

1 **Interspecific plant competition mediates the metabolic and ecological**
2 **signature of a plant-herbivore interaction under warming and elevated**
3 **CO₂**

4 Helena Van De Velde^{1,2 †}, Hamada AbdElgawad^{3,4 †}, Han Asard³, Gerrit T. S. Beemster³,
5 Samy Selim⁵, Ivan Nijs² and Dries Bonte^{1*}

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7 *¹Terrestrial Ecology Unit, Department of Biology, Ghent University, K.L. Ledeganckstraat*
8 *35, B-9000 Ghent, Belgium*

9 *²Centre of Excellence Plants and Ecosystems, Department of Biology, University of Antwerp,*
10 *Universiteitsplein 1, B-2610 Wilrijk, Belgium*

11 *³Integrated Molecular Plant Physiology Research Group (IMPRES), Department of Biology,*
12 *University of Antwerp, Groenenborgerlaan 171, B-2020 Antwerp, Belgium*

13 *⁴Department of Botany and Microbiology, Faculty of Science, University of Beni-Suef, Beni-*
14 *Suef 62511, Egypt.*

15 *⁵Department of Clinical Laboratory Sciences, College of Applied Medical Sciences, Aljouf*
16 *university, Sakaka, P.O. 2014, Saudi Arabia*

17

18 † joint first author

19 *Corresponding author

20 Tel.: +32 (0)9 264 52 13

21 E-mail address: dries.bonte@ugent.be

22 Email addresses other authors: helena.vd.velde@gmail.com; han.asard@uantwerpen.be,

23 ivan.nijs@uantwerpen.be, Gerrit.beemster@uantwerpen.be,

24 Hamada.AbdElgawad@uantwerpen.be, sabdulsalam@ju.edu.sa

25 Abstract

- 26 1. Biotic interactions shape community evolution, but we lack mechanistic insights on
27 how metabolic and ecological processes under climate change are altered by biotic
28 interactions.
- 29 2. We used a two-trophic model community consisting of the aphid *Dysaphis*
30 *plantaginea* feeding on the forb *Plantago lanceolata*, and a grass competitor *Lolium*
31 *perenne* that does not experience herbivory by the aphid. Monocultures and mixtures
32 were exposed to the herbivory treatment and to three relevant simulated environmental
33 changes as prevalent under current climate change (increased temperature, CO₂, and
34 increased temperature and CO₂)
- 35 3. Elevated CO₂ reduced the nitrogen content of *P. lanceolata*, while simultaneous
36 increases of CO₂ and temperature modified the plant metabolic component and the
37 magnitude of these responses in different directions. Elevated CO₂ enhanced defence
38 systems in *P. lanceolata*, but these effects were not altered by warming. Interspecific
39 plant competition did, however, neutralise these responses. There were no indications
40 for indirect effects of climate on aphid population growth as mediated by changes in
41 plant defence, nutritional quality or biomass.
- 42 4. We thus demonstrate interactions between abiotic and biotic processes on plant
43 metabolite profiles, but more importantly, that climate change effect on a selection of
44 the metabolic pathways are altered by herbivory and competition. Our experiment
45 under semi-natural conditions thus demonstrates the non-additive and often
46 neutralizing effects of biotic interactions on plant metabolism and species performance
47 under climate-associated environmental change.

48 **Introduction**

49 Interactions between insect herbivores and host plants are influenced by primary and
50 secondary metabolites. The concentrations and types of these molecules in plants is likely
51 affected by climate change (Bidart-Bouzat & Imeh-Nathaniel, 2008; Zvereva & Kozlov,
52 2006). For example, plants exposed to elevated CO₂ show higher concentrations of many
53 organic molecules, including carbohydrates (fertilization effect), but generally lower or
54 unaltered nitrogen levels, and consequently higher C:N ratios (Bezemer & Jones, 1998;
55 Lincoln *et al.*, 1993; Robinson *et al.*, 2012; Stiling & Cornelissen, 2007). Lower nitrogen
56 concentrations imply lower levels of leaf protein and amino acids and reduced nutritive value
57 for herbivores (Lincoln *et al.*, 1986). Consequently, the performance of insects generally
58 reduces under high CO₂ (Bezemer & Jones, 1998; Hunter, 2001; Lincoln *et al.*, 1993).
59 Contrary to elevated CO₂, moderate warming has positive effects on insect herbivore
60 performance by reducing development time (Bale *et al.*, 2002; Van De Velde *et al.*, 2016) and
61 increasing fecundity (Meisner *et al.*, 2014). Climate change may thus paradoxically constrain
62 rather than facilitate insect herbivore population growth by lowering plant quality (Bauerfeind
63 & Fischer, 2013; Jamieson *et al.*, 2015). These resource-quality mediated effects are,
64 however, anticipated to not fully counterbalance the direct effects of warming on
65 development, rendering the overall herbivore response to warming positive (Bauerfeind &
66 Fischer, 2013; Zvereva & Kozlov, 2006).

67

68 Secondary metabolites (e.g. lignin, tannins, phenolics and terpenoids), greatly affect tissue
69 quality by determining the nutritive value, palatability, digestibility and/or toxicity of foliage
70 (see Table 1 for a synthesis). Compared to primary metabolites, responses of secondary
71 metabolites to elevated CO₂ and warming are more variable and less understood (Bidart-
72 Bouzat & Imeh-Nathaniel, 2008; Robinson *et al.*, 2012). According to the carbon-nutrient

73 balance hypothesis (Bryant *et al.*, 1983), elevated CO₂ is expected to increase secondary
74 metabolite levels as a result of the ‘excess’ carbon (fertilizing). Yet, because of the relatively
75 lower N-availability, the increase of nitrogen-containing metabolites will be lower than
76 expected (Robinson *et al.*, 2012). On the other hand, trade-offs between growth and
77 secondary metabolisms are anticipated to promote primary metabolic activity under
78 warming and therefore favour photosynthesis and growth relative to defence. This
79 expectation corresponds to the growth-differentiation balance hypothesis (Herms & Mattson,
80 1992). The different components associated with climate change thus induce direct, and often
81 orthogonal responses in chemical defences (Bidart-Bouzat & Imeh-Nathaniel, 2008; Zavala *et*
82 *al.*, 2013; Zvereva & Kozlov, 2006).

83

84 Biotic interactions have a strong impact on life histories and physiology of species, and these
85 interactions may shift responses of plants towards climate change (Van de Velde *et al.* 2016).
86 Competition for limited nutrients for example, enhances the availability of carbon in a focal
87 plant relative to its demand. This, in turn, can increase carbon-based defences and thus reduce
88 herbivory compared with plants that are less constrained by competitors (Bryant *et al.*, 1983).
89 Also the identity of a neighbouring plant may affect the resistance of a focal plant by a
90 different outcome of the plant-plant competition (Barton & Bowers, 2006; Broz *et al.*, 2010).
91 Conspecific competition, in particular, may result in stronger decline of plant growth and
92 increased defence compared to heterospecific competition. Indeed, Broz *et al.* (2010) for
93 instance showed that focal plants with conspecific neighbours allocated more resources
94 towards production of carbon-based defence molecules, whereas those grown with
95 heterospecific neighbours allocated more resources towards growth. Hence, elevated CO₂ as
96 well as warming is anticipated to indirectly affect the defence of a focal plant, through species
97 specific stimulation of plant growth and thus plant-plant competition.

