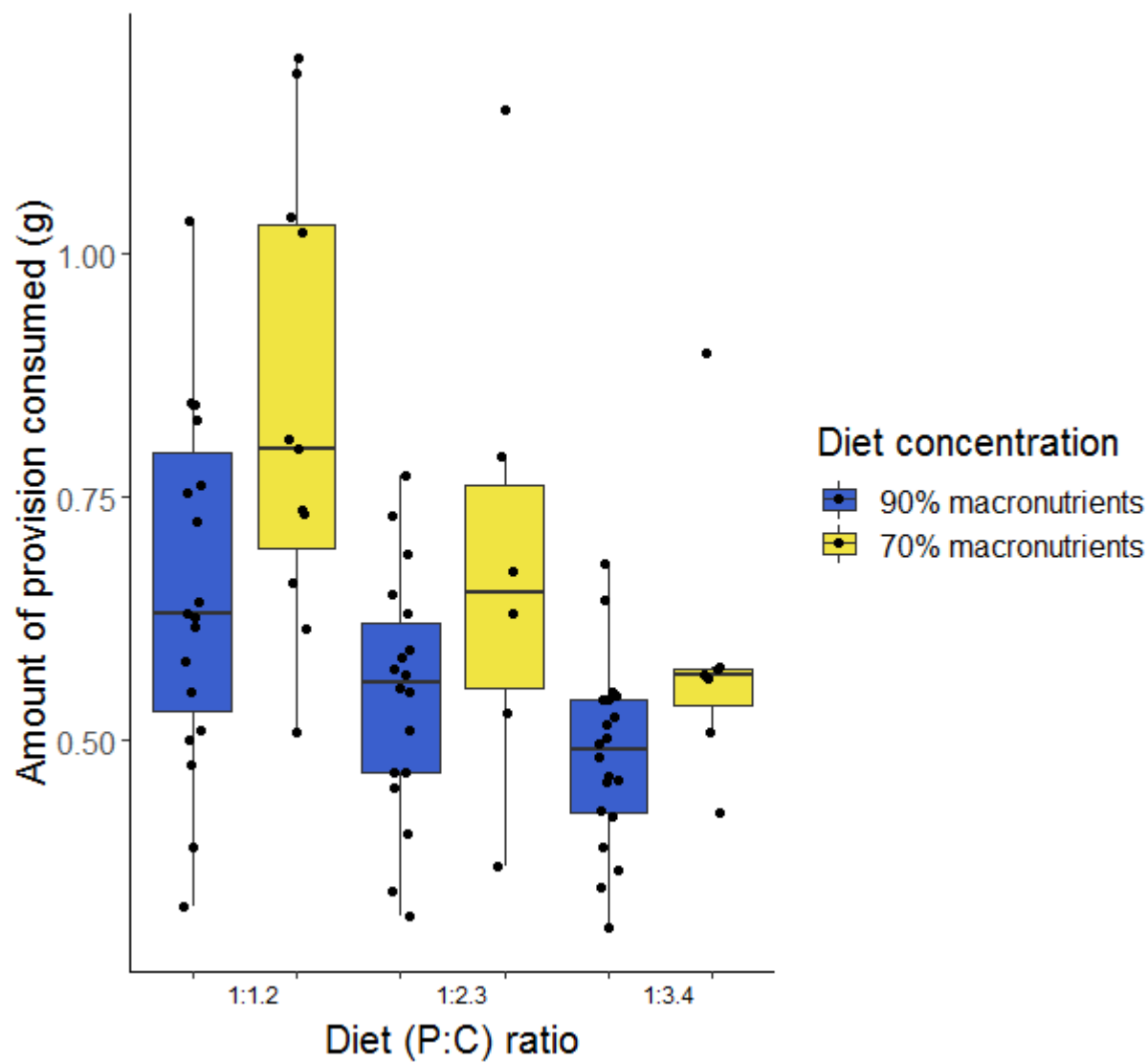
714 **Figure 6. (a)** The mean (\pm SE) amount of protein (P) and carbohydrate (C) eaten by
715 larvae in the choice experiment. Each point label denotes a choice of two diets, one A and
716 one C; black labels show observed intake, red labels show expected intake under random
717 consumption. Letters represent diet P:C ratio (A = 1:1.2, C = 1:3.4); numbers represent diet
718 concentration (1 = 90%, 2 = 70%), hence, for example, “A2C1” represents the pairing of
719 diet A2 with diet C1. Solid lines represent dietary P:C ratios (Top line = Diet C, Bottom line
720 = Diet A). Dashed red line shows expected average P:C ratio based on random consumption.
721 Dashed black line shows average P:C ratio of observed intake across larvae. (b) Deviation
722 from random intake of protein and carbohydrate for larvae in different treatment groups
723 during the choice phase. Treatment groups are given in order of overall diet concentration.
724 Bars with similar letters displayed above or below are not statistically significantly
725 different (Tukey’s post-hoc comparisons).

