bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Deacetylation of catalytic lysine in CDK1 is essential for Cyclin-B binding and cell cycle

Shaunak Deota[1], Sivasudhan Rathnachalam[2], Kanojia Namrata[1], Mayank Boob[2], Amit Fulzele[3], Radhika S[2], Shubhra Ganguli[4, 5], Chinthapalli Balaji[1], Stephanie Kaypee[6], Krishna Kant Vishwakarma[2], Tapas Kumar Kundu[6], Rashna Bhandari[4], Anne Gonzalez de Peredo[3], Mithilesh Mishra[1], Ravindra Venkatramani[2*] and Ullas Kolthur-Seetharam[1*]

1) Department of Biological Sciences, Tata Institute of Fundamental Research (TIFR), Mumbai 400005, India, 2) Department of Chemical Sciences, Tata Institute of Fundamental Research (TIFR), Mumbai 400005, India, 3) Institut de Pharmacologie et de Biologie Structurale (IPBS), Toulouse 31400, France, 4) Laboratory of Cell Signalling, Centre for DNA Fingerprinting and Diagnostics (CDFD), Hyderabad 500039, India, 5) Graduate Studies, Manipal Academy of Higher Education, Manipal 576104, India, 6) Transcription and Disease Laboratory, Molecular Biology and Genetics Unit, Jawaharlal Nehru Centre for Advanced Scientific Research (JNCASR), Bengaluru 560064, India.

*** Co-corresponding**

Correspondence: ullas@tifr.res.in, ullaskolthur@gmail.com, ravi.venkatramani@tifr.res.in

Running title: Deacetylated CDK1 binds Cyclin-B

Key words: CDK1; Cyclin-B; Decetylation; Cell cycle; SIRT1

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Abstract

Cyclin-dependent-kinases (CDKs) are essential for cell cycle progression. While dependence of CDK activity on Cyclin levels is established, molecular mechanisms that regulate their binding are less studied. Here, we show that CDK1:Cyclin-B interactions are regulated by acetylation, which was hitherto unknown. We demonstrate that cell cycle dependent acetylation of the evolutionarily conserved catalytic lysine in CDK1 or eliminating its charge state abrogates Cyclin-B binding. Opposing activities of SIRT1 and P300 regulate acetylation, which marks a reserved pool of CDK1. Our high resolution structural analyses into the formation of kinase competent CDK1:Cyclin-B complex have unveiled long-range effects of catalytic lysine in configuring the CDK1 interface for Cyclin-B binding. Cells expressing acetylation mimic mutant of Cdc2 in yeast are arrested in G2 and fail to divide. Thus, by illustrating cell cycle dependent deacetylation as a determinant of CDK1:Cyclin-B interaction, our results redefine the current model of CDK1 activation and cell cycle progression.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Introduction

Cyclin-dependent kinases (CDKs) are evolutionarily conserved protein kinases, which are indispensible for eukaryotic cell division cycles, differentiation and transcription [1, 2]. Among these, CDK1 (or yeast Cdc2 and Cdc28), CDK2, CDK4 and CDK6 orchestrate cell cycle progression. These constitutively expressed CDKs form cognate pairs with Cyclins, which have cell cycle phase dependent expression [1]. Besides Cyclin binding, our current understanding of the temporal control of CDK activity is based on inhibitory and activatory phosphorylations, brought about by a regulatory loop involving other kinases and phosphatases [3]. Aberrant activation or inhibition of CDKs are known to cause cancer, developmental disorders and senescence [4].

Protein regions that are required for CDK-Cyclin interactions have been mapped [5-7]. Cyclin binding induces the movement of PSTAIRE-helix in CDK and thus the formation of an active kinase conformation [6, 7]. Interestingly, activatory phosphorylation, for example in CDK1, has been shown to stabilize the CDK1:Cyclin-B complex, although, this phosphorylation is itself dependent upon Cyclin-B binding to CDK1 [5, 8]. Given the essentiality of Cyclin binding for CDK activation and hence cell cycle progression, physiologically relevant mechanisms that determine complex formation remain to be addressed. Specifically, deterministic residues within CDKs that may regulate Cyclin binding/unbinding and therefore contribute to cell division are poorly understood. It is important to emphasize that Cyclin binding to CDKs have been generally thought to be default.

Almost all eukaryotic protein kinases (EPKs), including the CDKs, have a conserved catalytic triad consisting of N-lobe-β3 lysine, glutamate in C-helix and aspartate in DFG motif, which are essential for kinase activity [9, 10]. Several

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

structure-function studies, mostly on cAMP dependent Protein Kinase A (PKA), have attempted to investigate the roles played by these catalytic triad residues in rendering the kinase active and in mediating phosphoryl-transfer. Uniquely in CDKs, binding of Cyclin induces a structural change to bring together these catalytic triad residues, which are otherwise far apart in apo-CDKs [6, 11]. Moreover, N-lobe-β3 containing the catalytic lysine is contiguous with the PSTAIRE-/C-helix, which not only harbors the catalytic glutamate but also forms the interface where Cyclin binds [6, 12]. Therefore, given that Cyclin binding brings a distinct mode of activation of CDKs, identification of regulatable residues or structural elements, which lead to the formation of the kinase competent complex will be crucial to our understanding of cell division cycles.

EPKs, including CDKs, have been reported to be acetylated [13-19]. It is important to note that in many of these cases, acetylation happens on the catalytic lysine. Substitution of the catalytic lysine to acetyl-mimic glutamine, for example in CDK2, CDK5 and CDK9, leads to a complete loss of kinase activity [14-16, 20]. This raises the possibility of acetylation as a key determinant of CDK activity, i.e. in addition to phosphorylation. Despite these studies, if and how acetylation and deacetylation dynamics regulate CDK activity has not been elucidated.

Here, we report that CDK1 is acetylated at lysine-33 by the acetyltransferase P300 and is deacetylated by NAD^+-dependent deacetylase SIRT1. We show that while lysine-33 is dispensable for ATP binding, the conformation and interactions of ATP in the active site are altered upon its acetylation. Surprisingly, we find that perturbing the charge state of catalytic lysine impacts CDK1 and Cyclin-B complex formation. Moreover, we also show

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

that CDK1 lysine-33 acetylation is dynamically regulated across cell-cycle stages and negatively correlates with CDK1:Cyclin-B complex formation. Substitution of the lysine to acetyl-mimic glutamine in Cdc2 failed to compliment different temperature-sensitive *cdc2* mutants in fission yeast. Together, this study identifies deacetylation as a regulatory mechanism for activation of CDK1, which was hitherto unknown.

Results

SIRT1 and P300 regulate acetylation of the conserved catalytic lysine in CDK1

Given that CDKs are acetylated, if and how acetylation impinges on their structural and functional states is poorly understood. Specifically, the impact of lysine acetylation on CDK1 function, which in general cannot be compensated by any of the other CDKs [21], has not been addressed thus far. In this context, we probed CDK1 acetylation in mammalian cells. We found immunoprecipitated CDK1-HA to be acetylated (Fig 1A) consistent with our global LC-MS/MS analyses of the acetylated sub-proteome (unpublished results). This analysis revealed lysine-33 as the sole acetylation site in endogenous CDK1 (Fig 1B), as reported earlier [13]. To further characterize the importance of this acetylation, we generated antibodies specific to the lysine-33 acetylated peptide sequence ^{30}VAM**K^{Ac}**KIRLESE^{40} in CDK1 (α-CDK1-K33Ac/α-K33Ac). This region is largely conserved across CDKs and specifically in CDK1 from yeast to humans (Figs S1A-S1C). Immunoaffinity purified α-K33Ac antibodies were able to detect endogenous CDK1 acetylation at lysine-33 (Fig S1E) and did not react with CDK1-K33R mutant (Fig S1D). Further, we could detect acetylated species of

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

CDK1 using both pan-acetyl-lysine and α-K33Ac antibodies (Fig 1C). These clearly corroborated the LC-MS/MS results vis-à-vis acetylation of CDK1 at lysine-33. Our results suggest that endogenous CDK1 acetylation is low in abundance (Fig S1E) and it is important to note that even phosphorylation, the dominant and deterministic modification on CDK1, is sub-stoichiometric, across cell cycle phases [22, 23].

Protein acetylation is regulated by activities of lysine acetyltransferases (KATs) and lysine deacetylases (KDACs). Hence, to identify the acetyltransferase responsible for acetylating CDK1 at lysine-33, we expressed CDK1-HA along with KATs: P300, PCAF and GCN5. As can be seen from Fig 1D, CDK1-acetylation was significantly enhanced in response to P300 expression when compared to PCAF and GCN5. Moreover, P300 was able to acetylate CDK1 *in-vitro* (Fig S1F). These results are consistent with earlier reports showing CDK1-P300 interaction and CDK1 mediated phosphorylation of P300 [24].

To identify the KDAC, we treated cells with nicotinamide (NAM) and Trichostatin-A (TSA), which non-overlappingly inhibit Sirtuins (NAD^+-dependent deacylases) and non-Sirtuin KDACs, respectively. We found that there was a robust increase in CDK1 acetylation following NAM treatment, clearly indicating an involvement of Sirtuins (Fig 1E). Moreover, earlier reports have shown that CDK1 and SIRT1 interact with each other [25] (Fig S1G), although the biological relevance of this interaction has not been investigated. Given this, we indeed observe that SIRT1 overexpression led to a reduction in CDK1 acetylation (Figs 1F and 1G). Taken together, these results clearly illustrate that P300 is the major KAT for CDK1 at lysine-33 and that SIRT1 is involved in its deacetylation.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Acetylation of CDK1 at lysine-33 impairs its kinase activity

Evolutionarily conserved lysine-33 lies in the catalytic pocket of CDK1 where ATP binds, and analogous active site lysines in other CDKs and in fact all other kinases have been implicated in kinase activity [14-17, 26-33]. Hence, to check the importance of lysine-33 acetylation for CDK1 function, we employed K33R and K33Q mutants of CDK1, which have typically been used as deacetylation and acetylation mimics, respectively. *In-vitro* kinase assays of CDK1-WT and -K33Q, immunoprecipitated from G2/M synchronized cells co-expressing Cyclin-B1 (Fig S2B), showed that K33Q mutation completely abrogated the CDK1 activity (Figs 2A and 2B). Interestingly, CDK1-K33R also abolished kinase activity (Fig S2A) despite retaining the charge state, as with other kinases with similar arginine substitutions [17, 28-33]. It is important to note that although mutations of the catalytic lysine have been shown to abolish CDK activity, the underlying molecular mechanism is still unknown [14-16, 20, 27]. Notably, these results indicated that both the lysine side-chain and its charge state are essential determinants of kinase activity.

Mutations of tyrosine-15 and threonine-161 (inhibitory and activatory phosphorylation sites, respectively) or aspartate-146 (in DFG motif) in CDK1 affect cell cycle progression [26, 34-38]. Unlike these mutants, overexpression of CDK1-K33Q did not lead to a dominant negative phenotype (Figs S2C and S2D). Possibly owing to the kinase dead CDK1-K33Q mutant, we could not generate a cell culture system to assay for the acetyl-mimic mutant in a background wherein the endogenous wild-type CDK1 was absent (Data not shown). In this context, it is important to note that mutating an orthologous lysine residue to the acetyl-mimic glutamine (K40Q) in Cdc28 results in lethality [13]. Therefore, we

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

resorted to using ectopically expressed CDK1-K33Q mutant to dissect out the molecular mechanism of loss of kinase activity.

Inhibitory phosphorylation at tyrosine-15 prevents precocious activation and is an indicator of inactive CDK1. Upon assaying for phospho-tyrosine-15, we did not find any significant difference between CDK1-WT and -K33Q (Figs 2C and 2D). This clearly rules out inhibitory phosphorylation as causal for inactivation of CDK1-K33Q mutant. Moreover, these motivated us to investigate a hitherto unknown mechanism of CDK1 activity regulation that is dependent on the catalytic lysine residue.

Modeling and simulations of WT, K33Q and K33Ac CDK1:Cyclin-B:ATP complexes

Next, we investigated how lysine-33, its acetylation or the mutation to acetyl-mimic glutamine affected CDK1:Cyclin-B:ATP active complex, through modeling and computational analyses. To this end, we used the crystal structure of CDK1:Cyclin-B complex (PDB ID: Y472), reported recently [12], and docked an ATP molecule into the catalytic pocket. We generated the wild type CDK1:Cyclin-B:ATP ternary complex model and validated it using the ATP bound CDK2:Cyclin-A crystal structure (Fig 3A), given their active site structural similarities. We also generated models for CDK1-K33Q and CDK1-K33Ac (lysine-33 replaced with acetyl-lysine) (Table S1). We would like to point out that the initial binding mode of ATP for all three models was the same. The atomistically detailed models were solvated, equilibrated at standard temperature (300 K) and pressure (1 bar), and subjected to 200 ns Classical Molecular Dynamics (CMD) simulations to relax the complexes to their native ternary form. Importantly, given that acetylated lysine

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

is bulkier than lysine (Fig S3A), we employed three different equilibration protocols to study the impact of K33Ac on CDK1:Cyclin-B:ATP complex. Specifically, we obtained three distinct structurally relaxed conformations of the ternary complex after equilibration: CDK1-K33Ac-1 (ATP, E51 both constrained during equilibration), CDK1-K33Ac-2 (ATP unconstrained, E51 constrained during equilibration), and CDK1-K33Ac-3 (ATP constrained, E51 unconstrained during equilibration) (Figs 3B-3F and Figs S3B-S3F). We stress that no constraints were employed in the 200 ns production simulations. In the case of CDK1-K33Ac-1, but not in others, the acetyl-lysine residue flipped out of the active site pocket during production runs. In CDK1-K33Ac-2 and -K33Ac-3, the uncharged residue was observed to interact with the ATP nucleobase (Figs 3G-3K and Fig S3G-S3K).

