

Converging roles of glutamate receptors in domestication and prosociality

Thomas O'Rourke^{1,2} and Cedric Boeckx^{1,2,3}

¹Universitat de Barcelona

²Universitat de Barcelona Institute of Complex Systems

³ICREA

October 10, 2018

Abstract

Building on our previous work and expanding the range of species considered, we highlight the prevalence of signals of positive selection on genes coding for glutamate receptors (most notably kainate and metabotropic receptors) in domesticated species and anatomically modern humans. Relying on their expression in the central nervous system and phenotypes associated with mutations in these genes, we claim that regulatory changes in kainate and metabotropic receptor genes have led to alterations in limbic function and Hypothalamic-Pituitary-Adrenal axis regulation, with potential implications for the emergence of unique social behaviors and communicative abilities in (self-)domesticated species.

1 Introduction

Under one account of recent human evolution, selective pressures on prosocial behaviors led not only to a species-wide reduction in reactive aggression and the extension of our social interactions [1], but also left discernible physical markers on the modern human phenotype, including our characteristically “gracile” anatomy [2, 3].

It has long been noted that these morphological differences resemble those of domesticated species when compared with their wild counterparts [4]. Experimental observation of domestication unfolding in wild farm-bred silver foxes has unequivocally shown that selection for tameness alone can affect developmental trajectories to bring about a suite of physiological and behavioral traits indicative of the “domestication syndrome” [5]. This raises the possibility that morphological changes in *Homo sapiens* resulted from selective pressures on reduced reactivity to encounters with conspecifics. This predicts overlapping

regions of selection and convergent physiological effects in the genomes of domesticated species and modern humans.

In a recent comparative study, we have shown that genes with signals of positive selection pooled across different domesticated species — dog, cattle, cat, and horse — have above-chance overlap with genes exhibiting signals of selection in AMH, suggestive of convergent evolutionary processes [3]. Signals on glutamate receptor genes were identified more consistently than any other gene class across human and domesticate selective-sweep studies, and will be the focus of this paper.

Glutamate is the primary excitatory neurotransmitter in the vertebrate nervous system, essential for fast synaptic transmission and plasticity, learning, memory, and modulation of Hypothalamic-Pituitary-Adrenal (HPA) activity. [6]. The 26 glutamate receptors are primarily localized at synaptic nerve terminals in the brain and are divisible into two broad families (ionotropic and metabotropic). There are various widely used names for each receptor and corresponding gene in the literature. For ease of reference, see Table 1.

Ionotropic								
NMDA			AMPA			Kainate		
HUGO	Aliases		HUGO	Aliases		HUGO	Aliases	
GRIN1	NR1	GluN1	GRIA1	GLUR1	GLUA1	GRIK1	GLUR5	GLUK1
GRIN2A	NR2A	GluN2A	GRIA2	GLUR2	GLUA2	GRIK2	GLUR6	GLUK2
GRIN2B	NR2B	GluN2B	GRIA3	GLUR3	GLUA3	GRIK3	GLUR7	GLUK3
GRIN2C	NR2C	GluN2C	GRIA4	GLUR4	GLUA4	GRIK4	KA1	GLUK4
GRIN2D	NR2D	GluN2D				GRIK5	KA2	GLUK5
GRIN3A	NR3A	GluN3A						
GRIN3B	NR3B	GluN3B						
						Delta		
						GRID1	GLUD1	
						GRID2	GLUD2	

Metabotropic								
Group I			Group II			Group III		
HUGO	Aliases		HUGO	Aliases		HUGO	Aliases	
GRM1	mGluR1	mGlu1	GRM2	mGluR2	mGlu2	GRM4	mGluR4	mGlu4
GRM5	mGluR5	mGlu5	GRM3	mGluR3	mGlu3	GRM6	mGluR6	mGlu6
						GRM7	mGluR7	mGlu7
						GRM8	mGluR8	mGlu8

Table 1: Glutamate receptors. (Throughout the present study we use HUGO nomenclature to refer to both receptor genes and proteins.)

Given their importance for the normal functioning of the organism, glutamate receptors are rarely subject to extensive structural changes from one species to the next [7, 8]. Despite this high structural conservation, there are significant differences between humans and chimpanzees in the cortical expression of glutamate receptor genes [9, 10], suggesting that changes to regulatory regions may have had important functional consequences for the emergence of the human cognitive phenotype. Similarly, the vast majority of selective sweeps or high-frequency changes on glutamate receptor genes in AMH relative to archaic *Homo* are found in regulatory regions that control gene expression [11, 12].

Like the tameness of domesticates towards carers, prosociality among humans necessitates a reduction in fearful and aggressive reactions to encounters with conspecifics. Fear, anxiety, and aggression are stress responses, mediated across vertebrates by the hypothalamic-pituitary-adrenal (HPA) axis [13, 14], the hypofunction of which has been proposed to be a key mechanism in the development of tame behaviors in domesticates [15, 16]. Here we articulate the hypothesis that increased modulatory actions of glutamate receptors have brought about attenuation of the HPA stress response in ours and domesticated species. We first characterize the extent to which certain (sub)families of glutamate receptor genes exhibit shared signals of selection in domesticates and modern humans. We then discuss the likely functional implications of these changes with reference to gene expression data and evidence from developmental and psychiatric disorders.

2 Results

In earlier work, we have pointed out the intersection of glutamate receptor genes showing signals of selection in humans, dogs, cats, cattle, and horses [3]. Here we take into account a much broader range of human and domestication studies showing changes in AMH, dogs, cats, cattle, horses, foxes, sheep, pigs, rabbits, yaks, goats, guinea pigs, chickens, and ducks. (Although we focus almost exclusively on signals of selection on glutamate receptor genes, changes to other glutamatergic signaling genes have also been identified in modern human and domestication studies. These include glutamate transporter, accessory subunit, and G-protein signaling genes. We review some of the most noteworthy of these in Supplementary section S1.)

At least one glutamate receptor gene shows signals of selection in all the species in Table 2. Among all the domesticated animals examined here, the goat is perhaps the most conspicuous absence in terms of signals of selection of glutamate receptor genes. Despite this, alterations to glutamatergic signaling seem to have played an important role in the goat domestication process. In a recent study of convergent signals of selection on domesticated goats and sheep, Neurobeachin (*NBEA*), a gene that regulates glutamate and GABA receptor expression at synapses, was highlighted as being one of the most likely genes to be implicated in behavioral changes under domestication [17].

	AMH [18, 11, 19, 20, 12, 21]	Dog [22, 23, 24, 25, 26]	Cattle [27]	Cat [28]	Horse [29]	Fox [30, 31]	Yak [32, 33]	Sheep [34]	Pig [35, 36, 37]	Rabbit [38]	Guinea Pig [39]	Chicken [40]	Duck [41]
GRIN1													
GRIN2A							▲						
GRIN2B						■	▲						
GRIN2C	■												
GRIN2D						◆							
GRIN3A							▲						●
GRIN3B													
GRIA1				●					●				
GRIA2				●									
GRIA3													
GRIA4							▲						
GRIK1									●				
GRIK2		●	◆				▲			●			●
GRIK3	●	●	●	●	◆		▲	●					
GRIK4	■												
GRIK5	●	■											
GRM1												●	
GRM2	●	■											
GRM3	●					■	◆						
GRM4							▲				◆		
GRM5													
GRM6	■					■							
GRM7	■								●				
GRM8	(●)	●						●	●				
GRID1					●								
GRID2	●												

Table 2: Signals of selection, high-frequency changes, introgression, and differential expression of glutamate receptor genes in AMH and domesticated species.

- Selective sweep study identifying differences between AMH and archaics or domesticates and their wild ancestors
- (●) Selective sweep study identifying signals of selection in AMH (not relative to archaics as an outgroup)
- High frequency changes differentiating AMH and archaics or domesticates and their wild ancestors
- ◆ Differential brain expression between domesticated species and wild ancestors
- ▲ Introgression study

Overlapping signals of selection were most consistently detected on kainate and Group II and III metabotropic receptor genes across modern human and animal domestication studies (see Table 2 and Figure 1).

Out of the twenty-eight instances where differences were detected on functioning ionotropic receptor genes (NMDA, AMPA, or kainate), seventeen were detected among the kainate receptor genes, and thirteen of these occurred on either on *GRIK2* or *GRIK3*. *GRIK3* exhibits signals of selection in modern humans, dogs, cattle, sheep, and of introgression in yaks, while *GRIK2* shows signals of selection in dogs, rabbits, and ducks, and of introgression in yaks.

Metabotropic receptors are the other major subclass of glutamate receptor genes that display consistently convergent signals among domesticated species and modern humans, with members of Group II (*GRM2* and *GRM3*) and, in particular, Group III (*GRM4*, *GRM6*, *GRM7*, and *GRM8*) exhibiting signals of selection across domesticate and human selective-sweep studies. Signals on *GRM8* have been detected in dogs, sheep, pigs, and humans, although the human study did not have archaics as an outgroup [21].

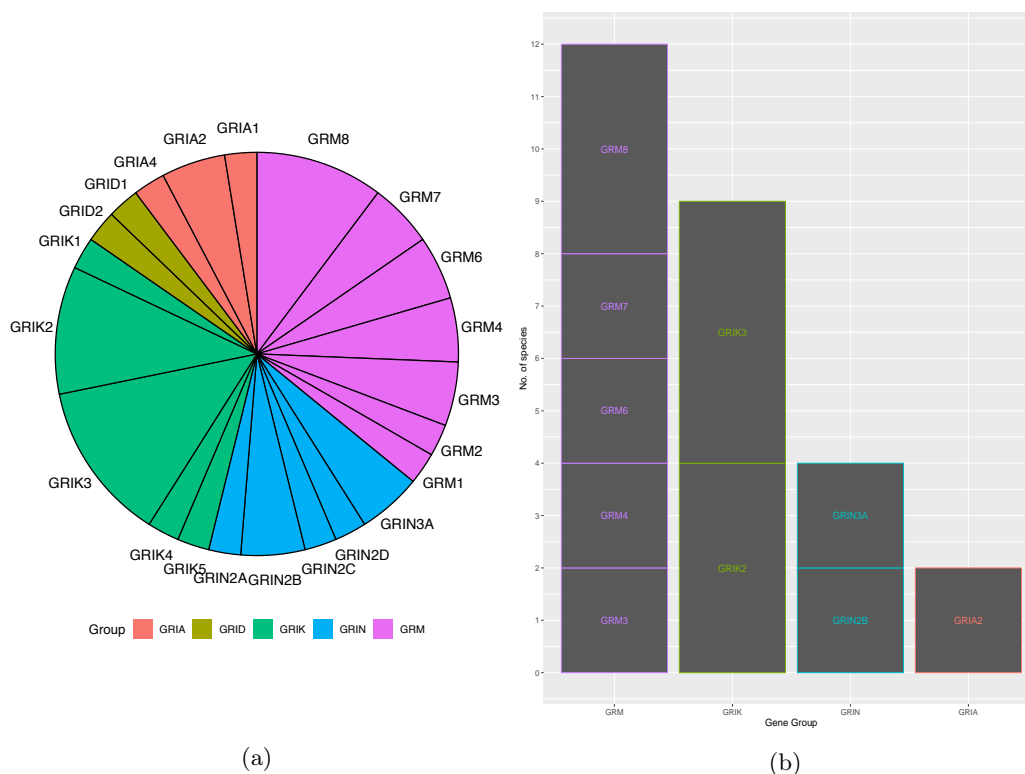


Figure 1: Signals of selection on glutamate receptor genes, plotted by gene group. (a): Number of species for which signals of selection are detected on at least one gene. (b): Number of species for which potential signals of convergent selection are detected in at least two species per gene.

Relative to Neanderthals and Denisovans, modern humans show signals of selection on *GRM2* and *GRM3* (this latter gene detected on the KhoeSan terminal branch of our lineage). Together, *GRM2* and *GRM3* make up the Group II metabotropic receptor gene subfamily.

Fifteen out of the sixteen instances where signals of selection, expression changes, high-frequency missense changes, or introgression were detected on metabotropic glutamate receptor genes, these occurred in Group II or Group III subfamilies. The only exception to this pattern is the detection of a selective sweep on *GRM1* in the chicken. Group II and III metabotropic receptors share structural and functional similarities, which differentiate them from Group I receptors. Perhaps the most striking difference is that Group II and III receptors primarily inhibit adenylyl-cyclase signaling, while Group I receptors potentiate this signaling cascade [42]. We discuss the differences between metabotropic receptor subfamilies further below.

