Reduced insulin/IGF-1 signalling in adult parents increases offspring fitness

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Summary

Reduced expression of the insulin/insulin-like nutrient-sensing signalling (IIS) pathway gene *daf-2* in adult *Caenorhabditis elegans* nematode worms increases longevity without affecting fecundity, but the effect of parental lifespan extension on adult offspring is largely unknown. Here we show that reduced IIS signalling in parental generation increases offspring fitness. We used RNA interference (RNAi) to silence *daf-2* expression in sexually mature *C. elegans* hermaphrodites from three different genotypes: N2 wildtype, as well as *ppw-1* and *rrf-1* mutants that are deficient for RNAi in germline and soma, respectively. Long-lived *daf-2* RNAi parents showed normal fecundity as self-fertilizing hermaphrodites and improved late-life reproduction when mated to males. Remarkably, the offspring of *daf-2* RNAi parents produced more progeny and had higher Darwinian fitness across all three genotypes. Thus, reduced IIS signalling in adulthood improves offspring quality supporting the emerging view that suboptimally high levels of nutrient-sensing signalling in late-life lie at the heart of ageing.

Introduction

Understanding mechanisms underpinning healthy ageing is fundamental to improving quality of life in an increasingly long-lived society. The long-standing paradigm postulates that energy trade-offs between reproduction and somatic maintenance underlie organismal ageing [1, 2]. However, the discoveries of environmental interventions that dramatically increase healthy lifespan in model organisms without the cost of reduced reproduction have challenged the current paradigm and revolutionized our understanding of the biology of ageing [3-8]. Specifically, experimental downregulation of nutrient-sensing insulin/IGF-like (IIS) signalling pathway that governs biosynthesis in response to nutrient availability can achieve increased longevity without a concomitant decrease in reproduction in model organisms [6, 9, 10].

Since cost-free lifespan extension contradicts the traditional view on how ageing evolves, several studies investigated the fitness consequences of reduced IIS signalling [11-15]. Indeed, mutations that reduce nutrient-sensing signalling, as well as environmental interventions aimed at mimicking the mutational effect, often have detrimental pleiotropic effects on key life-history traits, such as development, growth rate, body size and early-life reproduction resulting in reduced Darwinian fitness even if total reproduction is unaffected [11, 16]. The first longevity mutant discovered in *C. elegans, age-1*, is a good example because increased longevity, stress resistance and late-life reproduction come at a cost of reduced early-life reproduction and total individual fitness [15]. Moreover, a recent literature survey suggests that all classic longevity-extending mutations across taxa from worms to flies to mice detrimentally affect life-history traits resulting in reduced fitness [16]. Similarly, experimental evolution studies showed that when longevity and fecundity are increased simultaneously through selection, the organisms pay the price in slow development and delayed sexual maturation, again resulting in reduced individual fitness [17]. These results suggest that genes with antagonistically pleiotropic effects between early-life and late-life fitness play an important role in the evolution of ageing [18]. However, the mechanisms of antagonistic pleiotropy (AP) remain elusive. The leading hypothesis, the "disposable soma" theory of ageing (DS) suggests that ageing results from competitive energy allocation between somatic maintenance and reproduction [1, 2, 19].

However, this hypothesis suffered several setbacks in recent years, with many empirical studies challenging the importance of energy trade-offs in organismal senescence [reviewed in 4, 5, 8, 10, 20]. Instead, several authors proposed that ageing can result from molecular signalling networks being optimized for development, growth and early-life reproduction rather than for late-life reproduction and longevity [5, 9, 20-22]. For example, the hyperfunction theory maintains that ageing is driven by excessive nutrient-sensing molecular signalling in late-life, which results in cellular hypertrophy leading to age-related pathologies [21-23]. These ideas can be traced back to the original AP theory by George Williams, who suggested that the same physiological processes that are beneficial for fitness early in life can become detrimental for organismal fitness with age because of the reduced strength of natural selection on late-life function [18].

