

1 **Title:** Gut Microbiota in male patients with chronic traumatic complete spinal cord

2 injury

3 **Running title:** Gut microbiota in spinal cord injury patients

4 **Authors:**

5 Chao Zhang,<sup>1,2,3,4,5</sup> Wenhao Zhang,<sup>1,2,3,4,5</sup> Jie Zhang<sup>1,2,3,4,5</sup>, Yingli Jing,<sup>1,3,4,5,6</sup>

6 Mingliang Yang,<sup>1,2,3,4,5</sup> Liangjie Du,<sup>1,2,3,4,5</sup> Feng Gao<sup>1,2,3,4,5</sup>, Huiming Gong,<sup>1,2,3,4,5</sup>, Liang

7 Chen<sup>1,2,3,4,5</sup>, Jun Li,<sup>1,2,3,4,5</sup> Hongwei Liu<sup>1,2,3,4,5</sup>, Chuan Qin<sup>1,2,3,4,5</sup>, Yanmei Jia<sup>1,2,3,4,5</sup>, Jiali

8 Qiao<sup>1,2,3,4,5</sup>, Bo Wei<sup>1,3,4,5,7</sup>, Yan Yu<sup>1,3,4,5,6</sup>, Hongjun Zhou<sup>1,3,4,5,7</sup>, Zhizhong Liu<sup>1,3,4,5,8</sup> Degang Yang,

9 <sup>1,2,3,4,5\*</sup> Jianjun Li,<sup>1,2,3,4,5\*</sup>

10 <sup>1</sup>School of Rehabilitation Medicine, Capital Medical University, Beijing 100068,

11 China

12 <sup>2</sup>Department of Spinal and Neural Function Reconstruction, China Rehabilitation

13 Research Center, Beijing 100068, China

14 <sup>3</sup>Center of Neural Injury and Repair, Beijing Institute for Brain Disorders,

15 Beijing 100068, China

16 <sup>4</sup>China Rehabilitation Science Institute, Beijing 100068, China

17 <sup>5</sup>Beijing Key Laboratory of Neural Injury and Rehabilitation, Beijing 100068,

18 China

19 <sup>6</sup>Institute of Rehabilitation medicine, China Rehabilitation Research Center,

20 Beijing 100068, China

21 <sup>7</sup>Department of Spinal Cord Injury Rehabilitation, China Rehabilitation Research

22 Center, Beijing 100068, China

23 <sup>8</sup>Laboratory medicine, China Rehabilitation Research Center, Beijing 100068,  
24 China

25 **\* Corresponding authors:**

26 1 Jianjun Li and 2 Degang Yang.

27 School of Rehabilitation Medicine, Capital Medical University, No. 10 Jiaomen  
28 North Road, Fengtai District, 100068 Beijing, China

29 13718331416@163.com; yzydg2006@126.com.

30 **Author Disclosure Statement**

31 No competing interests exist.

32 **Abstract:**

33 This study examined the diversity and structure of gut microbiota in healthy adults  
34 and chronic traumatic complete spinal cord injury (SCI) patients, documented  
35 neurogenic bowel management of SCI patients. The V3-V4 region of 16S rRNA gene  
36 from DNA of 91 fecal samples of 48 healthy and 43 diseased subjects was amplified  
37 and sequenced. There was difference in gut microbiota between healthy adult males  
38 and females. Neurogenic bowel dysfunction (NBD) was common in patients with  
39 chronic traumatic complete SCI, patients with quadriplegia have longer time to  
40 defecate than paraplegic patients, with higher NBD scores and heavier neurogenic  
41 bowel symptoms. Gut microbiota dysbiosis existed in SCI patients. The abundance of  
42 Veillonellaceae and Prevotellaceae increased while Bacteroidaceae and Bacteroides  
43 decreased in SCI group. The abundance of Bacteroidaceae, Bacteroides in  
44 quadriplegia group and Acidaminococcaceae, Blautia in paraplegia group were

45 significant high than the health male group. Serum biomarkers GLU, HDL, CR and  
46 NBD symptoms defecation time, COURSE had significant correlation with microbial  
47 community structure. This study presents a comprehensive landscape of gut  
48 microbiota in adult male patients with chronic traumatic complete SCI and documents  
49 their neurogenic bowel management. The gut microbiota dysbiosis of SCI patients  
50 was correlation with serum biomarkers and NBD symptoms.

51 **IMPORTANCE:** Neurogenic bowel dysfunction is a major physical and  
52 psychological problem in patients with spinal cord injury, which can seriously affect  
53 the quality life of them. Gut dysbiosis are highly likely to occur in spinal cord injury  
54 patients There are few studies on intestinal microecology after spinal cord injury, and  
55 the clinical studies are fewer. It is importance to document their neurogenic bowel  
56 management and present a landscape of gut microbiota in them. We found the gut  
57 microbiota dysbiosis of spinal cord injury patients was correlation with serum  
58 biomarkers and neurogenic bowel dysfunction symptoms. These results may have  
59 implications in the next study about metagenomics and precision treatment of  
60 neurogenic bowel dysfunction in spinal cord injury patients.

61 **Keywords:** gut microbiota dysbiosis; chronic traumatic complete SCI; neurogenic  
62 bowel management, NBD symptoms, serum biomarkers;

63 **Running title:** Gut microbiota in spinal cord injury patients

64 **Introduction:**

65 After complete spinal cord injury, the loss of descending control over sympathetic  
66 preganglionic neurons causes autonomic reflex circuitry to become dysfunctional

67 creating pathology including autonomic dysreflexia and SCI-immune depression  
68 syndrome (1,2,3,4,5), it causes an autonomic imbalance in the gastrointestinal tract,  
69 which leads to deficits in colonic motility, mucosal secretions, and vascular tone (6,7).  
70 The early survival rate of such patients has been significantly improved, but the  
71 quality of life of such patients is still not satisfactory. Among them, neurogenic bowel  
72 dysfunction (NBD) is a major physical and psychological problem in patients with  
73 SCI, which can seriously affect the quality life of patients. The two main  
74 manifestations of NBD are constipation and fecal incontinence, with the prevalence of  
75 constipation in these patients reported to be 40–58%, and fecal incontinence from 2 to  
76 61% (8,9,10,11). Because of these problems, patients with chronic SCI tend to spend  
77 more time in the toilet while evacuating their bowels, use suppositories, laxatives and  
78 supplemental dietary fiber more frequently to improve bowel evacuation and require  
79 manual removal of feces much more frequently when compared with their matched  
80 control population (12,13,14,15). One of the aims of our study was to document  
81 neurogenic bowel management of chronic traumatic completed SCI male patients in  
82 our center.

83 Human intestinal tract is colonized by thousands of different genera of bacterial  
84 species whose number and genetic content exceed that of the host by a factor of ten  
85 and 150-fold, respectively (16). That is critical for normal digestion, nutrient  
86 absorption, and the development, metabolism, and function of cells throughout the  
87 body (17,18). Recent studies have shown that an imbalance of the normal gut  
88 microbiota (dysbiosis) is associated with inflammatory bowel diseases (19), irritable



89 bowel syndrome and some other diseases (20,21).

90 Elin O et al reported that sex hormones affected the gut microbiota composition in  
91 male and female mice in a controlled environment; Francesca Borgo et al reported  
92 that body mass index and gender affect microbial flora in different parts of the gut  
93 (22,23). One of the aims of this study was to explore whether there is a difference of  
94 gut microbiota in healthy adult males and females.

95 Common causes of gut dysbiosis include antibiotic use, prolonged stress, and  
96 gastrointestinal dysfunction (17,24,25). Because most patients with acute complete  
97 SCI have changed the intestinal transit time and destructed the intestinal mucosal  
98 function barrier after injury, the displacement of the intestinal flora making the  
99 intestines to be the largest "endotoxin pool" in the human body. The use of antibiotics  
100 must affect healthy intestinal micro-ecological systems (26,27,28,29). Therefore, gut  
101 dysbiosis are highly likely to occur in SCI.

102 There are few studies on intestinal microecology after SCI in clinical studies.  
103 Kigerl KA et al have shown that traumatic SCI can cause intestinal disorders, and that  
104 dysbiosis can impair functional recovery through stool samples from traumatic SCI  
105 mice (30). Bilgi Gungor et al reported a clinical study of 30 patients with SCI, showed  
106 that the number of butyrate communities in patients with SCI was significantly lower  
107 than that in the normal population (29). More researches are needed to determine  
108 whether intestinal dysbiosis after SCI changes in a range of clinically relevant  
109 variables (31).

110 The study of this article is to explore the difference of healthy adult males and

111 females in gut microbiota, document neurogenic bowel management of chronic  
112 traumatic complete SCI male patients in our center; To investigate the comparative  
113 analysis of intestinal gut microbiota in chronic traumatic complete SCI male patients  
114 and healthy males. Exploring the association between intestinal microbiota with  
115 serum biomarkers and neurogenic bowel symptoms.

## 116 **Results:**

### 117 **Baseline characteristics of the samples in the health male and female groups**

118 The mean age for 23 healthy adult males and 25 females was  $40\pm 9$  years and  $37\pm 8$   
119 years (18–60 years), there was no statistically significant differences (one-way  
120 ANOVA,  $p=0.255$ ). The BMI in males was significantly high ( $24.8\pm 2.777$ ) than  
121 females ( $22.8\pm 2.763$ ), (one-way analysis of variance,  $p = 0.015$ ).

### 122 **Diversity and taxonomic analysis in the health male and female groups**

123 16S rRNA gene sequences were generated using Illumina's MiSeq platform.  
124 Briefly, a total of 1010832 sequences were obtained. Reads were clustered in OTUs at  
125 97% of identity. The rarefaction curves showed clear asymptotes and the Good's  
126 coverage for the observed OTUs was 99.46%, which together indicate a  
127 near-complete sampling of community. No significant difference in either OTU  
128 abundance or OTU diversity index was observed between the male and female  
129 populations (Fig. 1A and Supplementary File 1). The discrete case of sample points  
130 distribution in PLS-DA on genus level showed differences in the composition of the  
131 gut microbiota between the two groups (Fig. 1B) .

132 STAMP analysis indicates there were 15 OTUs showed a significant difference

133 (P<0.05) among two groups (Welch's t-test). There were 4 of top 15 genus showed  
134 a significant difference (P<0.05) among two groups (Welch's t-test). The abundance  
135 of Megamonas and Dialister in male group were significant high than the female  
136 group (P< 0.01, P< 0.01, Mann-Whitney U test); the abundance of Bacteroides and  
137 Phascolarctobacterium in female group were significant high than the male group (P<  
138 0.01, P< 0.01, Mann-Whitney U test) (Fig.1C). We can find that there was a  
139 difference in fecal flora between healthy adult males and females.

#### 140 **Characteristics and neurogenic bowel management of male patients with chronic** 141 **traumatic complete SCI**

142 In all, 43 patients with chronic SCI fulfilling the enrollment criteria were  
143 interviewed and completed the survey form (Table 1). The causes of injury were  
144 traffic accidents (37.2%), bruised by heavy object (20.9%), fall from height (20.9%),  
145 in that order. The mean score of NBD was 10.02±5.11. The mean defecation time was  
146 35.33±16.766 minutes. Most patients (60.5%) took bowel care not daily but more than  
147 twice every week, the others (39.5%) frequency of bowel care were once daily. Main  
148 techniques for fecal evacuation was suppository (88.4%), manual evacuation  
149 (23.3%), digital stimulation (16.3%), spontaneous (4.7%), in that order.  
150 Supplementary interventions for fecal evacuation were abdominal massage (58.1%),  
151 digital anus-rectal stimulation (48.8%), digital evacuation (9.4%), taking cathartic  
152 drug (9.4%).

153 More than a half patients bowel care time was in afternoon (62.8%), the other  
154 patients' bowel care time was in evening (20.9%), and in morning (9.4%). The

155 location of bowel care was bed (44.2%), potty chair (37.2%) and toilet seat (18.8%).  
156 53.5% patients need all help during the defecation time, 25.6% patients need partial  
157 help, 20.9% patients can defecation independent. 62.8% patients had an abdominal  
158 discomfort symptom, 67.4% patients had a constipation symptom, 74.4% patients had  
159 a bloating symptom, 88.4% patients had flatus incontinence. The most common top 3  
160 complications that patients wanted to solved were neurogenic bowel  
161 dysfunction(100%), neurogenic bladder(83.7%), sexual dysfunction(44.2%).

162 The quadriplegia SCI patients had a significant high BMI ( $23.586\pm 3.35$ ) than  
163 paraplegia SCI patients ( $22.697\pm 2.31$ ) ( $P<0.001$ ). There were statistical differences  
164 between the two groups in HDL、UREA and CRP ( $P<0.001$ )( Table3). Compared  
165 with paraplegia SCI patients, the quadriplegia SCI patients had longer defecation time,  
166 higher NBD score, lower defecation frequency, need more supplementary  
167 interventions to complete the bowel care.

168 Most of the defecation locations in the quadriplegia SCI patients were in the bed.  
169 Almost all the quadriplegia SCI patients require total help to complete the bowel care,  
170 but most paraplegia SCI patients could finish the bowel care independently or only  
171 need partially help. Most of SCI patients have abdominal discomfort such as  
172 constipation, bloating, and flatus incontinence. More than half of the patients in the  
173 two groups have a serious impact on lifestyle, and the most common complication  
174 they want to resolve is NBD.

175 **Composition of the gut microbiome of health male group and male chronic**  
176 **traumatic complete SCI groups.**

177 To exclude the effect of gender on gut microbiota results, we selected 23 male  
178 healthy individuals and 43 male patients with spinal cord injury to perform a  
179 comparative analysis. Demographics and serum biomarkers between male healthy and  
180 patients with chronic traumatic completed SCI were showed in Table2.

181 Briefly, a total of 2247802 sequences were obtained. Reads were clustered in  
182 OTUs at 97% of identity. The rarefaction curves showed clear asymptotes and the  
183 Good's coverage for the observed OTUs was 99.88%, which together indicate a  
184 near-complete sampling of community. 798 OTUs are recognized in total. No  
185 significant difference in OTU abundance (ace, chao1 index) was observed between  
186 health male and SCI populations. In genus level, OTU diversity index Simpson  
187 showed a significant difference between two groups ( $P=0.03635$ ) (Fig.2A). This  
188 indicates a decrease in intestinal flora diversity in patients with SCI.

189 The PCA on phylum level and the NIMDS on OTU and genus level of  
190 beta-diversity analysis showed there were significant differences in bacterial  
191 community composition between two groups. ANOSIM/Adonis revealed significant  
192 differences in the structure of gut microbiota among the two groups ( $p<0.05$ )  
193 (Supplementary File 2). PLS-DA revealed that there were significant differences in  
194 bacterial community composition between two groups on OTU, phylum and genus  
195 level ( $p<0.05$ ) (Fig.2B).

196 STAMP analysis indicates there were 9 of top 15 genus showed a significant  
197 difference ( $P<0.05$ ) among two groups (Welch's t-test). The abundance of  
198 *Megamonas*, *Prevotella\_9*, (*Eubacterium*)\_rectale\_group, *Dialister*, *Subdoligranulum*

199 in male group were significant high than the SCI group ( $p < 0.05$ , Mann-Whitney U  
200 test); the abundance of Bacteroides, Blautia, Lachnoclostridium, Escherichia-Shigella  
201 in SCI group were significant high than the male group ( $P < 0.05$ , Mann-Whitney U  
202 test) (Fig.2C, D). By LEfSe analysis (LDA threshold of 2), it was found that  
203 Veillonellaceae and Prevotellaceae were significantly enriched in SCI group compared  
204 with Bacteroidaceae and Bacteroides enriched in healthy male group.

205 According to NBD constipation symptom, we divided the SCI patients into  
206 constipation group and without constipation group. STAMP analysis showed a  
207 significant difference ( $P < 0.05$ ) among two groups (Welch's t-test) in  
208 Bifidobacterium on Genus level (Fig.3A). We also divided the SCI patients into  
209 bloating group and without bloating group according to bloating symptom, STAMP  
210 analysis showed Megamonas had a significant high ( $P < 0.05$ ) in bloating group and  
211 Alistipes had a significant high ( $P < 0.05$ ) in without bloating group on genus level  
212 (Fig.3B).

213 The selected environmental factors: BMI, AGE, ALT, AST, GLU, TG, TCHO,  
214 HDL, LDL, UREA, CR, UA for RDA analysis. One-way ANOVA showed statistically  
215 significant differences in BMI, GLU, TCHO, LDL and UA between the two groups ( $P$   
216  $< 0.05$ ). RDA/CCA showed that GLU ( $p = 0.017$ ,  $r^2 = 0.1315$ ), HDL  
217 ( $p = 0.028$ ,  $r^2 = 0.1121$ ), CR ( $p = 0.017$ ,  $r^2 = 0.1349$ ) significantly affected bacterial  
218 composition in phylum level; In top 20 genus, BMI ( $p = 0.04$ ,  $r^2 = 0.0971$ ), GLU  
219 ( $p = 0.044$ ,  $r^2 = 0.108$ ) and HDL ( $p = 0.001$ ,  $r^2 = 0.3044$ ) significantly affected bacterial  
220 composition. We can found that Serum biomarkers GLU, HDL and CR had significant

221 correlation with microbial community structure ( $p < 0.05$ ).

222 Correlation heatmap analysis of different environmental factors on the  
223 community composition of two groups showed that Proteobacteria was positively  
224 correlated with UA (Pearson  $r = 0.26$ ,  $p = 0.035$ ) ; Cyanobacteria were positively  
225 correlated with AST (Pearson  $r = 0.355$ ,  $p = 0.003$ ) ; Fusobacteria were negatively  
226 correlated with AGE (Pearson  $r = -0.342$ ,  $p = 0.005$ ) in phylum level(Fig.4A). In top  
227 20 genus, Bacteroides was negative correlated with HDL (Pearson  $r = -0.418$ ,  
228  $p < 0.001$ ) ; Megamonas was negatively correlated with GLU (Pearson  $r = -0.513$ ,  
229  $p < 0.001$ ) ; Blautia was positively correlated with UA (Pearson  $r = 0.274$ ,  $p = 0.026$ ) ;  
230 Dialister was negatively correlated with UA, LDL, TG and TCHO (Pearson  $r = -0.32$ ,  
231  $P = 0.009$ ;  $r = -0.289$   $P = 0.019$ ;  $r = -0.258$ ,  $P = 0.037$ ;  $r = -0.303$ ,  $P = 0.013$  respectively.)  
232 (Fig.4B and Supplementary File 3-4).

