

1 **Noninvasive prenatal test of methylmalonic academia cblC type through targeted**
2 **sequencing of cell-free DNA in maternal plasma**

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24 **Abstract:**

25 Methylmalonic acidemia (MMA) cbIC type is the most frequent inborn error of
26 intracellular cobalamin metabolism which is caused by mutations of *MMACHC* gene.
27 Non-invasive test of MMA for pregnant women facilitates safe and timely prenatal
28 diagnosis of the disease. In our study, we aimed to design and validate a
29 haplotype-based noninvasive prenatal test (NIPT) method for cbIC type of MMA.
30 Targeted capture sequencing using customized hybridization was performed utilizing
31 gDNA (genomic DNA) of trios including parents and an affected proband to
32 determine parental haplotypes associated with the mutant and wild allele. The fetal
33 haplotype was inferred later based on the high depth sequencing data of maternal
34 plasma as well as haplotype linkage analysis. The fetal genotypes deduced by NIPT
35 were further validated by amniocentesis. Haplotype-based NIPT was successfully
36 performed in 21 families. The results of NIPT of 21 families were all consistent with
37 invasive prenatal diagnosis, which was interpreted in a blinded fashion. Three fetuses
38 were identified as compound heterozygosity of *MMACHC*, 9 fetuses were carriers of
39 *MMACHC* variant, and 9 fetuses were normal. These results indicated that the
40 haplotype-based NIPT for MMA through small target capture region sequencing is
41 technically accurate and feasible.

42

43 **Introduction**

44 Methylmalonic acidemia (MMA) is caused by a deficiency of methylmalonyl-coA
45 mutase or coenzyme adenosylcobalamin (AdoCbl). The cblC type combined with
46 methylmalonic acidemia and homocystinuria is a kind of autosomal recessive
47 hereditary disease and the incidence was found to be in 1 in 48,000 to 1 in 250,000
48 worldwide (Carrillo-Carrasco *et al.* 2012; Wang *et al.* 2010). Newborn screening for
49 MMA in Shandong province of China showed an estimated prevalence of 1 in 3920
50 during birth (Han *et al.* 2016). The MMA cblC type caused by mutations in the
51 *MMACHC* gene (NM_015506.2) located on chromosome 1p34.1 is the most frequent
52 congenital error in intracellular co-amine metabolism. Prenatal diagnosis of MMA
53 was essential due to age of onset in neonatal period and serious symptoms such as
54 multiple system damage and lethal possibility. It can not only contribute to early
55 medical management of infants, but also allows treatment of affected foetus as soon
56 as possible to reduce irreversible organ damage associated with metabolic acidosis
57 and high blood ammonia. (Huemer *et al.* 2005; Trefz *et al.* 2016).

58 Recently, several noninvasive prenatal test (NIPT) methods aimed to avoid
59 miscarriage or infection risk method have been developed (Evans *et al.* 2002;
60 Mujezinovic and Alfirevic 2007). Besides large-scale clinical application in fetal
61 aneuploidies screening, NIPT for dominant single-gene disorders confined to detect
62 paternally inherited and de novo mutations has also been introduced to clinical trials
63 (Lo *et al.* 1998; Fan *et al.* 2008). However, NIPT for recessive single gene disorders
64 are still at laboratory research stage. Our study group has developed a
65 haplotype-based method of NIPT for recessive inherited single gene diseases using
66 linkage analysis of trios' members and has been validated in several single gene
67 disorders (Xu *et al.* 2015; Meng *et al.* 2014; Lam *et al.* 2012; Ma *et al.* 2014; Ye *et al.*

68 2018; Chen *et al.* 2017). The feasibility and accuracy of this method depends on the
69 informative SNPs used to construct the haplotype and the selection of the targeted
70 region. NIPT of MMA based on haplotype analysis method has not been reported yet,
71 and so this study has been designed.

72 In our study, the NIPT of MMA can be accomplished using a 141.39 kb
73 customized probe, including *MMACHC* and 1125 surrounding highly heterozygous
74 SNPs ($0.3 < \text{minor allele frequency} < 0.5$) distributed within the 1 Mb on chromosome
75 1 (Figure 1). Our study indicated that the haplotype-based NIPT for MMA through
76 small target capture region sequencing is technically accurate and feasible, and also
77 further highlighted the feasibility of NIPT of monogenic diseases.