Because the main cost of longevity appears to be associated with reduced early-life function, it seems plausible that age-specific modification of gene expression can potentially circumvent this problem. In their landmark study, Dillin et al. (2002) used age-specific RNA interference (RNAi) approach to knock down *daf-2* gene expression in *C. elegans* nematodes across the life cycle of the worms. While early-life feeding with bacteria expressing *daf-2* double-stranded (ds) RNA resulted in reduced early-life reproduction, there was no detrimental effect of *daf-2* RNAi in adult worms, which enjoyed two-fold lifespan extension without any cost to reproduction [6]. This study provides the strongest support to date for the hypothesis that ageing results from molecular nutrient-sensing signalling that is optimized for early-life function but is suboptimal for late-life function.

Nevertheless, while this study provided a powerful example for the cost-free lifespan extension, it is possible that certain fitness costs have been overlooked. One possibility is that fecundity costs become apparent only in mated hermaphrodites. In nature, *C. elegans* live in populations with small (~0.3%) yet appreciable number of males living among self-fertilising hermaphrodites with sometimes high levels of outcrossing [24], and mating, as well as mere presence of male-derived pheromones, has pronounced effects of the life-history of hermaphrodites [25-27]. Perhaps more importantly, it is possible that while

fecundity is not affected, the fitness of the offspring and, therefore, Darwinian fitness of the parents are compromised. The trade-off between offspring number and quality is well known from a number of study systems [28], and is a potential explanation for the apparent lack of fitness costs in the previous studies. To investigate this possibility, we need to understand how late-life reduction in nutrient-sensing signalling affects longevity, offspring number and offspring quality. Here we show that *daf-2* RNAi in adult *C. elegans* results in increased offspring fitness across three genetic backgrounds. We discuss these findings in the light of the emerging new theories of ageing and suggest that they support the hypothesis that functional trade-offs between early-life fitness and late-life fitness shape the evolution of ageing.

Results

We confirmed that *daf-2* RNAi significantly extended the lifespan of unmated N2 wild-type hermaphrodite worms (censoring matricide: z = -4.94, df = 1, p <0.001, Fig. 1A; including matricide as dead: z = -4.97, df = 1, p <0.001), as expected from previous studies [6]. In addition, for mated N2, *daf-2* RNAi extended lifespan when matricide was censored (z = -2.42, df = 1, p = 0.016, Fig. 1B) but not if matricidal worms were included as dead (z = 0.16, df = 1, p = 0.87) because of an increase in matricide in the late reproducing mated *daf-2* RNAi N2.

We did not find any effect of *daf-2* RNAi on total reproduction (unmated: F = 0.32, df = 1, p = 0.58; mated: χ^2 = 1.11, df = 1, p = 0.29) or individual fitness λ_{ind} (unmated: F = 0.30, df = 1, p = 0.59; mated: χ^2 = 0.43, df = 1, p = 0.51) for neither unmated nor mated N2 (Table 1, Fig. 2). However, *daf-2* RNAi had a positive effect on late (day 5+) reproduction for mated hermaphrodites (χ^2 = 24.76, df = 1, p < 0.001, Fig. 2B).

In a second experiment, using unmated hermaphrodites only, we investigated the effect of *daf-2* RNAi on parent lifespan and offspring lifespan and reproduction across three genetic backgrounds (N2 wild-type and the mutants *ppw-1* and *rrf-1*, that are deficient for germline

and somatic RNAi, respectively). Parental treatment with *daf-2* RNAi increased lifespan across all genetic backgrounds, both when matricide was censored (treatment: $\chi^2 = 90.39$, df = 1, p <0.001; strain: $\chi^2 = 21.8$, df = 2, p <0.001; treatment × strain: $\chi^2 = 10.46$, df = 2, p = 0.005, Fig. 3A) and included as dead (treatment: $\chi^2 = 85.25$, df = 1, p <0.001; strain: $\chi^2 =$ 20.45, df = 2, p <0.001; treatment × strain: $\chi^2 = 9.43$, df = 2, p = 0.009). In accordance with previously published research [29], parental *daf-2* RNAi increased egg size (treatment: $\chi^2 =$ 5.11, df = 1, p =0.024; strain: $\chi^2 = 13.89$, df = 2, p <0.001; treatment × strain: $\chi^2 = 2.68$, df = 2, p = 0.262, Fig. 3B). However, we found that the effect was most pronounced in N2 wildtype worms, and relatively weak in both somatic and germline *daf-2* knockdown (see Fig. 3B), suggesting that daf-2 knockdown in both somatic and reproductive tissues is required to maximize the effect on egg size.

