Extended dengue virus panel: application to antiviral compounds studies.

- 1
- 2
- -3

4 Franck Touret^{1*}, Cécile Baronti¹, Xavier de Lamballerie¹ and Gilles Querat¹

6 Méditerranée Infection), Marseille, France

- 7 ^{*}Corresponding author: franck.touret@hotmail.fr
- 8

9 Abstract

10 Dengue is the most widespread arboviral disease with approximately 1/3 of the world 11 population at risk. Dengue viruses belong to the genus Flavivirus (Flaviviradea family) and 12 are divided into four closely related serotypes (1-4) that share 60 to 75 % identity at the amino 13 acid level. This viral diversity complicates the development of antivirals and currently they 14 are still no approved treatment. With the aim of providing an efficient tool for dengue virus 15 research particularly to evaluated compounds efficacy, we developed a panel of 19 dengue 16 viruses covering nearly all genotypes in the four serotypes. After a phylogenetic analysis we 17 selected relevant strains from our collection and design reverse genetic system based on the 18 ISA technic to generate the missing ones. Finally, we evaluated our dengue panel against 19 three, already published, compounds. We demonstrated that NITD008, which targeted the 20 very conserved active site of the polymerase, exhibited very similar EC50s regardless of the 21 dengue genotypes. But compounds targeting either directly or indirectly less conserved 22 proteins such as the capsid (ST-148) or NS4B (SDM25N) exhibited large differences of 23 activity toward the various genotypes of dengue viruses. These data reinforce our conviction 24 that there is a real need to evaluate compounds activity against a large panel of dengue 25 viruses.

26 Article

27 Dengue virus is a major threat to human health with approximately 1/3 of the world 28 population at risk of being infected. Dengue virus is the causative agent of the dengue fever 29 and the more severe Dengue Hemorrhagic Fever (DHF)¹ and Dengue Shock syndrome 30 (DSS). It belongs to the genus Flavivirus (Flaviviridae family) which comprises other 31 clinically important human pathogens such as Yellow fever virus, West Nile Virus and the recently emerging Zika Virus². Dengue is an arthropod born virus transmitted by the bites of 32 33 mosquitoes from the genus Aedes (Stegomya). This virus has a transmission cycle that 34 alternates between a sylvatic enzootic cycle using arboreal mosquitoes as vectors and an 35 urban cycle which include humans using *Aedes aegypti* and *albopictus* mosquitoes³. Dengue

^{5 &}lt;sup>1</sup>: Unité des Virus Émergents (UVE : Aix-Marseille Univ – IRD 190 – Inserm 1207 – IHU

bioRxiv preprint doi: https://doi.org/10.1101/439695; this version posted October 11, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

is a positive single strand RNA virus with a 10.7 kb genome coding for a single polyprotein
which is processed in three structural proteins (C, PrM and ENV), and seven nonstructural
proteins (NS1, NS2A, NS2B, NS3, NS4A, NS4B and NS5)⁴. Dengue is divided into four
closely relative serotypes (1-4) that share 60 to 75 % identity at the amino acid level⁵. Inside
this serotype classification, Dengue virus are also classified as genotypes with a variable
terminology between authors^{6,7}.

For researchers studying dengue virus, it can be really challenging to develop Direct Active Antivirals (DAA) against such a large viral diversity. Despite the huge effort to provide an antiviral therapy ⁸⁻¹¹, they are still no approved drugs. At this time treatments are only supportive ¹².

46 With the aim of providing an efficient tool for dengue virus research particularly to evaluated

47 compounds efficacy, we developed a panel of dengue viruses covering near all genotypes in

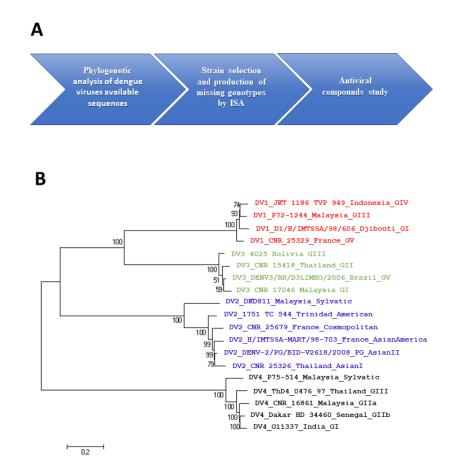
48 the four serotypes (figure 1). Whenever possible, we chosen to select strains that exhibited

49 low passages in cell cultures and were issued from recent outbreaks. The *ad. hoc.* strains were

50 selected from the European Virus Archive (EVA) collection ¹³, the French National Reference 51 center for arbovirus (CNR) and using our in house reverse genetic technic that allow us to

51 center for aboving (eff(x) and using out in house reverse generic technic that 2

52 generate infectious viruses from published full length genome sequences 14,15 .



