Genomic insights into evolutionary journey of the porcine endogenous retroviruses

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Abstract

Background: Xenotransplantation may overcome significant shortage of human allotransplant. Porcine organs are considered favorable for xenotransplantation duo to similar size and function to human organ. However, porcine endogenous retroviruses (PERVs) are potential infectious agents during xenotransplantation as they are able to infect and horizontally transfer among human cells. Furthermore, PERVs can be endogenized in pig genomes and are transmitted genetically in a Mendelian fashion. Here, we depict a complex evolutionary history of modern PERVs.

Results: We *in silico* mined 142 mammalian genomes and 14 pig genomes. This led to the documentation of 185 PERVs and a new viral cluster. Large-scale genomic alterations were found in most PERVs including many insertion-deletion events and which are suggestive of ancient origins, and pig genomes have been shaped by PERV-mediated genomic rearrangement during evolution. Notably, we found that lesser Egyptian jerboa and rock hyrax harbor ancestral PERV-related elements indicative of ancient cross-species transmission events from none-porcine species to pigs. A comprehensive analysis of these viral "fossils" suggested that recombination among none-porcine endogenous retroviruses led to the origination of PERVs.

Conclusion: For the first time, using large scale genomic mining, we decipher a complex evolutionary history for the PERVs. These new findings help us to understand the past of PERVs which pose the potential risk in clinical trials of xenotransplantation and provide novel insights into the origin and evolution of a human-infecting pathogen.

37 **Keywords:**   porcine endogenous retrovirus, genomic rearrangement, cross-species

38 transmission, evolution, origin, recombination

40 Background

Xenotransplantation, the transplantation of tissues and organs from one species to another,

may alleviate shortages of human donor organs(1, 2). Porcine organs are suitable for

xenotransplantation due to their similar size and function to human organs, and the fact

that pigs can be bred in large numbers(3). However, porcine endogenous retroviruses

(PERVs) are able to infect and horizontally transfer among human cells, raising major

concerns about the safety of xenotransplantation(3, 4), particularly given that retroviruses

are often associated with severe infectious disease or oncogenic disease(3, 5-7).

PERVs are endogenous gammaretroviruses, and exist in the genomes of all pig strains(3,

8). The envelope (*env*) genes of three PERV classes (PERV-A, -B and -C) differ with

respect to the receptor-binding domain (RBD)(9). Although there is no evidence of PERV

transmission in patients receiving encapsulated pig islets(10-12), PERV-A and -B have

been observed to infect both human cells and pig cells while PERV-C infects only pig

cells(13). PERVs may also integrate into the human genome *in vitro*(14, 15). In pig cells,

PERV-C can recombine with the *env* of PERV-A to produce A/C recombinants, which can

infect   human   cells   more   efficiently(13).   This   increases   the   inherent   risk   in

xenotransplantation and xenogeneic cell therapies.

While several studies have examined the evolutionary relationships between PERVs and

other viruses, the origin and evolution of PERVs remains uncertain, particularly why they

are unable to infect humans(16, 17). At least two species that belong to the same order as

pigs, *Tayassu pecari* (of Eocene origin) and *Babyrousa babyrussa* (of Miocene origin)

lack PERVs(18). However, the common warthog (*Phacochoerus africanus*) carries

PERVs, suggesting that an ancestral porcine species carried PERVs(18). PERVs have two

different types of long terminal repeat (LTRs), one with a 39-bp repeat structure in the U3

region, and the other without this repeat structure(19, 20). The 39-bp repeats carried by

PERV-A and -B confer strong promoter activity and thus increase transcription(19, 20).

However, the 39-bp repeat structure is absent in some PERV-A and all PERV-C. These

PERVs thus have low transcriptional activity(19, 20). BLAST search analysis confirmed

that the R and U5 regions of the PERV LTRs are highly conserved in the pig and mouse

genomes (74–87% identity)(21). Indeed, the LTRs of PERV-A, -B and LTR-IS (a LTR

family found solely in the mouse genome) have similar structures(21). The conserved LTR

sequences found in pigs and mice might have originated from a common exogenous viral

element, but evolved independently(21). Using genome mining with all available

mammalian genomes, we clearly illustrate the evolutionary journey of the modern PERVs

and reveal that the genesis of PERVs is much more complex than previously thought.