In several of the studies cited in Table 2, signals of selection on glutamate receptor genes have been suggested as potentially important for behavioral changes during the domestication process [23, 27, 28, 29, 30, 36, 41]. However, such suggestions form part of broader discussion on changes to genes related to the central nervous system (CNS) under domestication, and no mechanistic details are provided. Furthermore, discussion is usually limited to highlighting the potential importance of glutamatergic-signaling changes in learning, memory, or excitatory transmission in a single domesticated species under study, and tends to focus on cortical regions.

To our knowledge, no study to date has sought to explore the extent to which glutamate receptor genes come under selection across a large number of domestication studies. More significantly, we are unaware of any previous study that has identified kainate and Group II and III metabotropic receptor genes as the predominant targets of selective sweeps under domestication and in recent human evolution. The consistency with which these receptor genes are identified across domestication studies makes them prime candidates for being involved in the emergence of tameness.

If an argument is to be made for the convergent involvement of kainate and metabotropic glutamate receptor genes in domesticated tameness and human prosociality, evidence should point to the shared participation of these receptors in modulating the stress response across these species. Table 3 summarizes evidence from human, model organism, and animal gene-behavior-correlation studies that these kainate and metabotropic receptors are implicated in developmental, neuropsychiatric, stress, and mood disorders, as well as divergent tame and agonistic phenotypes in non-human species. (Detailed discussion of these studies can be found in Supplementary section S2.)

GENE	Associated Human phenotypes									Tame and aggressive animal phenotypes
	DD/ID	ADHD	ASD	SCZ	BPD	OCD	ANX	MDD	SR/Fear	
GRIK2	*	*	*	*	*	*	*	*	*	Retention of dog-like polymorphisms on GRIK2 in tame Czechoslovakian Wolf-Dog hybrid [43]
GRIK3	*		*	*	*		*	*	*	Signals of selection on GRIK3 differentiating cattle with agonistic and tame behaviors [44]
GRIK5			*	*	*		*			
GRM2		*	*	*		*	*	*	*	
GRM3		*	*	*	*	*	*	*	*	Fixed missense mutation on GRM3 differentiating aggressive from tame foxes [30]
GRM4		*	*	*	*		*	*	*	
GRM7		*	*	*	*	*	*	*	*	Lower nucleotide variability on GRM7 in docile Apennine Brown Bear [45]
GRM8	*	*	*	*			*	*	*	Lower nucleotide variability on GRM8 in docile Apennine Brown Bear [45]

Table 3: Human and Domesticated phenotypes associated with kainate and metabotropic receptor genes. Detailed discussion of these associations, including references can be found in Supplementary section S2. DD/ID - Developmental Delay/Intellectual Disability, ADHD - Attention Deficit Hyperactivity Disorder, ASD - Autism Spectrum Disorder, SCZ - Schizophrenia, BPD - Bipolar Disorder, OCD - Obsessive Compulsive Disorder, ANX - Anxiety Disorder, MDD - Major Depression, SR/Fear - Startle Response/Fear.

Note: Widespread use of non-selective Group II agonists that act on both GRM2 and GRM3 subunits in model organisms makes it difficult to always separate subunit associations with disorders. Where a disorder has been associated with Group II receptors, an asterisk is placed in the row for both GRM2 and GRM3.

Schizophrenia is among the disorders most regularly associated with mutations to the metabotropic and kainate receptor genes that we have reviewed here. In humans, prenatal stress is a risk factor for the development of schizophrenia in adult offspring [46, 47]. Pharmacological agonists of Group II metabotropic receptors reduce schizophrenia-like phenotypes in adult offspring of prenatally stressed mice [48]. Furthermore, high glucocorticoid inputs to the hippocampus reduce the expression of kainate receptors, and schizophrenics have been found express significantly reduced kainate receptors in this region [49, 50]. More broadly, heightened stress experienced during pregnancy can lead to a “persistently hyperactive” HPA axis in offspring, increasing children’s propensity to develop Attention Deficit Hyperactivity Disorder (ADHD) and adult anxiety and reactivity to stress, while in rats, prenatal stress decreases the propensity to play in juvenile offspring and impairs sociality and extinction of conditioned fear lasting into adulthood [51, 52, 53].

Prenatal stress can, then, contribute to the emergence of the same neurodevelopmental, neuropsychiatric, stress, and mood disorders commonly associated with altered expression of metabotropic and kainate receptor genes. We hypothesize that selective sweeps on these genes are markers of convergent positive selection on an attenuated stress response in both archaic humans and domesticated species. We propose that enhanced prenatal modulation by these receptors of stress responses to human contact in (pre-)domesticated and archaic human females provided an important first step in the emergence of tameness and prosociality. In section 3, we explore the neurobiological evidence for this proposal, focusing on the roles of kainate and metabotropic glutamate receptors in the stress-response cascade.

3 Discussion

Alterations to the HPA axis are considered to be essential for the emergence of tameness in different (indeed competing) theories of domestication [16, 54]. Glutamatergic signaling acts as a prominent regulator of HPA activity and has been identified among the top enriched pathways across studies of aggression [6, 55, 56, 57]. The association of kainate and Group II/III metabotropic receptor genes with multiple stress disorders implicates them in altered HPA-axis activity. Given the predominantly modulatory, as opposed to excitatory, functions of both kainate and Group II and III metabotropic receptors [42, 58], we argue that selective sweeps on their respective genes are markers of decreased HPA reactivity in humans and (pre-)domesticated species.

Below, we propose a mechanism for how these receptor subfamilies modulate glutamatergic signaling to alter developmental trajectories and, subsequently, the HPA stress response in both humans and domesticated species. Our argument relies on three pieces of evidence: First, evidence that alterations to the HPA axis are common across domesticated species versus their wild counterparts, and in non-reactive versus reactively aggressive humans; second, evidence that the genes highlighted here are extensively expressed in limbic and

hypothalamic brain regions crucial for controlling the stress response; and third, evidence that disturbance of this expression alters the stress response.

Alterations to the stress response in domesticated species and modern humans

In response to stress, corticotrophin releasing hormone (CRH) is synthesized in the paraventricular nucleus (PVN) of the hypothalamus. This induces adrenocorticotrophin (ACTH) release from the anterior pituitary gland, which, in turn, stimulates the release of glucocorticoids (GCs: primarily cortisol and corticosterone) from the adrenal gland [59]. GCs are “the principal end-products of the HPA axis”, which help to maintain homeostatic balance in the organism [60]. They also provide feedback directly to neurons in the PVN [13, 61], or via other brain regions, particularly limbic structures, including the hippocampus, thereby modulating CRH release and the HPA stress response [62, 60]. Thus, GC measures can be an accurate indicator of stress-response in vertebrates, once basal and stress-response measures can be differentiated [63].

Domesticated foxes, sheep, bengalese finches, and ducks have lower basal GC levels than their wild ancestors or other closely related wild comparators [64, 65, 66, 67, 68, 69, 70]. In the duck and the fox, differences have been shown to be particularly marked in prenatal and juvenile development, respectively [69, 70, 64]. Compared to their wild ancestors, Guinea pigs and chickens have a lower spike in GCs in response to stress [71, 72]. Although there is no extant ancestral comparator of neuroendocrine function in AMH, our species has considerably lower basal plasma cortisol levels than chimpanzees and most other primates [73].

Within the human population, variability in GC levels correlate with different individual stress responses, which mirrors findings in laboratory rats. Acute GC increases accompany bouts of reactive aggression, while chronically high basal levels have been found to correlate with increased anxiety and major depression, and may be implicated in reduced aggressive tendencies [14]. Chronically low GC levels can correlate with antisocial personality disorder, callous, unemotional tendencies, and externalizing behaviors in children, as well as aggressive delinquency in adults. Proactively aggressive or non-aggressive children tend to have a lower spike in GC levels in response to frustrating tasks than reactively aggressive children [74]. Psychopathic adults (who often exhibit pathological proactive aggression) tend to have no cortisol reactivity to frustrating tasks [14].

The above studies suggest that, from early development into adulthood, lower basal GC levels are shared by domesticates and modern humans relative to closely related extant wild species. Moreover decreased GC spikes in response to stress are common to domesticates and non-aggressive or proactively aggressive modern humans versus reactively aggressive individuals. These findings are consistent with the view that prosocial selective pressures have led to a reduction in reactive over and above proactive aggression in recent human evolution [1]. It could be considered that proactive aggressors within modern human populations

exhibit a pathological version of the non-reactive phenotype that has been under positive selection and is associated with HPA-axis hypofunction under stress.

Kainate and metabotropic receptor expression in brain regions crucial for HPA regulation

The HPA axis is centrally regulated by the limbic system, primarily through amygdalar processing of perceptual inputs, which are relayed via the bed nucleus of the stria terminalis (BNST) to the paraventricular nucleus (PVN) in the hypothalamus. The limbic system also mediates feedback mechanisms, whereby glucocorticoids and mineralocorticoids act upon receptors in the hippocampus and medial prefrontal cortex, which connect to the PVN via the BNST and lateral septum. Feedback also occurs directly on cells in the PVN to modulate HPA reactivity. Feedforward mechanisms, further potentiating the stress response, are relayed from the amygdala to the PVN via the BNST. [75, 62, 61].

Glutamatergic and GABAergic signaling are the central mediators of each of these aspects of HPA (re)activity, and the kainate and metabotropic receptor subfamilies discussed here play prominent roles in modulating release of both neurotransmitters. These receptors are extensively expressed in limbic regions crucial for modulating the stress response. Figure 2 highlights these expression patterns. A detailed overview of what is known about kainate and Group II and III metabotropic receptor expression in the developing and adult brain can be found in Supplementary section S3. In the subsection that follows, we propose a mechanism by which metabotropic and kainate receptors modulate HPA activity.

Control of HPA function by kainate and metabotropic glutamate receptors

The PVN is the crucial hypothalamic mediator of psychogenic stressors that drive HPA activity. Glutamate acting directly on parvocellular neurons of the PVN stimulates CRH release, whereas GABA inhibits this [6]. This means that modulation of glutamatergic signaling by both kainate and metabotropic glutamate receptors may serve to inhibit direct activation of the PVN.

GRIK2, *GRIK3*, and *GRIK5* are all expressed in the PVN and surrounding regions in adult rats, although *GRIK1* is the most abundant kainate receptor in the PVN proper [76]. *GRIK5* is extensively expressed on parvocellular neurons [77]. Presynaptic activation of *GRIK1* subunits in the PVN has been shown to modulate HPA activity by inhibiting CRH from parvocellular neurons [55]. Similarly, agonism of presynaptic kainate receptors in hypothalamic neurons facilitates inhibitory GABAergic signaling [78].

In vitro antagonism of Group II metabotropic receptors in hypothalamic slices has been shown to increase CRH signaling, whereas no other metabotropic receptor agonists or antagonists had this effect. Mice administered with Group II antagonists *in vivo* experienced an increase in corticosterone that mimicked

Kainate (KAR) and Metabotropic Receptor (mGluR) Expression in Limbic and Stress Response Networks

Expression Key

- Low - Moderate mGluR
- Abundant mGluR
- Low - Moderate KAR
- Abundant KAR
- PreN Prenatal
- PosN Postnatal