53

54 **Figure 1: Dengue panel construction** A: Structure of the workflow used for the panel construction. B: 55 phylogenetic tree of the panel using full nucleic acid coding sequence and the maximum likelihood method

bioRxiv preprint doi: https://doi.org/10.1101/439695; this version posted October 11, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

56 (GTR+G+I model) with 500 bootstraps. Strains information: DENV-1: Genotype I: (Djibouti, AF298808,); 57 Genotype III (Malaysia, EF457905.1); Genotype IV (Indonesia, EUO7031); Genotype V (France, MF004384); 58 DENV-2: Genotype AsianAmerica (France Martinique, AF208496); Genotype American (Trinidad, 59 EU073981.1); Genotype Cosmopolitan (France, GenBank MF004385,); Genotype Asian1 (Thailand, 60 MH888331); Genotype Asian 2 (Papua New Guinea, FJ906959.1); Genotype Sylvatic (Malaysia, FJ467493.1); 61 DENV-3: Genotype I (Malaysia, MF004386, Genotype II (Thailand, MH888332); Genotype III (Bolivia, 62 MH888333); Genotype V (Brazil, JN697379.1); DENV-4 : Genotype IIb (Senegal, MF004387,); Genotype IIa 63 (Malaysia, MH888334); Genotype III (Thailand, AY618988.1); Genotype Sylvatic (Malaysia, JF262779.1); 64 Genotype I (INDIA, JF262783.1). Complete information relative to the strains of the panel are more fully 65 detailed in the supplemental material.

66

67 For this purpose, we collected dengue complete genome sequences from the NCBI database 68 and complemented this large database with those of our still unpublished "in house" and CNR 69 strains. We performed phylogenetic reconstructions with the maximum likelihood method to assign all available genome sequences to a genotype in a serotype (reduced versions of 70 71 phylogenetic trees are provided in Supplemental material fig 1,2,3 and 4). From this 72 phylogenetic analysis, we noticed that six dengue genotypes were available only as complete 73 genome sequences in the NCBI database but without biological strain counterparts (DENV-1 74 genotype III, DENV-2 genotype sylvatic and Asian II, DENV-3 genotype V and DENV-4 75 genotype III and sylvatic). To obtain these strains, we designed reverse genetics systems 76 based on ISA technic and generated synthetics overlapping DNA fragments that cover each of 77 the whole genome, bordered by a CMV promoter on the 5' end and a Ribozyme and poly-78 adenylation signal on the 3' end. The overlapping fragments were co-transfected in 79 permissible cell lines and allowed us to recover the missing biological strains to complete our 80 panel. The viral stocks were grown on Vero E6 cells and were fully sequenced. All these 81 dengue strains have been made available through the EVA collection.

82 Various specific dengue inhibitors that targeted several viral proteins involved in different 83 replication steps have been developed. ST-148, an inhibitor targeting the capsid structural 84 protein, has been reported to inhibit all dengue serotypes in cultures, although with varying efficiency, and seems promising in AG-129 mouse model infected with a Dengue 2 strain ¹⁶. 85 86 NITD008, a adenosine analog inhibitor that target the RNA-dependent RNA polymerase 87 activity was shown to be efficient against all dengue serotypes as well as other flaviviruses 11 . SDM25N a ∂ opioid receptor antagonist has been reported to target, probably indirectly, the 88 NS4 B protein. So far it has only been shown to be active against a dengue 2 strain 10 . 89 90 Depending on the target and its sequence variability and the various mechanisms of action of 91 these compounds their efficiency is likely to differ a lot regarding all the genotypes of dengue 92 viruses. To evaluate our reference dengue panel, we assess the antiviral activity of these three 93 compounds using a single common protocol based on a viral RNA yield reduction assay that 94 do not depend on the cytopathogenic potential of the strain and allow the inclusion of any 95 dengue strain in the panel. The compounds were assayed from 10 to 0.005µM with 3-fold 96 step-dilution in triplicates and the amounts of viral RNA in the supernatant during the log-97 growth curve was quantified by qRT-PCR so as to determine the EC50 (table1).