Results

In silico **characterization of putative PERVs.**

Using previously reported PERV sequences as queries, we mined 14 pig genomes

(Additional files 2: Table S1) available in GenBank, and showed a detailed genome-wide

distribution of full-length PERVs (i.e. containing two LTRs). We initially compiled a

PERV data set that included 185 putative PERVs (containing at least one LTR) (Additional

files 2: Table S2). A total of 84 classified (30 PERV-A, 39 PERV-B and 15 PERV-C), 18

unclassified full-length PERVs (i.e., lacking of env gene) were retrieved from pig

genomes. We identified 2–10 full-length PERVs in each of 12 other pig breeds, including

Meishan, Goettingen, and Large White. We removed 19 previously classified PERV

sequences that were low quality fragments (> 200 "N" bases). Hence, the final data set

comprised 65 high quality classified PERVs (27 PERV-A, 29 PERV-B, and 9 PERV-C).

Their viral genomic structures are summarized in Figure 1.

Most of the PERVs exhibited large-scale genetic alterations induced by indels and stop

codons (Fig. 1), suggestive of a relatively long evolutionary history. PERV LTRs were

classified by the presence (LTR B) or absence (LTR A) of the 18 bp and 21 bp repeat

structure reported previously(9, 19, 22). Three different type B LTRs in the PERV were

identified, distinguished by the number of 18 bp and 21 bp repeat sequences: LTR B1 (two

18-bp and one 21-bp repeats), LTR B2 (three 18-bp and two 21-bp repeats), and LTR B3

(four 18 bp and three 21 bp repeats). Of the 65 high-quality PERVs we analyzed, we

assigned 57, of which 32 (>55%) carried LTR A, 10 carried LTR B1, 13 carried LTR B2,

and 2 carried LTR B3. LTR A was identified in PERV-A and -C, and LTR B1 was

identified in PERV-A and -B. LTR B2 and LTR B3 were only identified in PERV-B. The

remaining eight PERVs contained different types of 5'- and 3'- LTR which may reflect

recombination between PERVs with different LTR types (Fig. 1). For example, we

discovered one PERV (AEMK02000536.1) with LTR B2 at the 5' end and LTR A at the 3'

end (Fig. 1).

Potential genomic rearrangement via PERVs.

Retrovirus integration creates a short duplication called target site duplication (TSD)

flanking the LTR(23, 24). If chromosomal rearrangement through homologous

recombination between distant proviruses occurred, the flanked TSDs should be different,

as mentioned in a previous study of primate genomic rearrangement via ERVs (25). To

identify pig genomic rearrangement via PERVs, we firstly constructed a maximum

likelihood (ML) tree representing the 5'- and 3'- LTR sequences of full-length PERVs

(Additional files 1: Fig. S1). The phylogenetic tree was divided into three large clusters

(Additional files 1: Fig. S1), suggesting that three major integration events had occurred.

We collected PERVs with 5'- and 3'- LTR sequences did not clustered together in the

phylogenetic. Remarkably, 11 PERVs did not share the same TSD (4bp in length) (Table 1,

Additional files 2: Table S3), thus these PERVs could reflect porcine genomic

rearrangement during evolution.

Table 1. PERVs with different target site duplications (TSDs).

Name	Accession number	Divergent LTRs based on structure [*]	Divergent LTR based on tree	Flanking TSD [**]	
				5'	3'
AEMK02000536.1	AEMK02000536.1	yes	yes	AGCC	CTTT
CM000818.5_a	CM000818.5	no	yes	GTTC	CTTC

CM000826.5_c	CM000826.5	no	no	ACCA	AATC
CM000828.5_d	CM000828.5	no	yes	CCAC	CACC
KQ001967.1	KQ001967.1	no	yes	CCAC	CACC
LIDP01000017.1	LIDP01000017.1	no	yes	CCAC	CACC
LUXR01004647.1	LUXR01004647.1	yes	yes	GTTC	CTTC
LUXR01022139.1	LUXR01022139.1	yes	yes	CCAC	CACC
LUXX01045907.1	LUXX01045907.1	no	yes	CCAC	CACC
LUXX01080744.1	LUXX01080744.1	no	yes	GTTC	CTTC
LUXY01101100.1	LUXY01101100.1	no	yes	CCAC	CACC

[*]LTRs of PERVs comprise four types (LTR A, B1, B2, and B3). If two different types of

LTRs are flanking the PERV, the LTRs are considered "divergent".