Catalytic lysine in CDK1 plays distinct roles in stabilizing ATP and influencing interactions with Cyclin-B

We examined, *in-silico*, the effect of lysine-33 perturbations on ATP binding in terms of the following two measures: a) Solvent accessibility of ATP in the catalytic pocket relative to its accessibility in free solution (relative SASA), and b) Non-bonded (electrostatic and van der Waals) interaction energy of ATP with residues in the catalytic pocket (Fig 4B). As shown in Fig 4A, more than 60 % of the ATP surface area is buried in the active site in all five complexes (CDK1-WT, -K33Q, and the three -K33Ac models). Although in the CDK1-K33Ac-2 model, we observed transient fluctuations in the relative SASA for ATP during the 200 ns MD trajectory, the averaged values over the entire timescale show that ATP is buried in the active site as in the other models. Nevertheless, we

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

found that with the loss of electrostatic interactions of ATP tail with lysine-33, as in CDK1-K33Q and -K33Ac complexes, the conformation adopted by ATP was different when compared to CDK1-WT (Fig 4D). We further assessed the non-bonded interaction energy (E_{NB}) of ATP with residues of the active site pocket within a 3-7 Å radius around the ATP. In the wild type complex, for residues within 3 Å, E_{NB} was about -400 kcal/mol and become slightly more favorable (to around -450 kcal/mol) when residues up to 5 Å were included (Fig 4B and Fig S3L). In CDK1-K33Q and -K33Ac complexes, the ATP E_{NB} values were comparable and were less favorable (by about 100-200 kcal/mol), vis-à-vis CDK1-WT for interactions with residues within a 3-7 Å radius. Moreover, our analyses clearly suggested that interactions of ATP with residues beyond 5 Å appear to be insignificant, leading to negligible changes in E_{NB} for both CDK1-WT and mutant complexes.

An analysis of ATP interactions with the core catalytic triad (residue-33, aspartate-146, and glutamate-51) showed that lysine-33 was one of the major contributors in terms of binding energetics (Fig 4C). However, it should be noted that despite the loss of favorable non-bonded interactions, as in the case of CDK1-K33Q and -K33Ac, the overall interactions (E_{NB} < -200 kcal/mol) of ATP with the residues of active site pocket remain highly favorable (Fig 4B). In summary, ATP appears to be stably bound to the same extent, albeit with different binding modes in CDK1-WT, -K33Q and -K33Ac complexes. In particular, we find that the Mg^{2+} coordination sphere, which includes oxygens of ATP, acidic residues at the active site and water molecules to be different for CDK1-WT, -K33Q and -K33Ac complexes (Fig 4D). Based on these results on lysine-33 perturbations in CDK1, and the previous report of lysine-72 mutations

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

in PKA [33], we propose that the catalytic lysine may be dispensable for stable ATP binding in EPKs. Nevertheless, our analyses raise the possibility of lysine-33 to play a key role in phosphoryl-transfer reaction, which needs to be addressed in the future.

Interactions between the catalytic residues lysine and glutamate on N-lobe-β3 and C-helix, respectively, have been proposed to be important for kinase activity in almost all EPKs [10, 39]. Specifically, in a complex kinase system like the CDKs, Cyclin binding induces a salt-bridge interaction between the catalytic lysine and PSTAIRE-/C-helix glutamate [7, 12, 40]. Importantly for CDKs, the impact of the loss of lysine charge state on salt-bridge with glutamate and PSTAIRE-/C-helix interactions has not been investigated. In this regard, for CDK1-WT we found that lysine-33 interacted with both glutamate-51 and ATP during the time course of the simulations, showing more prominent interactions with the latter (Figs S3M-S3O). The change in the lysine-33 charge state caused a small rotation of the PSTAIRE-helix axis by ~5 degrees in the CDK1:Cyclin-B interfacial plane (Figs S4A and S4B), and also changed the position and conformation of the T-loop (activation segment) in the CDK1 catalytic domain (Figs S4C and S4D). However, we did not find any movement of either the PSTAIRE-/C-helix or glutamate-51 towards the ATP because of the loss of lysine-33 charge state (Figs S3P-S3R), unlike what was recently reported for PKA [33]. This indicates that the catalytic lysine, while being indispensable, plays differential roles in rendering kinases active across EPKs.

Mutational analyses and structural evidences have indicated that PSTAIRE-/C-helix in CDK1 is involved in binding to Cyclin-B, similar to CDK2:Cyclin-A [6, 7, 12, 41]. However, the forces/interactions that stabilize the

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

CDK:Cyclin complex remain unclear. In our attempts to examine the consequences of loss of lysine-33 charge state, we discovered a change in the distribution of center-of-mass (COM) distances between CDK1 and Cyclin-B proteins in the CDK1-K33Q and -K33Ac complexes relative to the CDK1-WT. As can be clearly seen in Fig 4E, the distribution of CDK1:Cyclin-B COM separations for the mutant lysine-33 complexes (CDK1-K33Q, -K33Ac-1 and -K33Ac-2) distinctly shift to larger values relative to CDK1-WT. For CDK1-K33Ac-3, while there is no significant shift in peak COM values, the distribution broadens significantly, indicating higher flexibility in the binding geometry of the two proteins. In fact, the widths of distribution of the inter-protein separations along our MD trajectories are the smallest for CDK1-WT. These observations suggest that the perturbations of lysine-33 could be involved in remodeling CDK1:Cyclin-B interface, which was unknown thus far.

To summarize, our *in-silico* structural analyses using multiple CDK1:Cyclin-B:ATP ternary complexes led us to hypothesize that: a) While the charge state of lysine-33 in the catalytic pocket was necessary for orienting ATP, it may be dispensable for its binding and b) Perturbing lysine-33 could influence CDK1 interactions with Cyclin-B. We therefore set out to validate these hypotheses using biochemical approaches.

Catalytic lysine-33 is dispensable for ATP binding

To check if lysine-33 is required for ATP binding, we used the following approaches: a) binding of 5'-(4-Fluoro-sulfonyl-benzoyl)-adenosine (FSBA), an analog of ATP and b) ATP-agarose in which ATP is conjugated to the beads in different orientations. Consistent with previous reports from CDK9 [14] and

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Cdc2 [42], CDK1-K33Q and -K33R mutants had reduced FSBA binding (Fig 5A). FSBA covalently binds to the active site lysine with high affinity [43]. Moreover unlike FSBA, ATP is non-covalently coordinated in the catalytic pocket. Thus FSBA binding that is equivalently affected in CDK1-K33Q and -K33R does not reveal the importance of lysine in ATP binding.

We found that CDK1-WT bound 6-AH ATP-beads better than the others and were used for further analyses (Fig S5A). Next, we assessed the relative binding efficiency of CDK1-WT, -K33Q and -K33R to the ATP conjugated beads. In contrast to our results with FSBA, we found that steady-state ATP binding is not affected by these mutations in CDK1 (Figs 5B and 5C and Figs S5B and S5C). Nevertheless, in binding competition assays using ATP in solution, we found small differences between CDK1-WT, -K33Q and -K33R (Figs S5D and S5E). These results clearly indicate that the catalytic pocket lysine or its charge state are dispensable for steady-state ATP binding and importantly corroborate our predictions based on the *in-silico* analyses.

Non-default binding of Cyclin-B to CDK1 is determined by the charge state of catalytic lysine

Having ruled out all known possible mechanisms that led to CDK1 inactivation due to the alterations of catalytic lysine and motivated by our *in-silico* results, we wondered if Cyclin-B binding itself is affected. To test this, we probed for the interaction of CDK1-WT, -K33Q and -K33R with Cyclin-B1 by reversible co-immunoprecipitations. Remarkably, we found that CDK1-K33Q, which takes away the charge and is typically used as an acetylation mimic, reduced the interaction with Cyclin-B1 (Figs 5D-5G). On the contrary, CDK1-

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

K33R, which retains the charge state and mimics constitutively deacetylated state, remained bound to Cyclin-B1 (Figs S5F and S5G).

Given that we found endogenous CDK1 lysine-33 to be acetylated, which would naturally mask its charge state, we wanted to assess if acetylation is a determinant of Cyclin-B binding in a cell cycle dependent manner. Despite our best attempts, we could not map differential acetylation of endogenous CDK1 during different phases of cell cycle using α-K33Ac antibodies (Data not shown). As is known for most posttranslational modifications, including key regulatory phosphorylations [22, 23], stoichiometry of protein acetylation has been shown to be relatively low [44, 45]. Therefore, our inability to quantitatively estimate cell cycle dependent changes in acetylation could be due to low abundance of acetylated CDK1 combined with the low sensitivity of α-K33Ac antibodies. Nevertheless, using over-expressed CDK1, we investigated if the acetylation and deacetylation potential was indeed regulated in a cell cycle dependent manner. As shown clearly in Fig 5H and Fig S5H, we found hypoacetylated CDK1 in G2/M synchronized cells when compared to cells arrested in G1/S. Importantly, overexpression of Cyclin-B1, which increases G2/M population, also resulted in reduced CDK1 lysine-33 acetylation (Fig 5I). This was consistent with our hypothesis of deacetylation of catalytic lysine as a determinant of Cyclin-B binding. We further assayed for CDK1:Cyclin-B1 complexes under conditions, which led to increased CDK1 lysine-33 acetylation, by immunoprecipitation. As anticipated, we found that hyperacetylated CDK1 showed significantly less Cyclin-B1 association (Fig 5J). We consistently failed to express and purify soluble CDK1 and Cyclin-B1, independently (Data not shown), given that recombinant CDK1:Cyclin-B are only obtained by co-expressing them together.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Due to this limitation, we could not carry out *in-vitro* binding studies to assess strength of interaction of CDK1-K33Q or -K33Ac with Cyclin-B, which are complexation defective. Together, we have found CDK1:Cyclin-B interactions to be regulated, which was hitherto unknown. Importantly, our findings establish that the positive charge of catalytic lysine-33 in CDK1 is essential for Cyclin-B binding and its loss either by acetylation or a mutation to glutamine abrogates the interaction.

Electrostatic tethering of catalytic lysine-33 dictates surface interactions between CDK1 and Cyclin-B

Next, we wanted to investigate the mechanistic underpinnings of lysine-33 dependent association of CDK1 and Cyclin-B proteins. Examining the structure of ternary WT CDK1:Cyclin-B:ATP complex revealed that residues 40-60 of CDK1 interface directly with Cyclin-B. Specifically, in addition to the PSTAIRE-helix (residues 45-58), which has been previously described [6, 12], we found that the random coil segment (residues 40-44) in CDK1 also interacts with Cyclin-B, as implicated in CDK2:Cyclin-A [6]. Importantly, the random coil segment is contiguous with the N-lobe-β3 (residue 28-36), containing lysine-33. It should be noted that combination mutants of residues in each of these segments in CDK1 have been shown to reduce Cyclin-B binding [5]. However, the forces that orient these segments and help in stabilizing interactions between the proteins are still unknown.

These observations prompted us to investigate if the local electrostatic interactions of catalytic lysine-33 impinge on interface interactions of CDK1 with Cyclin-B. Specifically, we carried out a statistical analysis of salt-bridge

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

interactions in CDK1-WT, -K33Q and -K33Ac ternary complexes (Table S2) during the initial (first 1 ns) stages of our 200 ns MD trajectory (Fig 6C). We chose the 1 ns period of the trajectory since destabilization of the CDK1:Cyclin-B interface, as assessed by inter-protein COM separations, occurred early in our MD production runs in CDK1-K33Q and -K33Ac systems. Our analysis clearly shows that the number of interfacial salt-bridges is halved in the case of CDK1-K33Q and -K33Ac complexes, as compared to the CDK1-WT, during the initial phase of our MD simulations (Fig 6A), presaging the increase in inter-protein separations (Fig 4E). Further, we quantified the distributions of E_{NB} of interfacial salt-bridge forming residues of CDK1 and Cyclin-B (Table S2). Interestingly, we found the E_{NB} values to be significantly lower (by ~150-200 kcal/mol) and also highly variable when the charge state of lysine-33 is lost (Fig 6B). Moreover, a more detailed analyses revealed that interfacial salt-bridge interactions involving both, acidic (D/E) and basic (K/R) residues of CDK1 (Fig S6A), as well as side chain-backbone salt-bridges (Fig S6B) were reduced in K33Q and K33Ac CDK1:Cyclin-B:ATP complexes.

Our comprehensive analysis further demonstrated that, upon perturbing lysine-33, the largest changes in E_{NB} were observed for residue glutamate-41 (by ~100 kcal/mol) at the CDK1:Cyclin-B interface (Fig 6D). Interestingly, we found negligible perturbations in salt-bridge interactions formed by acidic and basic residues in the PSTAIRE-helix (arginine-50 and glutamate-57). Specifically, our results illustrate that the effects mediated by catalytic lysine-33 on glutamate-41 in the N-lobe-β3/C-helix linker loop of CDK1 may play a deterministic role in enabling CDK1:Cyclin-B interactions. These findings are also illustrated in the form of movies (Movies S1-S5).

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Cdc2/CDK1 lysine-33 mutation blocks cell-cycle progression in *S. pombe*

To better understand physiological consequence of lysine-33 acetylation for CDK1 function, we turned to fission yeast, *Schizosaccharomyces pombe*, which shares the cell cycle regulatory mechanisms with higher eukaryotes including humans [46, 47] and is one of the best studied system for mitotic control. Indeed the first human CDK gene was cloned based on its ability to compliment a temperature sensitive mutation in *cdc2* gene, *cdc2*-33 [46]. Unlike mammalian cells, the fission yeast cell cycle is controlled by a single CDK, Cdc2, which drives both G1/S and G2/M transitions, making it a simple model to study cell cycle progression [48-50]. Moreover a single CDK/Cyclin pair (Cdc2p/Cdc13p) is sufficient to drive progression through the different phases of cell cycle [51].

Wild type fission yeast cells grow to a length of about 14 microns before dividing. Temperature-sensitive (ts) *cdc2* mutants that undergo cell-cycle arrest at G2 and are unable to divide or form colonies, continue to grow when incubated at the restrictive temperature of 36°C producing abnormally elongated cells with a single nucleus (Fig 7A). Two independent ts alleles of *cdc2*, *cdc2-33* (A177T) and *cdc2-L7* (P208S), [48, 52] arrest as elongated mononucleate cells when incubated at restrictive temperature of 36°C (Figs 7B and 7C and Figs S7A-S7D), and fail to form viable colonies (Figs 7D and S7E). Expressing wild type *cdc2* gene under moderate strength thiamine repressible promoter (pnmt41) rescues the cell cycle arrest in these mutants. These cells grow and divide at wild type length distribution and are viable at the restrictive temperature of 36°C (Figs 7B-7D and Figs S7A-S7E). Unlike the cells rescued with Cdc2-WT, inducible expression of both Cdc2-K33Q and Cdc2-K33R failed to

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

compliment the two *cdc2* ts mutants. These cells were elongated, arrested in G2 and also failed to form viable colonies (Figs 7B-7D and Figs S7A-S7E). Although, an earlier study in *S. cerevisiae* had also shown that Cdc28-K40Q and Cdc28-K40R failed to complement *cdc28* null cells vis-à-vis viability [13], the effect on cell cycle was not addressed. Importantly, the findings described in the current study are consistent with our hypothesis about the significance of the catalytic lysine-33, which is evolutionarily conserved, in rendering CDK1/Cdc2 kinase competent and hence its role in cell cycle progression.

