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- 2 CHARACTERISING GENETIC DIVERSITY IN CASSAVA BROWN STREAK
- 3 VIRUS

# Characterising Genetic Diversity in Cassava Brown Streak Virus

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Abstract.— Plant viruses represent a significant threat to food security for many global populations. Cassava Brown Streak Virus (CBSV) causes immense damage to cassava crops in Eastern, Central and Southern Africa. The eradication of CBSV is a difficult challenge, as it has been shown to be fast-evolving and it is transmitted by flying insects that are ubiquitous in cassava growing regions. In this paper we demonstrate the ability of two new developments in bioinformatics that can be used to increase our understanding of CBSV and ultimately inform strategies for its combat. We reconstruct the phylogeny of 29 whole-genome virus isolates using the GHOST model. This phylogeny identifies three distinct clades among the viruses and highlights a section of the genomes that is highly influential in their divergence. We also perform Multiple Correspondence Analysis on the alignment which is consistent in recovering the three clades, and offers insight on the significance of the influence of a variety of external variables on the evolution of the viruses. Knowledge and information from this analysis will be used as a base on which to formulate sustainable Cassava Brown Streak Disease (CBSD) management strategies in Africa.

### Introduction

It is difficult to overemphasize the importance of agriculture to people living in

sub-Saharan Africa. The population of sub-Saharan Africa exceeds one billion people, and more than 60% of the population live in rural areas (United Nations, 2015). Beyond the obvious benefit of nutrition, agriculture is also the primary source of income for the majority of sub-Saharan African families, and so access to essentials such as health care and education depend indirectly on agriculture. As such, the productivity of the agricultural industry in a given country and year is a key indicator of the health and wellbeing of the population and the economy. One of the major threats to agricultural productivity comes in the form of crop pests and diseases. Of particular significance are the viruses of the genus *Ipomovirus.* They have the potential to devastate crops, and they infect some of the most economically important, commonly grown food staples across sub-Saharan Africa, such as cassava and sweet potato. Cassava Brown Streak Disease (CBSD) is caused by two closely related *Ipomovirus* species, Cassava Brown Streak Virus (CBSV) and Ugandan Cassava Brown Streak Virus (UCBSV). CBSD is widespread and causes significant reduction in both the quality and the yield of cassava crops, making it a strong barrier to food security and economic prosperity in the regions in which it is grown. Efforts to combat CBSV and UCBSV are ongoing, but there are many 58

Efforts to combat CBSV and UCBSV are ongoing, but there are many challenges. The viruses are transmitted between plants courtesy of whiteflies

(Bemesia tabaci) and other potential vectors (Ateka et al., 2017), so it is critical
that infected plants are removed from crops swiftly. This indicates a critical need
for early detection and positive diagnosis. However, traditional sequencing methods
such as PCR and Sanger sequencing are slow and costly, with a positive diagnosis
taking weeks or months, far too late to prevent the virus from spreading. A recent
advance in sequencing technology has seen researchers use nanopore sequencing to
take samples from plants, and then sequence and identify pathogens in just a few
hours, without the need to leave a farmer's field (Boykin et al., 2018). This enables
the swift quarantine of infected material, protecting the rest of their crop and
neighbouring crops.

Efforts to isolate and destroy infected plant material are on the frontline of
the battle against CBSV and UCBSV, but they are reactive strategies in that they
combat the virus after plant infection has already occurred. It is necessary to
simultaneously pursue proactive strategies, such as breeding resistant forms of
cassava. This is not straightforward though, as it has already been shown that
these viruses are fast-evolving (CBSV moreso than UCBSV) (Alicai et al., 2016).
Mbewe et al. (2017) showed that in the gene tree of the P1 gene, a third distinct
clade is found in addition to CBSV and UCBSV, which they tentatively labelled
Tanzanian CBSV (CBSV-TZ). Critical to winning the fight against CBSD is

ongoing research into the evolutionary forces acting on these viruses, at both the molecular level and geographically.

We made use of two new bioinformatics tools to analyse a sequence 81 alignment consisting of 29 whole virus genomes, 14 CBSV and 15 UCBSV. We first performed phylogenetic inference under the GHOST model of sequence evolution (Crotty et al., 2017). The GHOST model is a mixture model which enables the fitting of multiple classes to a single alignment. The tree topology is common across all classes, but each class has its own set of branch lengths and model parameters. This enables subtle phylogenetic signals to be extracted from the data, which is especially useful when there exists a clear dominant signal, such as that distinguishing the two virus species in our alignment. We also applied Multiple Correspondence Analysis (MCA) to the alignment, following the method outlined in Rohrlach et al. (2018). They demonstrated MCA as an effective tool for decomposing the variability within categorical sequence data for visualization in two dimensions. Furthermore, they showed that it was possible to meaningfully associate the genetic diversity with geographical coordinates, allowing for informed hypotheses relating to the path of evolution to be constructed.

Methods

Data

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The alignment consisted of 14 CBSV and 15 UCBSV whole genome sequences. The alignment was obtained from Alicai et al. (2016), in which details of its assembly can be found.

