Theory of cytoskeletal reorganization during crosslinker-mediated mitotic spindle assembly

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Abstract

Cells grow, move, and respond to outside stimuli by large-scale cytoskeletal reorganization. A prototypical example of cytoskeletal remodeling is mitotic spindle assembly, where microtubules nucleate, undergo dynamic instability, bundle, and organize into a bipolar spindle. Key mechanisms of this process include regulated filament polymerization, crosslinking, and motor-protein activity. Remarkably, fission yeast can assemble a bipolar spindle using only passive crosslinkers. We develop a torque-balance model that describes this reorganization due to dynamic microtubule bundles, spindle-pole bodies, the nuclear envelope, and crosslinkers to predict spindle-assembly dynamics. We compare these results to those obtained with kinetic Monte Carlo-Brownian dynamics simulations, which introduce crosslinker-binding kinetics and other stochastic effects. Our results show that rapid crosslinker reorganization to MT overlaps facilitates crosslinker-driven spindle assembly, a testable prediction for future experiments. Combining these two modeling techniques, we illustrate a general method for studying cytoskeletal network reorganization.

Introduction

Cell survival depends on cells' ability to divide, move, grow, and respond to changing conditions, biological functions enabled by flexible and rapid remodeling of the cellular cytoskeleton. Cytoskeletal remodeling is essential for polarized growth, both of single cells [1,2] and tissues [3]. During cell crawling and adhesion, turnover of the actin and microtubule cytoskeletons is tuned by signaling events [4,5]. Large-scale cellular volume changes required for phagocytosis, exocytosis, and endocytosis require actin remodeling [6]. Proper organization and localization of organelles, including mitochondria [7,8] and the endoplasmic reticulum [9], depend on dynamic interactions with the cytoskeleton. During mitosis and cytokinesis, the cytoskeleton undergoes large rearrangements to construct the mitotic spindle [10] and cytokinetic ring [6]. Given the ubiquity of cytoskeletal remodeling for cellular behavior, it is not surprising that aberrant cytoskeletal dynamics or regulation are associated with many diseases, including cancer and developmental defects. The ability of the cytoskeleton to undergo rapid and large structural rearrangements is therefore of broad importance in biology.

The dynamic reorganization of cytoskeletal assemblies is facilitated by a small number of building blocks—filaments, molecular motors, crosslinkers, and associated proteins—which work in concert to generate force [4]. Cytoskeletal filaments, including microtubules (MTs) and actin, undergo dynamic cycles of polymerization and depolymerization that allow rapid turnover. Filament nucleation is controlled by cellular factors (such as γ-tubulin for MTs [10] and the Arp2/3 complex for actin [6]), which spatiotemporally tune filament localization. Crosslinking proteins, including diverse actin crosslinkers [5, 11] and the Ase1/PRC1/MAP65 family of MT crosslinkers [12–18], bundle cytoskeletal filaments to organize higher-order assemblies. These proteins can also have a preferred polarity for crosslinking parallel or antiparallel filament pairs [5, 13]. Motor proteins such as kinesins and myosins can walk on filaments to transport cargo, or crosslink and slide filaments to reorganize them [4]. MT- and actin-binding proteins often regulate filament length and dynamics, allowing further control of cytoskeletal architecture [19–23]. While the flexible, dynamic nature of the cytoskeleton is essential for its biological function, we currently lack predictive understanding of cytoskeletal reorganization.

Mathematical modeling of cytoskeletal systems can reveal molecular mechanisms essential for self-assembly, give insight into the physical requirements for a given behavior, and can be dissected in more detail than is possible in experiments. For example, mathematical modeling has been used to study traveling waves of actin polymerization and protrusions in cell motility [24, 25], cortical MT organization in plants [26–29], mitosis [30, 31] and cytokinesis [32, 33]. A frontier in cytoskeletal modeling is the development of general methods to handle three-dimensional systems in which large structural rearrangements occur. Early mathematical modeling of the cytoskeleton focused on one-dimensional problems such as muscle contraction [34–36] and mitotic-spindle length regulation [37–45]. However, higher-dimensional models are required to capture significant rearrangements, such as the actin protrusions of motile cells, cytokinetic ring formation, and cell cleavage.

A prototypical example of cytoskeletal reorganization is mitotic spindle assembly, during which spindle MTs reorganize to form a bipolar structure as the spindle poles separate (Figure 1) [46]. This requires significant structural rearrangement, even in yeasts, which enter mitosis with side-by-side spindle-pole bodies (SPBs), the nucleating centers for spindle MTs (Figure 1B) [47, 48]. In the closed mitosis of yeasts, the SPBs are embedded in the nuclear envelope and therefore undergo constrained motion [49]. Steric interactions between MTs, SPBs, and the nuclear envelope, along with fluid drag from the nucleoplasm, resist large-scale rearrangement of spindle components [50, 51]. Motor proteins and crosslinkers that bundle and slide MTs are key force generators for spindle assembly that create, extend, and stabilize MT bundles [13, 17, 40, 47, 48, 50, 52–62].

In contrast to the previously established importance of molecular motors in spindle assembly, recent work showed that fission-yeast spindles can assemble in the absence of mitotic motors if the passive crosslinking protein Ase1 is present. In most organisms, kinesin-5 motors that crosslink antiparallel MTs and slide them apart are essential for spindle-pole separation and the establishment of a bipolar spindle [47, 48, 52, 63–69]. Therefore, it was surprising that in fission yeast, kinesin-5 mutants become viable when the kinesin-14s are deleted, because spindle assembly is rescued [53, 70, 71]. In fact, spindle assembly can occur with all mitotic motors deleted or inhibited [51, 72]. In this context, the MT-crosslinking protein Ase1 is essential for spindle assembly [51, 72]. Our previous modeling work showed that antiparallel crosslinking by Ase1 and the stabilization of crosslinked MT dynamics are sufficient for model bipolar spindle assembly in the absence of motors [51]. Related previous theory has studied mitotic MT bundling by motors and crosslinkers [73, 74]. Here, we develop new theory to understand the minimal requirements for spindle structural reorganization and bipolar spindle assembly and

Figure 1: Model schematic. A, Ase1 crosslinker (left) and mitotic spindle (right), including spindle-pole bodies (SPBs, blue), MTs (purple and green), and Ase1 (black). B, Steps of spindle assembly starting from side-by-side SPBs (left), which may lead to a kinetically trapped aberrant state (right, top) or a bipolar spindle (right, bottom)

make quantitative predictions for experiments.

In order to describe the requirements for crosslinkers to align spindle MTs into a bipolar state, we developed a minimal model that accounts for rotation generated by MT polymerization, crosslinkers, drag, and steric interactions. We find that spindle assembly is sensitive to the initial angle at which MT bundles crosslink: smaller initial angle and increased SPB separation facilitates spindle bipolarity. The model we develop includes the general features of dynamic cytoskeletal filaments, forces and torques generated by crosslinkers, friction, and steric interactions. Therefore, this torque-balance model can be applied generally to model crosslinked filament networks driven by polymerization dynamics, contraction, extension, and filament alignment.

Results

Minimal model of microtubule reorientation during spindle assembly

The forces and torques that drive cytoskeletal reorganization can be modeled and simulated using low-Reynolds-number dynamics and statistical mechanics. To create a predictive, tractable model of spindle

assembly, we considered forces from interactions of MT bundles, SPBs, the nuclear envelope, and the nucleoplasm (Figures 1, 2, S1). MTs are nucleated by SPBs and subsequently bundled by crosslinkers. While in principle multiple bundles may be nucleated from one SPB, there is typically one dominant bundle. SPBs are embedded in the nuclear envelope, which is roughly spherical and confines the SPBs to a position on that spherical shell. Motion of the SPB in the shell encounters a viscous drag (Figures 2, S1). The shell prevents MT bundles and other elements of the nucleoplasm from exiting the nucleus. Nucleoplasmic viscosity produces a drag force that opposes the lateral motion of MT bundles, and controls crosslinker diffusion (Figures 2A).

As an MT bundle grows, steric interactions with the nuclear envelope cause the bundle end to slide along the edge of the nucleus, which moves the bundle. We assume that the longest MT within the bundle touches the nuclear envelope, and that MT ends slide freely along the surface of the nuclear envelope, neglecting friction in sliding studied previously [75–77]. MTs undergo dynamic instability with polymerization speed v_p and depolymerization speed v_d. However, within a bundle, we neglect single-MT catastrophe and consider an average bundle growth speed v_pavg of

$$v_\mathrm{pavg} = \frac{f_\mathrm{r} v_\mathrm{p} - f_\mathrm{c} v_\mathrm{s}}{f_\mathrm{r} + f_\mathrm{c}} > 0, \tag{1}$$

where f_r and f_c are the MT rescue and catastrophe frequencies [78]. MT polymerization speed slows in response to compressive force along the MT axis, which we model as in previous work (Equation S7) [79]

$$v_\mathrm{p} = \frac{v_\mathrm{o}}{\eta - 1}(\eta^{1 - L/2R} - 1), \tag{2}$$

where $v_\mathrm{o} = v_\mathrm{p}(F_\mathrm{w,\parallel} = 0)$ is the zero-force polymerization speed and the tubulin association constant $\eta = k_\mathrm{2,on}/k_\mathrm{2,off} = \exp(\delta F_\mathrm{s}/k_\mathrm{B} T N)$ where δ is the tubulin dimer length of 8 nm, F_s is the force at which MT polymerization stalls, $N = 13$ is the number of protofilaments in an MT, k_B is the Boltzmann constant, and T is the temperature.