Parental *daf-2* RNAi treatment did not, however, influence the lifespan of their offspring, neither when matricidal worms were censored (treatment: $\chi^2 = 0.04$, df = 1, p = 0.85; strain: $\chi^2 = 24.2$, df = 2, p <0.001; treatment × strain: $\chi^2 = 0.61$, df = 2, p = 0.74, Fig. 4A) nor when included as dead (treatment: $\chi^2 = 0.01$, df = 1, p = 0.92; strain: $\chi^2 = 21.8$, df = 2, p <0.001; treatment × strain: $\chi^2 = 0.48$, df = 2, p = 0.79).

In contrast, parental *daf-2* RNAi treatment significantly increased offspring total reproduction (treatment: F = 15.9, df = 1, p <0.001; strain: F = 33.7, df = 2, p <0.001; treatment × strain: F = 0.09, df = 2, p = 0.91, Fig. 4B-C) and individual fitness λ_{ind} (treatment: F = 11.8, df = 1, p <0.001; strain: F = 13.1, df = 2, p <0.001; treatment × strain: F = 0.18, df = 2, p = 0.84, Fig. 4D) across all genetic backgrounds. Importantly, there was no correlation between the effect of parental *daf-2* RNAi on egg size (see above) and offspring total reproduction / individual fitness, suggesting that factors beyond the amount of resources in the egg contribute to increased fitness of offspring of *daf-2* RNAi parents.

Discussion

The "disposable soma" theory of ageing postulates that senescence results from competitive energy allocation between key life-history traits, such as growth, reproduction and somatic maintenance [1, 30]. This theory predicts that genetic and environmental manipulations that increase energy allocation to growth and somatic maintenance will result in detrimental effects to reproduction. This is why the findings by Dillin et al. (2002), which suggested that adult-only downregulation of insulin/IGF-1 by daf-2 RNAi can substantially increase lifespan without any detrimental effect to reproduction, were subsequently scrutinized in an attempt to find the hidden costs of longevity [7, 13]. Nonetheless, both the original findings [6] and our results here, suggest that adult-only daf-2 RNAi can more than double longevity without any negative effect on reproduction. Moreover, when supplied with sperm from males, *daf-2* RNAi-treated parents have improved fecundity in late-life. It is possible, however, that treatments that improve parental performance have negative effects on their offspring. The trade-off between offspring number and offspring quality is a well known concept in life-history evolution [28] but is rarely considered in biogerontological research [reviewed in 8]. Germline maintenance is costly [8, 31, 32], and increased investment into somatic maintenance can, in theory, result in increased mutation rate and reduced fitness of progeny.

Alternatively, it is possible that instead of energy trade-offs, the evolution of senescence is governed by functional trade-offs. Functional trade-offs can occur because the physiological requirements of a young organism can differ substantially from those of a mature one [18]. In his classic 1957 paper, George Williams [18] described a hypothetical example of a mutation that positively affects bone calcification in a developing young organism but increases calcification of the connective tissues of arteries in a mature one with detrimental consequences. More recently, it has been suggested that nutrient sensing IIS/TOR molecular signalling pathways that govern growth and development result in excessive biosynthesis in late-life leading to different pathologies and increased mortality [20-23]. These proximate explanations rest on the fundamental assumption that the strength of natural selection declines with age because of environmental mortality from a range of biotic and abiotic hazards (e.g. predation, pathogens, competition, starvation) [18]. Because of such

environmental mortality, immediate reproduction is more valuable than future reproduction, and optimizing development, growth and early-life reproduction is more important for organismal fitness that optimizing late-life survival and reproduction [18, 33]. Thus, while natural selection is predicted to modify nutrient-sensing signalling in an adult organism, the weak selection in late-life may result in suboptimal levels of IIS/TOR signalling leading to pathology and senescence [22]. Unlike the classic energy trade-off theory, the functional trade-off approach predicts that it should be possible to modify adult physiology to improve both longevity and fitness.

Here we found that reduced insulin/IGF-1 signalling in adult worms not only improved longevity and late-life reproduction, but also increased reproduction and Darwinian fitness of the resulting offspring in three different genetic backgrounds. This result contradicts the hypothesis that improved longevity and postponed ageing of *daf-2* RNAi parents comes at the cost of offspring fitness. Instead, our findings are in line with the hypothesis that suboptimal levels of nutrient-sensing signalling in adult life accelerate ageing, curtail lifespan and reduce individual fitness. This result was not caused by direct inheritance of *daf-2* RNAi, since we did not recover the lifespan extension effect of *daf-2* knockdown in these offspring.