#### Phylogenetic Analysis

We used IQ-TREE (Nguyen et al., 2015) to fit a GHOST model to the sequence alignment. We used the model selection procedure outlined in Crotty et al. (2017) to choose the model of sequence evolution and number of classes. After performing the inference we analysed the site-wise probabilities of evolving under each inferred class to identify influential sites, contiguous regions and genes within the genome.

#### Multiple Correspondence Analysis

We conducted MCA following the procedure outlined in Rohrlach et al. (2018).

Due to the fast rate of evolution present we removed all singleton sites: sites where

the nucleotide was conserved across all but one taxon. We carried out MCA on the

entire sequence alignment, as well as on the 14 CBSV taxa and 15 UCBSV taxa

separately.

RESULTS

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#### Phylogenetic Analysis

Model selection.— In order to determine the optimal model of sequence evolution and number of classes when fitting the GHOST model, we experimented with a 117 wide variety of substitution models, fitting each one with between two and twelve 118 classes. We then identified the best fitting combinations according to both Bayesian 119 Information Criterion (BIC) (Schwarz et al., 1978) and Akaike's Information 120 Criterion (AIC) (Akaike, 1974). Results can be found in Supplementary Figures S1 121 and S2. The smallest BIC score was obtained using the General Time Reversible 122 (GTR) model and three classes, while the smallest AIC score was obtained using 123 GTR with ten classes. It is not unexpected to see such a discrepancy with the parameter-rich GHOST model. BIC places a relatively heavy penalty on additional 125 parameters and is therefore prone to underfitting, whereas AIC imposes a relatively 126 light penalty and is therefore prone to overfitting (Burnham and Anderson, 2003; 127 Posada and Buckley, 2004; Dziak et al., 2012).

Examining the recovered trees from the 3-class GHOST model (Figure 1)
preferred by BIC, distinct and reasonable biological interpretations can be made for
all three classes. The first class has very short branch lengths, indicating strong

conservation of nucleotides across all taxa. Since these viruses are closely related, 132 we can think of this class as capturing the component of the phylogenetic signal 133 that is common to CBSV and UCBSV. The second class shows only one branch 134 length of any significance, that separating the CBSV and UCBSV clades. Within 135 the two clades the branch lengths are again very short, indicating that this class 136 captures the component of the phylogenetic signal relating to the divergence of 137 CBSV and UCBSV. The third class distinguishes itself from the second by a clade 138 of four CBSV replicates that diverges from both UCBSV and the remaining CBSV 139 replicates. 140

Unlike the trees of the 3-class model, the trees of the 10-class model 141 (Supplementary Figure S3) preferred by AIC bear the hallmarks of overfitting, as 142 described in Crotty et al. (2017). Many of the classes bear strong similarity to each 143 other, most noticeably the first three classes all appear strongly conserved across all 144 taxa, much like the first class of the 3-class model. In fact, it is not unreasonable to 145 suggest that all 10 class trees could be loosely categorised as falling into one of the 146 three categories defined above by the trees of the 3-class model. No significant new 147 phylogenetic signal appears to be captured by this model, beyond those which were 148 found by the 3-class model. Consequently, we concluded that the 3-class GHOST 149 model with a GTR model of sequence evolution provided the best fit to this

Figure 1: Trees inferred by IQ-TREE using the GHOST model with three GTR classes.

alignment.

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Identifying regions of interest.— Having chosen the optimal GHOST model, we can use the results to identify regions within the genome that may be influential in the 153 patterns of evolution that we observe. For convenience, we refer to the first class as 154 the Common Class, since there is no significant divergence among any of the taxa; 155 we refer to the second class as the UCBSV Class, since this class is characterised by the divergence between the CBSV and UCBSV replicates; and we refer to the third class as the CBSV-TZ Class. The reason for this is that the four CBSV replicates that diverge from the remainder in this class are also present in the study of the P1 159 gene (Mbewe et al., 2017) referred to earlier. These four replicates are found in the 160 third clade which they tentatively labelled CBSV-TZ, and so it seems probable 161 that the phylogenetic signal captured in the third class is related to the divergence 162 of CBSV-TZ from CBSV. It is worth noting here that we do not make the claim 163 that CBSV-TZ is a new species of CBSV, but use these terms for convenience in 164 labelling the classes, and consistency with the work of Mbewe et al. (2017). Further 165 research is required in order to establish CBSV-TZ as a new vspecies of the 166 virus. 167