726

727 **Figure 7. (a, c)** Mean expected intake over successive diet swaps assuming random
728 consumption of diets (\pm 1 SE, inner ribbon, and SD, outer ribbon) of protein (red, lower
729 ribbons) and carbohydrate (blue, upper ribbons), irrespective of the concentrations of the
730 diet choices, for larvae starting on (a) diet A or (c) diet C. (b, d) Nutrient intake actually
731 observed for larvae starting on (b) diet A or (d) diet C (\pm 1 SE, inner ribbon, and SD, outer
732 ribbon). For details of calculations of expected consumption, see text. Swap 11 lacks
733 confidence intervals because only one larva in each group reached this stage.



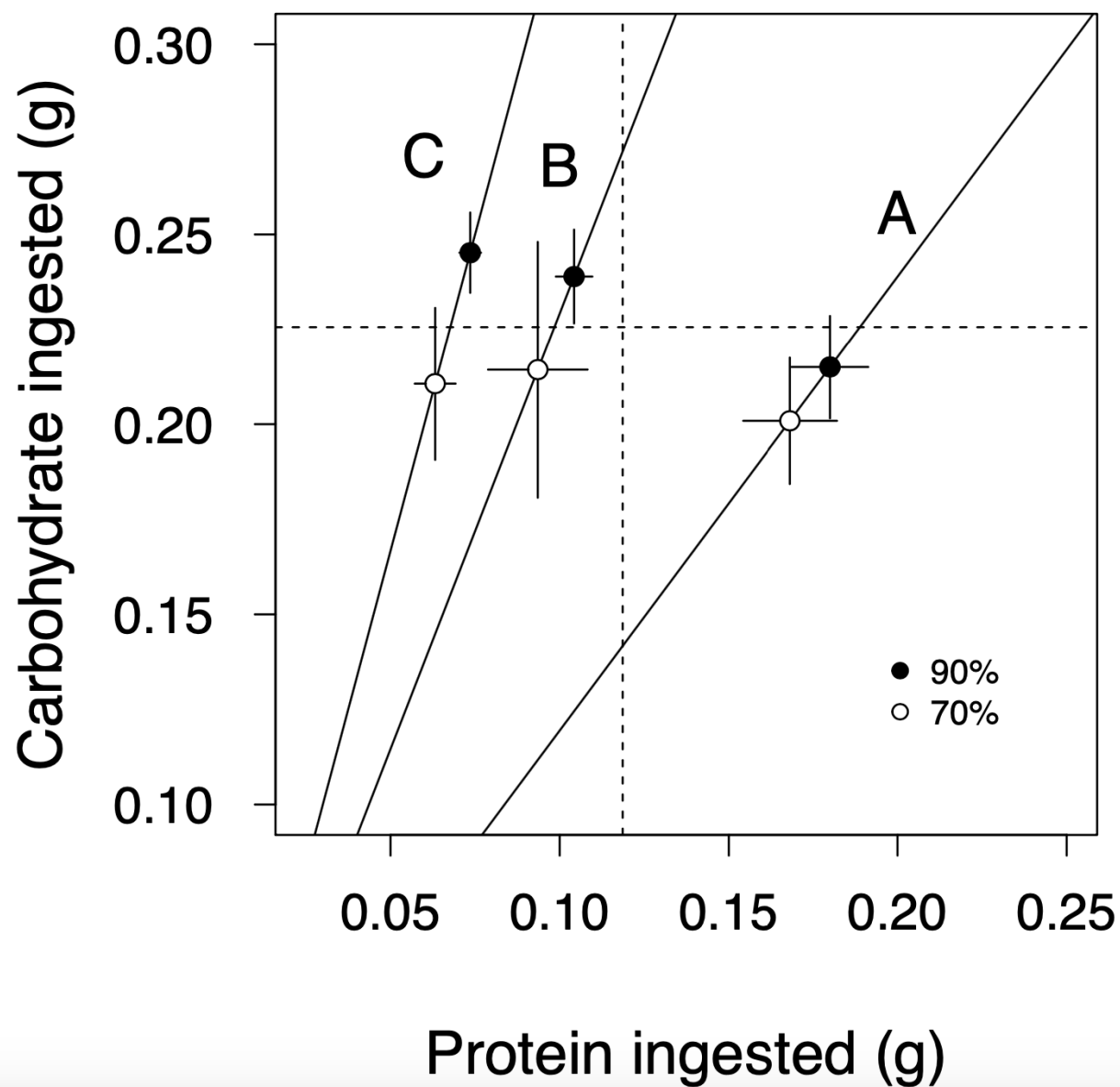
734
735 Fig. 1.



