The effects of training population design on genomic prediction accuracy in wheat

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17 Abstract

Genomic selection offers several routes for increasing genetic gain or efficiency of plant breeding programs. In various species of livestock there is empirical evidence of increased rates of genetic gain from the use of genomic selection to target different aspects of the breeder's equation. Accurate predictions of genomic breeding value are central to this and the design of training sets is in turn central to achieving sufficient levels of accuracy. In summary, small numbers of close relatives and very large numbers of distant relatives are expected to enable accurate predictions.

To quantify the effect of some of the properties of training sets on the accuracy of 25 genomic selection in crops we performed an extensive field-based winter wheat trial. In 26 summary, this trial involved the construction of 44 F_{2:4} bi- and triparental populations, from 27 which 2992 lines were grown on four field locations and yield was measured. For each line, 28 genotype data were generated for 25,000 segregating single nucleotide polymorphism 29 markers. The overall heritability of yield was estimated to 0.65, and estimates within 30 individual families ranged between 0.10 and 0.85. Within cross genomic prediction accuracies 31 of yield BLUEs were 0.125 - 0.127 using two different cross-validation approaches, and 32 generally increased with training set size. Using related crosses in training and validation sets 33 generally resulted in higher prediction accuracies than using unrelated crosses. The results of 34 this study emphasize the importance of the training set design in relation to the genetic 35 material to which the resulting prediction model is to be applied. 36

37 Keywords: genomic selection, wheat, population design

39 Introduction

Genomic selection in plant breeding offers several routes for increasing the genetic gain 40 or efficiency of plant breeding programs (e.g., Bernardo and Yu, 2007; Hickey et al., 2014; 41 Gaynor et al., 2017). Genomic selection based strategies can achieve this by reducing breeding 42 cycle time, increasing selection accuracy and increasing selection intensity; three of the four 43 factors in the breeder's equation. Genomic prediction can reduce breeding cycle time because 44 individuals can be selected and crossed without being phenotyped. It can increase the selection 45 accuracy because genomic data enables more powerful statistical models and experimental 46 designs using more observations than can be phenotyped in a single trial round. By reducing 47 48 the cost of evaluating individuals via reducing the numbers phenotyped and/or reducing their replication, application of genomic selection can increase selection intensity. A final advantage 49 is that the prediction models may be cumulatively updated with data of trials from previous 50 years and become more accurate, enabling individuals to be "evaluated" across a broader 51 range of environments and years. 52

In livestock there is empirical evidence of increased rates of genetic gain from the use of 53 genomic selection to target different aspects of the breeder's equation. For example the first 54 seven years of genomic selection in US dairy cattle has delivered ~50 - 100% increases in rates 55 of genetic gain (García-Ruiz et al., 2016). Much of this gain has emanated from a reduction in 56 generation interval. In commercial pig breeding, genomic selection has driven a 35% increase 57 in rate of genetic gain in the breeding program that supplies the genetics in 25% of the 58 intensively raised pigs globally. This gain came from increased accuracy of selection and a 59 better alignment of selection accuracy with the breeding goal (W. Herring, personal 60 communication). 61

Genomic selection uses genotype data to calculate the realised relationship between individuals, and in a standardized statistical framework uses data from phenotyped relatives to estimate genetic values of the selection candidates. The usefulness of genomic selection to a breeder is a function of its accuracy. This is affected by the relatedness between the phenotyped individuals in the training set and the individuals that are to be predicted (Habier
et al., 2007, 2010; Meuwissen, 2009; Clark et al., 2012; Hickey et al., 2014; Liu et al., 2016),
which may or may not be phenotyped themselves. In addition to the level of relatedness, the
sample size of the phenotyped individuals is an important factor in determining accuracy
(Zhang et al., 2017).

In summary, small numbers of close relatives and very large numbers of distant relatives enable accurate predictions. Small or modest numbers of distant relatives do not enable accurate predictions, as they share only a small proportion of genome with the selection candidates, and thus provide less reliable predictions (de los Campos et al., 2013). Finally, the training set should also comprise a diverse set of individuals to produce reliable predictions (Calus, 2010; Pszczola et al., 2012; Pszczola and Calus, 2015), as supported by recent research in both cattle (Jenko et al., 2017) and simulated barley (Neyhart et al., 2017).

The objective of this study was to explore the effect of level of relatedness between 78 training set and validation set on genomic prediction accuracy using data from a large set of 79 field experiments. To do this, 44 bi-parental or three-way crosses were obtained from four 80 commercial wheat breeders in the United Kingdom, as described for the GplusE Project 81 (Mackay et al., 2015). The crosses had different degrees of relatedness among each other and 82 there were many shared parents. 68 F_{2:4} lines from each cross were genotyped and phenotyped 83 for yield. As this data set is of substantial size, it enabled genomic predictions while masking 84 85 specific fractions to assess the impact on genomic selection accuracy of training sets: (i) of different sizes; and (ii) that comprise close or distant relatives, or combinations thereof. 86

88 Materials and Methods

89 Germplasm

Thirty-nine bi-parental and 5 triparental populations were used to develop 2992 $F_{2:4}$ lines (68 per cross). The parents of these populations were elite breeders' germplasm consisting of both hard and soft winter wheat cultivars adapted to the United Kingdom. A total of 27 parents were used, of which 5 parents were used in 6 or more crosses, 6 parents were used in 3 or 4 crosses, and 1 parent was used in 2 crosses. The remaining 15 parents were only used in a single cross.