Discussion

In this study, we establish the molecular mechanism of acetylation-dependent inhibition of CDK1 and its role in cell cycle progression. Protein acetylation has been largely shown to occur on lysine residues in proteins [53-55] and our results indicate CDK1 to be mono-acetylated. We identified P300 and SIRT1 as key enzymes that regulate CDK1 acetylation. Importantly, our results illustrate that CDK1 is hypoacetylated in G2/M synchronized cells as compared to those in G1/S. Earlier reports have shown that both SIRT1 and P300 interact with CDK1:Cyclin-B and the latter interaction led to phosphorylation-dependent degradation of P300 [24]. Our results now provide physiological significance for interaction between these key molecular components in regulating CDK1 acetylation and thus cell division cycles.

Capturing the importance of CDK1 deacetylation, mutating lysine-33 to glutamine that mimics a constitutively acetylated state, resulted in a kinase dead CDK1. Although, CDK1-K33R retained the charge state, it was inactive and mechanistically it was distinct from CDK1-K33Q (see below). These suggested

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

that the charge state and/or shape complementarity of the catalytic lysine are necessary for kinase activity. Notably, overexpression of CDK1-K33Q in the background of endogenous CDK1 did not lead to a dominant negative effect, unlike the phosphorylation defective mutants [36]. Given that CDK1 knockdown has been shown to have variable effects on mammalian cells [56] and to eliminate possible compensatory role of CDK2 [57], we addressed the importance of catalytic lysine acetylation using *S. pombe*. As stated earlier and well established in the field, Cdc2 and CDK1 are highly conserved both at structural and functional levels [46]. Highlighting the physiological relevance of catalytic lysine and its charge state, we found that mutating Cdc2 lysine-33 led to cell cycle arrest and reduced viability in *S. pombe*.

Structural studies and evidence based on ATP analogues have led to the assumption that the catalytic lysine is critical for both ATP binding and phosphoryl-transfer reaction [14-17]. In this context, our high resolution computational analyses and ATP-binding assays show that ATP is stably bound to CDK1:Cyclin-B even when lysine-33 is mutated to glutamine or replaced with acetyl-lysine. Given that most of the current understanding of kinase catalytic triads has come from PKA, including a very recent study [33], we compared the results on PKA with those on our CDK1:Cyclin-B complex. Unlike in PKA, our results show that perturbing lysine-33 does not alter the position of PSTAIRE-/C-helix vis-à-vis ATP. Nevertheless, we do show that in CDK1-K33Q and -K33Ac, ATP adopts distinct conformations in the active site pocket and the non-bonded interaction energies become less favorable relative to CDK1-WT. These insights distinguish the role of catalytic lysine in different kinase systems, which was previously unknown.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Oscillation of Cyclin levels during different cell cycle phases have been thought to be sufficient to form cognate complexes with CDKs to drive cell cycle progression [3]. Although, previous structural and mutational analyses of CDK:Cyclin complexes have elucidated the domains that mediate interactions [5, 7, 41], deterministic molecular mechanisms that enable complexation remained unclear. In this context, we have discovered a key role for catalytic lysine-33 in CDK1 in determining Cyclin-B binding using both computational and biochemical approaches, with an evident physiological effect on cell cycle progression. Specifically, mutating lysine-33 to acetyl mimic glutamine (K33Q) abrogated Cyclin-B1 binding to CDK1. Interestingly, this interaction was intact in CDK1-K33R, indicating that the charge state is necessary for enabling Cyclin-B1 binding. Further, this also shows that absence of kinase activity and cell cycle arrest in K-R mutants is independent of control via Cyclin-B binding. This could be due to abrogated phosphoryl-transfer reaction, as has been reported in other kinase systems including PKA [33]. We emphasize that none of the previous studies have dissected out the link between the catalytic lysine in CDKs and the interface interactions with Cyclins. Our exciting results highlight that intra-molecular electrostatic tethering of lysine-33 predisposes CDK1 to bind to Cyclin-B, which eventually leads to the formation of active kinase conformation. Thus, we establish that CDK:Cyclin interaction is regulated and determined by the catalytic lysine, contrary to our current understanding of this complexation to be default.

In conclusion, our results suggest that acetylation of CDK1, which is sub-stoichiometric and prevents Cyclin-B binding, marks a reserve pool of CDK1. Our findings highlight that deacetylation, which naturally unmasks the charge state

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

of lysine-33, is a key determinant of CDK1:Cyclin-B binding. Considering that other CDKs are also acetylated at similar active site lysine, acetylation dependent regulation might be a universal phenomenon of CDKs and could be true for other EPKs as well. Importantly, our study highlights how local perturbations involving a key catalytic residue have long-range effects on protein-protein interactions.

Materials and methods

Cell Culture and transfection: HEK293T, HeLa-TetOn and Mouse embryonic fibroblast (MEF) cells were grown in DMEM high glucose medium (Sigma-D777) supplemented with 10% fetal bovine serum (Gibco) or 10% newborn calf serum (Gibco) and 1% antibiotic-antimycotic solution (Gibco), and maintained under standard 5% CO_2 conditions. Cells were transfected using Lipofectamine 2000 (Life technologies) or Fugene 6 (Roche) as per manufacturer's instructions. For inducible expression, 2 μg/ml doxycycline (Sigma) was added to the medium as indicated. For inhibiting HDACs or Sirtuins, cells were treated with 400 nM Trichostatin A (TSA) or 5 mM Nicotinamide (NAM), respectively, for 16 h. Sf21 (*Spodoptera frugiperda*) insect cells (Thermo Fisher) were grown in Graces' Media containing antibiotic solution (Sigma) and 10% fetal bovine serum (Gibco) at 25^0C.

Yeast and bacterial culture: *Schizosaccharomyces pombe* strains were transformed with *pREP41* plasmid encoding wild type or mutant *cdc2* gene using fast lithium acetate transformation method [58]. All strains were cultured in YE or EMM (MP Biomedicals) media with necessary supplements. 10 μM thiamine in

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

EMM was used to suppress the *nmt41* promoter. For induction, cells initially cultured in EMM media containing thiamine were washed and subsequently cultured in EMM media without thiamine at 25°C. After 16 h of induction, the cultures were shifted to restrictive temperature of 36°C for 4 h. *E.coli DH5α* cells were cultured in LB media supplemented with 100 µg/ml ampicillin for amplification of plasmids. 2% agar was used for solid media.

Cell synchronization: Cells plated at 50% confluency were treated with 150 µM mimosine (Sigma) for 16 h to synchronize the cells in G1/S phase. To obtain cells synchronized in different phases, mimosine treated cells were washed twice with PBS and released into fresh medium. Cells collected at 2-3 h, 5-6 h and 7-8 h post release corresponded to S-phase, G2-phase and G2/M-phase respectively, as assessed by flow cytometery and phospho-H3 (S10) levels. To synchronize cells in G2 phase, 9 µM RO-3306 (Sigma) was used for 16 h and for G2/M synchronization, cells were treated with Nocodazole (100 ng/mL) for 8 h after mimosine release.

Flow cytometry: Cells were collected by trypsinization, washed with PBS and fixed using chilled 70% ethanol, overnight at -20^{0}C. The fixed cells were washed with thrice with PBS, treated with RNase A (Sigma) and stained using propidium iodide (Sigma) for 30 minutes at 37^{0}C. Cell-cycle profiles were acquired on a FACS Fortessa instrument (BD Biosciences) and analyzed using FACS Diva Software 6.0 (BD Biosciences).

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Cell lysate preparation: Cells were harvested and washed twice in ice cold PBS. Cells were lysed either in TNN (50 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.9% NP 40, 1 mM PMSF, PhosSTOP, Protease inhibitor cocktail) or RIPA buffer (50 mM Tris pH 8.0, 150 mM NaCl, 0.1% SDS, 0.5% Sodium deoxycholate, 1% Triton X 100, 0.1% SDS, 1 mM PMSF, PhosSTOP, Protease inhibitor cocktail) for 15 minutes on ice. When probing for acetylation, buffers were supplemented with 4 μM TSA and 5 mM Nicotinamide to inhibit the activity of deacetylases. The lysate was cleared by centrifugation at 12,000 rpm for 15 minutes at 4^0C and the supernatant collected as cell lysate. The protein concentration was measured using BCA kit (Sigma) as per manufacturer's protocol.

Immunoprecipitation and HA peptide elution: Cell lysates were pre-cleared with either Protein-A/-G sepharose beads (Sigma) for 1 h at 4^0C on rotor. Equal amount of protein was taken and the cell lysates were incubated overnight with the indicated antibody. Equal volume of Protein-A/-G beads were added and incubated for 2 h at 4^0C on rotor to pull down the immune complexes and subsequently washed thrice with the TNN lysis buffer for 10 minutes at 4^0C on rotor. The washed beads were then boiled in SDS gel loading buffer to elute the bound proteins. Alternatively, for eluting CDK1-HA immunecomplexes, beads were incubated in HA peptide elution buffer (50 mM Tris pH 8.0, 150 mM NaCl, 100 μg/ml HA peptide) for 2 h at 27^0C and with 1000 rpm mixing. The beads were pelleted and supernatant was boiled in SDS gel loading buffer.

FSBA labeling: CDK1-HA immunoprecipitates on beads were treated with 50 μM FSBA (Sigma) in 0.1% PBS-Triton-X-100 for 15 minutes at 30^0C with constant

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

shaking, washed four times with 0.5% PBS-Triton-X-100 for 5 minutes at 4^0C on rotor and boiled in SDS gel loading buffer to elute the bound proteins. Western blotting was performed using -FSBA antibodies and normalized to the amount of immunoprecipitated CDK1.

CDK1 kinase activity assay: Myc-tagged Cyclin-B1 was co-expressed with either CDK1-WT/-K33Q/-K33R-HA in HEK293T cells. 24 hr post-transfection, the cells were treated with Nocodazole (100 ng/mL) for 12 hr and harvested in TNN lysis buffer. The lysate was incubated with α-HA antibody overnight at 4^0C, followed by Protein-A sepharose beads for 2 hr at 4^0C. The beads were washed thrice with TNN buffer for 10 min each at 4^0C, twice with kinase buffer (50 mM Tris-Cl pH 7.5, 15 mM $MgCl_2$) for 10 min each at 4^0C, and divided in two parts, one for western blotting to estimate the amount of CDK1, and the other for the kinase activity assay. Immunoprecipitated CDK1 on Protein-A beads was incubated in kinase buffer with 1 mM DTT, PhosSTOP (Roche), 200 μM Mg^{2+}-ATP, 3 μCi [γ-^{32}P]ATP and 2 μg recombinant H1 protein (Millipore) for 30 min at 30^0C. The reactions were terminated by the addition of 1X LDS sample buffer (NP0007, Invitrogen), and heated at 95^0C for 5 min. Proteins were resolved on a 4-12% NuPAGE Bis-Tris gel (Invitrogen) and transferred to a PVDF membrane (GE Life Sciences). The blot was subjected to autoradiography (Typhoon FLA-9500) to monitor H1 phosphorylation, and stained with Ponceau S to detect the amount of H1. Bands were quantified using Image J. The extent of phosphorylation of H1 was quantified by first expressing the amount of radiolabeled H1 as a fraction of total H1, and then normalizing this to the amount of immunoprecipitated CDK1 detected by western blotting.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

ATP binding and competition assays: The ATP binding assay was performed using the ATP affinity kit (Jena Bioscience) as per manufacturer's protocol. Briefly, the TNN cell lysate was dialyzed overnight at 4^0C against dialysis buffer (PBS, 1mM EDTA, 1mM DTT) with two buffer changes to remove the bound ATP-Mg^{2+}. The dialyzed lysate was diluted with the binding buffer containing protease inhibitor cocktail and incubated with equilibrated control or ATP beads for 2 h at 4^0C on rotor. For ATP competition, increasing amount of ATP was added in the dialyzed lysate before incubation with equilibrated control or ATP beads. The beads were washed thrice with the wash buffer for 15 minutes at 4^0C on rotor and boiled in SDS gel loading buffer to elute the bound proteins.

Recombinant 6XHis-P300 expression and purification

6XHis-tagged full-length P300 was expressed by transfection of the recombinant baculovirus into Sf21 insect ovary cells. 72 h post infection, the cells were harvested and lysed in 10 mM Tris-Cl, 500 mM NaCl, 10% glycerol, 0.1% NP-40, 15 mM imidazole, 2 mM PMSF, and 2 mM β-mercaptoethanol followed by nickel-nitriloacetic acid (Ni-NTA) affinity chromatography-based purification as described previously [59].

Acetyltransferase Assay: In the acetyltransferase assays, 25 nM full-length P300 was incubated with 500 ng recombinant CDK1-Cyclin B1 (Merck-Millipore #14-450) in the presence or absence of 1.2 mM acetyl-CoA lithium salt (Sigma). The reaction was carried out in HAT assay buffer (50 mM Tris–HCl, pH 7.5, 1 mM PMSF, 0.1 mM EDTA and 10% v/v glycerol) and 100 mM sodium butyrate at 30

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

°C for 90 minutes. The enzyme and acetyl-CoA were replenished twice in the reaction at an interval of 30 minutes to ensure complete acetylation of the substrate. The reaction was stopped on ice, followed by the addition of 5X SDS sample loading buffer (10 % w/v SDS, 250 mM Tris-Cl, pH 6.8, 50% glycerol, 0.5% Bromophenol Blue, 5% β-mercaptoethanol).

2D-PAGE: After immunoprecipitation, the beads were solubilized in rehydration buffer (7 M urea, 2 M thiourea, 2 % CHAPS, 50 mM DTT and 2 % IPG buffer) in a final volume of 100 µl. The IPG strips (IPG ReadyStrip™ pH 5-8, 7 cm, Biorad #163-2004) were rehydrated overnight and IEF focusing was done with the following program:- Step 1 - 250V for 30 minutes, step 2 - 4000V for 1 h and final step 3 - 15000 Vhrs. The strips were equilibrated with 2% DTT (Sigma Aldrich) and 2.5% IAA (Sigma Aldrich) before proceeding to second dimensional gel electrophoresis on 10% SDS-PAGE gel.