736

737 Fig. 2.

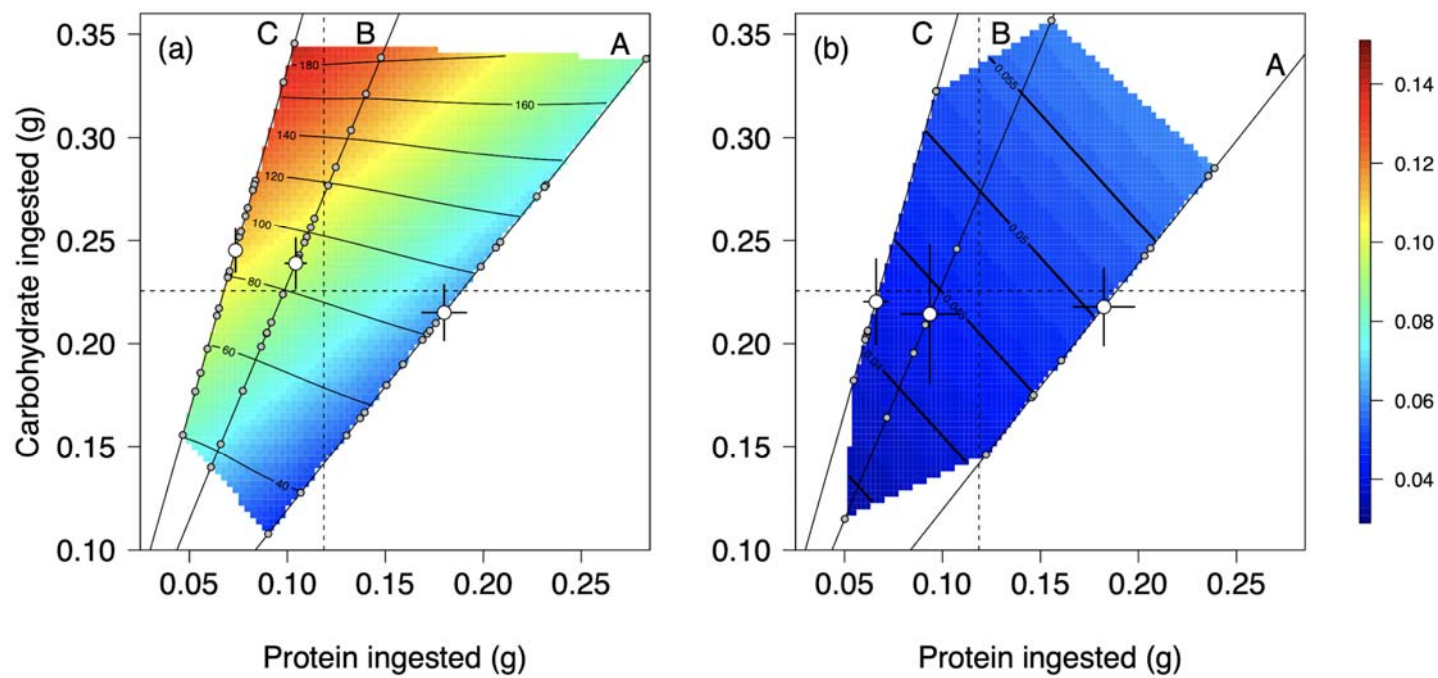
738



739