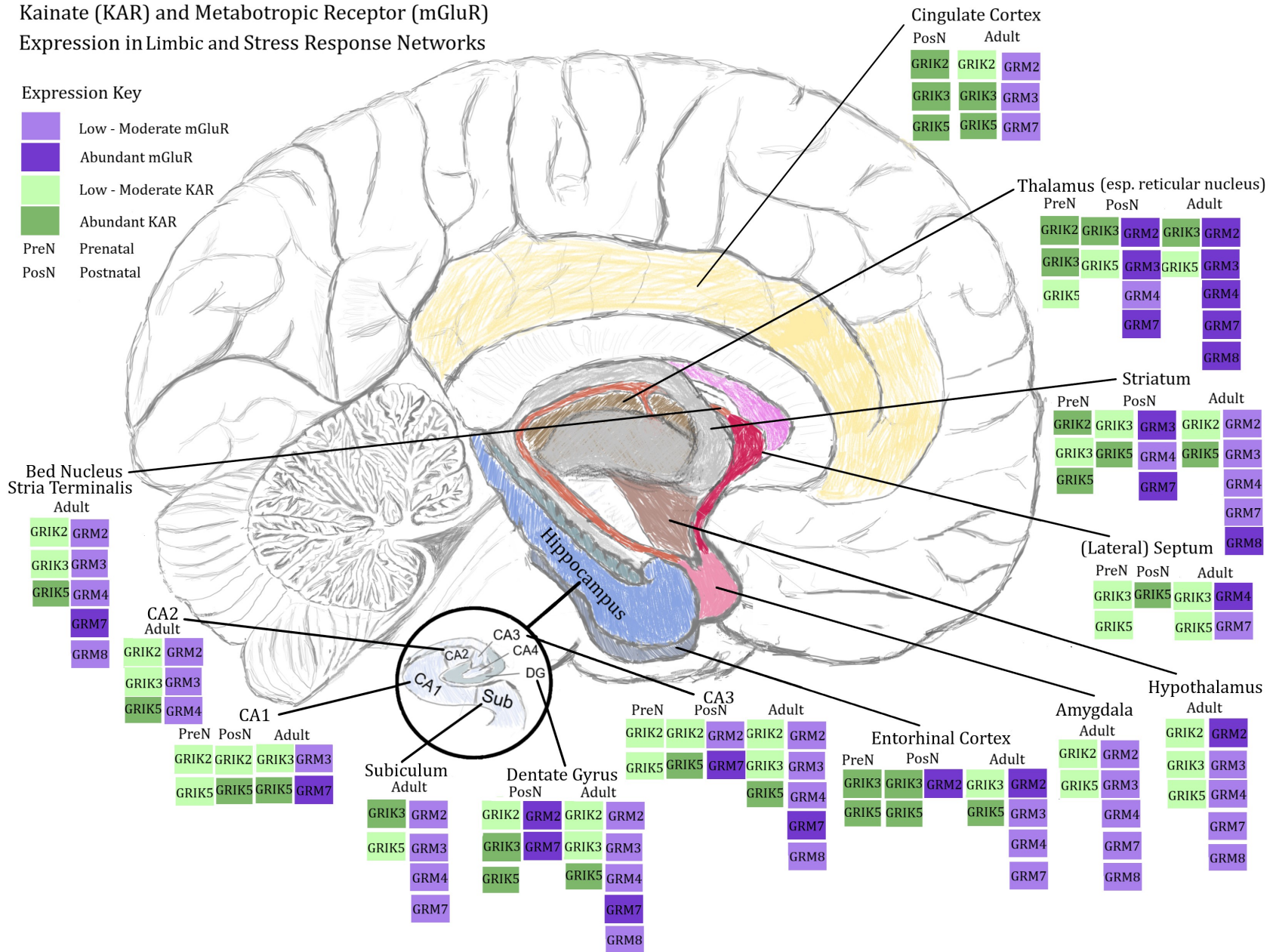


Figure 2: Kainate and metabotropic receptor expression in brain regions crucial for HPA regulation. (Most detailed brain-expression data come from rodent studies. Where available, we have used data from human fetal or postmortem studies. Broadly, there are cross-species parallels in kainate and metabotropic expression. Similarities and differences are discussed in Supplementary section S3.)

the response to the forced-swim (behavioral despair) test [79]. Given the predominant presynaptic and glial modulatory functions of Group II receptors, the above evidence implicates Group II receptors in the attenuation of central glutamatergic inputs to the hypothalamus (likely in the PVN), attenuating the HPA stress response. Group III metabotropic receptors have also been implicated in modulating excitatory inputs to the lateral hypothalamus [80], while Group I metabotropic receptors stimulate both oxytocin and vasopressin release from the SON [81].

Agonism of Group II metabotropic receptors disrupts the fear-potentiated startle response in mice, suggesting that they regulate the learning of fearful experiences [82]. Knockout of *GRM2* has been shown to correlate with increased stress in social interactions [83]. In macaques captured from the wild, six-week chronic intravenous administration of a Group II agonist reduced basal cortisol levels by as much as 50% compared to controls [84]. This same agonist has been shown to act on GRM3 receptors in adrenal gland cells, leading to a reduction in aldosterone and cortisol via inhibition of the adenylyl cyclase / cAMP signaling pathway [85]. Yet another Group II agonist attenuates aggressive tendencies, hyperactivity, and deficits in the inhibition of the startle response of mice reared in isolation [86].

It has been proposed that Group II metabotropic receptors in the central amygdala dampen the stress response by modulating the release of glutamate, in turn leading to an increase in GABAergic signaling and overall dampening of excitatory inputs to the PVN. Agonism of these receptors also leads to an increase in activity in the predominantly inhibitory BNST, and in the PVN in response to stress, suggesting modulation of the HPA by suppression of excitatory signaling. At the same time, activation of Group II receptors is downregulated in the hippocampus [87]. Excitatory feedback outputs from the hippocampus likely act on inhibitory neurons of the BNST that, in turn, relay to the hypothalamus [75]. Thus, decreased Group II modulation of these outputs in response to stress can have the effect of enabling increases of inhibitory signals to dampen the HPA cascade. Within the BNST itself, activation of Group II and III receptors has been shown to suppress excitatory transmission [88].

Mice lacking the Group III receptor *GRM8* display increased age- and sex-dependent anxiety-like behaviors and startle response [89, 90]. However, in contrast to *GRM2*, knockout of *GRM8* can enhance social interactions, suggesting that this receptor has opposing effects depending on the nature of the stressor. In another contrast with *GRM2* and the Group II subfamily as a whole, ablation of *GRM7* can make mice less fearful and less aggressive [91, 92, 93] correlating with a severe reduction in neuronal activity in the BNST. This suggests that GRM7 serves to enhance overall excitability of BNST neurons projecting to the PVN (and thus the stress response), perhaps through modulation of glutamatergic release innervating GABA inhibitory interneurons [93]. Activation of GRM4 reduces anxiety-like behaviors in mice, while knockout enhances fear-conditioning responses and increases anxiety in adult but not juvenile mice [94]. Such anxiety-related effects are thought to be brought about by alterations to amygdalar function, whereby either excitatory or inhibitory signaling is modu-

lated by Group III receptors.

Although the above behavioral correlates of Group II and III receptor (ant-)agonism and ablation are partially contrasting, they indicate that both sub-families are important for modulating the stress response, including aggressive reactivity. This tendency is clear in Group II metabotropic receptors, while the actions of Group III receptors are more varied according to the specific subunits and brain regions activated. Studies in rodents suggest that GRM8 and GRM7 have broadly opposite effects on anxiety levels, with GRM8 activation tending to be closer to Group II metabotropic receptors in its anxiolytic effects, while GRM7 seems to be more anxiety- (and aggression-) inducing [95]. This suggests that the numerous signals of selection on *GRM8* across domesticates and similar signals on Group II receptors in humans may be markers of convergent selection for a decreased stress response. This said, evidence also points more clearly towards activation of Group II receptors in potentiating prosocial behaviors. Future investigation of differences in brain region expression of Group II and III receptors in domesticated species may help to shed more light on their contributions of each to the modulation of the stress response.

The kainate receptors we have examined here are abundantly expressed in limbic regions that modulate HPA-axis function via glucocorticoid (GC) feedback (in particular the hippocampus and medial prefrontal cortex, but also more moderately in the amygdala [see Figure 2]). GCs promote glutamate release in these feedback regions, and the different affinities of mineralocorticoid receptors (MRs; bound by GCs at low concentrations) and glucocorticoid receptors (GRs; bound at higher concentrations) enable the modulation of stress feedback signaling from basal or moderate to acute levels [60, 61].

Glucocorticoids differentially modulate the expression of kainate receptor mRNA in the hippocampus depending on whether MRs or GRs are bound [96, 49]. Adrenalectomy (lowering corticosteroid levels) leads to increased expression of *GRIK2* in DG and CA3, and of *GRIK3* in DG [49] (although no change has also been reported for *GRIK2* in CA3 [96]). Single dose treatment with low levels of corticosterone following adrenalectomy — thought to bind MRs — increases *GRIK3* and high affinity subunit (*GRIK4* and *GRIK5*) mRNA in DG, as well as *GRIK5* across the hippocampus [96]. MR binding has been reported both to lower and raise *GRIK2* levels in the hippocampus [96, 49].

Acute corticosterone treatment in rats lowers kainate receptor mRNA expression to levels of untreated controls [96]. Similarly, chronic treatment leads to lower expression of *GRIK3* and *GRIK4* in hippocampal structures, although no changes were noted for *GRIK2* or *GRIK5* [49]. These divergent MR/GR-mediated patterns of expression can help to elucidate the mechanism by which the genes under selection in the domesticates and humans are expressed in a manner that can modulate the stress-induced feedback response.

Kainate receptor activation at CA3-CA1 synapses serves to inhibit glutamate transmission via $G_{i/o}$ signaling, especially when synapses are immature. At mossy fiber synapses connecting DG and CA3 (areas of high kainate receptor expression throughout life), kainate receptors inhibit glutamatergic signaling when glutamate is released at high levels, while facilitating release at lower lev-

els, again via a $G_{i/o}$ -coupled mechanism [97]. Similar biphasic modulation has been detected in the neocortex and amygdalae of rodents. Thus, when GCs are circulating at low levels, during basal or low stress, MR binding should lead to higher kainate receptor expression and facilitation of glutamatergic signaling in feedback regions. At higher GC levels (as when under acute stress) GR binding will tend to reduce kainate receptor expression, thus diminishing these receptors' potency in modulating glutamate release. In contrast, postsynaptic expression of AMPA and NMDA is enhanced under acute stress and corticosterone treatment [98].

Because there are no direct hippocampal, prefrontal, or amygdalar connections to the PVN, feedback from these regions are instead relayed via the BNST, lateral septum, and ventromedial hypothalamus (VMH), which are all predominantly GABAergic [99, 6]. For the amygdala, which emits primarily inhibitory outputs to intermediary regions, this results in "GABA-GABA disinhibitory" downstream signals, increasing excitatory inputs to the PVN [62]. In the case of kainate receptor expression, which is predominant in the hippocampus and medial prefrontal cortex, increased facilitation of glutamate release during basal or low-level stress is likely to primarily innervate GABAergic neurons along pathways relaying to the PVN. Similarly, downregulation of kainate receptor expression by GR binding during acute stress serves to diminish the alternate modulatory effect of kainate receptors during intense glutamatergic release. This, in turn, should allow for NMDA and AMPA receptor signaling to be potentiated, leading to a dampening of the HPA stress response, again via the innervation of inhibitory neurons of the BNST, septum and VMH, which relay to the PVN.

In contrast to prefrontal and hippocampal feedback regions, the effect of glucocorticoids on parvocellular and magnocellular neurons of the hypothalamus is to downregulate glutamatergic signaling via the release of endocannabinoids, in turn promoting the release of GABA [100]. Kainate receptors have been implicated in the mobilization of endocannabinoid signaling in distinct brain regions, as well as in the promotion of GABAergic signaling in the PVN [55, 101, 102].

Figure 3 presents a schema of the modulatory actions of metabotropic and kainate receptors in the stress-response cascade.

Increased metabotropic and kainate-mediated attenuation of central and feedback stress responses may plausibly have conferred selective advantages in human evolution, not only via the reduction of stress and enabling of prosocial cooperation, but also by enabling subsequent increases in *GRIK2*, *GRIK3*, and *GRIK5* expression: Firstly, Group II and III metabotropic glutamate receptor modulation of amygdalar fear processing in response to stressors, in combination with kainate and Group II metabotropic receptor inhibition of CRH release in the PVN can lead to a signaling cascade that results in lower glucocorticoid feedback in limbic structures. This, in turn may lead to increased expression of kainate receptors in the hippocampus, prefrontal cortex, and elsewhere, perhaps enabling subsequent selection on improvements in plasticity and learning, as well as further resources for limbic modulation of the stress response.

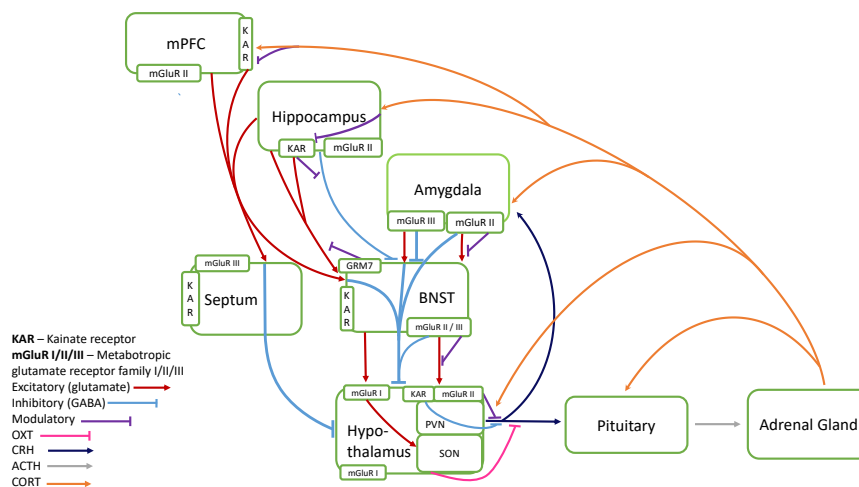


Figure 3: Modulatory Actions of Metabotropic and Kainate Receptors

4 Conclusion

We have argued here that, on balance, selective pressures have led to a modulation of glutamatergic signaling in order to attenuate the HPA stress response, and that this has had the corollary effect of increasing synaptic plasticity in limbic feedback regions that play a crucial role in memory and learning. We have not dealt in detail with G-protein signaling cascades activated by kainate, Group II, and Group III receptors. A more in-depth view of convergence may be gained by exploring the extent to which the same signaling cascades (in particular $G_{i/o}$ signaling) are activated in homologous brain regions across species regardless of the receptor subtype initially bound.

