ASSESSMENT OF GENETIC STRUCTURE OF THE ENDANGERED FOREST

SPECIES *BOSWELLIA SERRATA* ROXB. POPULATION IN CENTRAL INDIA

Vivek Vaishnav[1], Shashank Mahesh[2], Pramod Kumar[3]*

[1]Institute of Forest Productivity, Ranchi, Jharkhand, India, 835303

[2]National Institute for Research in Tribal Health, Jabalpur, Madhya Pradesh, 482001

[3]Tropical Forest Research Institute, Jabalpur, Madhya Pradesh, 482021

* pramod_kt@rediffmail.com

ABSTRACT

Boswellia serrata Roxb., a commercially important species for its pulp and pharmaceutical

properties was sampled from three locations representing its natural distribution in central

India for genetic characterization through 56 RAPD + 42 ISSR loci. The wood fiber

dimensions measured for morphometric characterization confirmed 11.36% of the variation

in the length and 8.75% of the variation in the width indicating its fitness for local adaptation.

Bayesian and non-Bayesian approach based diversity measures resulted moderate within

population gene diversity (0.26±0.17), Shannon's information index (0.40±0.22) and

panmictic heterozygosity (0.28±0.01). A high estimate for genetic differentiation measures

i.e. G_{ST} (0.31), G_{ST}-B (0.33±0.02) and θ-II (0.45) led to the distinct clusters of the sampled

genotypes representing their regional variability due to limited gene flow and total absence of

natural regeneration. We report the first investigation of the species for its molecular

characterization emphasizing the urgent need for the genetic improvement program for the

In-situ/Ex-situ conservation and sustainable commercialization.

Keywords: Burseraceae, θ-statistics, Shannon's information index, genetic improvement

INTRODUCTION

Boswellia serrata Roxb. (family- Burseraceae), commonly known as salai guggul or Indian frankincense (olibanum indicum) is a commercially important deciduous tree of India (Shah et al. 2008). It is found in the region having rainfall between 500mm -2000mm and temperature up to 45 °C. The species has the ability to thrive in the poorest and the shallowest soils (Bhat et al. 1952). In India, it is distributed in states Rajasthan, Maharashtra, Madhya Pradesh, Karnataka and Chhattisgarh (Pawar et al. 2012). The oleo-gum-resin of the tree contains boswellic acid, which is effective for the treatment of inflammatory disorders, arthritis, cardiovascular diseases (Ammon 2006), diarrhea, dysentery and other skin diseases (Khare 2004). Apart from these medicinal values, the resin of the species was found a more effective sizing agent in papermaking than the rosin obtained from *Pinus* species (Sharma et al. 1985). The convincing wood quality for pulp making led to the establishment of the first paper mill in India in Nepanagar of Madhya Pradesh (Khan 1972) as the region had been occupied by wide range of natural patch of the species.

The species is highly out-crossing supported by the self-incompatibility to selfing (Sunnichan et al. 2005). But, a poor fruit setting (2.6% - 10%) under open pollination condition, inadequate production of viable seeds and scanty seed germination (10% - 20%) limit the distribution of the species in nature (Ghorpade et al. 2010). The species population has been harvested for the frankincence also (Sharma 1983). The scarcity of protocols to regenerate it through seeds and clones makes the mass multiplication difficult (Purohit et al. 1995). This situation resulted in declined abundance of the species and the International Union for Conservation of Nature (IUCN) has enlisted it under the status of endangered. Therefore, the available natural patches of the species require keen attention for both *In-situ*

and *Ex-situ* conservation with actual knowledge of available genetic resources of the species

for management and breeding objectives.

Information on actual genetic variability and a cryptic number of the genetically

differentiated genetic resource of any species is an important aid for its conservation and

genetic improvement. For such purposes, DNA based markers are of value because unlike the

morphometric traits the molecular markers are independent of the influence of environment

and can be assessed at any growth stage. Among the various molecular markers employed to

assess diversity studies, PCR-based dominant markers such as RAPD (random amplified

polymorphic DNA, Williams et al. 1990) and ISSR (inter-simple sequence repeat,

Zietkiewiez et al. 1994) have become popular due to its polymorphism and discrimination

power, as their application does not need any prior sequence information. These primer

systems have been successfully applied for the genetic characterization of populations of

tropical tree species (Ansari et al. 2012, Abuduli et al. 2016, Vaishnav & Ansari 2018).

Bayesian statistics has been extended to dominant markers for precise estimates of population

genetic hierarchies equivalent to those obtained with codominant markers, circumventing

inbreeding estimate within the population (Holsinger et al. 2002). Therefore, the present

investigation was conducted to differentiate the morphometric and the genetic variability

exhibited by the natural population of *B. serrata* Roxb. in three agro-climatic regions of

central India applying dominant marker system.

MATERIAL AND METHODS

Population sampling

The information of the distribution of the patches or population of the species was obtained

from the old forest working plans of the concerned forest departments of the Indian state

Chhattisgarh, which is classified in three agro-climatic zones *viz.* Northern Hills,

Chhattisgarh plain and Bastar plateau (Table 1). Forests of these agro-climatic regions were

visited during July 2014 to September 2014 to collect samples from 20 trees of each region in

natural distribution of *B. serrata* respecting inter-tree distance of 100 m along the latitude

(Figure 1). The girth at breast height (GBH) was measured by measuring tape. A wood radial

core sample was extracted from each tree at breast height (1.34 m) and was stored in a tube

with 40% formaldehyde. A leaf sample was also collected from each tree. The samples were

brought to the laboratory in a cryo-box.