96 Genotypes

The $F_{2:4}$ lines were genotyped using the Wheat Breeders' 35K Axiom array (Allen et al., 2016). The DNA for genotyping was obtained by bulking leaves from approximately 6 F_4 plants per $F_{2:4}$ line. Genotype calling was performed using the Axiom Analysis Suite 2.0 with a modified version of the "best practices" workflow. Quality control threshold was reduced to 95 (97 normally), plate pass percent was changed to 90 (95 normally), and average call rate was changed to 97 (98.5 normally). After quality control and genotype calling, a total of 35,143 markers were brought forward with 24,498 segregating in the 44 crosses.

104 Phenotypes

The F_{2:4} lines and agronomic checks were evaluated in 2 by 4 meter harvested plots at 2 105 locations (Cambridge, UK and Duxford, UK) in the 2015-16 growing season, and 2 locations 106 (Hinxton, UK, and Duxford, UK) in the 2016-17 growing season. All locations were managed 107 for optimal yield by following best agronomic practice. All F_{2:4} lines were evaluated in 4 plots. 108 Seed for eleven of the populations was unavailable in the 2015-16 growing season. To 109 110 accommodate these populations and keep the number of plots per line constant, an allocation of F_{2:4} lines was devised that was highly unbalanced across both years and locations as 111 described below. 112

In the 2015-16 growing season, 33 of the 44 populations were planted at two locations 113 (Table 1). The experimental design for both locations was a modified α -lattice design 114 (Patterson and Williams, 1976). The design consisted of a traditional, replicated α -lattice 115 116 design with un-replicated lines added to the sub-blocks. The replicated portion of the alphalattice design was composed of the agronomic checks and half of the lines (34) from 22 of the 117 $F_{2:4}$ populations. These lines were planted in 2 blocks split into 151 sub-blocks each containing 118 5 lines. The remaining $F_{2:4}$ lines were randomly allocated to sub-blocks, bringing the total 119 number of lines per sub-block to either 9 or 10. Half of the F2:4 lines used for the replicated 120 portion of the design differed between locations. Thus lines from 22 of the $F_{2:4}$ populations 121 122 were evaluated in 3 plots split across both locations and the lines from the remaining populations were evaluated in 2 plots split across locations. 123

All 44 populations were planted in the 2016-17 growing season at two locations (Table 1); the experimental design was similar as in the previous season. The replicated portion of the α -lattice design was composed of the agronomic checks and the F_{2:4} lines from the 11 populations not planted in the 2015-16 growing season. These lines were planted in 2 blocks split into 156 sub-blocks each containing 5 lines. Additional F_{2:4} lines from the other populations were randomly allocated to sub-blocks, bring the total number of lines per subblock to 10.

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132 Yield Trial Analysis

Yield phenotypes were spatially adjusted for each trial separately. An AR1 x AR1 model
(Gilmour et al., 1997) was used to adjust spatial variation across both columns and rows as
implemented in ASREML 3.0.22 (Gilmour et al., 2009). A summary of line means after
adjusting for spatial effects is shown in Table 2.

Best linear unbiased estimates (BLUEs) for each line were estimated collectively across alltrials by fitting the following model:

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$$\mathbf{y} = \mathbf{X}\mathbf{b} + \mathbf{u} + \mathbf{e},$$
 (2)

where y was the response vector of spatially adjusted yield values, b site-specific means with
design matrix X, u line BLUEs to estimate, and e the model residual.

142 Genomic prediction

143 This study used the genomic best linear unbiased prediction (GBLUP) model to estimate144 heritabilities and predict line effects. The GBLUP model used was:

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$$\mathbf{y} = \boldsymbol{\mu} + \mathbf{g} + \mathbf{e},\tag{1}$$

where **y** was the response vector of yield BLUEs, μ the model intercept, **g** the vector of genetic values of genotyped F_{2:4} and **e** the model residual. We assumed that $\mathbf{g} \sim N(0, \mathbf{G}\sigma_g^2)$ with genomic relationship matrix calculated as $\mathbf{G} = \mathbf{W}\mathbf{W}'/2\sum p_i(1-p_i)$ (VanRaden, 2008) from the centred genotype matrix **W** and allele frequencies p_i estimated in the dataset. Further, we assumed that $\mathbf{e} \sim N(0, \mathbf{I}\sigma_e^2)$, which was assumed uncorrelated to **g**.

151 The Average-Information Restricted Maximum Likelihood (AI-REML) algorithm 152 (Madsen et al., 1994; Johnson and Thompson, 1995), as implemented in DMU v. 5.1 (Madsen 153 and Jensen, 2000), was used to fit the GBLUP model to a subset of the data (training set) and 154 predict line effects ($\hat{\mathbf{g}}$) in the validation set. We defined convergence of the AI-REML algorithm 155 based on the change of variance components, $|\theta^{(t+1)} - \theta^{(t)}| < 10^{-5}$, where $\theta^{(t)}$ is the vector of 156 normalised variance components estimated at step *t* (Jensen et al., 1997).