The contribution of a particular site in the alignment to the likelihood

function of a mixture model such as GHOST is simply the weighted sum of the partial likelihoods of that site under each class. We can make use of these partial 170 likelihoods to determine for every site the probability distribution of evolving under 171 each of the classes. Put simply, we can look at each site and identify the 172 probability with which it belongs to any of the classes. This allows us to locate 173 particular sites, contiguous regions and genes that belong to the CBSV-TZ class 174 with high probability, thereby being identified as influential in the evolutionary 175 divergence of CBSV-TZ and CBSV. In Table 1 we list the ten genes that 176 consititute the genome of the viruses, the length of the genes in nucleotides as well 177 as in percentage of the genome, and finally the percentage contribution of the genes 178 towards the CBSV-TZ Class. Most apparent from the Table 1 is that the P1 gene 179 appears to be of strong influence in the divergence of CBSV-TZ and CBSV. The 180 1086 sites that make up the P1 gene, representing 12.41% of the entire genome, 181 account for more than 22% of the weight of the CBSV-TZ Class. The remaining 182 genes contribute to the CBSV-TZ Class approximately in proportion with their 183 relative length in the genome, with the exception of the CP gene which constitutes 12.96% of the genome but only accounts for 6.57% of the CBSV-TZ Class. 185 In Figure 2, we show the cumulative probability of sites in the P1 gene 186

belonging to the CBSV-TZ Class. It displays graphically the observation made 187

Gene	Size	% Genome	% CBSV-TZ Class
P1	1086	12.41	22.08
Р3	882	10.08	10.37
6K1	156	1.78	1.91
CI	1890	21.61	20.27
6K2	156	1.78	1.86
Vpg	558	6.38	7.19
NIa	702	8.03	8.45
NIb	1506	17.22	15.61
Ham1	678	7.75	5.69
СР	1134	12.96	6.57
Total	8748	100	100

Table 1: A list of the 10 genes that make up CBSV and UCBSV. Also shown is the size of each gene in units of nucleotides and as a percentage of the entire genome. The final column shows the percentage of the CBSV-TZ Class attributable to each gene.

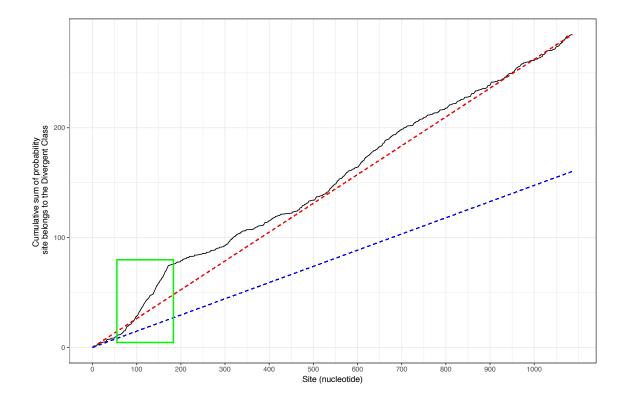


Figure 2: A cumulative sum plot of the probability of sites belonging to the CBSV-TZ Class, for the P1 gene. The average gradient in any contiguous region is representative of the contribution of this region to the CBSV-TZ Class. The red dashed line indicates the average probability of the sites in the P1 gene belonging to the CBSV-TZ Class. The blue dashed line indicates the average probability of all sites in the genome belonging to the CBSV-TZ Class. The green box highlights a section of the gene in which the gradient is particularly steep, indicating that this region contributes strongly to the CBSV-TZ Class.

from Table 1, that the average gradient of the P1 gene (represented by the red 188 dotted line) is nearly double that of the average gradient across the entire genome 189 (represented by the blue dotted line). Additionally, Figure 2 also demonstrates that 190 the contribution to the CBSV-TZ Class is not even across the entire gene. 191 Compared to the average for the entire gene, there is a noticeable increase in 192 gradient, roughly between nucleotide 70 and 170. We therefore surmise this to be 193 the region of highest influence within the gene of highest influence. Figures 3-5 194 show the alignment for the first 80 amino acids of the P1 gene. Examining the 195 region of interest identified above (amino acids 23 to 57, approximately), we notice 196 in Figure 3 that there is strong conservation among the CBSV isolates, yet the 197 UCBSV and CBSV-TZ isolates are relatively diverged. Similarly, Figure 4 shows 198 that there is also strong conservation among the 15 UCBSV isolates in this region. 199 In contrast, Figure 5 shows comparatively little conservation among the four 200 CBSV-TZ isolates. In summary, while all three strains are strongly divergent from each other in this region, within group genetic variability is low in CBSV and 202 UCBSV but high in CBSV-TZ.