98 99 100 Table 1: Dengue panel susceptibility to three antiviral compounds assessed by yield reduction assay. Anti-capsid ST-148, nucleoside analog NITD008 and anti-NS4B SDM25N were tested against the dengue panel from 10μ M to 0.005μ M. AA: Asian American, A: American, C: cosmopolitan.

-			ST-148	NITD008	SDM25N
	Virus	Genotype	EC50 (µM)	EC50 (µM)	EC50 (µM)
Dengue 1	D1/H/IMTSSA/98/606 Djibouti	Ι	>10	0,9± 0,1	>10
	JKT 1186 TVP 949 Indonesia	IV	>10	0,3± 0,03	5,5 ±3,67
	CNR_25329 France	V	>10	2,7± 4	7,4± 0,04
	P72-1244 Malaysia	III	3± 0,5	0,9± 0,2	>10
Dengue 2	H/IMTSSA-MART/98-703 France	AA	0,8 ±0,5	0,9± 0,3	2,9± 0,95
	_1751 TC 544 Trinidad	Α	1 ± 0,7	0,3± 0,06	2,9± 0,01
	CNR_25679 France	С	1,1 ± 0,3	0,2± 0,07	1,9± 0,03
	CNR 25326 Thailand	AsianI	0,1 ± 0,03	0,9± 0,2	7,7± 0,04
	DENV-2/PG/BID-V2618/2008 Papua New Guinea	Asian II	0,2± 0,16	0,3± 0,5	4,1± 0,02
	DKD811 Malaysia	Sylvatic	0,4 ±0,18	0,4± 0,1	>10
Dengue 3	DENV3/BR/D3LIMHO/2006 Brazil	V	>10	1± 0,09	>10
	4025 Bolivia	III	>10	1± 0,05	>10
	CNR 17046 Malaysia	I	>10	2,8± 0,3	>10
	CNR 15418 Thailand	II	>10	1,2± 0,3	>10
Dengue 4	G11337 India	Ι	>10	1,2± 0,03	>10
	Dakar HD 34460 Senegal	IIb	>10	0,9± 0,3	>10
	CNR_16861 Malaysia	IIa	>10	0,4± 0,01	>10
	ThD4_0476_97 Thailand	III	0,3 ±0,08	0,2± 0,08	>10
	P75-514 Malaysia	Sylvatic	>10	1± 0,05	>10

101

For NITD008 we measured EC50 ranging from 0.2μ M to 2.8μ M in accordance to previously described activity of the compound against different serotypes ¹⁷. All dengue strains in the panel were somewhat similarly sensitive to NITD008, which is fully consistent with the flaviviral broad spectrum activity of this nucleoside analog.

106 Concerning the capsid inhibitor ST-148, all genotypes of dengue 2 strains were sensitive with 107 EC50 ranging from 0.25 to 1.1 μ M. However, we found only one sensitive genotype for 108 DENV-1 (DENV-1 GIII at 0.5 μ M) as the EC50 of the other dengue 1 genotypes were all 109 above 10 μ M. Similarly, only one DENV-4 genotype was sensitive to the compound: (DENV-110 4 GIII at 0.3 μ M), the EC50s of the others were above 10 μ M. Lastly, all DENV3 genotypes 111 were resistant to the capsid inhibitor, with EC50 > 10 μ M. Although Byrd and co-workers 112 found that DENV-2 serotype was the most sensible serotype to this capsid inhibitor and showed some variability in the inhibition, they could not fully assess the poor and highly variable susceptibility of strains from other serotypes.

115 For SDM25N, most of the dengue 2 genotypes and half of dengue 1 genotypes were

- moderately sensitive to the compound with EC50 ranging from around 2 to 8 μ M and all
- 117 dengue 3 and 4 genotypes were less sensitive with EC50 above $10 \,\mu$ M.

Overall, these results showed that compounds, like NITD008, that targeted very conserved sites such as the active site of the polymerase exhibited very similar EC50s regardless of the dengue genotypes but compounds targeting either directly or indirectly less conserved proteins such as the capsid or NS4B exhibited large differences of activity toward the various genotypes of dengue viruses. These data reinforce our conviction that there is a real need to evaluate compounds activity against a larger panel of clinically relevant dengue viruses such as our reference panel of near all available dengue genotypes.