157 The heritability was calculated from the trial yield data per plot as $H^2 = \frac{\sigma_g^2}{\sigma_g^2 + \frac{v}{n}}$ in which n 158 is the number of locations in which the genotype was observed and v is the residual variance 159 (Piepho and Mohring, 2007).

160 Prediction accuracies

We applied several cross-validation strategies for investigating prediction accuracies of genomic selection with varying training set size and grouping of training sets and validation sets, as described in detail in the following sections. In all strategies, the GBLUP model was

used as described above. The prediction accuracies were calculated as the Pearson correlation(ρ) between the yield BLUEs and its prediction from the GBLUP model.

166 Cross-validation prediction accuracy

In the first approach, we used 10-fold cross-validation and leave-one-cross-out cross-167 validation (effectively 44-fold cross-validation; refer to Figure 1). Populations were randomly 168 assigned to either training or validation set, without considering that some crosses are more 169 closely related due to sharing a parent or other ancestors. The validation sets were entire 170 populations, which means that line means of a population was confined entirely to either 171 training set or validation set. Prediction accuracies were summarised on a per cross basis and 172 encapsulate the within cross genomic prediction accuracy (sometimes referred to as the within 173 family accuracy or the accuracy of predicting the Mendelian sampling term). For the 10-fold 174 cross-validation, 10 replicates were performed where the 10 folds were re-sampled. 175

To evaluate the effect of training set size, the two cross-validation methods described 176 above were repeated using a subset of the total training set. For the 10-fold cross-validation, 177 178 10%, 20%, ..., 80%, 90% of records were randomly removed from the training set, before estimating variance components and predicting line means of the validation set. For each 179 replicate and the proportion of training set masked, 10 repetitions were performed and the 180 resulting prediction accuracies encapsulate the joint across and within cross genomic 181 prediction accuracy. For the leave-one-cross-out cross-validation, 1-10, 15, 20, 30, 40 crosses 182 were randomly sampled to be used as training set. For each number of crosses sampled as 183 184 training sets, 10, 20, ..., 60, 65 records from each cross was sampled. Again, 10 repetitions were performed. We emphasise that the validation sets were always entire populations (from 185 3-4 crosses in 10-fold cross-validations, from single cross in leave-one-cross-out) and no 186 records of the validated populations were included in the training set. 187

188 Prediction accuracy with related or unrelated crosses

189 In the second approach, we evaluated the prediction accuracies under different levels of 190 relatedness between validation and training sets. The 6 crosses of the 4 most frequently used

191 parents were targeted as validation crosses and tested separately. In summary, the training sets consisted of varying proportions of sister-lines and half-sibs from offspring of either one 192 or both parents or unrelated crosses. Specifically, for each validation cross, training sets were 193 designed to consist of either one or several crosses of one parent, an equal number of crosses 194 from each parent, nominally unrelated crosses, or equal number of related and unrelated 195 crosses. To reduce computation time, for each training set of crosses, 5 combinations were 196 sampled from the large number of possible combinations. For each training set, the validation 197 cross contributed with 0, 1, 2, or 3 quarters of its lines. The prediction accuracies were 198 evaluated for the fourth quarter of lines that were not used in the training set. For each 199 combination of training set, 10 replicates were performed as well as cycling through all four 200 quarters of the validation cross as training set. 201

202 Results

44 bi- and tri-parental crosses from 27 parents were analysed for yield with a GBLUP model
(1), using BLUEs from 4 trials (2 trials in 2016, and 2 trials in 2017).

205 Trait heritability

The overall heritability of yield for all populations over all four trial locations was estimated at
0.65. Heritabilities estimated on single crosses were highly variable, ranging from as low as
0.1 to as high as 0.85 (Figure 2).

209 Cross-validation prediction accuracy

Within cross prediction accuracies were 0.125 – 0.127 using two different cross-validation approaches (Table 3). In these two approaches, all lines of the crosses used for validation were absent from the training set. Using a 10-fold cross-validation approach where individual lines, not all lines of a cross, were selected for validation sets, the prediction accuracy was slightly higher (0.142) when calculated on a per-cross basis ('10-fold, random', Table). Lastly, the prediction accuracy was higher when calculated across all crosses in the validation set, due to capturing variation within and between crosses (0.289 and 0.543, Table 3).

The prediction accuracy was found to increase with training set size. Figure 3 displays 217 the average prediction accuracy across all crosses with 10th and 90th percentile range shown as 218 the greyed area. The prediction accuracy varied greatly between the crosses (Supplemental 219 figure 1) with some accuracies as high as 0.45 (cross 7), as low as -0.20 (cross 30). For 31 220 crosses out of 44, significant positive prediction accuracies were found (Wald's test, p<0.05). 221 Crosses with higher phenotypic variance generally yielded higher predictions; in 222 Supplemental figure 1, prediction accuracy plots for individual crosses are sorted with 223 decreasing phenotypic variance. Finally, the two cross-validation approaches generally 224 produced similar results (Supplemental figure 1), but when the training sets were small, the 225 accuracy of predictions from leave-one-cross-out were less stable than from 10-fold cross 226

validation. The leave-one-cross-out sampled entire crosses in contrast to the 10-fold cross-validation, where lines across all crosses except the validated cross were sampled.