### 233 **Comparison of the gut microbiome in quadriplegia and paraplegic groups**

234 We divided the 43 SCI patients into 20 quadriplegia group and 23 paraplegic group, the  
235 characteristics and neurogenic bowel management were showed in Table 1 and 3. We found  
236 that the defecation time of quadriplegia patients ( $41.789 \pm 19.29$  minutes) was significant high  
237 than paraplegic patients ( $30 \pm 13.94$  minutes) ( $P = 0.026$ ).

238 The rarefaction curves showed clear asymptotes and the Good's coverage for the  
239 observed OTUs was 99.88%, which together indicate a near-complete sampling of  
240 community (Fig.5A). No significant difference in OTU abundance (ace, chao index)  
241 was observed between the three groups. Significant difference in genus abundance  
242 chao index was observed between quadriplegia group and paraplegic group

243 (P=0.02922), healthy male group and paraplegic group (P=0.02919), those indicates  
244 a difference in community richness in the two groups (Fig.5B). The Simpson index of  
245 health male group showed a significant high than paraplegic group (P=0.04094) in  
246 genus level, this indicates a decrease in intestinal flora diversity in patients with  
247 paraplegic spinal cord injury (Fig.5C and Supplementary File 5-6) .

248 ANOSIM/Adonis of beta-diversity analysis revealed significant differences in  
249 the structure of gut microbiota among the three groups (p=0.001, r<sup>2</sup>=0.233) in phylum  
250 level (Fig.5D and Supplementary File 7). PLS-DA revealed that there were significant  
251 differences in bacterial community composition between three groups on OTUs,  
252 phylum and genus level (Fig.5E).

253 STAMP analysis indicates there were 8 OTUs showed a significant difference  
254 (P<0.05) among three groups (Welch's t-test) in top 15 OTUs. There were 8 of top  
255 15 genus showed a significant difference (P<0.05) among three groups (Welch's  
256 t-test). The abundance of Firmicutes in paraplegic group and healthy male group were  
257 significant high than the quadriplegia group (P= 0.0251 和 P= 0.0185, One-way  
258 ANOVA test) . In top 15 genus, the abundance of Bacteroides, Faecalibacterium,  
259 Blautia, Prevotella\_9, Phascolarctobacterium, Parabacteroides, (Eubacterium)\_rectale  
260 showed a significant difference between the three groups (P< 0.05, One-way ANOVA  
261 test) (Fig.6).

262 The selected 16 environmental factors were: BMI, ALT, AST, GLU, TG, TCHO,  
263 HDL, LDL, UREA, CR, UA, AGE, COURSE, CRP, NBD-score, Defecation time for  
264 RDA analysis in quadriplegia group and paraplegic group. One-way ANOVA showed



265 statistically significant differences in BMI, HDL, UREA, APOA1 and defecation time  
266 between the two groups (Table3).

267 RDA/CCA showed that GLU ( $p=0.014$ ,  $r^2=0.1969$ ), HDL ( $p=0.009$ ,  $r^2=0.2274$ ),  
268 CR ( $p=0.006$ ,  $r^2=0.2306$ ), significantly affected bacterial composition in phylum  
269 level; In OTU level, TG ( $p=0.042$ ,  $r^2=0.2192$ ), CR ( $p=0.007$ ,  $r^2=0.2388$ ), Defecation  
270 time ( $p=0.022$ ,  $r^2=0.2009$ ) significantly affected bacterial composition; In genus level,  
271 HDL ( $p=0.001$ ,  $r^2=0.4675$ ), CR ( $p=0.001$ ,  $r^2=0.3209$ ) significantly affected bacterial  
272 composition.

273 Correlation heatmap analysis of different environmental factors on the  
274 community composition of quadriplegia and paraplegic groups showed that Alistipes  
275 were negatively correlated with defecation time (Pearson  $r=-0.363$ ,  $p=0.017$ ),  
276 negatively correlated with course (Pearson  $r=-0.375$ ,  $p=0.013$ ). In phylum level:  
277 Firmicutes was negatively correlated with CRP (Pearson  $r=-0.491$ ,  $p=0.001$ ),  
278 positively correlated with HDL (Pearson  $r=0.419$ ,  $p=0.005$ ). At the genus and OUT  
279 levels, the results indicated that the HDL, LDL, CR, UA, AGE have an effect on the  
280 intestinal microbiota of two groups (Fig7, Supplementary File 8-9).

## 281 **Discussion:**

282 In this study, after verified the differences in gut microbiota between healthy adult  
283 males and females, we compared the gut microbiome between healthy adult males and  
284 male patients with chronic traumatic complete SCI. The neurogenic bowel  
285 management of SCI patients in our center were firstly reported through cross-sectional  
286 interviews. We try to explore the association between gut microbiota and

287 environmental factors in quadriplegia and paraplegic groups; analyse the correlation  
288 between gut microbiota and neurogenic bowel symptoms. The results of neurogenic  
289 bowel symptoms in SCI patients were related to some gut microbiota and it may help  
290 explain the potential link between gut dysbiosis and NBD symptoms in SCI patients.

291 Acute traumatic SCI (ASCI) used to appear mostly in adults (21—69 years), the  
292 causes of injury were fall from height (37.5%), traffic accidents (26.9%) (32,33). This  
293 was also consistent with traffic accidents (37.2%), fall from height (20.9%) in this  
294 study. The average age of SCI patients in this study was 39.9 years old, which was in  
295 the prime of life and was more susceptible to accidental injuries such as high-energy  
296 trauma. After chronic course, the most common complication those patients hope to  
297 resolve was NBD. Gut microbiota may be a potential method to improve this problem.

298 Li et al has reported the male/female ratio of ASCI was 3.1/1(32), most of the  
299 patients with chronic traumatic complete SCI admitted to our center were males, so  
300 we chose male patients as our research objects. Elin O et al had reported the different  
301 in gut microbiota between male and female before (22-23). A recent study by Haro et  
302 al highlighted differences between men and women in the luminal microbial  
303 population. Their results suggest that these differences may be influenced by the grade  
304 of obesity (34). The abundance of Bacteroides was higher in females than in males  
305 was same with Haro's research (34). In fact, gut microbiota composition seems to be  
306 more influenced by ambient and dietary cues than by genetic factors and  
307 inter-individual heterogeneity (35,36). We compared the gut microbiota of healthy  
308 males and male SCI patients to eliminate the gender impact.

309 Julia et al illustrated the practice and outcomes of bowel care in the community of  
310 individuals with SCI in Malaysia (37). R Yasmeeen had reported that 43 of 50 adult  
311 patients with SCI in Pakistan gave a history of occasional or regular fecal  
312 incontinence (38). The prevalence of NBD in SCI patients was 80% in previous study,  
313 97.3% of motor complete SCI patients had chronic NBD complaints in their study  
314 (10), which was similar with patients had a constipation in our study. The patients in  
315 our study had to spend much time on defecation in their long course of SCI to deal  
316 with NBD.

317 Injury level has been shown not to be related to gastro-intestinal complaints in  
318 SCI patients detected no significant relation between gastro-intestinal symptom  
319 prevalence and SCI level in their study (39,40,41). Most patients with quadriplegia  
320 have no active exercise capacity but paraplegic patients can complete all upper limb  
321 movements. The autonomic nervous system that supports the gastrointestinal tract in  
322 quadriplegia SCI patients remains relatively intact and has less effect on the function  
323 of the gastrointestinal tract. Patients with paraplegia had damaged the sympathetic  
324 center or defecation center would had a greater impact on intestinal function. This  
325 may explain the different between the two groups.

326 Surveys among the SCI population often rank colorectal, bladder and sexual  
327 dysfunction as significant obstacles and prioritize recovery of bowel function above  
328 the ability to walk (42,43,44). In this study, the most common top 3 complications that  
329 patients wanted to solved were NBD, neurogenic bladder, sexual dysfunction, the gut  
330 microbiota may be a potential solution strategy.

331 Rajilić-Stojanović observed an increase in the phylum Bacteroidetes, which was  
332 reflected by increased Bacteroides at the genus level. The phylum Bacteroidetes  
333 encompasses a diverse and abundant group of gram-negative commensal bacteria in  
334 the gut (45). The major outer membrane component of gram-negative bacteria is  
335 lipopolysaccharide (LPS), which is capable of triggering systemic inflammation and  
336 the release of pro-inflammatory cytokines after translocation from the gut to systemic  
337 circulation (46). In our study, we found the Bacteroides was significant high in SCI  
338 group and negatively correlated with HDL, especially enriched in quadriplegia group.  
339 HDL levels are positively correlated with amount of exercise, and lack of exercise can  
340 result in lower HDL levels (47). Reduced exercise in quadriplegia patients made the  
341 lowers HDL levels, and the Bacteroides were increased in quadriplegia patients.  
342 Those indicated that Bacteroides may be a harmful flora and was associated with the  
343 exercise v and serum HDL levels.

344 Nicholas M. Vogt had reported that Dialister showed the strongest correlations in  
345 non-demented participants, with greater abundance of these bacteria associated with  
346 less Alzheimer's disease (AD) pathology, suggesting these bacterial taxa may be  
347 protective against development or progression of AD pathology (48). In our study, the  
348 Dialister was significant high in health males, and these bacteria were negatively  
349 correlated with UA, LDL, TG and TCHO. Those elevated serum markers represent  
350 high blood lipids, which are harmful to the health. We found that the decreased  
351 Dialister in SCI patients may aggravated the symptoms of the NBD.

352 Ming et al reported the positive association between bean consumption and the

353 Megamonas genus discovered in their study may implicate Megamonas as a beneficial  
354 microbe(49).The abundance of Megamonas was decreased in SCI group and was  
355 negatively correct with GLU, positively with NBD scores, implicate that Megamonas  
356 may have a positive effect on the body in terms of carbohydrate metabolism. The  
357 decreased Megamonas exacerbates NBD symptoms. We found that in bloating group  
358 the relative abundance of Megamonas had a significant high than without bloating  
359 group, this may be due to the fact that some carbohydrates in food cannot be digested  
360 and absorbed by intestinal digestive enzymes, but they can be metabolized by  
361 Megamonas in the large intestine, that producing gas and causing bloating,  
362 exacerbates NBD symptoms.

363 Genera of Alistipes were already reported to be altered in irritable bowel  
364 syndrome, animal-based diet or vegetables consumption (50,51,52). Alistipes were  
365 associated with the phenotype of frequently recurrent abdominal pain (50). We found  
366 the relative abundance of Alistipes was significant decreased in bloating group, and  
367 Alistipes were negatively correlated with defecation time, this may be a factor that  
368 affect the defecation time between quadriplegia and paraplegic group because the  
369 abundance of Alistipes and the defecation time in quadriplegia group were significant  
370 high than the paraplegic group. But there is still a big gap in knowledge to explain  
371 biochemical and functional roles of Alistipes.

372 Members of the Bifdobacterium genus, are an important bacterial inhabitant of  
373 the human gut across the lifespan, and their beneficial health effects have been  
374 well-documented (53,54). But certain species of Bifdobacterium are associated with

375 decreased intestinal permeability. In our study the abundance of Bifidobacterium and  
376 Bacteroides were increased in SCI group, it may be because the increased bacterial  
377 translocation effect of Bacteroides played a more important role than Bifidobacterium.  
378 The relative abundance of Bifidobacterium was significantly high in constipation group  
379 in our study, we thought in the chronic course of constipation symptom, the patients  
380 may have used the probiotics which including Bifidobacterium. More investigation is  
381 needed to determine the interaction of these bacterial.

382 We also examined the association between serum biomarkers and the gut  
383 microbiota. Prevotella is considered a beneficial microbe, but it is also linked with  
384 chronic inflammatory conditions (55,56,57,58). Our study found a decreased  
385 Prevotella level in SCI group and negatively correlated with GLU, which implicates that  
386 Prevotella has a positive effect on the body in terms of carbohydrate metabolism  
387 supporting Prevotella as a beneficial microbe.

388 A strong point of this study was the inclusion of only complete SCI male patients.  
389 This approach excluded probable confounding effects of gender and residual nerves  
390 on gut functions, and therefore other incomplete injuries were not included in this  
391 study. Individual diet-associated flora differences could not be determined and  
392 remains as a major weakness of this study. The continuing analyses of genomic and  
393 metagenomic changes in gut microbiota will allow scientists to map the dynamic  
394 patterns of dysbiosis caused by SCI.

395 Only male SCI patients were enrolled in our study, and future studies should  
396 include female patients to identify gender disparities. Further work, including animal

397 experiments and longitudinal human studies, will be needed to determine the  
398 cause-effect relationship between gut microbiota and SCI. Determining the role of gut  
399 microbiota in the progression or maintenance of SCI may lead to novel interventional  
400 approaches that alter or restore healthy gut bacterial composition, or identification of  
401 microbial metabolites that are protective against SCI.

402 In conclusion, this study presents a comprehensive landscape of gut microbiota in  
403 adult male patients with traumatic complete SCI and documents their neurogenic  
404 bowel management. We found a difference in fecal flora between healthy adult males  
405 and females; Dysbiosis of SCI patients was correlation with Serum biomarkers and  
406 NBD symptoms.

## 407 **Materials and methods:**

### 408 **Ethics statement**

409 Approval of hospital ethics committee was obtained before commencing the study.

### 410 **Patients and controls**

411 A total of 43 chronic traumatic complete SCI male patients (20 with quadriplegia  
412 and 23 with paraplegia) in our center from March 2017 to October 2017 were enrolled  
413 to face-to-face clinical questionnaire survey. Signed informed consent before the  
414 assessment, and use " International Spinal Cord Injury Core Data Set ", " International  
415 bowel function basic spinal cord injury data set " and " International bowel function  
416 extended spinal cord injury data set " to get the NBD symptoms dates (59,60,61).

417 Patients were included if they met the following criteria: 1) neurologically  
418 complete SCI (ASIA grade A) occurring 6 or more months prior to study, 2) 18-60

419 years of age, 3) traumatic spinal cord injury, 4) male patients. The exclusion criteria: 1)  
420 patients who can not cooperate for questionnaire survey, 2) with a history of antibiotic  
421 use in the first month before enrollment, 3) patients with diabetes, gastrointestinal  
422 system diseases, multiple sclerosis, and immune metabolic diseases.

423 A total of 43 SCI patients and 48 healthy adults (23 males and 25 females) were  
424 enrolled in to collect clinical dates of the subjects and fresh stool specimens, extract  
425 fecal genomic DNA, amplify the V3-V4 region of 16S rDNA, and sequence 11mmola  
426 MiSeq platform to analyze the gut microbiota of healthy male with female and healthy  
427 male with SCI patients.

428 The healthy control group included criteria: 1) 18-60 years of age, 2) without a  
429 history of antibiotics or probiotics use 1 month prior to study, 3) without the history of  
430 diabetes, gastrointestinal system diseases, multiple sclerosis, and immune metabolic  
431 diseases. All subjects selected before sampling and signing informed consent fully  
432 understand the sampling process and research options. To exclude probable effects of  
433 diet on microbiota, all patients and healthy subjects were fed with standard hospital  
434 food 2 weeks before stool collection.

## 435 **16S Diversity materials and methods**

### 436 Microbial diversity analysis

#### 437 1. Stool sampling

438 91 fresh specimens were collected, including 23 healthy male, 25 healthy female,  
439 43 SCI patients. Fresh fecal samples were collected and transferred to the laboratory.  
440 Their 200 mg sample was placed in a new 2-mL sterile centrifuge tube, quickly placed



441 on ice, and transferred to a refrigerator-80 °C cryostat for cryopreservation. The entire  
442 sampling process is completed in 30 minutes.

## 443 2. DNA extraction and PCR amplification

444 Microbial DNA was extracted from stool samples using the E.Z.N.A.® Stool DNA Kit  
445 (Omega Bio-tek, Norcross, GA, U.S.) according to manufacturer's protocols. The V3-V4  
446 region of the bacteria 16S rRNA gene were amplified by PCR (95 °C for 2 min, followed by  
447 25 cycles at 95 °C for 30 s, 55 °C for 30 s, and 72 °C for 30 s and a final extension at 72 °C  
448 for 5 min) using primers 338F 5'-ACTCCTACGGGAGGCAGCA-3' and 806R 5'-  
449 GGACTACHVGGGTWTCTAAT-3'. PCR reactions were performed in triplicate 20 µL  
450 mixture containing 4 µL of 5 × FastPfu Buffer, 2 µL of 2.5 mM dNTPs, 0.8 µL of each primer  
451 (5 µM), 0.4 µL of FastPfu Polymerase, and 10 ng of template DNA.

## 452 3. Illumina MiSeq sequencing

453 Amplicons were extracted from 2% agarose gels, purified by using the AxyPrep  
454 DNA Gel Extraction Kit (Axygen Biosciences, Union City, CA, U.S.), and quantified  
455 by using QuantiFluor™ -ST (Promega, U.S.). Purified amplicons were pooled in  
456 equimolar and paired-end sequenced (2 × 300 bp) on an Illumina MiSeq platform  
457 according to the standard protocols. The raw reads were deposited into the NCBI  
458 Sequence Read Archive database (Accession Number: SRP158549).

## 459 4. Processing of sequencing data

460 Raw fastq files were quality-filtered by Trimmomatic and merged by FLASH with  
461 the following criteria: (i) The reads were truncated at any site receiving an average  
462 quality score < 20 over a 50 bp sliding window. (ii) Sequences whose overlap being

463 longer than 10 bp were merged according to their overlap with mismatch no more  
464 than 2 bp. (iii) Sequences of each sample were separated according to barcodes  
465 (exactly matching) and Primers (allowing 2 nucleotide mismatching) and reads  
466 containing ambiguous bases were removed.