Given the important roles that serotonin and oxytocin play in promoting social and empathetic behaviors across different species, it has been proposed that convergent tameness in domesticates and prosociality in modern humans is driven by alterations to these systems [2]. The present analysis of modern human-archaic and domesticate-wild differences in glutamatergic receptor genes suggests that any such modifications (particularly to oxytocinergic signaling) are more likely to be dependent on upstream changes to glutamatergic signaling. Glutamate mediates the release of oxytocin and vasopressin from the SON and PVN [103].

Serotonin modulates glutamatergic activity in the brain, often inhibiting excitatory potentials and stimulating GABAergic inhibitory signaling [104]. This regulation from outside the glutamatergic system could potentially produce comparable modulation of the stress-response to the regulation from within that we propose for metabotropic and kainate receptors. In studies of genetic differences between tame and aggressive foxes, changes to serotonergic signaling accompany those of the glutamatergic system, with genetic changes often po-

tentially relevant to both synapses [30, 31]. Similarly, domestication of the pig appears to have involved important changes across both systems [36]. Signals of selection associated with serotonergic signaling have also been identified in a tame strain of rat and the domesticated goat [105, 106].

One may reasonably ask whether Group II and III metabotropic and kainate receptors share functional or structural qualities that have made them more likely to come under selection than NMDA, AMPA, or Group I metabotropic receptor (sub)families in domesticate species and modern humans. These receptors, particularly NMDA, have been implicated in many disorders and phenotypes reviewed here for metabotropic and kainate receptors [107, 108, 109, 110, 111, 112, 113, 56]. Even within receptor families, specific subunits (and combinations of them) or splice variants can have diametrically opposing functional properties from others that are nominally similar [114, 115].

The similar modulatory properties, overlapping expression patterns, and shared phenotypical associations of those genes that most consistently detected appear across domestication and modern human studies has led us to hypothesize an overarching role for kainate and Group II and III metabotropic receptors in the modulation of the stress response. However, there is no reason, in principle, why other glutamate receptor families could not have contributed to the emergence of tame behaviors. In fact, signals of selection on NMDA receptors are detected for two domesticate species on each of *GRIN2B* (fox and yak) and *GRIN3A* (yak and duck, see Table 2). Both of these genes' respective receptors are associated with the maintenance of immature dendritic spines and impeding long-term potentiation (LTP) during development: GRIN2B subunits are highest expressed during embryonic development and are gradually replaced at the synapse by GRIN2A as spines mature [108]. Similarly, when *GRIN3A* is genetically deleted, the maturation of spines is sped up, and GRIN2A and GRIA1 subunits — typically enriched at adult synapses — are recruited to the membrane earlier [116]. If similar patterns of selection should be detected on NMDA receptors across other domesticates, a case could be feasibly be made for their contributing to the juvenile cognitive phenotype typically retained by domesticate species into adulthood. On the other hand, the only case of potentially convergent selection on an AMPA receptor gene occurs on *GRIA2* (cat and pig), which is a crucial AMPA subunit at mature synapses [117].

The diversity of roles played by kainate receptors in the CNS, including extensive excitatory, inhibitory, ionotropic, metabotropic, and modulatory activities [58], provides a wide range of possible functions upon which natural selection can act. Selective pressures seem to have honed varied specializations for kainate receptors in different brain regions. Group II and III metabotropic receptor subunits can have opposing actions within a single brain region, and to stressful stimuli being processed. This suggests that these subunits often have complementary modulatory functions within their subfamilies. Nonetheless, there are considerable overlaps of function, and these subunits primarily inhibit adenylyl-cyclase signaling. Each receptor subfamily acts, broadly, to dampen stress responses. These overlaps in function undoubtedly contribute to numerous signals of selection being spread across different metabotropic recep-

tor genes in distinct species.

In the future, it may be worthwhile examining the extent to which the glutamatergic changes discussed here could have impacted the vocal abilities of the relevant species, including ours. Although the extent of archaic humans' vocal-learning abilities is not known, this capacity is highly striatum-dependent, as can be seen in the neurology of Tourette's syndrome. Glutamatergic signaling alterations in the striatum have been implicated in Tourette's syndrome, including genes with AMH-specific changes. Vocal learning deficits in humans who carry a mutation on the *FOXP2* gene are thought to arise, in part, from abnormalities in the striatum [118]. Knock-in experiments of the humanized *FOXP2* allele in mice have shown the principal structural changes to take place in the striatum, where MSN dendrites are longer and respond to stimulation with increased long term depression [119]. *FOXP2* is also expressed in glutamatergic projection neurons of the motor cortex and is highly expressed during the development of corticostriatal and olivocerebellar circuits, important for motor control [119, 120]. Several glutamate receptor genes discussed above, such as *GRIK2* and *GRM8*, have been identified as a transcriptional target of *FOXP2* in the developing human brain [121]. Significant changes to glutamatergic expression have also been found in the vocal nuclei of vocal-learning birds, and in the domesticated bengalese finch compared to its wild ancestor the white-rumped munia [8, 122, 123]. These changes are thought to play a role in the more complex singing capabilities of the bengalese finch. Glutamate receptor genes form part of the most highly co-expressed module in the periaqueductal gray of bats, a mammalian species that displays strong evidence of vocal-learning capabilities [124].

Finally, an important question arising from the evidence presented above is how glutamatergic involvement in the modulation of the stress response relates to the hypofunction of the neural crest, proposed to account for the suite of phenotypic changes that make up the domestication syndrome [16]. At this point, the evidence from selective sweep studies points more unequivocally to the involvement of glutamatergic signaling in domestication than to hypofunction of the neural crest, in line with other arguments against the universality of the effects predicted by the neural crest hypothesis [125]. We argue that changes to glutamatergic signaling are central to the unifying domesticated trait of tameness and that changes to NCC proliferation are subsequent to this. There are various possible explanations for how glutamatergic signaling may interact with, or even bypass, the neural crest to bring about the broader phenotypical features of the domestication syndrome. We present two alternatives here that future work could put to the test.

One possibility is that modulation of glutamatergic signaling attenuates HPA-axis signaling, and ultimately glucocorticoid output, from the adrenal cortex in (pre)domesticated females during pregnancy. Modified hormonal concentrations would then affect embryonic development, altering neural crest cell inputs to different tissues prenatally. In cell cultures, glucocorticoids are essential for NCC survival and differentiation [126]. An increase in corticosteroid concentrations has been shown to increase NCC numbers and alter cell fates,

for example converting small intensely fluorescent cells, which are normally dopaminergic, into epinephrine-synthesizing cells. Removal of corticosteroids and replacement with nerve growth factor converts small intensely fluorescent cells and chromaffin cells into sympathetic neurons [127, 126]. Mice embryos lacking the glucocorticoid receptor cannot survive outside the womb due to widespread defects, including severe reductions in chromaffin cells from early development. Those cells remaining lose their ability to synthesize epinephrine [128]. The above studies suggest that lowered stress hormone levels in the womb, resulting from HPA hypofunction in the mother can have the knock-on effect of bringing about neural crest hypofunction and cell-differentiation changes in embryonic development. Under our hypothesis, mild reductions in NCC inputs are driven by increased glutamatergic modulation of the stress response during pregnancy resulting from selection for tameness.

As mentioned above, corticosterone treatment during embryonic development of domesticated ducks can bring about mallard-like behaviors in neonates [69, 70]. Glucocorticoid levels are lower during gestation in both tame rats and foxes compared with aggressive strains [129]. Suppression of glucocorticoid levels in pregnant aggressive females between days 12 and 14 of gestation leads to a concomitant reduction in embryonic glucocorticoids by day 20. This, in turn, leads to significant increases in depigmentation in neonate rats [129]. In both the rat and the duck studies, the effects of glucocorticoids on embryonic development take place long after neurulation and migration of neural crest cells away from the neural tube. This suggests that at least some of the most important phenotypical features of domestication need not depend on genetic changes to NCC development or migration, but, rather, on postmigratory regulation of expression at different tissue sites by glucocorticoids.

An alternative possibility is that alterations to glutamate receptor signaling can directly account for both behavioral and physical changes in domestication. Some evidence for this derives from the fact that glutamate receptor, transporter, and other signaling genes identified in different domestication studies are expressed in melanocytes, osteoblasts, osteoclasts, chondrocytes, and other tissues beyond the CNS [130, 131, 132, 133, 134]. Thus, traits such as floppy ears, shortened snouts, or depigmentation could feasibly result from alterations to glutamatergic receptor expression in these tissues. Bone is innervated by glutamatergic fibers and glutamate receptors are thought to play a role in osteoblastic differentiation and proliferation [135, 136]. *GRIK5*, *GRM4*, and *GRM8* along with NMDA and AMPA receptor genes are expressed *in vitro* in developing rat calvarial osteoblasts [137, 138]. Group II and III metabotropic receptors have been shown to prevent mineralization of chondrocytes, implicating them in cartilage development [130, 134]. NMDA receptors have been implicated in the maturation and differentiation of chondrocytes [139]. Glutamate receptor genes (among them *GRIK3*) are expressed on melanocytes, and inhibition of AMPA-mediated excitatory transmission has been found to decrease the expression of *MITF* (melanogenesis associated transcription factor) [131].

On considering these two alternative accounts of glutamatergic signaling in the emergence of the domestication syndrome (NCC-interacting versus NCC-

independent), we prefer to be cautious about attributing too many functional consequences to the signals of selection highlighted here. We consider that a mild neurocristopathy will almost certainly explain some physical trait changes in domesticated species, and that this may serve to entrench earlier selection for tameness via reduced inputs to stress-hormone cells in the adrenal glands. However, we predict that these reductions will most often result from upstream regulatory changes by glutamatergic signaling affecting stress responses. In some species, changes to glutamatergic signaling may also have acted to directly alter the development of physical traits, independently of genetic or epigenetic alterations to the neural crest, driven by domestication.

5 Materials and Methods

Based on our previous work on overlapping signals of selection on glutamatergic-signaling genes in domesticates [3], we extended our comparative analysis across a broad range of domesticated species. In order to delimit the comparison to a clearly-defined gene set within the glutamatergic system, we selected the 26 glutamate receptor genes as our cross-species comparator (Table 1).

For cross-species comparison, we included all domesticated species for which whole-genome sequences were available. Given the absence of such data for the domesticated camel, we excluded this species from our comparison. In species for which no studies of selective sweeps were available, we included studies of decreased heterozygosity differentiating tame and aggressive strains (fox) and/or studies detailing brain-expression differences between tame and wild or aggressive lineages (guinea pig, fox, and rat). We also included a study of introgressed genes from cattle to yaks, which identified genes likely to have been subject to positive selection. Finally, we included for comparison high-frequency (near-fixed) changes differentiating modern from archaic humans. (Table 2.)

We extracted all instances of selective sweeps, introgression, brain-expression differences, and high-frequency changes on glutamate receptor genes from the domestication and modern-human studies listed in Table 2. We compiled these genes for comparison according to the receptor (sub)families of which they are members.

In order to determine potentially shared functions of kainate and metabotropic receptor genes showing the strongest signals of convergence across humans and domesticates, we investigated associations with phenotypes relevant to tameness and prosociality by doing an exhaustive literature search (Pubmed). These included associations with the startle response and neurodevelopmental, neuropsychiatric, stress, and mood disorders, summarized in Table 3.

Finally, we carried out a meta-analysis (summarized in Figure 2, details in Supplementary Material) to determine the brain expression of kainate and Group II and III metabotropic receptor genes.

Author contributions

Data collection and analysis: TOR, CB; proposed mechanism: TOR; draft preparation: TOR, with input and revisions from CB; project supervision: CB.

Acknowledgements

We would like to thank Pedro Tiago Martins for invaluable help in preparing Tables 1 and 2, and Figure 1. We would also like to thank Stefanie Sturm, Alejandro Andirkó, and Pedro Tiago Martins for helpful comments on earlier drafts of this text.

Funding statement

CB acknowledges support from the Spanish Ministry of Economy and Competitiveness (grant FFI2016-78034-C2-1-P/FEDER), Marie Curie International Reintegration Grant from the European Union (PIRG-GA-2009-256413), Fundació Bosch i Gimpera, MEXT/JSPS Grant-in-Aid for Scientific Research on Innovative Areas 4903 (Evolinguistics: JP17H06379), and Generalitat de Catalunya (2017-SGR-341). TOR acknowledges support from the Generalitat de Catalunya (FI fellowship).

