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Adaptive evolution of sperm proteins depends on sperm competition in a pair of Lepidoptera

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26 **Abstract**

27 Sperm are among the most variable cells in nature. Both within and between species, sperm display
28 more diversity in form than one would expect of an ostensibly single-function cell. Morphologically,
29 some of this variation within species has been demonstrated to be non-adaptive, but for many species
30 that consistently produce multiple sperm morphs, the significance of variable sperm remains unknown.
31 Here, we investigate the molecular evolution of dimorphic sperm in Lepidoptera, the butterflies and
32 moths; males of this order produce both fertilizing eupyrene sperm and a secondary apyrene type that
33 lacks DNA. Based on population genetic analyses in two species, the monandrous Carolina sphinx moth
34 and the highly polyandrous monarch butterfly, we see evidence for increased selection in fertilizing
35 sperm, but only in the polyandrous species. This signal comes primarily from a decrease in non-
36 synonymous polymorphism in sperm proteins compared to the rest of the genome, indicative of strong
37 purifying selection. Additionally, rates of non-synonymous divergence are comparable between sperm
38 genes and the rest of the genome, suggesting that many alleles reach fixation owing to positive selection
39 as well. Investigation of the distribution of fitness effects of new non-synonymous mutations in monarch
40 sperm confirms stronger selection on sperm proteins in monarchs, with very few neutral variants and a
41 preponderance of deleterious and beneficial variants. These results mirror findings on sperm evolution
42 in other taxa; increased sperm competition decreases within-population morphological variation. Our
43 results suggest that sperm competition can be a powerful selective force at the sequence level as well.

44 **Introduction**

45 Sperm cells display remarkable diversity throughout the animal kingdom (Pitnick et al. 2009), from small
46 and plentiful to gigantic (Pizzari 2006) or super-structure-forming (Higginson et al. 2012). Indeed, this
47 variation exists at every level, from fixed differences between species to variability within individual
48 males (Buckland-Nicks 1998; Swallow and Wilkinson 2002; Tavares-Bastos et al. 2002; Marks et al. 2008;
49 Sasakawa 2009). In many independently evolved cases, males consistently produce two different sperm
50 types in a phenomenon known as sperm dimorphism. Intriguingly, in all cases examined, only one of the
51 two is capable of fertilization (Wilms 1986; Eckelbarger et al. 1989; Bressac et al. 1991; Carcupino et al.
52 1999; Sasakawa 2009). The evolutionary causes and consequences of variation in sperm morphology
53 both within and between morphs are immediately intriguing. As gametes, these cells are the final step in
54 the long chain of events leading to reproductive success. Why should such important components of
55 fitness be so variable? Different fields of research have pursued largely independent explanations.

56 Morphological and behavioral reproductive biologists note that much of this diversity, at least within
57 sperm morphs, can be attributed to deleterious variation, e.g. genetic defects (Chenoweth 2005) or age-
58 related decline in sperm quality (Preston et al. 2015). This deleterious variation has also been shown to
59 be inversely correlated with rates of sperm competition between species (Kleven et al. 2008), *i.e.* taxa
60 that experience more sperm competition tend to have less morphologically variable sperm at both
61 population and individual levels. From this perspective, sperm vary *in spite of* constraint imposed by
62 their reproductive importance, with postcopulatory selection through sperm competition and cryptic
63 female choice weeding out the suboptimal sperm variants in species with high rates of polyandry
64 (Birkhead 1998; Immler et al. 2008). By extension of this paradigm, non-fertilizing sperm are often
65 posited as specialized agents of sperm competition, acting as final combatants in male-male competition
66 (Buckland-Nicks 1998; Swallow and Wilkinson 2002; Buckland-Nicks et al. 2010), though conclusive
67 evidence to demonstrate as much is less abundant.

68 Separately, molecular biologists and geneticists have been mostly concerned with understanding the
69 diversity of sperm (and other reproductive) proteins between species. One common observation is that
70 reproductive proteins diverge rapidly between species. These proteins often appear as outliers
71 compared to the rest of the genome and, in many cases, adaptive evolution is implicated as the cause of
72 this elevated divergence (Civetta and Singh 1995; Willie J. Swanson and Vacquier 2002; Dorus et al.
73 2004). Typical reasons proposed include roles in sexual antagonism or speciation through post-
74 copulatory, pre-zygotic isolation (Willie J Swanson and Vacquier 2002; Martin and Hosken 2004). These
75 arguments posit that sperm and other reproductive proteins vary precisely *because of* their importance
76 to fitness. However, experimental evidence to demonstrate the role of adaptation here is far less
77 common (but see Swanson and Vacquier 1998). Moreover, high rates of non-adaptive variation within
78 species could also generate elevated divergence in evolutionary time when coupled with relaxed
79 selection (Willie J Swanson and Vacquier 2002; Willie J. Swanson and Vacquier 2002; Gershoni and
80 Pietrokovski 2014). Only recently have theoreticians attempted to unify these patterns of molecular
81 evolution with the pressures exerted by sperm competition (Dapper and Wade 2016).

82 As explained by Nearly Neutral Theory (Ohta 1992), the efficacy of selection on a set of proteins
83 depends on the effective population size on which selection can act. Genes encoding reproductive
84 proteins are typically expressed in only half of a given population (males or females), thus cutting in half
85 the size of the population under selection (Whitlock and Wade 1995; Barker et al. 2005). Furthermore,
86 proteins involved in postcopulatory events like sperm competition may only experience selection in the

87 presence of a competitor's proteins, which substantially decreases the opportunity for selection if
88 females seldom mate more than once in a breeding season. This logic predicts that reproductive
89 proteins can diverge more quickly than the rest of the genome due to relaxed selection and that
90 adaptive evolution, particularly in sperm proteins, should only be a significant force in species with high
91 rates of polyandry (Dapper and Wade 2016). Few molecular evolutionary studies have sought to
92 disentangle the signal of adaptive evolution from the more easily observed patterns of divergence, and
93 no studies have addressed the function of dimorphic sperm from this paradigm. Here, we present the
94 first molecular evolutionary investigation of dimorphic sperm, integrating perspectives from both
95 morphological reproductive biology and theoretical population genetics to better understand some of
96 the most perplexing dimorphic sperm observed in the animal kingdom.

97 *Sperm Dimorphism in Lepidoptera*

98 Males of almost all species in the order Lepidoptera (butterflies and moths) produce two distinct sperm
99 types: one fertilizing sperm type (**eupyrene**) and a second type (**apyrene**) which lacks a nucleus and
100 nuclear DNA (Meves 1902). The function of apyrene sperm is poorly understood, but it is incapable of
101 fertilizing eggs. Nevertheless, its production is hormonally regulated and occurs in a developmentally
102 predictable way, implying a novel gain of function rather than loss of fidelity in spermatogenesis
103 (Friedlander 1997), and evidence from organismal studies suggests that it plays some functional role(s)
104 in reproduction (Takemura et al. 2006). Males can apparently control the ratio of the two sperm types in
105 their ejaculate and typically transfer to females 10-20 times as much apyrene sperm as eupyrene sperm
106 (Oberhauser 1988), leading some to suggest that apyrene sperm play a role in sperm competition
107 (Silberglied et al. 1984; Swallow and Wilkinson 2002). Here, we assess patterns of both polymorphism
108 and divergence among sperm proteins using two newly published proteomic datasets from both
109 eupyrene and apyrene sperm of two species: the monarch butterfly, *Danaus plexippus* (Whittington et al.
110 2017), and the Carolina sphinx moth, *Manduca sexta* (Whittington et al. 2015).