The prediction accuracy increased with an increasing number of crosses in training set or increasing number of lines per cross in training set. Figure displays the average prediction accuracy when sampling a number of lines from a number of crosses (x-axis). Adding an additional 10 or 15 lines to a training set of 50 lines per cross generally led to a low increase in prediction accuracy as compared to adding them to training sets of \leq 40 lines per cross, irrespective of the number of crosses included in the training set.

235 Prediction accuracies with related or unrelated crosses

236 Using related crosses as a training set generally resulted in higher prediction accuracies compared to using unrelated crosses. This is shown in Figure 5, where the green lines (related 237 238 training sets) are above the purple lines (unrelated training sets). Using both related and unrelated crosses in equal proportions (blue lines, Figure 5) led generally to similar 239 correlations to those for related crosses. At approximately 700 to 800 lines in the training set, 240 the prediction accuracy using both related and unrelated crosses plateaued; this was where 241 additional crosses in the training set were unrelated to the validation cross. The level of 242 prediction accuracy of the training set comprising both related and unrelated crosses (lower 243 blue line, Figure) was higher than that in Figure because results in Figure are averages over 244 just 6 crosses rather than over all crosses as in Figure 3. 245

Using only 1, 2, or 3 quarters of the validation cross as training set (grey, horizontal lines, Figure 5) generally led to prediction accuracies that were higher than using a few unrelated or related crosses as the training set. Adding three quarters of the validation cross to the training sets of other crosses generally increased the prediction accuracy, as shown with the upper thick lines in Figure . The gradual increase in prediction accuracy when adding 1, 2, or 3 quarters of the validation cross to the training set is shown in the inserted plot in Figure 5.

252 Discussion

In this study, we have demonstrated the impact of training set size and relatedness on genomic prediction in wheat, using $F_{2:4}$ lines from 44 bi- and tri-parental crosses. The results were consistent with expectations from existing literature (as discussed in the next sections). Specifically, we found that increasing the size of the genomic prediction training set increased accuracy. We also found that training sets composed of lines more closely related to the validation set produce higher prediction accuracies than equivalently sized training sets of more distantly related lines.

It is important for genomic prediction of a complex trait that it displays a reasonable heritability. Our estimate of broad sense heritability for yield (0.65) is well within range of similar studies in wheat (Poland et al., 2012; Combs and Bernardo, 2013; Michel et al., 2016; Schopp et al., 2017; Norman et al., 2017). We note that the heritability values within individual families (Figure) cover the whole range of heritability for this trait reported in the literature.

The various strategies of data subset masking applied in this study has enabled us to 265 266 demonstrate both training set size and relatedness as parameters that influence successful 267 genomic prediction. Generally, increasing the training set size increased the prediction accuracy, as expected from existing theory (Daetwyler et al, 2008, Goddard, 2009, Hickey et 268 al., 2014) and field reports (Liu et al., 2016; Zhang et al., 2017). However, we can add three 269 270 observations that put some nuance to this general conclusion. First (1), with a fixed training set size, it is better to increase the number of populations (crosses) rather than number of lines 271 per population (cross). Second (2), the prediction accuracy plateaus when adding additional 272 crosses that are unrelated to the predicted cross (Figure). Third (3), prediction accuracies vary 273 greatly between individual crosses and this could not be explained by neither the crosses' 274 phenotypic variance nor heritability. 275

For item 1), we showed that, for example, using 10 crosses with 40 lines per cross gave prediction accuracy of ≈ 0.06 , while 40 crosses with 10 lines per cross gave prediction accuracy of ≈ 0.075 (Figure). We assume that in both strategies different processes increase the accuracy with the addition of extra lines: In the first case, entire crosses were masked simulating the future prediction of an unphenotyped cross. In comparison, increasing the number of lines instead of number of crosses (while constraining the training set size) did not necessarily improve the prediction accuracy. The lines capture the crosses' variance, and there will be a limit to how much more variance that additional lines will capture, hence no additional gain. The exception to this was adding fractions of the validation cross' lines to the training set (Figure).

286 For item 2), we saw in Figure that using training sets comprised of exclusively unrelated crosses resulted in lower prediction accuracies than training sets that included related crosses. 287 288 Using training sets comprised of either exclusively related crosses or related and unrelated crosses (half-and-half) both resulted in approximately the same prediction accuracy. The 289 290 comparison between these three sets stops at about 800 lines in the training set, because beyond this point, additional crosses were no longer distinctively related or unrelated. 291 Therefore, after this point the slope of increase in prediction accuracy is less steep, as the 292 crosses added to the training set are less related. 293

For item 3), there was no observable connection between how well the cross could be predicted and the cross' heritability or the observed phenotypic variance. Likewise, these values did not correspond to how well the data from the cross could be used to predict breeding values in other crosses.