78

79 **Materials and Methods**

80 **Patient recruitment**

81 Twenty-one at-risk families including singleton pregnant woman, her husband, and
82 proband diagnosed with MMA were enrolled at Shanghai Xin Hua Hospital with
83 genetic counseling and informed consent. The institutional review board (IRB) of BGI
84 and Shanghai Xinhua Hospital approved our study with approval number BGI-IRB
85 NO. FT15195 and XHEC-C-2015-025.

86 Ten causative mutations in four exons of *MMACHC* gene in probands and their
87 parents were already identified prior to performing NIPT of MMA during pregnancy.
88 Before amniocentesis, blood samples were drawn from each pregnant woman at
89 16–20 weeks of gestation and her family members was used for NIPT of MMA. The
90 maternal plasma should be separated as soon as possible. Amniotic fluid (AF) samples
91 for routine prenatal diagnosis were obtained at 16–18 weeks of gestation. The data
92 analysis of fetal DNA sample was blinded to NIPT. The AF samples of F01- F06

93 family were also sent to BGI to evaluate the accuracy of inferred paternal and
94 maternal specific loci of fetus compared with the AF standard haplotype.

95 **Targeted sequencing**

96 The extraction of gDNA and maternal plasma DNA was accomplished using the
97 commercial kits from QIAGEN. The NGS library of gDNA was constructed according
98 to the Illumina standard protocol. The construction of cell-free DNA library as a result
99 of its micro inputs nature was performed by the Kapa Biosystem library preparation
100 kit. The hybrid captures of gDNA and cf-DNA libraries were separately carried out
101 using the same probe. The post-capture libraries were sequenced using PE 101 bp on
102 Illumina platform (Hiseq 2500).

103 **Variation calling**

104 The raw data were aligned to the human reference sequence (Hg19, GRCh37) by
105 BWA software (0.7.12) in the paired end mode. After removal of low-quality reads
106 including duplicated reads and multiple aligned reads using Picard Tools, Variation
107 calling was accomplished through GATK software. Only the variations with depth
108 greater than 50x will be analyzed in the next step.

109 **Estimation of fetal concentration and plasma sequencing error**

110 The fetal genotype should be heterozygous state with different homozygous
111 genotype of parents according to the Mendel's laws. So, it could be estimated as two
112 times of the percentage of the minor allele depth to the total depth of this allele. The
113 sequencing error could be described as the ratio of the count of different loci reads to
114 total count of this SNP reads when the genotypes of parents are same homozygous.

115 **NIPT for MMA**

116 Haplotypes linked with wild and mute allele were constructed using SNP
117 information within flanking and coding region of *MMACHC* gene (You et al. 2014).

118 Hap 0 was defined as the pathogenic haplotype and Hap1 was defined as the
119 wild-type haplotype. The fetal inheritance from father was determined using the set of
120 SNPs that were heterozygous in father but homozygous in mother. SNPs that were
121 heterozygous in mother but homozygous in father were used to determine fetal
122 inheritance from mother. Hidden Markov Model (HMM) and Viterbi algorithm was
123 used to deduce the fetal haplotypes using the target region data of plasma (Ma *et al.*
124 2017).

125 **Accuracy of NIPT for MMA**

126 The Sanger sequencing and standard haplotypes of fetal gDNA obtained using
127 amniotic fluid was further operated to validate the uniformity of NIPT result.

128

129 **Results**

130 **Sequencing data of recruited families**

131 Sanger sequencing of *MMACHC* was operated to determine the variation of
132 pathogenic mutation (Table 1). The mean depth of gDNA and cf-DNA was about
133 147.48x (63.41x–348.05x) and 237.97x (89.67x–396.23x), respectively. The coverage
134 with more than 20x was approximately 98.14% (90.09–99.68%). The mean capture
135 efficiency and duplicate rate was 50.19% (27.12%–75.23%) and 7.37%
136 (4.98%–44.09%) in all the samples (Table S1).