Measurement of wood fiber dimensions

78   In the laboratory, the wood samples were macerated following the method described in

79   Mahesh et al. (2015). Five slides were prepared for each tree and the wood fiber length

80   (WFL) and width (WFW) of five fibers per slide were measured through the compatible

81   program integrated with the compound microscope (Leica, EC3, Switzerland).

DNA isolation and amplification

DNA from the leaf samples was isolated following modified CTAB method (Deshmukh et al.

2007). To select the reproducible markers, 10 RAPD and 10 ISSR primers were cross-

amplified on set of 12 genotypes of the species (four from each region) and finally, only five

from each of RAPD and ISSR markers were selected based on their polymorphism and

reproducibility for the final amplification of all sixty genotypes (Table 3). For RAPD/ISSR

amplification, the final reaction mixture for PCR assay was consisted of 15 µl/10 µl

containing 50 ng/ 40ng genomic DNA, 0.66 µM/0.8 µM of primers, 0.2 mM/ 0.1mM of

dNTP mix, 1.5 mM/2.5 mM $MgCl_2$, 1X buffer with KCl and 1 unit of Taq polymerase. PCR

parameters included an initial 4 minutes denaturation step at 94 °C followed by 35 cycles

of30s at 94 °C, 30s at the annealing temperature of 35 °C/50 °C, 45s extension at 72 °C

followed by a final extension of 5 minutes at 72 °C. The amplified products were

electrophoresed on 1.5% agarose gel containing 0.5µg/ml ethidium bromide (EtBr) in 0.5 X

TBE (pH 8.0). The separation was carried out by applying constant 100V for 3 hours. The

fractionated amplified products on agarose gel were visualized on gel documentation system

under UV light. To avoid the homology, the amplification profile of all sixty genotypes was

evaluated through the molecular weight of the bands for each primer and the bands were

scored in a Microsoft Excel sheet in binary data format.

Data analysis

The measures of central tendency and the coefficient of variation (CV) were calculated for

GBH, WFL, and WFW. A genetic profile of 60 genotypes on 10 markers was generated and

analyzed following both band-based and allele frequency-based approaches as suggested in

Bonin et al. (2007). The information from the markers was evaluated estimating mean allelic

frequency (AF), gene diversity (GD), and polymorphic information content (PIC) by program

POWERMARKER v3.1 (Liu & Muse 2005). Resolving power (RP) of the markers was

calculated following the formula given by Prevost and Wilkinson (1999).

The genetic variability and the differentiation of the sampled population were evaluated

applying both non-Bayesian and Bayesian approaches. The program POPGENE v1.32 (Yeh

et al. 1999) was applied for the non-Bayesian estimates of genetic diversity measures i.e.

Nei's gene diversity (GD), Shannon's information index (I) and the percentage of

polymorphism (P%) for each location and G_{ST} was calculated to evaluate the level of genetic

differentiation. The genetic relationship among the genotypes was calculated based on

Jaccard's genetic similarity coefficient by the program DARWIN v5.0 (Perrier &

Jacquemoud-Collet 2006). The principal coordinate analysis (PCoA) was performed to

cluster the genotypes in different axes applying program GENALEX v6.5 (Peakall & Smouse

2012). Analysis of molecular variance (AMOVA) was performed applying program

ARLEQUIN v3.11 (Excoffier & Lischer 2010) to estimate the hierarchical variation. The

isolation by distance (IBD) was confirmed calculating the correlation coefficient between two

pairwise matrices of genetic and geographical distances among genotypes through Mantel's

test performed by program Alleles in Space (Miller 2005) in independent runs (with and

without logarithmic transformation) on 10,000 permutations.

For the Bayesian approach based genetic estimates we preferred the θ-statistics (*viz.* θ-I,

θ-II, θ-III and G_{ST}-B) implemented in program HICKORY v1.1 (Holsinger & Lewis 2007)

that allows direct estimation of genetic differentiation measure (F_{ST}) from dominant markers

without assumption of prior knowledge of the extent of inbreeding and Hardy –Weinberg

Equilibrium (HWE) within population even with small size and number. Region-wise

panmictic heterozygosity (Hs) and total panmictic heterozygosity (Ht) were also estimated.

All these estimations were performed with 50,000 steps of burn-in, 500,000 replicates,

'thinning' = 20 following the best-suited model for the data amongst the all four models (*viz.*

full, f = 0, θ = 0 and f-free models) implemented in the program based on the minimum DIC

(deviance information criterion) value as suggested by Spiegelhalter et al. (2002). To

determine the most suitable number of cryptic populations (K) for the dataset, the program

STRUCTURE v2.3.1 (Pritchard et al. 2000) was run applying model combinations of

admixture/ no-admixture model with correlated/ independent allele frequencies among the

samples on 100000 burn-in and 1000000 MCMC repeats with three-run from K=1 to K=9,

for number of K $2 \leq K \leq 8$. The most suitable model and the best-suited 'K' was determined

based on the highest Delta-K value resulted by online program STRUCTURE HARVESTER

(Earl and VonHoldt 2012).