References

- [1] Richard W. Wrangham. Two types of aggression in human evolution. *Proceedings of the National Academy of Sciences*, 115(2):245–253, January 2018.
- [2] Brian Hare. Survival of the Friendliest: Homo sapiens Evolved via Selection for Prosociality. *Annual Review of Psychology*, 68(1):155–186, 2017.
- [3] Constantina Theofanopoulou, Simone Gastaldon, Thomas O’Rourke, Bridget D. Samuels, Pedro Tiago Martins, Francesco Delogu, Saleh Alamri, and Cedric Boeckx. Self-domestication in Homo sapiens: Insights from comparative genomics. *PLOS ONE*, 12(10):e0185306, October 2017.
- [4] Franz Boas. *The Mind of Primitive Man*. The MacMillan Company, United States of America, 1938.
- [5] Lee Alan Dugatkin and Lyudmila Trut. *How to Tame a Fox (and Build a Dog): Visionary Scientists and a Siberian Tale of Jump-Started Evolution*. University of Chicago Press, March 2017.
- [6] James P. Herman, Nancy K. Mueller, and Helmer Figueiredo. Role of GABA and glutamate circuitry in hypothalamo-pituitary-adrenocortical

- stress integration. *Annals of the New York Academy of Sciences*, 1018:35–45, June 2004.
- [7] Hiroki Goto, Kazunori Watanabe, Naozumi Araragi, Rui Kageyama, Kunika Tanaka, Yoko Kuroki, Atsushi Toyoda, Masahira Hattori, Yoshiyuki Sakaki, Asao Fujiyama, Yasuyuki Fukumaki, and Hiroki Shibata. The identification and functional implications of human-specific "fixed" amino acid substitutions in the glutamate receptor family. *BMC Evolutionary Biology*, 9:224, September 2009.
- [8] Kazuhiro Wada, Hironobu Sakaguchi, Erich D. Jarvis, and Masatoshi Hagiwara. Differential expression of glutamate receptors in avian neural pathways for learned vocalization. *Journal of Comparative Neurology*, 476(1):44–64, August 2004.
- [9] Xiling Liu, Mehmet Somel, Lin Tang, Zheng Yan, Xi Jiang, Song Guo, Yuan Yuan, Liu He, Anna Oleksiak, Yan Zhang, Na Li, Yuhui Hu, Wei Chen, Zilong Qiu, Svante Pääbo, and Philipp Khaitovich. Extension of cortical synaptic development distinguishes humans from chimpanzees and macaques. *Genome Research*, February 2012.
- [10] Gerard Muntané, Julie E. Horvath, Patrick R. Hof, John J. Ely, William D. Hopkins, Mary Ann Raghanti, Albert H. Lewandowski, Gregory A. Wray, and Chet C. Sherwood. Analysis of Synaptic Gene Expression in the Neocortex of Primates Reveals Evolutionary Changes in Glutamatergic Neurotransmission. *Cerebral Cortex*, 25(6):1596–1607, June 2015.
- [11] Stéphane Peyrégne, Michael James Boyle, Michael Dannemann, and Kay Prüfer. Detecting ancient positive selection in humans using extended lineage sorting. *Genome Research*, 27(9):1563–1572, September 2017.
- [12] Martin Kuhlwilm and Cedric Boeckx. Genetic differences between humans and other hominins contribute to the "human condition". *bioRxiv*, April 2018.
- [13] Robert John Denver. Structural and Functional Evolution of Vertebrate Neuroendocrine Stress Systems. *Annals of the New York Academy of Sciences*, 1163(1):1–16, April 2009.
- [14] József Haller. The glucocorticoid/aggression relationship in animals and humans: An analysis sensitive to behavioral characteristics, glucocorticoid secretion patterns, and neural mechanisms. In Klaus A. Miczek and Andreas Meyer-Lindenberg, editors, *Neuroscience of Aggression*, volume 17 of *Current Topics in Behavioral Neurosciences*, pages 73–109. Springer, Berlin, Heidelberg, February 2014.

- [15] Lyudmila Trut, Irina Oskina, and Anastasiya Kharlamova. Animal evolution during domestication: the domesticated fox as a model. *BioEssays : news and reviews in molecular, cellular and developmental biology*, 31(3):349–360, March 2009.
- [16] Adam S. Wilkins, Richard W. Wrangham, and W. Tecumseh Fitch. The “Domestication Syndrome” in Mammals: A Unified Explanation Based on Neural Crest Cell Behavior and Genetics. *Genetics*, 197(3):795–808, July 2014.
- [17] Florian J. Alberto, Frédéric Boyer, Pablo Orozco-terWengel, Ian Streeter, Bertrand Servin, Pierre Villemereuil, Badr Benjelloun, Pablo Librado, Filippo Biscarini, Licia Colli, Mario Barbato, Wahid Zamani, Adriana Alberti, Stefan Engelen, Alessandra Stella, Stéphane Joost, Paolo Ajmone-Marsan, Riccardo Negrini, Ludovic Orlando, Hamid Reza Rezaei, Saeid Naderi, Laura Clarke, Paul Flicek, Patrick Wincker, Eric Coissac, James Kijas, Gwenola Tosser-Klopp, Abdelkader Chikhi, Michael W. Bruford, Pierre Taberlet, and François Pompanon. Convergent genomic signatures of domestication in sheep and goats. *Nature Communications*, 9(1):813, March 2018.
- [18] Fernando Racimo. Testing for Ancient Selection Using Cross-population Allele Frequency Differentiation. *Genetics*, 202(2):733–750, February 2016.
- [19] Swapan Mallick, Heng Li, Mark Lipson, Iain Mathieson, Melissa Gymrek, Fernando Racimo, Mengyao Zhao, Niru Chennagiri, Susanne Nordenfelt, Arti Tandon, Pontus Skoglund, Iosif Lazaridis, Sriram Sankararaman, Qiaomei Fu, Nadin Rohland, Gabriel Renaud, Yaniv Erlich, Thomas Willems, Carla Gallo, Jeffrey P. Spence, Yun S. Song, Giovanni Polletti, Francois Balloux, George van Driem, Peter de Knijff, Irene Gallego Romero, Aashish R. Jha, Doron M. Behar, Claudio M. Bravi, Cristian Capelli, Tor Hervig, Andres Moreno-Estrada, Olga L. Posukh, Elena Balanovska, Oleg Balanovsky, Sena Karachanak-Yankova, Hovhannes Sahakyan, Draga Toncheva, Levon Yepiskoposyan, Chris Tyler-Smith, Yali Xue, M. Syafiq Abdullah, Andres Ruiz-Linares, Cynthia M. Beall, Anna Di Rienzo, Choongwon Jeong, Elena B. Starikovskaya, Ene Metspalu, Jüri Parik, Richard Villems, Brenna M. Henn, Ugur Hodoglugil, Robert Mahley, Antti Sajantila, George Stamatoyannopoulos, Joseph T. S. Wee, Rita Khusainova, Elza Khusnutdinova, Sergey Litvinov, George Ayodo, David Comas, Michael F. Hammer, Toomas Kivisild, William Klitz, Cheryl A. Winkler, Damian Labuda, Michael Bamshad, Lynn B. Jorde, Sarah A. Tishkoff, W. Scott Watkins, Mait Metspalu, Stanislav Dryomov, Rem Sukernik, Lalji Singh, Kumarasamy Thangaraj, Svante Pääbo, Janet Kelso, Nick Patterson, and David Reich. The Simons Genome Diversity Project: 300 genomes from 142 diverse populations. *Nature*, 538(7624):201–206, October 2016.

- [20] Kay Prüfer, Fernando Racimo, Nick Patterson, Flora Jay, Sriram Sankararaman, Susanna Sawyer, Anja Heinze, Gabriel Renaud, Peter H. Sudmant, Cesare de Filippo, Heng Li, Swapan Mallick, Michael Danneberg, Qiaomei Fu, Martin Kircher, Martin Kuhlwilm, Michael Lachmann, Matthias Meyer, Matthias Ongyerth, Michael Siebauer, Christoph Theunert, Arti Tandon, Priya Moorjani, Joseph Pickrell, James C. Mullikin, Samuel H. Vohr, Richard E. Green, Ines Hellmann, Philip L. F. Johnson, Hélène Blanche, Howard Cann, Jacob O. Kitzman, Jay Shendure, Evan E. Eichler, Ed S. Lein, Trygve E. Bakken, Liubov V. Golovanova, Vladimir B. Doronichev, Michael V. Shunkov, Anatoli P. Derevianko, Bence Viola, Montgomery Slatkin, David Reich, Janet Kelso, and Svante Pääbo. The complete genome sequence of a Neanderthal from the Altai Mountains. *Nature*, 505(7481):43–49, January 2014.
- [21] Eric T. Wang, Greg Kodama, Pierre Baldi, and Robert K. Moyzis. Global landscape of recent inferred Darwinian selection for *Homo sapiens*. *Proceedings of the National Academy of Sciences*, 103(1):135–140, January 2006.
- [22] Guo-dong Wang, Weiwei Zhai, He-chuan Yang, Ruo-xi Fan, Xue Cao, Li Zhong, Lu Wang, Fei Liu, Hong Wu, Lu-guang Cheng, Andrei D. Poyarkov, Nikolai A. Poyarkov Jr, Shu-sheng Tang, Wen-ming Zhao, Yun Gao, Xue-mei Lv, David M. Irwin, Peter Savolainen, Chung-I. Wu, and Ya-ping Zhang. The genomics of selection in dogs and the parallel evolution between dogs and humans. *Nature Communications*, 4:1860, May 2013.
- [23] Yan Li, Guo-Dong Wang, Ming-Shan Wang, David M. Irwin, Dong-Dong Wu, and Ya-Ping Zhang. Domestication of the Dog from the Wolf Was Promoted by Enhanced Excitatory Synaptic Plasticity: A Hypothesis. *Genome Biology and Evolution*, 6(11):3115–3121, November 2014.
- [24] Erik Axelsson, Abhirami Ratnakumar, Maja-Louise Arendt, Khurram Maqbool, Matthew T. Webster, Michele Perloski, Olof Liberg, Jon M. Arnemo, Ake Hedhammar, and Kerstin Lindblad-Toh. The genomic signature of dog domestication reveals adaptation to a starch-rich diet. *Nature*, 495(7441):360–364, March 2013.
- [25] Adam H. Freedman, Rena M. Schweizer, Diego Ortega-Del Vecchyo, Eun-jung Han, Brian W. Davis, Ilan Gronau, Pedro M. Silva, Marco Galaverni, Zhenxin Fan, Peter Marx, Belen Lorente-Galdos, Oscar Ramirez, Farhad Hormozdiari, Can Alkan, Carles Vilà, Kevin Squire, Eli Geffen, Josip Kusak, Adam R. Boyko, Heidi G. Parker, Clarence Lee, Vasisht Tadigotla, Adam Siepel, Carlos D. Bustamante, Timothy T. Harkins, Stanley F. Nelson, Tomas Marques-Bonet, Elaine A. Ostrander, Robert K. Wayne, and John Novembre. Demographically-Based Evaluation of Genomic Regions under Selection in Domestic Dogs. *PLOS Genetics*, 12(3):e1005851, April 2016.