137 **Estimation of fetal concentration and plasma sequencing error**

138 For these twenty-one pregnant women, the cf-DNA concentrations varied from
139 4.62% to 18.96% during the second trimester (Table 2), showing significant
140 differences between the individuals. The mean sequencing error rate of plasma was
141 0.41% (0.02–1.18%) (Table 2). The data suggested high experimental quality for next
142 analysis.

143 **Construction of Parental haplotype**

144 The parental haplotype was constructed by the genotyping information from a
145 trio strategy of father, mother and proband. Clean data with more than 20-fold
146 coverage were about 97.88% and about 1041 SNPs on the target region were detected.

147 **Noninvasive prenatal diagnosis of fetal MMA**

148 The number of SNPs identified ranged from 854 to 1211. The number of
149 informative SNPs which were used to predict the combination of fetal haplotype
150 inherited from mother and father were 122 (20–323) and 113 (24–290). Parental
151 haplotypes were successfully constructed in F01 family. 155 SNPs was used to
152 determine that the fetus inherited wild allele from the father and none SNP supported
153 that fetus inherited the pathogenic haplotype. 86 SNPs was used to determine that the
154 fetus inherited wild allele from the mother and none SNP supported that fetus
155 inherited the pathogenic haplotype. So, the fetus of F01 was normal because of the
156 F1+M1 haplotype (Table 3 and Figure 2). Based on this strategy, nine fetuses were
157 diagnosed as carriers, nine fetuses were normal and three fetuses were affected by
158 cblC type of MMA due to the compound heterozygous mutation of *MMACHC* (Table
159 3 and Figure 2).

160 **Accuracy of NIPT for MMA**

161 The Sanger results of fetal DNA were 100% consistent with NIPT result (Table 2)
162 and SNPs deduced using NIPT were 100% consistent with standard haplotypes of
163 fetal gDNA in F01 to F06 (Table S2).

164

165 **Discussion**

166 The cumulative incidence of monogenic disease accounted for 7% of birth defects,
167 while the chromosomal abnormalities accounted for 6% according to the March of

168 Dimes Global Report on Birth Defects 2006. A growing number of birth defects and
169 diseases can be diagnosed prenatally and treated before birth in some cases. With the
170 large-scale clinical application of NIPT for fetal aneuploidies based on massively
171 parallel sequencing approach (Benn *et al.* 2012; Norton *et al.* 2012; Dar *et al.* 2014),
172 NIPT of monogenic disease remains the next frontier. In the recent years, NIPT of
173 monogenic disorders has been actively investigated and it has a great clinical
174 application prospect, providing requisite prognostic data for clinical intervention.
175 Initially, NIPT of monogenic disorders relied on the detection or exclusion of
176 paternally inherited mutations or de novo mutations by direct detection method based
177 on PCR. PCR-based method such as ddPCR (Lun *et al.* 2008), QPCR (Guissart *et al.*
178 2017), PCR-RED (Chitty *et al.* 2015) and cSMART (Chen *et al.* 2016; Han *et al.* 2017)
179 have been reported in β thalassemia, cystic fibrosis, thanatophoric dysplasia, Wilson
180 disease and autosomal recessive nonsyndromic hearing loss, respectively. PCR-based
181 method involves the advantage of simple operation. However, the design of primer or
182 probe at the mutation site that is confined only to SNP and indel remains difficult.
183 NIPT based on PCR cannot be applied to the major mutation type with copy number
184 variance (CNV). The sensitivity and specificity of PCR-based NIPT is affected by
185 fetal DNA fraction and quality of sample.