Because previous research found that both dietary restriction and reduction in insulin-like signalling by *daf-2* RNAi knockdown increased embryo size in *C. elegans* nematodes [29], we replicated these results to test whether increased fitness of adult progeny results from increased resource allocation to eggs by *daf-2* RNAi mothers. While *daf-2* knockdown increased egg size to a different degree in N2, *ppw-1* and *rrf-1* strains, there was no correlation between the effect of parental *daf-2* RNAi on egg size and offspring reproductive performance. We provisionally conclude that increased egg size under reduced maternal insulin-like signalling can contribute to increased offspring fitness, but it is likely not the sole source of variation in this trait. Recent work has identified oocyte-specific IIS targets that are different from soma-specific IIS targets suggesting that IIS signalling regulates reproduction and longevity through different mechanisms [34]. In the future, it will be interesting to test

for individual fitness of offspring produced via genetic and pharmacological manipulation of oocyte-specific targets of IIS signalling pathway.

Methods

Strains

We used the *Caenorhabditis elegans* strains Bristol N2 wild-type (Brenner, Genetics 1974), as well as the mutants *ppw-1(pk2505)* (REF) and *rrf-1(pk1417)* (REF), obtained from Caenorhabditis Genetics Center (CGC, Missouri, USA).

Maintenance

Before each assay, worms were recovered from freezing and synchronised by bleaching for two generations to remove any freezing effects. The nematode populations were maintained at 20°C and 60% relative humidity in an environmental test chamber. For regular maintenance, the worms were kept on NGM agar supplemented with the antibiotics streptomycin, kanamycin and nystatin (following Lionaki & Tavernarakis [35]), seeded with the antibiotic-resistant *E. coli* strain OP50-1 (pUC4K).

Outline of the study

The study was run in three separate experiments. In the first experiment, we investigated lifespan and reproduction of mated (n=75 per treatment) and unmated (n = 25 per treatment) N2 hermaphrodites reared from sexual maturity onwards on *daf-2* RNAi or empty vector (EV, control) plates. For logistic reasons, this experiment was conducted in two blocks for mated worms and one block for unmated worms. In the second experiment, we investigated the lifespan and egg size of unmated N2, *rrf-1(pk1417)* and *ppw-1(pk2505)* hermaphrodites (n = 25 per treatment for lifespan, n = 60 for egg size) on raised from sexual maturity onwards on *daf-2* RNAi or EV plates. In a separate experiment, we collected one egg from each parent at their second day of adulthood (from *daf-2* RNAi and EV treatments)

and investigated the lifespan and reproduction of these offspring on control plates (n = 25 per parental treatment). Because different experiments differed in setup time, daily reproduction values (and calculations based upon these, such as λ_{ind}) are only meaningful for comparison between treatments within each experiment.

RNAi

RNase-III deficient, IPTG-inducible HT115 *Escherichia coli* bacteria with empty plasmid vector (L4440) was used as control [36] and the same HT115 bacteria with *daf-2* RNAi construct from the Vidal library was used as RNAi treatment. RNAi treatment started from sexual maturity, and continued until the death of the individual. During the experiments, worms were maintained on 35 mm NGM agar plates (supplemented with 1 mM IPTG and 50 μ g/ml ampicillin) seeded with 0.1 ml L4440 empty vector control or *daf-2* bacteria grown in LB supplemented with 50 μ g/ml ampicillin for 16-20 hours and seeded (incubated) on the NGM agar plates again for 24 hours (following Hinas et al. [37]).

Lifespan Assays

Lifespan assays were set up for all treatment combinations described above. In the lifespan assays, the individual age-synchronised L4 worms were placed on separate 35 mm plates and the plates were checked daily to record any instances of death. The surviving worms were moved to new plates daily until their death. Fertile worms, which showed odd developmental characteristics and low offspring numbers (<36 offspring), were excluded from the final analysis (3 mated control worms and 7 mated *daf-2* worms).