- [26] Amanda L. Pendleton, Feichen Shen, Angela M. Taravella, Sarah Emery, Krishna R. Veeramah, Adam R. Boyko, and Jeffrey M. Kidd. Comparison of village dog and wolf genomes highlights the role of the neural crest in dog domestication. *BMC Biology*, 16:64, June 2018.
- [27] Saber Qanbari, Hubert Pausch, Sandra Jansen, Mehmet Somel, Tim M. Strom, Ruedi Fries, Rasmus Nielsen, and Henner Simianer. Classic Selective Sweeps Revealed by Massive Sequencing in Cattle. *PLOS Genetics*, 10(2):e1004148, February 2014.
- [28] Michael J. Montague, Gang Li, Barbara Gandolfi, Razib Khan, Bronwen L. Aken, Steven M. J. Searle, Patrick Minx, LaDeana W. Hillier, Daniel C. Koboldt, Brian W. Davis, Carlos A. Driscoll, Christina S. Barr, Kevin Blackistone, Javier Quilez, Belen Lorente-Galdos, Tomas Marques-Bonet, Can Alkan, Gregg W. C. Thomas, Matthew W. Hahn, Marilyn Menotti-Raymond, Stephen J. O'Brien, Richard K. Wilson, Leslie A. Lyons, William J. Murphy, and Wesley C. Warren. Comparative analysis of the domestic cat genome reveals genetic signatures underlying feline biology and domestication. *Proceedings of the National Academy of Sciences*, 111(48):17230–17235, December 2014.
- [29] Mikkel Schubert, Hákon Jónsson, Dan Chang, Clio Der Sarkissian, Luca Ermini, Aurélien Ginolhac, Anders Albrechtsen, Isabelle Dupanloup, Adrien Foucal, Bent Petersen, Matteo Fumagalli, Maanasa Raghavan, Andaine Seguin-Orlando, Thorfinn S. Korneliussen, Amhed M. V. Velazquez, Jesper Stenderup, Cindi A. Hoover, Carl-Johan Rubin, Ahmed H. Alfarhan, Saleh A. Alquraishi, Khaled A. S. Al-Rasheid, David E. MacHugh, Ted Kalbfleisch, James N. MacLeod, Edward M. Rubin, Thomas Sicheritz-Ponten, Leif Andersson, Michael Hofreiter, Tomas Marques-Bonet, M. Thomas P. Gilbert, Rasmus Nielsen, Laurent Excoffier, Eske Willerslev, Beth Shapiro, and Ludovic Orlando. Prehistoric genomes reveal the genetic foundation and cost of horse domestication. *Proceedings of the National Academy of Sciences*, 111(52):E5661–E5669, December 2014.
- [30] Xu Wang, Lenore Pipes, Lyudmila N. Trut, Yury Herbeck, Anastasiya V. Vladimirova, Rimma G. Gulevich, Anastasiya V. Kharlamova, Jennifer L. Johnson, Gregory M. Acland, Anna V. Kukekova, and Andrew G. Clark. Genomic responses to selection for tame/aggressive behaviors in the silver fox (*Vulpes vulpes*). *Proceedings of the National Academy of Sciences*, page 201800889, September 2018.
- [31] Anna V. Kukekova, Jennifer L. Johnson, Xueyan Xiang, Shaohong Feng, Shiping Liu, Halie M. Rando, Anastasiya V. Kharlamova, Yury Herbeck, Natalya A. Serdyukova, Zijun Xiong, Violetta Beklemischeva, Klaus-Peter Koepfli, Rimma G. Gulevich, Anastasiya V. Vladimirova, Jessica P. Hekman, Polina L. Perelman, Aleksander S. Graphodatsky, Stephen J. O'Brien, Xu Wang, Andrew G. Clark, Gregory M. Acland, Lyudmila N.

- Trut, and Guojie Zhang. Red fox genome assembly identifies genomic regions associated with tame and aggressive behaviours. *Nature Ecology & Evolution*, 2(9):1479–1491, September 2018.
- [32] Qiang Qiu, Lizhong Wang, Kun Wang, Yongzhi Yang, Tao Ma, Zefu Wang, Xiao Zhang, Zhengqiang Ni, Fujiang Hou, Ruijun Long, Richard Abbott, Johannes Lenstra, and Jianquan Liu. Yak whole-genome resequencing reveals domestication signatures and prehistoric population expansions. *Nature Communications*, 6:10283, December 2015.
- [33] Ivica Medugorac, Alexander Graf, Cécile Grohs, Sophie Rothhammer, Yondon Zagdsuren, Elena Gladyr, Natalia Zinovieva, Johanna Barbieri, Doris Seichter, Ingolf Russ, André Eggen, Garrett Hellenthal, Gottfried Brem, Helmut Blum, Stefan Krebs, and Aurélien Capitan. Whole-genome analysis of introgressive hybridization and characterization of the bovine legacy of Mongolian yaks. *Nature Genetics*, 49(3):470–475, March 2017.
- [34] Marina Naval-Sanchez, Quan Nguyen, Sean McWilliam, Laercio R. Portoneto, Ross Tellam, Tony Vuocolo, Antonio Reverter, Miguel Perez-Enciso, Rudiger Brauning, Shannon Clarke, Alan McCulloch, Wahid Zamani, Saeid Naderi, Hamid Reza Rezaei, Francois Pompanon, Pierre Taberlet, Kim C. Worley, Richard A. Gibbs, Donna M. Muzny, Shalini N. Jhangiani, Noelle Cockett, Hans Daetwyler, and James Kijas. Sheep genome functional annotation reveals proximal regulatory elements contributed to the evolution of modern breeds. *Nature Communications*, 9(1):859, February 2018.
- [35] Kai Wang, Pingxian Wu, Qiang Yang, Dejuan Chen, Jie Zhou, Anan Jiang, Jideng Ma, Qianzi Tang, Weihang Xiao, Yanzhi Jiang, Li Zhu, Xuwei Li, and Guoqing Tang. Detection of Selection Signatures in Chinese Landrace and Yorkshire Pigs Based on Genotyping-by-Sequencing Data. *Frontiers in Genetics*, 9(119), April 2018.
- [36] Jordi Leno-Colorado, Nick J. Hudson, Antonio Reverter, and Miguel Pérez-Enciso. A Pathway-Centered Analysis of Pig Domestication and Breeding in Eurasia. *G3: Genes/Genomes/Genetics*, 7(7):2171–2184, July 2017.
- [37] Sunjin Moon, Tae-Hun Kim, Kyung-Tai Lee, Woori Kwak, Taeheon Lee, Si-Woo Lee, Myung-Jick Kim, Kyuho Cho, Namshin Kim, Won-Hyong Chung, Samsun Sung, Taesung Park, Seoae Cho, Martien AM Groenen, Rasmus Nielsen, Yuseob Kim, and Heebal Kim. A genome-wide scan for signatures of directional selection in domesticated pigs. *BMC Genomics*, 16:130, February 2015.
- [38] Miguel Carneiro, Carl-Johan Rubin, Federica Di Palma, Frank W. Albert, Jessica Alföldi, Alvaro Martinez Barrio, Gerli Pielberg, Nima Rafati, Shumaila Sayyab, Jason Turner-Maier, Shady Younis, Sandra Afonso, Bronwen Aken, Joel M. Alves, Daniel Barrell, Gerard Bolet, Samuel Boucher,

- Hernán A. Burbano, Rita Campos, Jean L. Chang, Veronique Duranton, Luca Fontanesi, Hervé Garreau, David Heiman, Jeremy Johnson, Rose G. Mage, Ze Peng, Guillaume Queney, Claire Rogel-Gaillard, Magali Ruffier, Steve Searle, Rafael Villafuerte, Anqi Xiong, Sarah Young, Karin Forsberg-Nilsson, Jeffrey M. Good, Eric S. Lander, Nuno Ferrand, Kerstin Lindblad-Toh, and Leif Andersson. Rabbit genome analysis reveals a polygenic basis for phenotypic change during domestication. *Science (New York, N.Y.)*, 345(6200):1074–1079, August 2014.
- [39] Frank W. Albert, Mehmet Somel, Miguel Carneiro, Ayinuer Aximu-Petri, Michel Halbwax, Olaf Thalmann, Jose A. Blanco-Aguilar, Irina Z. Plyusnina, Lyudmila Trut, Rafael Villafuerte, Nuno Ferrand, Sylvia Kaiser, Per Jensen, and Svante Pääbo. A Comparison of Brain Gene Expression Levels in Domesticated and Wild Animals. *PLOS Genetics*, 8(9):e1002962, September 2012.
- [40] Carl-Johan Rubin, Michael C. Zody, Jonas Eriksson, Jennifer R. S. Meadows, Ellen Sherwood, Matthew T. Webster, Lin Jiang, Max Ingman, Ted Sharpe, Sojeong Ka, Finn Hallböök, Francois Besnier, Orjan Carlborg, Bertrand Bed'hom, Michèle Tixier-Boichard, Per Jensen, Paul Siegel, Kerstin Lindblad-Toh, and Leif Andersson. Whole-genome resequencing reveals loci under selection during chicken domestication. *Nature*, 464(7288):587–591, March 2010.
- [41] Zebin Zhang, Yaxiong Jia, Pedro Almeida, Judith E. Mank, Marcel van Tuinen, Qiong Wang, Zhihua Jiang, Yu Chen, Kai Zhan, Shuisheng Hou, Zhengkui Zhou, Huifang Li, Fangxi Yang, Yong He, Zhonghua Ning, Ning Yang, and Lujiang Qu. Whole-genome resequencing reveals signatures of selection and timing of duck domestication. *GigaScience*, 7(4), April 2018.
- [42] Colleen M. Niswender and P. Jeffrey Conn. Metabotropic Glutamate Receptors: Physiology, Pharmacology, and Disease. *Annual review of pharmacology and toxicology*, 50:295–322, July 2010.
- [43] Romolo Caniglia, Elena Fabbri, Pavel Hulva, Barbora Černá Bolfíková, Milena Jindřichová, Astrid Vik Stronen, Ihor Dykyy, Alessio Camatta, Paolo Carnier, Ettore Randi, and Marco Galaverni. Wolf outside, dog inside? The genomic make-up of the Czechoslovakian Wolfdog. *BMC Genomics*, 19(533), July 2018.
- [44] P. G. Eusebi, O. Cortés, C. Carleos, S. Dunner, and J. Cañon. Detection of selection signatures for agonistic behaviour in cattle. *Journal of Animal Breeding and Genetics = Zeitschrift Fur Tierzucht Und Zuchtungsbiologie*, April 2018.
- [45] Andrea Benazzo, Emiliano Trucchi, James A. Cahill, Pierpaolo Maisano Delser, Stefano Mona, Matteo Fumagalli, Lynsey Bunnefeld, Luca Cornetti, Silvia Ghirotto, Matteo Girardi, Lino Ometto, Alex Panziera, Omar

- Rota-Stabelli, Enrico Zanetti, Alexandros Karamanlidis, Claudio Groff, Ladislav Paule, Leonardo Gentile, Carles Vilà, Saverio Vicario, Luigi Boitani, Ludovic Orlando, Silvia Fuselli, Cristiano Vernesi, Beth Shapiro, Paolo Ciucci, and Giorgio Bertorelle. Survival and divergence in a small group: The extraordinary genomic history of the endangered Apennine brown bear stragglers. *Proceedings of the National Academy of Sciences*, 114(45):E9589–E9597, November 2017.
- [46] Jim van Os and Jean-Paul Selten. Prenatal exposure to maternal stress and subsequent schizophrenia: The May 1940 invasion of the Netherlands. *The British Journal of Psychiatry*, 172(4):324–326, April 1998.
- [47] James I. Koenig, Brian Kirkpatrick, and Paul Lee. Glucocorticoid Hormones and Early Brain Development in Schizophrenia. *Neuropsychopharmacology*, 27(2):309–318, August 2002.
- [48] Francesco Matrisciano, Patricia Tueting, Stefania Maccari, Ferdinando Nicoletti, and Alessandro Guidotti. Pharmacological Activation of Group-II Metabotropic Glutamate Receptors Corrects a Schizophrenia-Like Phenotype Induced by Prenatal Stress in Mice. *Neuropsychopharmacology*, 37(4):929–938, March 2012.
- [49] Richard G. Hunter, Rudy Bellani, Erik Bloss, Ana Costa, Katharine McCarthy, and Bruce S. McEwen. Regulation of Kainate Receptor Subunit mRNA by Stress and Corticosteroids in the Rat Hippocampus. *PLOS ONE*, 4(1):e4328, January 2009.
- [50] Francine M. Benes, Mark S. Todtenkopf, and Paul Kostoulakos. GluR5,6,7 subunit immunoreactivity on apical pyramidal cell dendrites in hippocampus of schizophrenics and manic depressives. *Hippocampus*, 11(5):482–491, October 2001.
- [51] L. a. M. Welberg and J. R. Seckl. Prenatal Stress, Glucocorticoids and the Programming of the Brain. *Journal of Neuroendocrinology*, 13(2):113–128, February 2001.
- [52] Maria G. Motlagh, Liliya Katsoyich, Nancy Thompson, Haiqun Lin, Young-Shin Kim, Lawrence Scahill, Paul J. Lombroso, Robert A. King, Bradley S. Peterson, and James F. Leckman. Severe psychosocial stress and heavy cigarette smoking during pregnancy: an examination of the pre- and perinatal risk factors associated with ADHD and Tourette syndrome. *European Child & Adolescent Psychiatry*, 19(10):755–764, October 2010.
- [53] M. K. Green, C. S. S. Rani, A. Joshi, A. E. Soto-Piña, P. A. Martinez, A. Frazer, R. Strong, and D. A. Morilak. Prenatal stress induces long term stress vulnerability, compromising stress response systems in the brain and impairing extinction of conditioned fear after adult stress. *Neuroscience*, 192:438–451, September 2011.