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One of the major practical implications of this study is that increased prediction accuracies can be obtained by balancing the training set for genomic selection with phenotypic and genomic data of multiple related crosses, which could be taken into account in advance when designing the training population, as previously proposed by Rincent et al., 2012. For existing data sets, a strategy may be applied of supplementing these with phenotypic data from previous trials (provided genotype-by-environment interaction is limited or can be accounted for by use of trait data for control lines). Although such data might be present within the

context of a rolling breeding program, obtaining genomic data presents a bottleneck as this
requires genotyping of (old) biological material that might not be readily available, and will
require investment in at least low-density genotyping. In case high density genotype data sets
are available for the parental lines, high density genotype information for their offspring
populations can subsequently be obtained by imputation, as reported by Hickey et al. (2015),
Gorjanc et al. (2017) and others.

312 Conclusions

Genomic predictions of yield across 44 populations resulted in modest correlations between observed and predicted values. The correlations did increase with training set size, but by selecting training sets that comprised related crosses improved the correlation more than increasing training set size. The results also showed that if the training set size is fixed, using few lines from more crosses, rather than many lines from few crosses, resulted in higher correlations.

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320 Authors' contributions

Wheat crosses were made by JL, EB, CB, PJ, SB, EF, BP, SS, CH; wheat yield trials were conducted by RJ, PH, EO and IJM; ARB co-ordinated genotyping; SME, RCG and GG performed data analysis; SME, JBB, RCG and JMH wrote the manuscript; JMH and IJM conceived the study, designed the experiment and led the project.

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334 Competing interests

335 The authors declare that they have no competing interest.

336 References

- Allen, A.M., M.O. Winfield, A.J. Burridge, R.C. Downie, H.R. Benbow, G.L.A. Barker, P.A.
 Wilkinson, J. Coghill, C. Waterfall, A. Davassi, G. Scopes, A. Pirani, T. Webster, F.
 Brew, C. Bloor, S. Griffiths, A.R. Bentley, M. Alda, P. Jack, A.L. Phillips, and K.J.
 Edwards. 2016. Characterization of a Wheat Breeders' Array suitable for highthroughput SNP genotyping of global accessions of hexaploid bread wheat (Triticum
 aestivum). Plant Biotechnol. J.Available at http://doi.wiley.com/10.1111/pbi.12635.
- Bernardo, R., and J. Yu. 2007. Prospects for Genomewide Selection for Quantitative Traits in
 Maize. Crop Sci. 47(3): 1082.
- Calus, M.P.L. 2010. Genomic breeding value prediction: methods and procedures. animal 4(02): 157–164.
- de los Campos, G., A.I. Vazquez, R. Fernando, Y.C. Klimentidis, and D. Sorensen. 2013.
 Prediction of Complex Human Traits Using the Genomic Best Linear Unbiased
 Predictor. PLoS Genet 9(7): e1003608.
- Clark, S.A., J.M. Hickey, H.D. Daetwyler, and J.H. van der Werf. 2012. The importance of
 information on relatives for the prediction of genomic breeding values and the
 implications for the makeup of reference data sets in livestock breeding schemes.
 Genet. Sel. Evol. 44(1): 4.
- Combs, E., and R. Bernardo. 2013. Accuracy of Genomewide Selection for Different Traits with
 Constant Population Size, Heritability, and Number of Markers. Plant Genome 6(1).
- Daetwyler H.D., B. Villanueva, J.A. Woolliams (2008) Accuracy of Predicting the Genetic Risk
 of Disease Using a Genome-Wide Approach. PLoS ONE 3(10): e3395
- García-Ruiz, A., J.B. Cole, P.M. VanRaden, G.R. Wiggans, F.J. Ruiz-López, and C.P. Van
 Tassell. 2016. Changes in genetic selection differentials and generation intervals in US
 Holstein dairy cattle as a result of genomic selection. Proc. Natl. Acad. Sci. 113(28):
 E3995-E4004.
- Gaynor, R.C., G. Gorjanc, A.R. Bentley, E.S. Ober, P. Howell, R. Jackson, I.J. Mackay, and J.M.
 Hickey. 2017. A two-part strategy for using genomic selection to develop inbred lines.
 Crop Sci. 57: 1404-1420
- Gilmour, A.R., B.R. Cullis, and A.P. Verbyla. 1997. Accounting for Natural and Extraneous
 Variation in the Analysis of Field Experiments. J. Agric. Biol. Environ. Stat. 2(3): 269–
 293.
- Gilmour, A.R., B.J. Gogel, B.R. Cullis, and R. Thompson. 2009. ASReml User Guide Release
 3.0. VSN International Ltd, Hemel Hempstead, UK.
- Goddard, M. 2009. Genomic selection: prediction of accuracy and maximisation of long term
 response. Genetica 136(2): 245-57
- Gonen, S., R. Ros-Freixedes, M. Battagin, G. Gorjanc, and J.M. Hickey. 2017. A method for the
 allocation of sequencing resources in genotyped livestock populations. Genet. Sel.
 Evol. 49(1)Available at http://gsejournal.biomedcentral.com/articles/10.1186/s12711017-0322-5 (verified 22 May 2017).