## MCA analysis

When applied to the entire dataset, Figure 6 shows that the distinction between

	101	201	201	405	501	cav	701	16
	10V	201		405	50L	60K	701	1
CBSV_KR108828	MTTIQLFKTVQFGSF							YCRSY
CBSV_LG577537		D .	. V Y		A N			
CBSV_LG577538			. V Y		A NN .			
CBSV_KR108832		V D\	/ Y	S	A N			
CBSV_GQ329864					A N			
CBSV_KR108829		V D .	Y	V	. S A L G			
CBSV_KR108830								
CBSV_KR108831								
CBSV_FN434436		. L V D .	Y		R . A NN .	.Q		
CBSV_KR108833	. A	V K D .	L T N	. X X K . V .	T . K			
CBSV-TZ_GU563327	. A V I	V K . DN E F	QN . K F	NEELTVVSEVE	. KKVCEK.Q.ST	VXXXX	. F V	1
CBSV-TZ_KR108834	. A V I	V K . D S E F	VL.D.KI	TEETTPIGEVE	KKEVCGK.Q.LT	TXXXX	. F V	1
CBSV-TZ_FN434437	. A V I	V K . D S E F	V N . K F	NEE.KITSEVE	KKEVCEK.Q.LI	TXXXX	. F V	1
CBSV-TZ_HG965221	. A S V . V I	V K F D D E S	S . G . T . E A	. K . L S K A S G E K	E E HQ T Q Q . E R	VXXXX.SV.	V V	1
UCBSV_KR108836	. S I R	VKLDEGNN\	/ . E . V D	L L A G N D G S G P E	. QT EQKQHRKES	GESWR.VT.	V V	1
UCBSV_KR108838	. S I T	VKLDEGNNI	I . E V D	LLTGND.FGPE.	. Q S EQ K Y H R K E S	GESWR.VT.	V V	1
UCBSV_KR108837	. S I T	VKLDEGNNI	I . E V D	LLTGND.FGPE	. Q S EQKYHRKES	GESWR.VT.	V V	1
UCBSV_KR108839	. S I T	VKLDEGNNI	I . E T D	LQTGNDGSGPE	. Q S EQ K Y H R K E S	GESWR.VT.	V V	1
UCBSV_KR108835	. S I R	VKLDEGNN\	/ . E . V D	LLAGNDGSGPE	.QSEQKHH.GES	G.SWR.VT.	V	1
UCBSV_HG965222	. S I R	VKLDEGNN\	/ . E . V . I D	L L A G N D G S G P E	. Q S EQ K Y H R K E S	GESWR.VT.	V V	1
UCBSV_LT577539	. S I R	VKLDEGNN\	/ . E . V . I D	LLAGNDGSGPE	. Q S EQKYHRKES	GESWR.VT.	V V	1
UCBSV_NC014791	. S I T	K L D E E N N I	I . E T D	LLAGNDGSGPE	GQSEQKYHRKES	GESWR.VT.	V V	1
UCBSV_FN434109	. S I R	VKLDEGNN\	/ V E V D	LLAGNDGSGPE	. Q S EQKYHRKES	GESWR.VT.	V V	1
UCBSV_FN433931	. S I K	VKLDEGNN\	/ . E . V . V D	LLAGNDGSGPE	. Q S EQ K H H R K E S	GESWR.VT.	V V	1
UCSBV_FN433930	. S I R	VKLDEGNN\	/VE.V.VD	L L A G N D G S G P E	. Q S EQKYHRKES	GESWR.VT.	V V	1
UCBSV_FN433932	. S I R	VKLDEGNN\	/VE.V.VD	LLAGNDGSGPE	. Q S EQ K Y H R K E S	GESWR.VT.	V V	1
UCBSV_FN433933	. S I R	VKLDEGNN\	/ V E V D	L L A G N D G S G P E	. Q S EQKYHRKES	GESWR.VT.	V V	1
UCBSV_FJ039520	. S I T							
UCBSV_FJ185044	. S I T	KLDEENN	I . E T D	LLAGNDGSGPE	GQSEQKYHRKES	GESWR.VT.	V V	1

Figure 3: The first 80 amino acid residues in the alignment of the P1 gene of the 29 CBSV and UCBSV sequences. The top CBSV sequence is used as a reference, with the conserved residues represented as dots.

10I 20L	301	40G	50K	60W	701
CBSV_KR108828 .TVQTIECTT		NA EG SRNA L EN \		KVKF.PI.	
CBSV LG577537 .TVQTIECTT					
CBSV_LG577538 . T VQ T   ECTT	GEVPAYIAN	NAEGSRNALENA	AP E L V S G N N N C	KVKF.PI.	
CBSV KR108832 . T VQ VT   ECTT	DV . P AY I A	SAEGSRNALENA	PELVSGNDNO	KVKF.PI.	
CBSV_GQ329864 .TVQTIECTT	DEVP AY I A!	NAEGSRNALENA	PELVSGNDNO	KVKF.PI.	1 1
CBSV KR108829 .TVQVT   ECTT	DE.PAYIAN	N V E G S R N A L E S A	LELVSGGDNO	KVKF.PI.	
CBSV_KR108830 . T VQ T   ECTT	DEVLAYIA!	NAEGSRNALENA	PELVSGNDNO	KVKF.PI.	1 1
CBSV_KR108831 .TVQVT   ECTT	DE.PAYIAN	NAEGSRNALESA	A P E L V S G N X N C	KVKF.PV.	l l
CBSV_FN434436 .TVQLVTIECTT	DE.PAYIAN	NAEGSRNALRNA	APELVSGNNNO	QVKF.PI.	1 1
CBSV_KR108833 .AVQVTIKCTT	DE.LNIX)	X A E G S K N V L E N \	PELVTG. DNO	KVKF.PI.	l l
CBSV-TZ_GU563327 . A V Q V T I K C D .	EP.PQN.KFNEI	ELTVV.EV.EK	V C E K S Q S S T \	XXXX.PI.	F . I
CBSV-TZ_KR108834 .AVQVT   K C D S	EPVL.D.KITE	ETTPIGEV.KKI	V C G K S Q S L T 1	TXXXX.PI.	F . I
CBSV-TZ_FN434437 . A V Q V T I K C D S	EPVP.N.KFNEI	EAKIT.EV.KKI	V C E K S Q S L I T	TXXXX.PI.	F . I
CBSV-TZ_HG965221 .ASV.VQVTIKFDD	ES.G.T.EAIK!	NLSKAEKENI	. HQTQQS . R \	XXXX.SV.	
UCBSV_KR108836	. V V . A	E . 1	Q		
UCBSV_KR108838	V 1	T S F E			
UCBSV_KR108837	V 1	T S F E			
UCBSV_KR108839	Q1	T E			
UCBSV_KR108835 R V	. V V . A	E	H . GG	. K	1
UCBSV_HG965222	. V V . I	E			
UCBSV_LT577539	. V V . I	E			
UCBSV_NC014791					
UCBSV_FN434109	. V V V	E			
UCBSV_FN433931	. V V . V	E	H		
UCSBV_FN433930					
UCBSV_FN433932 R V	. V V V . V	E			
UCBSV_FN433933RVG.					
UCBSV_FJ039520	V M1	T . D . S F E	Кн		
UCBSV_FJ185044 MSTIQLFKTITFGSFEPIKLDEEN	NIIEKIPTDLLA	A G N D G S G P E G Q S	EQKYHRK E S (	GESWRKVT	DLYSVIGNSVYCRSY