RESULTS

Variation in wood fiber dimension

The WFL of the species ranged between 0.803 mm to 1.397 mm with average of 0.968±0.11

mm (11.36% CV) and the WFW of the species ranged between 0.019 mm to 0.030 mm with

average of 0.025±0.002 mm (8.75% CV). Both traits followed normal distribution and no

significant correlation was observed between them. WFL and WFW both were evenly

distributed on the standard deviation (σ) based classes and no significant variation was

observed among the locations based on WFL and WFW (Table 2).

Genetic informativeness of markers

The five RAPD primers amplified 826 bands and 56 loci (14.75 bands/locus). The diversity

measures AF and GD were 0.81±0.065 and 0.27±0.075 respectively. PIC and RP were

0.22±0.054 and 5.51±3.17 subsequently (Table 3). For RAPD markers, the diversity

measures and discriminatory power measures were found in significant ($p<0.05$) linear

correlation. The five ISSR primers amplified 1004 bands for 42 loci (23.90 bands/locus). In a

significant linear correlation ($p<0.01$) to each other, the AF was 0.76±0.068 and the GD was

0.33 ±0.073. The PIC and the RP were 0.27 ±0.052 and 6.69±1.69 subsequently (Table 3).

Genetic diversity and differentiation

Among the non-Bayesian estimates of genetic diversity measure for the species population,

100% polymorphism, 0.26±0.17 GD, and 0.40±0.22 I were observed among the regional

locations (Table 4). For all measures, no significant difference was found among the three

regions. Dhamtari was found slightly more diverse and polymorphic than the other two

regions (Table 4). The G_{ST} was 0.31. The Bayesian model implemented in Hickory estimated

the lowest DIC value (896.96) for the full model and the panmictic heterozygosity was

0.28±0.01. No significant variation in panmictic heterozygosity was found among the three

regional sub-populations (Table 4). The θ-I, θ-II, and θ-III values were 0.50±0.04, 0.45±0.04

and 0.25±0.01 subsequently and the G_{ST}-B was 0.33±0.02.

The dendrograph based on Jaccard's genetic similarity coefficient (average 0.67) grouped

the genotypes following their regions (Figure 2). The genotypes sampled from Sarguja

further bifurcated into two different but conjoint clusters supported by the bootstrap values

>60 (Figure 2). The PCoA accounting for 68.97% cumulative separation based on the same

genetic distance separated the genotypes of Narayanpur from rest of the sampled genotypes

and the equal proportion of the genotypes from Dhamatari and Sarguja was in the separate

group from their sampled locations making overall four small clusters (Figure 3). The

Bayesian program STRUCTURE assigned K=2 as the most appropriate number of cryptic

population for the sampled genotypes based on the highest deltaK value in STRUCTURE

HARVESTER. The bar plot generated (Figure 4) shows admixing of genotypes assigned to

the two cryptic populations (K=2) with >80% of ancestry coefficient. The genotypes from

Dhamtari were found admixed with both of the cryptic populations, the genotypes from

Narayanpur and Dhamatri were assigned to be in the same cluster. On the other hand,

genotypes from Sarguja clustered distinctly (Figure 4). AMOVA resulted 46.03±0.65%

variation among groupings suggested by dendrograph, PCoA, and STRUTURE (K=2) and

53.38±0.15% variation within these groups. Mantel's test found significant (p<0.001)

correlation between pair-wise genetic and geographical distances among the genotypes.

DISCUSSION

B. serrata Roxb. a member of family Burseraceae, bifurcated from "Terebinthaceae" along

with Anacardiaceae during the Miocene epoch due to lesser intrinsic ability to adopt the

changing conditions across the climatic niches (Weeks et al. 2014). Therefore, the

distribution of genus *Boswellia* limited within the mediterranian and tropical regions. *B.*

serrata Roxb. is found only in central India and the species is renowned for its olio-gum-resin

and pulp content. Due to uncontrolled harvesting, it has become scarce in nature and the

literature regarding its distribution is also not adequate to locate it. During the investigation,

the species was found in the temperature range of 19.02±6.35 °C to 33.02±4.07 °C and the

annual rainfall of 1595.33±95.30 mm confirming its suitability for moist conditions along

with the arid regions. For molecular characterization of the species, the RAPD and ISSR

markers are preferred due to the absence of background genomic information of the species

to design microsatellites and low-cost applicability. The problems with the dominant markers

such as low reproducibility and uncertain homology of alleles have been handled during

amplification and data profiling. Bayesian model-based θ-statistics has been applied based on

its successful estimation of the genetic variability and the differentiation of population

applying dominant markers system (Holsinger & Wallace 2004) in forest species as well (Ci

et al. 2008,Vaishnaw et al. 2015).

Variation in wood fiber dimension

The wood fiber length along with the thickness is considered to be the most important

characteristics for the mechanical strength of wood (Migneault et al. 2008) and for the

making of high-quality groundwood in paper industries (Zobel & Jett 1995). The comparison

with the other tropical trees employed in pulp making process establishes that the wood fiber

dimensions of *B. serrata* Roxb. obtained in our investigation is much higher than those of

Ailanthus altissima (WFL= 0.747±0.035 mm and WFW= 0.023±0.0004 mm) and *Eucalyptus*

globulus (WFL = 0.785±0.012 mm and WFW= 0.018±0.0005) reported in Baptista et al.

(2014). The WFL is lower than of *Eucalyptus grandis* (1.01±0.14 mm) reported in Bhat et al.