Western Blotting: Equal amount of protein was run on SDS-PAGE and transferred to PVDF membrane (Merck-Millipore). The blots were probed with the indicated primary and secondary antibodies. The chemiluminescence signal was detected using SuperSignal West Pico PLUS and SuperSignal West Femto ECL kit (Thermo Scientific) and imaged using AI600 imager (GE-Amersham).

Microscopy: Cells were fixed using 3.7% formaldehyde for 12 minutes and permeabilized with 1% Triton. Cells were stained using 1µg/ml DAPI and 1 mg/ml Aniline Blue to visualize nucleus and cell wall respectively. Images were taken on Olympus 1X-71 Epifluorescence system with a 100X (NA 1.40) Olympus

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

objectiveand AndoriXon EM+ EMCCD camera with using Micro manager software.

MS/MS analysis: HEK293T cells were lysed by sonication in 20 mM HEPES containing 6 M urea and protease inhibitors (Complete, Roche). Proteins were reduced (5mM DTT), alkylated (10mM iodoacetamide), then digested with 1% trypsin (w/w) after dilution of urea to a concentration of 1M. Resulting peptides were desalted (Sep-pak C18), and acetylated peptides were enriched through immunoprecipitation with anti-Acetyl beads (PTM Scan kit, Cell Signaling Technology). Acetylated peptides were analyzed by nanoLC-MS/MS using an UltiMate 3000 RSLCnano system (Dionex, Amsterdam, The Netherlands) coupled to a LTQ-Velos Orbitrap mass spectrometer (Thermo Scientific, Bremen, Germany). Separation was performed on a C-18 column (75 μm ID x 15 cm, Reprosil C18) equilibrated in 95% solvent A (5% acetonitrile, 0.2% formic acid) and 5% solvent B (80% acetonitrile, 0.2% formic acid), using a gradient from 5 to 50% gradient of solvent B over 180 min at a flow rate of 300 nL/min. The LTQ Orbitrap Velos was operated in data-dependent acquisition mode with Xcalibur software. Survey scan MS was acquired in the Orbitrap on the 350–2,000 m/z range, with the resolution set to a value of 60,000. The 20 most intense ions survey scans were selected for fragmentation by collision-induced dissociation, and the resulting fragments were analyzed in the linear trap. Dynamic exclusion was used within 60 s to prevent repetitive selection of the same peptide. Raw mass spectrometry files were processed with the Mascot software. Peptide identification results were validated by the target-decoy approach at a FDR of 1%.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Modelling of CDK1:Cyclin-B:ATP ternary complexes: Our computational modelling studies were initiated using a crystal structure of Cyclin-B bound CDK1 (PDB ID: 4Y72) solved by Endicott and co-workers. The 4Y72 structure was bound to an inhibitor as well as the accessory protein CKS2, both of which were removed. The resultant structure lacked ATP and C-terminal residues (resid: 290-297) for CDK1. We modelled the missing C-terminal CDK1 residues (resid: 290-297) using the loop modeling module in Modeller version 9 [60]. An ATP molecule was then introduced in the active site of the CDK1:Cyclin-B binary complex using the crystal structure of CDK2:Cyclin-A:ATP ternary complex as a reference (PDB ID: 1FIN) to create a WT CDK1:Cyclin-B:ATP ternary complex model. Specifically, we used the ATP coordinates from the 1FIN structure and inserted it at the active site of the CDK1:Cyclin-B complex retaining the same interactions of ATP with the active site aspartate-146 residue as in the 1FIN structure. After this step, we created two more mutant CDK1:Cyclin-B:ATP complex models by replacing the lysine-33 in the WT model with either glutamine (K33Q) or acetyl lysine (K33Ac). For all 3 model complexes (WT, K33Q, K33Ac), hydrogen atoms were added using the psfgen utility in VMD [61]. Then the modelled complexes were immersed in a large rectangular water box (explicit solvent model) of dimension 126x145x128 $Å^3$. The net charge of the model complexes was +3 for WT and +2 for K33Q/K33Ac. Therefore 3 Cl^- ions were added to WT and 2Cl^- ions to K33Q/K33Ac solvent boxes to neutralize the systems.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Equilibration protocol and Simulations: All simulations were carried out using the NAMD program [62] (version 2.9) and the CHARMM36 force field (C36 FF) [63]. Acetyl lysine was parametrized using the FFTK toolkit [64] in VMD (see parametrization procedure below). The water model used in the simulations was TIP3P. Our simulations employed periodic boundary conditions with the Particle Mesh Ewald method for describing electrostatic interactions and a switching function for van der Waals interactions (10Å switching distance and 12Å cutoff). Each of the three CDK1:Cyclin-B:ATP models (WT, K33Q and K33Ac) were subjected to an initial 10000 step energy minimization. Subsequently, the system was gradually heated at a rate 6K/1ps from 0K to 300K followed by an equilibration at 300K for 50 ps. The minimization and heating protocol was carried out 3 times initially with all the protein heavy atoms, ATP and Mg^{2+} fixed (with free hydrogens, solvent), then with all the protein heavy atoms other than those from modelled residues (resid 290-297), ATP and Mg^{2+} fixed, and finally with harmonic constraints of 25 kcal/mol/$Å^2$ on the protein heavy atoms, ATP and Mg^{2+}. These steps were then followed by constant pressure and constant temperature (NPT) equilibration runs for 150 ps to stabilize the density of the system at 1atm pressure and temperature 300 K. The pressure of the system was maintained by the Nose-Hoover method in combination with Langevin dynamics to control the temperature of the system. The NPT protocol was repeated 3 more times by lowering the harmonic constraints on the system to 12, 6 and 3 kcal/mol/$Å^2$. Finally an unconstrained ~1 ns NPT (NPT-free) run was performed at 300K to arrive at the starting point of our production runs. The time step for all these simulations was 1 fs. In the K33Ac model, the larger size of acetyl-lysine relative to the WT lysine-33 lead to significant steric clashes during residue

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

placement at the active site. We therefore employed three different protocols on the K33Ac model which differed in terms of constraints on ATP and E51 during the optimization, temperature equilibration, and pressure equilibration steps described above to create three distinct structurally relaxed conformations of the K33Ac ternary complex: K33Ac-1 (ATP, E51 both constrained), K33Ac-2 (ATP unconstrained, E51 constrained), and K33Ac-3 (ATP constrained, E51 unconstrained). Following the equilibration protocol, 200 ns production runs were carried out for all five systems (WT, K33Q, K33Ac-1, K33Ac-2, K33Ac-3) which were then used for subsequent analysis. Table S1 summarizes all systems and simulations in the present manuscript.

Charmm36 parameters for acetyl-lysine: In acetyl-lysine, the side-chain amino group of lysine is altered by the attachment of an acetyl (-$COCH_3$) group at the epsilon Nitrogen (NE) position. Since no standard parameters exist in C36 FF for this modification, we parametrized the side chain acetyl head group of acetyl lysine attached to the epsilon carbon atom (CE) of Lysine (Fig S3A). This was done to give a fair description of the change in charges and bonded parameters on the side chain NZ and CE atoms, upon modifying the positively charged amino group to a neutral acetyl group. The acetylated head-group fragment is capped with an extra H atom at the lysine terminal for epsilon C to account for the absence of the bond between delta C and epsilon C. The charges on H are absorbed into the epsilon C when the acetylated head-group fragment is patched onto the lysine residue. The C36 FF comprises of non-bonded van der Waals and electrostatic interactions and covalent interactions comprising of bond, angle, and dihedrals terms. We assigned van der Waals parameters for atoms by

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

analogy with existing C36 FF parameters. Atomic partial charges for electrostatic interactions were assigned to reproduce ab-initio interaction energies of the acetylated head-group fragment obtained from an electronic structure calculation at HF/6-31G* level of theory. In these calculations the geometry of the acetylated head-group fragment was optimized at an ab-initio HF/6-31G* level of theory. Force constants for bonded interactions (covalent bonds and angles) were obtained from Hessian calculations on the optimized geometry of the acetylated fragment. Dihedral parameters were assigned to reproduce target potential energy surfaces (PES) scans as a function of torsion angle calculated at the HF/6-31G* level of theory. Below we provide the final topology and parameters for acetyl-lysine residue derived using FFTK.

CHARMM36 Topology file

```
RESI ALY       0.00
GROUP              !
ATOM N    NH1   -0.47 !    |
ATOM HN   H      0.31 ! HN-N
ATOM CA   CT1     0.07 !    |  HB1 HG1 HD1 HE1 HZ  OCD HM1
ATOM HA   HB1     0.09 !    |  |  |  |  |   |  |  |
GROUP              ! HA-CA--CB--CG--CD--CE--NZ--CO--CM--HM3
ATOM CB   CT2   -0.18 !    |  |  |  |  |          |
ATOM HB1  HA2     0.09 !    |  HB2 HG2 HD2 HE2        HM2
ATOM HB2  HA2     0.09 !  O=C
GROUP              !    |
ATOM CG   CT2   -0.18 !
```

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

```
ATOM HG1  HA2    0.09
ATOM HG2  HA2    0.09
GROUP
ATOM CD   CT2   -0.18
ATOM HD1  HA2    0.09
ATOM HD2  HA2    0.09
GROUP
ATOM CE   CT2   -0.598
ATOM HE1  HA2    0.227
ATOM HE2  HA2    0.227
GROUP
ATOM NZ   NZ    -0.244
ATOM HZ   HZ     0.247
GROUP
ATOM CM   CM    -0.418
ATOM HM1  HM     0.133
ATOM HM2  HM     0.133
ATOM HM3  HM     0.133
GROUP
ATOM CO   CO     0.822
ATOM OCD  OCD   -0.662
GROUP
ATOM C    C      0.51
ATOM O    O     -0.51
```

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

```
BOND CB CA   CG CB   CD CG   CE CD   NZ CE  CE HE1  CE HE2

BOND N  HN   N  CA    C  CA

BOND C  +N   CA HA   CB HB1  CB HB2  CG HG1

BOND CG HG2  CD HD1  CD HD2

DOUBLE   O  C

BOND NZ HZ

BOND CO NZ  CM CO  CM HM1  CM HM2  CM HM3

DOUBLE OCD CO

IMPR N -C CA HN  C CA +N O

CMAP -C  N  CA  C   N  CA  C  +N

DONOR HN N

DONOR HZ

ACCEPTOR O C

ACCEPTOR OCD NZ

CHARMM36 Parameter file:

BONDS

!V(bond) = Kb(b - b0)**2

!

!Kb: kcal/mole/A**2

!b0: A

!

!atom type Kb b0

!

NZ  HZ  489.677    1.010

NZ  CO  462.890    1.361
```

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

```
NZ  CT2 417.386    1.418

CO  OCD 767.853    1.236

CO  CM  296.309    1.536

CE  HE1 345.280    1.090

CE  HE2 345.280    1.090

CM  HM  363.627    1.093


ANGLES

!

!V(angle) = Ktheta(Theta - Theta0)**2

!

!V(Urey-Bradley) = Kub(S - S0)**2

!

!Ktheta: kcal/mole/rad**2

!Theta0: degrees

!Kub: kcal/mole/A**2 (Urey-Bradley)

!S0: A

!

!atom types     Ktheta    Theta0   Kub     S0

!

!

NZ  CT2 HA2 71.626   107.750

NZ  CO  CM  117.947  115.101

NZ  CO  OCD 122.322  122.967

HZ  NZ  CT2 59.174   119.740
```

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

```
HZ  NZ  CO  75.065   124.254

CO  CM  HM  66.097   110.245

CO  NZ  CT2 96.613   117.960

HE1 CT2 HE2 78.469   109.472

CM  CO  OCD 113.730  124.779

HM  CM  HM  45.989   107.983

CT2 CT2 NZ   70.000    113.5000 ! ALLOW    ALI PEP POL ARO! from NH1   CT1

CT2, for lactams, adm jr.



DIHEDRALS

!

!V(dihedral) = Kchi(1 + cos(n(chi) - delta))

!

!Kchi: kcal/mole

!n: multiplicity

!delta: degrees

!

!atom types          Kchi    n   delta

!

HZ  NZ  CO  CM  1.2250   2  0.00

HZ  NZ  CO  CM  1.2100   3  0.00

CT2 NZ  CO  OCD 0.5240   3  0.00

CT2 NZ  CO  OCD 2.6160   2  180.00

OCD CO  CM  HM  2.9810   2  0.00
```

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

```
OCD CO  CM  HM  0.3220   3  180.00
HZ  NZ  CO  OCD 0.7860   3  180.00
HZ  NZ  CO  OCD 2.3940   2  180.00
CT2 NZ  CO  CM  2.4350   3  180.00
CT2 NZ  CO  CM  3.0000   2  180.00
NZ  CO  CM  HM  0.4860   3  180.00
NZ  CO  CM  HM  2.4860   2  0.00
HZ  NZ  CT2 HA2 1.7510   2  0.00
HZ  NZ  CT2 HA2 0.0500   3  0.00
CO  NZ  CT2 HA2 0.0450   2  0.00
CO  NZ  CT2 HA2 0.0160   3  0.00
CO  NZ  CT2 HA1 0.0450   2  0.00
CT2 CT2 NZ  HZ  0.0000   1  0.00 ! ALLOW PEP ! from H    NH1  CT2  CT3, for
lactams, adm jr.
CT2 CT2 NZ  CO  1.8000   1  0.00 ! ALLOW PEP ! from CT2  CT1  NH1  C, for
lactams, adm jr.

IMPROPER
!
!V(improper) = Kpsi(psi - psi0)**2
!
!Kpsi: kcal/mole/rad**2
!psi0: degrees
!note that the second column of numbers (0) is ignored
!
```

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

```
!atom types        Kpsi              psi0
!


NONBONDED nbxmod  5 atom cdiel shift vatom vdistance vswitch -
cutnb 14.0 ctofnb 12.0 ctonnb 10.0 eps 1.0 e14fac 1.0 wmin 1.5
!
!V(Lennard-Jones) = Eps,i,j[(Rmin,i,j/ri,j)**12 - 2(Rmin,i,j/ri,j)**6]
!
!epsilon: kcal/mole, Eps,i,j = sqrt(eps,i * eps,j)
!Rmin/2: A, Rmin,i,j = Rmin/2,i + Rmin/2,j
!
!atom  ignored   epsilon    Rmin/2  ignored  eps,1-4     Rmin/2,1-4
!
CE  0.0  -0.056000  2.010000  0.0  -0.010000  1.900000
CM  0.0  -0.078000  2.040000  0.0  -0.010000  1.900000
CO  0.0  -0.110000  2.000000
HE  0.0  -0.034000  1.340000
HM  0.0  -0.024000  1.340000
HZ  0.0  -0.046000  0.224500
NZ  0.0  -0.200000  1.850000  0.0  -0.200000  1.550000
OCD 0.0  -0.120000  1.700000  0.0  -0.120000  1.400000


END
```

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Statistical analysis: Statistical analyses for biochemical data were performed using GraphPad Prism (Version 6.0) and Microsoft Excel 2013. For comparing two groups, Student's t-test was performed. Statistical analyses for CMD data were performed using MATLAB (version R2014a).