[**]Only TSDs flanking the intact 5' and 3' LTRs sequences were analyzed

Detection of PERV-related sequences in mammalian genomes.

After screening 142 mammalian genomes (Additional files 2: Table S4) available on

GenBank using tBLASTn and choosing three major proteins (Gag, Pol and Env) of

PERVs as queries, a significant sequence (accession number: NW_004504334.1) in the

genome of lesser Egyptian jerboa (*Jaculus jaculus*) that exhibited strong sequence

similarity (for *gag* and *pol*: >75% nucleotide identity over 95% region; for *env*: >75%

nucleotide identity over 55% region) to PERVs. Using this PERV-related sequence as

query, three other possible PERV related sequences were identified in *J. jaculus* (accession

number: NW_004504375.1, NW_004504378.1, and NW_004504445.1) with >85%

nucleotide identity over 80% of the query sequence. The four PERV-related sequences

identified in *J. jaculus* were designated as eJJRVs. These four significant hits located in

large scaffolds > 5 Mb in length (Additional files 2: Table S5), with several host genes

138 identified, indicated that the eJJRV sequences were relatively reliable.

139 We were only able to identify one pair of eJJRV LTRs. This full-length eJJRV (containing

140 2 LTRs) (accession number: NW_004504334.1) is annotated in Additional files 1: Fig. S2.

141 The length of 3'- LTR of this eJJRV is 674 bp while 5'- LTR is 932 bp with a 258 bp

142 insertion. We aligned eJJRV LTRs with PERV LTRs. The start of the U3 region and the

143 end of the U5 region were distinct and not included in the alignment (Additional files 1:

144 Fig. S3). The eJJRV LTRs included a repeat structure (three 18 bp and two 21 bp repeat

145 sequences) in the U3 region, identical to that of the PERV LTR B2. Furthermore, 3' LTR

146 of eJJRV had high identity with LTR B2 of PERV (~73%). Alignment analysis revealed a

147 closer relationship between LTRs of the eJJRV and LTR B2 of PERVs (Additional files 1:

148 Fig. S3). Notably, the alignment of the conserved R region supported a close evolutionary

149 relationship between the eJJRV and PERVs (Fig. 2a). To highlight the similarity between

150 PERVs and eJJRVs we generated pairwise alignments of eJJRV and PERV nucleotides

151 using the full-length ERVs, and performed a sliding window analysis of these pairwise

152 alignments (Fig. 2b)(26, 27). For comparison, we determined the similarity of HIV-1

153 provirus sequence to that of its closest relative (chimpanzee SIVcpz) (28, 29).

154 Interestingly, *gag* and *pol* were more similar between eJJRV and PERV-A, -B and -C than

155 HIV-1 and SIVcpz (Fig. 2b). Therefore, our results indicate that eJJRVs and PERVs are

156 homologous. However, the RBD and the proline rich-region (PRR) of the surface subunit

157 (SU) of *env* were dissimilar between eJJRV and PERV-A, -B and -C, as also seen in HIV

158 and SIVcpz. As RBD determined host range(30-33), this observation suggests that eJJRVs

159 and PERVs have distinct host ranges.

We used the RBD amino acid sequences from PERV-A, -B and -C as queries to screen for

homologous viral elements. The eight significant hits (>60% amino acid identity over 80%

region) were obtained in rock hyrax (*Procavia canpensis*) of *Procaviidae*, and all eight

hits flanking with genes located in large scaffolds >0.3 Mb in length (accession number:

KN678690.1, KN676491.1, KN678005.1, KN677924.1, KN676905.1, KN676182.1,

KN680906.1, and KN676638.1) (Additional files 2: Table S5). We examined the

sequences flanking the eight hits (especially *pol*), and found that ERVs including these hits

were endogenous gamma-retroviruses. These hits were therefore designated ePCRVs. We

aligned the RBDs of PERVs and ePCRVs, and found that ePCRVs were highly similar to

PERVs (Fig. 3). Pairwise comparisons revealed that ePCRV_1 and ePCRV_2 had a high

identity to PERV-B (63%) but a low identity with PERV-A, -C and PERV-IM (40–43%).

Therefore, the RBD of ePCRVs and PERV-B are homologous.