The nuclear envelope exerts equal and opposite radially inward forces on MT bundle ends, which produce a net force that moves the bundle center of mass toward the center of the nuclear envelope, but no net torque (Figure S1B). MT bundle minus ends interact with SPBs; this coupling to SPBs with their large drag breaks the symmetry between plus- and minus-ends and produces a net torque about the bundle centers. SPB drag on the minus ends of MT bundles tends to rotate the bundles away from alignment, leading to a torque

$$\tau_\mathrm{spb}(\dot{\phi}, L) = -\gamma_\mathrm{spb} \frac{L}{4} \left(\dot{\phi} \sqrt{R^2 - (L/2)^2} - \dot{L} \right), \tag{3}$$

where γ_spb is the friction coefficient of SPBs, R is the radius of the nuclear envelope, ϕ is the bundle crossing angle, and $\dot{\phi}$ the time derivative of ϕ (Figures 2A, S1A). The magnitude of the SPB torque τ_spb is proportional to γ_spb, the average MT polymerization speed $\dot{L}$, and bundle length L (Equation S13). However, longer MTs experience a greater parallel force (along the MT axis) from the nuclear envelope than perpendicular force (perpendicular to the MT axis), which slows MT polymerization, reduces SPB velocity, and therefore lowers the antialigning torque (Figure S1).

We calculate the average force and torque exerted by crosslinkers on MT bundles by considering the statistical mechanics of crosslinker binding, unbinding, and stretching/compression in a two-dimensional geometry. Because crosslinker binding and unbinding kinetics are relatively fast (time scale of seconds)

compared to spindle assembly (time scale of minutes), we make a quasi-steady-state assumption that crosslinker binding equilibrates given the instantaneous MT bundle positions. Therefore, we can determine crosslinker-induced forces and torques on MTs by computing the grand partition function and its derivative of crosslinker binding. The single-crosslink partition function between two filaments is the integral of the Boltzmann factor of its bound energy $e^{\beta u_{cl}}$, where u_{cl} is the crosslinker binding free energy, over all possible binding configurations [50, 51]

$$q(\phi, L) = c \int_0^{L/2} \int_0^{L/2} dr_1 dr_2 e^{-\frac{\beta k}{2}(h(r_1, r_2, \phi) - h_{cl})^2}, \tag{4}$$

where c is the crosslinker concentration, the integrals over crosslinker endpoints r_1, r_2 extend over the filament length from 0 to L, k is the crosslinker spring constant, ϕ is the angle between the two filaments, $h(r_1, r_2, \phi) = \sqrt{r_1^2 + r_2^2 - 2r_1 r_2 \cos(\phi)}$ is the crosslinker length when the endpoints are at r_1 and r_2 along the two filaments, h_{cl} is the crosslinker rest length (at which it is neither stretched nor compressed), and $\beta = 1/(k_B T)$ is the inverse temperature (Figure 2B, Equation S17). We assume that crosslinkers do not interact with each other, which motivates the use of the grand canonical ensemble of a non-interacting gas with partition function

$$\Xi = \sum_{n=0}^{\infty} \frac{1}{n!} z^n q^n = e^{zq}. \tag{5}$$

Here n is the number of bound crosslinkers and z is the fugacity $e^{\mu_{o,cl}\beta}$, where $\mu_{o,cl}$ is the chemical potential of crosslinkers binding to the filaments. We can then determine crosslinker force and torque from derivatives of the grand potential

$$\Phi = \frac{-1}{\beta} \ln(\Xi) = \frac{-zq}{\beta}. \tag{6}$$

For example, the torque from all crosslinkers on MT bundles is

$$\tau(\phi, L) = -(\partial_\phi \Phi) = -zkc \sin(\phi) \int_0^L \int_0^L dr_1 dr_2 r_1 r_2 \left(1 - \frac{h_{cl}}{h(r_1, r_2, \phi)}\right) e^{-(k\beta/2)(h(r_1, r_2, \phi) - h_{cl})^2}. \tag{7}$$

Torque due to crosslinkers can promote or oppose antiparallel alignment of the MTs, because the magnitude and direction of the torque depends on MT length and the crossing angle (Figure 2). Crosslinkers can bind above, below, to the left, and to the right of the crossing point. Symmetry allows us to consider only the left/right and above/below cases: the angle in Equation 7 is ϕ for left/right and $\pi - \phi$ for above/below. As expected, a small angle between bundles produces greater crosslinker attachment, and therefore greater torque. Left/right binding typically produces aligning torque, although there can be exceptions when the crosslinkers are too compressed upon binding.

By using the Langevin equations for translational and rotational motion of filaments we can derive a system of integro-differential equations for ϕ and L (Section S1, Equations S21, S22)

$$\dot{\phi}(\dot{L}, L, \phi, t) = \frac{\gamma_{spb} L \dot{L} + 2\tau_{cl,-} - 2\tau_{cl,+}}{2} \gamma_{rot} + \gamma_{spb} L \sqrt{R^2 - (L/2)^2} \tag{8}$$

$$\dot{L}(L) = v_p(\dot{\phi}, \phi, L, t) \tag{9}$$

where $\gamma_{rot}(L)$ is the rotational drag coefficient of a filament of length L [80]. These equations can be solved numerically given initial ϕ_0 and d_0 (Figure 2A, Equation S2). The time evolution predicts the end configuration, either bipolar or aberrant (Figure 2C).

Figure 2: Schematic of and results from the torque-balance model. A, Schematic of forces and torques. MT bundles of length L are attached to SPBs, separated by distance d and crossing angle ϕ. Forces arise from crosslinkers, drag, and confinement within the nuclear envelope. B, Schematic of crosslinking geometry. Two MT bundles of length L cross at angle ϕ, with crosslinker heads binding at distance r_1 and r_2 from the MT bundle ends. C, MT bundle crosslinking angle ϕ as a function of time in the torque-balance model for two initial conditions. Red (top) larger initial crossing angle becomes larger, leading to an aberrant final state. Blue (lower) smaller initial crossing angle decreases, leading to an aligned bipolar spindle. D, Phase diagram of bipolar spindle assembly in the torque-balance model as a function of initial SPB separation and crossing angle. Blue, bipolar spindles form. Red, aberrant spindles form. Parameters are $v_{\mathrm{p}} = 4\ \mu\mathrm{m/min}$, $h_{\mathrm{cl}} = 53$ nm, $k_{\mathrm{cl}} = 0.2055$ pN/nm, $R_{\mathrm{NE}} = 1.375$ nm, $c_{\mathrm{cl}} = 2.88 \times 10^{-4}\ \mathrm{nm}^{-2}$, $z = 1$, $d_o = 0.55\ \mu\mathrm{m}$, $D_{\mathrm{spb}} = 4.5 \times 10^{-4}\ \mu\mathrm{m}^2/\mathrm{sec}$, initial angle $\phi_o = 6.07$ degrees for bipolar final state in panel C, 9.1 degrees for aberrant final state in panel C. See table 1.

Phase diagram of spindle assembly

To compute a phase diagram for spindle assembly as a function of initial conditions, we numerically integrate Equations 8 and 9 (Supplementary Material). We characterize dynamics that reach a bipolar final state by a decreasing crossing angle ϕ at the end of the integration, $d\phi/dt < 0$ (Figures 2C, S2, S3, S4). Aberrant states occur for larger initial crossing angle because the aligning torque from crosslinkers

Figure 3: Kinetic Monte Carlo-Brownian dynamics simulation snapshots, typical final states of simulations, and phase diagram. A, Simulation snapshots of bipolar spindle assembly from an initial condition with adjacent SPBs. Times shown are in minutes, seconds, and tenths of seconds. B, Schematics of final states of simulations. C, Phase diagram of spindle formation failure: red indicates an aberrant end-state, blue the formation of a bipolar structure. Bipolar spindle fraction determined from 24 simulations at each data point. See Movie S1.

is not able to overcome the anti-aligning torque from SPB drag (Figure 2D). For larger SPB separation, longer MTs provide more sites for crosslinker binding, while steric forces from the nuclear envelope are more parallel to the MT axis, slowing polymerization and reducing SPB drag. For MTs that span the nucleus, bipolarity occurs whenever $\phi_o < \pi/2$, since crosslinker aligning torque dominates. In fission-yeast spindle assembly, bundles tend to crosslink initially at short SPB separation [81]. Our results show that model parameters that favor bipolarity for small SPB separation will favor spindle assembly for a range of initial conditions.