Acknowledgements: The following laboratories were supported by the indicated research grants. UK (TIFR/DAE 12P0122 and CEFIPRA 4503-1), RV (TIFR/DAE 12P0155), MM (TIFR/DAE 12P0128), RB (HFSP RGP0025/2016), TKK (JNCASR and Sir JC Bose Fellowship, DST-India; SR/S2/JCB-28/2010), AGP (Investissement d'Avenir Infrastructures Nationales en Biologie et Sant. program; ProFI, Proteomics French Infrastructure project, ANR-10-INBS-08). We thank Prof. Matthias Wymann (University of Basel, Switzerland) for providing α-FSBA antibodies.

Author Contributions: UKS/RV conceptualized the project with inputs from SD, MM, TKK, RB & AGP. SD, SR, NK, MB, AF, RS, SG, CB, SK, KKV, AGP, RV and UKS performed the experiments. SD, RV and UKS wrote the manuscript. All authors have contributed towards analysis and interpretation of data.

Conflict of interest: The authors declare no competing interests.

References:

1. Morgan DO (1997) Cyclin-dependent kinases: engines, clocks, and microprocessors. *Annu Rev Cell Dev Biol* **13**: 261-91

2. Malumbres M (2014) Cyclin-dependent kinases. *Genome Biol* **15**: 122

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

3. Crosby ME (2007) Cell Cycle: Principles of Control. *Yale J Biol Med* **80**: 141-2

4. Asghar U, Witkiewicz AK, Turner NC, Knudsen ES (2015) The history and future of targeting cyclin-dependent kinases in cancer therapy. *Nat Rev Drug Discov* **14**: 130-46

5. Ducommun B, Brambilla P, Félix MA, Franza BR, Karsenti E, Draetta G (1991) cdc2 phosphorylation is required for its interaction with cyclin. *EMBO J* **10**: 3311-9

6. Jeffrey PD, Russo AA, Polyak K, Gibbs E, Hurwitz J, Massague J, Pavletich NP (1995) Mechanism of CDK activation revealed by the structure of a cyclinA-CDK2 complex. *Nature* **376**: 313-20

7. Morris MC, Gondeau C, Tainer JA, Divita G (2002) Kinetic mechanism of activation of the Cdk2/cyclin A complex. Key role of the C-lobe of the Cdk. *J Biol Chem* **277**: 23847-53

8. Russo AA, Jeffrey PD, Pavletich NP (1996) Structural basis of cyclin-dependent kinase activation by phosphorylation. *Nat Struct Biol* **3**: 696-700

9. Kornev AP, Taylor SS (2010) Defining the conserved internal architecture of a protein kinase. *Biochim Biophys Acta* **1804**: 440-4

10. Endicott JA, Noble ME, Johnson LN (2012) The structural basis for control of eukaryotic protein kinases. *Annu Rev Biochem* **81**: 587-613

11. De Bondt HL, Rosenblatt J, Jancarik J, Jones HD, Morgan DO, Kim SH (1993) Crystal structure of cyclin-dependent kinase 2. *Nature* **363**: 595-602

12. Brown NR, Korolchuk S, Martin MP, Stanley WA, Moukhametzianov R, Noble ME, Endicott JA (2015) CDK1 structures reveal conserved and unique features of the essential cell cycle CDK. *Nat Commun* **6**: 6769

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

13. Choudhary C, Kumar C, Gnad F, Nielsen ML, Rehman M, Walther TC, Olsen JV, Mann M (2009) Lysine acetylation targets protein complexes and co-regulates major cellular functions. *Science* **325**: 834-40

14. Sabo A, Lusic M, Cereseto A, Giacca M (2008) Acetylation of conserved lysines in the catalytic core of cyclin-dependent kinase 9 inhibits kinase activity and regulates transcription. *Mol Cell Biol* **28**: 2201-12

15. Mateo F, Vidal-Laliena M, Canela N, Zecchin A, Martinez-Balbas M, Agell N, Giacca M, Pujol MJ, Bachs O (2009) The transcriptional co-activator PCAF regulates cdk2 activity. *Nucleic Acids Res* **37**: 7072-84

16. Lee J, Yun N, Kim C, Song MY, Park KS, Oh YJ (2014) Acetylation of cyclin-dependent kinase 5 is mediated by GCN5. *Biochem Biophys Res Commun* **447**: 121-7

17. Pillai VB, Sundaresan NR, Samant SA, Wolfgeher D, Trivedi CM, Gupta MP (2011) Acetylation of a conserved lysine residue in the ATP binding pocket of p38 augments its kinase activity during hypertrophy of cardiomyocytes. *Mol Cell Biol* **31**: 2349-63

18. Sarikhani M, Mishra S, Desingu PA, Kotyada C, Wolfgeher D, Gupta MP, Singh M, Sundaresan NR (2018) SIRT2 regulates oxidative stress-induced cell death through deacetylation of c-Jun NH. *Cell Death Differ*

19. Sarikhani M, Mishra S, Maity S, Kotyada C, Wolfgeher D, Gupta MP, Singh M, Sundaresan NR (2018) SIRT2 deacetylase regulates the activity of GSK3 isoforms independent of inhibitory phosphorylation. *Elife* **7**

20. Zhang H, Park SH, Pantazides BG, Karpiuk O, Warren MD, Hardy CW, Duong DM, Park SJ, Kim HS, Vassilopoulos A, *et al.* (2013) SIRT2 directs the

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

replication stress response through CDK9 deacetylation. *Proc Natl Acad Sci U S A* **110**: 13546-51

21. Santamaria D, Barriere C, Cerqueira A, Hunt S, Tardy C, Newton K, Caceres JF, Dubus P, Malumbres M, Barbacid M (2007) Cdk1 is sufficient to drive the mammalian cell cycle. *Nature* **448**: 811-5

22. Mayya V, Rezual K, Wu L, Fong MB, Han DK (2006) Absolute quantification of multisite phosphorylation by selective reaction monitoring mass spectrometry: determination of inhibitory phosphorylation status of cyclin-dependent kinases. *Mol Cell Proteomics* **5**: 1146-57

23. Coulonval K, Kooken H, Roger PP (2011) Coupling of T161 and T14 phosphorylations protects cyclin B-CDK1 from premature activation. *Mol Biol Cell* **22**: 3971-85

24. Wang SA, Hung CY, Chuang JY, Chang WC, Hsu TI, Hung JJ (2014) Phosphorylation of p300 increases its protein degradation to enhance the lung cancer progression. *Biochim Biophys Acta* **1843**: 1135-49

25. Sasaki T, Maier B, Koclega KD, Chruszcz M, Gluba W, Stukenberg PT, Minor W, Scrable H (2008) Phosphorylation regulates SIRT1 function. *PLoS One* **3**: e4020

26. Norbury C, Blow J, Nurse P (1991) Regulatory phosphorylation of the p34cdc2 protein kinase in vertebrates. *Embo j* **10**: 3321-9

27. Leroy D, Birck C, Brambilla P, Samama JP, Ducommun B (1996) Characterisation of human cdc2 lysine 33 mutations expressed in the fission yeast Schizosaccharomyces pombe. *FEBS Lett* **379**: 217-21

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

28. Hannink M, Donoghue DJ (1985) Lysine residue 121 in the proposed ATP-binding site of the v-mos protein is required for transformation. *Proc Natl Acad Sci U S A* **82**: 7894-8

29. Kamps MP, Sefton BM (1986) Neither arginine nor histidine can carry out the function of lysine-295 in the ATP-binding site of p60src. *Mol Cell Biol* **6**: 751-7

30. Weinmaster G, Zoller MJ, Pawson T (1986) A lysine in the ATP-binding site of P130gag-fps is essential for protein-tyrosine kinase activity. *EMBO J* **5**: 69-76

31. Ebina Y, Araki E, Taira M, Shimada F, Mori M, Craik CS, Siddle K, Pierce SB, Roth RA, Rutter WJ (1987) Replacement of lysine residue 1030 in the putative ATP-binding region of the insulin receptor abolishes insulin- and antibody-stimulated glucose uptake and receptor kinase activity. *Proc Natl Acad Sci U S A* **84**: 704-8

32. Iyer GH, Moore MJ, Taylor SS (2005) Consequences of lysine 72 mutation on the phosphorylation and activation state of cAMP-dependent kinase. *J Biol Chem* **280**: 8800-7

33. Meharena HS, Fan X, Ahuja LG, Keshwani MM, McClendon CL, Chen AM, Adams JA, Taylor SS (2016) Decoding the Interactions Regulating the Active State Mechanics of Eukaryotic Protein Kinases. *PLoS Biol* **14**: e2000127

34. Gould KL, Nurse P (1989) Tyrosine phosphorylation of the fission yeast cdc2+ protein kinase regulates entry into mitosis. *Nature* **342**: 39-45

35. Gould KL, Moreno S, Owen DJ, Sazer S, Nurse P (1991) Phosphorylation at Thr167 is required for Schizosaccharomyces pombe p34cdc2 function. *EMBO J* **10**: 3297-309

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

36. Krek W, Nigg EA (1991) Mutations of p34cdc2 phosphorylation sites induce premature mitotic events in HeLa cells: evidence for a double block to p34cdc2 kinase activation in vertebrates. *Embo j* **10**: 3331-41

37. Den Haese GJ, Walworth N, Carr AM, Gould KL (1995) The Wee1 protein kinase regulates T14 phosphorylation of fission yeast Cdc2. *Mol Biol Cell* **6**: 371-85

38. van den Heuvel S, Harlow E (1993) Distinct roles for cyclin-dependent kinases in cell cycle control. *Science* **262**: 2050-4

39. Taylor SS, Kornev AP (2011) Protein kinases: evolution of dynamic regulatory proteins. *Trends Biochem Sci* **36**: 65-77

40. Bayliss R, Fry A, Haq T, Yeoh S (2012) On the molecular mechanisms of mitotic kinase activation. *Open Biol* **2**: 120136

41. Child ES, Hendrychová T, McCague K, Futreal A, Otyepka M, Mann DJ (2010) A cancer-derived mutation in the PSTAIRE helix of cyclin-dependent kinase 2 alters the stability of cyclin binding. *Biochim Biophys Acta* **1803**: 858-64

42. Atherton-Fessler S, Parker LL, Geahlen RL, Piwnica-Worms H (1993) Mechanisms of p34cdc2 regulation. *Mol Cell Biol* **13**: 1675-85

43. Anostario M, Jr., Harrison ML, Geahlen RL (1990) Immunochemical detection of adenine nucleotide-binding proteins with antibodies to 5'-p-fluorosulfonylbenzoyladenosine. *Anal Biochem* **190**: 60-5

44. Baeza J, Dowell JA, Smallegan MJ, Fan J, Amador-Noguez D, Khan Z, Denu JM (2014) Stoichiometry of site-specific lysine acetylation in an entire proteome. *J Biol Chem* **289**: 21326-38

45. Gil J, Ramírez-Torres A, Chiappe D, Luna-Peñaloza J, Fernandez-Reyes FC, Arcos-Encarnación B, Contreras S, Encarnación-Guevara S (2017) Lysine

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

acetylation stoichiometry and proteomics analyses reveal pathways regulated by sirtuin 1 in human cells. *J Biol Chem* **292**: 18129-18144

46. Lee MG, Nurse P (1987) Complementation used to clone a human homologue of the fission yeast cell cycle control gene cdc2. *Nature* **327**: 31-5

47. Norbury C, Nurse P (1992) Animal cell cycles and their control. *Annu Rev Biochem* **61**: 441-70

48. Nurse P, Thuriaux P, Nasmyth K (1976) Genetic control of the cell division cycle in the fission yeast Schizosaccharomyces pombe. *Mol Gen Genet* **146**: 167-78

49. Nurse P, Bissett Y (1981) Gene required in G1 for commitment to cell cycle and in G2 for control of mitosis in fission yeast. *Nature* **292**: 558-60

50. Nurse P (1985) Cell cycle control genes in yeast. *Trends in genetics* **1**: 51-55

51. Coudreuse D, Nurse P (2010) Driving the cell cycle with a minimal CDK control network. *Nature* **468**: 1074-9

52. Fantes PA (1983) Control of timing of cell cycle events in fission yeast by the wee 1+ gene. *Nature* **302**: 153-5

53. Choudhary C, Weinert BT, Nishida Y, Verdin E, Mann M (2014) The growing landscape of lysine acetylation links metabolism and cell signalling. *Nat Rev Mol Cell Biol* **15**: 536-50

54. Verdin E, Ott M (2015) 50 years of protein acetylation: from gene regulation to epigenetics, metabolism and beyond. *Nat Rev Mol Cell Biol* **16**: 258-64

55. Ali I, Conrad RJ, Verdin E, Ott M (2018) Lysine Acetylation Goes Global: From Epigenetics to Metabolism and Therapeutics. *Chem Rev* **118**: 1216-1252

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

56. Cai D, Latham VM, Zhang X, Shapiro GI (2006) Combined depletion of cell cycle and transcriptional cyclin-dependent kinase activities induces apoptosis in cancer cells. *Cancer Res* **66**: 9270-80

57. L'Italien L, Tanudji M, Russell L, Schebye XM (2006) Unmasking the redundancy between Cdk1 and Cdk2 at G2 phase in human cancer cell lines. *Cell Cycle* **5**: 984-93

58. Keeney JB, Boeke JD (1994) Efficient targeted integration at leu1-32 and ura4-294 in Schizosaccharomyces pombe. *Genetics* **136**: 849-56

59. Kundu TK, Wang Z, Roeder RG (1999) Human TFIIIC relieves chromatin-mediated repression of RNA polymerase III transcription and contains an intrinsic histone acetyltransferase activity. *Mol Cell Biol* **19**: 1605-15

60. Sali A, Blundell TL (1993) Comparative protein modelling by satisfaction of spatial restraints. *J Mol Biol* **234**: 779-815

61. Humphrey W, Dalke A, Schulten K (1996) VMD: visual molecular dynamics. *J Mol Graph* **14**: 33-8, 27-8

62. Phillips JC, Braun R, Wang W, Gumbart J, Tajkhorshid E, Villa E, Chipot C, Skeel RD, Kalé L, Schulten K (2005) Scalable molecular dynamics with NAMD. *J Comput Chem* **26**: 1781-802

63. MacKerell AD, Bashford D, Bellott M, Dunbrack RL, Evanseck JD, Field MJ, Fischer S, Gao J, Guo H, Ha S, *et al.* (1998) All-atom empirical potential for molecular modeling and dynamics studies of proteins. *J Phys Chem B* **102**: 3586-616

64. Mayne CG, Saam J, Schulten K, Tajkhorshid E, Gumbart JC (2013) Rapid parameterization of small molecules using the Force Field Toolkit. *J Comput Chem* **34**: 2757-70

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Figure legends

Figure 1 – In-vivo CDK1 acetylation at lysine-33 is regulated by opposing actions of P300 and SIRT1.