(1990). The coefficient of variation of wood fiber dimension in *B. serrata* is higher than *A.*

altissima (WFL= 4.76% and WFW= 1.66%) and *E. globulus* (WFL= 1.5% and WFW=

2.72%) and lower than *E. grandis* (WFL= 14.45%) in above studies. In terms of adaptability,

such high variation confirms the adaptive fitness of the population to the local ecological

conditions (MacPherson et al. 2015).

Genetic informativeness of the primers

With an approach to handle the limitation of the non-reproducibility, the dominant markers

(RAPD/ISSR) have exhibited moderate values for AF and GD. It indicates that the approach

avoiding null alleles have controlled the informativeness of the primers helping genetic

differentiation (Lynch & Milligan 1994). The AF exhibited by the RAPD/ISSR markers are

slightly higher to the information resulted in populations of *Tectona grandis* L. f. sampled

from India on ISSR markers (0.73±0.02, Vaishnav & Ansari 2018). On the other hand,

negligible differences in AF among the markers of RAPD and ISSR systems ensure the

qualitative support for the diversity assessment (Lynch & Milligan 1994, Nybom & Bartish

2000). The discriminatory power measures exhibit a strong and linear relationship between

the ability of a primer to distinguish genotypes (Prevost & Wilkinson 1999). The values of

the PIC and RP are comparable to those obtained with dominant markers in other species, e.g.

Podophyllum hexandrum (Naik et al. 2010), *Jatropha curcas* (Grativol et al. 2011) and

Pongamia pinnata (Sharma et al. 2014).

Genetic diversity and differentiation

In order to estimate the genetic diversity of *B. serrata* Roxb. population, we applied both;

band-based and allele frequency-based approach with non-Bayesian and Bayesian statistics

so that the under-estimation or over-estimation of the diversity measures caused by the

dominant marker systems could be avoided. A significant correlation between diversity

measures i.e. P%, GD, I and Hs; with slight differences among them for different regions,

confirms almost equal heterogeneity of the species population. The gene diversity of the

sampled population has been found slightly higher than that reported by Nybom (2004) for

out-crossing species (0.18). Nevertheless, Holsinger & Wallace (2007) suggested avoiding

the comparison among the diversity measures resulted by the neutral markers (AFLP/RAPD)

as these markers only show the differences in rates of mutations at their loci rather than

reflecting different migration rates. However, there are few studies based on the tree species

of the central Indian regions *viz.* Wang et al. (2011) investigated the genetic diversity of *D.*

sissoo sampled from Indian regions and found 89.11% of polymorphism, 0.27 ± 0.16 GD and

0.41 ± 0.23 I. Ansari et al. (2012) sampled *Tectona grandis* L. f. from all along the central and

peninsular Indian regions and found 80.30% polymorphism, 0.32 GD and 0.45 I. The

comparison of the results from present investigation with above studies indicates that the

species natural distribution and the level of genetic diversity exhibited are somehow equally

influenced by the geographical conditions and the landscapes. The significant correlation

found between genetic and geographical distances of the genotypes in our result also supports

the fact.

The genetic structure of a population reflects the long-term evolutionary history of the

species along with its mating system (Duminil et al. 2007), reproductive biology and gene

flow influenced by the adaptive selection and fragmentation (Slatkin 1987, Nybom & Bartish

2000). In our investigation, the results of Bayesian model-based investigation through

HICKORY found full-model as the most suitable for our data determining the influence of

inbreeding within the populations. The low level of panmictic heterozygosity (Hs) for

different regions and for overall population also supports it. Like other out-crossing species

(Hamrick & Godt 1989; Nybom 2004) the *B. serrata* Roxb also exhibited higher within-

population variability than among populations but the genetic variation within the population

resulted by AMOVA was found comparatively lower than those reported in other out-

crossing forest species (Nybom 2004, Wang et al. 2011, Ansari et al. 2012). A limited gene

flow in *B. serrata* Roxb. is one of the important factors that have led to the high

differentiation among sampled populations and the genotypes. The species has an

entomophilous mode of pollination and the self-incompatible flowers support only cross-

pollinated pollen to grow the pollen tubes. Moreover, only up to 10% fruit set has been

observed in open-pollination condition (Sunnichan et al. 2005). In our field observation also,

we noticed earlier abscission of unripe fruits and ripen fruit with empty seeds led to the total

absence of natural regeneration in all sites of sampling. These bottlenecks contribute to

genetic drift and signatory inbreeding depression to the species population reflected through

different estimates.

The genetic differentiation measures G_{ST} and G_{ST}-B have resulted in comparatively equal

values indicating the cautions taken to employ the dominant markers for the investigation

could be equivalent to the Bayesian model correction. The θ-statistics measures have

exhibited differences in their values due to a low number of populations sampled for the

investigation (Holsinger & Wallace 2004). The θ-I is supposed to be equivalent to Wright's

F_{ST} (Wright 1969) and its higher value (0.50±0.04) is expected because it results in the scaled

allele frequency measured across the evolutionary time. The θ-II is a more appropriate

measure for a small number of admixed population and its higher value (0.45±0.004) reflects

the problematic gene exchange among the genotypes leading to high differentiation.