111 While monarchs are typically studied to understand migration, insect-plant interactions, and disease
112 ecology (de Roode et al. 2007; de Roode et al. 2008; Merlin et al. 2013), and Carolina sphinx moths are
113 studied for physiology, immunology, and pest management (Kanost et al. 2004; Soberón et al. 2007;
114 Sears et al. 2012), the pair also present an interesting contrast in mating ecology. North American
115 monarchs spend time at incredibly high density in overwintering colonies in Mexico (Urquhart 1976).
116 Owing to these unique population dynamics, female remating rates are among the highest observed in
117 Lepidoptera. Females mate up to 14 times in the wild (Hill Jr. et al. 1976; Smith 1984), creating ample

118 opportunity for sperm competition. In contrast, Carolina sphinx moths are typically monandrous (Snow
119 et al. 1974), making sperm competition rarely relevant as a selective force.

120 We investigate the evolutionary genetics of these insects' sperm proteomes with two goals: to examine
121 patterns of selection on sperm proteins between two species with differing mating systems and, if
122 possible, to detect signatures of differing selection between sperm morphs within species. We also
123 tested the general predictions for relaxed selection in sex-limited proteins using RNA-seq datasets for
124 these two species from previously published data for *Manduca sexta* (Cao and Jiang 2017) and newly
125 generated data for the monarch butterfly (summarized in Table S2). To do complete population genetic
126 analyses, we have also generated the first published set of whole-genome resequencing data for
127 *Manduca sexta* from a wild population.

128 **Results**

129 *Differences Between Sperm Proteins and the Genome Background*

130 First, we considered the sperm proteome as a whole and compared genes found in sperm to those in
131 the genome background (here defined as all protein coding genes not present in the sperm proteome).
132 Due to the lower gene count of the Z sex chromosome compared to the autosomes and the
133 complications of mixed-sex sampling creating differing allele counts between autosomes and sex
134 chromosomes, estimates from Z-linked genes were not investigated in these or subsequent analyses. In
135 both species this subset contained >90% of the sperm proteome (Mongue and Walters 2017). Using
136 permutation tests, we found no difference in the proportion of adaptive substitutions (α) between the
137 sperm proteome and the rest of the genome in *Manduca sexta* ($p = 0.40892$, Figure 1A, left); for
138 monarchs, however, the sperm proteome showed a significantly greater proportion of adaptive
139 substitutions than the rest of the genome ($p = 0.00006$, Figure 1A, right).

140 To better understand the source of these results, we investigated the individual components of α : non-
141 synonymous polymorphism (P_n), synonymous polymorphism (P_s), non-synonymous divergence (D_n),
142 and synonymous divergence (D_s). We compared the scaled estimates of each (e.g. non-synonymous
143 polymorphisms per non-synonymous site) to the genome background within each species using a
144 Wilcoxon-Mann-Whitney test (Figure 1B). We found no differences between sperm and the genome
145 background for any class of variants in *M. sexta* (P_n : $W = 3014100$, $p = 0.5964$; P_s : $W = 2879300$, $p =$
146 0.1830 ; D_n : $W = 3068300$, $p = 0.2009$; D_s : $W = 2895700$, $p = 0.2686$). The signal for elevated α in
147 monarchs came primarily from P_n , which was greatly depressed in sperm ($W = 3062400$; $p = 3.224 \times 10^{-6}$).

148 ¹¹) while other classes were comparable between sperm and the genome background (Ps: W = 2684200,
149 p = 0.2720; Dn: W = 2506400, p = 0.1300; Ds: W = 2544400, p = 0.3437).

150 Next, we leveraged orthology, as established by Whittington *et al.* (2017), to further examine the
151 apparent difference in selection occurring in monarch sperm proteins. A substantial portion of the
152 monarch sperm proteome (~42%, 216 genes, Figure 2A) shares an ortholog in the sperm proteome of *M.*
153 *sexta*; reciprocally, there are 236 genes in the *Manduca* sperm proteome that share an ortholog in the
154 monarch sperm proteome (due to a few one-to-many orthologs). These proteins, hereafter referred to
155 as *sperm homologs*, offer the opportunity to directly assess the selective pressures experienced by the
156 same genes with conserved function but found in species with different levels of postcopulatory
157 selection. We tested for differences in adaptive evolution between sperm homologs (containing an
158 ortholog in the other species' sperm proteome) and proteins unique to one sperm proteome (orthology
159 outside of sperm or no detectable orthology).

160 In *Manduca*, genes of these two classes did not differ in the proportion of adaptive substitutions with
161 permutation testing (p = 0.6174, Figure 2B). In monarchs, we detected an increased proportion of
162 adaptive substitution in the sperm homologs (p = 0.0372, Figure 2B). In fact, comparing between species,
163 these sperm homologs had much higher α values in monarchs than in Carolina sphinx moths (p =
164 0.00008), while genes with unique expression in either species did not show differences between
165 species (p = 0.5922).

166 *Site-frequency-based methods*

167 Based on different patterns of selection between sperm proteins and the genome background in
168 monarch butterflies, we investigated the distribution of fitness effects (DFE) of new non-synonymous
169 mutations in these genes. Using the same samples as above, we generated site frequency spectra to
170 estimate the DFE and α for both the Carolina sphinx and monarch genome background, whole sperm
171 proteome, and sperm homolog subset using a more complex likelihood model in the program polyDFE
172 (Tataru *et al.* 2017).

173 The distribution of fitness effects of new non-synonymous mutations suggests stronger selection on
174 sperm genes in monarchs but not Carolina sphinx moths. The DFE is largely unchanged from the genome
175 background to whole sperm proteome to sperm homologs in the Carolina sphinx moth (Figure 3, left
176 panels). For monarchs, the sperm proteome shows a dearth of weakly deleterious and effectively
177 neutral variants, with a concurrent increase in strongly deleterious and beneficial variants. This pattern

178 is even more exaggerated in the sperm homologs, where almost no neutral variants are detectable
179 (Figure 3, right).

180 With polyDFE, we see a slightly higher α for the sphinx moth sperm proteome compared to the genome
181 background, but this pattern is not localized in the sperm homologs. Moreover, we see a much larger
182 difference in the way selection acts on sperm protein variants compared to the rest of the genome only
183 in monarch butterflies. Here, upwards of 90% of substitutions are inferred to be a result of adaptive
184 evolution in both the whole sperm proteome and the shared sperm orthologs (Supplemental Figure 1).
185 We note that estimates of α are influenced by the ways in which demography are (or are not) accounted
186 for, so the values obtained with this more complex likelihood method differ from our estimates based
187 solely on count-data. Likely, the point-estimates here are closer to the true proportions of adaptive
188 substitutions for these species, but we are more interested in relative patterns between classes of genes
189 than accurately estimating the true, absolute value of α per se.

190 *Molecular evolution in dimorphic sperm*

191 The two sperm proteomic datasets consisted of three classes of sperm proteins: unique to eupyrene
192 sperm, unique to apyrene sperm, or found in both cell types (henceforth “shared”). We assessed
193 differences in selective pressures between the sperm morphs with another series of permutation tests,
194 both comparing parts of the sperm proteome to the genome background and comparing parts of the
195 proteome to each other.

196 As expected based on the whole-proteome results, neither eupyrene-specific ($p = 0.55912$), shared ($p =$
197 0.4647), nor apyrene-specific proteins ($p = 0.96496$) differed from the genome background in the
198 Carolina sphinx (Figure 4). In monarchs, both eupyrene-specific proteins ($p = 0.00018$) and shared
199 proteins ($p = 0.01038$) showed elevated α , but apyrene-specific proteins did not evolve differently from
200 the rest of the genome ($p = 0.55934$).

201 In *Manduca sexta*, α did not vary between apyrene-specific and eupyrene-specific proteins ($p = 0.7271$),
202 between apyrene-specific and shared ($p = 0.7176$) or eupyrene-specific and shared proteins ($p = 0.9979$).
203 Similarly, neither apyrene nor eupyrene sperm differed significantly from the shared set in monarchs (p
204 $= 0.6332$ & $p = 0.6234$, respectively). There was, however, a trend for increased α in eupyrene-specific
205 proteins compared to apyrene-specific proteins ($p = 0.0986$). In these analyses, we did not investigate
206 the role of orthology due to a loss of statistical power that would result from further subdividing our
207 datasets.