740 Fig. 3.

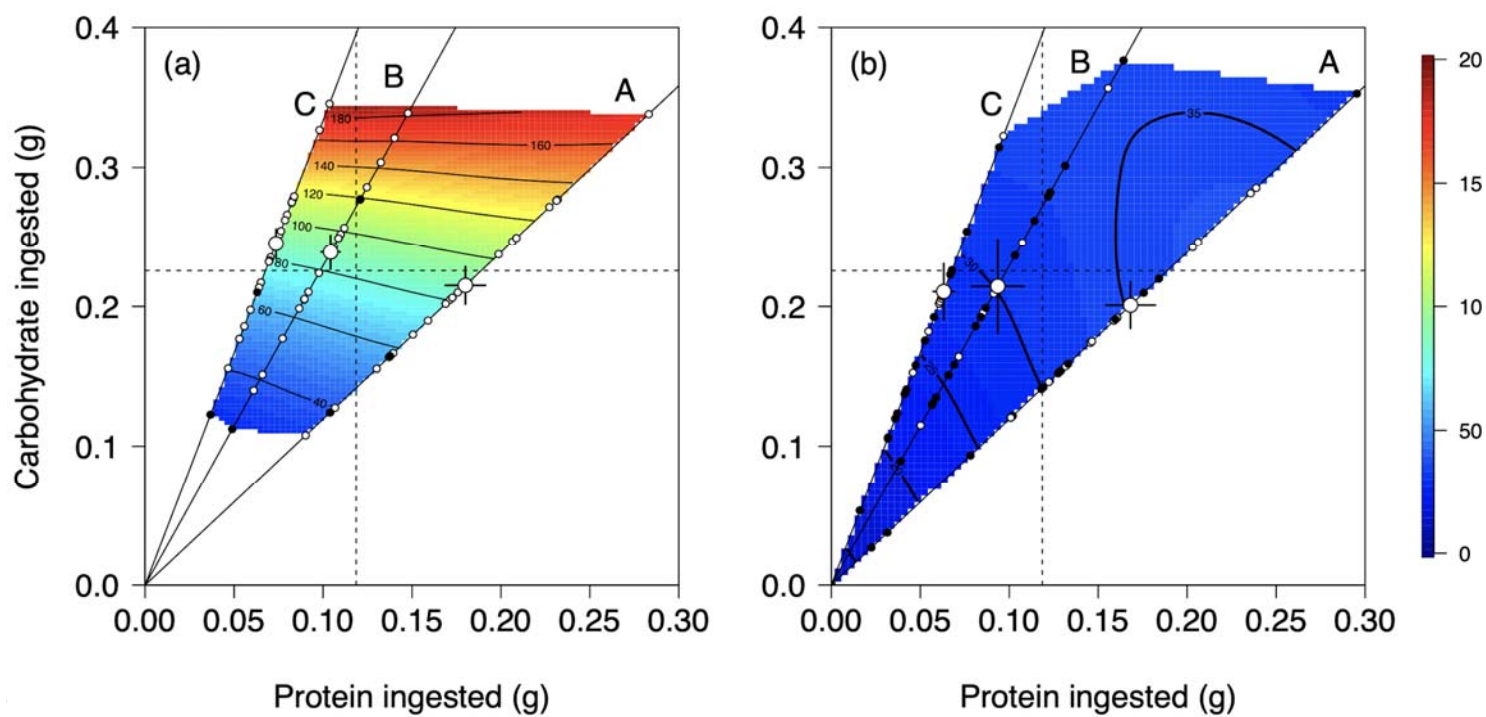
741



74;
743 Fig. 4.

