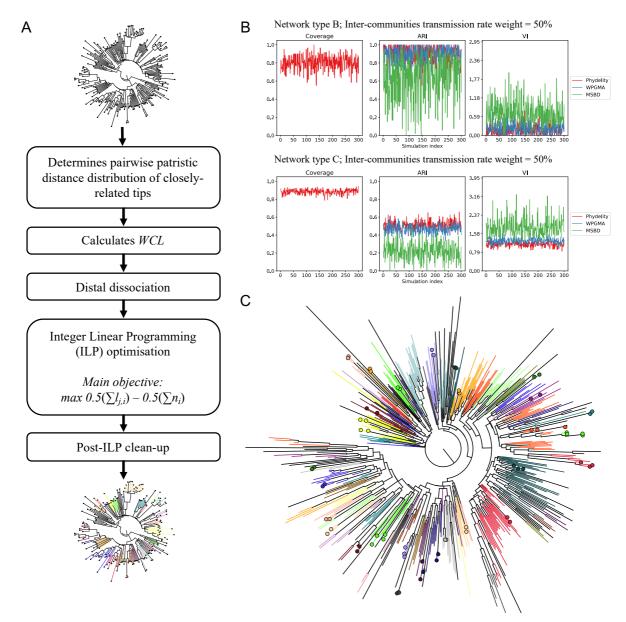
Inferring putative transmission clusters with Phydelity 1 2 Alvin X. Han^{1,2,3*}, Edyth Parker^{3,4}, Sebastian Maurer-Stroh^{1,5} and Colin A. Russell^{3*} 3 4 5 ¹Bioinformatics Institute, Agency for Science, Technology and Research (A*STAR), 30 Biopolis Street, Singapore 138671 6 ²NUS Graduate School for Integrative Sciences and Engineering, National University of 7 8 Singapore (NUS), 21 Lower Kent Ridge, Singapore 119077 9 ³Laboratory of Applied Evolutionary Biology, Department of Medical Microbiology, Academic Medical Centre, Meibergdreef 9, 1105 AZ Amsterdam-Zuidoost, The Netherlands, 10 11 ⁴Department of Veterinary Medicine, University of Cambridge, Madingley Rd, Cambridge 12 CB3 0ES, United Kingdom 13 ⁵Department of Biological Sciences, National University of Singapore, 16 Science Drive 4, 14 Singapore 117558 15 16 *Corresponding authors: hanxc@bii.a-star.edu.sg or c.a.russell@amc.uva.nl 17 Abstract: Current phylogenetic clustering approaches for identifying pathogen transmission 18 19 clusters are centrally limited by their dependency on arbitrarily-defined genetic distance 20 thresholds for within-cluster divergence. Incomplete knowledge of a pathogen's underlying 21 dynamics often reduces the choice of distance threshold to an exploratory, ad-hoc exercise 22 that is difficult to standardise across studies. Phydelity is a new tool for the identification of transmission clusters in pathogen phylogenies. It identifies groups of sequences that are more 23 24 closely-related than the ensemble distribution of the phylogeny under a statisticallyprincipled and phylogeny-informed framework, without the introduction of arbitrary distance 25 26 thresholds. In simulated phylogenies, Phydelity achieves higher rates of correspondence to 27 ground-truth clusters than current model-based methods, and comparable results to 28 parametric methods without the need for parameter calibration. 29 30 Availability and implementation: Phydelity is available at http://github.com/alvinxhan/Phydelity. 31 32 33