Among the Bayesian clusterings resulted by STRUCTURE, K=2 has been found the most

suitable for our data on the basis of deltaK value. Nevertheless, the presence of inbreeding in

sampled population and deviation from the HWE suggest preferring the dendrograph and the

PCoA over the two admixed cryptic populations resulted by the program STRUCTURE

because of its prior assumption of HWE. In PCoA, We have found Sarguja population closely

related to Dhamtari population and the Narayanpur population has represented itself

distinctly even after geographical closeness to Dhamatri. It might be due to the gene flow

among these populations follows geographical limits along with their own biological

constraints. On the other hand, the Narayanpur population might represent a different source

of genetic origin of southern Indian region that could not be sampled. An almost equal

polymorphism, genetic diversity, and heterozygosity but with highly differentiable genetic

resource resulting to the distinctly different clusters indicate that the sampled populations

may belong to a large metapopulation of the species that has been fragmented with due

course of time due to its own intrinsic inability to cope up with natural and anthropogenic

pressures later leading to geographical isolation.

B. serrata Roxb. is a highly out-crossing species with self-incompatibility and therefore,

its own constraint to exchange the genes and the fragmentation of the population due to

geographical isolation and commercial harvesting might be the reason for detected inbreeding

pressure. The estimates of genetic diversity measures, genetic structure, and level of

differentiation of a species population depend on the molecular marker system employed for

the investigation. In our investigation, the dominant markers might be a reason behind low to

moderate estimates of gene diversity, panmictic heterozygosity, and within-population

variability. On the other hand, the Bayesian model-based analysis has given a higher value of

θ-II because the θ-statistics has been developed specifically for the dominant markers

(Spiegelhalter et al. 2002) and has been found more appropriate than G_{ST} and G_{ST}-B to

estimate the inbreeding and population differentiation (Holsinger et al. 2002, Holsinger &

Wallace 2004). The comparatively higher variation in morphometric traits also indicates the

existence of higher variability in the population. The northernmost region Sarguja and the

southernmost region Narayanpur might represent different centers of genetic origin of a large

metapopulation of the species and the genes from Sarguja are immigrating to the central

region Dhamtari. For a better understanding of the influence of landscape and other

geographical features on distribution and gene diversity of *B. serrata* Roxb. an investigation

can be conducted with a larger number of locations representing its entire range of natural

distribution applying co-dominant multiallelic marker system to cross verify the results of our

investigation.

CONCLUSION

B. serrata Roxb. has been over-exploited due to its significant quality of wood for pulp and

paper industries. The technology in paper industries to use the species as a raw material has

become obsolete and nowadays the oleo-gum-resin obtained from its wood is of significant

medicinal value to heal arthritis and asthma. The species is endemic to India and

unfortunately, there is not a single source of sustainable supply for the raw material required

in pharmaceutical industries. Apart from all, the species is understudied and has not received

much scientific attention to report on population genetic. Through the present investigation,

we report that the available genetic resource of the species in central Indian region maintains

high variation in wood fiber dimension and exhibits higher genetic variability within and

among its sub-populations. In order to maintain the valuable forest genetic resource, the

species need prior attention for the conservation. The locations of its natural distribution can

be developed as strict nature reserves. On the other hand, a genetic improvement program

should be initiated for selection and multiplication of elite genotypes. There is need to

develop protocols for the artificial generation of the species through clonal and tissue culture

techniques.

ACKNOWLEDGEMENT

Authors are grateful to Director, Tropical Forest Research Institute, Jabalpur for Institutional

support and to Director General, Indian Council of Forestry Research and Education,

Dehradun for the financial support received under the project ID-175/TFRI/2011/Gen-4 (23).

REFERENCES

ABUDULI A, AYDIN Y, SAKIROGLU M. 2016. Molecular evaluation of genetic diversity
in wild-type mastic tree (*Pistacia lentiscus* L.). *Biochemical Genetics* 54 (5): 619-635.
doi:10.1007/s10528-016-9742-0

AMMON HP. 2006. Boswellic acid in chronic inflammatory disease. *Planta Medica* 72 (12):
1100-1116. doi:10.1055/s-2006-947227

ANSARI SA, NARAYANAN C, WALI SA, KUMAR R, SHUKLA N, KUMAR SR. 2012.
ISSR markers for analysis of molecular diversity and genetic structure of Indian teak
(*Tectona grandis* L f) locations. *Annals of Forest Research* 55 (1): 1-13.

BAPTISTA P, COSTA AP, SIMOES R, AMARAL ME. 2014. *Ailanthus altissima*: An
alternative fiber source for papermaking. *Industrial Crops & Products* 52: 32-37. doi:
10.1016/j.indcrop.2013.10.008

BHAT KM, BHAT KV, DHAMODARAN. 1990. Wood density and fiber length of
Eucalyptus grandis grown in Kerala, India. *Wood Fiber Science* 22 (1): 54-61.

BHAT RV, GUHA SRD, PANDA SP. 1952. Indigenous cellulosic raw materials for the
production of pulp, paper and board, part VIII- writing and printing papers from
Boswellia serrata Roxb. (Salai). *Indian Forester* 78 (4): 169 – 175.