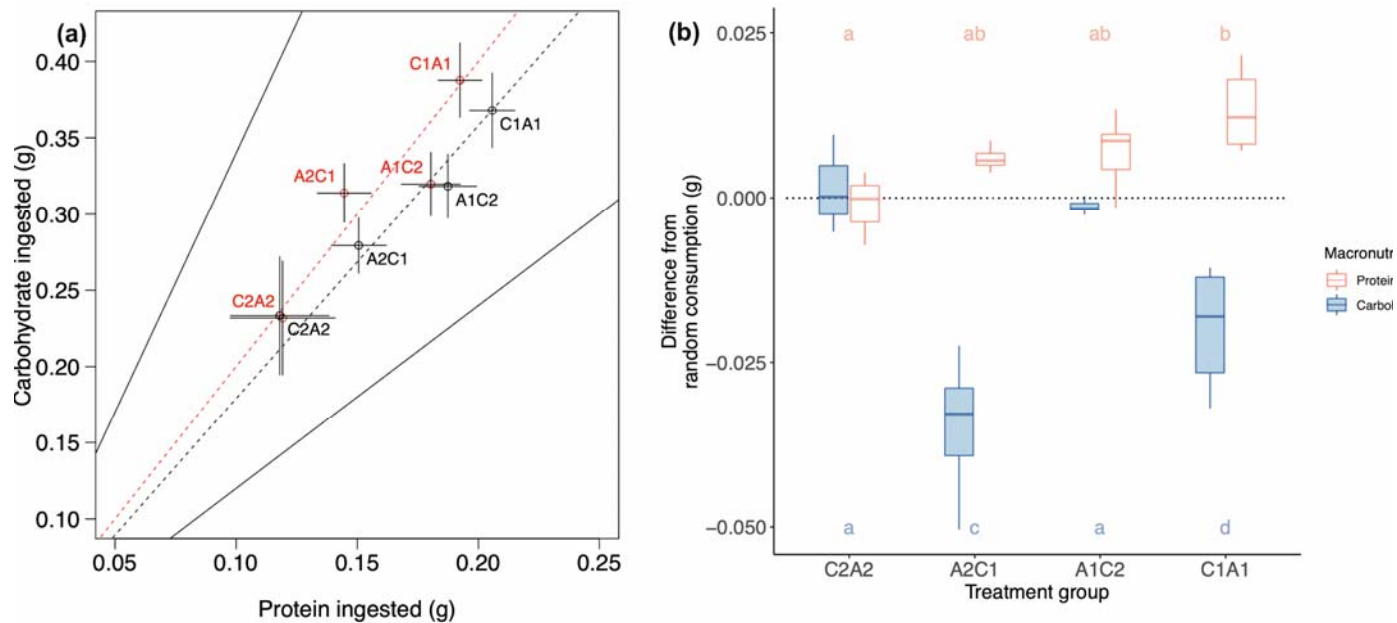
744

745



747 Fig. 5.

748