208 *Patterns of Adaptive Evolution in Sex-Specific Tissues*

209 Using RNA-seq data for a number of tissues in both monarchs and *Manduca sexta*, we calculated the
210 tissue specificity metric, SPM (Kryuchkova-Mostacci and Robinson-Rechavi 2017), of every gene in the
211 genome assembly with a sliding cutoff. Owing to the non-independence of estimates at each point, *e.g.*
212 all genes that pass a strict threshold of SPM > 0.9 are represented in every estimate at lower thresholds,
213 these data were not significance-tested. Nevertheless, the results from both species' sperm proteomes
214 hold at all SPM thresholds. In monarchs, sperm genes show consistently higher α than the whole
215 genome (Figure 5A). For the Carolina sphinx moth, sperm genes do not diverge from the whole genome,
216 except at very high specificity thresholds, where sperm genes show lower α (Figure 5B). Finally, we note
217 a general trend for increasing rates of adaptive evolution in more tissue-specific genes in monarchs but
218 not in the Carolina sphinx moth. This result mirrors the inferred DFEs; there is little evidence for
219 adaptive evolution, as indicated by positively selected variants, in the Carolina sphinx moth genome
220 background.

221 *Demographic estimates*

222 Finally, using site frequency from 4-fold degenerate sites in the two species' genomes, we estimated
223 population size history (Figure S2). Both have effective population sizes near 2,000,000, as expected of
224 herbivorous invertebrates with high dispersal potential, numerous host plants, and a large range over
225 North America. We also recover a population size increase in monarch butterflies in the recent past,
226 which has been previously reported with genomic data (Zhan et al. 2014). We note that our inferred
227 timing of this event differs from that of the previous authors, who used mutation rate estimates from
228 *Drosophila melanogaster*. Such parameter differences affect the estimated time of events, but not the
229 trajectories.

230 **Discussion**

231 We found elevated adaptive evolution in the sperm proteome of monarch butterflies compared to the
232 genome background, but not in *Manduca sexta* (Figure 1). In particular, this difference is greatest for
233 sperm genes with a sperm homolog in *M. sexta* (Figure 2B), suggesting that the same genes experience
234 stronger selection in the polyandrous species. Intriguingly, the source of this signal was not increased
235 divergence, but a reduction in non-synonymous polymorphism in sperm proteins (Figure 1B, top left).
236 This pattern suggests an increased role of purifying selection on sperm protein variants in monarchs.
237 These results echo the work of morphological reproductive biologists, who have shown that bird species

238 with higher rates of sperm competition show less intraspecific and intra-male variation in sperm length
239 (Immler et al. 2008; Kleven et al. 2008). On the sequence level, similarly increased purifying selection
240 has been observed in genes expressed in pollen, the main male-male competitors in flowering plants
241 (Arunkumar et al. 2013). Our results are, to our knowledge, the first indication that this pattern of
242 intensifying selection and decreasing intraspecific variation extends to the molecular level in sperm
243 specifically.

244 That stronger purifying selection results in increased adaptation may seem counterintuitive, but greater
245 selective scrutiny of within-population variation necessarily changes the patterns of fixation of alleles.
246 Stronger positive selection obviously increases the proportion of adaptive divergences, but stronger
247 purifying selection has a similar, albeit indirect effect. With greater purifying selection, fewer weakly
248 deleterious alleles can fix by drift, meaning that the alleles that do reach fixation and contribute to
249 divergence are more likely to be positively selected. Thus, theoretically, reproductive proteins
250 experiencing stronger selection should not have elevated rates of divergence (Dapper and Wade 2016).

251 This reasoning runs contrary to the conventional wisdom of reproductive protein evolution, but as
252 stated above, while observations of elevated divergence of reproductive proteins are common (Civetta
253 and Singh 1995; Willie J. Swanson and Vacquier 2002; Dorus et al. 2004; Martin and Hosken 2004),
254 evidence directly linking this pattern to adaptive processes is much more limited. In one recent
255 exception, a role for sperm competition in adaptive evolution was suggested for wrasses, but this study
256 assessed genes with male-biased expression rather than sperm genes directly and did not account for
257 demography in modeling evolution (Dean et al. 2017). Furthermore, interpretation of results in
258 spawning fish is complicated by the increased importance of abiotic selective pressures on sperm
259 associated with external fertilization, as observed by Liao *et al.* (2018). We directly assessed the
260 evolution of sperm proteins in two internally fertilizing species, avoiding such confounding
261 environmental variables.

262 *Targets of selection*

263 Without better functional annotation of genes in either genome, it is difficult to identify the exact roles
264 of the proteins under stronger selection in monarchs, but the signal from sperm homologs suggests that
265 these genes have had conserved sperm function since the divergence of the two species some 100
266 million years ago (Heikkila et al. 2012). Such genes relate to core traits in sperm, according to recent
267 gene ontology analyses (Whittington et al., in submission). In a similar vein, both proteins shared in the

268 two sperm types and those unique to fertilizing eupyrene sperm show an elevated α compared to the
269 genome background (Figure 4). Shared sperm proteins are enriched for structural proteins that give rise
270 to the sperm tail and thus impact motility (Whittington et al., in submission), while those expressed only
271 in eupyrene sperm doubtless include important mediators of fertilization. On the morphological scale,
272 variation in universal sperm traits like swimming ability, longevity, and overall viability are known to
273 affect sperm competition outcomes (Burness et al. 2004; Kim et al. 2017) and have been shown to have
274 a polygenic basis in other taxa (Hering et al. 2014). For traits like longevity and motility there is a
275 threshold below which fertilization becomes significantly impaired, but in the absence of competitor
276 alleles, there is likely a larger range of effectively-neutral trait-values, allowing for more variation to be
277 maintained in the population. In the presence of competitor alleles, however, marginal differences in
278 fertilization success come under selection, leading to the removal of deleterious variants.

279 Indeed, while the exact mechanisms of fertilization in Lepidoptera remain unclear, eggs are known to
280 possess multiple micropyle openings for sperm (Kumar et al. 2007) and eupyrene sperm possess
281 structures resembling an acrosome while their apyrene counterparts do not (Wolf 1992). This somewhat
282 rare combination of male and female gamete structures is also found in sturgeon, in which the multiple
283 micropyles give several sperm potential access to the egg nucleus and there is competition among
284 sperm to initiate karyogamy via the acrosome reaction (Psenicka et al. 2010). If a similar dynamic exists
285 in Lepidoptera, acrosomal proteins in eupyrene sperm would be likely targets for selection in
286 polyandrous systems.

287 Whatever the mechanics of fertilization are, paternity outcomes in polyandrous species are routinely
288 observed to be strongly bimodal (Simmons and Siva-Jothy 1998; Wedell and Cook 1998), including in
289 monarch butterflies (Mongue et al. 2015). In other words, one of the two males in a double mating
290 typically fathers most, if not all, of the observed offspring produced by the female, but there is little
291 consistency in whether it is the first or second male. With these dynamics, fitness differences between
292 winning and losing sperm phenotypes are large and selection can reliably remove less successful
293 genotypes.