Comparison to kinetic Monte Carlo-Brownian dynamics simulations

Our kinetic Monte Carlo-Brownian dynamics (kMC-BD) simulation model includes three-dimensional geometry, multiple sterically interacting MTs, and stochastic effects [50, 51], making it a useful point of comparison with the minimal torque-balance model (Figure 3, Movie S1). In contrast to the torque-balance model, kMC-BD simulations with the same parameters do not always end in the same state because of stochastic effects, including thermal motion, varying initial conditions, and binding kinetics. However, these stochastic effects also allow spindles to escape aberrant states and become bipolar (Figure 3B). The three-dimensional geometry adds another degree of freedom, increasing the number of ways to escape from kinetic traps. In previous work, we showed that the kMC-BD model leads to spindle self-assembly in simulations both with [50] and without [51] motor proteins. In the previous study of the model without motor proteins, we found that simulated spindles can form with crosslinkers only if crosslinkers stabilize the dynamics of crosslinked MTs. Here we extend previous modeling results to interrogate the requirements for crosslinker-mediated spindle assembly.

In our kMC-BD model, the total crosslinker number is fixed, and we compute the location of individual molecules, which allows crosslinker forces to fluctuate. Therefore, the overall force that crosslinkers can exert is constrained by the total crosslinker number. This means that crosslinkers must diffuse or

unbind and rebind to apply force to recently overlapped MTs, processes that are modulated by thermal fluctuations, force, and MT configuration. The kMC-BD model becomes more similar to the quasi-steady approximation used in the torque-balance model for high crosslinker turnover and/or high bound diffusion coefficient. The distribution of crosslinkers can be out of equilibrium, since small random thermal movements of MTs and SPBs occur on short time scales compared to the redistribution of crosslinkers. If out of equilibrium, crosslinkers exert restoring forces that maintain the network configuration despite random forces acting on MTs and SPBs. In this way the stochastic binding kinetics allow kMC-BD simulations to escape kinetic traps but, if slow compared to thermal motion, can increase the strength of kinetic traps and the frequency of aberrant states.

We quantified spindle assembly frequency as the fraction of simulations that end in a stable bipolar state after 24 minutes of simulated real time [51]. A higher spindle assembly frequency occurs in the phase diagram of the kMC-BD model (Figure 3C) for parameter sets that give bipolar spindle assembly in the torque-balance model (Figure 2D), demonstrating that the torque-balance model contains the key ingredients that control bipolar spindle assembly in the full kMC-BD model.

Lengthening crosslinkers inhibits bipolar spindle assembly by over-bundling parallel MTs

Crosslinkers have a characteristic length, but it is not well understood whether changes in this length might affect spindle assembly. We assume Ase1 is around 40-55 nm long, based on the length of the human homolog PRC1 (37 nm long [82]) and kinesin-5 crosslinking motors (53 nm long [83]). In our model, crosslinkers are springs, so that stretched crosslinkers pull MTs together, whereas compressed crosslinkers push them apart. Crosslinker length determines the distance from the MT crossing point at which crosslinkers prefer to bind, which affects the amount of splay in MT bundles.

Short crosslinker length inhibits spindle assembly because crosslinkers primarily bundle neighboring MTs, limiting interdigitation of antiparallel MTs (Figure 4, movie S2). MT bundles become tightly bound and are less likely to become interdigitated with MTs from the other SPB, which decreases the strength of the crosslinker aligning torque (Figure 4A, bottom left inset).

Increasing crosslinker length too far allows crosslinkers to interact with more MTs, which negatively impacts spindle assembly in two ways: crosslinking occurs when the SPB separation is small, and MT bundles form aberrant I- and X-shaped spindles (Figure 3B). Longer crosslinkers splay parallel bundles, limiting MT crosslinking near the dynamic ends of the bundles. Crosslinking between splayed bundles tends to trap the SPBs close together, leading to a narrow X- or I-shaped spindle (Figure 4A, insets, movie S2). These effects make I- and X-shaped spindles strong kinetic traps for long crosslinkers.

These results suggested that the defects of the model with long crosslinkers might be rescued by separating SPBs at the start of the simulation. To test this prediction, we began simulations with SPBs separated and allowed MTs and crosslinkers to interact while the SPBs were held in place for 1 second. Spindle assembly frequency increases sharply as initial SPB separation increases above around 1.2 μm, similar to the location of the phase boundary in our torque-balance model (Figure 4B,C).

Increased crosslinker turnover helps spindles escape kinetically trapped aberrant states

Because in previous work we found that crosslinker-only simulated spindles can become stuck in persistent monopolar states [51], we tested whether more rapid crosslinker turnover can promote bipolar spindle assembly by accelerating escape from aberrant states (Figure 5, Movie S3). Increasing turnover

Figure 4: Crosslinker length of 20-60 nm is optimal for spindle assembly. A, Fraction of simulations that assemble a bipolar spindle as a function of crosslinker length. Top, schematic of crosslinker length effects. Inset, simulation snapshots of typical aberrant final states. Red dotted line indicates reference parameter value. B, C, Effects of long crosslinker length on spindle assembly, for crosslinkers of length 140 nm. B, Fraction of simulations that assemble a bipolar spindle as a function of initial SPB separation. C, Spindle assembly phase diagram from the torque-balance model with long crosslinkers. See Movie S2.

does not alter the average number of bound crosslinkers, but increases binding and unbinding (between one- and two-head bound states). With increased turnover, stretched or compressed crosslinkers more rapidly detach and can re-bind in states closer to mechanical equilibrium of the spring. Rapid turnover therefore makes random thermal forces more effective at repositioning MT bundles.

We further examined whether changing the binding and unbinding rates (Figure 5A) individually has similar effects. Increasing the on-rate tends to lock in monopolar states, because it increases the total number of bound crosslinkers. For high binding-rate, nearly all crosslinkers are bound to two MTs, preventing them from reorienting. By contrast, for low binding-rate, crosslinkers can remain with one head bound to an MT while they diffuse, then reform a crosslink at another position. Our results therefore suggest that the rate of crosslinker rearrangement controls the rate of MT rearrangement, and thus the rate of bipolar spindle formation.

Increasing the unbinding rate of doubly-bound crosslinkers facilitates the transition to a bipolar spin-

Figure 5: High crosslinker turnover facilitates spindle assembly. A, Schematic effects of varying crosslinker kinetic rates. Changing turnover (left) changes both on- and off-rates for transitions between one and two heads bound. Changing the on-rate (center) changes the binding rate from one head to two. Changing off rate (right) changes the unbinding rate from two heads to one. B, Fraction of simulations that assemble a bipolar spindle as a function of crosslinker kinetic rates. See Movie S3.

dle by accelerating bundle reconfiguration. However, increasing the unbinding rate too much decreases the average number of crosslinks, leading to a critical value of unbinding rate above which the number of bound crosslinkers is too low to maintain bundle integrity (Figure 5B, Movie S3).

When crosslinker turnover is rapid, the kMC-BD simulation model more closely approximates the torque-balance model's quasi-steady binding approximation. In this limit, crosslinkers rapidly change their configuration to the most statistically probable. In this case, we expect that fluctuations and aligning torque from crosslinkers will allow X-shaped spindles to form a bipolar spindle, given sufficient time. While not every simulated spindle reaches a bipolar state with high crosslinker turnover, the trends match this expectation.

Crosslinker diffusion facilitates escape from kinetic traps by promoting crosslinker redistribution and MT reorientation

One-dimensional diffusion of bound crosslinkers repositions crosslinker heads on the MTs, which suggest that a higher diffusion coefficient might promote escape from aberrant states. Since diffusion is biased by force, when random thermal forces reorient MTs and extend/compress the crosslinkers, diffusion favors crosslinker motion that reduces this force. Therefore, diffusion modulates MT structural

Figure 6: Bound crosslinker diffusion facilitates crosslinker redistribution and spindle assembly. A, Fraction of simulations that assemble a bipolar spindle as a function of bound crosslinker diffusion coefficient. Red dotted line indicates reference parameter value. Top, schematic of bound diffusion coefficient effects. Inset, simulation snapshots of typical aberrant final states. B, Fraction of simulations that assemble a bipolar spindle as a function of crosslinker kinetic rates, in the absence of bound crosslinker diffusion. Bipolar spindle formation is rescued by high crosslinker turnover. C, Schematic showing how high crosslinker turnover can rescue MT reorientation in the absence of bound diffusion. See Movie S4.

rearrangement: slow diffusion tends to inhibit MT reorientation, while fast diffusion tends to promote it (Figure 6, Movie S4).