BONIN A, EHRICH D, MANEL S. 2007. Statistical analysis of amplified fragment length
polymorphism data: a toolbox for molecular ecologists and economists. *Molecular Ecology*
16: 3737–3758. Doi:10.1111/j.1365-294X.2007.03435.x

CI X, CHEN J, LI Q. 2008. AFLP and ISSR analysis reveals high genetic variation and inert-
population differentiation in fragmented populations of the endangered *Litsea szemaois*
(Lauraceae) from Southwest China. *Plant Systematics & Evolution* 273: 237-346.
https://doi.org/10.1007/s00606-008-0012-4

DESHMUKH VP, THAKARE PV, CHAUDHARI US, GAWANDE PA. 2007. A simple method for isolation of genomic DNA from fresh and dry leaves of *Terminalia arjuna* (Roxb.) Wight and Argot. *Electronic Journal of Biotechnology* 10: 468-472.

DUMINIL J, DAINOU K, KAVIRIRI DK, GILLET P, LOO J, DOUCET JL, HARDY OJ. 2016. Relationships between population density, fine-scale genetic structure, mating system and pollen dispersal in a timber tree from African rainforests. *Heredity* 166: 295-303. doi: 10.1038/hdy.2015.101

EARL DA, VONHOLDT BM. 2012. STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conservation Genetics Resources* 4: 359-361.

EXCOFFIER L, LISCHER HEL. 2010. ARLEQUIN Suite ver 3.5: A new series of programs to perform population genetics analyses under LINUX and WINDOWS. *Molecular Ecology Notes* 10: 564–567. doi: 10.1111/j.1755-0998.2010.02847.x

GHORPADE RP, CHOPRA A, NIKAM TD. 2010. *In vitro* zygotic embryo germination and propagation of an edangered Boswellia serrata Roxb., a source of boswellic acid. *Physiology & Molecular Biotechnology of Plants* 16 (2): 159-165. doi: 10.1007/s12298-010-0017-7

GRATIVOL C, DA FONSECA LIRA-MEDEIROS C, HEMERLY AS, FERREIRA PC. 2011. High efficiency and reliability of inter-simple sequence repeats (ISSR) markers for evaluation of genetic diversity in Brazilian cultivated *Jatropha curcas* L. accessions. *Molecular Biology Reporter* 38 (7): 4245-4256. doi: 10.1007/s11033-010-0547-7

HAMRICK JL, GODT MJW. 1989. Allozyme diversity in plant species. In: Brown AHD, Clegg MT, Kahler AL, Weir BS. (ed), Plant Population Genetics: Breeding and Genetic Resources. Sinauer, Sunderland; Massachusetts, USA, pp 43–63

HOLSINGER KE, LEWIS PO, DEY DK. 2002. A Bayesian approach to inferring population structure from dominant markers. *Molecular Ecology* 11: 1157 – 1164.

HOLSINGER KE, LEWIS PO. 2007. Hickory: A package for analysis of population genetic data, version 1.1. Computer program and documentation, Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, Connecticut, USA

HOLSINGER KE, WALLACE LE. 2004. Bayesian approach for analysis of population structure: an example from *Planthanthera leucophaea* (Orchidaceae). *Molecular Ecology* 13: 887–894.

KHAN MAW. 1972. Propagation of *Boswellia papyrifera* through branch-cuttings. *Indian Forester* 98 (7): 437-440.

KHARE CP. 2004. Encyclopedia of India, Rational Western Therapy, Ayurvedic and other Traditional Usage. Botany. Springer, Berlin

LIU K, MUSE SV. 2005. POWERMARKER: Integrated analysis environment for genetic marker data. *Bioinformatics* 21 (9): 2128-2129. doi:10.1093/bioinformatics/bti282

LYNCH M, MILLIGAN BG. 1994. Analysis of population genetic structure with RAPD markers. *Molecular Ecology* 3 (2) : 91-99. doi:10.1111/j.1365-294x.1994.tb00109.x.

MACPHERSON A, HOHENLOHE PA, NUISMER SL. 2015. Trait dimensionality explains widespread variation in local adaptation. *Proceedings of the Royal Society of London B* 282: 20141570. http://dx.doi.org/10.1098/rspb.2 014.1570

MAHESH S, KUMAR P, ANSARI SA. 2015. A rapid and economical method for the maceration of wood fibers in *Boswellia serrata* Roxb. *Tropical Plant Research* 2 (2): 108-111.

MIGNEAULT S, KOUBAA A, ERCHIQUI F, CHAALA A, ENGLUND K, KRAUSE C, WOLCOTT M. 2008. Effect of fiber length on processing and properties of extruded wood-fiber/HDPE composites. *Journal of Applied Polymer Science* 110 (2): 1085-1092. https://doi.org/10.1002/app.28720

MILLER MP. 2005. Alleles in space: computer software for the joint analysis of inter-individual spatial and genetic information. *Journal of Heredity* 96: 722–724. doi: 10.1093/jhered/esi119

NAIK PK, ALAM A, SINGH H, GOYAL V, PARIDA S. 2010. Assessment of genetic diversity through RAPD, ISSR and AFLP markers in *Podophyllum hexandrum*: a medicinal herb from the Northwestern Himalayan region. *Physiology & Molecular Biology of Plants* 16:135–148. doi: 10.1007/s12298-010-0015-9.