294 Evidence of this can be seen in the estimated distribution of fitness effects of new mutations in monarch
295 sperm proteins. Compared to the genome background, we see a decrease in the proportion of
296 effectively neutral and weakly deleterious mutations ($-10 \leq s \leq 0$) and an increase in both strongly
297 deleterious ($s \leq -10$) and beneficial ($s > 0$) mutations (Figure 3, right). The increase in apparently
298 beneficial mutations follows the logic above. In the absence of competition, not only are mildly

299 suboptimal variants effectively neutral, but novel beneficial variants have no point of comparison for
300 fertilization efficiency. Thus, more efficient sperm variants should have no selective advantage in
301 monandrous species unless they markedly increase fitness in a single mating. This reasoning is
302 supported by the estimated distribution of fitness effect for the complimentary gene sets in the Carolina
303 sphinx moth; in this species, we see little variation in the DFE between the genome background and the
304 sperm proteome (Figure 3, left).

305 *Sperm dimorphism*

306 We failed to detect a strong difference between eupyrene (fertilizing) and apyrene (non-fertilizing)
307 sperm in either species (Figure 4). Moreover, apyrene sperm did not show a distinct pattern of evolution
308 compared to the genome background. If these sperm were truly involved in interfering with competitors'
309 sperm, as some have suggested (Silberglied et al. 1984; Swallow and Wilkinson 2002; Solensky and
310 Oberhauser 2009), we would expect evidence for stronger selection in apyrene sperm of monarch
311 butterflies; if anything, however, there was a trend for more adaptive evolution in eupyrene-specific and
312 shared-sperm proteins (Figure 4, bottom), likely due to fertilization dynamics coupled with sperm
313 competition as discussed above. Moreover, there appears to be proportionally less detectable orthology
314 in apyrene-specific than eupyrene-specific proteins in both species (Whittington et al., in submission),
315 suggesting that the apyrene sperm proteome is less conserved and potentially less selectively
316 constrained over evolutionary time.

317 These results suggest a few possibilities for apyrene sperm. Firstly, it is possible that the function(s) of
318 apyrene sperm is governed by a small subset of apyrene-specific proteins. Because our methods
319 aggregate signal for selection across multiple genes or sites to counteract high variance in variant counts
320 within genes (Stoletzki and Eyre-Walker 2011), the importance of these few genes could be lost in the
321 heterogeneous selection on different proteins. Still, the observation that few or no apyrene specific
322 genes show mating-system-dependent evolution argues against an *active* role for apyrene sperm in
323 sperm competition, but apyrene sperm may still have adaptive significance without specialized
324 molecular function.

325 The filler hypothesis also relates to sperm competition, but posits that apyrene sperm are employed
326 proactively, to fill the female's sperm storage organ and delay remating, thus decreasing the risk of
327 sperm competition rather than impacting its outcome (Swallow and Wilkinson 2002). Indeed, both in
328 monarchs and another butterfly (*Pieris napi*), female time to remating increases with the number of

329 apyrene sperm received from males (Oberhauser 1988; Cook and Wedell 1999). Such observations are
330 somewhat confounded by the size of the spermatophore nuptial gift that males provide during mating,
331 but apyrene sperm themselves have been proposed as a form of nutritional nuptial gift, so these two
332 hypotheses are not mutually exclusive (He et al. 1995; Lamunyon 2000). Under both the nutrient and
333 filler hypothesis, the actual protein content of apyrene sperm should be less important than its physical
334 presence and abundance, so factors affecting the rate of apyrene sperm production would be more
335 likely targets for selection in polyandrous species than the proteins sequences themselves.

336 Finally, we consider the most intriguing hypothesis for apyrene sperm function: capacitation of
337 fertilization. This hypothesis is based on the observation that apyrene sperm appear to be necessary for
338 successful fertilization in *Bombyx mori* (Takemura et al. 2006); the mechanism here is unclear, but could
339 conceivably involve one or a few genes that interact with the female reproductive tract to make
340 conditions more favorable for eupyrene sperm. In such a case, these proteins would behave more akin
341 to the broader class of reproductive proteins with sex-limited expression and evolve independently of
342 rates of polyandry in a species. Moreover, the experiment demonstrating the capacitation role in
343 *Bombyx* used apyrene sperm from a different male to restore fertilization success to the experimental
344 male (Sahara and Takemura 2003; Takemura et al. 2006), so whatever the method of capacitation is, it
345 lacks the allorecognition to be involved in offensive sperm competition.

346 *Evolution of genes with sex-limited expression*

347 One corollary to the model of reproductive protein evolution proposed by Dapper and Wade is the
348 prediction that genes with sex-limited expression can diverge more quickly under relaxed selection due
349 to the smaller effective population size of males or females compared to the population as a whole
350 (2016). As discussed above, we recovered their predictions for variable selection on sperm proteins but
351 did not observe a strong pattern of difference in the evolution of genes with testes-specific expression,
352 our proxy for sex-limited expression (Figure 5). This pattern was not formally tested, but we did not see
353 relaxed selection for the apyrene-specific set of genes either, which, as discussed above, may behave
354 more like male-limited genes than sperm-competition genes.

355 Nearly Neutral Theory may provide an explanation for these discrepancies. As mentioned above, larger
356 populations have more efficient selection and a smaller range of slightly deleterious mutations that
357 behave neutrally (Ohta 1992). Formally, mutations with a selective effect less than $1/N_e$ are expected to
358 behave neutrally. For instance, one commonly cited estimate for human population size is $N_e \approx 10,000$

359 over our species' evolutionary history (Zhao et al. 2000). Based on this, mutations with selective effects
360 less than 0.0001 should behave neutrally for alleles expressed in both sexes, while those with effects of
361 0.0002 are effectively neutral for alleles only expressed in one sex. And indeed, there is evidence that
362 genes expressed only in men have a higher mutational load than those expressed in both sexes
363 (Gershoni and Pietrokovski 2014). Chimpanzees, another species with a similar effective population size
364 (Won and Hey 2005), also show increased non-synonymous divergence in reproductive proteins (Wong
365 2010). Moreover, male reproductive protein evolution appears to depend more on effective population
366 sizes than intensity of sperm competition in the great apes in general (Good et al. 2013).

367 Consider, however, that the effective population sizes of insects are orders of magnitude higher than
368 those of most mammals. Using neutral site frequency spectra, we estimate effective populations near
369 2,000,000 for both North American monarchs and Carolina sphinx moths (Figure S2). Consequently,
370 selection is much more effective in these populations; mutations with effects above 5×10^{-7} should be
371 subject to selection in both sexes and those with effects on the order of 1×10^{-6} under selection if
372 expression is sex-limited. In other words, even selection on alleles with sex-limited expression in these
373 insects should be 100 times stronger than selection on the entire human population. There could indeed
374 be a two-fold difference in selection, but the absolute magnitude of the difference would be miniscule,
375 and the effects of mating system would be more apparent.

376 Conclusions

377 Established literature on sperm morphology has long recognized the effects of sperm competition on
378 variation in sperm traits (Immler et al. 2008; Kleven et al. 2008). Whether this pattern extends to the
379 nucleotide level has been unclear because research on molecular evolution in sperm proteins has
380 focused less on non-adaptive variation and more on explaining positive selection. Our investigation of
381 the sperm proteome in two Lepidoptera sits at the intersection of these fields and provides important
382 insights for the influence of post-copulatory selection in molecular evolution of reproductive proteins. In
383 a polyandrous species, sperm genes experience a strikingly different selective environment than the rest
384 of the genome. When considering genes with a sperm homolog in both species, we see weaker selection
385 in the monandrous species, consistent with selection through sperm competition. The results on genetic
386 variation are visible with both straightforward count-based and sophisticated model-based population
387 genetic analyses. The evolution of dimorphic sperm, however, does not show such strong differences,
388 but patterns of increased selection in fertilizing-sperm-specific proteins are consistent with sperm
389 competition as a driver of selection.

390 Our findings alone do not confirm the function of apyrene sperm in Lepidoptera, but they do
391 contextualize established hypotheses. We suggest that apyrene sperm are unlikely to be active agents of
392 sperm competition, but rather, may play a passive role in reducing the risk of competition. The method
393 by which apyrene sperm capacitate fertilization in some species remains unclear based solely on
394 genomic approaches and will likely require functional experiments to completely understand.