Reproduction assays

Offspring production was scored in the reproduction assays using the same worms as those scored for lifespan, except for the parental N2, *ppw-1* and *rrf-1* worms in the second experiment, where only lifespan was recorded. Unmated individual hermaphrodites were moved to new plates daily and scored for offspring produced 2.5 days later. In the "mated" treatment, two male *C. elegans* (from the initial sample population of N2 strain) were

placed on a plate with a single hermaphrodite for two hours every day to allow time for mating. Offspring production was scored 2.5 days later, as in the "unmated" treatment.

Egg size assays

Egg size was measured in N2, *ppw-1* and *rrf-1* strains (unmated hermpahrodites) growing on either *daf-2* RNAi or empty vector (EV) plates. Two days after maturation, worms were placed individually on new plates and observed continually during five hours for the presence of newly laid eggs, of which the first two eggs were collected. Eggs were picked immediately after laying and placed under a Leica M165C microscope set on 12x magnification; photos were taken using a Lumenera Infinity 2-6C digital microscope camera. Egg size was analysed from photos using *ImageJ* (https://imagej.nih.gov/ij/). Only eggs laid during gastrulation stage (the normal developmental stage at egg laying) were included in the analyses. This resulted in the following total numbers: N2 on EV: 56, N2 on *daf-2*: 54, *ppw-1* on EV: 44, *ppw-1* on *daf-2*: 42, *rrf-1* on EV: 59, *rrf-1* on *daf-2*: 42.

Statistical analyses

Survival was analysed for each experiment in Cox proportional hazard models in *R 3.3.3*. Mated and unmated individuals were analysed separately, as they were run in different blocks. Unmated individuals were analysed using the *coxph* function in the package *survival*, with *daf-2* RNAi treatment as a fixed factor. For mated individuals, we used the *coxme* package in order to fit block as a random effect, in addition to the fixed effect of RNAi treatment. In the second experiment, in addition to RNAi treatment, we also fitted the fixed factor strain (N2, *ppw-1*, *rrf-1*) and its interaction with treatment using the *coxph* function in the *survival* package.

Reproduction was analysed as total reproduction as well as rate-sensitive individual fitness λ_{ind} , which encompasses the timing and number of offspring [38, 39]. λ_{ind} is estimated by solving the Euler-Lotka equation for each individual using the *lambda* function in the *popbio* package and is analogous to the intrinsic rate of population growth (Stearns 1992). For all unmated worms, we estimated the fixed effect of treatment (*daf-2* RNAi or empty vector).

For offspring of the three mutants, we also estimated the fixed effect or strain, using linear models. For the mated worms, we also estimated the random effect of block, in addition to RNAi treatment. These models were implemented as mixed effect models using the *Ime4* package in *R 3.3.3*, and chi-square tests of fixed effects were performed using the *car* package. Egg size was analysed in a mixed effect model in *Ime4*, treating strain and RNAi treatment as crossed fixed effects, and parent ID as well as block as random effects.

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Author contributions

MIL and AAM designed the study, with the aid of AH. SR, ZS, MIL and HC collected the data, MIL analysed the data, MIL and AAM drafted the manuscript. All authors contributed to the revision of the manuscript.

Competing interests

There are no competing interests.

References

- 1. Kirkwood T.B.L. 1977 Evolution of aging. *Nature* **270**(5635), 301-304.
- 2. Kirkwood T.B.L., Austad S.N. 2000 Why do we age? *Nature* **408**(6809), 233-238.
- Kenyon C. 2005 The plasticity of aging: Insights from long-lived mutants. *Cell* **120**(4), 449-460. (doi:10.1016/j.cell.2006.02.002).
- Flatt T. 2011 Survival costs of reproduction in Drosophila. *Experimental Gerontology* 46(5), 369-375. (doi:10.1016/j.exger.2010.10.008).