98 Although atmospheric CO₂ and temperature increase concurrently, empirical studies
99 investigating their combined effect on multi-trophic communities are surprisingly scarce and
100 insufficient to effectively guide theory or synthesis (Cornelissen, 2011; Dyer et al. 2013;
101 Zvereva & Kozlov, 2006). The current study therefore investigates effects of elevated CO₂ as
102 well as combined effects of elevated CO₂ and warming on a simple model community
103 consisting of three species: rosy apple aphid *Dysaphis plantaginea* Passerini (*Hemiptera*:
104 *Aphididae*) feeding on plantain, *Plantago lanceolata* L., and a heterospecific neighboring
105 plant species, perennial ryegrass, *Lolium perenne* L. The aphid does not feed on *L. perenne*.
106 The experimental design consisted of monocultures of *P. lanceolata* and mixtures of *L.*
107 *perenne* and *P. lanceolata* exposed to the two aforementioned simulated climate scenarios as
108 well as to a control scenario with the current climate. As the experiments were conducted
109 outdoors, we were also able to *post-hoc* study how the imposed climatic and competition
110 treatments affected mildew infections. Biotrophic parasites such as mildew extract nutrients
111 from living cells and have extended periods of physiological interaction with their hosts
112 (Agrios, 2005). Elevated CO₂ and warming have been shown to affect the severity of
113 pathogen infections (Mikkelsen *et al.*, 2015, Thomas & Blanford, 2003). We took advantage
114 of this unplanned infestation to study the impact of elevated CO₂ and warming on mildew
115 infestation and the putative additive effects of mildew infestation on plant-insect herbivore
116 interaction.

117

118 Based on the above, we anticipate that elevated CO₂ and warming will alter foliar nutrients
119 and defence molecules, that altered host quality and plant resistance will affect insect
120 herbivore performance, and that elevated CO₂ and warming should indirectly influence host
121 quality and plant resistance via effects on neighbouring plants. We predict investments in
122 defence to be higher under conspecific competition relative to heterospecific competition

123 because of increased resource competition. In addition, increased CO₂ is anticipated to
124 increase chemical defence by promoting the production of secondary metabolites, while
125 simultaneously increased temperature would in contrast promote plant growth. Depending on
126 the relative rates of these physiological changes, net impact on herbivore populations may
127 either remain constant, decline or increase if defence is respectively proportionally promoted
128 or constrained relative to metabolism.

129

130 We do, however, aim to move beyond this generic perspective and to acquire a deep
131 mechanistic understanding of the changing ecological interactions under elevated CO₂ and
132 warming. To this end, we selected metabolites and groups of metabolites as molecular
133 markers to evaluate defences and stress levels (see Table 1). In particular, we quantified an
134 indicator of oxidative cellular damage (lipid peroxidation). Oxidative stress in plants is
135 commonly induced under abiotic stress, but also by herbivory (Wu & Baldwin, 2010).
136 Elevated CO₂ on the other hand, often reduces abiotic stress impact (Zinta et al. 2018).
137 Herbivory damage also induces the production of secondary metabolites associated with the
138 salicylic acid and jasmonic acid pathway (Kuśnierczyk *et al.*, 2011; Morkunas & Gabryś,
139 2011). Therefore, as an indicator of plant-insect interaction, jasmonic acid levels were
140 determined. Simultaneously with the stress status, we evaluated the plant capacity to defend
141 itself by quantifying protective metabolites. Here we determined molecular antioxidants
142 (tocopherols, antioxidant capacity, carotenoids, proline) providing protection against
143 oxidative stress (Gómez-Ariza *et al.*, 2007, Peshev *et al.*, 2013). We also quantified several
144 groups of molecules that contribute to defence against insects (glycosides, tannins, lignin).
145 Finally, elevated CO₂ as well as elevated temperature are known to affect stomatal aperture
146 and therefore water transport. To evaluate the plant response to its water status we quantified
147 osmolytes (proline, soluble sugars). Increase in these molecules reduces the cellular water

148 potential, improving water uptake. Given the anticipated complex interplay between elevated
 149 CO₂, temperature and competition, we performed a multivariate grouping analysis to
 150 document concerted responses in the quantified metabolites.

151

152 Table 1: Overview of molecular markers determined in this study and their principal function.

METABOLITE	PRINCIPAL FUNCTION	FUNCTION DESCRIPTION	REF.
TOCOPHEROL (VITAMINE E)	Lipophilic antioxidant	Components of biological membranes; act as scavengers of oxygen radicals generated by stress, in particular singlet oxygen. Tocopherols also stabilize membrane structures.	Blokhina et al (2003)
CAROTENOIDS	Lipophilic antioxidant	Lipophilic antioxidants localized in plastids of photosynthetic and non-photosynthetic plant tissues. Carotenoids react with lipid peroxidation products to end the chain reactions and dissipate the excess excitation energy.	Bartley and Scolnik (1995)
PHENOLICS/ FLAVONOIDS	Antioxidant	Polyphenols have free radical scavenging activity and are involved in hydrogen peroxide neutralization. Flavonoids are also secondary ROS scavenging molecules in plants under excess excitation.	Blokhina et al (2003)
TOTAL ANTIOXIDANT CAPACITY (TAC)		Total antioxidant capacity (TAC) is the measure of the gross capacity of a plant extract to scavenge reactive oxygen species.	Das and Roychoudhury (2014)
MALONDIALDEHYDE, (MDA)	Lipid peroxidation marker	Oxidative stress induces lipid peroxidation disrupting membrane functions. Lipid peroxidation products (MDA) indicate membrane oxidation levels.	Das and Roychoudhury (2014)
SOLUBLE SUGARS (GLUCOSE, SUCROSE, FRUCTOSE)	Osmolytes	Soluble sugars protect plant cells through osmoregulation, improving water uptake and retention, and may impact cellular membranes through interaction with the lipid bilayer.	Shao et al., (2006)
PROLINE	Osmolytes/ antioxidant	Proline is an osmolyte, contributing to water uptake, as well as an antioxidant.	Szabados and Savoure (2010).
GLYCOSIDES: CATAPOL, AUCUBIN	Biotic stress defence	Secondary metabolites that help repelling herbivores, protects plant from UV and contributes to control of competitive interactions between plants.	Bowers (1991).
TANNINS	Biotic stress defence	Tannins are astringent bitter polyphenols and act as feeding deterrents to many insect pests.	Barbehenn et al., (2011)
LIGNIN	Biotic stress defence	A complex phenolic polymer that plant cell wall rigidity and hydrophobic properties, and is an important barrier against pests and pathogens.	Liu et al., (2018)
SALICYLIC ACID	Biotic stress	Salicylic acid is a phytohormone involved in regulation of	War et al.,

defence signal plant defence, and resistance to insect pests. (2012)

153

154 **Materials and methods**

155 *Experimental set-up (Fig S1.1 in supplementary material)*

156 The study was performed at the Drie Eiken Campus, University of Antwerp, Wilrijk, Belgium
157 (51° 09' N, 04° 24'E) in 12 sunlit, south-facing, climate-controlled chambers (details in
158 Naudts *et al.* 2014). Three climate scenarios (four chambers per scenario) were simulated in
159 an additive design: (1) current atmospheric CO₂ concentration and temperature (C); (2) future
160 atmospheric CO₂ and current temperature (CO₂); and (3) future atmospheric CO₂ and
161 temperature (TCO₂). Climate scenarios with elevated CO₂ had a target CO₂ concentration of
162 620 μmol mol⁻¹ and future temperature chambers simulated a continuous 3 °C warming above
163 fluctuating ambient temperatures. Climate manipulations were based on the IPCC-SRES B2-
164 scenario prediction of moderate change for the year 2100 (IPCC, 2001).