467 Operational taxonomic units (OTUs) were clustered with 97% similarity cutoff  
468 using UPARSE (version 7.1) and chimeric sequences were identified and removed  
469 using UCHIME. The taxonomy of each 16S rRNA gene sequence was analyzed by  
470 RDP Classifier algorithm against the Silva (SSU123) 16S rRNA database using  
471 confidence threshold of 70% Roche 454 (Roche, Switzerland) high-throughput  
472 sequencing of the PCR products was performed by Shanghai Majorbio Biological  
473 Technology Co. Ltd., Shanghai, China.

#### 474 **Bioinformatic and statistical analysis**

475 Sequencing reads were processed using QIIME (version 1.9.0), and included  
476 additional quality trimming, demultiplexing, and taxonomic assignments. Profiling of  
477 predictive urine microbiota was analyzed by using PiCRUST based on 13 August 2013  
478 Greengenes database (62). KW rank sum test and pairwise Wilcoxon test were used  
479 for the identification of the different markers, and LDA was used to score each feature  
480 in the LEfSe analysis. Index of alpha diversity was calculated with QIIME based on  
481 sequence similarity at 97%. Beta diversity was measured by unweighted UniFrac  
482 distance, which was also calculated by QIIME. Hierarchical clustering was performed,  
483 and a heatmap was generated using a Spearman's rank correlation coefficient as a  
484 distance measure and a customized script developed in the R statistical package. The

485 output file was further analyzed using Statistical Analysis of Metagenomic Profiles  
486 software package (version 2.1.3) (63).

487 Statistical analysis was performed using the SPSS data analysis program (version  
488 21.0) and Statistical Analysis of Metagenomic Profiles software. For continuous  
489 variables, independent t-test, Welch's t-test, White's nonparametric t-test, and  
490 Mann-Whitney U-test were applied. For categorical variables between groups, using  
491 either the Pearson chi-square or Fisher's exact test, depending on assumption validity.  
492 For taxon among subgroups, ANOVA test was applied (Tukey-Kramer was used in  
493 Post-hoc test, Effect size was Eta-squared) with Benjamini-Hochberg FDP false  
494 discovery rate correction (63,64). All tests of significance were two-sided and p  
495 <0.05.

#### 496 **Abbreviations:**

497 SCI: spinal cord injury; NBD: neurogenic bowel dysfunction; GLU: glucose; HDL: high  
498 density lipoprotein; LDL: low density lipoprotein; UA: Uric acid; CR: Creatinine; CPR:  
499 C-reactive protein; OUTs: Operational taxonomic units; PLS-DA: Partial least squares  
500 discrimination analysis; BMI: Body mass index; APOA1: Apolipoprotein A1; APOB:  
501 Apolipoprotein B; ALT: Alanine transaminase; AST: Aspartate transaminase; TG: Triglyceride;  
502 TCHO: Total Cholesterol; LPA: lipoprotein A; NEFA: non-esterified fatty acid; HCY:  
503 homocysteine; ASCI: acute spinal cord injury; LPS: lipopolysaccharide; AD: Alzheimer's  
504 disease. HM: healthy male; FM: healthy female; PU: quadriplegia SCI patient; PL: paraplegic  
505 SCI patient.

#### 506 **Acknowledgements**

507 This work was supported by the Special Fund for Basic Scientific Research of Central Public  
508 Research Institutes, grant number: 2016cz-1, 2018cz-8, and Beijing Municipal Science  
509 and Technology Commission (BSTC, No. Z171100001017076).

510 **Author Disclosure Statement**

511 No competing interests exist.

512 **References:**

- 513 1. Brommer B, Engel O, Kopp MA, Watzlawick R, Müller S, Prüss H, Chen Y, DeVivo MJ,  
514 Finkenstaedt FW, Dirnagl U, Liebscher T, Meisel A, Schwab JM. 2016. Spinal cord  
515 injury-induced immune deficiency syndrome enhances infection susceptibility dependent on  
516 lesion level. *Brain* 139:692-707.
- 517 2. Meisel C, Schwab JM, Prass K, Meisel A, Dirnagl U. 2005. Central nervous system  
518 injury-induced immune deficiency syndrome. *Nat Rev Neurosci* 6:775-786.
- 519 3. Rabchevsky AG. 2006. Segmental organization of spinal reflexes mediating autonomic  
520 dysreflexia after spinal cord injury. *Prog Brain Res* 152:265-274.
- 521 4. Ueno M, Ueno-Nakamura Y, Niehaus J, Popovich PG, Yoshida Y. 2006. Silencing spinal  
522 interneurons inhibits immune suppressive autonomic reflexes caused by spinal cord injury.  
523 *Nat Neurosci* 19:784-787.
- 524 5. Zhang Y, Guan Z, Reader B, Shawler T, Mandrekar-Colucci S, Huang K, Weil Z, Bratasz A,  
525 Wells J, Powell ND, Sheridan JF, Whitacre CC, Rabchevsky AG, Nash MS, Popovich  
526 PG. 2013. Autonomic dysreflexia causes chronic immune suppression after spinal cord injury.  
527 *J Neurosci* 33:12970-12981.
- 528 6. Cervi AL, Lukewich MK, Lomax AE. 2014. Neural regulation of gastrointestinal

- 529 inflammation: role of the sympathetic nervous system. *Auton Neurosci* 182:83-88.
- 530 7. Elenkov IJ, Wilder RL, Chrousos GP, Vizi ES.2000. The sympathetic nerve—an integrative  
531 interface between two supersystems: the brain and the immune system. *Pharmacol Rev*  
532 52:595-638.
- 533 8. Koo BI, Bang TS, Kim SY, Ko SH, Kim W, Ko HY.2016. Anorectal Manometric and  
534 Urodynamic Parameters According to the Spinal Cord Injury Lesion(J). *Ann Rehabil Med*  
535 40(3):528-533.
- 536 9. Tate DG, Forchheimer M, Rodriguez G, Chiodo A, Cameron A, Meade M, Krassioukov  
537 A.2016. Risk Factors Associated with Neurogenic Bowel Complications and Dysfunction in  
538 Spinal Cord Injury(J). *Arch Phys Med Rehabil* 97(10),1679-1686.
- 539 10. Ozisler Z, Koklu K, Ozel S, Unsal-Delialioglu S.2015. Outcomes of bowel program in  
540 spinal cord injury patients with neurogenic bowel dysfunction(J). *Neural Regen Res*  
541 10(7):1153-1158.
- 542 11. Longo WE, Ballantyne GH, Modlin IM.1989. The colon, anorectum and spinal cord  
543 patient. A review of the functional alternations of the denervated hindgut. *Dis Colon Rectum*  
544 32(3): 261–267.
- 545 12. Glickman S, Kamm MA.1996. Bowel dysfunction in spinal-cord-injury patients. *Lancet*  
546 347: 1651–1653.
- 547 13. Benevento BT, Sipski ML.2002. Neurogenic bladder, neurogenic bowel, and sexual  
548 dysfunction in people with spinal cord injury. *PhysTher* 82(6): 601–612.
- 549 14. Ng C, Prott G, Rutkowski S, Li Y, Hansen R, Kellow J, Malcolm A.2005. Gastrointestinal  
550 symptoms in spinal cord injury: relationships with level of injury and psychologic factors. *Dis*

- 551 Colon Rectum 48(8): 1562–1568.
- 552 15. Lynch AC, Anthony A, Dobbs BR, Frizelle FA.2000. Anorectal physiology following  
553 spinal cord injury. *Spinal Cord* 38: 573–580.
- 554 16. Dethlefsen L, McFall-Ngai M, Relman DA.2007. An ecological and evolutionary  
555 perspective on human microbe mutualism and disease. *Nature* 449(7164): 811–818.
- 556 17. Hooper LV, Littman DR, Macpherson AJ.2012. Interactions between the microbiota and  
557 the immune system. *Science* 336:1268-1273.
- 558 18. Yano JM, Yu K, Donaldson GP, Shastri GG, Ann P, Ma L, Nagler CR, Ismagilov RF,  
559 Mazmanian SK, Hsiao EY.2015. Indigenous bacteria from the gut microbiota regulate host  
560 serotonin biosynthesis. *Cell* 161(2):264-276.
- 561 19. Sartor RB. Microbial influences in inflammatory bowel diseases. *Gastroenterology*. 2008;  
562 134 (2): 577–594.
- 563 20. Collins S, Verdu E, Denou E, Bercik P.2009. The role of pathogenic microbes and  
564 commensal bacteria in irritable bowel syndrome. *Dig Dis* 27: 85–89.
- 565 21. Robles Alonso V, Guarner F.2013. Linking the gut microbiota to human health. *Br J Nutr*  
566 109: S21–26.
- 567 22.Elin O, Margarete M, Parks BW, Shipkova P, Liu X, Drake TA, Lusic AJ.2016. Sex  
568 differences and hormonal effects on gut microbiota composition in mice: (J). *Gut Microbes*  
569 7(4):313.
- 570 23. Borgo F, Garbossa S, Riva A, Severgnini M, Luigiano C, Benetti A, Pontiroli AE,  
571 Morace G, Borghi E.2018. Body Mass Index and Sex Affect Diverse Microbial Niches within  
572 the Gut. *Front Microbiol* 9: 213.

- 573 24. Hill DA, Artis D.2010. Intestinal bacteria and the regulation of immune cell homeostasis.  
574 *Annu Rev Immunol* 28:623-667.
- 575 25. El Aidy S, Dinan TG, Cryan JF.2015. Gut microbiota: the conductor in the orchestra of  
576 immune-neuroendocrine communication. *Clin Ther* 37:954-967
- 577 26. Balzan S, de Almeida Quadros C, de Clevea R, Zilberstein B, Ceconello I.2007. Bacterial  
578 translocation: overview of mechanisms and clinical impact. *J Gastroenterol Hepatol*  
579 22:464-471.
- 580 27. Khosravi A, Yáñez A, Price JG, Chow A, Merad M, Goodridge HS, Mazmanian SK.2014.  
581 Gut microbiota promote hematopoiesis to control bacterial infection. *Cell Host Microbe*  
582 15(3):374-381.
- 583 28. Mazo IB, Massberg S, von Andrian UH.2011. Hematopoietic stem and progenitor cell  
584 trafficking. *Trends Immunol* 32(10):493-503.
- 585 29. Gungor B, Adiguzel E, Gursel I, Yilmaz B, Gursel M.2016. Intestinal microbiota in  
586 patients with spinal cord injury. *PLoS ONE* 11(1): e0145878.
- 587 30. Kigerl KA, Hall JC, Wang L, Mo X, Yu Z, Popovich PG.2016. Gut dysbiosis impairs  
588 recovery after spinal cord injury. *J Exp Med* 213 (12):2603-2620.
- 589 31.Kigerl KA, Mostacada K, Popovich PG.2018. Gut Microbiota Are Disease-Modifying  
590 Factors After Traumatic Spinal Cord Injury. *Neurotherapeutics* 15(1):60-67.
- 591 32. J Li, G Liu, Y Zheng, C Hao, Y Zhang, B Wei, H Zhou, D Wang.2011.The  
592 epidemiological survey of acute traumatic spinal cord injury (ATSCI) of 2002 in Beijing  
593 municipality. *Spinal Cord* 49:777–782.
- 594 33. R Yang, L Guo, L Huang, P Wang, Y Tang, J Ye, K Chen, X Hu, Z Cai, C Lu, Y Wu, H

- 595 Shen.2016. Epidemiological Characteristics of Traumatic Spinal Cord Injury in Guangdong,  
596 China(J). Spine 42 (9): E555.
- 597 34. Haro C, Rangel-Zúñiga OA, Alcalá-Díaz JF, Gómez-Delgado F, PérezMartínez P,  
598 Delgado-Lista J, Quintana-Navarro GM, Landa BB, Navas-Cortés JA, Tena-Sempere M,  
599 Clemente JC, López-Miranda J, Pérez-Jiménez F, Camargo A.2016. Intestinal microbiota is  
600 influenced by gender and body mass index. PLOS ONE 11 (5): e0154090.
- 601 35.Moschen AR, Wieser V, Tilg H,2012. Dietary Factors: Major Regulators of the Gut's  
602 Microbiota. Gut Liver 6:411–416.
- 603 36. Carmody RN, Gerber GK, Luevano JM Jr., Gatti DM, Somes L, Svenson KL, Turnbaugh  
604 PJ.2015. Diet dominates host genotype in shaping the murine gut microbiota. Cell host &  
605 microbe17:72–84.
- 606 37. Engkasan JP, Sudin SS.2013. Neurogenic bowel management after spinal cord injury:  
607 Malaysian experience. J Rehabil Med 45: 141–144.
- 608 38. Yasmeeen R, Rathore FA, Ashraf K, Butt AW.2010. How do patients with chronic spinal  
609 injury manage their bowels? A cross-sectional survey of 50 patients. Spinal Cord 48:  
610 872–875.
- 611 39. Han TR, Kim JH, Kwon BS.1998. Chronic gastrointestinal problems and bowel  
612 dysfunction in patients with spinal cord injury. Spinal Cord 36:485-490
- 613 40.Kirshblum SC, Gulati M, O'Connor KC, Voorman SJ.1998. Bowel care practices in  
614 chronic spinal cord injury patients. Arch Phys Med Rehabil 79:20-23.
- 615 41.Stone JM, Nino-Murcia M, Wolfe VA, Perakash I.1990. Chronic gastrointestinal problems  
616 in spinal cord injury patients: a prospective analysis. Am J Gastroenterol 85:1114-1119.



- 617 42. Lynch AC, Antony A, Dobbs BR, Frizelle FA.2001. Bowel dysfunction following spinal  
618 cord injury. *Spinal Cord* 39:193-203.
- 619 43. Anderson KD.2004. Targeting Recovery: Priorities of the Spinal Cord Injured Population.  
620 *J. Neurotrauma* 21: 1371-1383.
- 621 44. Simpson LA, Eng JJ, Hsieh JT, Wolfe DL, and the SCIRE Research Team.2012. The  
622 health and life priorities of individuals with spinal cord injury: A systematic review. *J.*  
623 *Neurotraum* 29:1548-1555.
- 624 45. Rajilić-Stojanović M, Vos WM.2014. The first 1000 cultured species of the human  
625 gastrointestinal microbiota. *FEMS Microbiol. Rev* 38: 996–1047.
- 626 46. Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F,  
627 Tuohy KM, Chabo C, Waget A, Delmée E, Cousin B, Sulpice T, Chamontin B, Ferrières J,  
628 Tanti JF, Gibson GR, Casteilla L, Delzenne NM, Alessi MC, Burcelin R.2007. Metabolic  
629 endotoxemia initiates obesity and insulin resistance. *Diabetes* 56(7):1761–1772.
- 630 47. Myers J, Lee M, Kiratli J.2007.Cardiovascular disease in spinal cord injury: an overview  
631 of prevalence, risk, evaluation, and management(J). *Am J Phys Med Rehabil* 86(2):142-152.
- 632 48. Vogt NM, Kerby RL, Dill-McFarland KA, Harding SJ, Merluzzi AP, Johnson SC,  
633 Carlsson CM, Asthana S, Zetterberg H, Blennow K, Bendlin BB, Rey FE.2017. Gut  
634 microbiome alterations in Alzheimer’s disease. *Sci Rep* 7: 13537.
- 635 49. Liao M, Xie Y, Mao Y, Lu Z, Tan A, Wu C, Zhang Z, Chen Y, Li T, Ye Y, Yao Z, Jiang Y,  
636 Li H, Li X, Yang X, Wang Q, Mo Z.2018. Comparative analyses of fecal microbiota in  
637 Chinese isolated Yao population, minority Zhuang and rural Han by 16sRNA sequencing. *Sci*  
638 *Rep* 8:1142.

- 639 50. Saulnier DM, Riehle K, Mistretta TA, Diaz MA, Mandal D, Raza S, Weidler EM, Qin X,  
640 Coarfa C, Milosavljevic A, Petrosino JF, Highlander S, Gibbs R, Lynch SV, Shulman RJ,  
641 Versalovic J.2011. Gastrointestinal microbiome signatures of pediatric patients with irritable  
642 bowel syndrome. *Gastroenterology* 141: 1782–1791.
- 643 51. Li F, Hullar MA, Schwarz Y, Lampe JW.2009. Human gut bacterial communities are  
644 altered by addition of cruciferous vegetables to a controlled fruit- and vegetable-free diet. *J*  
645 *Nutr* 139:1685–1691.
- 646 52. David LA, Maurice CF, Carmody RN, Gootenberg DB; Button JE; Wolfe BE, Ling AV,  
647 Devlin AS, Varma Y, Fischbach MA, Biddinger SB, Dutton RJ, Turnbaugh PJ.2014. Diet  
648 rapidly and reproducibly alters the human gut microbiome. *Nature* 505:559–563.
- 649 53. O’Callaghan A, van Sinderen D.2016. Bifidobacteria and their role as members of the  
650 human gut microbiota. *Front Microbiol* 7: 1–23.
- 651 54. Arboleya S, Watkins C, Stanton C, Ross RP.2016. Gut bifidobacteria populations in human  
652 health and aging. *Front Microbiol* 7: 1–9.
- 653 55. Martínez I, Stegen JC, Maldonado-Gómez MX, Eren AM, Siba PM, Greenhill AR, Walter  
654 J.2015. The gut microbiota of rural papua new guineans: composition, diversity patterns, and  
655 ecological processes. *Cell Rep* 11:527–538.
- 656 56. Wu GD, Chen J, Hoffmann C, Bittinger K, Chen YY, Keilbaugh SA, Bewtra M, Knights  
657 D, Walters WA, Knight R, Sinha R, Gilroy E, Gupta K, Baldassano R, Nessel L, Li H,  
658 Bushman FD, Lewis JD.2011. Linking long-term dietary patterns with gut microbial  
659 enterotypes. *Science* 334: 105–108.
- 660 57. Kovatcheva-Datchary P, Nilsson A, Akrami R, ShiuanLeeY, Vadder FD, Arora T, Hallen A,

- 661 Martens E, Björck I, Bäckhed F.2015. Dietary Fiber-Induced Improvement in Glucose  
662 Metabolism Is Associated with Increased Abundance of Prevotella. *Cell Metab*  
663 22(6):971–982.
- 664 58. Hofer U.2014. Microbiome: pro-inflammatory Prevotella? *Nat Rev Microbiol* 12 (1):5.
- 665 59. DeVivo M, Biering-S F, Charlifue S, Noonan V, Post M, Stripling T, Wing P.2006.  
666 International Spinal Cord Injury Core Data Set. *Spinal Cord* 44, 535-540.
- 667 60. Krogh K, Perakash I, Stiens SA, Biering-Srensen F.2009. International bowel function  
668 basic spinal cord injury data set(J). *Spinal Cord* 47(3):230-4.
- 669 61. Krogh K,Perakash I,Stiens SA, Biering-Srensen F.2009. International bowel function  
670 extended spinal cord injury data set(J). *Spinal Cord* 47(3),235-41.
- 671 62. Langille MG, Zaneveld J, Caporaso JG, McDonald D, Knights D, Reyes JA, Clemente JC,  
672 Burkepille DE, Vega Thurber RL, Knight R, Beiko RG, Huttenhower C.2013. Predictive  
673 functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nat*  
674 *Biotechnol* 31(9):814-821.
- 675 63. Parks DH, Tyson GW, Hugenholtz P and Beiko RG.2014. STAMP: statistical analysis of  
676 taxonomic and functional profiles. *Bioinformatics* 30(21):3123-3124.
- 677 64. White JR, Nagarajan N and Pop M.2009. Statistical methods for detecting differentially  
678 abundant features in clinical metagenomic samples. *PLoS Comput Biol* 5(4): e1000352.