To characterize the relationship within eJJRVs, ePCRVs and PERVs, we inferred

phylogenetic trees of Gag, Pol and Env, first removing the variable RBD (Fig. 2c-e). Our

maximum likelihood (ML) phylogenetic tree revealed that eJJRVs and PERVs clustered

together with high bootstrap supports in three phylogenies (Fig. 2c-e), suggesting that they

shared the most recent common ancestry. However, the Gag of ePCRVs clustered with

eJJRVs and PERVs while Pol and Env (without RBD) of ePCRVs were distant related to

eJJRVs and PERVs which might indicate recombinant events among ancient retroviruses

(Fig. 2c-e).

Remarkably, a new lineage close to PERV-A and PERV-C was observed, named PERV-IM

(as designation of PERV-intermediate type), and presented in all 14 pig genomes (Fig. 2e,

Additional files 2: Table S2). The Env proteins of PERV-IMs showed relatively low

similarity to PERV-A, -B, and -C, and they were clearly distinct in the RBD region (Fig.

3).

Discussion

Using systematic large-scale genome mining, we analyzed the origin and evolution of the

modern PERVs. We found homologous LTRs of PERVs (~73% identity) in eight

Muroidea species (*Mus caroli*, *M. pahari*, *M. musculus*, *M. spretus*, *Apodemus speciosus*,

A. sylvaticus, *Rattus norvegicus*, and *Phodopus sungorus*). The coding genes (*gag*, *pol*,

and *env*) near these homologous LTRs were identified (Additional files 2: Table S6). Also,

ERVs in two *Muroidea* species (*M. musculus* and *R. norvegicus*) found in previous study

were also presented(34). However, phylogenetic analysis of the Gag (>220 amino acids),

Pol (>560 amino acids) and Env (>330 amino acids) sequences of these ERVs suggested

that they were distantly related to the modern PERVs (Additional files 1: Fig. S7). We

provided robust phylogenetic results that eJJRV shared the most recent ancestry with the

PERVs (Fig. 2), and ePCRV also contributed the origin of Gag (Additional files 1: Fig.

S7). Failure to detect any other eJJRV and PERV-related elements in the remaining rodent

genome screening suggests that the virus was not vertically transmitted, suggesting

ancient horizontal transmission occurred during evolution.

PERVs, eJJRVs, and ePCRVs integrated into *Suidae*, *Dipodidae*, and *Procaviidae*, respectively. Miocene (23 - 5.33 MY) *Suidae* fossils have been found in East Africa, Europe and Asia (http://fossilworks.org/?a=taxonInfo&taxon_no=42381). Miocene *Dipodidae* fossils have been found in North Africa, Europe and Asia (http://fossilworks.org/?a=taxonInfo&taxon_no=41695); Pliocene (5.3 - 2.59 MY) *Dipodidae* fossils have been identified in East Africa, thus suggesting that the *Dipodidae* may have spread to East Africa during the Miocene. Miocene *Procaviidae* fossils have been found in South of Africa and East Africa (http://fossilworks.org/?a=taxonInfo&taxon_no=43293). According to the current fossil records, the only shared region for Miocene *Dipodidae*, *Procaviidae* and *Suidae* fossils is East Africa. It is likely that ancestral PERV might be derived from two ancient retroviruses carried by *Dipodidae* and *Procaviidae*, denoted as JJRV and PCRV in the illustration (Fig. 4), in East Africa during the Miocene time. The ancestral PERV then split into different classes (PERV-A ,-B, -IM and PERV-C), in which the PERV-B diverged earlier than other classes of PERVs (Fig. 4).

We also discovered a new class of PERV named PERV-IM and they are presented in all pig genomes we scanned (Additional files 2: Table S1). It remains unknown whether PERV-IMs are able to infect human cells. However, it is possible that non-human trophic PERV was able to participate in recombination, leading to human trophic retroviruses(35, 36). The identification of the new PERVs is crucial for future assessment of pathogen

226   transmission during xenotransplantation.

Conclusions

For the first time, we decipher a complex evolutionary history for the PERVs. The

ancestral PERV might be derived from two ancient retroviruses carried by non-porcine

species. We also suggest that pig genomes have been shaped by PERVs, as specifically

reflected by PERV-associated genomic rearrangement that have occurred during porcine

evolution. In a word, modern PERVs have a complex evolutionary history than previously

thought prior to their appearance in pigs.