395 **Materials and Methods**

396 *Sources of data*

397 We used gene sets from the published genomes of each species (Zhan et al. 2011; Kanost et al. 2016)
398 with sperm genes identified from their respective proteomes (Whittington et al. 2015; Whittington et al.
399 2017). We inferred selection from patterns of polymorphism and divergence from congeners using
400 whole genome Illumina resequencing data for both species: a previously published dataset for North
401 American monarch butterflies (Zhan et al. 2014) and a new dataset of North Carolinian sphinx moths.
402 Focal moths were collected with a mercury vapor light trap in July of 2017 in Rocky Mount, North
403 Carolina (see supplemental table S1 for sequencing summary statistics). Divergences were called by
404 comparison to the queen butterfly (*Danaus gilippus*, previously published) for monarchs, and the five-
405 spotted hawkmoth (*Manduca quinquemaculata*, sequenced for this project) for the Carolina sphinx
406 moth.

407 In both focal species, we used twelve wild-caught individuals for sampling of polymorphism. In the case
408 of Carolina sphinx moths, these were twelve males caught over the course of three nights. The sex-
409 biased sampling reflects a sex bias in dispersal and collection at the light trap. In the case of monarchs,
410 samples were selected based on depth of sequencing coverage in the published dataset and included 8
411 females and 4 males from the panmictic North American migratory population. These samples added
412 the complication of unequal sampling between the autosomes ($n = 24$) and Z sex chromosome ($n = 16$).
413 Despite the Z's enrichment in the sperm proteome however, the vast majority of sperm genes (92% in
414 the Carolina sphinx, 90% in the monarch) are autosomally linked in both species (Mongue and Walters
415 2017). Due to the lowered power for statistical testing and limited inference to be gained from Z-linked
416 genes, we focused on the autosomal genes in both species in subsequent analyses.

417 *SNP-based methods*

418 After quality-trimming, we aligned sequenced reads with bowtie2 for conspecifics to their reference
419 genome (Langmead and Salzberg 2012) or stampy with an increased allowance for substitution for
420 heterospecific alignments (Lunter and Goodson 2011). Alignments were taken through GATK's best
421 practices pipeline (McKenna et al. 2010), including hard filtering, to yield a set of high quality SNPs both
422 within and between species. Effect-class of each polymorphism and divergence (synonymous, non-
423 synonymous, intergenic, etc.) was determined using custom databases for the two species created with
424 SnpEff (Cingolani et al. 2012). Annotated SNPs were curated to remove false divergences (ancestral
425 polymorphism) and then differences in adaptive evolution were calculated using an unbiased estimator
426 of the neutrality index to calculate α , the proportion of substitutions driven by adaptive evolution
427 (Stoletzki and Eyre-Walker 2011); this form of α corrects the inherent bias in a ratio of ratios while also
428 allowing summation across multiple genes to reduce noise associated with small numbers in count data.
429 For any set of i genes with non-zero counts of synonymous polymorphism and divergence:

$$430 \quad \alpha = 1 - \frac{\sum(Dsi * Pni)/(Psi + Dsi)}{\sum(Dni * Psi)/(Psi + Dsi)}$$

431 This statistic was calculated with custom R scripts in R version 3.3.3 (R Core Team 2017).

432 *Assessment of adaptive evolution and statistical significance*

433 In each analysis we calculated α for a biologically meaningful set of genes, e.g. the sperm proteome and
434 the genome background, and generated a test statistic from the absolute difference of the two point-
435 estimates. To determine significance, we combined the two sets and randomly assigned genes into two
436 new sets of sizes equal to the originals. The difference of these two datasets was determined and the
437 process was repeated for 50,000 permutations to build a distribution of differences between the point
438 estimates of two gene sets of these relative sizes. The p-value was taken as the proportion of times a
439 greater absolute difference was observed between the two random data sets than the original sets.

440 These analyses were applied within-species at several levels: differences between the sperm proteome
441 and genome background, differences between sperm homologs and unique proteins in the proteome,
442 and, finally, differences between the two sperm morphs. The whole proteome comparison is relatively
443 straightforward. For the sperm homolog to novel proteins comparison, we considered orthology in the
444 same manner as we did in a previous investigation of genomic architecture in these two species
445 (Mongue and Walters 2017). Sperm proteins were divided into one of two classes based on orthology
446 and expression: either sperm homolog or unique if the ortholog was not found in the sperm proteome

447 or there was no detectable ortholog. In these analyses, the sperm proteome was taken as a whole,
448 agnostic of sperm dimorphism. To examine differences in dimorphic sperm, we finally considered the
449 gene product's location in the proteome, either unique to eupyrene sperm, unique to apyrene sperm, or
450 shared in both types. For these analyses, we did not consider orthology status owing to the reduction in
451 power that would accompany multiple layers of subdivision of the dataset.

452 *Site-frequency-based methods*

453 We also investigated molecular evolution by leveraging site-frequency-spectrum-based approaches as
454 complimentary evidence. We used the population genetics software suite ANGSD (Korneliussen et al.
455 2014) to generate site frequency spectra at putatively neutral (four-fold degenerate) and selected (zero-
456 fold-degenerate) sites in the genome. We unfolded site frequency spectra using parsimonious inference
457 of ancestral state of alleles. These unfolded spectra were fed into the sfs-based tool polyDFE (Tataru et
458 al. 2017) to examine rates of adaptive evolution with a more complex likelihood model that corrects for
459 effects of demography and misattribution of ancestral state. We compared sites from the genome
460 backgrounds to sites from the sperm proteomes to see if estimates of α or the distribution of fitness
461 effects of new mutations differed between these two gene sets in each species. Divergence counts were
462 omitted here to simplify the likelihood computation for these large datasets and remove any error for
463 misattributed divergence. To test the robustness of results, input data were bootstrapped 100 times to
464 obtain confidence intervals for parameter estimates. Processing of model inputs and outputs was
465 accomplished with custom R scripts.

466 *Investigation of sex-limited and tissue-specific expression*

467 Next, we investigated the robustness of results from the sperm proteome analyses using RNA-seq data
468 in these taxa. For *Manduca sexta*, there exists a wealth of tissue-specific data at multiple developmental
469 timepoints (Cao and Jiang 2017). Because we were primarily interested in sperm involvement, we
470 focused on sequencing from adult males, specifically RNA from the testes, head, thorax, and gut.
471 Expression (as measured by FPKM) was averaged across biological replicates where available in this
472 species. Monarchs had no comparable published data, so we sequenced the head, thorax, gut, testes,
473 and accessory gland of three adult males.

474 In order to localize the signal for specific expression, we calculated tissue-specificity as SPM (specificity
475 metric), a ratio ranging from 0 to 1 on the proportion of gene expression limited to a focal tissue
476 (Kryuchkova-Mostacci and Robinson-Rechavi 2017). For instance, a gene with an SPM value of 0.8 for

477 the testes shows 80% of its total expression in the sampled tissues in the testes. Rather than strictly
478 defining a single threshold for tissue-specific expression compared to general expression, we
479 implemented a sliding cut-off. For a series of SPM thresholds ranging from 0 to 1 in increments of 0.5, all
480 genes of a given class that showed specificity higher than the threshold were included in a calculation of
481 α . This methodology created substantial non-independence between point estimates and precluded
482 significance testing, but still allowed us to investigate whether results from the sperm proteome were
483 sensitive to filtering of the included genes.

484 We first compared the effect of SPM threshold value on all genes in the genome that had non-zero
485 expression in sampled tissues. These genes were evaluated based on the maximum SPM across all
486 tissues. For sperm proteins, we considered only genes identified in the sperm proteome and ranked
487 them by SPM in the testes. Finally, for male-limited non-sperm genes, we excluded sperm proteome
488 genes and considered again those ranked by specificity in the testes (or testes and accessory glands for
489 monarchs). These analyses were completed with custom R scripts as well.