Increasing the doubly bound diffusion coefficient promotes spindle assembly, similar to the effects of increased turnover, but there are some differences. An increased diffusion coefficient by itself does not release crosslinked MTs, so does not lead to the separated asters seen for high crosslinker unbinding rate. In the opposite limit, when we completely remove doubly-bound crosslinker diffusion, bipolar spindles do not form, because crosslinkers do not rearrange quickly enough. Instead, these simulations produce long-lived I- or X-shaped spindles. In I-shaped spindles, the crosslinked bundles remain close together, since crosslinkers cannot migrate when SPBs fluctuate apart.

The similar increases in spindle assembly frequency for increasing turnover and diffusion suggest that changing turnover might be able to rescue the absence of diffusion: if binding kinetics are sufficiently rapid, one head can unbind and reattach in a different position, redistributing the crosslinkers. Consistent with this, high turnover rescues spindle assembly in simulations lacking bound diffusion (Figure 6B). Similarly, high crosslinker unbinding rate can rescue spindle bipolarity for a narrow range of values before reaching the critical value at which too few crosslinkers remain bound.

Bipolar spindles form most readily when the parallel-antiparallel binding ratio is low but non-zero

Crosslinkers of the Ase1/PRC1/MAP65 family have a binding preference for antiparallel MT overlaps [14, 15, 84], but it has not been determined whether this bias affects crosslinker-mediated bipolar spindle assembly. Previous work has found that crosslinkers in this family favor antiparallel MT binding over parallel by a factor of two to ten [13]. In our model, we implement this effect as a binding enhancement that changes when the angle between MT axes is greater or less than 90 degrees. We then examined whether varying this binding preference affects spindle assembly (Figure 7).

For low parallel-antiparallel binding ratio (α=0–0.01), spindles can form because crosslinkers inhibited in parallel bundling avoid the X-shaped spindle. In this limit, failure of bipolar spindle assembly occurs sometimes because antiparallel bundling for very short MTs from adjacent SPBs leads to trapped monopolar spindles (Figure 7, Movie S5). Parallel crosslinking can promote spindle assembly by forming X-shaped spindles that allow the SPBs to separate, then interdigitate.

For low-intermediate parallel-antiparallel binding ratio (α=0.01–0.1), predominantly antiparallel binding with some parallel binding allows X-shaped spindles to transition to bipolar spindles, leading to the highest frequency of spindle assembly observed. Monopolar spindles tend to be short-lived, because parallel binding allows crosslinkers to migrate away from short antiparallel overlaps, promoting SPB separation (Figure 7C, Movie S5). Bundles then either break up to form a bipolar spindle, or rotate until the bundles are antiparallel.

For larger parallel-antiparallel binding ratio (α=0.1–0.3), the modest increase in parallel crosslinking favors the X-shaped spindle, inhibiting bipolarity. The lifetime of the X-shaped spindle increases with α. Similarly, when parallel and antiparallel binding are equally likely (α=1), X-shaped spindles typically form. However, random thermal forces can occasionally lower the angle between bundles, allowing antiparallel crosslinks and a bipolar spindle to form. Fully extended bundles at right angles to each other have, on average, balanced torques promoting alignment and anti-alignment. This balance can be broken if a thermal fluctuation rotates bundles toward antiparallel alignment.

For purely antiparallel binding ($\alpha = 0$), increasing crosslinker turnover has a dramatic effect (Figure 7B). If turnover is sufficiently high, purely antiparallel-binding crosslinkers produce bipolar spindles for nearly every simulation. Varying turnover and unbinding rate show similar trends, except for the

Figure 7: The crosslinker binding preference for antiparallel MTs promotes spindle assembly. A, Fraction of simulations that assemble a bipolar spindle as a function of the parallel-to-antiparallel binding ratio α. The red dotted line indicates the reference parameter value. Top, schematic of binding ratio effects. Inset, simulation snapshots of typical aberrant final states. B, Fraction of simulations that assemble a bipolar spindle as a function of crosslinker kinetic rates in the absence of parallel binding. C, Schematic of spindle assembly pathway for the optimal binding ratio. MTs crosslink parallel bundles, SPBs separate, then bundles interdigitate at shallower angle to form a bipolar spindle. See Movie S5.

failure above the critical value for the unbinding rate at which most crosslinkers unbind. As for the other model perturbations discussed above, rapid rearrangement of crosslinkers allows spindles to escape kinetic traps and become bipolar.

Spindles typically transition from X-shaped to bipolar via two pathways: reconfiguration of crosslinkers, and aligning torques between bundled MTs (Figure 7C, Movie S5). Slow reconfiguration occurs when MT bundles move into antiparallel alignment due to thermal motion and crosslinker torque. The aligning torque pathway occurs more often when α is small, so that single polar MTs are common. When single MTs escape the main bundles, they can crosslink with MTs from the other SPB at a relatively shallow angle. This allows the MTs to push against the nuclear envelope and separate SPBs until the main MT bundles also align into a fully bipolar spindle.

Discussion

Here we have developed a physical theory for cytoskeletal reorganization during fission-yeast mitotic spindle assembly (Figure 1) that incorporates the key ingredients of filament polymerization and depolymerization, crosslinking, steric interactions, and drag (Figure 2). Comparison of our minimal torque-balance model to full kMC-BD simulations that incorporate all stochastic effects (Figure 3, Movie S1) shows good agreement, demonstrating that the torque-balance model can illuminate the physical constraints on crosslinker-mediated spindle assembly. We studied specific individual perturbations to crosslinker length (Figure 4, Movie S2), binding kinetics (Figure 5, Movie S3), bound diffusion (Figure 6, Movie S4), and parallel versus antiparallel binding preference(Figure 7, Movie S5). The results demonstrate that crosslinkers that favor binding to antiparallel MT pairs and rapid redistribution of crosslinkers are crucial for bipolar spindle assembly from initially side-by-side SPBs.

In crosslinker-mediated spindle assembly, the crosslinker binding preference to antiparallel rather than parallel MT pairs is important for spindle assembly: bipolar spindles have more possible binding states between antiparallel MT pairs, so the antiparallel binding preference energetically favors bipolarity. However, this state must still be kinetically reachable from an initial condition in which spindle MTs are predominantly parallel. Therefore, crosslinker-mediated spindle assembly is vulnerable to kinetic traps at key stages of assembly (Figure 3). If antiparallel crosslinking predominates when SPBs are close to each other and crosslinker redistribution is too slow, spindles become trapped in a monopolar state. If parallel crosslinking predominates and crosslinkers either cannot bind to antiparallel MTs or redistribute too slowly, the frozen parallel bundles of X- or I-shaped spindles predominate. If too few crosslinkers are present, the spindle can fall apart into separated asters.

Spindles avoid or escape these kinetic traps when aligning torques overcome anti-aligning torques early in spindle assembly. The torque-balance model demonstrates that aligning torques are strongest for separated SPBs and low crossing angle between MTs. In some cases, aligning torques must overcome the anti-aligning torques early in our model simulations if the spindle is to become bipolar. Important stochastic effects that promote escape from kinetic traps include crosslinker redistribution and random thermal forces. Crosslinker redistribution is faster when crosslinker binding kinetics and/or bound diffusion are more rapid. Remarkably, increasing crosslinker turnover or diffusion can rescue defects in bipolar spindle assembly caused by crosslinkers of non-optimal length or exclusively antiparallel crosslinker binding.

The modeling techniques we use are generally applicable to cytoskeletal reorganization in which crosslinkers facilitate reorientation of filaments. This area is a frontier of cytoskeletal theory and modeling, as the field confronts more challenging three-dimensional problems that were difficult to address computationally until recently. Despite the complex dynamics and large-scale filament rearrangements

Parameter	Symbol	Value	Notes
Crosslinker spring constant	k_{cl}	0.2 pN nm^{-1}	[17]
Crosslinker length	h_{cl}	53 nm	[17, 51]
Crosslinker fugacity	z_{cl}	1.0	Chosen to have $\sim$20 crosslinkers per MT at full overlap
Crosslinker binding affinity	c_{cl}	3×10^{-4} nm^{-2}	Chosen to have $\sim$20 crosslinkers per MT at full overlap
Nuclear envelope radius	R_{NE}	1.375 μm	[85]
SPB diffusion coefficient	D_{spb}	4×10^{-4} μm^2 sec^{-1}	[50]
MT stall force	$F_{stall,MT}$	14.8 pN	[79]
Base MT polymerization rate	v_o	4 μm min^{-1}	[51]
Time step	δt	0.0358 sec	Chosen for numerical stability

Table 1: Table of parameter values used in the torque-balance and kMC-BD simulations.

that occur during bipolar spindle assembly, our work shows that the key physical effects can be understood both in detailed simulations and a minimal torque-balance model. The principles and modeling methods we describe are broadly applicable to cytoskeletal systems.