- [54] D. K. Belyaev. Destabilizing selection as a factor in domestication. *Journal of Heredity*, 70(5):301–308, September 1979.
- [55] Nathan K. Evanson and James P. Herman. Role of Paraventricular Nucleus Glutamate Signaling in Regulation of HPA Axis Stress Responses. *Interdisciplinary information sciences*, 21(3):253–260, September 2015.
- [56] Aki Takahashi and Klaus A. Miczek. Neurogenetics of Aggressive Behavior: Studies in Rodents. In Klaus A. Miczek and Andreas Meyer-Lindenberg, editors, *Neuroscience of Aggression*, volume 17 of *Current Topics in Behavioral Neurosciences*, pages 3–44. Springer, Berlin, Heidelberg, December 2013.
- [57] Yanli Zhang-James, Noèlia Fernández-Castillo, Jonathan L. Hess, Karim Malki, Stephen J. Glatt, Bru Cormand, and Stephen V. Faraone. An integrated analysis of genes and functional pathways for aggression in human and rodent models. *Molecular Psychiatry*, page 1, June 2018.
- [58] Anis Contractor, Christophe Mulle, and Geoffrey T. Swanson. Kainate receptors coming of age: milestones of two decades of research. *Trends in Neurosciences*, 34(3):154–163, March 2011.
- [59] J. Axelrod and T. D. Reisine. Stress hormones: their interaction and regulation. *Science*, 224(4648):452–459, May 1984.
- [60] Ryan Jankord and James P. Herman. Limbic Regulation of Hypothalamo-Pituitary-Adrenocortical Function During Acute and Chronic Stress. *Annals of the New York Academy of Sciences*, 1148:64–73, December 2008.
- [61] J.P. Herman, J.M. McKlveen, M.B. Solomon, E. Carvalho-Netto, and B. Myers. Neural regulation of the stress response: glucocorticoid feedback mechanisms. *Brazilian Journal of Medical and Biological Research*, 45(4):292–298, March 2012.
- [62] James P. Herman, Michelle M. Ostrander, Nancy K. Mueller, and Helmer Figueiredo. Limbic system mechanisms of stress regulation: Hypothalamo-pituitary-adrenocortical axis. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 29(8):1201–1213, December 2005.
- [63] Lauren Jacobson and Robert Sapolsky. The Role of the Hippocampus in Feedback Regulation of the Hypothalamic-Pituitary-Adrenocortical Axis*. *Endocrine Reviews*, 12(2):118–134, May 1991.
- [64] I. Z. Plyusnina, I. N. Oskina, and L. N. Trut. An analysis of fear and aggression during early development of behaviour in silver foxes (*Vulpes vulpes*). *Applied Animal Behaviour Science*, 32(2):253–268, November 1991.

- [65] E. V. Naumenko, N. K. Popova, E. M. Nikulina, N. N. Dygalo, G. T. Shishkina, P. M. Borodin, and A. L. Markel. Behavior, adrenocortical activity, and brain monoamines in Norway rats selected for reduced aggressiveness towards man. *Pharmacology Biochemistry and Behavior*, 33(1):85–91, May 1989.
- [66] J. M. Bassett and N. T. Hinks. Micro-determination of corticosteroids in ovine peripheral plasma: effects of venipuncture, corticotrophin, insulin and glucose. *The Journal of Endocrinology*, 44(3):387–403, July 1969.
- [67] Jack C. Turner. Diurnal periodicity of plasma Cortisol and corticosterone in desert bighorn sheep demonstrated by radioimmunoassay. *Canadian Journal of Zoology*, 62(12):2659–2665, December 1984.
- [68] Kenta Suzuki, Maki Ikebuchi, Hans-Joachim Bischof, and Kazuo Okanoya. Behavioral and neural trade-offs between song complexity and stress reaction in a wild and a domesticated finch strain. *Neuroscience & Biobehavioral Reviews*, 46:547–556, October 2014.
- [69] James T. Martin. Hormonal Influences in the Evolution and Ontogeny of Imprinting Behavior in the Duck. In W. H. Gispen, Tj. B. van Wimersma Greidanus, B. Bohus, and D. de Wied, editors, *Progress in Brain Research*, volume 42 of *Hormones, Homeostasis and the Brain*, pages 357–366. Elsevier, January 1975.
- [70] James T. Martin. Embryonic Pituitary Adrenal Axis, Behavior Development and Domestication in Birds. *Integrative and Comparative Biology*, 18(3):489–499, August 1978.
- [71] Christine Künzli and Norbert Sachser. The Behavioral Endocrinology of Domestication: A Comparison between the Domestic Guinea Pig (*Cavia apereaf.porcellus*) and Its Wild Ancestor, the Cavy (*Cavia aperea*). *Hormones and Behavior*, 35(1):28–37, February 1999.
- [72] Maria Ericsson, Amir Fallahsharoudi, Jonas Bergquist, Mark M. Kushnir, and Per Jensen. Domestication effects on behavioural and hormonal responses to acute stress in chickens. *Physiology & Behavior*, 133:161–169, June 2014.
- [73] G. P. Chrousos, D. Renquist, D. Brandon, C. Eil, M. Pugeat, R. Vigersky, G. B. Cutler, D. L. Loriaux, and M. B. Lipsett. Glucocorticoid hormone resistance during primate evolution: receptor-mediated mechanisms. *Proceedings of the National Academy of Sciences*, 79(6):2036–2040, March 1982.
- [74] Nestor L. Lopez-Duran, Sheryl L. Olson, Nastassia J. Hajal, Barbara T. Felt, and Delia M. Vazquez. Hypothalamic Pituitary Adrenal Axis Functioning in Reactive and Proactive Aggression in Children. *Journal of Abnormal Child Psychology*, 37(2):169–182, February 2009.

- [75] James P Herman and William E Cullinan. Neurocircuitry of stress: central control of the hypothalamo–pituitary–adrenocortical axis. *Trends in Neurosciences*, 20(2):78–84, February 1997.
- [76] James P. Herman, Ozhan Eyigor, Dana R. Ziegler, and Lothar Jennes. Expression of ionotropic glutamate receptor subunit mRNAs in the hypothalamic paraventricular nucleus of the rat. *Journal of Comparative Neurology*, 422(3):352–362, June 2000.
- [77] Jean-Michel Aubry, Viktor Bartanusz, Sonia Pagliusi, Pierre Schulz, and Jozsef Z. Kiss. Expression of ionotropic glutamate receptor subunit mRNAs by paraventricular corticotropin-releasing factor (CRF) neurons. *Neuroscience Letters*, 205(2):95–98, February 1996.
- [78] Qing-Song Liu, Peter R. Patrylo, Xiao-Bing Gao, and Anthony N. van den Pol. Kainate Acts at Presynaptic Receptors to Increase GABA Release From Hypothalamic Neurons. *Journal of Neurophysiology*, 82(2):1059–1062, August 1999.
- [79] S. Scaccianoce, F. Matrisciano, P. Del Bianco, A. Caricasole, V. Di Giorgi Gerevini, I. Cappuccio, D. Melchiorri, G. Battaglia, and F. Nicoletti. Endogenous activation of group-II metabotropic glutamate receptors inhibits the hypothalamic–pituitary–adrenocortical axis. *Neuropharmacology*, 44(5):555–561, April 2003.
- [80] Claudio Acuna-Goycolea, Ying Li, and Anthony N. van den Pol. Group III Metabotropic Glutamate Receptors Maintain Tonic Inhibition of Excitatory Synaptic Input to Hypocretin/Orexin Neurons. *Journal of Neuroscience*, 24(12):3013–3022, March 2004.
- [81] D. J. Morsette, H. Sidorowicz, and C. D. Sladek. Role of metabotropic glutamate receptors in vasopressin and oxytocin release. *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology*, 281(2):R452–458, August 2001.
- [82] David L. Walker, Lisa M. Rattiner, and Michael Davis. Group II metabotropic glutamate receptors within the amygdala regulate fear as assessed with potentiated startle in rats. *Behavioral Neuroscience*, 116(6):1075–1083, December 2002.
- [83] Yosuke Morishima, Tsuyoshi Miyakawa, Tomoyuki Furuyashiki, Yasuhiro Tanaka, Hiroshi Mizuma, and Shigetada Nakanishi. Enhanced cocaine responsiveness and impaired motor coordination in metabotropic glutamate receptor subtype 2 knockout mice. *Proceedings of the National Academy of Sciences*, 102(11):4170–4175, March 2005.
- [84] Jeremy D. Coplan, Sanjay J. Mathew, Eric L. P. Smith, Ronald C. Trost, Bruce A. Scharf, Jose Martinez, Jack M. Gorman, James A. Monn, Darryle D. Schoepp, and Leonard A. Rosenblum. Effects of LY354740,

- a Novel Glutamatergic Metabotropic Agonist, on Nonhuman Primate Hypothalamic-Pituitary-Adrenal Axis and Noradrenergic Function. *CNS Spectrums*, 6(7):607–617, July 2001.
- [85] Saulo J. A. Felizola, Yasuhiro Nakamura, Fumitoshi Satoh, Ryo Morimoto, Kumi Kikuchi, Tomohiro Nakamura, Atsushi Hozawa, Lin Wang, Yoshiaki Onodera, Kazue Ise, Keely M. McNamara, Sanae Midorikawa, Shinichi Suzuki, and Hironobu Sasano. Glutamate receptors and the regulation of steroidogenesis in the human adrenal gland: the metabotropic pathway. *Molecular and Cellular Endocrinology*, 382(1):170–177, January 2014.
- [86] Yukio Ago, Ryota Araki, Koji Yano, Toshiyuki Kawasaki, Shigeyuki Chaki, Atsuro Nakazato, Hirotaka Onoe, Hitoshi Hashimoto, Akemichi Baba, Kazuhiro Takuma, and Toshio Matsuda. The Selective Metabotropic Glutamate 2/3 Receptor Agonist MGS0028 Reverses Isolation Rearing-Induced Abnormal Behaviors in Mice. *Journal of Pharmacological Sciences*, 118(2):295–298, 2012.
- [87] Yu Zhao, Christopher V. Dayas, Harinder Aujla, Marco A. S. Baptista, Rémi Martin-Fardon, and Friedbert Weiss. Activation of Group II Metabotropic Glutamate Receptors Attenuates Both Stress and Cue-Induced Ethanol-Seeking and Modulates c-fos Expression in the Hippocampus and Amygdala. *Journal of Neuroscience*, 26(39):9967–9974, September 2006.
- [88] Brad A. Grueter and Danny G. Winder. Group II and III Metabotropic Glutamate Receptors Suppress Excitatory Synaptic Transmission in the Dorsolateral Bed Nucleus of the Stria Terminalis. *Neuropsychopharmacology*, 30(7):1302–1311, July 2005.
- [89] Robert M. Duvoisin, Connie Zhang, Timothy F. Pfankuch, Heather O'Connor, Jacqueline Gayet-Primo, Salma Quraishi, and Jacob Raber. Increased measures of anxiety and weight gain in mice lacking the group III metabotropic glutamate receptor mGluR8. *European Journal of Neuroscience*, 22(2):425–436, July 2005.
- [90] Robert M. Duvoisin, Laura Villasana, Timothy Pfankuch, and Jacob Raber. Sex-dependent cognitive phenotype of mice lacking mGluR8. *Behavioural Brain Research*, 209(1):21–26, May 2010.
- [91] Robert M. Duvoisin, Laura Villasana, Matthew J. Davis, Danny G. Winder, and Jacob Raber. Opposing roles of mGluR8 in measures of anxiety involving non-social and social challenges. *Behavioural Brain Research*, 221(1):50–54, August 2011.
- [92] Miwako Masugi, Mineto Yokoi, Ryuichi Shigemoto, Keiko Muguruma, Yasuyoshi Watanabe, Gilles Sansig, Herman van der Putten, and Shigetada Nakanishi. Metabotropic Glutamate Receptor Subtype 7 Ablation