165

166 The CO₂ concentration in each chamber was continuously measured and maintained at the
167 target concentration with a CO₂ control group with an infrared analyser (WMA-4, PPSystems,
168 Hitchin, UK). Air temperature and relative humidity were monitored every 0.5 h with a
169 combined humidity–temperature sensor (Siemens QFA66, Erlangen, Germany), by averaging
170 instantaneous readings in half-hour mean values. During the experiment, the CO₂
171 concentration was 382 ± 55 μmol mol⁻¹ (SD) in the current climate, while it was 615 ± 70
172 μmol mol⁻¹ (SD) in the climate scenarios with future CO₂ concentration (CO₂ and TCO₂). The
173 monthly average air temperature in the C and CO₂ chambers was 16.2, 17.2 and 18.7 °C in
174 June, July and August, respectively. TCO₂ chambers were 2.9 ± 1.0 °C (SD) warmer than
175 current temperature chambers. Average vapour pressure deficit was 0.60 ± 0.34 and 0.64 ±
176 0.52 kPa (SD) in the climate treatments with ambient and warmed air, respectively. Irrigation

177 mimicked the rainfall outside (Naudts et al. 2014), with monthly totals equalling 64.4, 85.1
178 and 80.2 mm in June, July and August, respectively. Water freely drained while capillary rise
179 was prevented by a drainage system placed below the chambers. Future climate chambers
180 (TCO₂) received the same amount of water as current climate chambers, so that any increase
181 in water consumption would result in (aggravated) soil drought.

182

183 *Plant and insect communities*

184 We used two common co-occurring grassland species, *L. perenne* and *P. lanceolata*. *P.*
185 *lanceolata* is characterized by the presence of the iridoid glycosides aucubin and catalpol.
186 These compounds stimulate feeding and oviposition by specialist insects and can act as
187 defence against generalist herbivores (Bowers & Puttick, 1988; Puttick & Bowers, 1988).
188 Both plant species were sown at the end of March in a non-climate controlled greenhouse
189 with a time lag of one week to prevent size differences at the start of the experiment (Cotrufo
190 & Gorissen, 1997), and were watered twice a week. Four or five-week-old seedlings were
191 transplanted into PVC containers (24 cm inner diameter and 40 cm height), filled with sandy
192 soil (93.9% sand, 4.1% silt, 2.0% clay; pH 7.5; Kjeldahl-N 0.125 g kg⁻¹; 2.1% C in humus).
193 Each of the 12 chambers received 20 containers with two different compositions: (1) 10
194 monocultures of *P. lanceolata*, and (2) 10 mixtures of both plant species in a 50:50 ratio.
195 Each community contained 18 individuals planted in a hexagonal grid with 4.5 cm interspace.
196 Interspecific interactions were maximized by avoiding clumping. All communities were
197 fertilized with 10 g m⁻² NH₄NO₃, 5 g m⁻² P₂O₅, 10 g m⁻² K₂O and micro-elements (Fe, Mn,
198 Zn, Cu, B, Mo), given dissolved in water in two equal amounts.

199 When the plants were three months old, *P. lanceolata* was involuntarily infested with
200 powdery mildew *Podosphaera plantaginis*. *P. plantaginis* is a biotrophic fungal pathogen
201 which means that it feeds on living plant tissue, but does not kill the infected host. The

202 powdery mildew was found in all climate scenarios and plant compositions but not all the
203 containers were equally affected. We took advantage of this unplanned infestation to study the
204 impact of elevated CO₂ and warming on mildew infestation and the putative additive effects
205 of mildew infestation on plant-insect herbivore interaction. Because these infections occurred
206 post-hoc, we do not have proper controls to quantify mildew effects on metabolite
207 composition.

208

209 The rosy apple aphid *D. plantaginea* was used as an insect herbivore. It overwinters as eggs
210 on apple trees, the primary host plant, and migrates in spring to the obligate alternate hosts, *P.*
211 *major* L. and *P. lanceolata* (Alford, 2014). On *Plantago* spp., they give birth to apterous
212 (wingless) morphs that reproduce by parthenogenesis (Blommers *et al.*, 2004). *L. perenne* is
213 not a host plant for *D. plantaginea*. The aphids were reared in small cages on *P. lanceolata*
214 under laboratory conditions of 22 ± 1 °C. They were introduced on *P. lanceolata* when the
215 plants were 20 weeks old. At this time, two adult, apterous aphids were placed with a dry
216 paintbrush on the apex of each *P. lanceolata* plant in monocultures and mixtures.
217 Consequently, at the start of the infestation each container was inoculated with 36
218 (monocultures) or 18 (mixtures) aphids. In each chamber, four monocultures of *P. lanceolata*
219 and four mixtures were randomly chosen for aphid infestation All containers were meshed,
220 and those not receiving aphids acted as controls. The meshes consisted of an 85-cm-tall
221 cylinder of lightweight netting to ensure aphids did not migrate between pots. The
222 infrastructure did not physically limit plant growth and did not cause photosynthetic stress
223 effects (F_v/F_m , the intrinsic efficiency of PSII in controls equalled 0.84 which is an optimal
224 value (Johnson *et al.*, 1993).

225

226 *Data collection*

227 In the fourth week after the aphid introduction we determined the mildew infestation. The
228 degree of powdery mildew on *P. lanceolata* was categorized by a rating system: 1) healthy
229 (no visible lesions); 2) 1% - 25% of the leaves damaged; 3) 26% - 50% of the leaves
230 damaged; 4) 51% - 75% of the leaves damaged; 5) more than 75% of the leaves damaged. At
231 the same time, aphid populations of four replicate communities per plant composition in each
232 chamber were collected (totalling 4 replicate communities \times 2 plant compositions \times 12
233 chambers). Aphids were brushed directly into 70% ethanol with a dry paintbrush. It was not
234 possible to collect all the aphids because their number per container was either too high or too
235 low (hard to find), therefore a subsample of the population per container was collected: we
236 searched for and collected aphids for 30 minutes. The total number of aphids per community
237 was divided by the number of *P. lanceolata* individuals in that community. At the same time,
238 we also harvested the plants of two replicate communities per treatment in each chamber
239 (totalling 2 replicate communities \times 4 treatments \times 12 chambers). Alive aboveground plant
240 biomass in each community was separated from dead biomass by species, dried at 70 °C for
241 48 h and weighed. These biomass values per species were likewise divided by the number of
242 the species' individuals in the community, providing primary data for the statistical analysis.
243 The alive biomass samples of *P. lanceolata* plants of the two harvested communities per
244 treatment in each chamber (2 replicate communities \times 4 treatments \times 12 chambers) were
245 ground in a mill. Three subsamples per community were analysed for nitrogen and carbon
246 content using a NC element analyser (NC-2100 element analyser, Carlo Erba Instruments,
247 Milano, Italy), which were averaged prior to data analysis.

248 A separate subsample of the milled live aboveground biomass of *P. lanceolata* of the two
249 harvested communities per treatment in each chamber (2 replicate communities \times 4 treatments
250 \times 12 chambers) was taken to quantify biochemical parameters. These parameters are
251 indicators of stress in the organism (membrane damage as malondialdehyde levels, MDA),

252 indicators of antioxidant defences i.e. total antioxidant capacity (TAC) and antioxidant
253 molecules (tocopherols, carotenes, proline, phenols); indicators of biotic stress and defence,
254 i.e. glycosides (catapol, aucubin), tannin, lignin and salicylic acid; and metabolites involved in
255 plant water-deficit defence (proline, soluble sugars). We refer to Supplementary material 2 for
256 a detailed description of the methods used to quantify these metabolites.