Figure 4: The first 80 amino acid residues in the alignment of the P1 gene of the 29 CBSV and UCBSV sequences. The bottom UCBSV sequence is used as a reference, with the conserved residues represented as dots.

		101	201	30N	40T	50E	60X	701
								100
CBSV_TZ_Tan_70_FN434437	MATIQV	FKTIQFGSFEP	VTIKCDSEP	V P K N P K F N E E	EAKITSEVEKK	EVCEK SQ S L I T	XXXXKPIDL	FSIIGNSVYCRSY
CBSV_TZ_Tan_19_1_KR108834				. L . D I T	TTP   G	G T .		
CBSV_TZ_NaI_07_HG965221	S V		F . D . S	IG.T.EAIKN	ILSKA.GEKEN	. EHQTQ ERV	/ S V <sup>1</sup>	Y . V
CBSV_TZ_KoR6_GU_563327			N	I . Q	LTVVE.	K ST\	1	

Figure 5: The first 80 amino acid residues in the alignment of the P1 gene of the 4 CBSV-TZ sequences. The top CBSV-TZ sequence is used as a reference, with the conserved residues represented as dots.

the three strains is the dominant feature that is captured by MCA. The first
principal dimension clearly distinguishes between CBSV (including the four
CBSV-TZ sequences), while the second dimension distinguishes between CBSV and
CBSV-TZ. The signals that distinguish the three strains are so dominant that it is
difficult to observe anything additional from performing MCA on the entire
dataset. To search for more nuanced patterns in the alignment, we performed MCA
on meaningful subsets of the genomes.

Figure 7 displays four scatterplots of the first two dimensions of a MCA
performed on the 14 CBSV isolates. Each scatterplot is coloured according to a
different variable, to aid in highlighting any patterns in the data. Once again, the
first principal dimension captures the most dominant feature, the distinction
between the CBSV and CBSV-TZ strains. A clear pattern is not obvious in Figures
7 a) and b), which colour the points according to the geographic origin of the
isolates (latitude and longtitude). The points in Figure 7 c) are coloured according
to the sample collection date. Most samples in the dataset were collected either in

2013 (15 of 29), or in 2006 - 2008 (11 of 29). We therefore used binary colouring according to these two groups. The striking feature of Figure 7c), is that most of 222 the CBSV-TZ isolates were collected prior to 2009 (3 of 4), whereas most of the 223 CBSV isolates (8 of 10) were collected in 2013. That is, 60% of the samples 224 collected before 2009 were CBSV-TZ, whereas only 11% of the samples collected in 225 2013 were CBSV-TZ. A similar pattern emerges when observing Figure 7 d), in 226 which points are classified as either lowlands (less than 1000m above sea level) or 227 highlands (more than 1000m above sea level). The threshold was chosen as 228 historically, CBSD was not found in areas that were greater than 1000m above sea 229 level (Alicai et al., 2007). We see from Figure 7 d) that all of the CBSV-TZ isolates 230 came from lowland areas, while the CBSV isolates are approximately evenly split 231 between lowland and highland areas. Figure 8 is analogous to Figure 7 except that the four CBSV-TZ isolates 233 have been removed from the analysis. It is here that we can first see some 234 relationship between the MCA results and the geographic origin of the isolates. 235 Figures 8 a) and b) show that the cluster of four isolates in the top left corner of 236

Victoria, in Uganda and Northern Tanzania.