- 5. Antebi A. 2013 Regulation of longevity by the reproductive system. *Experimental Gerontology* **48**(7), 596-602. (doi:10.1016/j.exger.2012.09.009).
- Dillin A., Crawford D.K., Kenyon C. 2002 Timing requirements for insulin/IGF-1 signaling in C-elegans. *Science* 298(5594), 830-834. (doi:10.1126/science.1074240).
- Partridge L., Gems D., Withers D.J. 2005 Sex and death: What is the connection? *Cell* 120(4), 461-472.
- Maklakov A.A., Immler S. 2016 The expensive germline and the evolution of ageing. *Current Biology* 26(13), R577-R586.
- Kenyon C.J. 2010 The genetics of ageing. *Nature* 464(7288), 504-512.
 (doi:10.1038/nature08980).
- Kenyon C. 2011 The first long-lived mutants: discovery of the insulin/IGF-1 pathway for ageing. *Philosophical Transactions of the Royal Society B-Biological Sciences* 366(1561), 9-16. (doi:10.1098/rstb.2010.0276).
- Gems D., Sutton A.J., Sundermeyer M.L., Albert P.S., King K.V., Edgley M.L., Larsen P.L., Riddle D.L. 1998 Two pleiotropic classes of daf-2 mutation affect larval arrest, adult behavior, reproduction and longevity in Caenorhabditis elegans. *Genetics* 150(1), 129-155.
- Walker D.W., McColl G., Jenkins N.L., Harris J., Lithgow G.J. 2000 Natural selection -Evolution of lifespan in C-elegans. *Nature* 405(6784), 296-297.
- Jenkins N.L., McColl G., Lithgow G.J. 2004 Fitness cost of extended lifespan in Caenorhabditis elegans. *Proceedings of the Royal Society of London Series B-Biological Sciences* 271(1556), 2523-2526. (doi:10.1098/rspb.2004.2897).
- Savory F.R., Benton T.G., Varma V., Hope I.A., Sait S.M. 2014 Stressful environments can indirectly select for increased longevity. *Ecology and Evolution* 4(7), 1176-1185. (doi:10.1002/ece3.1013).
- Maklakov A.A., Carlsson H., Denbaum P., Lind M.I., Mautz B., Hinas A., Immler S.
 2017 Antagonistically pleiotropic allele increases lifespan and late-life reproduction

at the cost of early-life reproduction and individual fitness. *Proceedings of the Royal Society B-Biological Sciences* **284**(1856). (doi:10.1098/rspb.2017.0376).

- Briga M., Verhulst S. 2015 What can long-lived mutants tell us about mechanisms causing aging and lifespan variation in natural environments? *Experimental Gerontology* **71**, 21-26. (doi:10.1016/j.exger.2015.09.002).
- Lind M.I., Chen H.Y., Meurling S., Gil A.C.G., Carlsson H., Zwoinska M.K., Andersson J.,
 Larva T., Maklakov A.A. 2017 Slow development as an evolutionary cost of long life.
 Functional Ecology **31**(6), 1252-1261. (doi:10.1111/1365-2435.12840).
- Williams G.C. 1957 Pleiotropy, natural selection, and the evolution of senescence.
 Evolution 11(4), 398-411.
- 19. Kirkwood T.B.L., Holliday R. 1979 Evolution of aging and longevity. *Proceedings of the Royal Society of London Series B-Biological Sciences* **205**(1161), 531-546.
- 20. Gems D., Partridge L. 2013 Genetics of longevity in model organisms: debates and paradigm shifts. *Annual Review of Physiology* **75**, 621-644. (doi:10.1146/annurev-physiol-030212-183712).
- Blagosklonny M.V. 2010 Revisiting the antagonistic pleiotropy theory of aging TORdriven program and quasi-program. *Cell Cycle* 9(16), 3151-3156. (doi:10.4161/cc.9.16.13120).
- Ezcurra M, Benedetto A, Sornda T, Gilliat AF, Au C, Zhang Q, van Schelt S, Petrache AL, Wang H, de la Guardia Yila, et al. 2018 C. elegans Eats Its Own Intestine to Make Yolk Leading to Multiple Senescent Pathologies. *Current Biology* 28, 2544-2556.
- 23. Blagosklonny M.V. 2006 Aging and immortality Quasi-programmed senescence and its pharmacologic inhibition. *Cell Cycle* **5**(18), 2087-2102. (doi:10.4161/cc.5.18.3288).
- Sivasundar A., Hey J. 2005 Sampling from natural populations with RNAi reveals high outcrossing and population structure in Caenorhabditis elegans. *Current Biology* 15(17), 1598-1602. (doi:10.1016/j.cub.2005.08.034).