186 Maternally inherited alleles and alleles shared by both parents were detected by
187 more sophisticated techniques such as haplotype-based strategy called indirect
188 method. Haplotype-based strategy such as relative haplotype dosage (RHDO) (Lam *et al.*
189 2012) and proband-assisted haplotype phasing have been successfully reported in
190 several recessive monogenic diseases. These historical data were retrospectively
191 analyzed in our study (Table 4). Fifty-seven high risk families who had an affected
192 child were enrolled in the clinical research to validate the accuracy and feasibility of

193 NIPT. All the NIPT results were consistent with invasive prenatal diagnosis. Overall,
194 the data obtained from all the 57 families included in the test have shown sensitivity
195 and specificity rates of 100%, with 0% failure rate. The accuracy of inferred fetal
196 maternal alleles and paternal alleles using plasma sequencing data were almost 100%
197 compared with the standard haplotype obtained by the AF data without considering
198 the recombination point. Other experiments were necessary to determine the precise
199 positioning of recombination and whether the proband or fetus involve the
200 recombination. Ye *et al.* (2018) showed that when the plasma sequence depth was
201 200X, the accuracy of fetal inherited maternal haplotype was above 99% and when
202 the fetal fraction was between 5% and 10%, the mean number of SNP was about 20 to
203 reach 99% detection accuracy. Previous data showed that the haplotype-based NIPT
204 can be applied to genes with highly homologous sequences like CYP21A2 and SMN1,
205 which is almost impossible to be detected by directly sequencing the pathogenic
206 mutations.

207 MMA is one of the most common disorders of congenital organic acid metabolism.
208 Early prenatal treatment may have an impact on the long-term complications
209 associated with cbIC disease. Prior studies have evaluated the effects of prenatal
210 HOCbl administration and the results showed a decrease in the maternal metabolites
211 (Huemmer *et al.* 2005). Although the haplotype-based strategy was successfully
212 implemented in several autosomal recessive disorders, this has not been applied in
213 MMA till now. The feasibility and accuracy of this method depends on the
214 informative SNPs used to construct the haplotype and the selection of the targeted
215 region, and hence investigation of technical feasibility is necessary for each target
216 disease. Our study initially demonstrated the feasibility of haplotype-based NIPT for
217 *MMACHC*. In our study, the fetal genotype of 21 families was successfully

218 determined using haplotype-assisted NIPT. The result of this study was consistent
219 with the findings of invasive method. Based on these data, the feasibility and accuracy
220 of NIPT of MMA using haplotype strategy has been demonstrated. The sensitivity
221 and specificity rates were both 100%, with 0% failure rate. The accuracy of the fetal
222 alleles inherited from parents deduced by haplotype strategy were almost 100%
223 compared with the standard haplotype obtained by the AF data in families from F01
224 to F06 (Table S2). The other family results showed that the haplotype-based NIPT
225 method allowed 100% concordance with the invasive diagnostic approach.

226 In most of the reported cases, the targeted region is always relatively large, and the
227 sequencing cost remained exorbitant, hampering the clinical utility of this technology.
228 In this research, we have demonstrated the feasibility of NIPT of MMA by
229 sequencing a much smaller region. This method requires trio members of the family
230 for constructing the parental haplotype linked to the mutation allele. The sensitivity
231 and specificity of this method have been reported to be 100% validated by the
232 invasive result, which is the greatest advantage of haplotype-based strategy NIPT. A
233 fatal flaw of the haplotype-based strategy is that the feasibility is highly depend on the
234 availability of proband sample.

235 Linked reads method like 10xGenomics to specify the individual haplotype
236 directly without proband has been recently reported by Dennis Lo in NIPT of
237 monogenic diseases (Hui *et al.* 2017), but the technology has several limitations. The
238 most important restrictive factor for clinical application was the detection success
239 ratio. In this article, 12 of 13cases were successfully detected and the failed cases
240 were unable to determine the fetal genotype due to insufficient number of informative
241 SNPs. In our research, we used the coding region and highly heterozygous SNPs with
242 1M flanking sequence of *MMAHC* gene to construct the haplotype, and the number

243 of informative SNPs were sufficient to build the fetal haplotype based on HMM
244 model. Another limitation was that the phasing block size using 10x Genomics linked
245 reads. The block size was affected by the integrity of input DNA and rigorous
246 experimental operation. The phasing block stridden across the target gene and
247 sufficient SNPs surrounding this region coexisted to ensure the success rate. In
248 summary, 10x Genomics linked the reads have solved the problem of not relying on
249 the precursor, but the success rate, high cost and complex operation restricted its
250 clinical application. Further study is needed to develop cost-effective and simplified
251 technology for NIPT of monogenic disease.