257

258 *Data analysis*

259 We focused on *P. lanceolata* because this is the host plant for the aphid and powdery mildew.
260 In a first step we examined covariation in the metabolites responses to the three treatments
261 (aphid infestation, climate scenario and plant composition) by using a hierarchical clustering
262 analysis. Metabolites were normalized by Z-transformation and subjected to a hierarchical
263 clustering analysis with an Euclidean distance metric, and visualized as a heat map using
264 Multi Experiment Viewer (Mev) version 4.8 (Saeed *et al.*, 2003). A distance cut off of 0.3
265 was applied. The clusters were used in the structural equation models (see below).

266 In a second step, we fitted two piecewise Structural Equation Models (SEM), which combine
267 information from multiple separate linear models into a single causal network (Shipley,
268 2009). The first SEM investigated the effect of aphid infestation, climate scenario, plant
269 composition and mildew infestation on the metabolites and alive aboveground biomass of *P.*
270 *lanceolata*. In a second SEM, we separated the metabolites of *P. lanceolata* in control pots
271 from those with aphids. This allowed us to test the effects of the metabolites on the aphid
272 population, as well as to quantify effects of the aphid population on these metabolites. The
273 response of the aphid population was measured as their number at the end of the infestation.
274 The four clusters obtained by the clustering analysis were used to divide the metabolites in
275 separate groups and hence to avoid redundancy in the analyses. We standardized the
276 metabolites, the alive aboveground biomass of *P. lanceolata* and the number of aphids by

277 converting to Z-scores to equalize variances. To reduce the number of mildew categories, we
278 rearranged degree of mildew in two categories: (1) no or mild mildew infection (category 1 -
279 2) (2) severe mildew infection (category 3 - 5).

280 Traditional SEM estimation methods assume that all variables follow a normal distribution
281 and all observations are independent (Grace, 2006). In our analyses, we used piecewise SEM
282 that allows fitting general linear mixed effect models that can incorporate random effects.
283 Each mixed effects model was fitted using the “lme” function in the “nlme” package (version
284 3.1-128) in R. For each model, we fitted a random effect of chamber. The overall path model
285 (the SEM) was fitted using the “piecewiseSEM” package (version 1.2.1) in R (Lefcheck,
286 2016). Goodness of fit was estimated using Shipley’s test of d-separation, which yields a
287 Fisher’s C statistic that is Chi-squared distributed (Shipley, 2009). If the resulting P-value >
288 0.05, then the SEM can be said to adequately reproduce the hypothesized causal network.

289 As a final confirmatory step, all data were analysed with General Linear Mixed models
290 (GLM) in SAS (version 9.2, SAS Institute Inc., Cary, NC) (Littell *et al.*, 1996) with chamber
291 as a random factor nested within climate scenario. Climate scenario, plant composition, aphid
292 infestation and two-way and three-way interactions between these predictors were included as
293 fixed factors. Because mildew infestation did not have a significant effect on biochemical
294 plant responses, this treatment was excluded from the GLM analysis. Non-significant factors
295 were backwards-excluded from the model. In case of significant effects, *a posteriori* means
296 comparisons using the Tukey test, corrected for multiple comparisons, were made. Effects
297 were considered significant at $P \leq 0.05$. Repeated univariate analyses are subject to error
298 proliferations. We deliberately decided not to correct results from these analyses for multiple
299 testing, but instead interpret them in a conservative way in congruence with the higher
300 described grouping analyses. Results and discussion of the series of univariate analyses are
301 therefore provided in Supplementary Material 3.

302

303

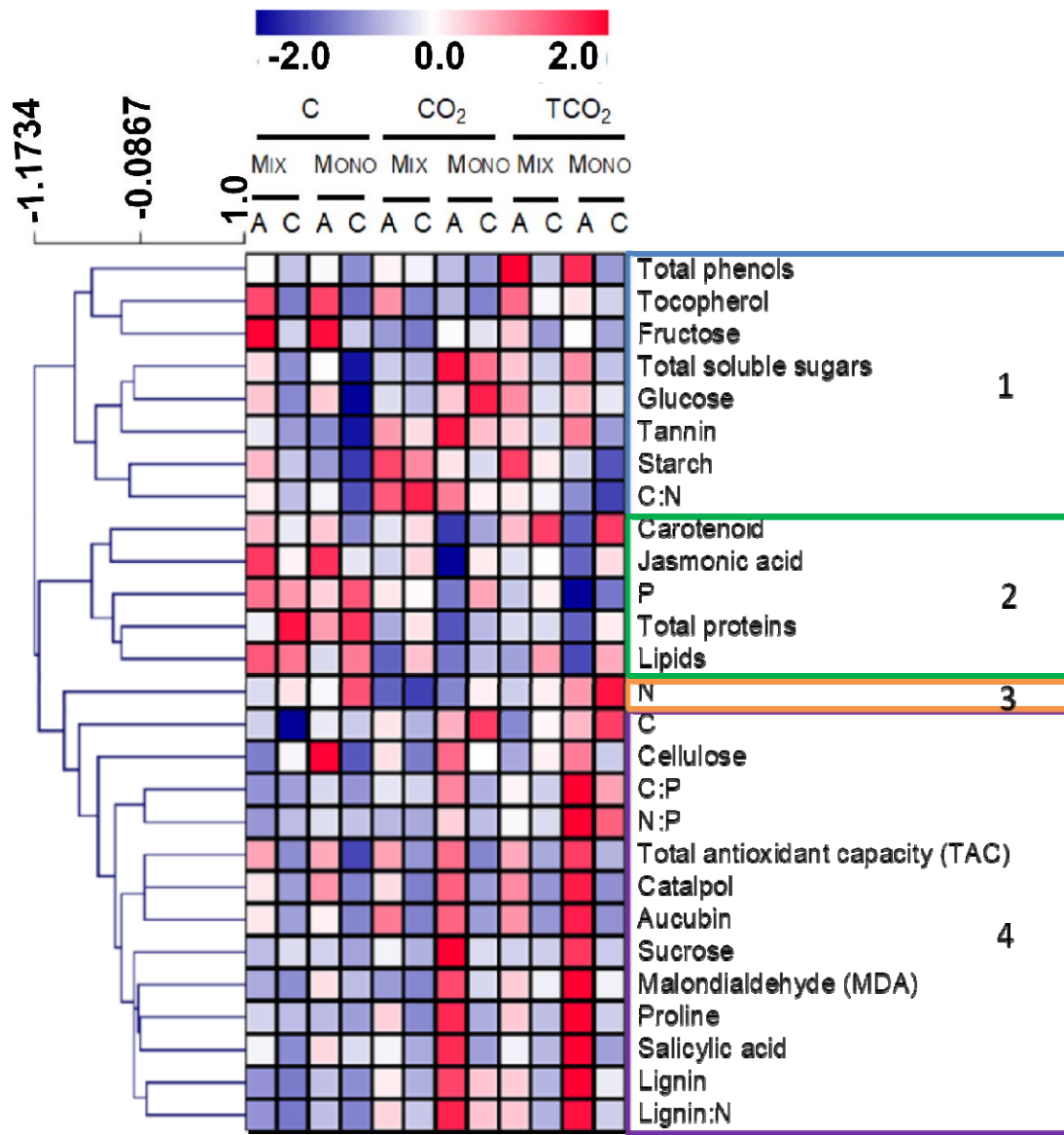
304 **Results**

305 *Hierarchical clustering analysis of metabolites*

306 The variation in primary and secondary metabolites of *P. lanceolata* was subjected to a
307 hierarchical clustering analysis in order to understand their grouping according to their
308 covariation in response to the imposed herbivory, climate and competition treatments. We
309 obtained four major clusters of metabolites (clusters 1 - 4 in Fig. 1).