(A) Immunoprecipitated CDK1-HA was probed with α-pan-acetyl lysine antibody to reveal acetylation.

(B) Identification of lysine-33 as the site of acetylation in CDK1 by LC-MS/MS analysis.

(C) 2D-PAGE of immunoprecipitated endogenous CDK1 from HEK 293T cells and immunoblot with α-pan-acetyl-lysine or α-CDK1-K33Ac antibodies confirms acetylation.

(D-E) Probing for acetylation of immunoprecipitated CDK1-HA from cells (D) co-transfected with P300, PCAF or GCN5, shows P300 as the major KAT and (E) treated with KDAC inhibitors indicates Nicotinamide (NAM) sensitive acetylation.

(F-G) Probing for acetylation of immunoprecipitated CDK1-HA from cells co-transfected with P300 or P300/SIRT1 shows SIRT1 mediated attenuation of P300 dependent CDK1 lysine-33 acetylation. (F) Representative immunoblot and (G) Quantification of pixel intensities from (F).

Error bars represent SEM for n=3 technical replicates from a representative experiment.

Figure 2 – CDK1 acetylation at lysine-33 inhibits kinase activity.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

(A-B) H1 phosphorylation by CDK1-K33Q expressed relative to the activity of CDK1-WT shows lysine-33 mutation abrogates kinase activity. Error bars represent SEM for n=3 technical replicates for two experimental repeats.

(C-D) Immunoprecipitated CDK1-WT-HA or -K33Q-HA probed with α-phospho-tyrosine-15 CDK1 (CDK1 Y15p) antibodies reveals no change in inhibitory CDK1-Y15 phosphorylation. Error bars represent SEM for n=3 technical replicates from a representative experiment.

Figure 3 – CMD simulations for WT, K33Q and K33Ac-1,-2,-3 CDK1:Cyclin-B:ATP ternary complexes.

(A) Workflow depicting modeling of WT, K33Q and K33Ac-1,-2,-3 CDK1:Cyclin-B:ATP ternary complexes.

(B-K) Structures for WT, K33Q and K33Ac-1,-2,-3 CDK1:Cyclin-B:ATP ternary complexes obtained from 200 ns NVT CMD simulations. (B-F) show CDK1 (multi-color) and Cyclin-B (orange) in secondary structure representation and ATP bound at the CDK1 active site is shown in atomic van der Waals spheres representation. (G-K) View of the active site showing key catalytic triad residues: lysine-33 (K33)/glutamine-33 (Q33)/acetyl-lysine (K33Ac), glutamate-51 (E51) and aspartate-146 (D146) interacting with the bound ATP and Mg^{2+} (green).

Figure 4 – Loss of CDK1 lysine-33 charge state impinges on active site ATP conformation and weakens CDK1:Cyclin-B complexation.

(A) Fractional Solvent Accessible Surface Area (SASA) for ATP (ratio of values in ternary complex and free solution) obtained during 200 ns NVT CMD trajectories for WT, K33Q and K33Ac CDK1:Cyclin-B:ATP systems are compared.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

(B) Comparison of non-bonded interaction energies (E_{NB}) of ATP with surrounding active site residues within a 3-7 Å radius in WT, K33Q and K33Ac CDK1:Cyclin-B:ATP systems. Error bars represent SD.

(C) A comparison of non-bonded interaction energy (E_{NB}) of ATP with catalytic triad residues, residue-33 (K33, Q33, K33Ac), aspartate-146 (D146), and glutamate-51 (E51) in WT, K33Q and K33Ac CDK1:Cyclin-B:ATP systems highlights significant contributions from K33, which are reduced upon mutation. Error bars represent SD.

(D) ATP orientations and Mg^{2+} (green) coordination spheres (including solvation) in WT, K33Q and K33Ac CDK1:Cyclin-B:ATP complexes. The ATP active site conformation and Mg^{2+} coordination are altered upon mutating lysine-33.

(E) Histograms of Centre of Mass (COM) distances between CDK1 and Cyclin-B reveal greater separations and broader distributions of distances in K33Q and K33Ac CDK1:Cyclin-B:ATP complexes relative to WT.

Figure 5 – CDK1 acetylation at lysine-33 does not affect ATP binding but reduces Cyclin-B1 interaction.

(A) Probing FSBA labeled CDK1-WT/-K33Q/-K33R-HA immunoprecipitates with α-FSBA antibodies revealed reduced binding upon K33 mutation.

(B) Probing for CDK1-WT-HA and -K33Q-HA binding (pull-down) by ATP conjugated agarose beads (6-AH beads) shows no change in steady-state binding. Coomassie staining of total bound proteins was used for normalization.

(C) Quantification of ATP binding in (B). Error bars represent SEM for n=5 technical replicates from two experimental repeats.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

(D-G) Reversible co-immunoprecipitations of CDK1-WT-HA or -K33Q-HA and Myc-Cyclin-B1 shows reduced CDK1:Cyclin-B1 binding upon K33Q mutation. Immunoprecipitation of complexes with (D) α-HA and (E) α-Myc antibodies. (F) and (G) Quantifications of results presented in (D) and (E), respectively. *marks the IgG heavy chain. Error bars represent SEM for n=3 technical replicates from a representative experiment.

(H) Probing for acetylation in immunoprecipitated CDK1-HA shows reduced CDK1 lysine-33 acetylation in G2/M synchronized cells as compared to G1/S phase. Phospho-H3 S10 (H3-S10p) was used as a marker for G2/M population.

(I) Immunoprecipitated CDK1-HA shows negative correlation between CDK1 lysine-33 acetylation and CDK1:Cyclin-B1 complexation on probing with α-K33Ac and α-Myc antibodies.

(J) Probing immunoprecipitated CDK1-HA with α-K33Ac and α-Myc antibodies from cells transfected with Myc-Cyclin-B1 or Myc-Cyclin-B1/P300 shows enhancing lysine-33 acetylation reduces Cyclin-B1 binding.

Figure 6 – Loss of CDK1 lysine-33 charge state affects surface electrostatic interactions with Cyclin-B.

(A) Reduction in the number of interfacial salt bridges between CDK1 and Cyclin-B in CDK1:Cyclin-B:ATP K33Q (top panel) and K33Ac systems (bottom panel) relative to WT.

(B) Reduction in non-bonded interaction energies between acidic and basic residues at CDK1:Cyclin-B interface in CDK1:Cyclin-B:ATP K33Q (top panel) and K33Ac systems (bottom panel) relative to WT.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

(C) View of the CDK1:Cyclin-B interface and its connectivity to the active site with catalytic triad residues (K33, D146, E51) and ATP. Key acidic and basic residues at the CDK1:Cyclin-B interface are also shown. In this representation, we show only residues 30-148 and 250-280 in CDK1.

(D) Comparison of non-bonded interaction energies (E_{NB}) of key acidic and basic residues of CDK1 with salt bridge partner residues of Cyclin-B shows maximum contribution by E41, which is dependent on K33 charge state. Error bars represent SD. The analysis is for data from the beginning of first 1 ns of NVT production runs for WT and mutant systems before CDK1-Cyclin COM-COM separations increase for the latter.

Figure 7 – Cdc2/CDK1 lysine-33 mutation blocks cell-cycle progression in *S. pombe* and fails to compliment *cdc2* ts mutants.

(A) Schematic representation of cell length observed in fission yeast temperature sensitive *cdc2* mutants at permissive temperature (normal cell cycle progression) and restrictive temperature (cell cycle arrest).

(B) Representative images of DAPI and aniline blue co-stained cells of fixed samples after 16 h of induction, before (top panel) and after (bottom panel) shift at restrictive temperature for 4 h. Cells fixed after 4 h incubation at 36°C.

(C) Quantification of cell length measurements observed in *cdc2-33 and cdc2-L7* strains over-expressing indicated plasmids at restrictive temperature for 4 h. Measurement indicates mean cell length and SEM for three independent experiments where n=500 for each sample.

(D) Plate growth assay after 16 h of induction. Samples were 1:4 serially diluted and plates were incubated at indicated temperature for 4-5 days.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Supplemental figure legends

Figure S1 – CDK1 is acetylated at lysine-33 in-vivo.

(A) Flow chart of the protocol for affinity purification of α-CDK1-K33Ac antibody.

(B) Sequence of the peptide used for generating α-CDK1-K33Ac antibody and the high conservation of that region across species.

(C) ELISA of purified antibodies after the 3rd immunization on acetylated and non-acetylated peptides shows high specificity of α-CDK1-K33Ac antibodies.

(D) Probing for acetylation of HA immunoprecipitates from cells expressing CDK1-WT-HA or -K33R-HA shows specificity of α-CDK1-K33Ac antibody.

(E) 2D-PAGE of immunoprecipitated endogenous CDK1 from mouse embryonic fibroblasts (MEFs) and immunoblot with α-CDK1-K33Ac antibodies confirms acetylation.

(F) In-vitro acetylation assay using recombinant CDK1:Cyclin-B1 complex and 6XHis-P300 shows P300 dependent acetylation of CDK1.

(G) Immunoblot of HA immunoprecipitates from cells co-transfected with SIRT1 shows CDK1-SIRT1 interaction.

Figure S2 – CDK1 lysine-33 acetylation abolishes kinase activity but does not affect cell cycle progression.

(A) H1 phosphorylation by CDK1-WT, -K33Q and -K33R shows lysine-33 mutation abrogates kinase activity.

(B) Flow cytometry analysis of cells used for the kinase assay in Fig 2A and Fig 2B.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

(C) Flow cytometry analysis of HeLa-pTetON cells transfected with CDK1-WT or -K33Q shows no change in cell cycle profile.

(D) Immunoblot showing the expression of CDK1-WT and -K33Q for cells used for flow cytometry analysis in (C).

Figure S3 – CMD simulations for WT, K33Q and K33Ac-1,-2,-3 CDK1:Cyclin-B:ATP ternary complexes and the impact of CDK1 lysine-33 mutation/acetylation on ATP-active site and ATP-PSTAIRE helix interactions.

(A) Acetylated head-group fragment for which Charmm36 parameters were derived using FFTK.

(B-K) Modeled pre-simulation structures for WT, K33Q and K33Ac-1,-2,-3 CDK1:Cyclin-B:ATP ternary complexes obtained from the crystal structure using the workflow depicted in Fig 3A.

(B-F) show CDK1 (multi-color) and Cyclin-B (orange) in secondary structure representation. ATP bound at the CDK1 active site is shown in atomic van der Waals spheres representation.

(G-K) View of the active site showing key catalytic triad residues, lysine-33 (K33)/glutamine-33 (Q33)/acetyl-lysine (K33Ac), glutamate-51 (E51) and aspartate-146 (D146) interacting with the bound ATP and Mg^{2+} (green).

(L) Non-bonded interaction energies of ATP with acidic (left panel) and basic (right panel) residues within 3-7Å in WT, K33Q and K33Ac CDK1:Cyclin-B:ATP systems are favorable. Error bars represent SD.

(M-O) Interactions of K33 with ATP and E51 during 200 ns MD trajectories for the WT CDK1:Cyclin-B:ATP complex (M) Non-bonded (electrostatic + van der

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Waals) interaction energies, (N) Distance between the sidechain amino nitrogen of lysine and the two carboxylate oxygens of glutamate, (O) distance between the sidechain amino nitrogen of lysine and three of the closest oxygens of the ATP phosphate tail.

(P-R) Interactions between the PSTAIRE helix and ATP do not change in WT, K33Q and K33Ac CDK1:Cyclin-B:ATP systems. (P) Distance between the PSTAIRE helix (COM) and ATP (COM), (Q) distance between E51 (COM) and ATP (COM), (R) Non-bonded interaction energy between ATP and E51.

Figure S4 – PSTAIRE helix and T-loop undergo a conformational change upon CDK1 lysine-33 mutation/acetylation.

(A) Relative orientation of PSTAIRE helix in a representative snapshot of CDK1-K33Q complex relative to its orientation in the first frame for CDK1-WT.

(B) Rotation of PSTAIRE helix during 200 ns MD trajectories for WT, K33Q and K33Ac CDK1:Cyclin-B:ATP systems. The angle made by the PSTAIRE helix axis vector during each frame relative to its direction in the first frame of the CDK1-WT trajectory (CDK1 backbone alignment) was computed.

(C) Percentage solvent accessibility (fractional SASA of T-loop with and without surrounding protein) computed for the T-loop segment (residues 152-171) along the 200 ns MD trajectory.

(D) Representative conformations of the T-loop during 200 ns MD production runs for WT, K33Q and K33Ac CDK1:Cyclin-B:ATP systems. The full CDK1:Cyclin-B complex is shown on the left and the view on the right zooms in on the T-loop and its neighbourhood.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Figure S5 – CDK1 acetylation at lysine-33 does not affect ATP binding but reduces Cyclin-B1 interaction.

(A) Probing for pull-down of CDK1-WT-HA using blank or different ATP conjugated agarose beads shows highest binding to 6-AH beads.

(B) Flow cytometry analysis of cells used for the ATP binding assay in Fig 5B and Fig 5C.