- Gorjanc, G., M. Battagin, J.-F. Dumasy, R. Antolin, R.C. Gaynor, and J.M. Hickey. 2017.
 Prospects for Cost-Effective Genomic Selection via Accurate Within-Family
 Imputation. Crop Sci. 57(1): 216.
- Gorjanc, G., M.A. Cleveland, R.D. Houston, and J.M. Hickey. 2015. Potential of genotyping by-sequencing for genomic selection in livestock populations. Genet. Sel. Evol. 47(1):
 12.
- Habier, D., R.L. Fernando, and J.C.M. Dekkers. 2007. The impact of genetic relationship
 information on genome-assisted breeding values. Genetics 177(4): 2389–2397.
- Habier, D., J. Tetens, F.-R. Seefried, P. Lichtner, and G. Thaller. 2010. The impact of genetic
 relationship information on genomic breeding values in German Holstein cattle.
 Genet. Sel. Evol. 42(1): 5.
- Hickey, J.M., S. Dreisigacker, J. Crossa, S. Hearne, R. Babu, B.M. Prasanna, M. Grondona, A.
 Zambelli, V.S. Windhausen, K. Mathews, and G. Gorjanc. 2014. Evaluation of genomic selection training population designs and genotyping strategies in plant breeding programs using simulation. Crop Sci. 54: 1476–1488.
- Hickey JM, G. Gorjanc, T.K. Varshney and C. Nettelblad (2015) Imputation of Single
 Nucleotide Polymorphism Genotypes in Biparental, Backcross, and Topcross
 Populations with a Hidden Markov Model Crop Science 55: 1934-1946

- Jenko, J., G.R. Wiggans, T.A. Cooper, S. a. E. Eaglen, W.G. de L. Luff, M. Bichard, R. PongWong, and J.A. Woolliams. 2017. Cow genotyping strategies for genomic selection in a
 small dairy cattle population. J. Dairy Sci. 100(1): 439–452.
- Jensen, J., E.A. Mantysaari, P. Madsen, and R. Thompson. 1997. Residual Maximum
 Likelihood Estimation of (Co) Variance Components in Multivariate Mixed Linear
 Models using Average Information. J. Indian Soc. Agric. Stat. 49: 215–236.
- Johnson, D.L., and R. Thompson. 1995. Restricted Maximum Likelihood Estimation of
 Variance Components for Univariate Animal Models Using Sparse Matrix Techniques
 and Average Information. J. Dairy Sci. 78(2): 449–456.
- Liu, G., Y. Zhao, M. Gowda, C.F.H. Longin, J.C. Reif, and M.F. Mette. 2016. Predicting Hybrid
 Performances for Quality Traits through Genomic-Assisted Approaches in Central
 European Wheat (L Lukens, Ed.). PLOS ONE 11(7): e0158635.
- Mackay, I., E. Ober, and J. Hickey. 2015. GplusE: beyond genomic selection. Food Energy
 Secur. 4(1): 25–35.
- 410 Madsen, P., and J. Jensen. 2000. A User's Guide to DMU. A Package for Analysing
 411 Multivariate Mixed Models. Version 6, release 5.1.: 32.
- Madsen, P., J. Jensen, and R. Thompson. 1994. Estimation of (co)variance components by
 REML in multivariate mixed linear models using average of observed and expected
 information. p. 455–462. *In* 5th WCGALP. Guelph, Canada.
- 415 Meuwissen, T.H. 2009. Accuracy of breeding values of "unrelated" individuals predicted by
 416 dense SNP genotyping. Genet. Sel. Evol. 41(1): 35.
- 417 Michel, S., C. Ametz, H. Gungor, D. Epure, H. Grausgruber, F. Löschenberger, and H.
 418 Buerstmayr. 2016. Genomic selection across multiple breeding cycles in applied bread
 419 wheat breeding. Theor. Appl. Genet. 129(6): 1179–1189.

- Neyhart, J.L., T. Tiede, A.J. Lorenz, and K.P. Smith. 2017. Evaluating Methods of Updating
 Training Data in Long-Term Genomewide Selection. G3amp58
 GenesGenomesGenetics 7(5): 1499–1510.
- 423 Norman, A., J. Taylor, E. Tanaka, P. Telfer, J. Edwards, J.-P. Martinant, and H. Kuchel. 2017.
 424 Increased genomic prediction accuracy in wheat breeding using a large Australian 425 panel. Theor. Appl. Genet. 130(12): 2543–2555.
- Patterson, H.D., and E.R. Williams. 1976. A new class of resolvable incomplete block designs.
 Biometrika 63: 83–92.
- Piepho, H. and J. Mohring. 2007. Computing heritability and selection response from unbalanced plant breeding trials. Genetics 177: 1881–1888.
- Poland, J.A., J. Endelman, J. Rutkoski, S. Wu, Y. Manes, S. Dreisigacker, J. Crossa, H.
 Sánchez-Villeda, M. Sorrells, and J.-L. Jannink. 2012. Genomic Selection in Wheat
 Breeding using Genotyping-by-Sequencing. Plant Genome J. 5(3): 103.
- Pszczola, M., and M.P.L. Calus. 2015. Updating the reference population to achieve constant
 genomic prediction reliability across generations. animal: 1–7.
- Pszczola, M., T. Strabel, H.A. Mulder, and M.P.L. Calus. 2012. Reliability of direct genomic values for animals with different relationships within and to the reference population.
 J. Dairy Sci. 95(1): 389–400.
- R. Rincent, D. Laloë, et al. 20112. Maximizing the reliability of genomic selection by
 optimizing the calibration set of reference individuals: Comparison of methods in two
 diverse groups of maize inbreds (*Zea mays* L.). Genetics 192: 715-728
- 441
 442 Ros-Freixedes, R., S. Gonen, G. Gorjanc, and J.M. Hickey. 2017. A method for allocating low443 coverage sequencing resources by targeting haplotypes rather than individuals. Genet.
 444 Sel. Evol. 49(1): 78.
- Schopp, P., D. Müller, Y.C.J. Wientjes, and A.E. Melchinger. 2017. Genomic Prediction Within
 and Across Biparental Families: Means and Variances of Prediction Accuracy and
 Usefulness of Deterministic Equations. G3amp58 GenesGenomesGenetics:
 g3.300076.2017.
- 449 VanRaden, P.M. 2008. Efficient methods to compute genomic predictions. J. Dairy Sci. 91(11):
 450 4414–23.
- Zhang, A., H. Wang, Y. Beyene, K. Semagn, Y. Liu, S. Cao, Z. Cui, Y. Ruan, J. Burgueño, F. San
 Vicente, M. Olsen, B.M. Prasanna, J. Crossa, H. Yu, and X. Zhang. 2017. Effect of Trait
 Heritability, Training Population Size and Marker Density on Genomic Prediction
 Accuracy Estimation in 22 bi-parental Tropical Maize Populations. Front. Plant Sci.
 8Available at http://journal.frontiersin.org/article/10.3389/fpls.2017.01916/full
 (verified 22 November 2017).
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458