34 Introduction

- 35 Recent advancements in high-throughput sequencing technologies have led to the widespread
- 36 use of sequence data in infectious disease epidemiology (Gardy and Loman, 2017). In
- 37 particular, phylogenetics is frequently used to infer genetic clusters underlying the structure
- 38 of transmission networks (Ambrosioni et al., 2012; Bezemer et al., 2015; de Oliveira et al.,
- 39 2017). Current phylogenetic approaches for inferring transmission clusters (primarily
- 40 'cutpoint-based' methods) are centrally limited by the need to define arbitrary, absolute
- 41 cluster divergence thresholds (Ragonnet-Cronin et al., 2013; Prosperi et al., 2011). The lack
- 42 of a consensus definition of a phylogenetic transmission cluster (Grabowski and Redd, 2014)
- 43 coupled with incomplete knowledge of a pathogen's underlying epidemiological dynamics
- 44 often reduces the choice of cutpoints to an *ad hoc* exploratory exercise resulting in subjective
- 45 cluster definitions.
- 46
- 47 Phydelity is a new tool for inferring putative transmission clusters through the identification
- 48 of groups of sequences that are more closely-related than the ensemble distribution under a
- 49 statistically-principled framework. Notably, Phydelity only requires a phylogeny as input,
- 50 negating the need to define arbitrary cluster divergence thresholds, and also only has a single
- 51 parameter that can either be user defined or determined directly by Phydelity. Phydelity is
- 52 freely available at http://github.com/alvinxhan/Phydelity.
- 53



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Figure 1. (A) Phydelity algorithm pipeline. (B). Clustering correspondence metrics for clustering algorithms
(Phydelity, WPGMA and MSBD) applied to phylogenies generated from simulations of HIV epidemics of
hypothetical MSM sexual contact network types B and C, with inter-communities transmission rates weighted at
50% of within-community rate (Villandre *et al.*, 2016). (C) Clustering results of Phydelity on HIV-1 subtype A *env* sequences collected from the Rakai Community Cohort Study (Grabowski *et al.*, 2014). Tips are coloured
by Phydelity clusters and those marked with • were phylogenetic clusters identified by Grabowski *et al.*

61

62 Method

63 Phydelity considers the input phylogeny as an ensemble of putative clusters, each consisting

- of an internal node *i* and the leaves it subtends. The within-cluster diversity of node *i* is
- 65 measured by its mean pairwise patristic distance (μ_i). Phydelity then determines the pairwise
- 66 patristic distance distribution of closely-related tips, which comprises the pairwise distances
- 67 of sequence *j* and its closest *k*-neighbouring tips, where the closest *k*-neighbours includes
- 68 sequence j. The user can input the desired k parameter or Phydelity can automatically scale k

69 to the value that yields the supremum distribution with the lowest overall divergence. All 70 tests of Phydelity presented in this work were performed using the autoscaled value of k. 71 Regardless of how the distance distribution of closely-related tips is determined, Phydelity 72 73 uses this distribution to calculate the within-cluster divergence limit (WCL), an upper bound 74 to μ_i of putative clusters: $WCL = \bar{\mu} + \sigma$ 75 76 where $\bar{\mu}$ is the median pairwise distance of the closely-related tips distance distribution and σ 77 is the corresponding robust estimator of scale without assuming symmetry about $\bar{\mu}$. 78 79 This is followed by distal dissociation of distantly-related descendant subtrees/sequences to 80 any ancestral node with $\mu_i > WCL$, thereby facilitating identification of both monophyletic as well as nested, paraphyletic clusters (Han et al., 2018). Phydelity filters outlying tips from 81 putative clusters under the assumption that viruses infecting individuals in a quick 82

transmission chain are ultimately descended from the same source and are highly similar

84 genetically. An outliers is defined by a node-to-tip distance more than three deviations from

85 the median distance. An integer linear programming model is implemented and optimised

86 under a blended objective of equal weights to maximise the number of sequences clustered

87 within the lowest number of clusters. Lastly, clean-up steps are taken to remove any

topologically outlying singletons that were spuriously clustered as described in Han *et al.*

89 (2018). The full algorithm description and mathematical formulation of Phydelity is detailed

90 in Supplementary Materials.

91

92 For computational performance, Phydelity can process a phylogeny of 1000 tips, on an

93 Ubuntu 16.04 LTS operating system with an Intel Core i7-4790 3.60 GHz CPU, in ~3

94 minutes using a single CPU core and 253 MB of peak memory usage.