125 Acknowledgments

We would thank the Dr Robert Tesh from the UTMB for providing us the Dengue 4 Genotype
I strain G11337 from India. We also like to thank the Dr Stéphane Priet for his careful reading
of the manuscript. As well as Magali Gilles, Fiona Baudino and Raphaelle Klitting from the
UMR UVE (Marseille, France) for excellent technical help. We also thank Geraldine
Piorkowski and Karine Barthelemy from the UMR UVE (Marseille, France) for the
sequencing. This work was supported by the research Agreement ICD#1041950 between
JANSSEN and the UMR UVE (Marseille, France).

133 Contributions

134 FT, XDL and GQ conceived the experiments. XDL allowed the funding of this study. FT, CB,

and GQ performed the experiments. FT and GQ analyzed the results. FT and GQ wrote thepaper. FT, CB, GQ and XDL reviewed and edited the paper.

137

bioRxiv preprint doi: https://doi.org/10.1101/439695; this version posted October 11, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

138 Bibliography

- 139 1. Messina, J. P. et al. Global spread of dengue virus types: mapping the 70 year history.
- 140 *Trends Microbiol.* **22**, 138–146 (2014).
- 141 2. Vasilakis, N. & Weaver, S. C. Flavivirus transmission focusing on Zika. Curr. Opin.
- 142 *Virol.* **22,** 30–35 (2017).
- 143 3. Chen, R. & Vasilakis, N. Dengue Quo tu et quo vadis? *Viruses* **3**, 1562–1608 (2011).
- 144 4. Gebhard, L. G., Filomatori, C. V. & Gamarnik, A. V. Functional RNA Elements in the
- 145 Dengue Virus Genome. *Viruses* **3**, 1739–1756 (2011).
- 146 5. Guzman, M. G. & Harris, E. Dengue. *The Lancet* **385**, 453–465 (2015).
- 147 6. Carrillo-Valenzo, E. et al. Evolution of dengue virus in Mexico is characterized by
- 148 frequent lineage replacement. Arch. Virol. 155, 1401–1412 (2010).
- 149 7. Weaver, S. C. & Vasilakis, N. Molecular evolution of dengue viruses: Contributions of
- phylogenetics to understanding the history and epidemiology of the preeminent arboviral
 disease. *Infect. Genet. Evol.* 9, 523–540 (2009).
- 152 8. Canard, B. Antiviral research and development against dengue virus. *Available Accessed*153 9, (2012).
- Coutard, B. *et al.* The VIZIER project: Preparedness against pathogenic RNA viruses.
 Antiviral Res. 78, 37–46 (2008).
- 156 10. van Cleef, K. W. R. *et al.* Identification of a new dengue virus inhibitor that targets the
 157 viral NS4B protein and restricts genomic RNA replication. *Antiviral Res.* 99, 165–171
 158 (2013).
- 159 11. Yin, Z. *et al.* An adenosine nucleoside inhibitor of dengue virus. *Proc. Natl. Acad. Sci. U.*160 *S. A.* **106**, 20435–20439 (2009).
- 161 12. Kaptein, S. J. & Neyts, J. Towards antiviral therapies for treating dengue virus infections.
- 162 *Curr. Opin. Pharmacol.* **30**, 1–7 (2016).

- 163 13. Romette, J. L. et al. The European Virus Archive goes global: A growing resource for
- 164 research. Antiviral Res. (2018). doi:10.1016/j.antiviral.2018.07.017
- 165 14. Aubry, F. et al. Single-stranded positive-sense RNA viruses generated in days using
- 166 infectious subgenomic amplicons. J. Gen. Virol. 95, 2462–2467 (2014).
- 167 15. Aubry, F. et al. 'ISA-Lation' of Single-Stranded Positive-Sense RNA Viruses from Non-
- 168 Infectious Clinical/Animal Samples. *PloS One* **10**, e0138703 (2015).
- 169 16. Byrd, C. M. et al. A Novel Inhibitor of Dengue Virus Replication That Targets the Capsid
- 170 Protein. Antimicrob. Agents Chemother. 57, 15–25 (2013).
- 171 17. Xie, X., Zou, J., Wang, Q.-Y. & Shi, P.-Y. Targeting dengue virus NS4B protein for drug
- 172 discovery. Antiviral Res. **118**, 39–45 (2015).
- 173
- 174
- 175