252 In conclusion, we demonstrated the feasibility of haplotype-based NIPT of MMA
253 by sequencing a much smaller region. The sequencing depth of plasma and the
254 number of informative SNPs were 200x and 20x, respectively. The proposition
255 couples who have been diagnosed as monogenic carriers and have an affected child,
256 our method is applicable to assess the repregnant fetal genotype to clinical
257 intervention. We here provided the evidence that the same approach can be applied to
258 other autosomal-recessive disorders, and the overall sensitivity and specificity was
259 100% on 78 patients tested from the previous and our study.

260

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265

266 **Competing financial interests**

267 The authors declare no conflict of interest.

268

269 **Author Contributions**

270 L.H., J.Y., W.Q., H.Z., L.L., and F.G, obtained patient materials; Y.W., H.W., and
271 X.J. conducted gene analysis in blood or amniotic fluid cell DNA; C.C., J.S. and Z.P.
272 designed the study and drafted the paper. Y.W., F.G. and W.L analyzed the data,
273 interpreted the data and wrote a part of the paper.

274 **Data Availability**

275 The authors affirm that all data necessary for confirming the conclusions of the article
276 are present within the article, figures, and tables. Supplemental materials reported in
277 this study are also available in the CNGB Nucleotide Sequence Archive (CNSA,
278 <ftp://ftp.cngb.org/pub/CNSA/CNP0000164/Supplementation/>)

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368 **Figures Legends**

369 **Figure 1 Target region of *MMACHC* gene and SNPs used for haplotyping.**

370 Custom NimbleGen probes within the 141.39 kb region was designed including the
371 *MMACHC* gene coding region and 1125 surrounding highly heterozygous SNPs
372 distributed within 1 Mb region.

373 **Figure 2 Fetal Haplotype Prediction.**

374 X-axis represents the locus on target region; Y-axis represents the logarithm of the
375 ratios of different fetal haplotype combinations. The black lines above zero (cyan
376 lines) indicate that the fetus inherited the pathogenic haplotype (Hap0), and the black
377 lines below zero indicate that the fetus inherited the normal haplotype (Hap1). Two
378 purple vertical lines represent the region of the *MMACHC* gene. Left chart and right
379 chart represents the results of fetal-inherited paternal haplotype and maternal
380 haplotype, respectively.

Table 1. Molecular Diagnosis Results in 21 Families.

Family	Gene	Mutation Type			GW ^b
		Father	Mother	Proband	
F01	MMACH	c.609G>A/N	c.658_660delAAG/N ^a	c.609G>A/c.658_660delAAG	17
F02	MMACH	c.609G>A/N	c.567dupT/N	c.567dupT/c.609G>A	18
F03	MMACH	c.441TG[2]/N	c.609G>A/N	c.609G>A/c.441TG[2]	17
F04	MMACH	c.609G>A/N	c.80A>G/N	c.80A>G/c.609G>A	17
F05	MMACH	c.441TG[2]/N	c.609G>A/N	c.441TG[2]/c.609G>A	17
F06	MMACH	c.609G>A/N	c.80A>G/N	c.80A>G/c.609G>A	18
F07	MMACH	c.609G>A/N	c.609G>A/N	c.609G>A/ c.609G>A	18
F08	MMACH	c.609G>A/N	c.217C>T/N	c.217C>T/c.609G>A	17
F09	MMACH	c.609G>A/N	c.315 C>G/N	c.609G>A/c.315 C>G	16
F10	MMACH	c.609G>A/N	c.609G>A/N	c.609G>A/ c.609G>A	18
F11	MMACH	c.658_660delAAG/N	c.609G>A/N	c.609G>A/c.658_660delAAG	17
F12	MMACH	c.609G>A/N	c.609G>A/N	c.609G>A/ c.609G>A	18
F13	MMACH	c.609G>A/N	c.80A>G/N	c.609G>A/c.80A>G	17
F14	MMACH	c.658_660delAAG/N	c.609G>A/N	c.609G>A/ c.658_660delAAG	17
F15	MMACH	c.609G>A/N	c.609G>A/N	c.609G>A/ c.609G>A	19
F16	MMACH	c.609G>A/N	c.656_658delA GA/N	c.656_658delA GA/c.609G>A	18
F17	MMACH	c.658_660delAAG/N	c.609G>A/N	c.609G>A/c.658_660delAAG	16
F18	MMACH	c.445_446delITG/N	c.609G>A/N	c.609G>A/c.445_446delITG	17
F19	MMACH	c.394C>T/N	c.656_658delA GA/N	c.394C>T/c.656_658delA GA	17
F20	MMACH	c.609G>A /N	c.609G>A /N	c.609G>A/ c.609G>A	17+5
F21	MMACH	c.445_446delITG/N	c.482G>A/N	c.482G>A/c.445_446delITG	17