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Author contributions. ARL, MAG, and MDB formulated the torque-balance model and code; ARL, CE, and MAG wrote the kMC-BD simulation code; ARL ran kMC-BD simulations; ARL, CE and MDB analyzed model results; ARL and MDB drafted the manuscript; ARL, CE, MAG, and MDB edited the manuscript.

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Data and materials availability. All data and computer code for this study are available on request from the authors.

References and Notes

[1] W. James Nelson. Adaptation of core mechanisms to generate cell polarity. *Nature*, 422(6933):766–774, April 2003.

[2] Reinhard Fischer, Nadine Zekert, and Norio Takeshita. Polarized growth in fungi – interplay between the cytoskeleton, positional markers and membrane domains. *Molecular Microbiology*, 68(4):813–826, 2008.

[3] David M. Bryant and Keith E. Mostov. From cells to organs: Building polarized tissue. *Nature reviews. Molecular cell biology*, 9(11):887–901, November 2008.

[4] D. Bray. *Cell Movements: From Molecules to Motility*. Routledge, 2000.

[5] Laurent Blanchoin, Rajaa Boujemaa-Paterski, Cécile Sykes, and Julie Plastino. Actin Dynamics, Architecture, and Mechanics in Cell Motility. *Physiological Reviews*, 94(1):235–263, January 2014.

[6] Thomas D. Pollard and John A. Cooper. Actin, a Central Player in Cell Shape and Movement. *Science*, 326(5957):1208–1212, November 2009.

[7] A. Rendon, D. Jung, and V. Jancsik. Interaction of microtubules and microtubule-associated proteins (MAPs) with rat brain mitochondria. *The Biochemical Journal*, 269(2):555–556, July 1990.

[8] Chuanhai Fu, Deeptee Jain, Judite Costa, Guilhem Velve-Casquillas, and Phong T. Tran. Mmb1p binds mitochondria to dynamic microtubules. *Current biology: CB*, 21(17):1431–1439, September 2011.

[9] C. M. Waterman-Storer and E. D. Salmon. Endoplasmic reticulum membrane tubules are distributed by microtubules in living cells using three distinct mechanisms. *Current biology: CB*, 8(14):798–806, July 1998.

[10] J. Richard McIntosh, Maxim I. Molodtsov, and Fazly I. Ataullakhanov. Biophysics of mitosis. *Quarterly Reviews of Biophysics*, 45(02):147–207, 2012.

[11] Yiider Tseng, Thomas P. Kole, Jerry S. H. Lee, Elena Fedorov, Steven C. Almo, Benjamin W. Schafer, and Denis Wirtz. How actin crosslinking and bundling proteins cooperate to generate an enhanced cell mechanical response. *Biochemical and Biophysical Research Communications*, 334(1):183–192, August 2005.

[12] Isabelle Loïodice, Jayme Staub, Thanuja Gangi Setty, Nam-Phuong T. Nguyen, Anne Paoletti, and P. T. Tran. Ase1p Organizes Antiparallel Microtubule Arrays during Interphase and Mitosis in Fission Yeast. *Molecular Biology of the Cell*, 16(4):1756–1768, January 2005.

[13] Marcel E. Janson, Rose Loughlin, Isabelle Loïodice, Chuanhai Fu, Damian Brunner, François J. Nédélec, and Phong T. Tran. Crosslinkers and Motors Organize Dynamic Microtubules to Form Stable Bipolar Arrays in Fission Yeast. *Cell*, 128(2):357–368, January 2007.

[14] Radhika Subramanian, Elizabeth M. Wilson-Kubalek, Christopher P. Arthur, Matthew J. Bick, Elizabeth A. Campbell, Seth A. Darst, Ronald A. Milligan, and Tarun M. Kapoor. Insights into Antiparallel Microtubule Crosslinking by PRC1, a Conserved Nonmotor Microtubule Binding Protein. *Cell*, 142(3):433–443, August 2010.

[15] Amanda Tulin, Sheri McClerklin, Yue Huang, and Ram Dixit. Single-Molecule Analysis of the Microtubule Cross-Linking Protein MAP65-1 Reveals a Molecular Mechanism for Contact-Angle-Dependent Microtubule Bundling. *Biophysical Journal*, 102(4):802–809, February 2012.

[16] Virginie Stoppin-Mellet, Vincent Fache, Didier Portran, Jean-Louis Martiel, and Marylin Vantard. MAP65 Coordinate Microtubule Growth during Bundle Formation. *PLoS ONE*, 8(2):e56808, February 2013.

[17] Zdenek Lansky, Marcus Braun, Annemarie Lüdecke, Michael Schlierf, Pieter Rein ten Wolde, Marcel E. Janson, and Stefan Diez. Diffusible Crosslinkers Generate Directed Forces in Microtubule Networks. *Cell*, 160(6):1159–1168, March 2015.

[18] Elizabeth H. Kellogg, Stuart Howes, Shih-Chieh Ti, Erney Ramírez-Aportela, Tarun M. Kapoor, Pablo Chacón, and Eva Nogales. Near-atomic cryo-EM structure of PRC1 bound to the microtubule. *Proceedings of the National Academy of Sciences*, page 201609903, August 2016.

[19] Arshad Desai and Timothy J. Mitchison. Microtubule Polymerization Dynamics. *Annual Review of Cell and Developmental Biology*, 13(1):83–117, 1997.

[20] C. G. Dos Remedios, D. Chhabra, M. Kekic, I. V. Dedova, M. Tsubakihara, D. A. Berry, and N. J. Nosworthy. Actin Binding Proteins: Regulation of Cytoskeletal Microfilaments. *Physiological Reviews*, 83(2):433–473, April 2003.

[21] Hui-Shun Kuan and M. D. Betterton. Biophysics of filament length regulation by molecular motors. *Physical Biology*, 10(3):036004, June 2013.

[22] Lishibanya Mohapatra, Bruce L. Goode, Predrag Jelenkovic, Rob Phillips, and Jane Kondev. Design Principles of Length Control of Cytoskeletal Structures. *Annual Review of Biophysics*, 45(1):85–116, 2016.

[23] Thomas D. Pollard. Actin and Actin-Binding Proteins. *Cold Spring Harbor Perspectives in Biology*, page a018226, March 2016.

[24] Jun Allard and Alex Mogilner. Traveling waves in actin dynamics and cell motility. *Current opinion in cell biology*, 25(1):107–115, February 2013.

[25] Erin L. Barnhart, Jun Allard, Sunny S. Lou, Julie A. Theriot, and Alex Mogilner. Adhesion-Dependent Wave Generation in Crawling Cells. *Current Biology*, 27(1):27–38, January 2017.

[26] Ram Dixit and Richard Cyr. Encounters between Dynamic Cortical Microtubules Promote Ordering of the Cortical Array through Angle-Dependent Modifications of Microtubule Behavior. *The Plant Cell Online*, 16(12):3274–3284, January 2004.

[27] Jun F. Allard, J. Christian Ambrose, Geoffrey O. Wasteneys, and Eric N. Cytrynbaum. A Mechanochemical Model Explains Interactions between Cortical Microtubules in Plants. *Biophysical Journal*, 99(4):1082–1090, August 2010.

[28] Ezgi Can Eren, Ram Dixit, Natarajan Gautam, and Fred Chang. A Three-Dimensional Computer Simulation Model Reveals the Mechanisms for Self-Organization of Plant Cortical Microtubules into Oblique Arrays. *Molecular Biology of the Cell*, 21(15):2674–2684, June 2010.

[29] Ezgi Can Eren, Natarajan Gautam, and Ram Dixit. Computer simulation and mathematical models of the noncentrosomal plant cortical microtubule cytoskeleton. *Cytoskeleton*, 69(3):144–154, February 2012.

[30] A. Mogilner and E. Craig. Towards a quantitative understanding of mitotic spindle assembly and mechanics. *Journal of Cell Science*, 123(20):3435–3445, 2010.

[31] Gul Civelekoglu-Scholey and Daniela Cimini. Modelling chromosome dynamics in mitosis: A historical perspective on models of metaphase and anaphase in eukaryotic cells. *Interface Focus*, 4(3):20130073, June 2014.

[32] Matthew Akamatsu, Julien Berro, Kai-Ming Pu, Irene R. Tebbs, and Thomas D. Pollard. Cytokinetic nodes in fission yeast arise from two distinct types of nodes that merge during interphase. *J Cell Biol*, 204(6):977–988, March 2014.

[33] Matthew R. Stachowiak, Caroline Laplante, Harvey F. Chin, Boris Guirao, Erdem Karatekin, Thomas D. Pollard, and Ben O'Shaughnessy. Mechanism of Cytokinetic Contractile Ring Constriction in Fission Yeast. *Developmental Cell*, 29(5):547–561, June 2014.