Materials and methods

In silico identification of PERV and PERV-related proviruses.

To identify PERV-related elements in *Sus scrofa*, tBLASTn (37) was used and the amino

acid sequences of Gag, Pol and Env of 20 representative PERV proviruses (accession

numbers: HQ536016.1, HQ536015.1, HQ536013.1, KC116220.1, AY570980.1,

HQ540592.1, HQ536007.1, AX546209.1, AF435967.1, AY953542.1, HQ540591.1,

AY099323.1, AJ133817.1, EU523109.1, EF133960.1, AY056035.1, AY099324.1,

A66553.1, HQ536011.1, and HQ536009.1) were chosen as queries to screen the 14 pig

genomes available in GenBank that were released before November 2017. A 50% identity

over 50% of the match region was used to filter significant hits. It has been shown that

PERVs harbor two LTR structures, one with and one without a repeat structure in the U3

region(9, 19). Using two typical LTRs as queries we extended flanking sequences of

coding domains of PERVs to identify LTRs with BLASTn, and TSDs were used to define

PERV boundaries. LTR lengths were defined as 100–1,000 bp. PERVs with at least one

LTR and one coding gene were used in the evolutionary analysis.

To identify PERV-related proviruses in mammals, tBLASTn was used with the queries

described above in 20 representative PERV proviruses to search the 142 mammal genomes

available in GenBank as of November 2017. A 50% identity over 80% region was used to

filter significant hits. LTRs were identified using LTR finder(38), LTRharvest (39) and

BLASTn. LTR lengths were also defined as 100–1,000 bp.

Detection of potential genomic rearrangement via PERVs.

To search for proviruses involved in recombination and genomic rearrangement, we

constructed a maximum likelihood (ML) tree of the 5'- and 3'- LTRs of full-length PERVs

using PhyML 3.1(40) with GTR+I+Γ nucleotide substitution model. LTRs less than 250

bp were not considered. Sequence alignment was performed with MAFFT 7.222(41).

Phylogenetic analyses.

To determine the evolutionary relationship among PERVs, eJJRVs, ePCRVs and

representative gammaretroviruses (Additional files 2: Table S7), phylogenetic trees were

inferred using the amino acid sequences of full-length PERVs and PERVs with one LTR

and at least one coding gene. All Gag, Pol and Env protein sequences (Dataset S1) were

aligned in MAFFT 7.222 and confirmed manually in MEGA7(42). The phylogenetic

history of these gammaretroviruses was then determined using the maximum likelihood

(ML) method available in PhyML 3.1(40), incorporating 100 bootstrap replicates to assess

node robustness. The best-fit JTT+Γ amino acid substitution model was selected for Gag,

Pol and JTT+Γ+I for Env using the ProtTest 3.4.2(43).

Additional Files

Additional files 1. Fig. S1. Maximum likelihood (ML) tree of the 5' and 3' LTRs of all

full-length PERVs. **Fig. S2.** Detailed descriptions of eJJRV genome. **Fig. S3.** The

alignment of LTRs of PERVs and eJJRV. **Fig. S4.** The complete phylogenetic tree of Gag.

Fig. **S5.** The complete phylogenetic tree of Pol. **Fig. S6.** The complete phylogenetic tree of

Env. **Fig. S7.** The phylogenetic trees of Gag, Pol and Env of Muroidea ERVs, eJJRVs,

ePCRVs, PERVs and representative retroviruses.

Additional files 2. Table S1. The information of pig, rodent, and rock hyrax genomes

used for data mining. **Table S2.** The information of full-length and near full-length

PERVs. **Table S3.** The recombination-related information of full-length PERVs shown in

Fig. S1. **Table S4.** The information of 142 mammals used for PERV-like sequences

mining. **Table S5.** The information of genes flanking the eJJRVs and ePCRVS. **Table S6.**

The information of full-length and near full-length Muroidea ERVs. **Table S7**. The

information of representative retroviruses used for phylogenetic analysis.

Additional files 3. Data set S1. The alignments used to build the phylogenetic trees of

Gag, Pol and Env represented in Fig. S4, S5, and S6, respectively. **Data set S2.** The

alignments used to build the phylogenetic trees of Gag, Pol and Env represented in Fig.