490 *Demographic estimates*

491 Finally, to contextualize the previous analyses, we characterized present and historical population size
492 from genomic data. Using 4-fold degenerate frequency spectra, we estimated neutral coalescence
493 patterns with the program stairway plot (Liu and Fu 2015). For estimated generation time, we used the
494 widely cited four generations per year for monarchs and three for the Carolina sphinx moth. Finally, for
495 mutation rate, we chose the estimate 2.9×10^{-9} from the butterfly *Heliconius melpomene*, the closest
496 relative with a spontaneous mutation rate estimate (Keightley et al. 2015).

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697 [abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=17204&tool=pmcentrez&rendertype=abstract)

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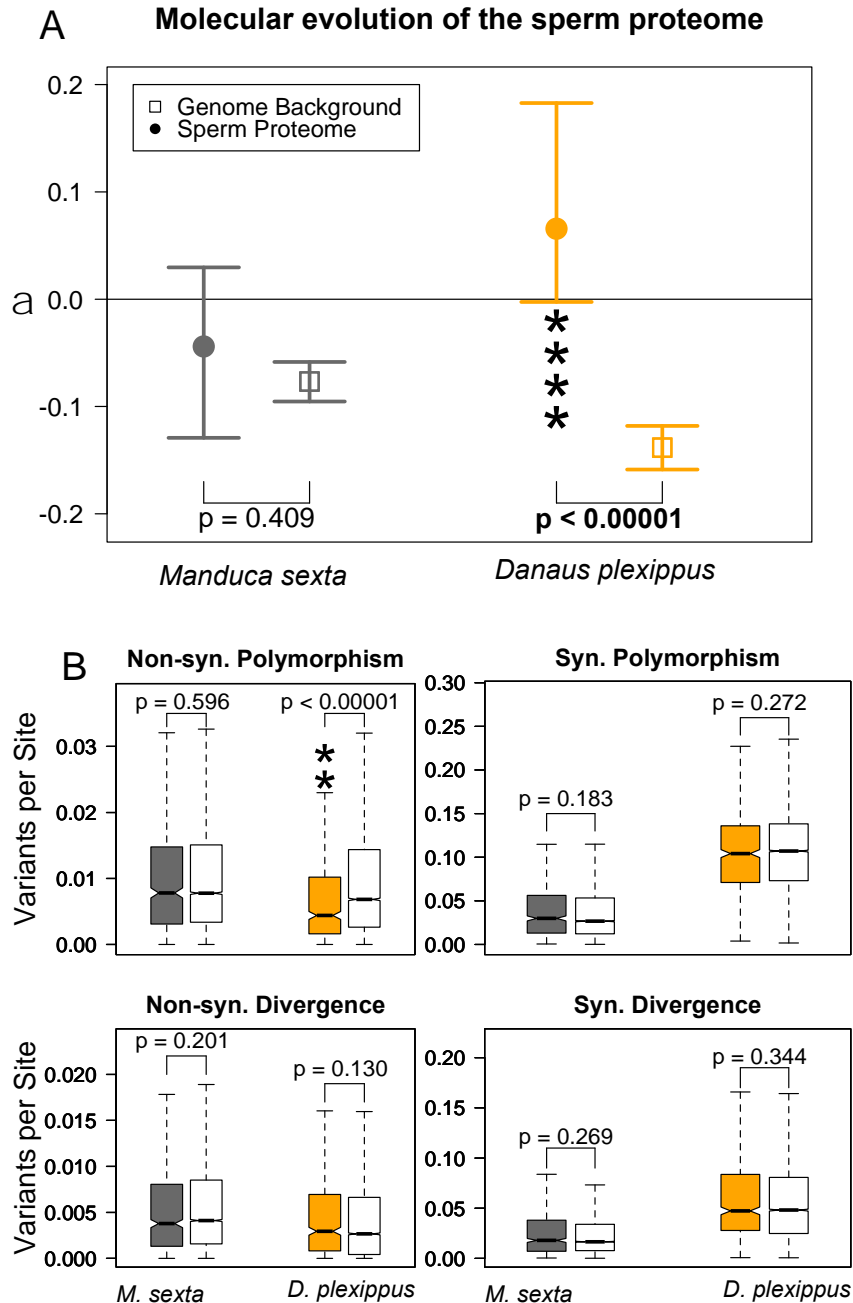
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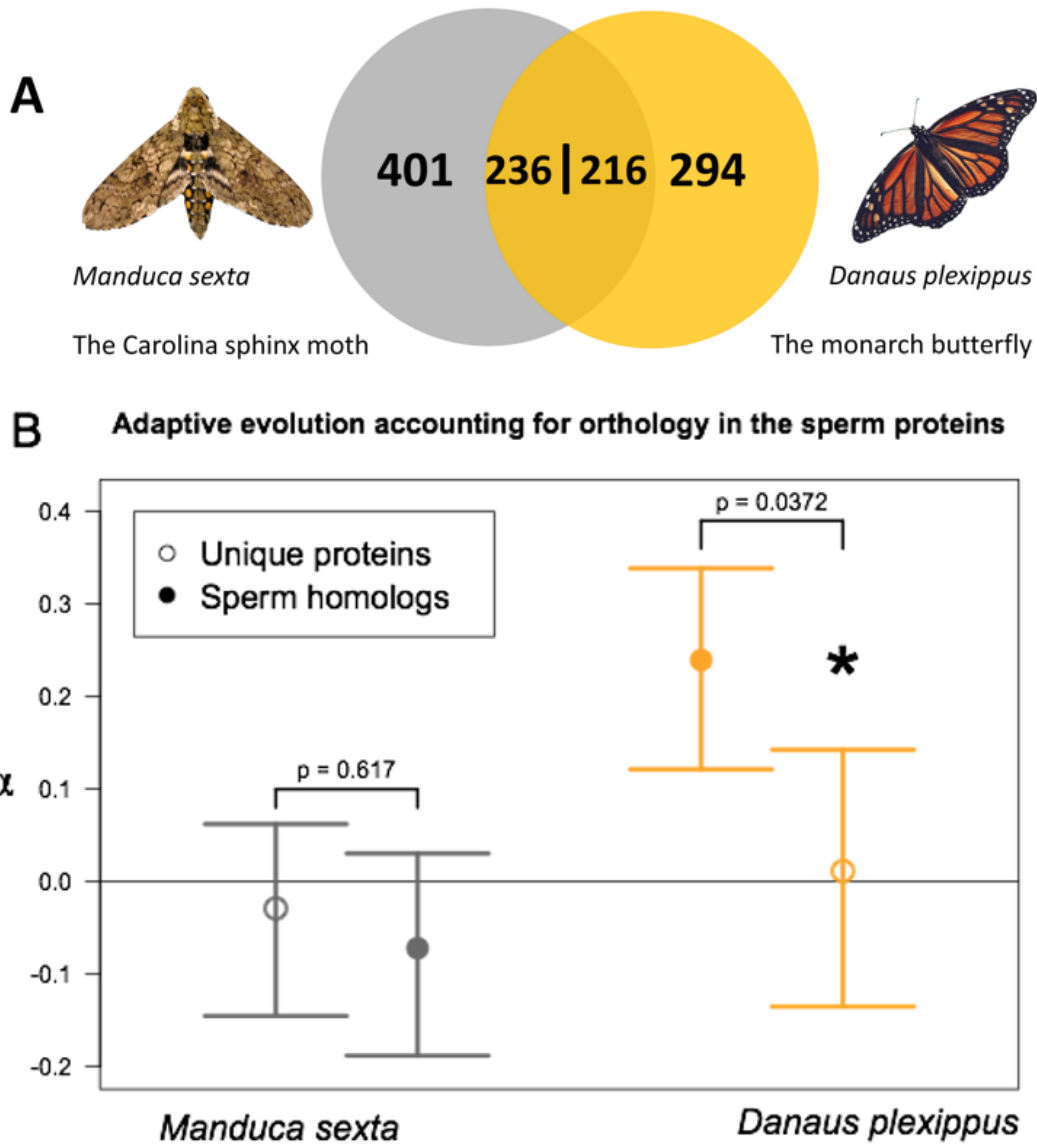