95

96 **Results**

97 Phydelity was evaluated on phylogenetic trees derived from simulated HIV epidemics of two

98 hypothetical MSM sexual contact network types (network types B & C) produced by

99 Villandre et al. (2016), wherein quick transmission chains (i.e. transmission clusters) could

100 be attributed to sexual contact among individuals belonging to the same community. Network

101 type B, which corresponded best with the assumption of monophyletic clusters, consisted of a

102 main contact network of 60 individuals with single linkages to 25 disjoint subnetworks of 20 subjects. Conversely, the more realistic network type C included 100 communities of sizes 103 104 sampled from an empirical distribution obtained from the Swiss HIV Cohort Study. For both network types, inter-community transmission rates were weighted at 25%, 50%, 75% or 105 100% of the within-community rate. These simulated datatsets were also tested by Barido-106 107 Sottani et al using their multi-state birth-death (MSBD) method which infers transmission 108 clusters by detecting significant changes in transmission rates (Barido-Sottani et al., 2018). 109 More information on the simulated epidemics are included in Supplementary Materials. 110

111 Clustering results from Phydelity were compared to those generated by the MSBD method

and a cutpoint method based on the weighted pair-group method of analysis (WPGMA)

113 (Villandre *et al.*, 2016) (Figure 1B, Supplementary Fig. 1 and Supplementary Table 1). Both

the adjusted rand index (ARI) and variation of information (VI) were calculated to quantify

the correspondence between the actual network communities and clustering results. Villandre

116 *et al.* (2016) assessed four different commonly used cutpoint-based methods, including

arbitrarily varying patristic distance thresholds between any two tips (Brenner *et al.*, 2007),

118 ClusterPicker (varying standardised number of nucleotide changes; Ragonnet-Cronin *et al.*,

119 2013), PhyloPart (changing arbitrary percentile of pairwise patristic distance distribution;

120 Prosperi *et al.*, 2011) and agglomerative hierarchical clustering methods such as the

121 WPGMA method. WPGMA methods achieved the best overall ARI. As such, only WPGMA

122 clustering results derived from the optimal cutpoint parameter (i.e. maximum ARI) were

123 compared.

124

125 Owing to Phydelity's definition of a closely-related neighbourhood, its distal dissociation

approach and outlier detection, its mean coverage of sequences clustered ranges from 70.8–

127 80.0% for B networks and 87.5-87.9% for C networks. Phydelity (mean ARI = 0.90-0.91,

128 mean VI = 0.17-0.18) consistently performed as well as optimised WPGMA (mean ARI =

129 0.88-0.96, mean VI = 0.09-0.25) for B networks. Phydelity was the best performing method

for C networks (Phydelity: mean ARI = 0.49-0.59, mean VI = 0.95-1.18; WPGMA: mean

131 ARI = 0.44-0.56, mean VI = 1.07-1.33; MSBD: mean ARI = 0.19-0.22, mean VI = 1.80-2.02;

132 Supplementary Table 1). Notably, even though results generated by WPGMA are comparable

to those from Phydelity, this was only possible for WPGMA when the optimal cutpoint could

be determined by calibration with the simulated ground-truth. However, ground truth

clustering is largely unavailable in epidemiological studies. Phydelity, on the other hand,does not require this calibration step.

- 137
- 138 Phydelity was also tested on an empirical dataset of HIV-1 subtype A *env* sequences obtained
- 139 from the Rakai Community Cohort Study (Grabowski *et al.*, 2014). Grabowski *et al.*
- identified 35 clusters by phylogenetic analysis, of which 18 constituted individuals of the
- same community and 12 clusters were confirmed to be from the same household. As
- 142 community and household information was blinded for privacy reasons, the available
- sequence data could not be matched for the exact clusters identified by Grabowski *et al.*
- 144 However, the stringent cluster definitions used by Grabowski et al. (≥90% bootstrap support
- 145 (1000 iterations), median genetic pairwise distance $\leq 2.6\%$) restricts most of these clusters to
- pairs, with non-pair clusters made up of no more than 5 individuals. As such, we found the
- same number of clusters of similar size distribution through visual inspection when we
- recapitulated the phylogeny using the same methods (GTR+I+G substitution model, Garli)
- and cluster definition as Grabowski et al. Phydelity identified 33 out of the 35
- 150 epidemiologically-identified clusters as distinct transmission clusters (Figure 1C).
- 151

152 Conclusion

- 153 Phydelity is a statistically-principled and phylogeny-informed tool capable of identifying
- 154 putative transmission clusters in pathogen phylogenies without the introduction of arbitrary
- 155 distance thresholds. It is fast, generalizable, and freely available at
- 156 https://github.com/alvinxhan/Phydelity.
- 157

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