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the plots are at the extreme ends of the latitude (northernmost) and longtitude

(westernmost) scales. These four isolates come from the areas that surround Lake

Figure 9 displays the MCA results for the 15 UCBSV isolates. The most 240 obvious feature is that two isolates strongly distinguish themselves from the 241 remaining 13 along the first principal dimension. Even though these two isolates 242 are clearly distinct from the others, we can see in Figure 6 that they obviously still 243 belong to the UCBSV cluster. What is most intriguing about these two outliers, is that they appear to share very little in common. Figure 9 indicates that they are 245 separated temporally, geographically and altitudinally. There doesn't appear to be 246 any other strong patterns emerging from the MCA on the UCBSV isolates. This 247 may hint to the hypothesis that UCBSV is a more versatile virus. Isolates that are 248 genetically very similar are able to infect plants in relatively different 249 environments. 250

It must be noted here that the patterns arising from the various MCA
analyses are somewhat speculative. MCA has been shown to be a powerful tool in
population genetics, able to map the genetic diversity amongst a dataset to a
geographical path of evolution (Rohrlach et al., 2018). However, the sparsity of our
dataset ensures that strong conclusions are not possible in this case. We have 29
isolates to represent three distinct strains of a fast evolving virus, spread over a
wide and diverse geographical area and collected at various intervals over a 16 year
time period. Even with such a sparse dataset, we were able to notice some

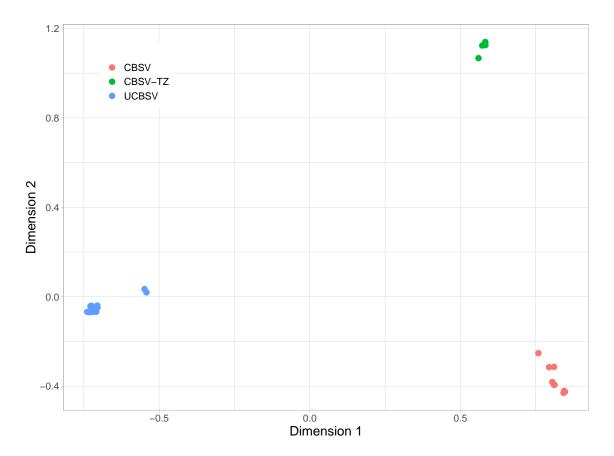


Figure 6: Results of Multiple Correspondence Analysis (MCA) on the full alignment. The three distinct clusters correspond to the three strains of CBSV.

associations between variables that can be used to generate testable hypotheses.

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# DISCUSSION

Our results confirm and extend the work of Mbewe et al. (2017), who found evidence for a third distinct clade of CBSV by analysing a subsection of the P1

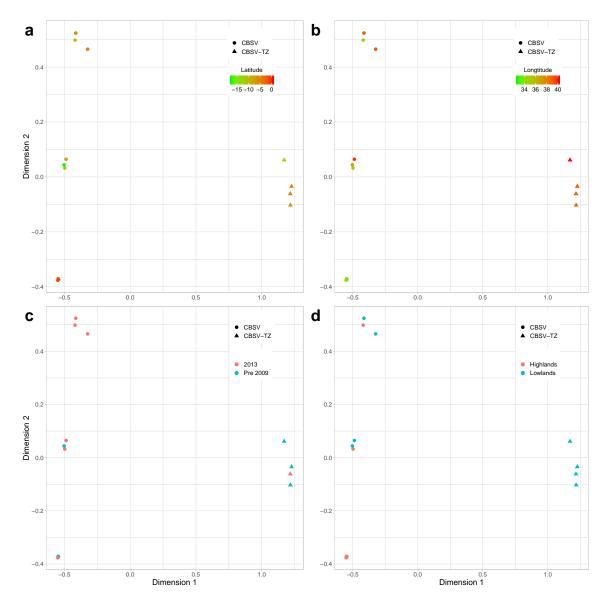


Figure 7: Results of MCA on the fourteen CBSV sequences. The shape of the points indicates the strain of the isolate, either CBSV or CBSV-TZ. The isolates are coloured by: a) latitude of their collection sites; b) longtitude of their collection sites; c) time of their collection (either before 2009 or 2013); and d) altitude of their collection sites (lowlands or highlands).

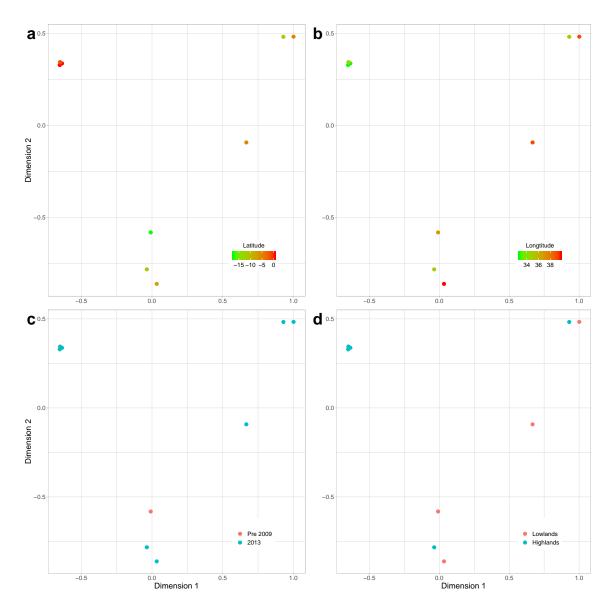


Figure 8: Results of MCA on the ten sequences from the CBSV strain (excluding the CBSV-TZ isolates). The isolates are coloured by: a) latitude of their collection sites; b) longitude of their collection sites; c) time of their collection (either before 2009 or 2013); and d) altitude of their collection sites (lowlands or highlands).