(C) Probing for CDK1-WT/-K33Q/-K33R-HA binding (pull-down) by 6-AH beads shows no change in steady-state binding. S6K binding was used for normalization.

(D, E) ATP competition assay of CDK1-WT/-K33Q/-K33R-HA using blank or 6-AH beads shows small differences. Coomassie staining was used for normalization. Error bars represent SEM for three experimental replicates.

(F, G) Reversible co-immunoprecitations of CDK1-WT/-K33Q/-K33R/-D146N-HA and Myc-Cyclin-B1 shows reduced CDK1:Cyclin-B1 binding upon K33Q mutation. Immunoprecipitation of complexes with (F) α-Myc and (G) α-HA antibodies. * marks the IgG heavy chain.

(H) Flow cytometry analysis of cells used for assaying CDK1 acetylation in Fig 5H.

Figure S6 – Loss of CDK1 lysine-33 charge state affects surface interactions with Cyclin-B.

(A, B) Histograms of interfacial salt bridges between (A) sidechains and (B) side chain-backbone of acidic and basic residues of CDK1 and Cyclin-B show reduced number of salt-bridges in K33Q and K33Ac CDK1:Cyclin-B:ATP systems relative to WT.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Figure S7 – Phenotypic characterization of fission yeast *cdc2-33* and *cdc2-L7* mutants in presence of plasmid expressing Cdc2-WT/-K33Q/-K33R.

(A) Quantification of cell length measurements observed in *cdc2-33* and *cdc2-L7* strains transformed with indicated plasmid at permissive temperature of 25°C.

(B) Quantification of binucleate cells after 16 h of induction at 25°C.

(C) Quantification of binucleate cells observed when shifted to restrictive temperature of 36°C for 4 h after 16 h of induction. Error bar indicates SEM for three technical replicates of independent experiments where n=500 for each sample.

(D) Representative images of DAPI and aniline blue co-stained cells at 25°C (top panel) and 36°C (lower panel) respectively.

(E) Plate viability assay of *cdc2-L7* strains over-expressing indicated plasmids at 25°C and 36°C respectively.

Supplementary tables

Table S1 – Summary of all CDK:Cyclin-B:ATP ternary complexes and their simulation trajectories.

Table S2 – Salt bridges between acidic and basic residues of CDK1 and Cyclin-B within 4 Å of the binding interface formed during the first 1 ns of the 200 ns MD production (NVT) simulations for the 5 different model complexes. Salt bridges in the mutant (K33Q and K33Ac models) complexes, also present in the WT, are highlighted in green while those that are distinct from WT are highlighted in red.

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a CC-BY-ND 4.0 International license.

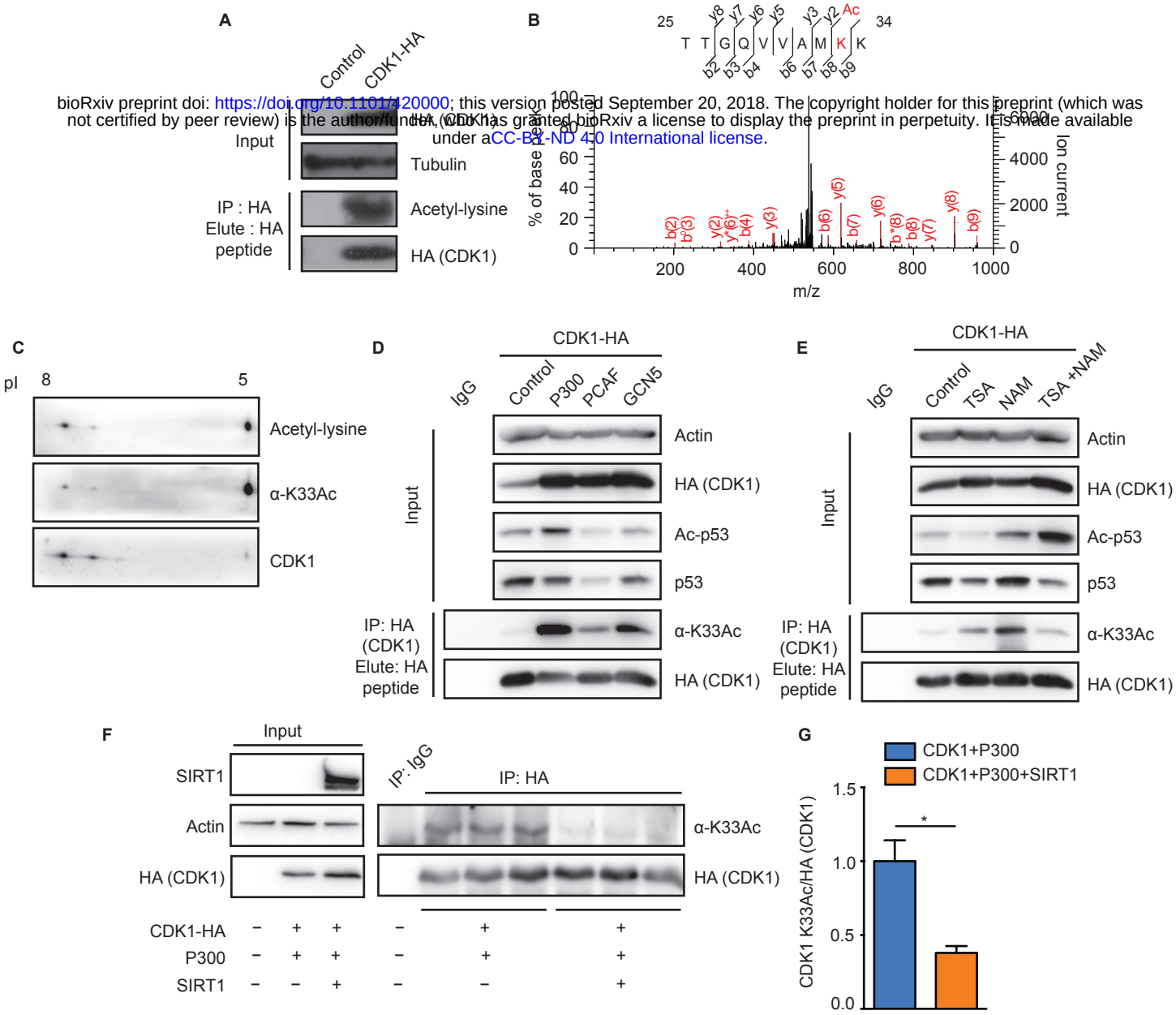


Figure 1

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

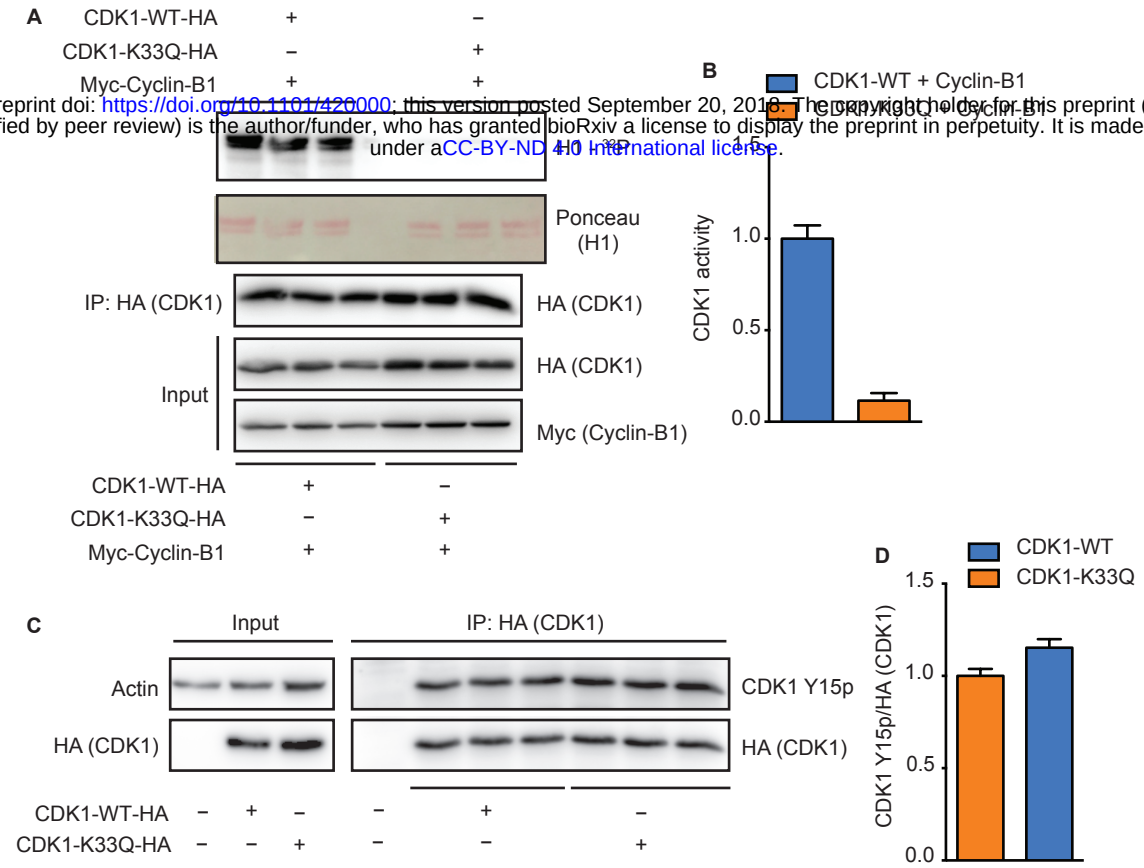


Figure 2

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

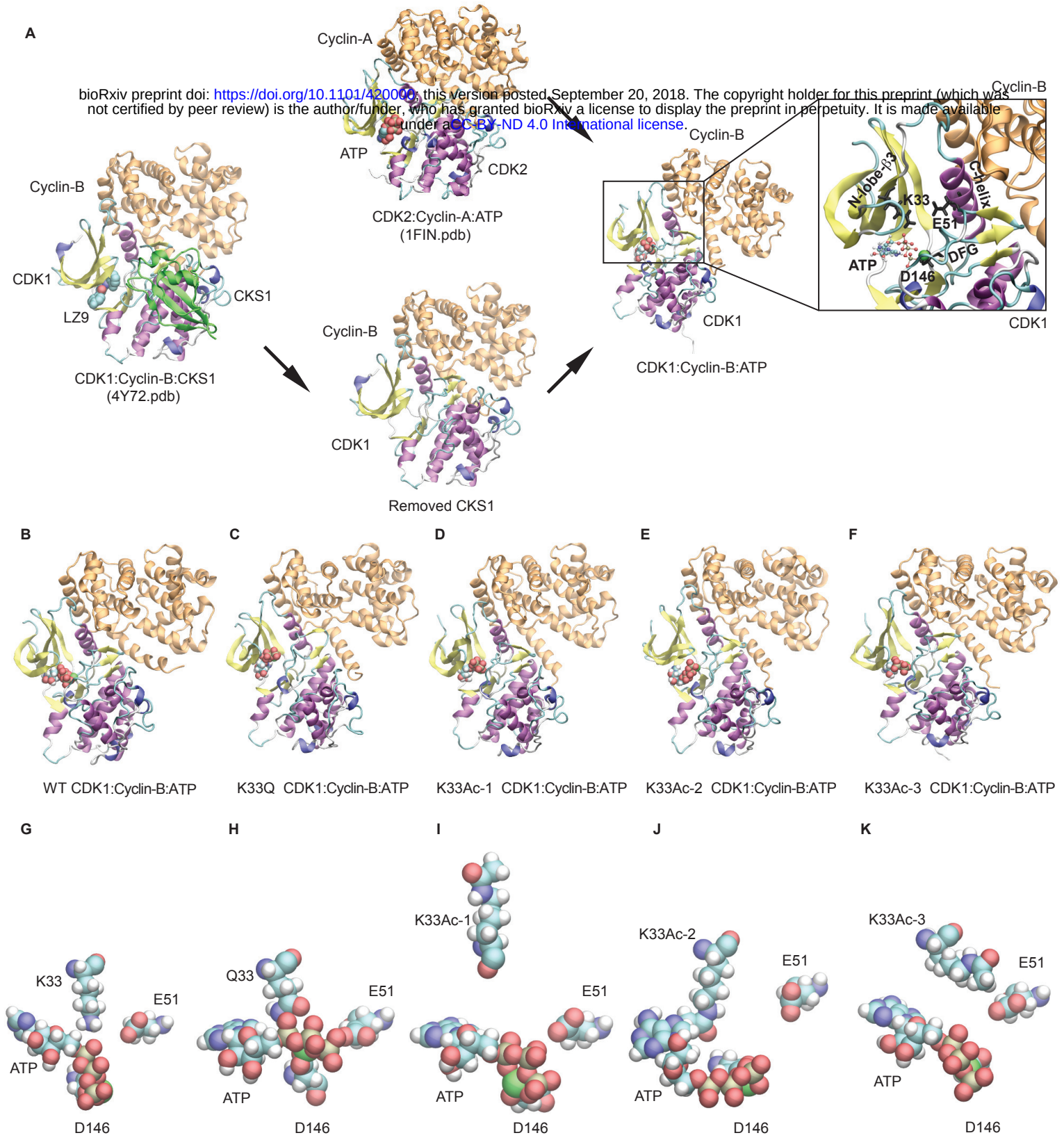
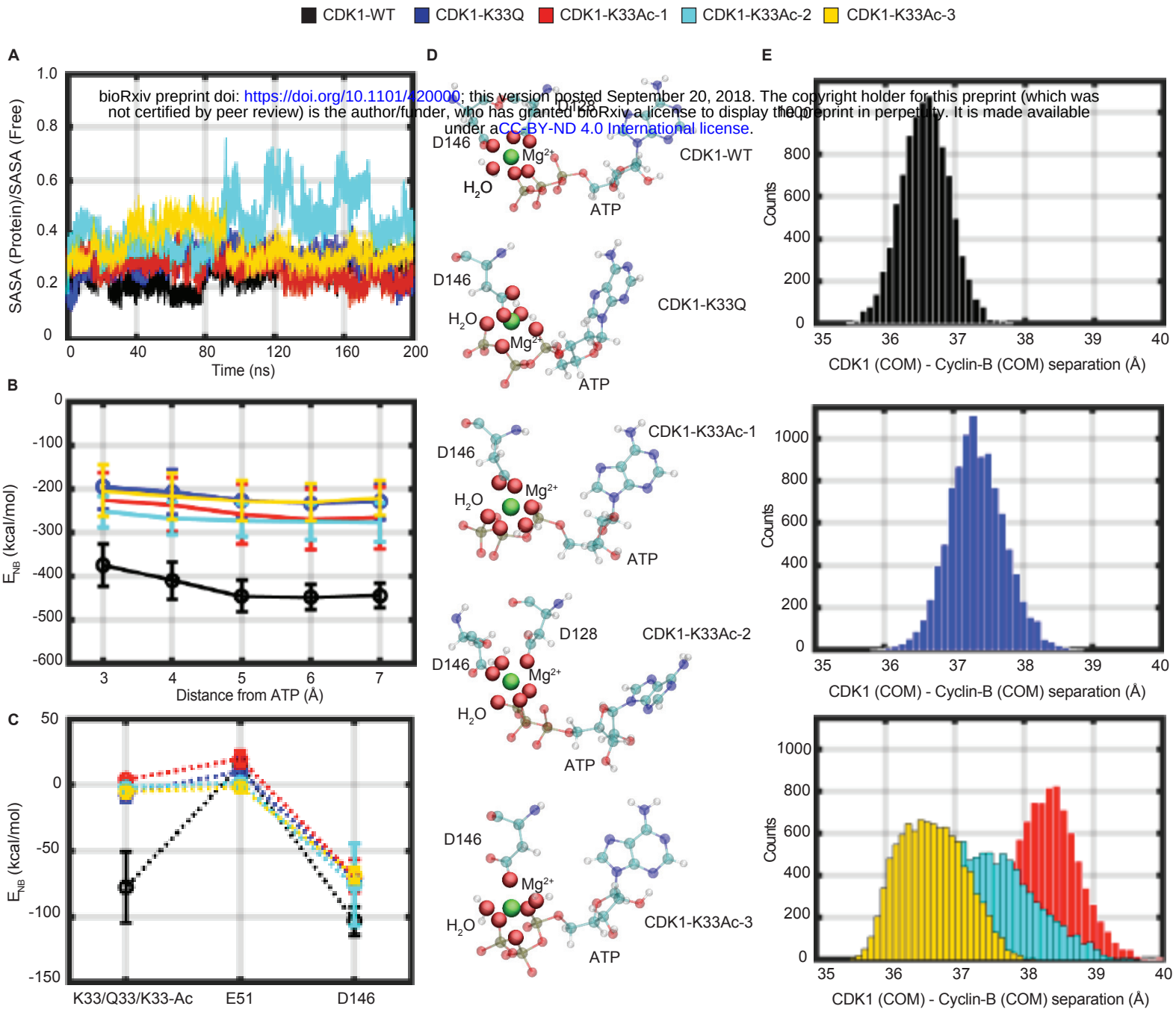