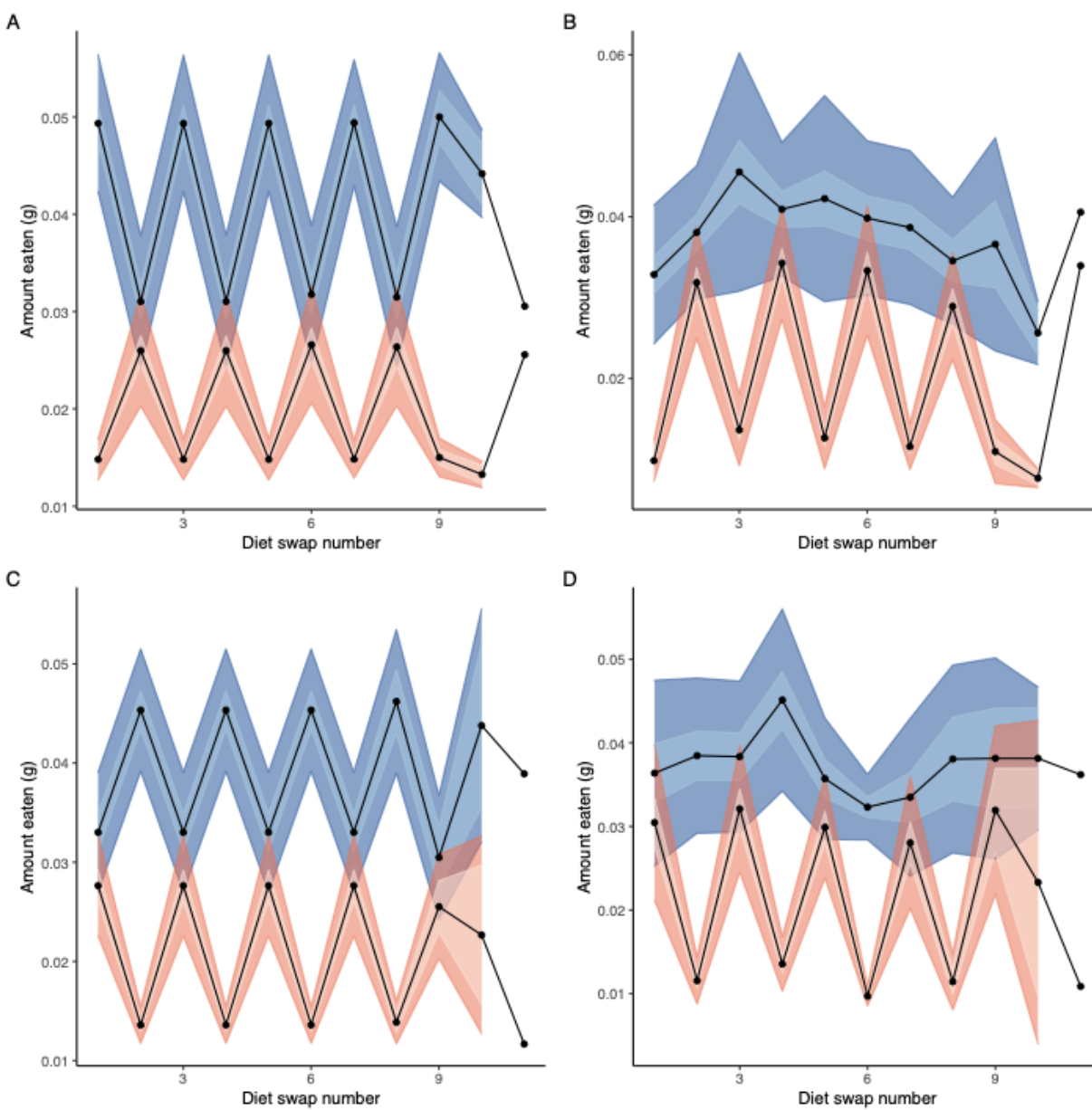


749

750 Fig. 6.

751

752



753

754 Fig. 7.

755

756