Abbreviations: N^a, normal; GW^b, gestational weeks.

Table 2. Statistics of Error Rate and Fetal DNA Fraction.

Pedigree	Type 1 SNP^a	Type 2 SNP^b	Error Rate	Fetal DNA fraction
O1	86	5	0.0200%	7.64%
F02	104	463	1.1788%	11.45%
F03	79	430	0.1936%	16.23%
F04	107	215	0.2437%	9.07%
F05	85	14	0.3360%	7.66%
F06	99	43	1.0153%	18.96%
F07	125	1	0.1631%	14.89%
F08	66	421	0.7156%	10.93%
F09	80	23	0.0941%	4.62%
F10	110	61	0.4655%	11.41%
F11	115	4	0.2434%	17.84%
F12	82	149	0.3810%	6.85%
F13	329	39	0.2170%	11.70%
F14	64	24	0.4847%	9.86%
F15	66	7	0.5687%	11.66%
F16	70	165	0.3680%	10.77%
F17	71	441	0.2319%	13.03%
F18	49	166	0.8472%	10.52%
F19	238	141	0.2406%	5.90%
F20	66	23	0.3877%	5.85%
F21	143	8	0.2170%	8.20%

SNPs^a that were homozygous with the same type of parents but different bases in the plasma, which were used to calculate the sequencing error rate of plasma; SNPs^b that were homozygous in both parents but different types, which were used to calculate fetal DNA fractions.

Family	Gene	SNPs For F0 ^a	SNPs For F1 ^a	SNPs For M0 ^a	SNPs For M1 ^a	Fetal Haplotype	Fetal Genotype	NIPT Results	Invasive Test Results	Consistency ^b
F01	MMACHC	0	155	0	86	F1+M1	N	Normal	N	Y ^c
F02	MMACHC	37	0	0	86	F0+M1	c.609G>A/N	Carrier	c.609G>A/N	Y
F03	MMACHC	0	60	73	0	F1+M0	c.609G>A/N	Carrier	c.609G>A/N	Y
F04	MMACHC	0	42	94	0	F1+M0	c.80A>G/N	Carrier	c.80A>G/N	Y
F05	MMACHC	109	0	122	0	F0+M0	c.441TG[2]/c.609G>A	affected	c.441TG[2]/c.609G>A	Y
F06	MMACHC	0	325	20	0	F1+M0	c.80A>G/N	Carrier	c.80A>G/N	Y
F07	MMACHC	0	140	0	91	F1+M1	N	Normal	N	Y
F08	MMACHC	129	0	42	0	F0+M0	c.217C>T/c.609G>A	affected	c.217C>T/c.609G>A	Y
F09	MMACHC	0	160	77	0	F1+M0	c.315 C>G/N	Carrier	c.315 C>G/N	Y
F10	MMACHC	0	24	0	273	F1+M1	N	Normal	N	Y
F11	MMACHC	0	81	0	149	F1+M1	N	Normal	N	Y
F12	MMACHC	0	43	203	0	F1+M0	c.609G>A/N	Carrier	c.609G>A/N	Y
F13	MMACHC	0	26	0	117	F1+M1	N	Normal	N	Y
F14	MMACHC	220	0	96	0	F0+M0	c.609G>A/c.658-660delAAG	affected	c.609G>A/c.658-660delAAG	Y
F15	MMACHC	0	27	86	0	F1+M0	c.609G>A/N	Carrier	c.609G>A/N	Y
F16	MMACHC	290	0	0	111	F0+M1	c.609G>A/N	Carrier	c.609G>A/N	Y
F17	MMACHC	0	109	0	84	F1+M1	N	Normal	N	Y
F18	MMACHC	0	54	0	323	F1+M1	N	Normal	N	Y
F19	MMACHC	108	0	0	128	F0+M1	c.394C>T/N	Carrier	c.394C>T/N	Y
F20	MMACHC	0	27	0	230	F1+M1	N	Normal	N	Y
F21	MMACHC	0	274	0	73	F1+M1	N	Normal	N	Y