679 **Figure legends:**

680 Figure 1 Diversity and taxonomic analysis in the health male and female groups

681 A: No significant difference in OTU abundance (Simpson index) was observed between the  
682 male and female populations ( $p=0.147$ ).

683 B: The discrete case of sample points distribution in PLS-DA on Genus level showed  
684 differences in the composition of the gut flora between male and female groups.

685 C: Genus-level operational taxonomic units different between healthy male and female groups.  
686 STAMP software was used to calculate the genus proportions in the two groups. There were 4  
687 of top 15 genus showed a significant difference ( $P < 0.05$ ) among two groups (Welch's t-test).

688 Figure 2: Diversity and taxonomic analysis in the health male and SCI groups

689 A: In genus level, Simpson index showed a significant difference between healthy male and  
690 SCI groups ( $P = 0.03635$ ).

691 B. Plot of principal coordinate analysis (PCA) on Phylum level of the fecal microbiota based  
692 on the unweighted UniFrac metric in healthy male and SCI groups.

693 STAMP analysis on Phylum and Genus level showed differences between healthy male and  
694 SCI groups. There were 2 of top 15 phylum (C) and 9 of top 15 genus (D) showed a  
695 significant difference ( $P < 0.05$ ) among two groups (Welch's t-test).

696 Figure 3 STAMP analysis on NBD symptoms

697 A. STAMP analysis showed a significant difference ( $P < 0.05$ ) among two groups (Welch's  
698 t-test) in Bifidobacterium on Genus level.

699 B. STAMP analysis showed Megamonas had a significant high ( $P < 0.05$ ) in bloating group  
700 and Alistipes had a significant high ( $P < 0.05$ ) in without bloating group on Genus level.

701 Figure 4. Correlation heatmap analysis of different environmental factors on the community  
702 composition of the healthy male and SCI groups in phylum level (A) and genus level (B).

703 Figure 5. Diversity and taxonomic analysis in the health male, quadriplegia and paraplegic  
704 SCI groups.

705 A. Sobs index of Rarefaction curves for healthy male, quadriplegia and paraplegic groups of  
706 samples based on OTUs detected using a similarity threshold of 97%.

707 B. Significant difference in genus chao index(B) and Simpson index(C) were observed  
708 between the three populations( $P < 0.05$ ).

709 D. ANOSIM/Adonis of beta-diversity analysis revealed significant differences in the structure  
710 of gut microbiota among the three groups ( $p=0.001$ ,  $r^2=0.233$ ) in phylum level.

711 E.PLS-DA revealed that there were significant differences in bacterial community  
712 composition between three groups on OTUs, phylum and genus level.

713 Figure 6. STAMP analysis indicates the significant difference phylum(A) and genus (B-H)  
714 among three groups.

715 Figure 7. Correlation heatmap analysis of different environmental factors on the community  
716 composition of the quadriplegia and paraplegic groups in phylum level (A) and genus  
717 level(B).

718 **Supplementary File Figure legends:**

719 Supplementary File 1:

720 Sobs index of Rarefaction curves for healthy male and female groups of samples based on  
721 OTUs detected using a similarity threshold of 97%.

722 Supplementary File 2:

723 ANOSIM/Adonis on OUT, phylum and genes level revealed significant differences in the  
724 structure of gut microbiota among the healthy male and SCI groups ( $p < 0.05$ ) ..

725 Supplementary File 3

726 Correlation heatmap analysis chart of different environmental factors on the community

727 composition of healthy male and SCI groups in genus level.

728 Supplementary File 4

729 Alpha-diversity index inter-group difference test chart between healthy male, quadriplegia  
730 and paraplegic SCI cohorts in OTUs level.

731 Supplementary File 5:

732 Alpha-diversity index inter-group difference test chart between quadriplegia and paraplegic  
733 SCI cohorts in genus level.

734 Supplementary File 6:

735 Alpha-diversity index inter-group difference test chart between healthy male and paraplegic  
736 SCI cohorts.

737 Supplementary File 7:

738 ANOSIM/Adonis distances box plot on phylum level revealed significant differences in the  
739 structure of gut microbiota among the healthy male, quadriplegia and paraplegic SCI groups  
740 ( $p < 0.05$ ) .

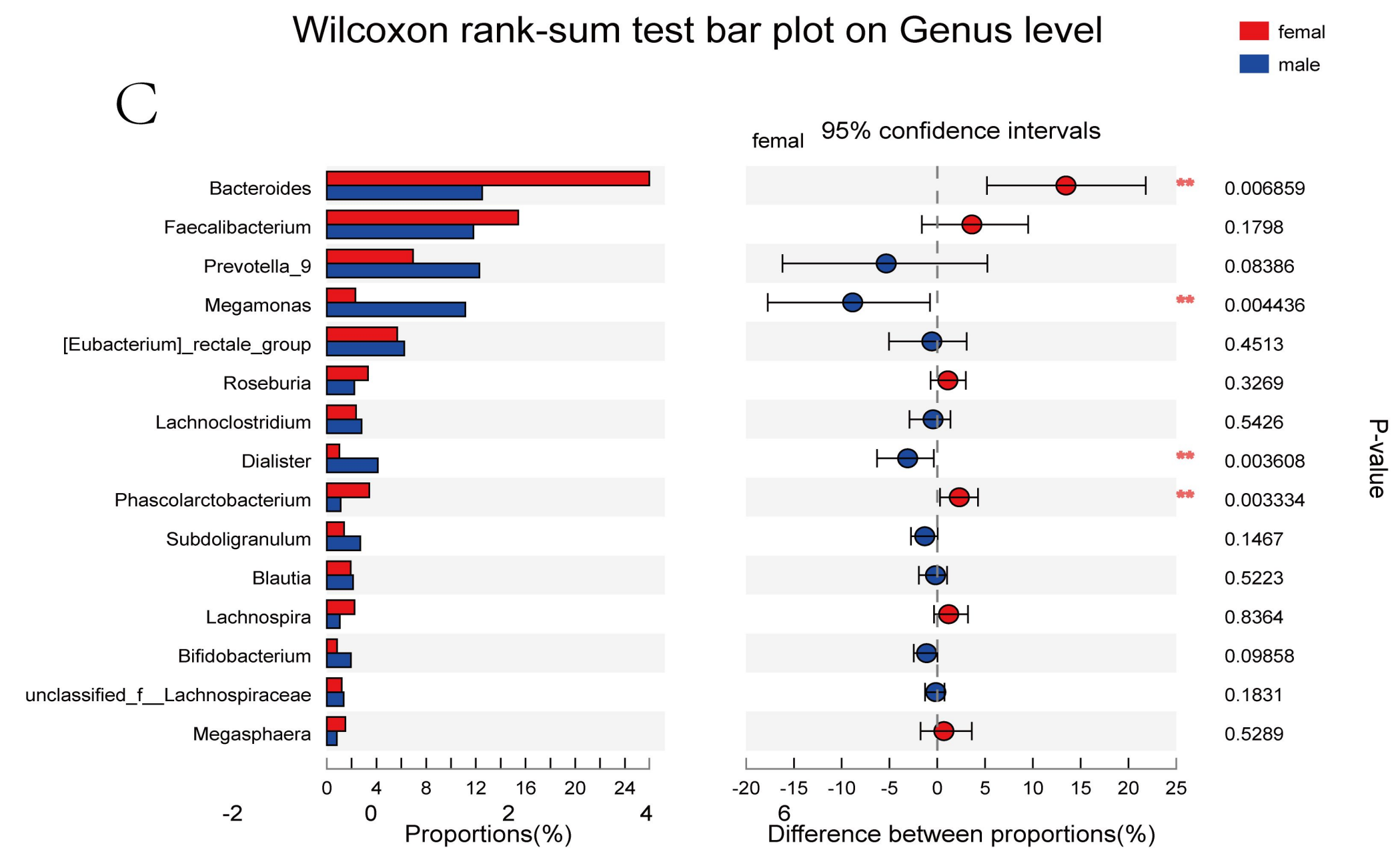
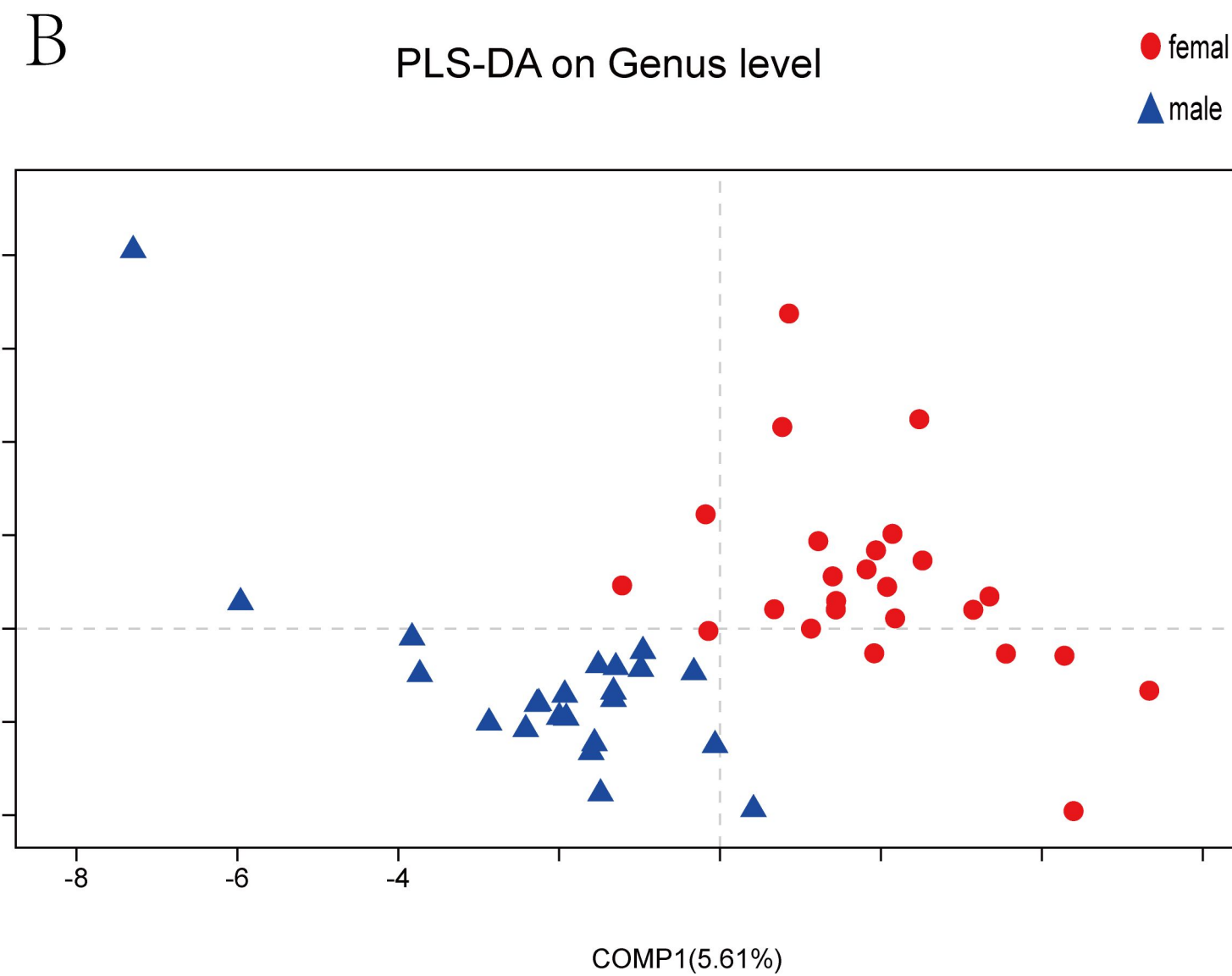
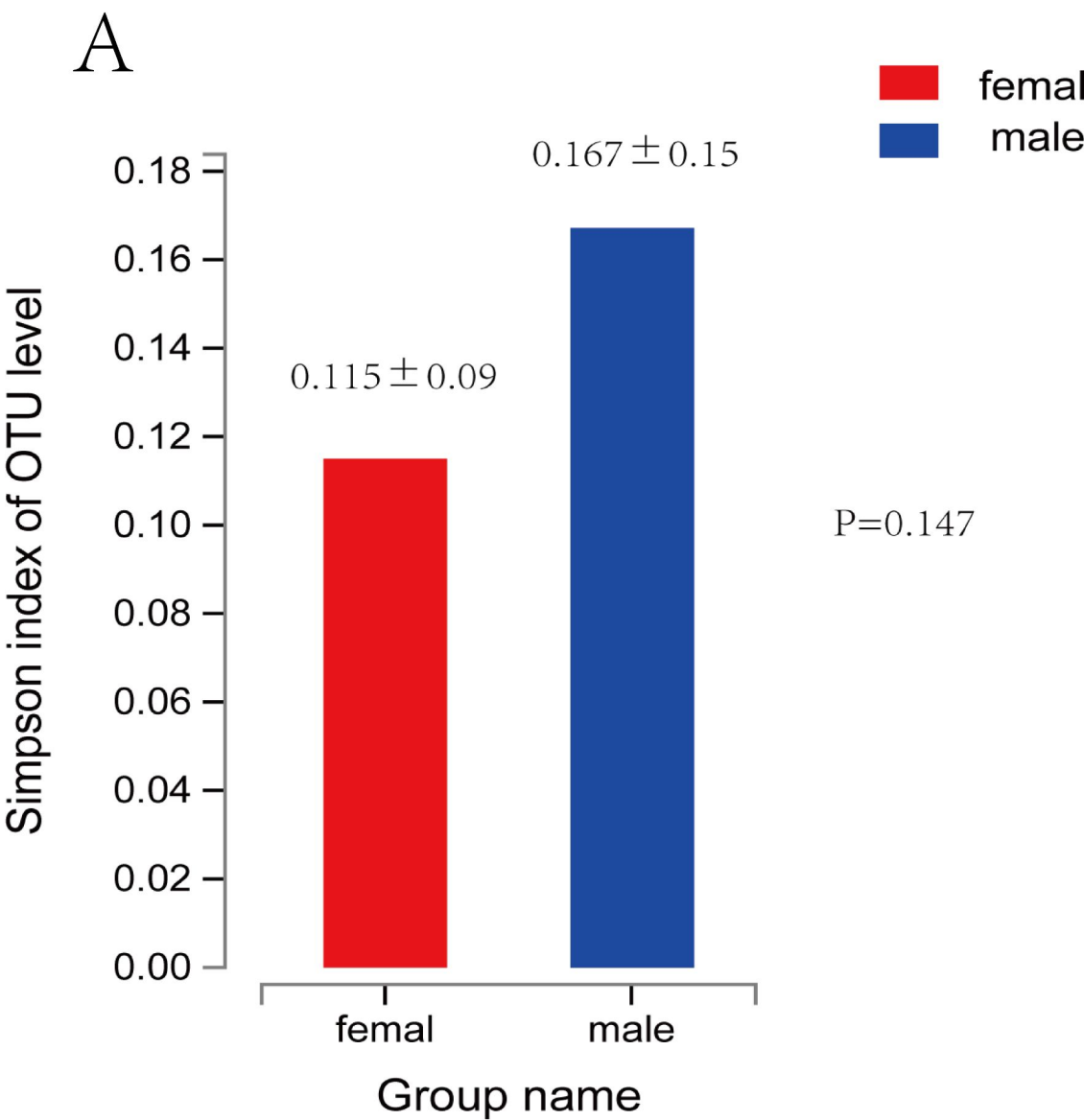
741 Supplementary File 8

742 Correlation heatmap analysis chart of different environmental factors on the community  
743 composition of quadriplegia and paraplegic SCI groups in phylum level.