- Causes Deficit in Fear Response and Conditioned Taste Aversion. *Journal of Neuroscience*, 19(3):955–963, February 1999.
- [93] Miwako Masugi-Tokita, Peter J. Flor, and Mitsuhiro Kawata. Metabotropic Glutamate Receptor Subtype 7 in the Bed Nucleus of the Stria Terminalis is Essential for Intermale Aggression. *Neuropsychopharmacology*, 41(3):726–735, February 2016.
- [94] Matthew J. Davis, Tammie Haley, Robert M. Duvoisin, and Jacob Raber. Measures of anxiety, sensorimotor function, and memory in male and female mGluR4^{-/-} mice. *Behavioural Brain Research*, 229(1):21–28, April 2012.
- [95] Chad J. Swanson, Mark Bures, Michael P. Johnson, Anni-Maija Linden, James A. Monn, and Darryle D. Schoepp. Metabotropic glutamate receptors as novel targets for anxiety and stress disorders. *Nature Reviews Drug Discovery*, 4(2):131–144, February 2005.
- [96] M. Joëls, A. Bosma, H. Hendriksen, P. Diegenbach, and W. Kamphuis. Corticosteroid actions on the expression of kainate receptor subunit mRNAs in rat hippocampus. *Molecular Brain Research*, 37(1):15–20, April 1996.
- [97] José V. Negrete-Díaz, Talvinder S. Sihra, Gonzalo Flores, and Antonio Rodríguez-Moreno. Non-canonical Mechanisms of Presynaptic Kainate Receptors Controlling Glutamate Release. *Frontiers in Molecular Neuroscience*, 11, April 2018.
- [98] Maurizio Popoli, Zhen Yan, Bruce McEwen, and Gerard Sanacora. The stressed synapse: the impact of stress and glucocorticoids on glutamate transmission. *Nature reviews. Neuroscience*, 13(1):22–37, November 2011.
- [99] James P. Herman, Jeffrey G. Tasker, Dana R. Ziegler, and William E. Cullinan. Local circuit regulation of paraventricular nucleus stress integration: Glutamate–GABA connections. *Pharmacology Biochemistry and Behavior*, 71(3):457–468, March 2002.
- [100] Shi Di, Marc M. Maxson, Alier Franco, and Jeffrey G. Tasker. Glucocorticoids Regulate Glutamate and GABA Synapse-Specific Retrograde Transmission via Divergent Non-Genomic Signaling Pathways. *Journal of Neuroscience*, 29(2):393–401, January 2009.
- [101] Joana Lourenço, Isabel Matias, Giovanni Marsicano, and Christophe Mulle. Pharmacological Activation of Kainate Receptors Drives Endocannabinoid Mobilization. *Journal of Neuroscience*, 31(9):3243–3248, March 2011.
- [102] John J. Marshall, Jian Xu, and Anis Contractor. Kainate receptors inhibit glutamate release via mobilization of endocannabinoids in striatal direct

- pathway Spiny Projection Neurons (dSPNs). *Journal of Neuroscience*, pages 1788–17, March 2018.
- [103] Cristiane Busnardo, Carlos C. Crestani, Leonardo B. M. Resstel, Rodrigo F. Tavares, José Antunes-Rodrigues, and Fernando M. A. Corrêa. Ionotropic Glutamate Receptors in Hypothalamic Paraventricular and Supraoptic Nuclei Mediate Vasopressin and Oxytocin Release in Unanesthetized Rats. *Endocrinology*, 153(5):2323–2331, May 2012.
- [104] Klaus-Peter Lesch and Jonas Waider. Serotonin in the Modulation of Neural Plasticity and Networks: Implications for Neurodevelopmental Disorders. *Neuron*, 76(1):175–191, October 2012.
- [105] F. W. Albert, E. Hodges, J. D. Jensen, F. Besnier, Z. Xuan, M. Rooks, A. Bhattacharjee, L. Brizuela, J. M. Good, R. E. Green, H. A. Burbano, I. Z. Plyusnina, L. Trut, L. Andersson, T. Schöneberg, Ö Carlborg, G. J. Hannon, and S. Pääbo. Targeted resequencing of a genomic region influencing tameness and aggression reveals multiple signals of positive selection. *Heredity*, 107(3):205–214, September 2011.
- [106] Yang Dong, Xiaolei Zhang, Min Xie, Babak Arefnezhad, Zongji Wang, Wenliang Wang, Shaohong Feng, Guodong Huang, Rui Guan, Wenjing Shen, Rowan Bunch, Russell McCulloch, Qiye Li, Bo Li, Guojie Zhang, Xun Xu, James W. Kijas, Ghasem Hosseini Salekdeh, Wen Wang, and Yu Jiang. Reference genome of wild goat (*capra aegagrus*) and sequencing of goat breeds provide insight into genic basis of goat domestication. *BMC Genomics*, 16:431, June 2015.
- [107] John W. Olney and Nuri B. Farber. Glutamate Receptor Dysfunction and Schizophrenia. *Archives of General Psychiatry*, 52(12):998–1007, December 1995.
- [108] Stuart Cull-Candy, Stephen Brickley, and Mark Farrant. NMDA receptor subunits: diversity, development and disease. *Current Opinion in Neurobiology*, 11(3):327–335, June 2001.
- [109] John W Olney, John W Newcomer, and Nuri B Farber. NMDA receptor hypofunction model of schizophrenia. *Journal of Psychiatric Research*, 33(6):523–533, November 1999.
- [110] Carlos A. Zarate, Jaskaran B. Singh, Paul J. Carlson, Nancy E. Brutsche, Rezvan Ameli, David A. Luckenbaugh, Dennis S. Charney, and Hussein K. Manji. A Randomized Trial of an N-methyl-D-aspartate Antagonist in Treatment-Resistant Major Depression. *Archives of General Psychiatry*, 63(8):856–864, August 2006.
- [111] E. Scarr, G. Pavey, S. Sundram, A. MacKinnon, and B. Dean. Decreased hippocampal NMDA, but not kainate or AMPA receptors in bipolar disorder. *Bipolar Disorders*, 5(4):257–264, August 2003.

- [112] Andrew Alt, Eric S. Nisenbaum, David Bleakman, and Jeffrey M. Witkin. A role for AMPA receptors in mood disorders. *Biochemical Pharmacology*, 71(9):1273–1288, April 2006.
- [113] Paul D. Arnold, David R. Rosenberg, Emanuela Mundo, Subi Tharmalingam, James L. Kennedy, and Margaret A. Richter. Association of a glutamate (NMDA) subunit receptor gene (GRIN2b) with obsessive-compulsive disorder: a preliminary study. *Psychopharmacology*, 174(4):530–538, August 2004.
- [114] B. Bettler and C. Mulle. AMPA and kainate receptors. *Neuropharmacology*, 34(2):123–139, February 1995.
- [115] David Perrais, Julien Veran, and Christophe Mulle. Gating and permeation of kainate receptors: differences unveiled. *Trends in Pharmacological Sciences*, 31(11):516–522, November 2010.
- [116] Maile A. Henson, Rylan S. Larsen, Shelikha N. Lawson, Isabel Pérez-Otaño, Nobuki Nakanishi, Stuart A. Lipton, and Benjamin D. Philpot. Genetic deletion of NR3a accelerates glutamatergic synapse maturation. *PLoS One*, 7(8):e42327, 2012.
- [117] John T. R. Isaac, Michael C. Ashby, and Chris J. McBain. The Role of the GluR2 Subunit in AMPA Receptor Function and Synaptic Plasticity. *Neuron*, 54(6):859–871, June 2007.
- [118] Faraneh Vargha-Khadem, David G. Gadian, Andrew Copp, and Mortimer Mishkin. *FOXP2* and the neuroanatomy of speech and language. *Nature Reviews Neuroscience*, 6(2):131–138, February 2005.
- [119] S. Reimers-Kipping, W. Hevers, S. Pääbo, and W. Enard. Humanized *Foxp2* specifically affects cortico-basal ganglia circuits. *Neuroscience*, 175:75–84, February 2011.
- [120] Cecilia S. L. Lai, Dianne Gerrelli, Anthony P. Monaco, Simon E. Fisher, and Andrew J. Copp. *FOXP2* expression during brain development coincides with adult sites of pathology in a severe speech and language disorder. *Brain*, 126(11):2455–2462, November 2003.
- [121] Zhimin Shi, Zoe Piccus, Xiaofang Zhang, Huidi Yang, Hannah Jarrell, Yan Ding, Zhaoqian Teng, Ofer Tchernichovski, and XiaoChing Li. miR-9 regulates basal ganglia-dependent developmental vocal learning and adult vocal performance in songbirds. *eLife*, 7:e29087, January 2018.
- [122] Kazuo Okanoya. Evolution of song complexity in Bengalese finches: Sexual selection and domestication as two factors. *The Journal of the Acoustical Society of America*, 138(3):1880–1880, September 2015.

- [123] Kazuo Okanoya. Sexual communication and domestication may give rise to the signal complexity necessary for the emergence of language: An indication from songbird studies. *Psychonomic Bulletin & Review*, 24(1):106–110, February 2017.
- [124] Pedro Rodenas-Cuadrado, Xiaowei Sylvia Chen, Lutz Wiegrebe, Uwe Firzlaff, and Sonja C. Vernes. A novel approach identifies the first transcriptome networks in bats: a new genetic model for vocal communication. *BMC Genomics*, 16:836, October 2015.
- [125] Marcelo R. Sánchez-Villagra, Madeleine Geiger, and Richard A. Schneider. The taming of the neural crest: a developmental perspective on the origins of morphological covariation in domesticated mammals. *Royal Society Open Science*, 3(6):160107, June 2016.
- [126] A. J. Doupe, S. C. Landis, and P. H. Patterson. Environmental influences in the development of neural crest derivatives: glucocorticoids, growth factors, and chromaffin cell plasticity. *Journal of Neuroscience*, 5(8):2119–2142, August 1985.
- [127] A. J. Doupe, P. H. Patterson, and S. C. Landis. Small intensely fluorescent cells in culture: role of glucocorticoids and growth factors in their development and interconversions with other neural crest derivatives. *Journal of Neuroscience*, 5(8):2143–2160, August 1985.
- [128] T. J. Cole, J. A. Blendy, A. P. Monaghan, K. Kriegstein, W. Schmid, A. Aguzzi, G. Fantuzzi, E. Hummler, K. Unsicker, and G. Schütz. Targeted disruption of the glucocorticoid receptor gene blocks adrenergic chromaffin cell development and severely retards lung maturation. *Genes & Development*, 9(13):1608–1621, July 1995.
- [129] I. N. Oskina, L. A. Prasolova, I. Z. Plyusnina, and L. N. Trut. Role of glucocorticoids in coat depigmentation in animals selected for behavior. *Cytology and Genetics*, 44(5):286–293, October 2010.
- [130] Liyang Wang, Eiichi Hinoi, Akihiro Takemori, Takeshi Takarada, and Yukio Yoneda. Abolition of chondral mineralization by group III metabotropic glutamate receptors expressed in rodent cartilage. *British Journal of Pharmacology*, 146(5):732–743, November 2005.
- [131] M. J. Hoogduijn, I. S. Hitchcock, N. P. M. Smit, J. M. Gillbro, K. U. Schallreuter, and P. G. Genever. Glutamate receptors on human melanocytes regulate the expression of MiTF. *Pigment Cell Research*, 19(1):58–67, February 2006.
- [132] Sulochana Devi, Yogananda Markandeya, Nityanand Maddodi, Anuradha Dhingra, Noga Vardi, Ravi C. Balijepalli, and Vijayasradhi Setaluri. Metabotropic glutamate receptor 6 signaling enhances TRPM1 calcium channel function and increases melanin content in human melanocytes. *Pigment Cell & Melanoma Research*, 26(3):348–356, May 2013.

- [133] Marcela Julio-Pieper, Peter J. Flor, Timothy G. Dinan, and John F. Cryan. Exciting Times beyond the Brain: Metabotropic Glutamate Receptors in Peripheral and Non-Neural Tissues. *Pharmacological Reviews*, January 2011.
- [134] Csaba Matta. Calcium signalling in chondrogenesis implications for cartilage repair. *Frontiers in Bioscience*, 5(1):305–324, 2013.
- [135] Kevin B Jones, Anthony V Mollano, Jose A Morcuende, Reginald R Cooper, and Charles L Saltzman. Bone and Brain. *The Iowa Orthopaedic Journal*, 24:123–132, 2004.
- [136] Wenjie Xie, Silvia Dolder, Mark Siegrist, Antoinette Wetterwald, and Willy Hofstetter. Glutamate Receptor Agonists and Glutamate Transporter Antagonists Regulate Differentiation of Osteoblast Lineage Cells. *Calcified Tissue International*, 99(2):142–154, August 2016.
- [137] Eiichi Hinoi, Sayumi Fujimori, Yoichi Nakamura, and Yukio Yoneda. Group III Metabotropic Glutamate Receptors in Rat Cultured Calvarial Osteoblasts. *Biochemical and Biophysical Research Communications*, 281(2):341–346, February 2001.
- [138] Eiichi Hinoi, Sayumi Fujimori, Akihiro Takemori, Hiroaki Kurabayashi, Yoichi Nakamura, and Yukio Yoneda. Demonstration of expression of mRNA for particular AMPA and kainate receptor subunits in immature and mature cultured rat calvarial osteoblasts. *Brain Research*, 943(1):112–116, July 2002.
- [139] Yoshifumi Takahata, Takeshi Takarada, Masato Osawa, Eiichi Hinoi, Yukari Nakamura, and Yukio Yoneda. Differential regulation of cellular maturation in chondrocytes and osteoblasts by glycine. *Cell and Tissue Research*, 333(1):91–103, July 2008.