- Maures T.J., Booth L.N., Benayoun B.A., Izrayelit Y., Schroeder F.C., Brunet A. 2013 Males shorten the life span of C. elegans hermaphrodites via secreted compounds. *Science* 343(6170):536-540.
- 26. Shi C., Murphy C.T. 2014 Mating induces shrinking and death in Caenorhabditis mothers. *Science* Express, 1-7.
- 27. Aprison E.Z., Ruvinsky I. 2016 Sexually antagonistic male signals manipulate germline and soma of C. elegans hermaphrodites. *Current Biology* **26**, P2827-2833.
- 28. Stearns S.C. 1992 *The Evolution of Life Histories*. New York, Oxford University Press.
- Hibshman JD, Hung A, LR B. 2016 Maternal Diet and Insulin-Like Signaling Control Intergenerational Plasticity of Progeny Size and Starvation Resistance. *PLoS Genetics* (doi: 10.1371/journal.pgen.1006396).
- Kirkwood T.B.L. 2017 The Disposable Soma Theory of Ageing. In *The Evolution of Senescence in the Tree of Life* (eds. Shefferson RP, Jones OR, R S.-G.), pp. 23-39.
 Cambridge, United Kingdom, Cambridge University Press.
- Agrawal A.F., Wang A.D. 2008 Increased transmission of mutations by low-condition females: Evidence for condition-dependent DNA repair. *Plos Biology* 6(2), 389-395. (doi:10.1371/journal.pbio.0060030).
- 32. Sniegowski P.D., Gerrish P.J., Johnson T., Shaver A. 2000 The evolution of mutation rates: separating causes from consequences. *Bioessays* **22**(12), 1057-1066.
- Maklakov A.A., Rowe L., Friberg U. 2015 Why organisms age: Evolution of senescence under positive pleiotropy? *Bioessays* 37(7), 802-807. (doi:10.1002/bies.201500025).
- Templeman N.M., Luo S.J., Kaletsky R., Shi C., Ashraf J., Keyes W., Murphy C.T. 2018 Insulin Signaling Regulates Oocyte Quality Maintenance with Age via Cathepsin B Activity. *Current Biology* 28(5), 753-+. (doi:10.1016/j.cub.2018.01.052).
- Lionaki E., Tavernarakis N. 2013 High-throughput and longitudinal analysis of aging and senescent decline in Caenorhabditis elegans. In *Cell Senescence* (eds. Lorenzo Galluzzi, Ilio Vitale, Oliver Kepp, Kroemer G.), pp. 485-500.

- Timmons L., Court D.L., Fire A. 2001 Ingestion of bacterially expressed dsRNAs can produce specific and potent genetic interference in Caenorhabditis elegans. *Gene* 263(1-2), 103-112. (doi:10.1016/s0378-1119(00)00579-5).
- Hinas A., Wright A.J., Hunter C.P. 2012 SID-5 Is an Endosome-Associated Protein Required for Efficient Systemic RNAi in C. elegans. *Current Biology* 22(20), 1938-1943. (doi:10.1016/j.cub.2012.08.020).
- Brommer J.E., Merila J., Kokko H. 2002 Reproductive timing and individual fitness.
 Ecology Letters 5(6), 802-810. (doi:10.1046/j.1461-0248.2002.00369.x).
- Lind M.I., Zwoinska M.K., Meurling S., Carlsson H., Maklakov A.A. 2016 Sex-specific trade-offs with growth and fitness following lifespan extension by rapamycin in an outcrossing nematode, Caenorhabditis remanei. *Journals of Gerontology Series a-Biological Sciences and Medical Sciences* 71(7): 882-890 (doi: 10.1093/gerona/glv1174).

Table 1. Total reproduction and individual fitness (λ_{ind}) for unmated and mated *C. elegans*

N2 wild-type treated with either empty vector (Control) or daf-2 RNAi from adulthood

onwards. All values expressed as mean ± SE.