744 Supplementary File 9

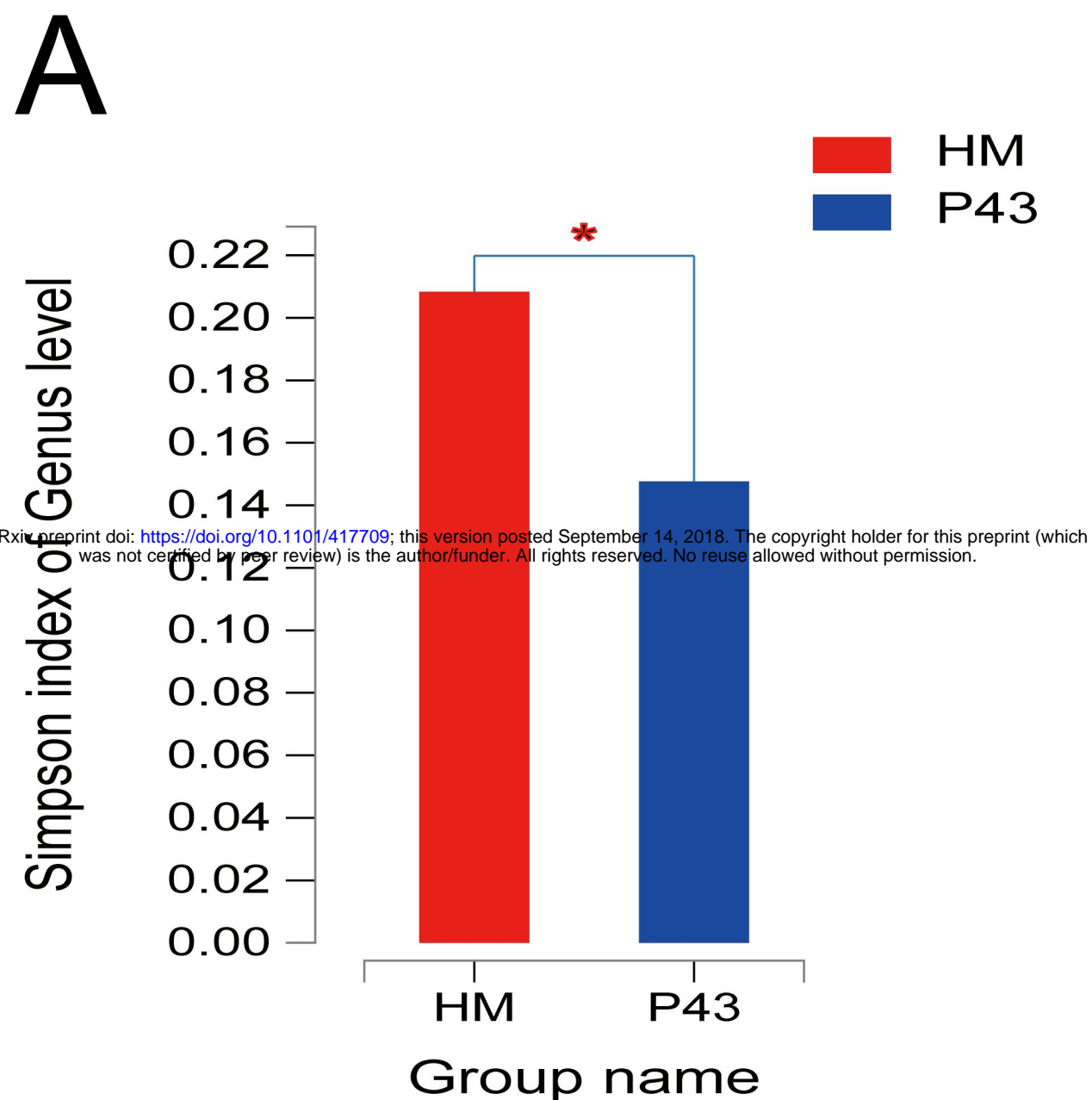
745 Correlation heatmap analysis chart of different environmental factors on the community  
746 composition of quadriplegia and paraplegic SCI groups in genus level.

# Student's t-test for estimator

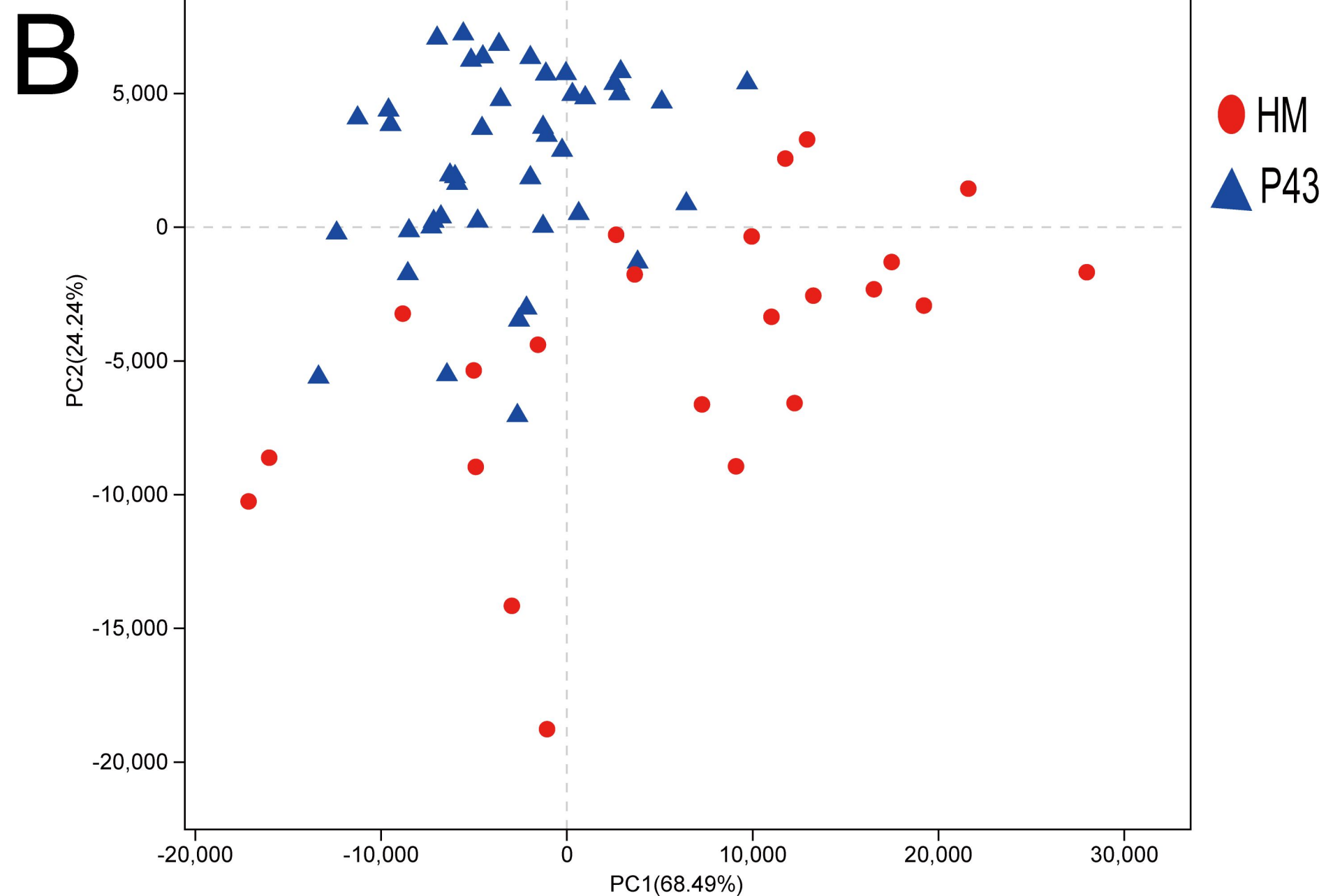




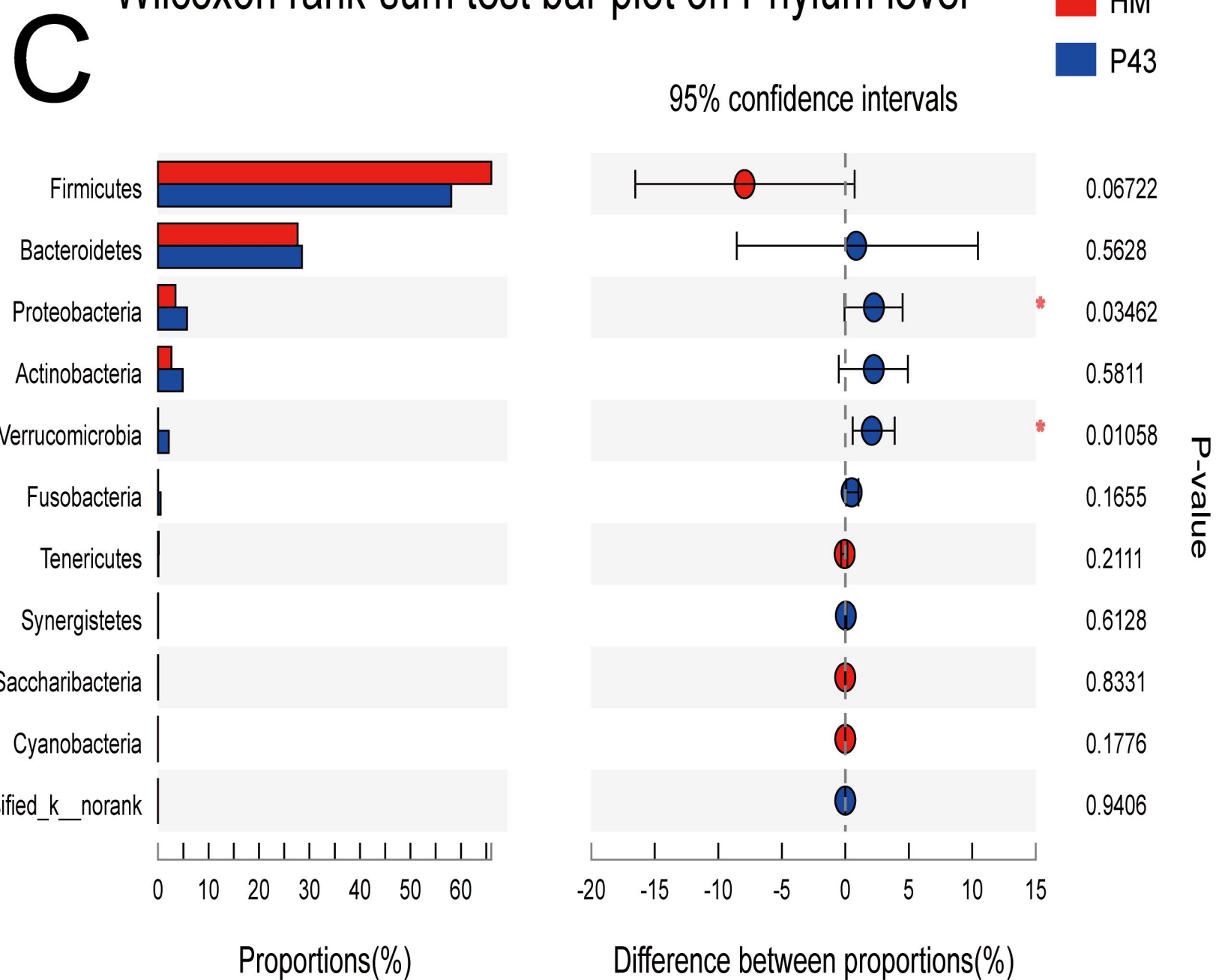
# Student's t-test for estimator



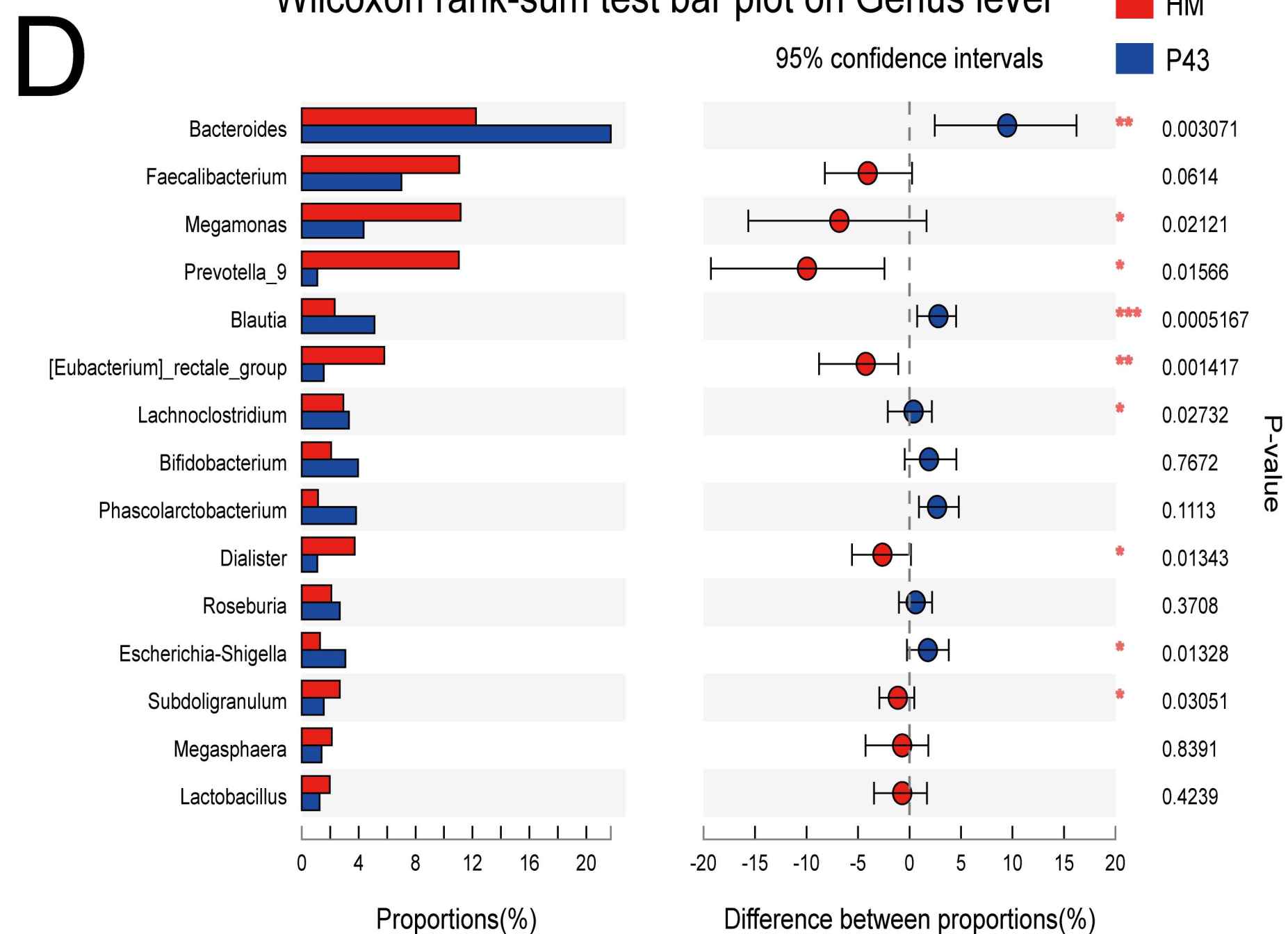
# PCA on Phylum level



# Wilcoxon rank-sum test bar plot on Phylum level



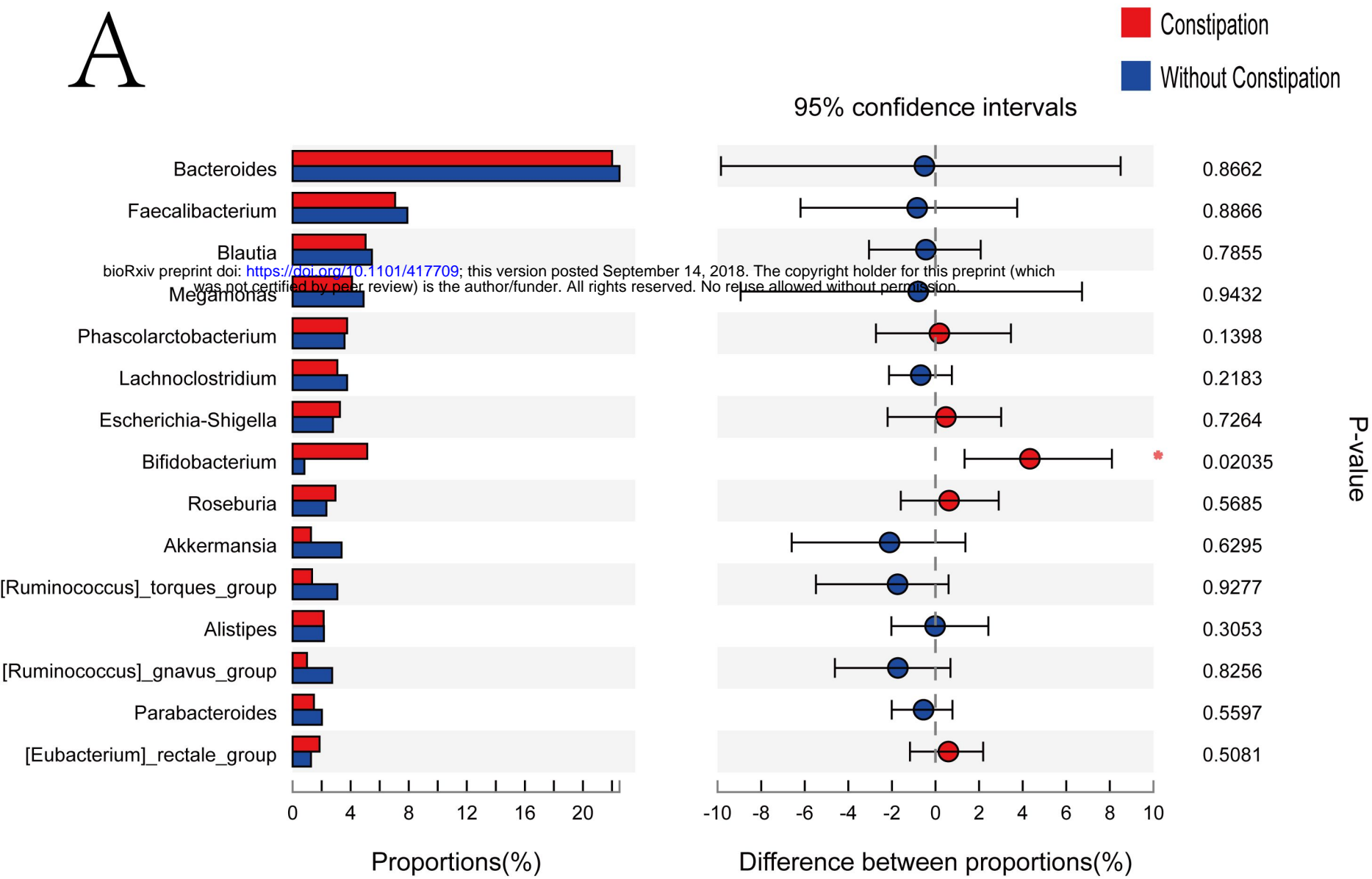
# Wilcoxon rank-sum test bar plot on Genus level