460 Figures

	2015/2016		2016/2017	
# Lines	Cambridge	Duxford	Duxford	Hinxton
367	2	1	1	0
381	2	1	0	1
381	1	2	1	0
367	1	2	0	1
748	1	1	1	1
748	0	0	2	2
Total plots	2992	2992	2992	2992

461 Table 1: Trial design summary showing number of plots per tested line per location

462

463 Table 2: Summary of line means per location after adjusting for spatial effects.

		No. lines	Avg. value	Coef. Variation	Correlation [†]
2016	Cambridge	2,247	8.58	6.1%	0.63
2016	Duxford	2,248	10.82	6.3%	0.81
2017	Hinxton	2,249	4.64	10.3%	0.71
2017	Duxford	2,235	8.24	6.6%	0.62

464 [†]: Correlation between moisture corrected yield values and spatially adjusted values.

465 Table 3: Prediction accuracies using the largest training sets by cross-validation

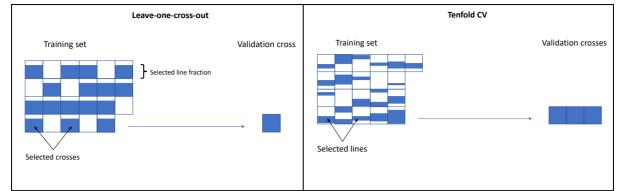
466 **approach.**

	Correlation metric	Training set size	Correlation ⁺
Leave-one-cross- out	By cross	2,787	0.127 0.222
10-fold, crosses	By cross	2,563	0.125 0.193
10-fold, random‡	By cross	2,567	0.142 0.195
10-fold, crosses	Across all ?	2,567	$0.289_{\ 0.259}$
10-fold, random ‡	Across all ?	2,567	0.543 0.009

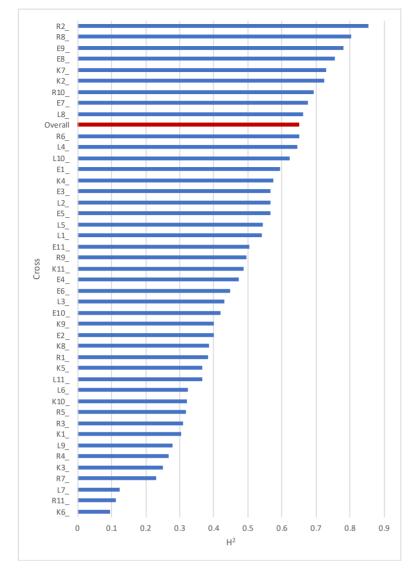
467 *†*: Average across all replicates. Small font displays inter-quantile range for correlations.

468 ***: 10-fold cross-validation where validation and training sets were grouped by lines instead of crosses.

469 ?: Correlations were calculated across multiple crosses in validation set.

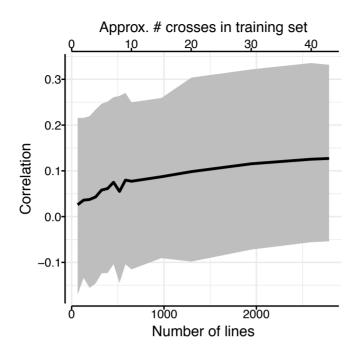


- 471 **Figure 1**: Resampling strategies applied to assess the impact of training set design. Leave-
- 472 one-cross out strategy (left) tests the impact of inclusion of the amount of crosses as well as
- training set size, while the ten-fold cross validation (right) tests training set size only.



476 Figure 2: Yield heritabilities when estimated per cross. Crosses (blue bars) are ordered by

477 heritability value, overall heritability for this trait is shown in red.