[34] A. F. Huxley. Muscle structure and theories of contraction. *Progress in Biophysics and Biophysical Chemistry*, 7:255–318, 1957.

[35] A. F. Huxley and S. Tideswell. Filament compliance and tension transients in muscle. *Journal of Muscle Research & Cell Motility*, 17(4):507–511, August 1996.

[36] A. F. HUXLEY and S. TIDESWELL. Rapid regeneration of power stroke in contracting muscle by attachment of second myosin head. *Journal of Muscle Research & Cell Motility*, 18(1):111–114, February 1997.

[37] Melissa K. Gardner, Chad G. Pearson, Brian L. Sprague, Ted R. Zarzar, Kerry Bloom, E. D. Salmon, and David J. Odde. Tension-dependent Regulation of Microtubule Dynamics at Kinetochores Can Explain Metaphase Congression in Yeast. *Molecular Biology of the Cell*, 16(8):3764–3775, January 2005.

[38] Melissa K. Gardner, David J. Odde, and Kerry Bloom. Kinesin-8 molecular motors: Putting the brakes on chromosome oscillations. *Trends in Cell Biology*, 18(7):307–310, July 2008.

[39] Jeremy M. Chacón, Soumya Mukherjee, Breanna M. Schuster, Duncan J. Clarke, and Melissa K. Gardner. Pericentromere tension is self-regulated by spindle structure in metaphase. *The Journal of Cell Biology*, 205(3):313–324, December 2014.

[40] Austin J. Hepperla, Patrick T. Willey, Courtney E. Coombes, Breanna M. Schuster, Maryam Gerami-Nejad, Mark McClellan, Soumya Mukherjee, Janet Fox, Mark Winey, David J. Odde, Eileen O'Toole, and Melissa K. Gardner. Minus-End-Directed Kinesin-14 Motors Align Antiparallel Microtubules to Control Metaphase Spindle Length. *Developmental Cell*, 31(1):61–72, October 2014.

[41] Jonathan J. Ward, Hélio Roque, Claude Antony, and François Nédélec. Mechanical design principles of a mitotic spindle. *eLife*, 3:e03398, January 2015.

[42] E.N. Cytrynbaum, J.M. Scholey, and A. Mogilner. A Force Balance Model of Early Spindle Pole Separation in Drosophila Embryos. *Biophysical Journal*, 84(2):757–769, February 2003.

[43] E. N. Cytrynbaum, P. Sommi, I. Brust-Mascher, J. M. Scholey, and A. Mogilner. Early Spindle Assembly in Drosophila Embryos: Role of a Force Balance Involving Cytoskeletal Dynamics and Nuclear Mechanics. *Molecular Biology of the Cell*, 16(10):4967–4981, January 2005.

[44] Roy Wollman, Gul Civelekoglu-Scholey, Jonathan M Scholey, and Alex Mogilner. Reverse engineering of force integration during mitosis in the Drosophila embryo. *Molecular Systems Biology*, 4, May 2008.

[45] Gul Civelekoglu-Scholey and Jonathan M. Scholey. Mitotic force generators and chromosome segregation. *Cellular and Molecular Life Sciences*, 67(13):2231–2250, July 2010.

[46] Edwin W. Taylor. Dynamics of Spindle Formation and Its Inhibition by Chemicals. *The Journal of Biophysical and Biochemical Cytology*, 6(2):193–196, January 1959.

[47] Iain Hagan and Mitsuhiro Yanagida. Novel potential mitotic motor protein encoded by the fission yeast cut7+ gene. *Nature*, 347(6293):563–566, October 1990.

[48] Iain Hagan and Mitsuhiro Yanagida. Kinesin-related cut 7 protein associates with mitotic and meiotic spindles in fission yeast. *Nature*, 356(6364):74, March 1992.

[49] R. Ding, K. L. McDonald, and J. R. McIntosh. Three-dimensional reconstruction and analysis of mitotic spindles from the yeast, Schizosaccharomyces pombe. *The Journal of Cell Biology*, 120(1):141–151, January 1993.

[50] Robert Blackwell, Christopher Edelmaier, Oliver Sweezy-Schindler, Adam Lamson, Zachary R. Gergely, Eileen O'Toole, Ammon Crapo, Loren E. Hough, J. Richard McIntosh, Matthew A. Glaser, and Meredith D. Betterton. Physical determinants of bipolar mitotic spindle assembly and stability in fission yeast. *Science Advances*, 3(1):e1601603, January 2017.

[51] Sergio A. Rincon, Adam Lamson, Robert Blackwell, Viktoriya Syrovatkina, Vincent Fraisier, Anne Paoletti, Meredith D. Betterton, and Phong T. Tran. Kinesin-5-independent mitotic spindle assembly requires the antiparallel microtubule crosslinker Ase1 in fission yeast. *Nature Communications*, 8:15286, May 2017.

[52] A. P. Enos and N. R. Morris. Mutation of a gene that encodes a kinesin-like protein blocks nuclear division in A. nidulans. *Cell*, 60(6):1019–1027, March 1990.

[53] A L Pidoux, M LeDizet, and W Z Cande. Fission yeast pkl1 is a kinesin-related protein involved in mitotic spindle function. *Molecular Biology of the Cell*, 7(10):1639–1655, October 1996.

[54] Jared C. Cochran, Christopher A. Sontag, Zoltan Maliga, Tarun M. Kapoor, John J. Correia, and Susan P. Gilbert. Mechanistic Analysis of the Mitotic Kinesin Eg5. *Journal of Biological Chemistry*, 279(37):38861–38870, October 2004.

[55] Akira Yamashita, Masamitsu Sato, Akiko Fujita, Masayuki Yamamoto, and Takashi Toda. The Roles of Fission Yeast Ase1 in Mitotic Cell Division, Meiotic Nuclear Oscillation, and Cytokinesis Checkpoint Signaling. *Molecular Biology of the Cell*, 16(3):1378–1395, January 2005.

[56] Megan T. Valentine, Polly M. Fordyce, Troy C. Krzysiak, Susan P. Gilbert, and Steven M. Block. Individual dimers of the mitotic kinesin motor Eg5 step processively and support substantial loads in vitro. *Nature Cell Biology*, 8(5):470–476, May 2006.

[57] Scott V. Bratman and Fred Chang. Stabilization of Overlapping Microtubules by Fission Yeast CLASP. *Developmental Cell*, 13(6):812–827, December 2007.

[58] Marcus Braun, Zdenek Lansky, Gero Fink, Felix Ruhnow, Stefan Diez, and Marcel E. Janson. Adaptive braking by Ase1 prevents overlapping microtubules from sliding completely apart. *Nature Cell Biology*, 13(10):1259–1264, 2011.

[59] Chun Ju Chen, Ken Porche, Ivan Rayment, and Susan P. Gilbert. The ATPase Pathway That Drives the Kinesin-14 Kar3Vik1 Powerstroke. *Journal of Biological Chemistry*, 287(44):36673–36682, October 2012.

[60] Masaki Edamatsu. Bidirectional motility of the fission yeast kinesin-5, Cut7. *Biochemical and Biophysical Research Communications*, 446(1):231–234, March 2014.

[61] Mishan Britto, Adeline Goulet, Syeda Rizvi, Ottilie von Loeffelholz, Carolyn A. Moores, and Robert A. Cross. Schizosaccharomyces pombe kinesin-5 switches direction using a steric blocking mechanism. *Proceedings of the National Academy of Sciences*, page 201611581, November 2016.

[62] Maja Novak, Bruno Polak, Juraj Simunić, Zvonimir Boban, Barbara Kuzmić, Andreas W. Thomae, Iva M. Tolić, and Nenad Pavin. The mitotic spindle is chiral due to torques within microtubule bundles. *Nature Communications*, 9(1):3571, September 2018.

[63] M. A. Hoyt, L. He, K. K. Loo, and W. S. Saunders. Two Saccharomyces cerevisiae kinesin-related gene products required for mitotic spindle assembly. *The Journal of Cell Biology*, 118(1):109–120, January 1992.

[64] Kenneth E. Sawin, Katherine LeGuellec, Michel Philippe, and Timothy J. Mitchison. Mitotic spindle organization by a plus-end-directed microtubule motor. *Nature*, 359(6395):540–543, October 1992.

[65] Anne Blangy, Heidi A. Lane, Pierre d'Hérin, Maryannick Harper, Michel Kress, and Erich A. Nigg. Phosphorylation by p34cdc2 regulates spindle association of human Eg5, a kinesin-related motor essential for bipolar spindle formation in vivo. *Cell*, 83(7):1159–1169, December 1995.

[66] Thomas U. Mayer, Tarun M. Kapoor, Stephen J. Haggarty, Randall W. King, Stuart L. Schreiber, and Timothy J. Mitchison. Small Molecule Inhibitor of Mitotic Spindle Bipolarity Identified in a Phenotype-Based Screen. *Science*, 286(5441):971–974, October 1999.