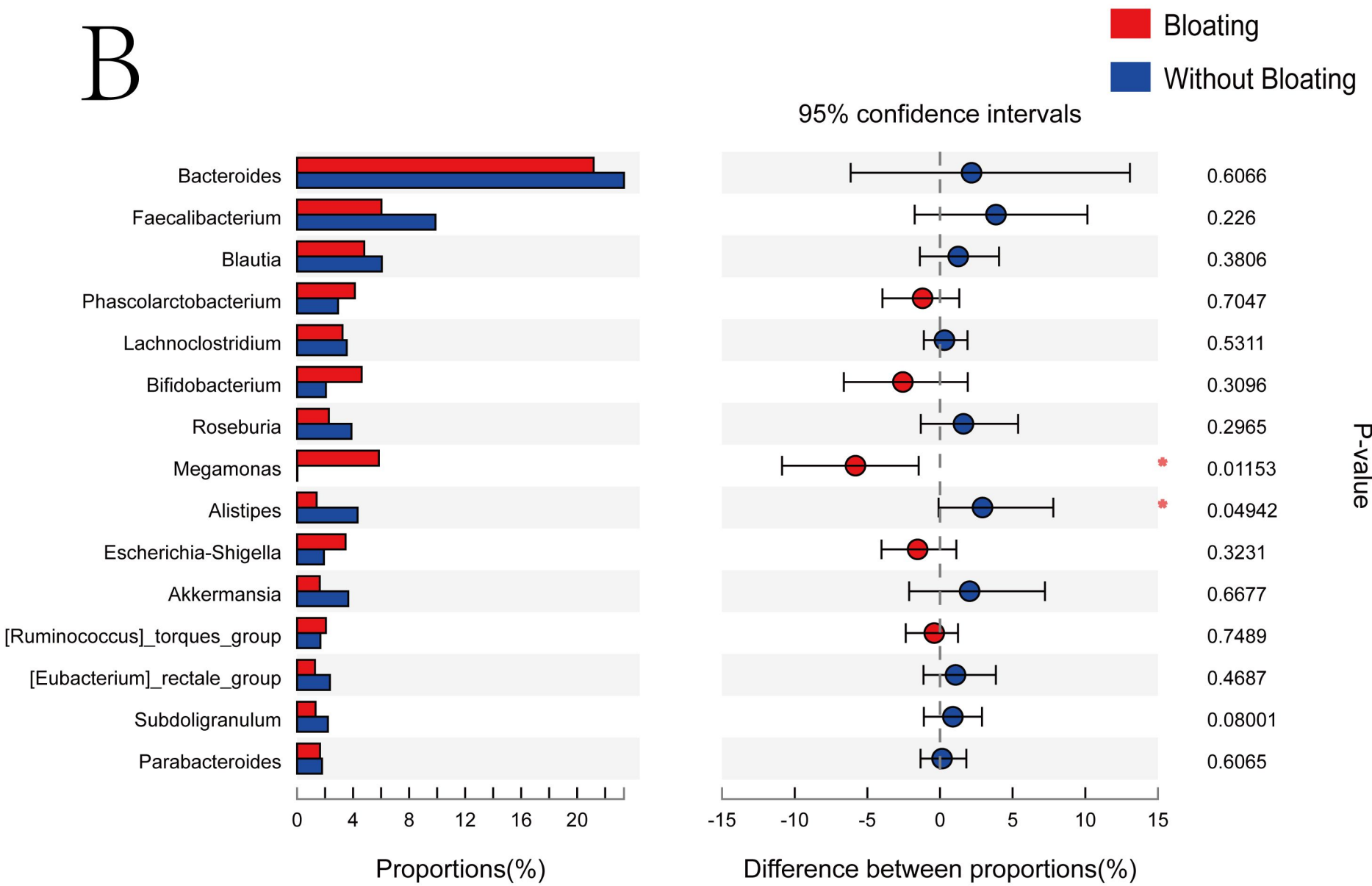
# Wilcoxon rank-sum test bar plot on Genus level

## A



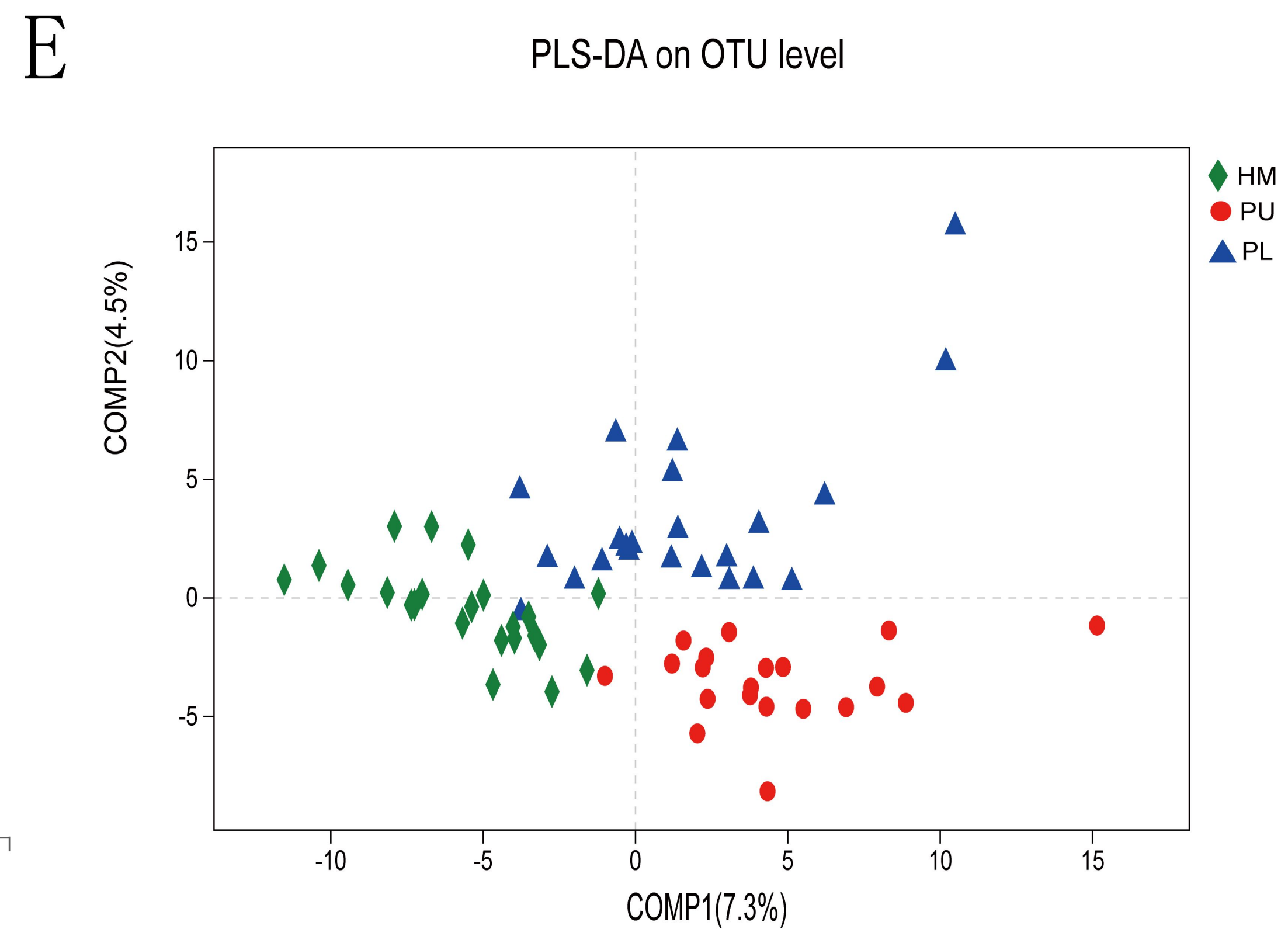
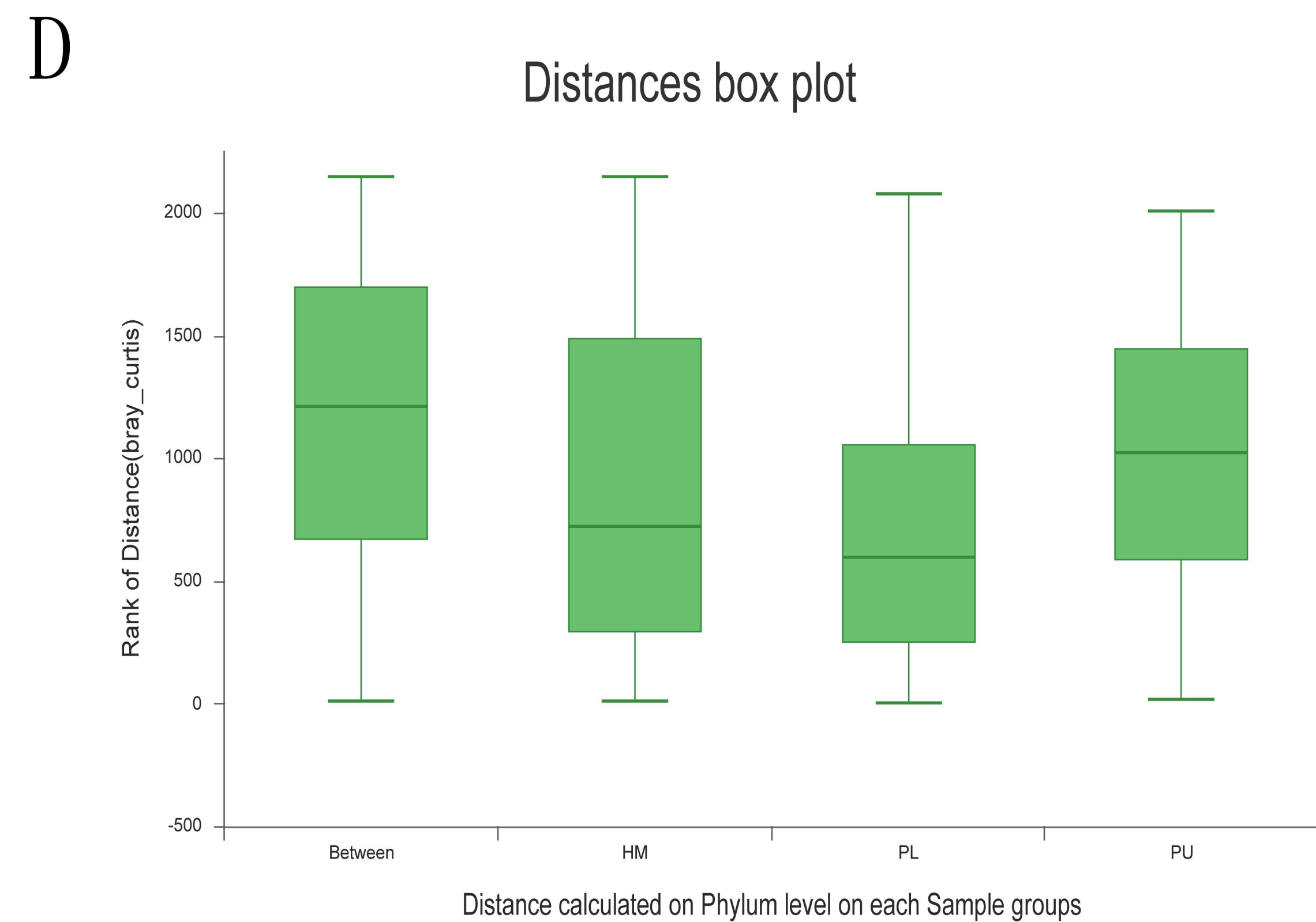
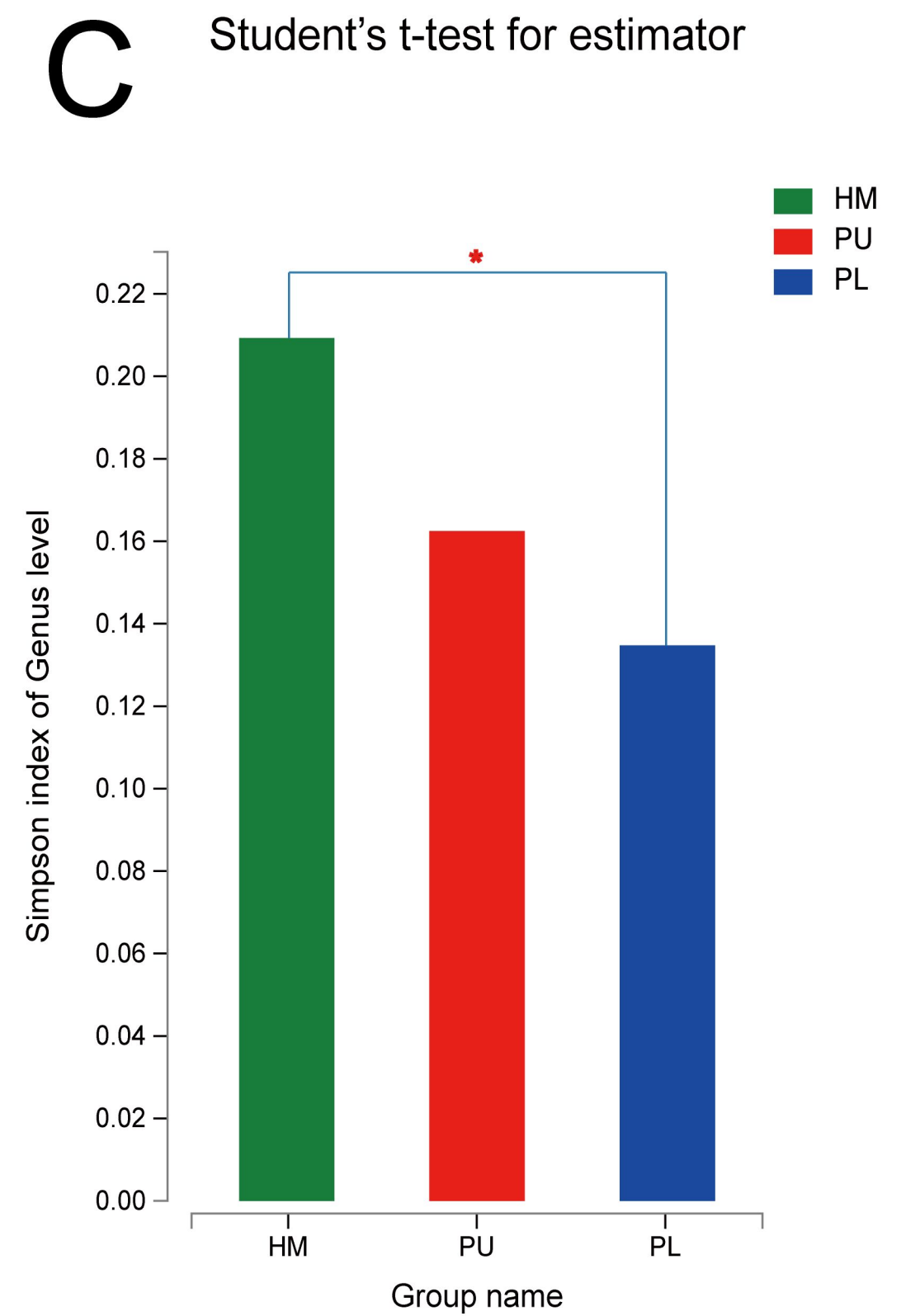
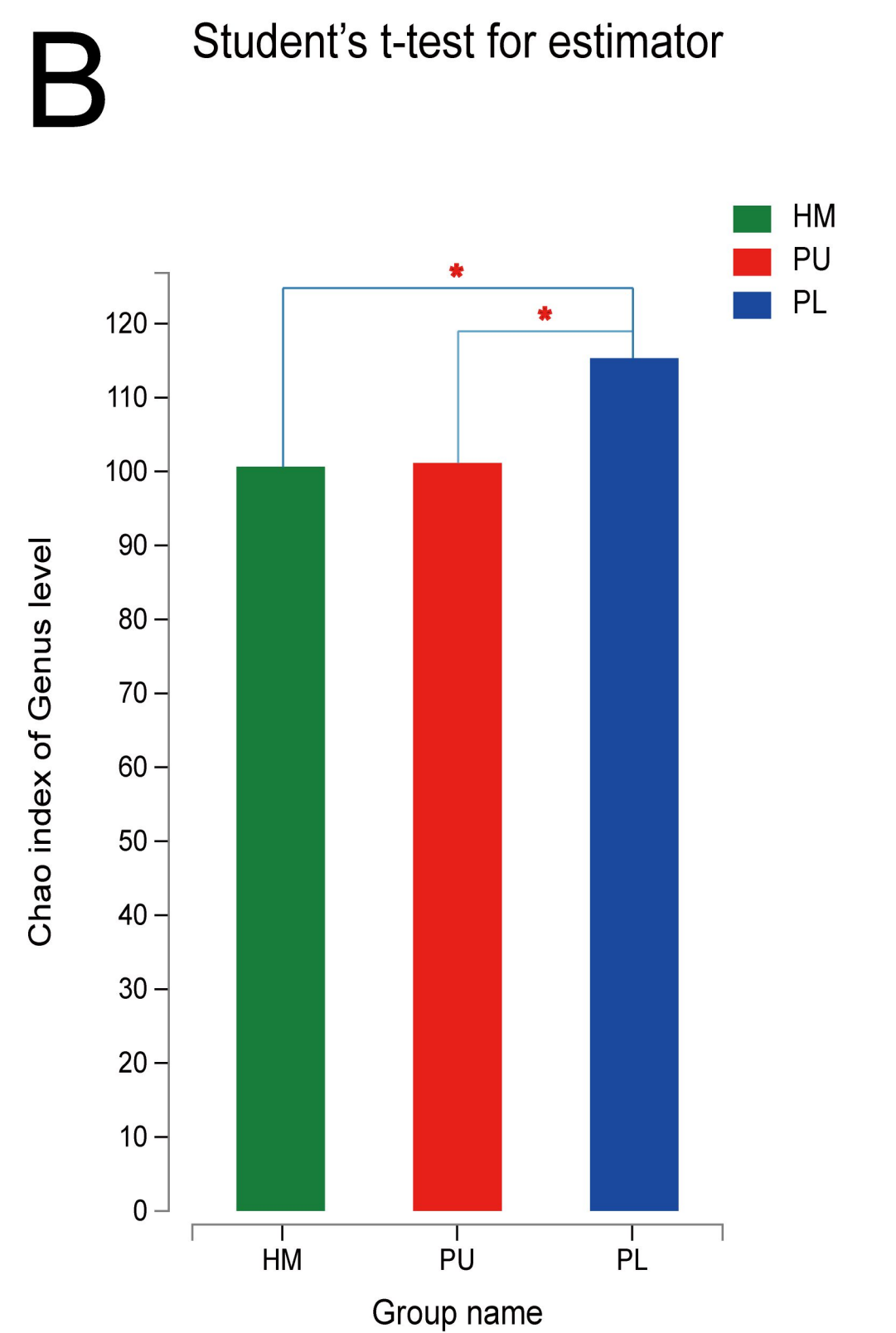
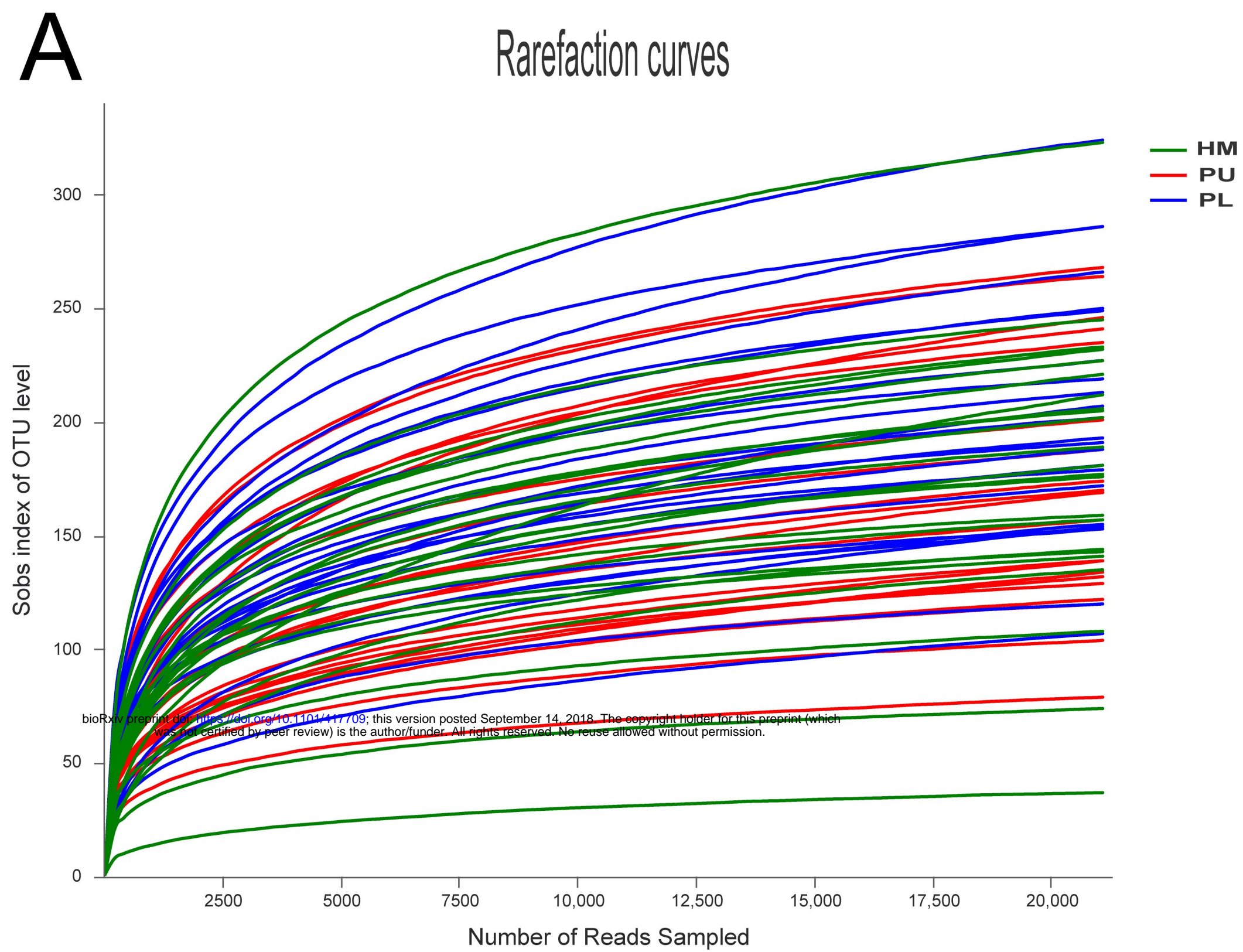
# Wilcoxon rank-sum test bar plot on Genus level