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718 **Figure 1. A.** Genes in the sperm proteome of monarch butterflies (*D. plexippus*) show a significantly
 719 higher proportion of adaptive substitutions (α) than the rest of the genome (right); compare to the
 720 Carolina sphinx moth (*M. sexta*) in which there is no difference between the sperm proteome and the
 721 rest of the genome. P-values come from permutation tests. Error bars represent 95% bootstrapped
 722 confidence intervals from the point estimates. **B.** Decomposing α into its components: Pn, Ps, Pn, and Ds
 723 and comparing the sperm proteome (filled boxes) to the genome background (open boxes). There were
 724 no strong differences between sperm genes and the genome background in Carolina sphinx moths. In
 725 monarch butterflies, the signal for increased adaptive substitution comes from a marginal increase in
 726 non-synonymous divergence (bottom left) combined with a great reduction in non-synonymous
 727 polymorphism in sperm genes compared to the rest of the genome (top left). P-values are derived from
 728 Wilcoxon-Mann-Whitney tests.

Sperm Proteome Overlap

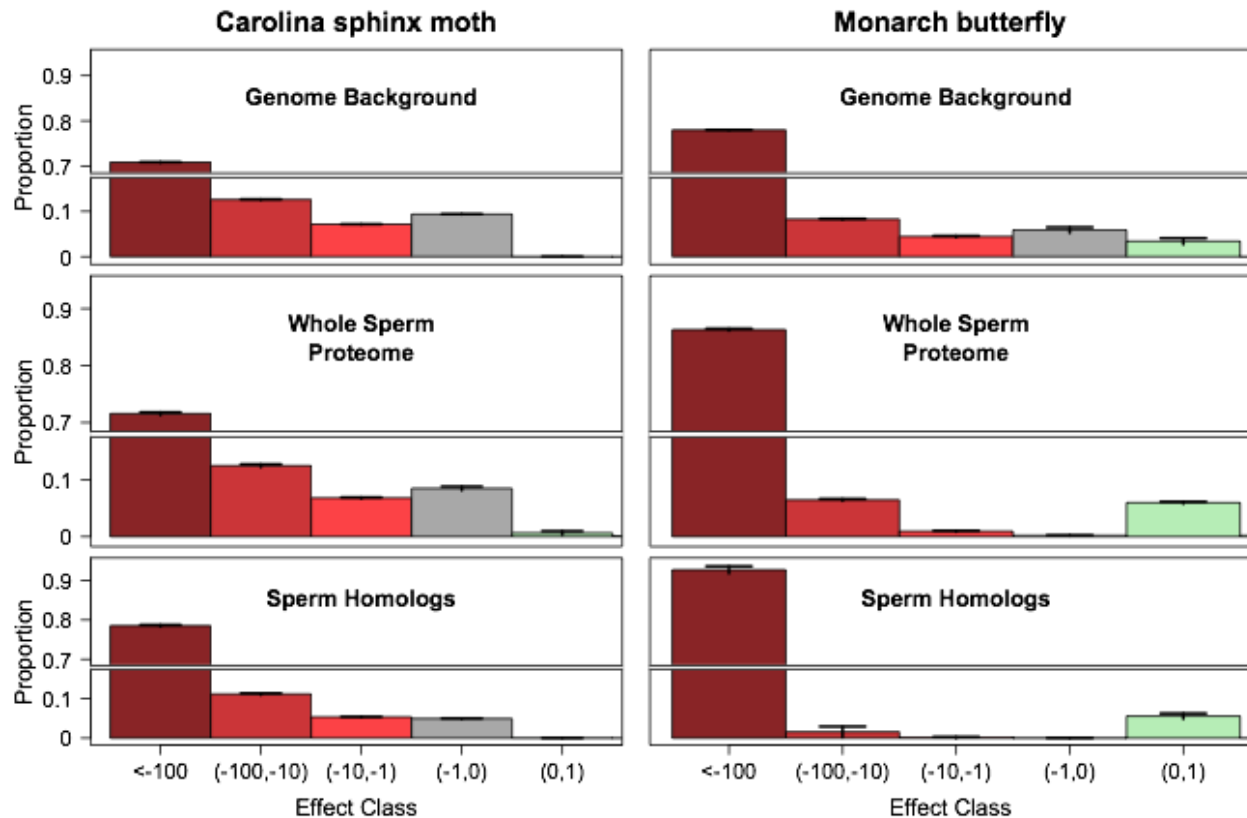


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730 **Figure 2. A.** Composition of the portion of the sperm proteomes analyzed in this study. Numbers
731 indicate counts of proteins unique to one species' sperm or with an ortholog in the other species' sperm
732 (sperm homologs). Note that the overlap number varies between species due to the presence of a few
733 one-to-many-orthologs. **B.** Sperm homologs show evidence for a greater proportion of adaptive
734 substitutions (α) in monarch butterflies, but not in Carolina sphinx moths. P-values are based on
735 permutation tests comparing the difference between two sets of genes randomly assigned from the
736 sperm proteome in each species; error bars are 95% bootstrap confidence intervals.