NYBOM H, BARTISH IV. 2000. Effects of life history traits and sampling strategies on
genetic diversity estimates obtained with RAPD markers in plants. *Perspectives in Plant*
Ecology Evolution & Systematics 3: 93-114. *https://doi.org/10.1078/1433-8319-00006*

NYBOM HD. 2004. Comparison of different nuclear DNA markers for estimating
intraspecific genetic diversity in plants. *Molecular Ecology* 13: 1143-1155.
https://doi.org/10.1111/j.1365-294X.2004.02141.x

PAWAR GV, SINGH L, JHARIYA MK, SAHU KP. 2012. Regeneration status in relation to
antropogenic disturbance in tropical deciduous forest of Chhattisgarh. *Ecoscan* 1: 281-
286.

PEAKALL R, SMOUSE PE. 2012. GenAlEx 6.5: genetic analysis in Excel. Population
genetic software for teaching and research—an update. *Bioinformatics* 28: 2537-2539.
doi: 10.1093/bioinformatics/bts460

PERRIER X, JACQUEMOUD-COLLET JP. 2006. DARwin software http://darwin.cirad.fr/

PREVOST A, WILKINSON MJ. 1999. A new system of comparing PCR primers applied to
ISSR fingerprinting of potato cultivars. *Theoretical and Applied Genetics* 98: 107–112.
https://doi.org/10.1007/s001220051046

PRITCHARD JK, STEPHENS M, DONNELLY P. 2000. Inference of population structure
using multilocus genotype data. *Genetics* 155: 945–959.

PUROHIT SD, SHARMA P, NAGORI R. 1995. Assessment of molecular diversity and
Invitro clonal fidelity in some Indian medicinal plants. *ISHS Acta Horticulturae* 1098: 47-
60. doi: 10.17660/ActaHortic.2015.1098.4

SHAH SA, RATHOD IS, SUHAGIA BN, PANDYA SS, PARMAR VK. 2008. A simple
high performance liquid chromatography for the estimation of boswellic acids from the
market formulations containing *Boswellia serrata* extract. *Journal of Chromatography*
Science 46(8): 735-738.

SHARMA S. 1983. A census of rare and endemic flora of South East Rajasthan. In: Jain,
S.K. and Rao, R.R. (eds.), Threatened Plants of India. Botanical Survey of India ,
Howrah, pp. 630-670.

SHARMA SS, AADIL K, NEGI MS, TRIPATHI SB. 2014. Efficacy of two dominant marker systems, ISSR and TE-AFLP for assessment of genetic diversity in biodiesel species *Pongamia pinnata*. *Current Science* 106: 1576-1580.

SHARMA YK, BHANDARI KS, SRIVASTAVA A. 1985 *Boswellia serrata* resin gum: a potential internal sizing agent for paper. *Indian Forester* 111(3): 149-157.

SLATKIN M. 1987. Gene flow and the geographic structure of populations. *Science* 236: 787–792.

SPIEGELHALTER DJ, BEST NG, CARLIN BP, VAN DER LINDE A 2002. Bayesian measures of model complexity and fit. *Journal of Royal Statistical Society Series B Statistical Methodology* 64: 583-689. https://doi.org/10.1111/1467-9868.00353

SUNNICHAN VG, MOHANRAM HY, SHIVANNA KR. 2005. Reproductive biology of *Boswellia serrata*, the source of salai guggul, an important gum-resin. *Botanical Journal of Linean Society* 147 (1): 73-82. https://doi.org/10.1111/j.1095-8339.2005.00349.x

VAISHNAV V, ANSARI SA. 2018. Genetic differentiation and adaptation in teak (*Tectona grandis* L.f.) metapopulations. *Plant Molecular Biology Reporter.* doi: 10.1007/s11105-018-1101-3

VAISHNAW V, MOHAMMAD N, WALI SA, KUMAR R, TRIPATHI SB, NEGI MS, ANSARI SA. 2015. AFLP markers for analysis of genetic diversity and structure of teak (*Tectona grandis* L f) in India. *Canadian Journal of Forest Research* 44: 297-306. https://doi.org/10.1139/cjfr-2014-0279

WANG BY, SHI L, RUAN ZY, DENG J. 2011. Genetic diversity and differentiation in *Dalbergia sissoo* (Fabaceae) as revealed by RAPD. *Genetics & Molecular Research* 10 (1): 114-120.

WEEKS A, ZAPATA F, PELL SK, DALY DC, MITCHELL JD, FINE PVA. 2014. To move or to evolve: contrasting patterns of intercontinental connectivity and climatic niche evolution in "Terbinthaceae" (Anacardeacea and Burseraceae). *Frontier Genetics*. doi: 10.3389/fgene.2014.0 0409

WRIGHT S. 1969. Evolution and the Genetics of Populations, vol. 2: The Theory of Gene
Frequencies. University of Chicago.

YEH FC, YANG RC, BOYLE T. 1999. POPGENE 32-version 1.32. Software.
http://www.ualberta.ca/~fyeh/fyeh/.

ZIETKIEWIEZ E, RAFALSKI A, LABUDA D. 1994. Genome fingerprinting by simple
sequence repeat (SSR)-anchored polymerase chain reaction amplification. *Genomics* 20
:176–183.

ZOBEL B, JETT JB. 1995. Genetics of wood production. Springer-Verlag, New York, pp
128.

Table Captions

Table 1 Geo-climatic conditions of the three regions of natural distribution of *B. serrata* Roxb. populations sampled for the investigation

Table 2 Variability in wood trait parameters of three sub-populations of *B. serrata* Roxb.