Table 3. The NIPT Results of 21 Studied Families.

^aSNPs for F/M represent the number of SNP supporting the inheritance of fetal haplotype from parents. F0/M0, fetal-inherited mutant haplotype; F1/M1, fetal-inherited normal haplotype; ^bConsistency represents the comparison of NIPT results with invasive testing results; ^c Y, yes; N, normal.

Table 4. Studies Reporting on NIPT of Monogenic Disease.

Author	Method	Disease	Case No.	NIPT Results	Consistency	Accuracy of maternal alleles	Accuracy of paternal alleles
Dennis Lo	RHDO	β -thalassemia	2	2 Carriers	Y	-	-
Dennis Lo	RHDO	CAH	14	5 Carriers+2 Normal+7 affected	Y	-	-
Yanqin You	PAHP	MSUD	1	affected	Y	-	-
Zhengfeng Xu	PAHP	CAH	1	Carriers	Y	96.41%	97.81%
Meng Meng	PAHP	GJB2	1	Carriers	Y	-	-
Yan Xu	PAHP	DMD	8	1 Carriers+2 Normal+5 affected	Y	99.98%	-
Min Chen	PAHP	SMA	5	2 Carriers+2 Normal+1 affected	Y	-	-
Zhengfeng Xu	PAHP	CAH	12	6 Carriers+6 affected	Y	100%	100%
Xuefan Gu	PAHP	PKU	13	4 Carriers+4 Normal+5 affected	Y	100%	100%
Total	-	-	57	22 Carriers+10 Normal+25 affected	Y		

RHDO, relative haplotype dosage; PAHP, proband assisted haplotype phasing; consistency means whether the NIPT result was consistent with the invasive prenatal testing, Y represents yes.

Figure 1

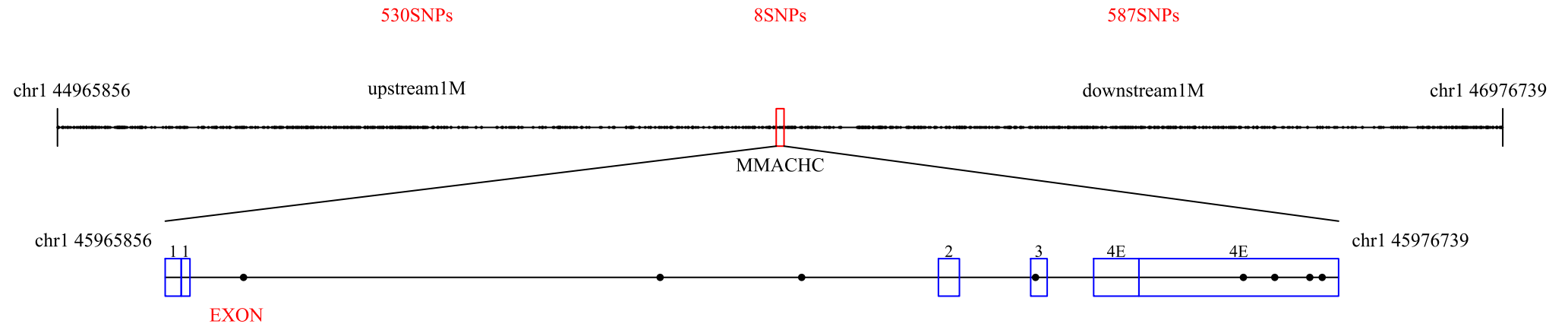


Figure 2