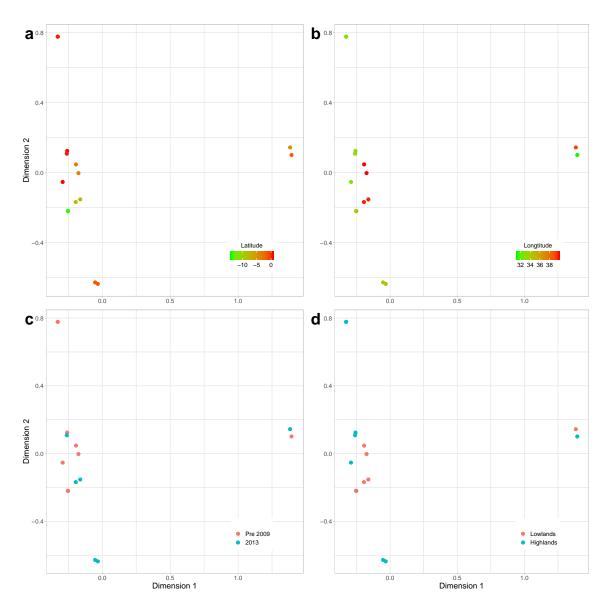


Figure 9: Results of MCA on the fifteen sequences from the UCBSV strain. The isolates are coloured by: a) latitude of their collection sites; b) longtitude of their collection sites; c) time of their collection (either before 2009 or 2013); and d) altitude of their collection sites (lowlands or highlands).

gene. Using two independent methods of sequence analysis we show that the
evidence for this third clade is not restricted to the P1 gene, but is in fact present
across the entire genome. That said, of the ten genes in the genome, the P1 gene is
the strongest relative contributor to the signal and focusing on this gene seems
apt.

There are practical implications of our findings that can have an immediate impact for cassava farmers in East Africa. The most vital step in securing the crops of farmers is the swift diagnosis of infected plants. It is vital not only to know that a plant is infected, but also to know which strain of the CBSV it has. This information is important on several fronts, it can be used to:

- select the most appropriate viral resistant cultivars for the next growing season.
- generate CBSV distribution maps that can be used to guide Agricultural

  Officers on where to screen (hotspot areas) or where to multiply (low pressure

  areas).
- inform where to deploy cassava material.
- strengthen phytosanitary regulations on movement of cassava germplasm.
- This work is undertaken by scientists working in the field directly with farmers.

Current practice when sequencing viral isolates is to focus on the CP gene, which
can effectively distinguish between CBSV and UCBSV (Monger et al., 2001;
Mbanzibwa et al., 2011). However, as indicated in Table 1, the CP gene is the least
influential of the ten genes in the CBSV-TZ Class. In order to effectively and
efficiently distinguish between CBSV, UCBSV and CBSV-TZ we recommend
biologists focus their attention on the P1 gene, in particular the first 60 amino
acids, as shown in Figures 3-5.

The results of the bioinformatic analyses presented here, when viewed in the 289 context of the history of CBSD in East Africa, allow a speculative hypothesis of the evolution of the virus to be constructed. We know that historically, the prevalence 291 of CBSD was negatively correlated with the altitude in which the crops were grown. The disease was prevalent in low-lying, coastal areas, and was never seen at 293 altitudes greater than 1000m. We know from the results of the MCA that the 294 CBSV-TZ strain (at least in the current dataset) has only been found at low 295 altitudes, and was much more prevalent among the pre 2009 samples in our dataset 296 than it was among the 2013 samples. Further, we know from previous research 297 (using the same dataset as we analyse here) by Alicai et al. (2016) that CBSV is 298 faster evolving than UCBSV. Finally, we showed that a relatively non-conserved 290 section of the P1 gene in CBSV-TZ is strongly conserved within both CBSV and

UCBSV, but strongly divergent between all three clades.

Let us denote the most recent common ancestor of the three strains as 302 CBSV-Anc. We hypothesise that given it's tendency to be found only at low 303 altitudes and its apparent decreased incidence in recent years, CBSV-TZ is the 304 closest of the three strains to CBSV-Anc. CBSV-Anc had low fitness at high 305 altitudes, and so mutations that increased the high-altitude fitness of the virus 306 would have been positively selected for on the fringes of its territory. Once these 307 alleles that increase high-altitude fitness gain a foothold in the population, the 308 geographical disparity in selective pressure may have precipitated a selective sweep of these alleles at high altitudes, but not at low altitudes. This effectively becomes the speciation event that spawns UCBSV.

As time passes UCBSV spreads through high altitude cassava growing
regions where it has no competition, but also populates low altitude areas, along
with CBSV-Anc. After some time the process then repeats itself. CBSV-Anc again
speciates into CBSV and CBSV-TZ. Like UCBSV before it, CBSV evolved the
ability to radiate into high-altitude regions. It also remains prevalent in the
low-altitude regions, resulting in the reduced prevalence of CBSV-TZ, as it now
competes with UCBSV and CBSV.