292   S7.

Declarations

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Fig. 1 PERV proviruses in porcine genomes. (**A**) Genomic structure of PERVs,

including *gag*, *pol* and *env* genes and LTRs. Proviruses of PERV-A and PERV-A like (**B**),

PERV-B and PERV-B like (**C**) and PERV-C and PERV-C like (**D**) groups depicted based

on reference PERV-A (accession number: AF435967.1), PERV-B (accession number:

EU523109.1) and PERV-C (accession number: HQ536015.1). LTRs of PERVs were

classified by the presence (type B) or absence (type A) of the 18 bp and 21 bp repeat

structure. Type B LTRs were divided into three subtypes (LTR B1, LTR B2 and LTR B3).

LTR A, B1, B2 and LTR B3 are presented in white, light gray, dark gray and black,

respectively. Insertions and deletions (< 50 bp) are depicted with blue and red flags,

respectively. Larger insertions (>50 bp) are labeled with green arrow. Large deletions (>50

bp) are shown without lines. Stop codons are showed with a black flag. (Du, Duroc pig;

Wu, Wuzhishan pig; El, Ellegaard pig; Ti, Tibetan pig; Go, Goettingen pig; Me, Meishan

pig; Ro, Rongchang pig; Ha, Hampshire pig; Lan, Landrace pig; Pi, Pietrain pig; Ba,

Bamei pig; Be, Bekshire pig; Lar, LargeWhite pig; Ji, Jinhua pig).

Fig. 2. Comparison of PERVs, eJJRVs and ePCRVs. (**A**) Alignment of R region of LTR

in eJJRV and PERVs. (**B**) Sliding window analysis of percent sequence identity along

pairwise alignments of proviruses without LTRs. Phylogenetic trees of Gag (**C**), Pol (**D**)

and Env (**E**) inferred using the amino acid sequences of PERVs, eJJRVs, ePCRVs and

other representative gammaretroviruses (Table S7). Bootstrap values < 70% are not shown

in phylogenetic trees. Trees were rooted to Reticuloendotheliosis virus (REV). The

complete phylogenetic trees of Gag (**C**), Pol (**D**) and Env (**E**) are presented in additional

files 1: Fig. S4-S6, respectively. All abbreviations can be found in Table S7.

Fig. 3. Amino acid sequence comparison of the RBD of PERVs and ePCRVs. Two

ePCRVs (ePCRV_1(accession number: KN678005.1) and ePCRV_2 (accession number:

KN678690.1)) were aligned to PERV-A, -B, -C, and the newly discovered PERV-IM.

Fig. 4. Scenario of genesis of PERVs. A schematic representation of PERV evolutionary

history is shown, summarizing our hypothesis regarding the origin and evolution of

PERVs. Arrow on the left indicates putative timeline of each type of ERVs. Different

background colors illustrate putative evolution periods of modern PERVs. JJRV, PCRV,

ancestral PERV represent the exogenous forms of the retroviruses from their hosts. Solid

blue arrows show origination and dotted blue arrows show speciation. Abbreviation: RBD,

receptor binding domain; LTR, long terminal repeat; Gag, group antigens gene.

a

pol

gag

env

LTR   LTR

b

Kb 0 1 2 3 4 5 6 7 8 9 10

Du_AEMK02000133.1
Du_AEMK02000536.1
Du_CM000812.5_c
Du_CM000812.5_b
Du_CM000814.5_a
Du_CM000816.5_b
Du_CM000817.5
Du_CM000818.5_b
Du_CM000819.5_b
Du_CM000823.5_b
Du_CM000824.5_b
Du_CM000824.5_d
Du_CM000826.5_a
Du_CM000828.5_c
Du_CM000828.5_a
Du_CM000830.5_c
Du_LT634572.1_a
Du_LT634572.1_b
Du_LT634572.1_d
Wu_AJKK01232762.1
Ha_LUXS01031219.1
Ha_LUXS01060324.1
Pi_LUXU01038095.1
Ba_LUXV01020898.1
Be_LUXW01074277.1
Lar_LUXX01006697.1
Lar_LUXX01090830.1