## B











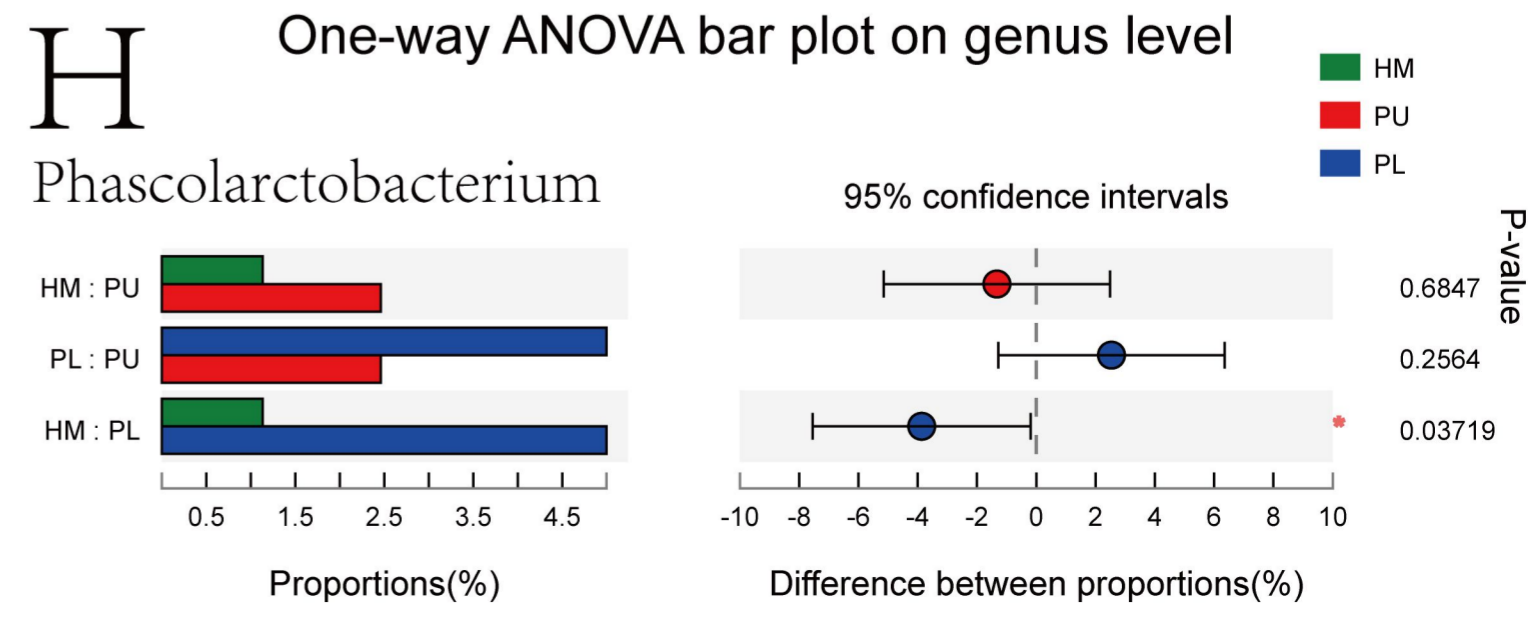
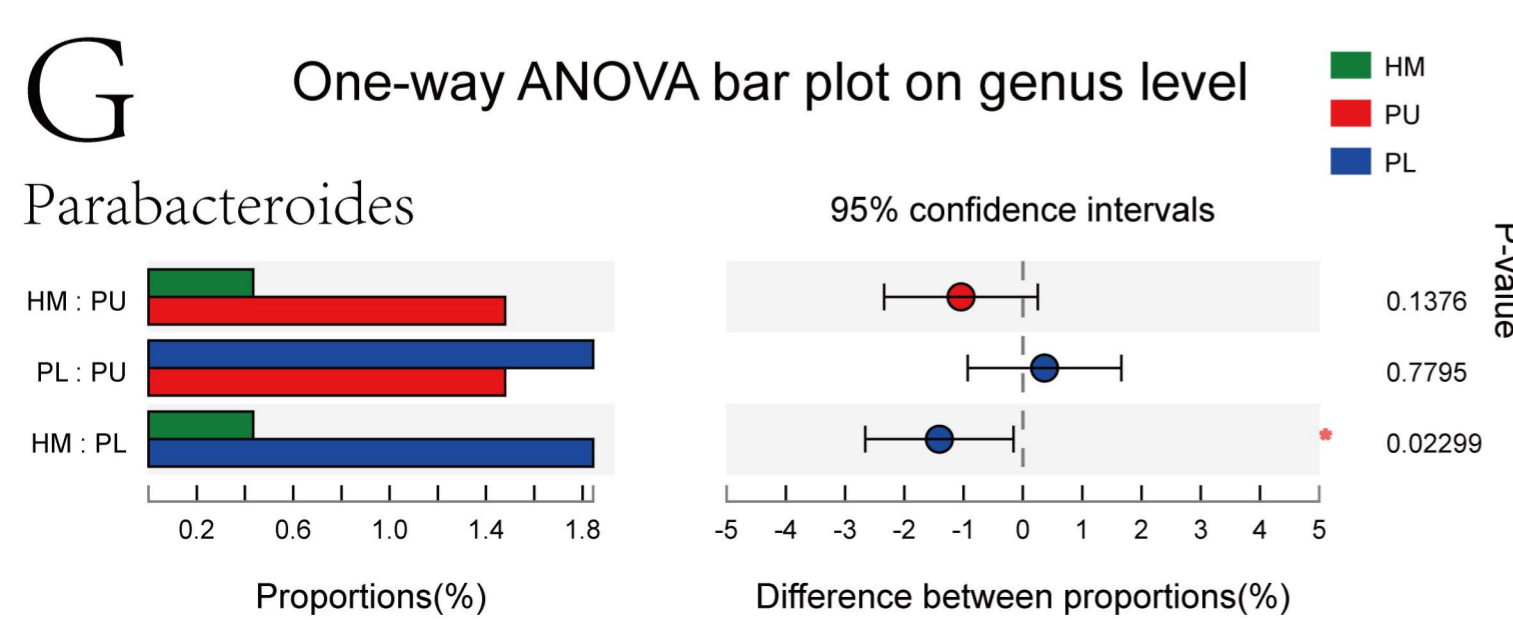
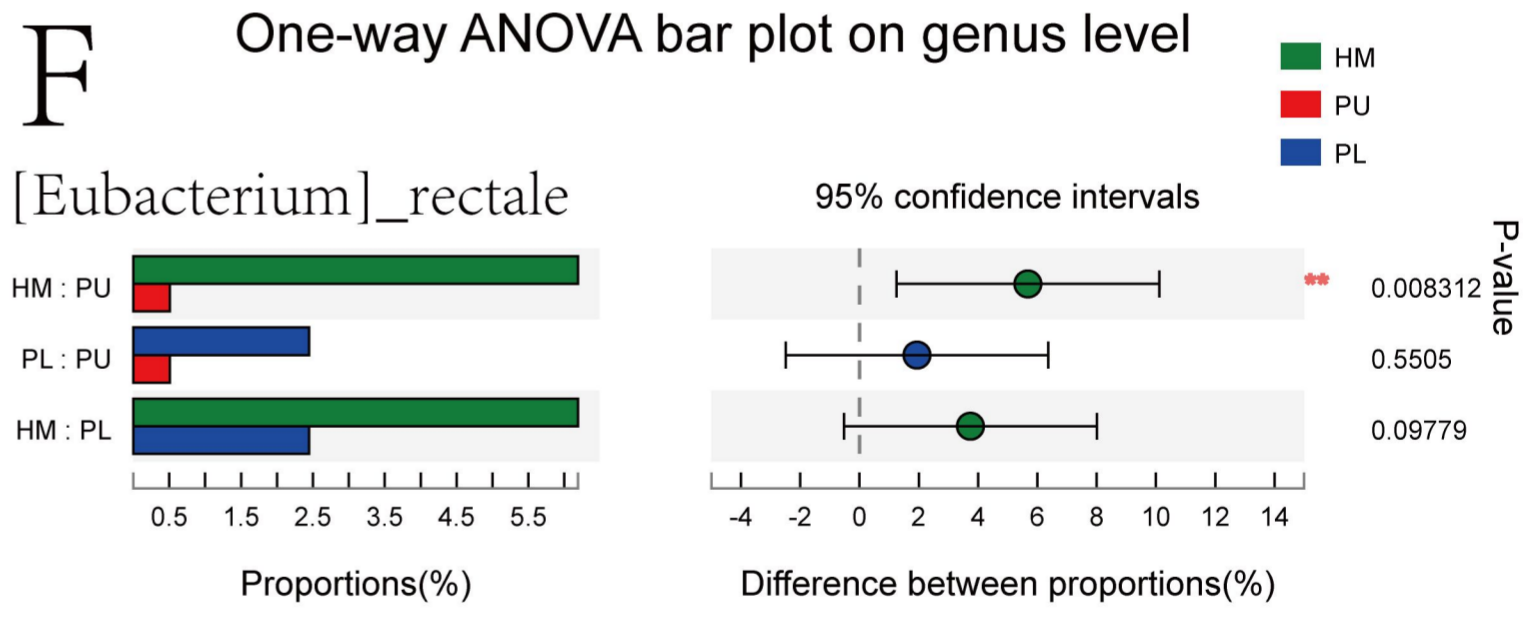
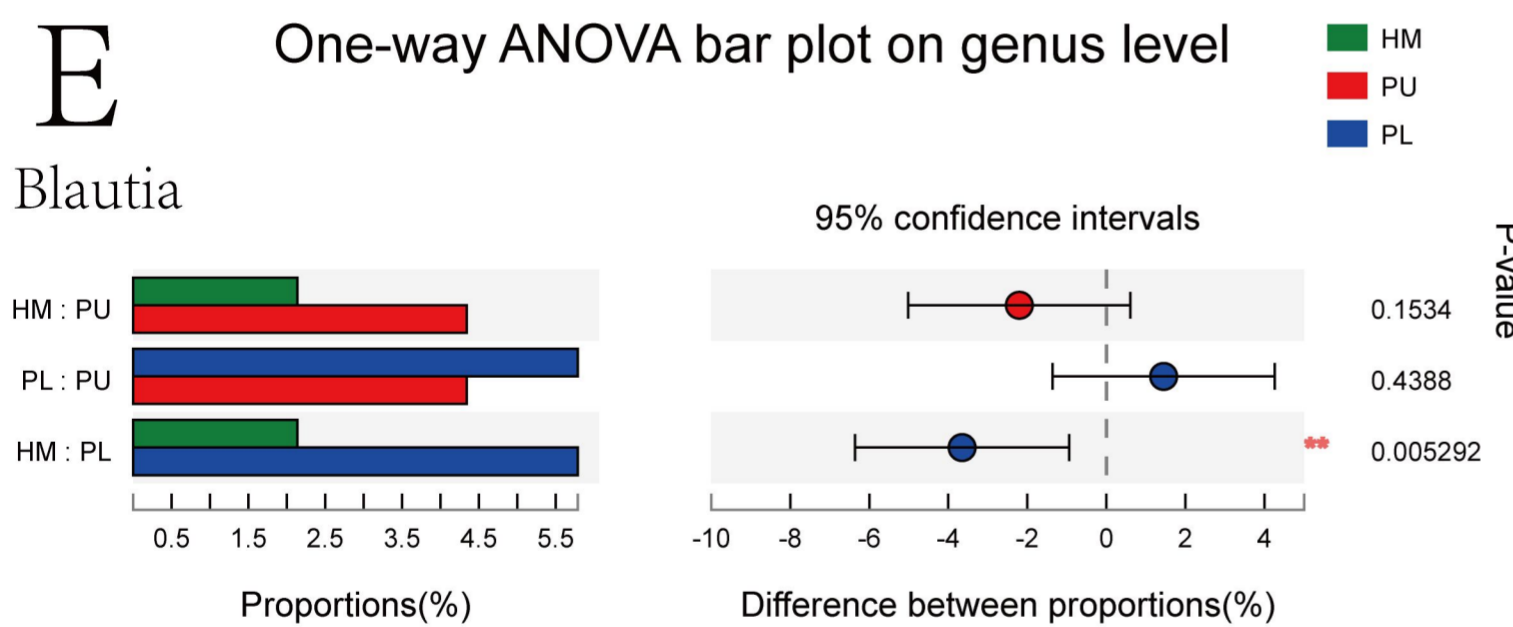
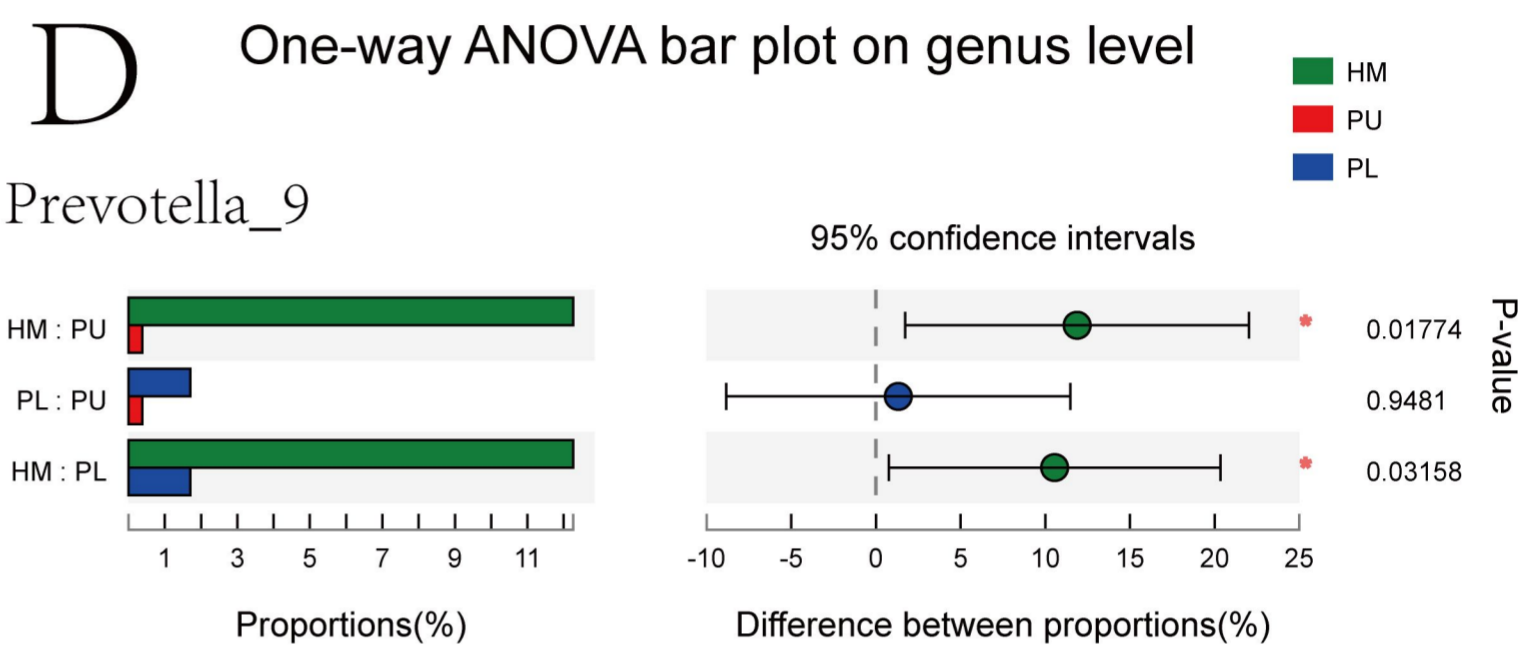
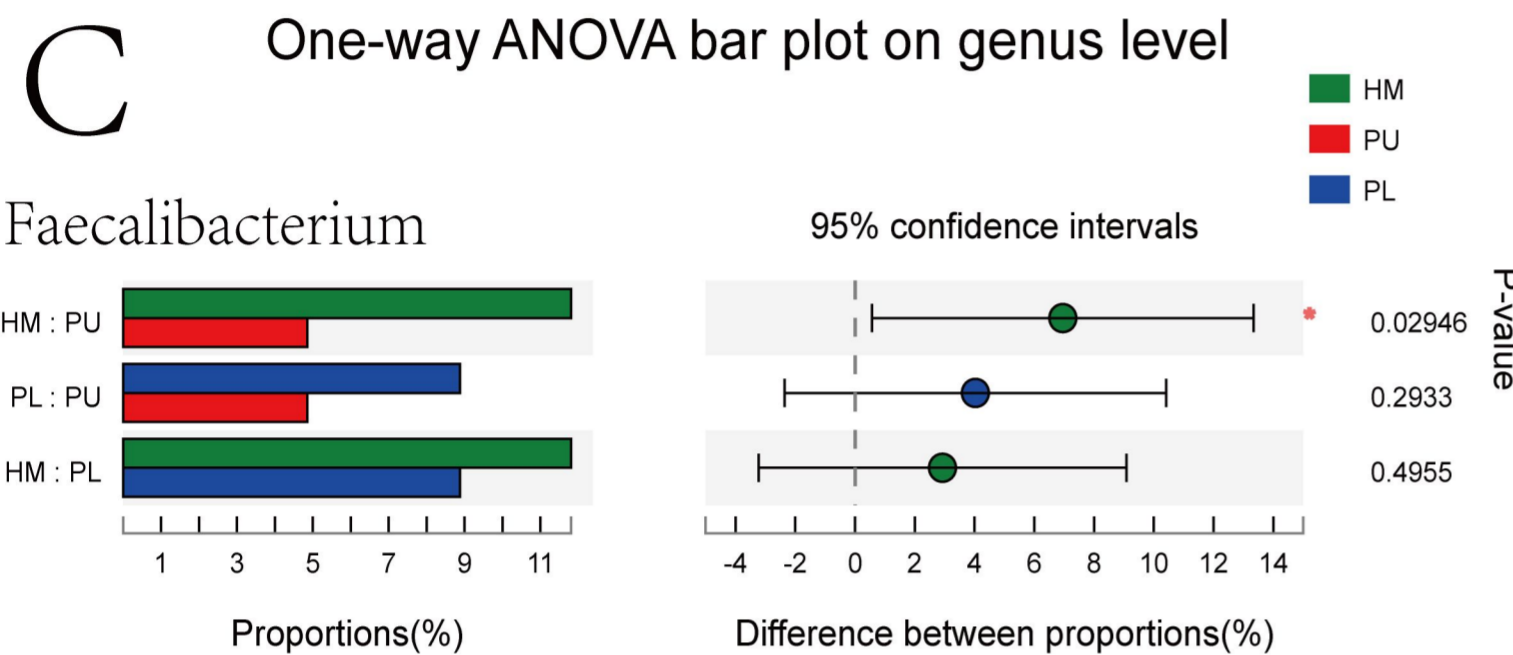
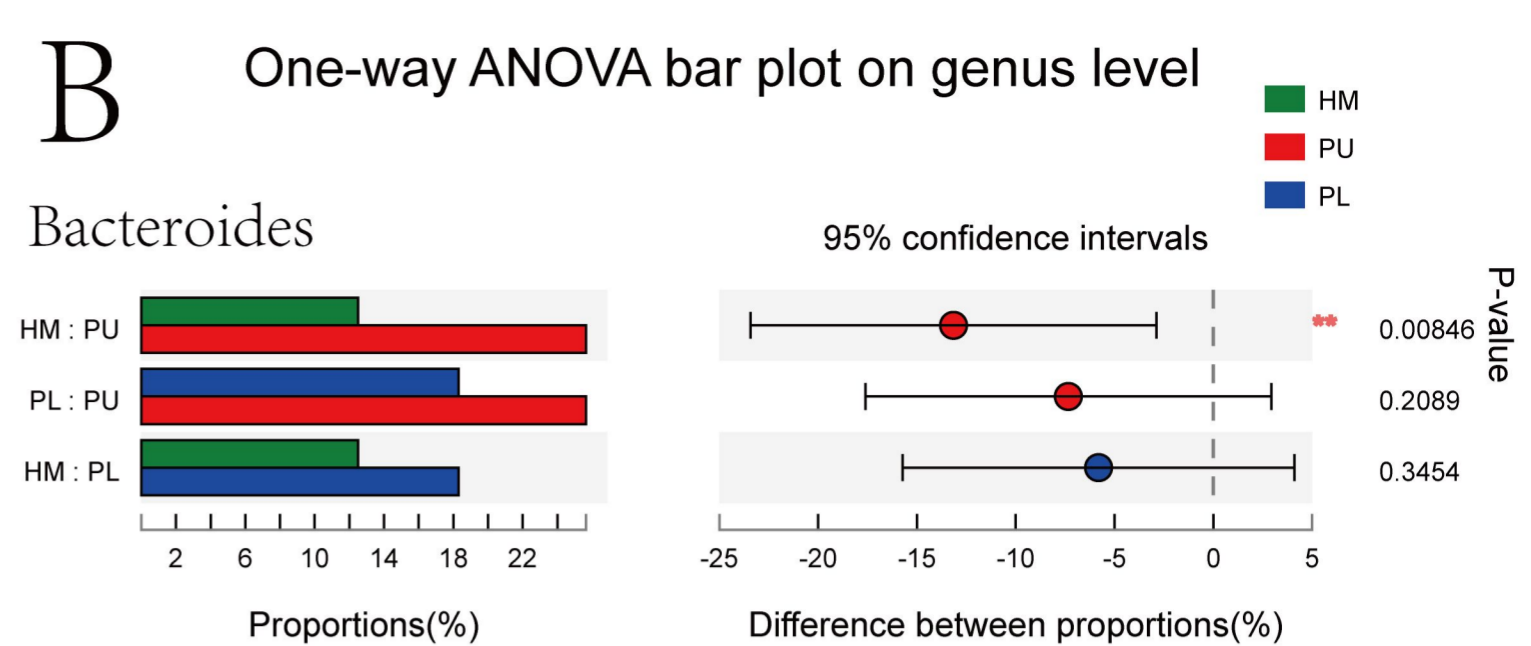
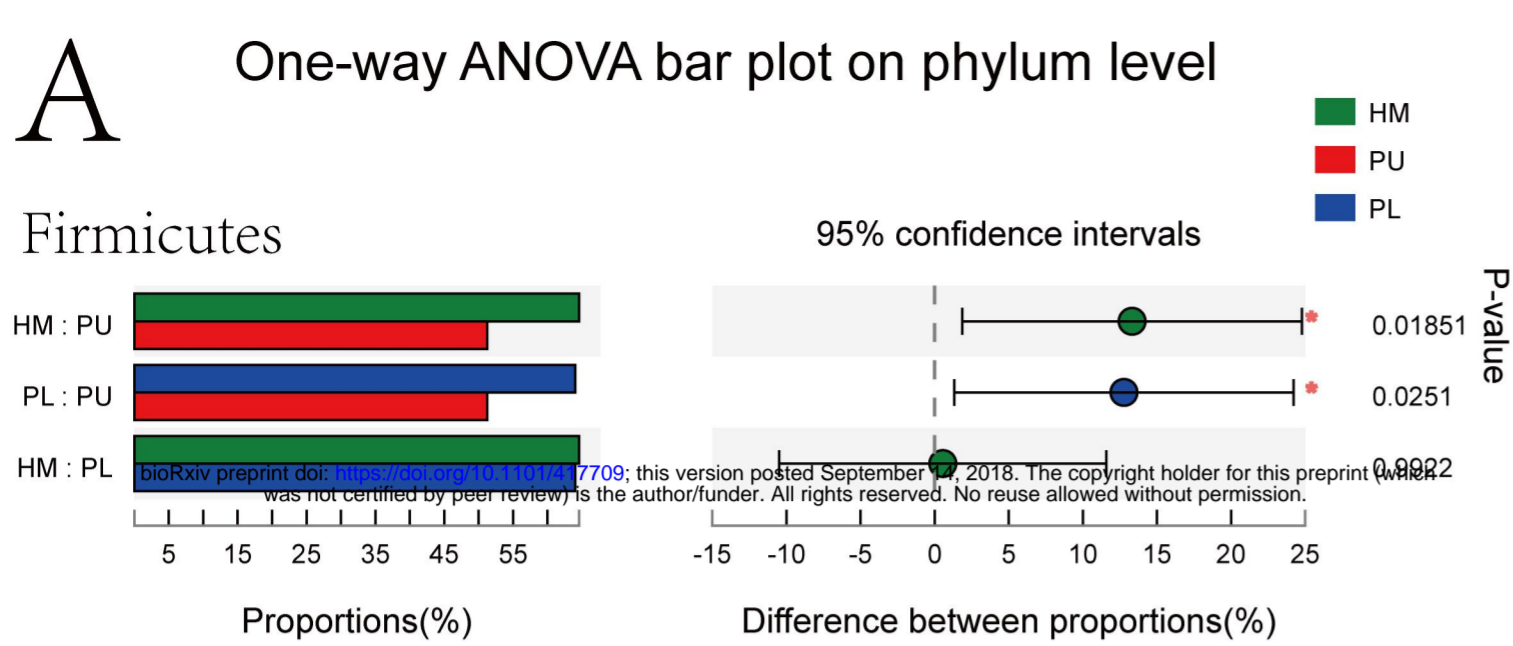