Figure 3



bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Figure 4

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

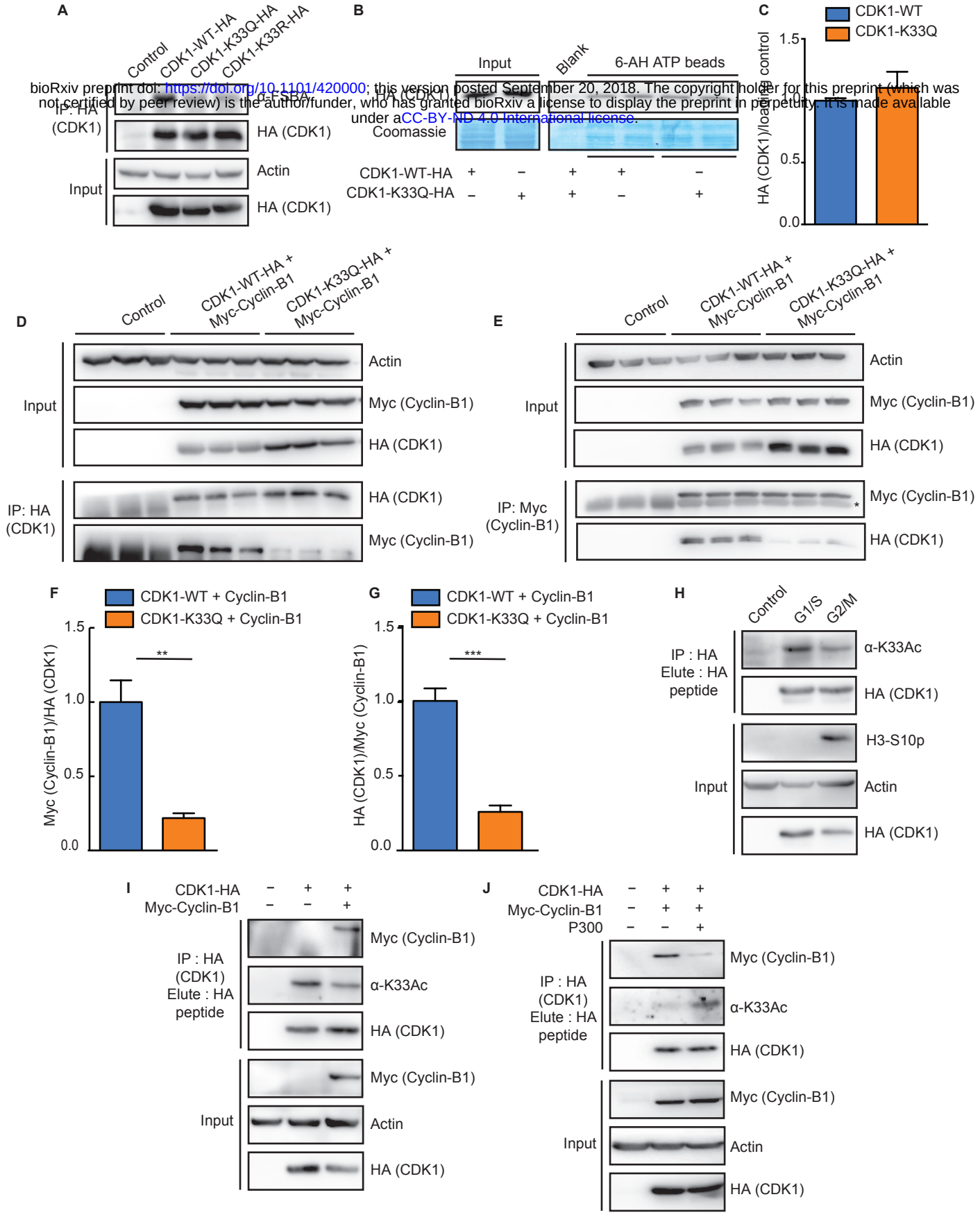
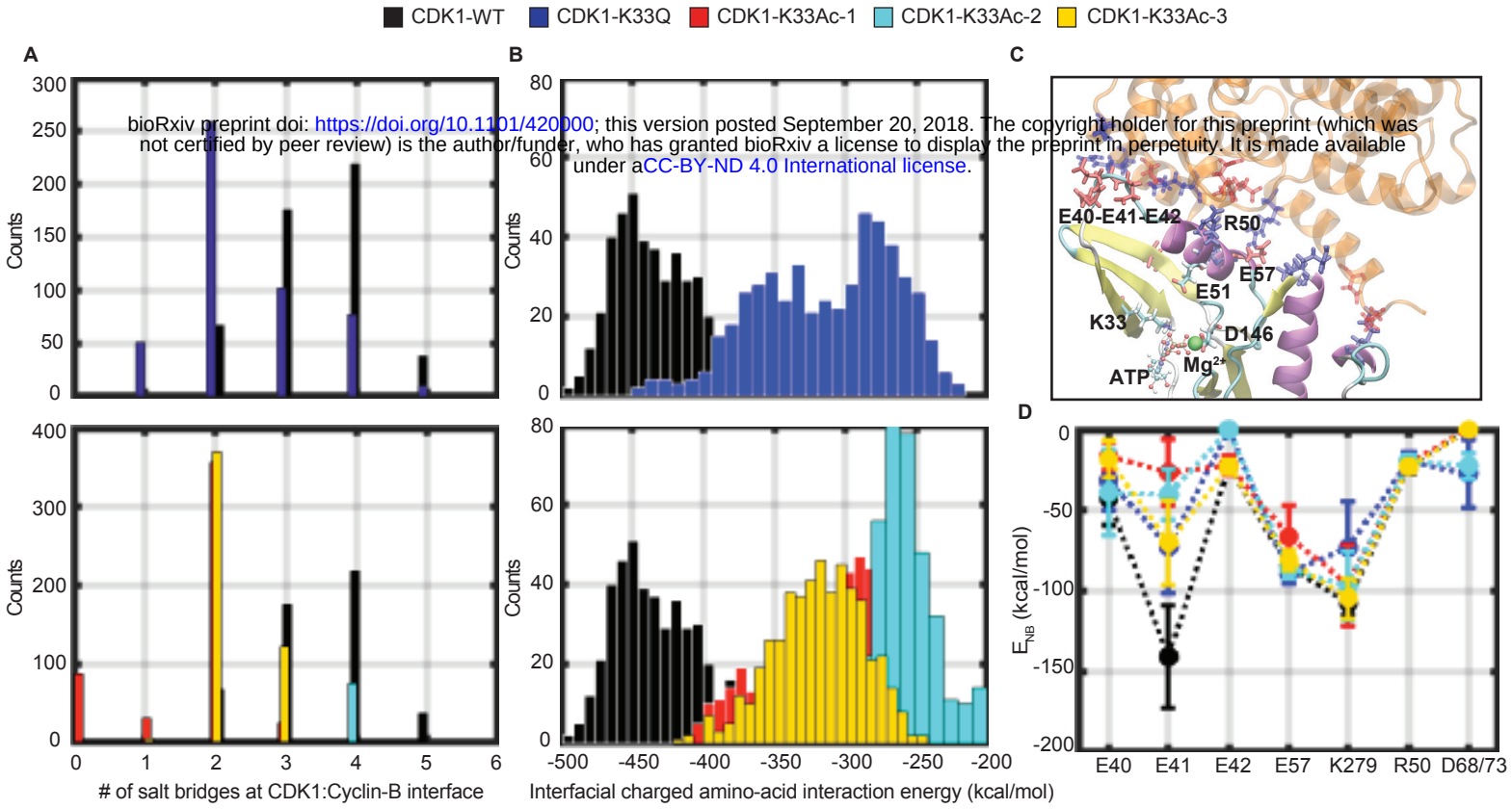


Figure 5



bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

Figure 6

bioRxiv preprint doi: https://doi.org/10.1101/420000; this version posted September 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a CC-BY-ND 4.0 International license.

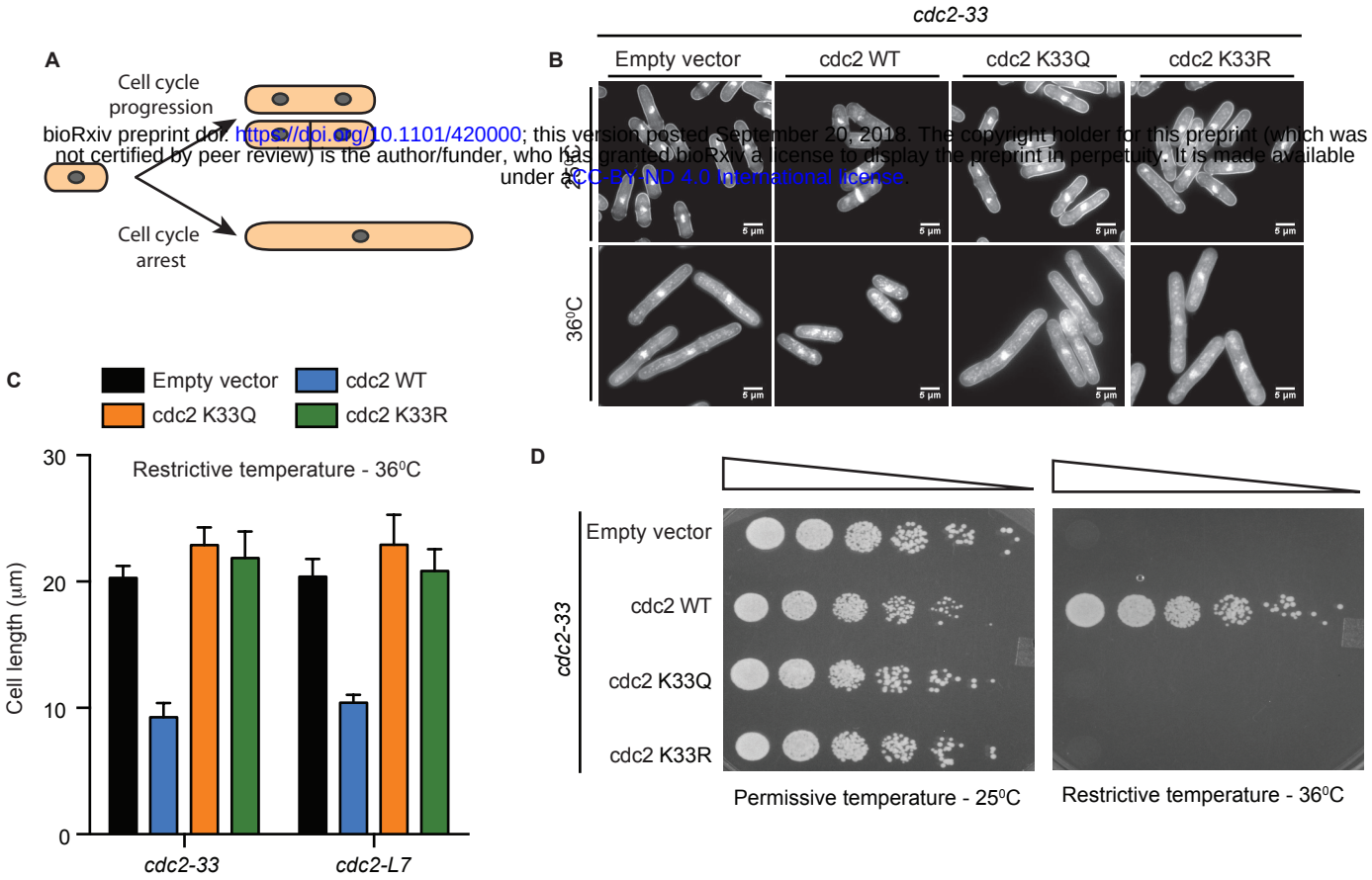


Figure 7