	Total reproduction		Fitness (λ _{ind})	
RNAi treatment	unmated	mated	unmated	mated
Control	311.0 ± 7.0	595.7 ± 24.3	4.66 ± 0.03	4.47 ± 0.05
<i>daf-2</i> RNAi	317.4 ± 8.9	630.5 ± 22.2	4.63 ± 0.05	4.50 ± 0.05

Figures

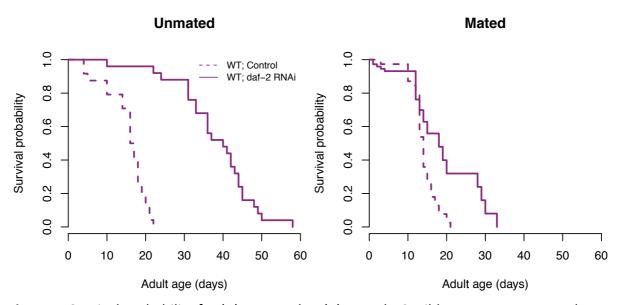


Figure 1. Survival probability for (A) unmated or (B) mated N2 wild-type worms, treated with either *daf-2* RNAi (solid lines) or control empty vector (broken lines) from adulthood onwards.

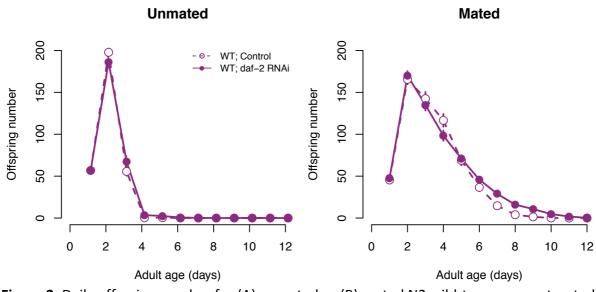
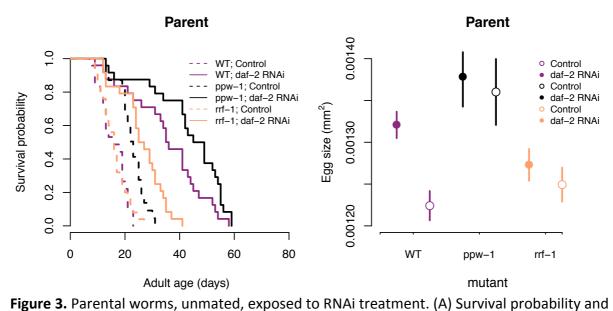


Figure 2. Daily offspring number for (A) unmated or (B) mated N2 wild-type worms, treated with either *daf-2* RNAi (solid lines, filled symbols) or control empty vector (broken lines, open symbols) from adulthood onwards. Symbols represent mean ± SE.



(B) egg size of N2 wild-type (purple), *ppw-1* (black) and *rrf-1* (orange) mutants, treated with either *daf-2* RNAi (solid lines, filled symbols) or control empty vector (broken lines, open symbols) from adulthood onwards. Symbols represent mean ± SE.

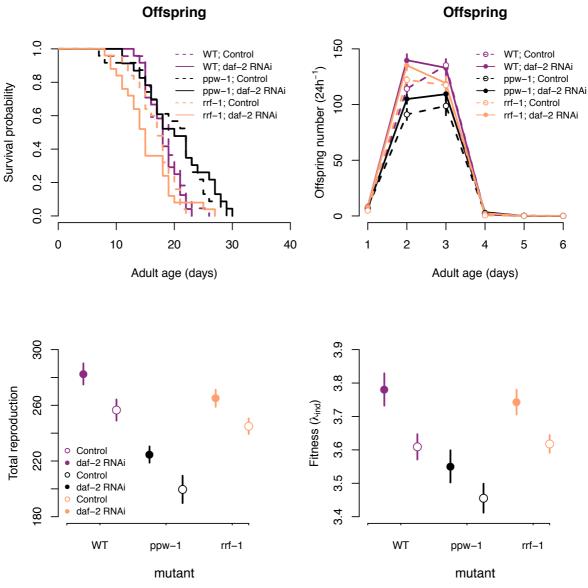


Figure 4. Offspring worms, unmated, on control (empty vector) plates from parents exposed to *daf-2* RNAi or control treatment. (A) Survival probability, (B) daily offspring number, (C) Total reproduction and (D) individual fitness (λ_{ind}) of offspring (on control plates) from parents either exposed to *daf-2* RNAi (solid lines, filled symbols) or control empty vector (broken lines, open symbols). The colors reflect N2 wild-type (purple), *ppw-1* (black) and *rrf-1* (orange) mutants. Symbols represent mean ± SE.