Table 3 Genetic informativeness of RAPD and ISSR marker systems applied on present investigation

Table 4 Population genetics parameters for 60 genotypes sampled from three sub-populations of *B. serrata* Roxb.

Table 1. Geo-climatic conditions of the three regions of natural distribution of *B. serrata* Roxb. populations sampled for the investigation

Regions	Agro-climatic zones	GPS coordinate ranges		Altitude (m)	Mean annual temperature (^{0}C)		Annual Precipitation (mm)
		Latitude (N)	Longitude (E)		Min	Max	
Dhamtari (DT)	Chhattisgarh Plain	20.37861-20.40511	81.96636-81.98122	445.80 ±5.44	21.60 ±5.22	33.02 ±3.56	1488±0
Narayanpur (NP)	Bastar Plateau	19.71365-19.71586	81.19447-81.19836	595.60 ±21.39	20.80 ±5.22	32.40 ±3.36	1628±0
Sarguja (SG)	Northern Hills	23.39300-23.39446	83.46277-83.46783	619.75 ±7.15	19.02 ±6.35	31.02 ±4.07	1670±0

± - Standard deviation (SD)

Table 2. Variability in wood trait parameters of three sub-populations of *B. serrata* Roxb.

Regions	GBH (m)	WFL (mm)	WFW (mm)
Dhamtari (DT)	0.99 ±0.25	0.919 ±0.097	0.025 ±0.001
Narayanpur (NP)	1.07 ±0.22	0.975 ±0.095	0.026 ±0.002
Sarguja (SG)	1.40 ±0.25	1.009 ±0.120	0.024 ±0.003
Average	1.15 ±0.30	0.968 ±0.11	0.025 ±0.002
CV (%)	26.01	11.36	8.75

GBH- girth at breast height, WFL- wood fiber length, WFW- wood fiber width, CV- coefficient of variation, ±- standard deviation (SD)

Table 3. Genetic informativeness of RAPD and ISSR markers

Marker System	Primers	Sequence (5' – 3')	N	AF	GD	PIC	RP
RAPD	OPA1	CAGGCCCTTC	12	0.77	0.32	0.26	6.10
	OPA2	TGCCGAGCTG	13	0.72	0.37	0.29	10.80
	OPY1	GTGGCATCTC	10	0.80	0.27	0.22	3.97
	OPY2	CATCGCCGCA	12	0.87	0.19	0.16	3.70
	OPP3	CTGATACGCC	9	0.87	0.21	0.18	2.97
	Average		11.20	0.81	0.27	0.22	5.51
ISSR	UBC840	GAGAGAGAGAGAGAGAAT	9	0.67	0.42	0.33	9.47
	UBC830	TGTGTGTGTGTGTGTGG	6	0.74	0.36	0.29	6.50
	UBC822	TCTCTCTCTCTCTCTCA	8	0.74	0.37	0.30	6.50
	UBC845	CTCTCTCTCTCTCTCTGG	8	0.84	0.24	0.20	6.17
	UBC836	AGAGAGAGAGAGAGAGTA	11	0.80	0.28	0.23	4.83
	Average		8.40	0.76	0.33	0.27	6.69

N- number of amplified loci, AF- allele frequency, GD- gene diversity, PIC- polymorphic information content, RP - resolving power

Table 4. Population genetics parameters for 60 genotypes sampled from three sub-populations of *B. serrata* Roxb.

Regions	P%	GD	I	Hs
Dhamtari (DT)	63.27	0.20 ±0.20	0.30 ±0.28	0.20 ±0.01
Narayanpur (NP)	57.14	0.19 ±0.19	0.28 ±0.28	0.19 ±0.01
Sarguja (SG)	55.10	0.14 ±0.16	0.22 ±0.24	0.17 ±0.01
Overall*	100	0.26 ±0.17	0.40 ±0.22	0.28 ±0.01

±- SD, P% - polymorphism%, GD - Nei (1973) gene diversity, I - Shannon's information index, Hs - panmictic heterozygosity resulted by Hickory, * considering all 60 genotypes as a group

Figures

Figure 1 Three regions i.e. Sarguja (SG), Dhamatri (DT) and Narayanpur (NP) of natural
distribution of *B. serrata* Roxb from central Indian region (Chhattisgarh) were sampled for
the investigation. Every location represents distinct agro-climatic zone namely Northern hills
(SG), Chhattisgarh plain (DT) and Bastar plateau (NP). An inset view of altitudinal variation
is given in the box and size differences among the location's legend dot depicts differences in
gene diversity of the species estimated employing the dominant markers

Figure 2 A Jaccard's similarity coefficient based radial dendrograph of 60 accessions from three natural populations i.e. Sarguja (SG), Dhamtari (DT), and Narayanpur (NP) of *B. serrata* Roxb. Bootstrap values (>60) are shown on nodes of accessions

513 **Figure 3** The principal coordinate analysis, showing closeness of genotypes of *B. serrata*
514 Roxb sampled from Dhamtari (DT) and Sarguja (SG) and the genotypes from Narayanpur
515 (NP) cluster distinctly, in two coordinates covering 68.97% of variation

Figure 4 The Bayesian clustering resulted by program STRUCTURE depicted two cryptic
populations (K) in 60 genotypes of *B. serrata* Roxb sampled from three locations of its
natural distribution. Each genotype is presented by a separate colored bar with corresponding
number. Green and Red color represents two different cryptic populations.