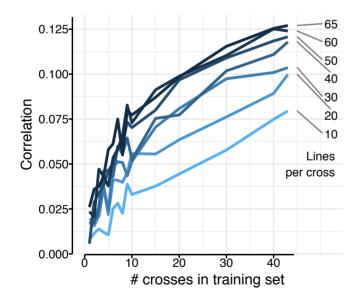




479 Figure 3: Increasing training set size increased prediction accuracy (correlation). Solid

480 line shows average of all leave-one-cross-out cross-validations with 10th and 90th percentile range shown

- 481 by greyed area.
- 482





484 Figure 4: Prediction accuracies increased with the increasing number of crosses or the
485 increasing number of lines per cross in training set. Right-hand numbers show number of lines
486 per cross in training set.

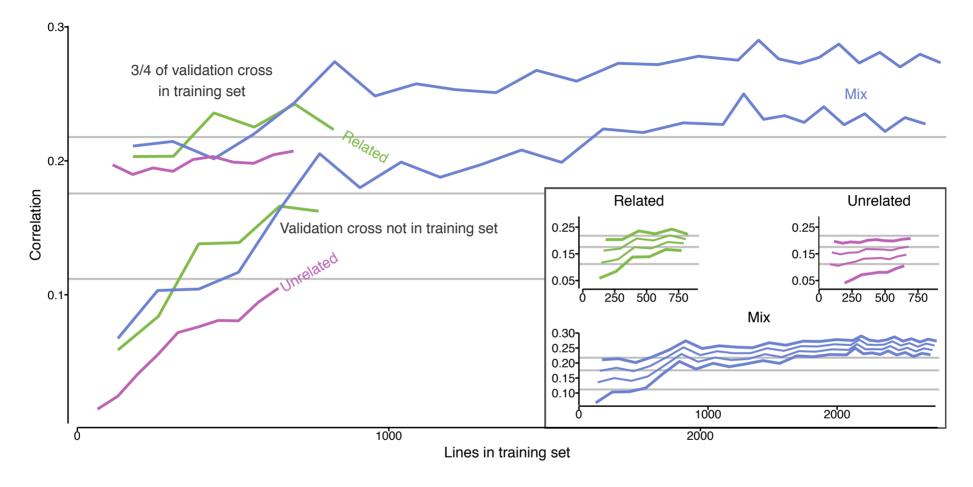
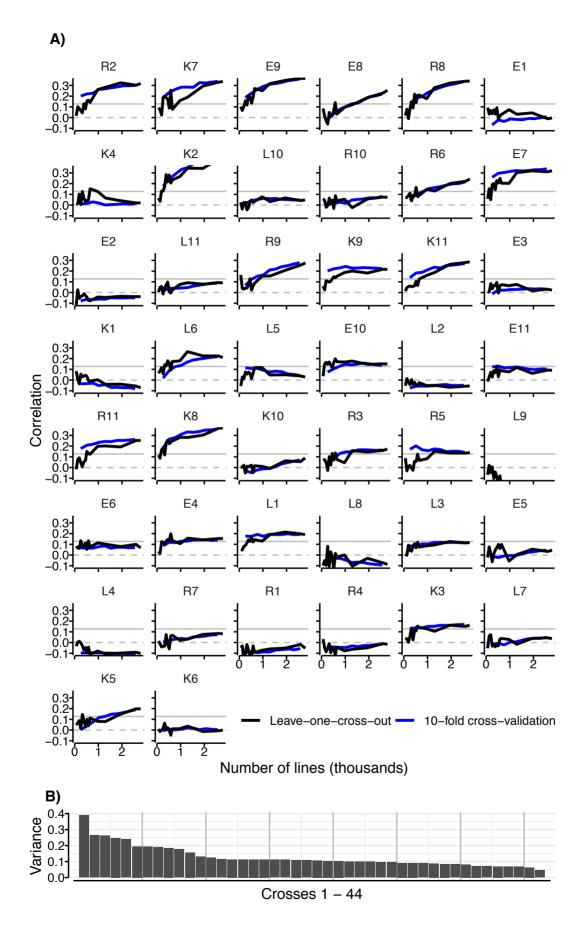


Figure 5: Prediction accuracies increased when the validation cross was partly in training set or had its related crosses in training set. Results show average prediction accuracies for 6 validation crosses. Lines show prediction accuracies when training set comprised of related crosses (green solid line), unrelated crosses (purple dashed line), or a mix of both (blue dotted line). Lower set of lines show prediction accuracies when validation crosses were not included on the training set; upper set of lines show prediction accuracies when validation crosses were included in the training set with 3/4 of lines. Grey horizontal lines show average prediction accuracy using *only* 1/4, 2/4, or 3/4 of validation cross as training set. Inserted figure shows the increase in accuracy when adding 1/4, 2/4, and 3/4 of the validation group to the training set. The thick lines in the inserted figure denote the lines of the main figure.

495 Supplementary materials



497 Supplemental figure 1: Per-cross correlation under two approaches (A), ordered by

- 498 decreasing variance of crosses' BLUEs (B). Grey, horizontal lines are guides for zero correlation
- 499 (dashed) and overall average correlation of 0.127 (solid). Crosses in A) are ordered with decreasing
- 500 variance of their BLUEs, same order as in B).