[67] David J. Sharp, Kristina R. Yu, John C. Sisson, William Sullivan, and Jonathan M. Scholey. Antagonistic microtubule-sliding motors position mitotic centrosomes in Drosophila early embryos. *Nature Cell Biology*, 1(1):51–54, May 1999.

[68] G. Goshima and R. D. Vale. The roles of microtubule-based motor proteins in mitosis. *The Journal of Cell Biology*, 162(6):1003–1016, 2003.

[69] Joshua S. Waitzman and Sarah E. Rice. Mechanism and regulation of kinesin-5, an essential motor for the mitotic spindle. *Biology of the Cell*, 106(1):1–12, January 2014.

[70] Cynthia L. Troxell, Mark A. Sweezy, Robert R. West, Karen D. Reed, Bryan D. Carson, Alison L. Pidoux, W. Zacheus Cande, and J. Richard McIntosh. Pkl1 +andklp2 +: Two Kinesins of the Kar3 Subfamily in Fission Yeast Perform Different Functions in Both Mitosis and Meiosis. *Molecular Biology of the Cell*, 12(11):3476–3488, January 2001.

[71] Zachary T. Olmsted, Andrew G. Colliver, Timothy D. Riehlman, and Janet L. Paluh. Kinesin-14 and kinesin-5 antagonistically regulate microtubule nucleation by γ-TuRC in yeast and human cells. *Nature Communications*, 5:5339, October 2014.

[72] Masashi Yukawa, Yusuke Yamada, Tomoaki Yamauchi, and Takashi Toda. Two spatially distinct Kinesin-14 Pkl1 and Klp2 generate collaborative inward forces against Kinesin-5 Cut7 in S. pombe. *J Cell Sci*, page jcs.210740, January 2017.

[73] Marcel Prelogović, Lora Winters, Ana Milas, Iva Tolic, and Nenad Pavin. Pivot-and-bond model explains microtubule bundle formation. *bioRxiv*, page 157719, August 2017.

[74] Lora Winters, Ivana Ban, Marcel Prelogovic, Nenad Pavin, and Iva M. Tolic. Pivoting of microtubules driven by minus end directed motors leads to their alignment to form an interpolar bundle. *bioRxiv*, page 347831, June 2018.

[75] Liedewij Laan, Nenad Pavin, Julien Husson, Guillaume Romet-Lemonne, Martijn van Duijn, Magdalena Preciado López, Ronald D. Vale, Frank Jülicher, Samara L. Reck-Peterson, and Marileen Dogterom. Cortical Dynein Controls Microtubule Dynamics to Generate Pulling Forces that Position Microtubule Asters. *Cell*, 148(3):502–514, February 2012.

[76] Nenad Pavin, Liedewij Laan, Rui Ma, Marileen Dogterom, and Frank Jülicher. Positioning of microtubule organizing centers by cortical pushing and pulling forces. *New Journal of Physics*, 14(10):105025, October 2012.

[77] Rui Ma, Liedewij Laan, Marileen Dogterom, Nenad Pavin, and Frank Jülicher. General theory for the mechanics of confined microtubule asters. *New Journal of Physics*, 16(1):013018, January 2014.

[78] F. Verde, M. Dogterom, E. Stelzer, E. Karsenti, and S. Leibler. Control of microtubule dynamics and length by cyclin A- and cyclin B-dependent kinases in Xenopus egg extracts. *The Journal of Cell Biology*, 118(5):1097–1108, September 1992.

[79] Marileen Dogterom and Bernard Yurke. Measurement of the Force-Velocity Relation for Growing Microtubules. *Science*, 278(5339):856–860, October 1997.

[80] M. Mercedes Tirado, Carmen López Martínez, and José García de la Torre. Comparison of theories for the translational and rotational diffusion coefficients of rod-like macromolecules. Application to short DNA fragments. *The Journal of Chemical Physics*, 81(4):2047–2052, August 1984.

[81] E. L. Grishchuk and J. R. McIntosh. Microtubule depolymerization can drive poleward chromosome motion in fission yeast. *The EMBO Journal*, 25(20):4888–4896, 2006.

[82] Radhika Subramanian, Shih-Chieh Ti, Lei Tan, Seth A. Darst, and Tarun M. Kapoor. Marking and Measuring Single Microtubules by PRC1 and Kinesin-4. *Cell*, 154(2):377–390, July 2013.

[83] A. S. Kashina, R. J. Baskin, D. G. Cole, K. P. Wedaman, W. M. Saxton, and J. M. Scholey. A bipolar kinesin. *Nature*, 379(6562):270–272, January 1996.

[84] Joshua Pringle, Amutha Muthukumar, Amanda Tan, Laura Crankshaw, Leslie Conway, and Jennifer L. Ross. Microtubule organization by kinesin motors and microtubule crosslinking protein MAP65. *Journal of Physics: Condensed Matter*, 25(37):374103, September 2013.

[85] Iana Kalinina, Amitabha Nandi, Petrina Delivani, Mariola R. Chacón, Anna H. Klemm, Damien Ramunno-Johnson, Alexander Krull, Benjamin Lindner, Nenad Pavin, and Iva M. Tolić-Norrelykke. Pivoting of microtubules around the spindle pole accelerates kinetochore capture. *Nature Cell Biology*, 2012.

Supplemental material

September 10, 2018

S1 Torque-balance model

We aim to create a systems of integro-differential equations for ϕ and l by balancing the forces and torques applied to MT bundles (Figure S1). This model includes three forces: steric force between the nuclear envelope and MT ends, drag from the nucleoplasm and SPB translation, and crosslinker force between MT bundles. We label MT bundle length L and MT crosslinking angle ϕ (Figure S1A). Then we find the normalized bundle length

$$l = \frac{L}{2R},\tag{S1}$$

SPB separation

$$d(l, \phi) = 2R\cos\left(\cos^{-1}(l) + \frac{\phi}{2}\right),\tag{S2}$$

and angle between SPB separation vector and SPB normal vector

$$\theta(l, \phi) = \cos^{-1}(l) + \frac{\phi}{2}.\tag{S3}$$

Due to symmetry we need only consider the equations of motion of a single bundle. We assume that MTs overlap at their centers. This approximation is valid for large d and small ϕ such that

$$\cos\left(\cos^{-1}\left(\frac{d}{2R}\right) - \frac{\phi}{2}\right) - \frac{d}{2R\cos(\phi/2)} \ll 1.\tag{S4}$$

Initial conditions are chosen so our simulations begin in this regime.

Force and torque balance

The first equation of motion for an MT bundle describes the force balance parallel to the bundle axis

$$0 = -\gamma_{\text{lin},\|}v_{\text{c},\|} - F_{+,\|} + F_{-,\|} + F_{\text{spb},\|},\tag{S5}$$

Figure S1: Schematic of the torque-balance model. A, Microtubule bundles, nuclear envelope, and spindle pole bodies. B, MT polymerization moves MT bundles toward the center of the nucleus. C, Forces exerted on crosslinked bundles. D, Crosslinked antiparallel microtubules.

where $F_{\pm,\parallel}$ is the parallel force component on the MT bundle ends due to the wall and $F_{\mathrm{spb},\parallel}$ is the parallel component of the SPB drag force (Figure S1B,C). The first term on the right hand side of Equation S5 is the drag force from the MT bundle moving parallel to its axis with $\gamma_{\mathrm{lin},\parallel}$ and $v_{\mathrm{c},\parallel}$ the parallel friction coefficient and bundle velocity. The bundle center of mass only moves parallel to its axis by polymerization; therefore, this drag force is approximately zero, i.e., $(\gamma_{\mathrm{lin},\parallel}v_{\mathrm{c},\parallel} \sim 0)$.

The second equation of motion balances the torque on the MT bundle

$$0 = -\tau_{\mathrm{drag,r}} + \tau_{+,\mathrm{wall}} - \tau_{-,\mathrm{wall}} + \tau_{\mathrm{spb}} + \tau_{\mathrm{cl},-} - \tau_{\mathrm{cl},+} \tag{S6}$$

where $\tau_{\mathrm{drag,r}}$ is the rotational drag from the nucleoplasm, $\tau_{\pm,\mathrm{wall}}$ is the torque from wall-end

steric interaction, τ_{spb} is the torque from the SPB on the bundle minus end, and $\tau_{\text{cl},-}$ and $\tau_{\text{cl},+}$ are the aligning and anti-aligning torque from crosslinkers. The nucleoplasm does not exert a torque since the bundle pivot point is at its center, the point of action of the total drag force, if Equation S4 holds.