310 The first cluster containing metabolites consistently upregulated by the presence of aphids
311 contains the phenol antioxidants, the lipophilic antioxidant tocopherol, the C:N ratio, tannins,
312 and the non-structural carbohydrates soluble sugars (including glucose, fructose) and starch
313 (Fig. 1). The second cluster contains another group of a.o., antioxidant and defence molecules
314 that are downregulated by elevated CO₂ and more so by aphids in mono-cultures, i.e. the
315 carotenoids, as well as the growth regulator, jasmonic acid, total proteins, phosphorous and
316 lipids (Fig. 1).

317 The nutrient N which contributes solely to the third cluster, is downregulated under elevated
318 CO₂. Carbon concentration, the C:P and N:P ratio, and lignin:N were classified in the fourth
319 cluster that is specifically upregulated by aphids in monocultures under elevated CO₂ (Fig. 1).
320 Cluster 4 also contains metabolites that play a role in plant defences against herbivores, such
321 as salicylic acid and the iridoid glycosides (catalpol and aucubin), and also contains TAC,
322 lignin, cellulose, proline and MDA. Notably in this cluster, metabolites are strongly induced
323 in CO₂ and TCO₂.



324

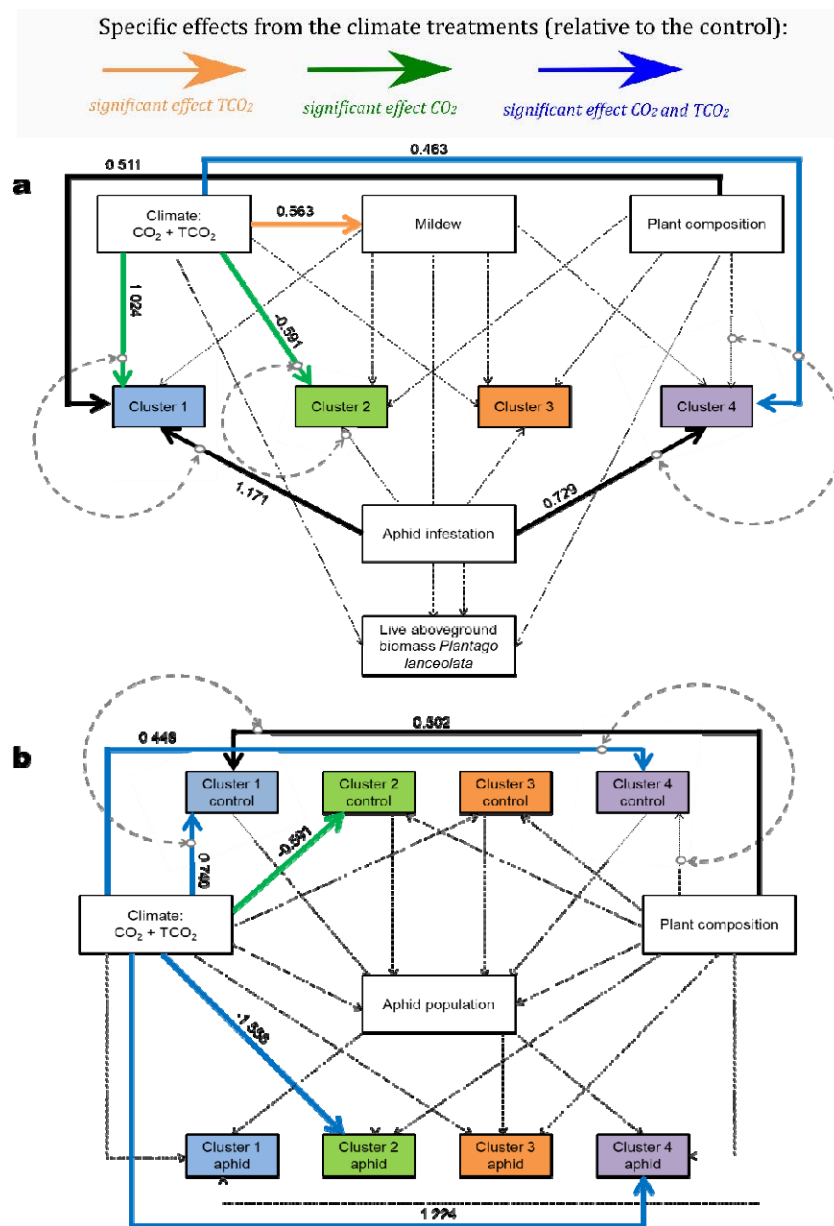
325 **Fig. 1** Heat map showing the metabolite levels in the leaves of *P. lanceolata*, normalized to Z-
 326 score for each metabolite (blue-white-red heat map). Blue and red colours indicate a low and
 327 high metabolite level, respectively. Clustering was based on the Euclidean distance for
 328 metabolites. Labels 1-4 and colours indicate the four prominent clusters. Labels C, CO₂ and
 329 TCO₂ indicate current climate, elevated CO₂ and combined warming and elevated CO₂,
 330 respectively. Plant communities consist of monocultures of *P. lanceolata* (mono) and mixtures
 331 of *Lolium perenne* and *P. lanceolata* (mix) with (A) and without aphids (C).

332

333 *Causal changes as inferred by structural equation models and univariate analyses*

334 SEMs quantify the strength of both the direct and indirect interactions between the
335 environmental scenarios related to climate change, herbivory and plant phenotype. The first
336 SEM presents the effect of aphid infestation (qualitative effect), climate scenario, plant
337 composition and mildew infestation on the chemical composition (the metabolites of *P.*
338 *lanceolata* were subdivided into the four groups obtained from the hierarchical clustering; see
339 Fig. 1) and the live aboveground biomass of *P. lanceolata* (Fig. 2A). The hypothesized
340 structural relationship with climate scenario affecting mildew infection severity, and climate
341 scenario, plant composition and aphid infestation interactively affecting metabolites and
342 biomass adequately fits the data ($\chi^2 = 42.53$, $df = 32$, $p = 0.101$). Fig. 2A and SEM statistics in
343 Table S2.1 (in supplementary material 2) provide us with three insights. Firstly, TCO₂
344 increased the mildew infestation compared to C, but the mildew infestation, in turn, did not
345 affect the metabolites and the alive aboveground biomass of *P. lanceolata*. Secondly, neither
346 climate scenario, nor aphid infestation or plant composition had a significant impact on the
347 live aboveground biomass of *P. lanceolata*. Univariate analyses showed that live biomass
348 tended to be higher in CO₂ than C and was significantly lower in TCO₂ than CO₂, which
349 combined lead to similar values in TCO₂ and C (Fig. 3; see also supplementary material 3.1).