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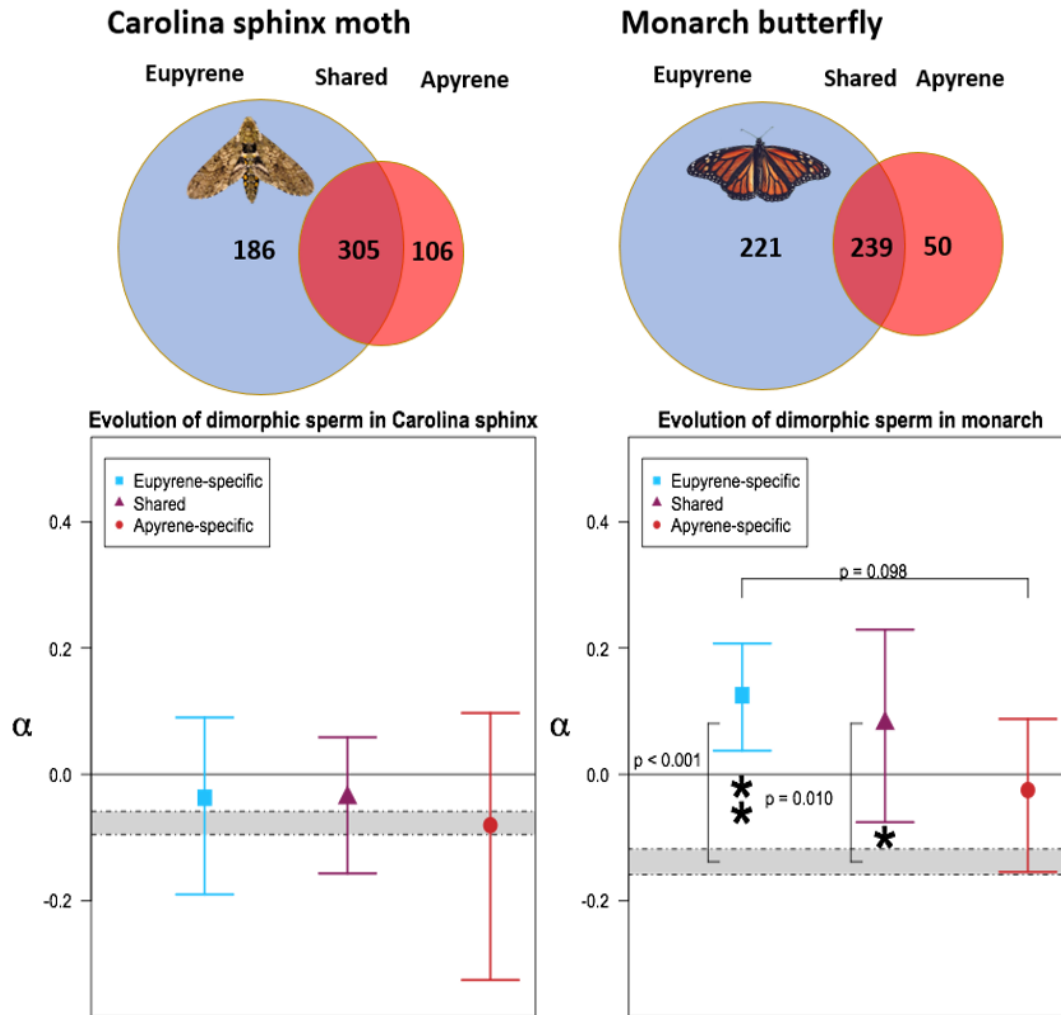
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741 **Figure 3.** Predicted distribution of fitness effects of new non-synonymous mutations for the gene sets
 742 investigated in Figures 1 and 2. From top to bottom: autosomal genome background, autosomal sperm
 743 proteome, and the subset of sperm homologs found in both species. Bars represent the mean
 744 proportion for each selective class, with error bars representing twice the standard error of the mean
 745 from 100 bootstrap replicates of the input data. Note the gap in the y-axis due to the preponderance of
 746 strongly deleterious ($s < -100$) mutations. **Left.** The DFE shows little variation between the background
 747 and the sperm data sets, barring a slight increase in the proportion of strongly deleterious mutations.
 748 **Right.** In monarch butterflies, note the increasingly bimodal distribution of fitness effects that coincides
 749 with increased selection inferred from earlier analyses. In the sperm proteome (middle), there is a
 750 decrease in effectively neutral (gray, $-1 < s < 0$) and weakly deleterious (light red, $-100 < s < -1$) variants,
 751 with a concomitant increase in both strongly deleterious (dark red, $s < -100$) and beneficial (green, $0 < s$
 752 < 1) variants. In sperm homologs this effect is even more pronounced, with nearly all variants coming
 753 under selection.

Composition of proteins in dimorphic sperm

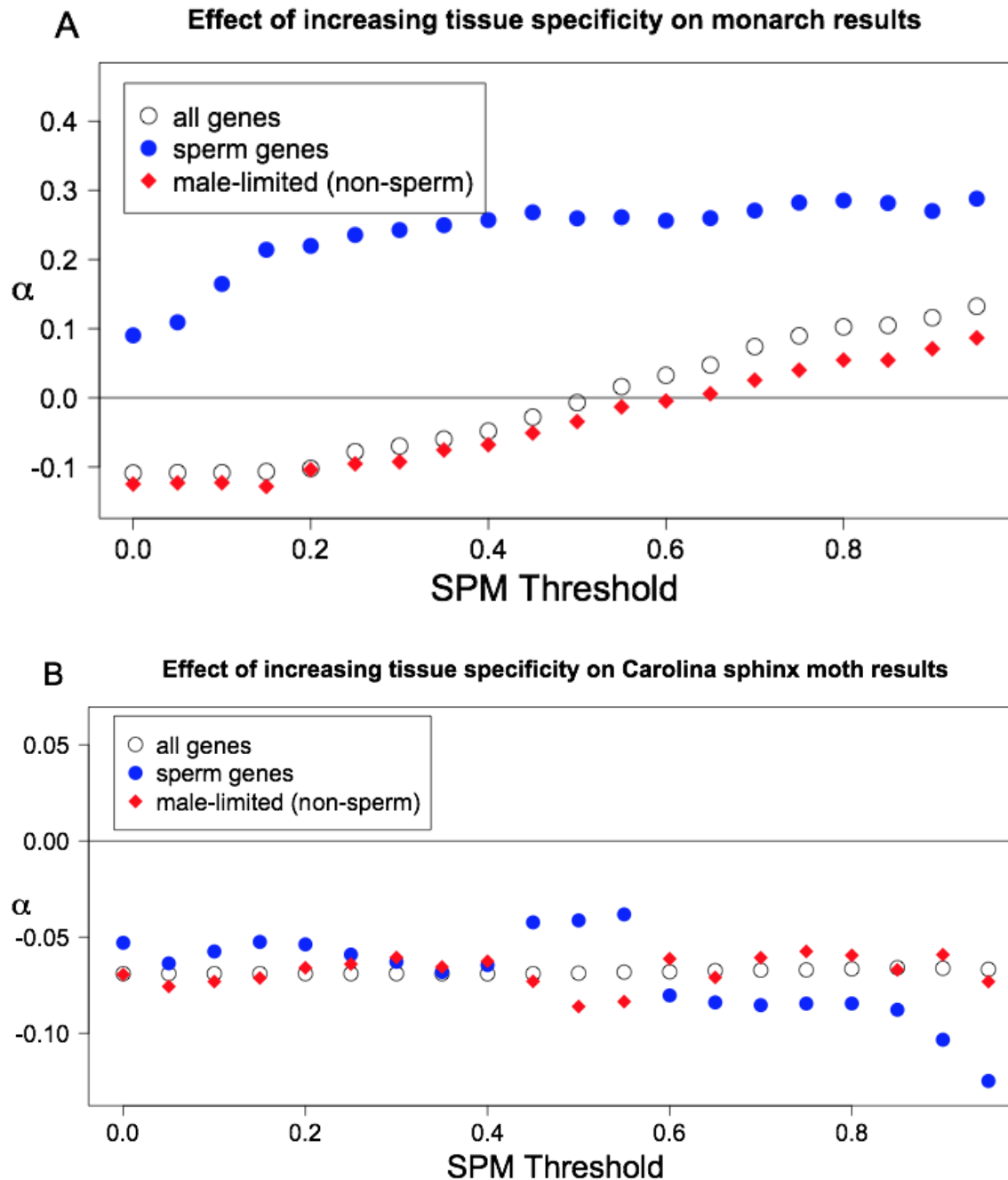


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755 **Figure 4. Top.** Composition of the sperm proteome with respect to dimorphic sperm. A majority of
756 identified proteins were shared between the two cell types, followed by the set unique to eupyrene
757 sperm, and finally the smallest set was the proteins found only in apyrene sperm. **Bottom.** None of the
758 sets of sperm proteins evolved either differently from each other or distinctly from the genome
759 background (shaded regions represent 95% confidence intervals of the genome background) in the
760 Carolina sphinx (left). In the monarch however (right), the signal for elevated α was localized to the
761 eupyrene-specific and shared proteins. There was also a trend for increased α in eupyrene-specific
762 proteins as compared to apyrene-specific. Error bars represent 95% confidence intervals from
763 bootstrapping.

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Figure 5. Estimated adaptive evolution as a function of specificity of gene expression, using a sliding cut-off for specificity (SPM). **A.** In monarchs, there is an overall increase in inferred α with increasing tissue specificity for all classes of genes. Adaptive evolution of the sperm proteome is higher than the genome as a whole at all cut-off thresholds. Interestingly, male-limited genes (as defined by high specificity of gonadal expression) appear to evolve similarly to somatic genes. **B.** Sperm proteins in the Carolina sphinx do not show differences from the genome for any but the strictest cut-off specificities, and even then, α slightly decreases compared to the rest of the genome. Genes with testes-limited expression again do not show differences from the genome background.