It can not be stated strongly enough that the above scenario is highly

speculative, but it is consistent with the known history of these viruses and the
evidence we can extract from the 29 whole-genome sequences. What we have
highlighted however, is that there are sophisticated bioinformatics tools now
available that can greatly assist in the fight against these viruses. What is needed
to turn our speculations into strong and robust evidence is more data. To combat
these fast-evolving viruses, we must increase research into them on two fronts:

- 1. We must amass much more systematic sequence data, covering the full geographic range of these viruses, with repeated sampling in each growing season. This will allow us to see in near real time how and where the virus is evolving, both geographically and on the molecular level. When sequencing virus isolates, detailed categorical information should be recorded, including the date; latitude, longtitude, and altitude of the collection site; cultivar the virus was found in; and detailed symptom description.
- 2. The structure of the viruses must be analysed on the molecular level. Here we highlight a region of the P1 gene that appears to be highly influential in increasing the high-altitude fitness of the virus. Understanding how and why this works on a molecular level is critical information that could assist in breeding highly resistant cultivars.

# Conclusion

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The fight to increase food security for the world's poorest citizens is of paramount 339 importance. Here we show that, given a sufficient amount of data, novel 340 bioinformatics tools have a vital role to play. Excellent work is already being done 341 to help farmers identify and contain virus outbreaks, but these are reactive 342 strategies. In order to eradicate these viruses we must understand how they work 343 and evolve. For this we need much more investment in data collection and analysis. 344 Ultimately, the cost of this investment is dwarfed by the potential gain. The benefits of achieving food security is not limited to nutrition. The resultant 346 economic boost would allow families and countries alike to invest heavily in health 347 and education, providing a dramatic increase to the standard of living across East Africa.

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# SUPPLEMENTARY MATERIAL

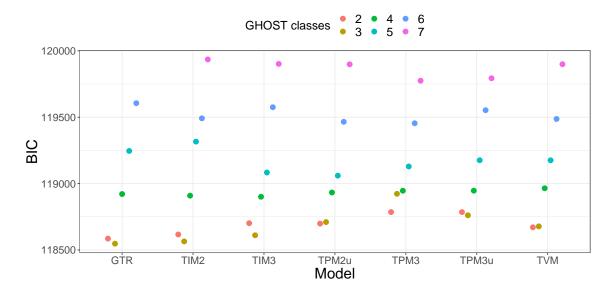


Figure S1: Results of model test procedure to select the optimal substitution model and number of classes, using BIC as the discriminating criterion. A total of 13 different nucleotide substitution models were tested, and each model was tested with between 2 and 12 classes. Poorly performing models (BIC > 120000) have been filtered out to improve resolution amongst the remaining models.

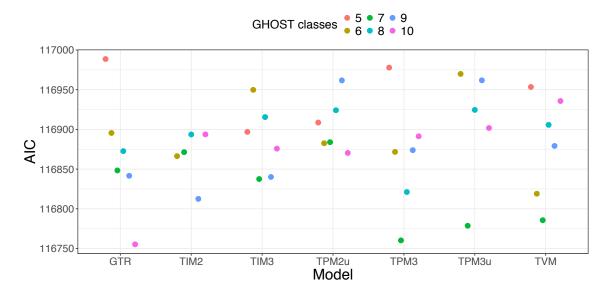


Figure S2: Results of model test procedure to select the optimal substitution model and number of classes, using AIC as the discriminating criterion. A total of 13 different nucleotide substitution models were tested, and each model was tested with between 2 and 12 classes. Poorly performing models (AIC > 117000) have been filtered out to improve resolution amongst the remaining models.

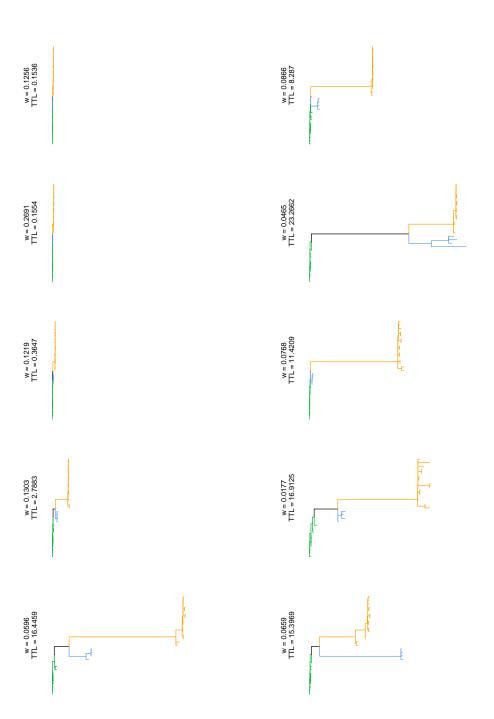


Figure S3: Trees inferred by IQ-TREE using the GHOST model with ten GTR classes. Tip labels are not shown for clarity. As in Figure 1, green edges form the CBSV clade, blue edges form the CBSV-TZ clade and orange edges form the UCBSV clade.