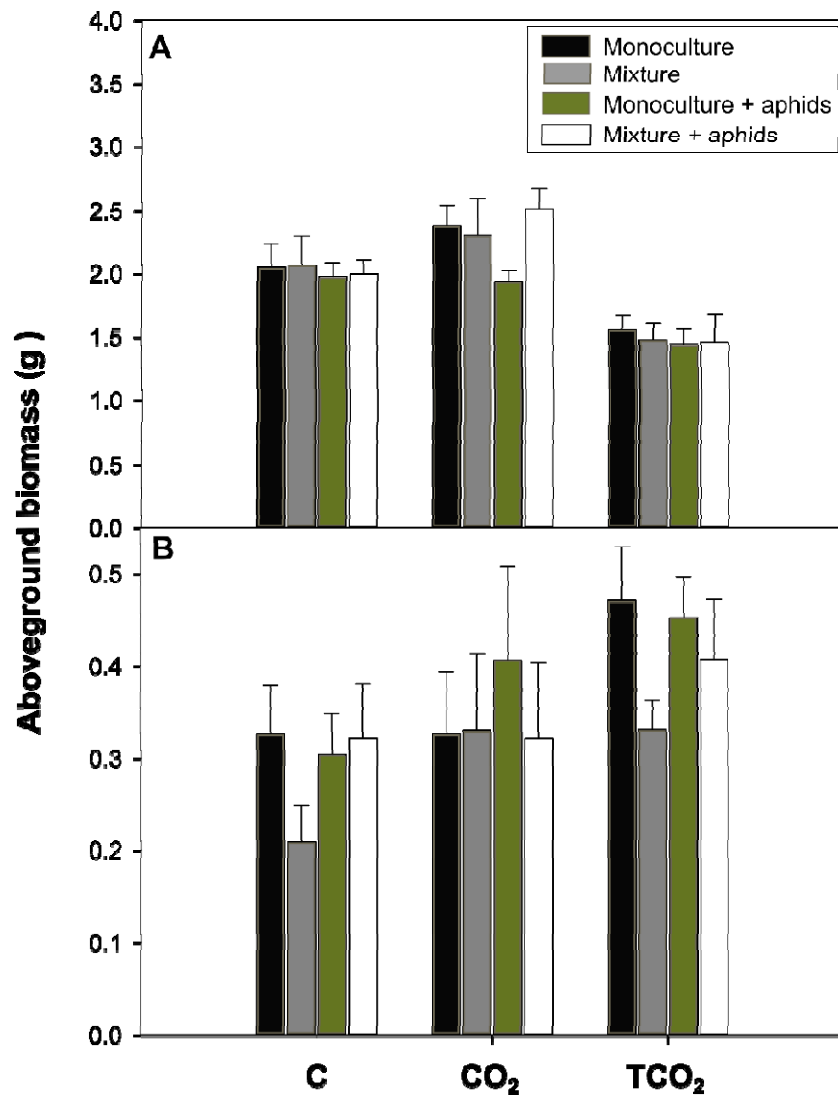
350



351

352 Fig. 2 (a) Structural equation model showing how climate scenario (CO_2 and TCO_2), mildew infestation, plant
 353 composition and aphid infestation affect the chemical composition and the live aboveground biomass of *P.*
 354 *lanceolata*. (b) Structural equation model showing how climate scenario, plant composition and chemical
 355 composition of *P. lanceolata* affect aphid population and how aphid population, in turn, affects the
 356 chemical composition of *P. lanceolata*. The four clusters refer to those obtained by the hierarchical clustering analysis
 357 (see Fig. 1). Solid black, green and orange arrows represent significant relationships ($P \leq 0.05$) and dashed
 358 light grey lines significant interactions. Blue lines stand for significant effects of both CO_2 and TCO_2 , green lines for
 359 significant effects of CO_2 and orange lines for significant effects of TCO_2 . Light grey arrows represent non-
 360 significant relationships. Standardized path coefficients are shown next to pathways. For the effect of CO_2 and
 361 TCO_2 , the average path coefficients are shown. The individual path coefficients of CO_2 and TCO_2 can be seen in
 362 Table S2.1 and Table S2.2 (see supplementary material section 2). Metabolites levels, live aboveground
 363 biomass of *P. lanceolata* and number of aphids were scaled before analysis.

364



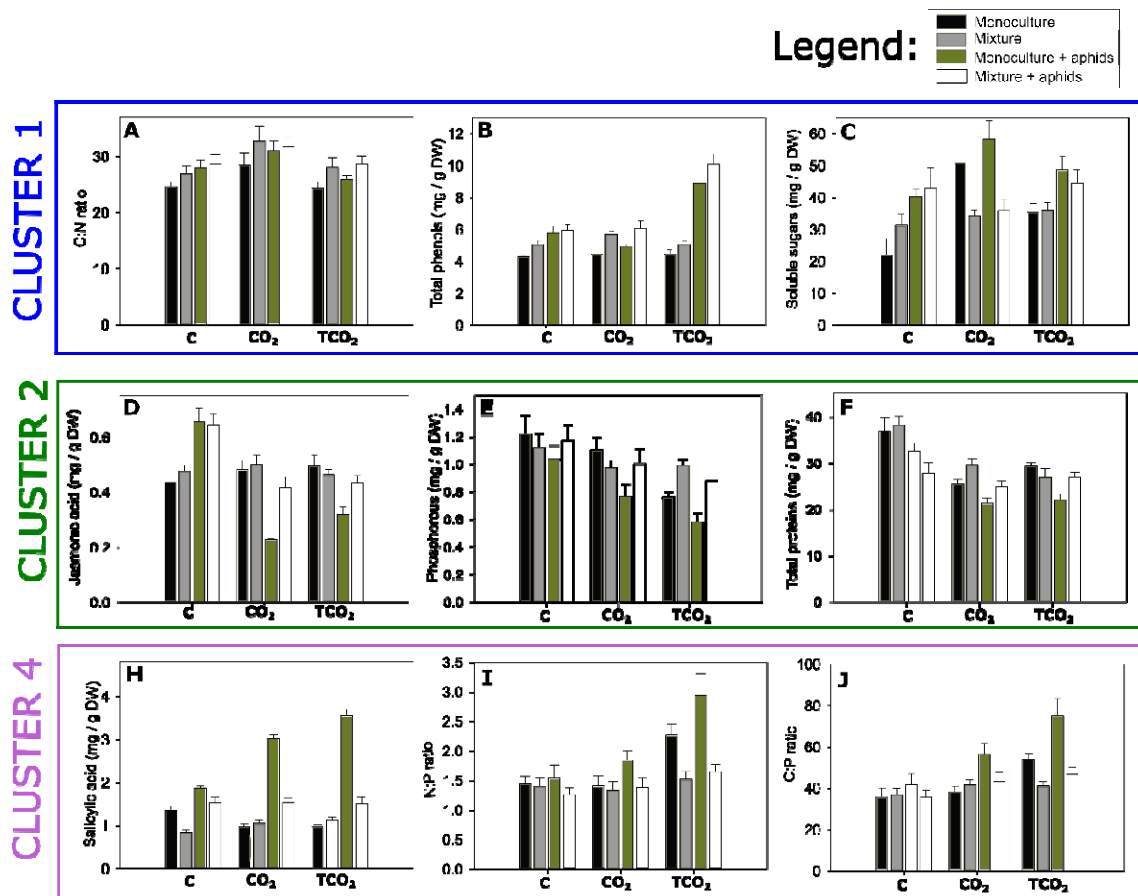
365

366 **Fig. 3** Effect of climate scenario (C, CO₂ and TCO₂), aphid infestation and plant composition on the live (A)
367 and dead (B) aboveground biomass of individual *P. lanceolata* plants. Bars represent means \pm SE. Plant
368 communities consist of monocultures of *P. lanceolata* and mixtures of *Lolium perenne* and *P. lanceolata*.