Table 1

Neurogenic Bowel Management table in male Patients with chronic traumatic completed SCI

	SCI-Male n (%)	SCI-cervical n (%)	SCI-thoracic and lumbar n (%)	P
Course	62.5±53.98	69.4±52.72	56.5±54.36	0.449
NBD Scores	10.02±5.11	11.17±5.16	8.7±4.72	0.119
Defecation time	35.33±16.766	41.789±19.29	30±13.94	0.026
Pathogenesis	Traffic accident 16 (37.2%) Bruised by heavy object 9 (20.9%) Falling down 9(20.9%) Other causes 9(20.9%)	Traffic accident 10 (50%) Bruised by heavy object 4(20%) Falling down 3(15%) Other causes 3(15%)	Traffic accident 6 (26.1%) Bruised by heavy object 5 (21.7%) Falling down 6 (26.1%) Other causes 6 (26.1%)	
Frequency of bowel care	Once a day: 17 (39.5%) Not daily but more than twice every week: 26 (60.5%)	Once daily: 5 (25%) Not daily but more than twice every week 15 (75%)	Once daily: 12 (52.2%) Not daily but more than twice every week 11 (47.8%)	
Main techniques for faecal evacuation	Suppository 38 (88.4%) Digital stimulation 7 (16.3%) Manual evacuation 10 (23.3%) Spontaneous 2 (4.7%)	Suppository 20 (100%) Digital stimulation 6 (30%) Manual evacuation 8 (40%)	Suppository 18 (78.3%) Digital stimulation 1 (2.3%) Manual evacuation 2 (4.6%) Spontaneous 2 (4.6%)	
Supplementary interventions	Abdominal massage 25 (58.1%) Digital anus-rectal stimulation 21 (48.8%) Digital evacuation 4 (9.3%) taking cathartic drug 4 (9.3%)	Abdominal massage 10 (50%) Digital anus-rectal stimulation 7 (35%) Digital evacuation 2 (10%) taking cathartic drug 1 (5%)	Abdominal massage 15 (65.2%) Digital anus-rectal stimulation 14 (60.9%) Digital evacuation 2 (8.7%) taking cathartic drug 3 (13%)	
Timing of bowel care	Morning 4 (9.3%) Afternoon 27 (62.8%) Evening 9 (20.9%) Inconsistent 3 (7%)	Morning 1 (5%) Afternoon 14 (70%) Evening 5 (25%)	Morning 3 (13%) Afternoon 13 (56.5%) Evening 4 (17.4%) Inconsistent 3 (13%)	
Location during	Bed	Bed	Bed	

evacuation	19 (44.2%) toilet seat 8 (18.6%) Potty chair 16 (37.2%)	12 (60%) toilet seat 2 (10%) Potty chair 6 (30%)	7 (30.4%) toilet seat 6 (26.1%) Potty chair 10 (43.5%)	
Degree of assistance needed	Need all help 23 (53.5%) Need partial help 10 (23.3%) Independent completion 9 (20.9%) Need special help 1 (2.3%)	Need all help 19 (95%) Need special help 1 (5%)	Need all help 4 (17.4%) Need partial help 10 (43.5%) Independent completion 9 (39.1%)	
Abdominal discomfort	27 (62.8%)	12 (60%)	15 (65.2%)	
Constipation	29 (67.4%)	14 (70%)	15 (65.2%)	
Bloating symptom	32 (74.4%)	16 (80%)	16 (69.6%)	
Flatus incontinence	38 (88.4%)	18 (90%)	20 (87%)	
Lifestyle alteration due to NBD	Major impact 25 (58.1%) Some impact 15 (43.9%) Little impact 3 (7%)	Major impact 12 (60%) Some impact 6 (30%) Little impact 2 (10%)	Major impact 13 (56.5%) Some impact 9 (39.1%) Little impact 1 (4.3%)	
Top 3 complication desired to be solved	Neurogenic bowel dysfunction 42 (97.7%) neurogenic bladder 36 (83.7%) Sexual Dysfunction 19 (44.2%) spasm 14 (32.6%) neuralgia 8 (18.6%)	Neurogenic bowel dysfunction 19 (95%) neurogenic bladder 16 (80%) Sexual Dysfunction 8 (40%) Spasm 7 (35%) neuralgia 3 (15%)	Neurogenic bowel dysfunction 23 (100%) neurogenic bladder 20 (87%) Sexual Dysfunction 11 (47.8%) Spasm 7 (30.4%) neuralgia 5 (21.7%)	

Table2 Demographics and serum biomarkers between male healthy and patients with chronic traumatic completed SCI

	Health male	SCI-Male	P
N	23	43	
AGE	40±9.03	39.9±10.57	0.998
BMI	24.8±2.677	23.11±2.876	0.022
ALT	26.791±16.367	26.2±19.303	0.903
AST	23.848±17.097	21±9.8	0.429
GLU	4.343±0.528	5.266±1.964	0.033
TG	1.436±1.319	1.928±1.207	0.137
TCHO	3.695±0.794	4.217±1.005	0.038
HDL	0.9152±0.2091	0.917±0.163	0.974
LDL	2.177±0.596	2.617±0.701	0.005
UREA	4.416±1.224	4.403±1.14	0.966
CR	64.3±12.701	60.7±11.8	0.265
UA	309±69.81	378.1±64.93	0.001



Table3 Demographics and serum biomarkers male Patients with chronic traumatic completed SCI

	SCI-Male	Sci-cervical	Sci-thoracolumbar	P
N	43	20	23	
AGE	39.9±10.57	41.5±8.30	38.5±12.04	0.369
BMI	23.11±2.876	23.586±3.35	22.697±2.31	<0.001
ALT	26.2±19.303	23.09±11.04	28.16±24.132	0.487
AST	21±9.8	21±9.32	22±10.2	0.796
GLU	5.266±1.964	5.766±2.68	4.83±0.747	0.125
TG	1.928±1.207	2.0325±1.259	1.837±1.15	0.607
TCHO	4.217±1.005	4.072±1.067	4.34±0.93	0.39
HDL	0.917±0.163	0.846±0.137	0.979±0.159	0.007
LDL	2.617±0.701	2.68±0.658	2.69±0.74	0.965
UREA	4.403±1.14	3.956±0.975	4.791±1.13	0.016
CR	60.7±11.8	61.2±11	60.3±12.3	0.815
UA	378.1±64.93	380.75±60.99	375.7±68.08	0.806
CRP	7.78±10.31	11.2±13.45	4.79±4.674	0.042