Force

How MT bundles interact with the nuclear envelope determines the steric force on MT ends. MT growth speed depends on the parallel force exerted on its plus end. We model MT dynamics as in previous work, with polymerization speed v_o and stall force F_s [76]

$$v_{\text{p}}(l, \phi) = \frac{v_o}{\eta - 1} \left(\eta^{1 - F_{+,\parallel}(l,\phi)/F_s} - 1 \right),$$
(S7)

where $\eta = e^{\delta F_s / N k_b T}$, δ is the size of a tubulin dimer, 8 nm, N the number of protofilaments in a microtubule, 13, k_b the Boltzmann constant, and T the temperature. Our model sets the nuclear envelope force to a constant directed toward the center of the nuclear envelope, $\vec{F}_+ = \text{const } \hat{r}$. The parallel component is

$$F_{+,\parallel} = F_+ l.$$
(S8)

As MTs grow, the angle between the nuclear envelope normal vector and the MT bundle axis decreases, which increases the parallel component of the wall force $F_{\pm,\parallel}$, and slows the polymerization and motion of the bundle (Figure S1B). We choose the wall force constant to be the stall force of MTs $F_+ = F_s$, which prevents unbounded MT growth since at $l = 1, v = 0$.

The SPB drag force is determined by the SPB velocity on the nuclear envelope (Figure S1C)

$$F_{\text{spb}} = \gamma_{\text{spb}} R \dot{\theta},$$
(S9)

where γ_{spb} is the friction coefficient [50]. We define F_{spb} so that decreasing θ produces a force with a negative component perpendicular to the MT bundle axis. With Equation S3 we re-write Equation S9 as

$$F_{\text{spb}} = \gamma_{\text{spb}} R \left(-\frac{\dot{l}}{\sqrt{1 - l^2}} + \frac{\dot{\phi}}{2} \right),$$
(S10)

where the parallel component is

$$F_{\text{spb},\parallel} = -\gamma_{\text{spb}} R \left(-\frac{\dot{l}}{\sqrt{1 - l^2}} + \frac{\dot{\phi}}{2} \right) l.$$
(S11)

This is negative because a decreasing ϕ (with no change in length) produces a positive parallel force along the MT axis.

The force on the MT bundle ends from the nuclear envelope is found using Equation S5 and $\gamma_{\text{lin},\parallel} v_{\text{c},\parallel} = 0$, so that

$$F_{-,\parallel} = F_{+,\parallel} - F_{\text{spb},\parallel}.$$
(S12)

Steric and drag torque

Torque changes the orientation of the MT bundles and thus changes ϕ. Positive torque increases ϕ. We calculate torque, with the exclusion of that produced by crosslinkers, from the cross product of forces and the displacement of the point of action from the pivot. Since we approximate MT centers to be the pivot point, perpendicular components of the forces mentioned above multiplied by half the bundle length ($L/2$) give the magnitude of the torque. The torque from SPBs is

$$\tau_{\mathrm{spb}} = -F_{\mathrm{spb},\perp} L/2 = -\gamma_{\mathrm{spb}} l R^2 \left(\frac{\dot{\phi}}{2}\sqrt{1-l^2} - \dot{l} \right) \tag{S13}$$

whereas the torque applied to MT plus and minus ends is

$$\tau_{+} = F_{+,\perp} L/2 = F_{+,\|} R\sqrt{1-l^2}, \tag{S14}$$

and

$$\tau_{-} = F_{-,\perp} L/2 = (F_{+,\|} - F_{\mathrm{spb},\|}) R\sqrt{1-l^2}. \tag{S15}$$

The rotational drag on MTs from the nucleoplasm is

$$\tau_{\mathrm{drag,rot}} = -\gamma_{\mathrm{rot}} \dot{\phi}, \tag{S16}$$

where γ_{rot} is the rotational friction coefficient for an MT bundle modeled as a spherocylinder of length L [77]. This leaves only the torque from crosslinkers to completely define Equation S6.

Crosslinker torque and molecule number

The statistical properties of an ensemble of passive crosslinkers (the average number bound, torque, force, etc.) are calculated from the grand canonical potential (Equation 6) for indistinguishable crosslinkers binding to two filaments pivoting around a common origin (Figure S1D). We assume crosslinkers do not interact with each other and bind to both bundles simultaneously. For the geometry of Figure S1D, the partition function is

$$q = c \int_0^{L/2} \int_0^{L/2} dr_1 dr_2 e^{-\frac{\beta k}{2}(h(r_1, r_2, \phi) - h_{\mathrm{cl}})^2} \tag{S17}$$

where c is the density of attachment sites and has the dimensions of inverse length squared, k is the spring constant of a crosslinker, $h = \sqrt{r_1^2 + r_2^2 - 2r_1 r_2 \cos(\phi)}$ is the crosslinker head separation, and h_o is the equilibrium length of each crosslinker. The crosslinker binding affinity to two microtubules is the product of the fugacity z (Equation 6) and c which are never separated in this model.

The torque produced on one MT bundle is the negative derivative of the grand potential Φ (Equation 7). The average crosslinker number comes from the derivative of Φ with respect to the chemical potential μ

$$\langle N \rangle = -\partial_\mu \Phi = (-\partial_\mu z)\partial_z \Phi = -\beta z \partial_z \Phi = zq. \tag{S18}$$

Crosslinkers also bind above and below the pivot point of Figure S1A. These crosslinkers produce the anti-aligning torque $\tau_{\mathrm{cl},+} = \tau_{\mathrm{cl}}(l, \pi - \phi)$ and contribute to the overall crosslinker number. Therefore, the total bound crosslinker number and torque are

$$\langle N_{\mathrm{tot}} \rangle = 2\langle N_+ \rangle + 2\langle N_- \rangle, \tag{S19}$$

and

$$\tau_{\mathrm{cl,tot}} = \tau_{\mathrm{cl},-} - \tau_{\mathrm{cl},+}. \tag{S20}$$

Equation S20 is then used in Equation S6 to complete the torque-balance model.

Equations of motion

Combining Equations S5, S6, and S7, we derive a system of equations for $\dot{\phi}$ and $\dot{l}$. By solving Equation S6 for $\dot{\phi}$ and plugging in the value $F_{-,\parallel}$ from Equation S12 we arrive at

$$\dot{\phi}(\dot{l}, l, \phi, t) = \frac{2R^2 \gamma_{\mathrm{spb}} l \dot{l} + \tau_{\mathrm{cl},-} - \tau_{\mathrm{cl},+}}{\gamma_{\mathrm{rot}} + R^2 \gamma_{\mathrm{spb}} l \sqrt{1 - l^2}}. \tag{S21}$$

Normalizing the MT polymerization speed from Equation S7 gives

$$\dot{l}(l) = \frac{v_o}{2R(\eta - 1)} \left(\eta^{1-l} \right). \tag{S22}$$

Equations S21 and S22 are numerically integrated in python using the `odeint` function from the `scipy.integrate` library. This code uses the Fortran `ODEPACK` library and LSODA integrator. The crosslinker torque from Equation 7 is computed by Gaussian quadrature using the `dblquad` method from `scipy.integrate`.

S2 Initialization of kMC-BD phase diagram simulations

Simulations that replicate the initial conditions of the torque-balance model start with SPBs set at separation d_o with the MT minus ends anchored at random locations on the SPBs. MTs are tilted away from the SPB normal vector so that the bundles cross at the desired ϕ_o. All MT lengths are initialized so that an MT attached to an SPB center makes contact with the nuclear envelope. The simulation then relaxes for 1 second so that MTs do not overlap and crosslinkers bind to MTs. During the relaxation, SPBs are held at their initial positions and MTs remain at their initial length. Afterward, SPBs are released and MTs become dynamically unstable.

Figure S2: Time evolution of the torque-balance model with the parameters of Figure 2C, with the exception of the crosslinker binding affinity c_{cl}. The SPBs are initially separated by 1 μm and MTs bundled at an angle of 5 degrees. Systems with $c_{\mathrm{cl}} > 2.9 \times 10^{-4}$ nm^{-2} assemble bipolar spindles because the bundling angle ϕ decreases while the SPB separation d increases.

Figure S3: Time evolution of the torque-balance model with parameters of Figure 2C and MTs initially bundled at 5 degrees. The SPBs separation is varied from 0.55 μm to 2.48 μm. The approximation that MTs bundle at their centers holds for systems of large SPB separation (> 0.5 μm) and small angle (< 20 degrees). Simulations that evolve to states outside this range give unphysical results at long time, as seen for $d_o < 0.96$ μm. However, these simulations have monotonically increasing ϕ, because aligning torque decreases with increasing ϕ after a certain ϕ. This monotonicity of ϕ before becoming unphysical implies an aberrant final state for these simulations.

Figure S4: Time evolution of the torque-balance model with parameters of Figure 2C and SPBs initially separated by 1 μm. MTs are bundled at angles from 0 to 20 degrees. MTs bundled at angles greater than 8 degrees fail to form bipolar spindles. At small angle and short distance, crosslinkers exert an anti-aligning torque, since they are compressed. As bundle length increases, the crosslinking torque becomes positive, and the spindle becomes bipolar.