369

370 Thirdly, climate scenario, aphid infestation and plant composition impacted the distinguished
371 clusters in a different manner. We noticed a significant interaction between climate scenario
372 and aphid infestation for metabolites associated with cluster 1 (see supplementary material
373 3.2; Table S3.1 and Fig. S3.1). Aphid infestation increased the concentrations of the
374 metabolites on average and pending on the environmental change scenario between 30 and
375 100% in cluster 1 and plant composition (except for C:N ratio; Fig. 4A-B). The

376 concentrations of glucose and soluble sugars increased at elevated CO₂ but only in
 377 monocultures, while TCO₂ did not alter the concentrations of glucose relative to the CO₂
 378 treatment but decreased the concentrations of soluble sugars in monocultures (Fig. 4C). In
 379 general CO₂, compared to C, increased the concentration of tannin while TCO₂ did not impose
 380 further differences. Elevated temperature and CO₂ reduced the concentration of starch (except
 381 for mixtures with aphids) and the C:N ratio compared to CO₂.



382

383 **Fig. 4** Effect of climate scenario (C, CO₂ and TCO₂), aphid infestation and plant composition on a selection of
 384 relevant metabolites from the four distinguished clusters. Bars represent means \pm SE. Plant communities
 385 consist of monocultures of *P. lanceolata* and mixtures of *Lolium perenne* and *P. lanceolata*. As cluster 3 only
 386 contained one metabolite, it is not depicted here.

387

388 Also the metabolites in cluster 2 (see supplementary material 3.2 – Table S3.2, Fig. S3.2)

389 differed according to climate scenario and aphid infestation. More specifically, univariate

390 analyses of the metabolites indicates that the concentration of jasmonic acid in C increased
391 with aphid infestation in monocultures and mixtures while aphid infestation in CO₂ and TCO₂
392 decreased it in monocultures (Fig. 4D). Interspecific interactions between *P. lanceolata* and *L.*
393 *perenne* mitigated the negative effect of aphid infestation in these climate scenarios, and
394 hence have a neutralizing effect. Aphid infestation reduced the concentration of P in
395 monocultures, but did not alter it in mixtures (Fig. 4E). Furthermore, also CO₂, compared to
396 C, reduced the concentration of P and the concentration was further reduced in TCO₂.
397 Furthermore, aphid infestation decreased the concentration of proteins in mixtures in C but
398 did not alter the concentrations in monocultures and mixtures in CO₂ and TCO₂ (Fig. 4F). In
399 general, CO₂ compared to C decreased the concentration of proteins irrespective of the plant
400 composition while aphid infestation and TCO₂, compared to CO₂, did not alter it. The
401 concentration of lipids decreased with aphid infestation in monocultures and mixtures in all
402 climate scenarios (except for mixtures in C). Also, CO₂, compared to C, decreased the
403 concentration of lipids in monocultures without aphids and mixtures with aphids, but in
404 TCO₂, compared to CO₂, this concentration increased again to similar levels as C in
405 monocultures without aphids. Carotenoid concentration increased in TCO₂, compared to C
406 and CO₂. Leaf nitrogen was not impacted by any of the treatments (cluster 3; see
407 supplementary material 3.2; Table S3.3).

408 Aphid infestation, CO₂ and TCO₂ increased the metabolites in cluster 4 (see supplementary
409 material 3.2; Table S3.4, Fig. S3.3 & Fig. S3.4) and specifically inhibited the expression of
410 salicylic acid under the two environmental change scenarios (Fig 4G). Besides these single
411 factor treatments effects, climate scenario, aphid infestation and plant composition also
412 interacted with each other in affecting the metabolites in cluster 4. CO₂ and TCO₂
413 strengthened (salicylic acid and lignin) or induced (proline) an effect of aphid infestation but
414 only in monocultures (Fig. 4G). TCO₂, compared to CO₂ and C increased the N:P and the C:P

415 ratio in monocultures (Fig. 4H,I). Cellulose levels were always higher in monocultures and
416 aphid infestation but the magnitude of increase was lower under the two climate-related
417 environmental change scenarios relative to the current conditions (control).

418

419 The second SEM presents the effect of climate scenario, plant composition and chemical
420 composition of *P. lanceolata* on the aphid population and whether the aphid population, in
421 turn, altered the chemical composition of *P. lanceolata* (quantitative effect). The mildew
422 infestation was left out of this analysis since it is a parasite of the plants, not of the aphids.
423 The results of this SEM model show the following goodness of fit statistics: $\chi^2 = 77.52$, $df =$
424 80 and $p = 0.558$ (Table S2.2 and Fig. 2b), which provides us with two insights. Firstly,
425 climate scenario, plant composition and the chemical composition of *P. lanceolata* did not
426 affect the number of aphids. Secondly, the aphid population size in turn, did not change the
427 chemical composition of *P. lanceolata*. Bringing together the findings from both SEM
428 models, we conclude that the treatment aphid infestation altered the chemical composition of
429 *P. lanceolata* but the number of aphids had no effect on it.

430 **Discussion**

431 A future climate clearly altered leaf nutritional quality and the expression of defence
432 molecules, while aphid infestation overall impaired nutritional requirements for insect
433 herbivores. Despite the strong metabolic changes as induced by climate-related environmental
434 change, no feedbacks on aphid population performance and herbivory were detected. More
435 importantly, we show that interspecific plant competition neutralizes the positive effects of
436 elevated CO₂ on the production of defence molecules in *P. lanceolata*. Induced effects of
437 climate-related environmental change on plant performance can thus be altered and even
438 nullified by competitive and food web interactions. We first discuss the general herbivore-
439 induced responses followed by an in-depth discussion of their dependency on the imposed
440 climate and competition treatments.

441

442 ***Herbivore-induced responses***

443 Our experiment, as conducted under unique semi-natural conditions confirmed in first
444 instance findings that aphid infestation reduces N, P, total proteins and lipids in *P. lanceolata*
445 leaves, thereby impairing nutritional requirements of insect herbivores (Schoonhoven *et al.*,
446 2005). In contrast to previous research (Walling, 2000), however, aphid infestation promoted
447 the salicylic acid and jasmonic acid pathway (Kuśnierczyk *et al.*, 2011; Morkunas & Gabryś,
448 2011) as parallelized by the increased production of catalpol, aucubin, tannin, lignin and
449 cellulose.

450

451 Aphid herbivory increased oxidative stress in plants as well (Wu & Baldwin, 2010). We
452 assessed the induction of oxidative stress by measuring MDA, which aphid herbivory
453 enhanced considerably. A common defence response of plants against oxidative stress

454 includes the increase of antioxidant metabolites. Indeed, TAC and the well-known antioxidant
455 molecules phenols and tocopherols increased due to aphid herbivory. Moreover, *P. lanceolata*
456 responded to herbivory by increasing soluble sugars. At higher concentrations, also sugars can
457 act as antioxidants and may play a signalling role in regulating stress and defence responses
458 (Gómez-Ariza *et al.*, 2007, Peshev *et al.*, 2013). The induced resistance against oxidative
459 stress associated with aphid feeding appeared sufficiently effective to constrain biomass loss
460 from herbivory.