c

Kb 0 1 2 3 4 5 6 7 8 9 10

Du_AEMK02000137.1
Du_AEMK02000197.1
Du_AEMK02000393.1_a
Du_AEMK02000476.1
Du_CM000812.5_a
Du_CM000814.5_c
Du_CM000815.5
Du_CM000818.5_a
Du_CM000819.5_a
Du_CM000820.5
Du_CM000822.5_b
Du_CM000823.5_a
Du_CM000825.5_b
Du_CM000826.5_b
Du_CM000827.5
Du_CM000828.5_d
Du_CM000830.5_d
Ti_AORO02006834.1
Me_LUXQ01001340.1
Go_LIDP01000017.1
Ro_LUXR01004647.1
Ro_LUXR01022139.1
Ro_LUXR01063446.1
Ha_LUXS01013084.1
Pi_LUXU01008622.1
Ba_LUXV01061131.1
Lar_LUXX01045907.1
Lar_LUXX01080744.1
Ji_LUXY001101100.1

d

Kb 0 1 2 3 4 5 6 7 8 9 10

Du_CM000825.5_a
Wu_KQ003791.1
El_AOCR01017097.1
El_AOCR01079940.1
El_AOCR01098665.1
El_AOCR01214680.1
Ro_LUXR01088996.1
Lan_LUXT01067056.1
Pi_LUXU01026052.1

| | Insertion | | Deletion | | LTR A | | LTR B2 |
| | Large Insertion | | Stop codon | | LTR B1 | | LTR B3 |

```
                1                                      40
ePCRV_1   P S A - - K S T P P N L M R S Y G F Y S C P G - H N K G R H C G G T G S H Y C G
ePCRV_2   P S A - - K S T P P N L V R S Y G F Y S C P G - N N K G R H C G G A G S Y Y C G
PERV-B    P A V - - K S T P P N L V R S Y G F Y C C P G - T E K E K Y C G G S G E S F C R
PERV-A    P G L N D Q A T P P D V L R A Y G F Y V C P G P P N N E E Y C G N P Q D F F C K
PERV-IM   P G L N T M S T P P D I L H A Y G F Y V C P G P P N N E K Y C G N P R D F F C K
PERV-C    P S L - - - T S P P D I L H A H G F Y V C P G P P N N G K H C G N P R D F F C K

                41                                     80
ePCRV_1   S W S C V T S N D G D W K W G I R K Q D L L K F S F V N P G P G K H V T M E - -
ePCRV_2   S W S C V T S N D G D W K W G V G N R D L L K F S F V N P G P G K H V T M E - -
PERV-B    R W S C V T S N D G D W K W P I S L Q D R V K F S F V N S G P G K Y K V M K - -
PERV-A    Q W S C I T S N D G N W K W P V S Q Q D R V S Y S F V N N P T S Y N Q F N Y G H
PERV-IM   E W S C V T S N - G R Y K W P T S R Q D R A S F S F V N N Y T S S D Q L N Y - -
PERV-C    Q W N C V T S N D G Y W K W P T S Q Q D R V S F S Y V N T Y T S S E Q F N Y - -

                81                                    120
ePCRV_1   - - - L - - - - - - - - - Y R D K P C S Q K D L D Y V K I Q F T E T G K T D
ePCRV_2   - - - L - - - - - - - - - Y R D K P C S Q K D L D Y V K I Q F T E T G K T D
PERV-B    - - - L - - - - - - - - - Y K D K S C S P S D L D Y L K I S F T E K G K Q E
PERV-A    G R W K D W Q Q R V Q K D V R N K Q I S C H S L D L D Y L K I S F T E K G K Q E
PERV-IM   - - - L D W - - - - Q R E G K Y K Q I S C S P S D L D Y L K I S F T E K G E Q Q
PERV-C    - - - L T W - - - - - - I R T G S P K C S P S D L D Y L K I S F T E K G K Q E

                121                                   158
ePCRV_1   P - S K W V D G L T W G I V F W K Y S R G E - G S G L H V R L N I K R - - P
ePCRV_2   T - S K W I N G L T W G V V F W K Y G G G V - G S S L H I R L K I E S - - P
PERV-B    N I Q K W I N G M S W G I V F Y K Y G G G A - G S T L T I R L R I E T G T E
PERV-A    N I Q K W V N G I S W G I V Y Y G G S G R K K G S V L T I R L R I E T Q M E
PERV-IM   N I - K W I N G M S W G I V F Y R G S G E E - R S I L T I H L R I E T Q M E
PERV-C    N I L K W V N G M S W G M V Y Y G G S G K Q P G S I L T I R L K I - N Q L E
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