461

462 *Climate dependency of the plant phenotype*

463 Elevated CO₂ and temperature did not change the impact of herbivory on *P. lanceolata*
464 biomass. This implies that warming and CO₂ did not affect the net interaction strength
465 between plants and herbivores under semi-natural conditions. In contrast to earlier studies
466 (Van De Velde *et al.*, 2016), our experiment allows a profound analysis of the underlying
467 biochemical processes.

468 First, we found elevated CO₂ to increase starch and soluble sugars (in monocultures) and to
469 lower leaf proteins, hereby increasing the C:N ratio. In addition, elevated CO₂ reduced P and
470 consequently increased the C:P ratio (in monocultures), thereby reducing the plant's nutritive
471 value to herbivores (Huberty & Denno, 2006; Lincoln *et al.*, 1986; Mattson, 1980;
472 Schoonhoven *et al.*, 2005). We demonstrate here that these effects can be countered when
473 elevated CO₂ is accompanied with a temperature rise. This contradicts findings based on a
474 limited set of primary metabolites (Murray *et al.* 2013), but urges to apply broader phenomic
475 approaches to fully understand plant responses to climate change.

476

477 Second, elevated CO₂ increased the defence molecules lignin and tannin, especially under
478 aphid infestation. The induction of plant chemical defences (Bidart-Bouzat & Imeh-Nathaniel,
479 2008; Bidart-Bouzat *et al.*, 2005) is thus not consistent for all defence molecules, and are
480 consequently anticipated to render prediction on future plant-insect herbivore interaction
481 highly complex (Agrawal, 1999). In our study, the higher C:N ratio reflected higher lignin
482 and tannin levels. While there is a consensus that the carbon-nutrient-balance hypothesis fails
483 as predictive framework (Hamilton *et al.*, 2001; Lindroth, 2012), alternative hypotheses have
484 been proposed stating that resource utilization for chemical defence is linked with
485 photosynthesis, hormone regulation and the control of gene expression (Zavala *et al.*, 2017).
486 We here provide evidence for this alternative mechanism as we found CO₂ to alter
487 phytohormones (i.e., salicylic acid and jasmonic acid) that play an important role in
488 promoting compounds responsible for herbivore defence (Wu & Baldwin, 2010). Since no
489 further quantitative changes were observed under higher temperature, we can confidently
490 conclude that climate-change-induced changes in metabolic profiles are more due to enhanced
491 CO₂ than to temperature increases acting in parallel.

492

493 ***Competition: a biotic interaction mediating climate-induced effects***

494 The presence and identity of neighbouring plants can influence the quality of the host plant by
495 altering primary and secondary chemistry (Barton & Bowers, 2006; Broz *et al.*, 2010, Lankau,
496 2012; Thorpe *et al.*, 2011). We provide evidence that these biotic interactions are additionally
497 able to alter climate-induced phenomic changes, and by that further impacting ecological
498 interactions under climate-related environmental change. Energetic growth-defence-trade-offs
499 have usually been attributed to these competition effects (Herms & Mattson, 1992), but recent
500 studies demonstrated the role of light limitation for plant defence downregulation (Ballaré,
501 2014; Campos *et al.*, 2016). More specifically, a reduced red to far-red ratio downregulates

502 defences by a simultaneous inhibition of the jasmonic acid and the salicylic acid pathway
503 (Wit *et al.*, 2013). The lower levels of defence molecules and salicylic acid in *P. lanceolata*
504 support this hypothesis. Competition did, however, also counter the CO₂-induced changes in
505 primary and secondary metabolites, as well as those induced by herbivores, rendering an
506 understanding of plant defences far from trivial (de Vries *et al.*, 2017). It is well-understood
507 that bottom-up effects from plant quality to herbivore abundance cannot be generalized across
508 the feeding guilds (Bezemer & Jones, 1998; Robinson *et al.*, 2012; Stiling & Cornelissen,
509 2007), but phloem feeders such as aphids were at least expected to be facilitated. Despite the
510 complex, high-dimensional changes in plant metabolomes under competition and climate
511 change, the overall effects on aphid population sizes were small to non-existent.

512

513 We here thus demonstrate that the joint action of atmospheric factors associated with climate
514 change and biotic interactions are able to induce pronounced changes in plant metabolomes
515 and biomass, but that translation of these biochemical changes towards ecological responses
516 appears non-trivial. Climate-change-imposed changes are shown to be completely neutralized
517 when common biotic interactions are taken into consideration. Plant-plant interactions thus
518 add a layer of complexity in mechanistic studies of climate change effects on plant-enemy
519 ecological interaction.

520 **Supplementary data:**

521 **Supplementary material 1:** Schematic representation of the experimental setup

522 **Supplementary material 2:** Metabolite quantification methods and SEM statistics

523 **Supplementary material 3:** Results from the univariate GLM analyses (statistics and
524 figures)

525

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533

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706 **Figure legends**

707

708 **Fig. 1** Heat map showing the metabolite levels in the leaves of *P. lanceolata*, normalized to Z-
709 score for each metabolite (blue-white-red heat map). Blue and red colours indicate a low and
710 high metabolite level, respectively. Clustering was based on the Euclidean distance for
711 metabolites. Labels 1-4 and colours indicate the four prominent clusters. Labels C, CO₂ and
712 TCO₂ indicate current climate, elevated CO₂ and combined warming and elevated CO₂,
713 respectively. Plant communities consist of monocultures of *P. lanceolata* (mono) and
714 mixtures of *Lolium perenne* and *P. lanceolata* (mix) with (A) and without aphids (C).

715 **Fig. 2** (a) Structural equation model showing how climate scenario (CO₂ and TCO₂), mildew
716 infestation, plant composition and aphid infestation affect the chemical composition and the
717 live aboveground biomass of *P. lanceolata*. (b) Structural equation model showing how
718 climate scenario, plant composition and chemical composition of *P. lanceolata* affect aphid
719 population and how aphid population, in turn, affects the chemical composition of *P.*
720 *lanceolata*. The four clusters refer to those obtained by the hierarchical clustering analysis
721 (see Fig. 1).

722 Solid black, green and orange arrows represent significant relationships ($P \leq 0.05$) and dashed
723 grey lines significant interactions. Blue lines stand for significant effects of both CO₂ and
724 TCO₂, green lines for significant effects of CO₂ and orange lines for significant effects of
725 TCO₂. Light grey arrows represent non-significant relationships. Standardized path
726 coefficients are shown next to pathways. For the effect of CO₂ and TCO₂, the average path
727 coefficients are shown. The individual path coefficients of CO₂ and TCO₂ can be seen in
728 Table S1 and Table S2 (see supplementary material section 2). Metabolites levels, live
729 aboveground biomass of *P. lanceolata* and number of aphids were scaled before analysis.

730 **Fig. 3** Effect of climate scenario (C, CO₂ and TCO₂), aphid infestation and plant composition
731 on the live (A) and dead (B) aboveground biomass of *Plantago lanceolata*. Bars represent
732 means \pm SE. Plant communities consist of monocultures of *P. lanceolata* and mixtures of
733 *Lolium perenne* and *P. lanceolata*.

734 **Fig. 4** Effect of climate scenario (C, CO₂ and TCO₂), aphid infestation and plant composition
735 on the on a selection of relevant metabolites from the four distinguished clusters. Bars
736 represent means \pm SE. Plant communities consist of monocultures of *P. lanceolata* and
737 mixtures of *Lolium perenne* and *P. lanceolata*.