

1 **Cortico-genetic mapping links individual brain maturity in youths to cognitive and** 2 **psychiatric traits**

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13
14 **Neurodevelopmental trajectories are shaped by interactions between coordinated biological**
15 **processes and individual experiences throughout ontogeny, yet the specific genetic and**
16 **environmental impact on brain development is enigmatic. Here, we map the genetic**
17 **architectures of cognitive traits and psychiatric disorders onto the brain, show that such**
18 **canonical genetic maps are associated with individual normative patterns in youths, and**
19 **provide evidence that trauma exposure and parental education may alter this relationship.**

20 Psychiatric, cognitive and brain imaging traits are highly heritable and polygenic¹⁻¹³. Individual
21 genetic architecture contributes to individual differences in neurodevelopmental trajectories and
22 subsequent scaffolding and maintenance of brain structure and function throughout ontogeny.
23 However, the links between the genetic and neural configurations and how their interplay shapes
24 individual differences in cognitive function and mental health remain poorly understood. This
25 knowledge gap has nurtured a debate on the extent of environmental influence and genetic
26 constraints on brain development. Here, we provide a comprehensive neuroanatomical mapping
27 of the genetic architecture of various cognitive traits and psychiatric disorders using brain
28 imaging and genetic data in a large population based sample (*UK Biobank*¹⁴), and link the
29 resulting canonical genetic maps to individual patterns of brain maturity in the *Philadelphia*
30 *Neurodevelopmental Cohort*¹⁵.

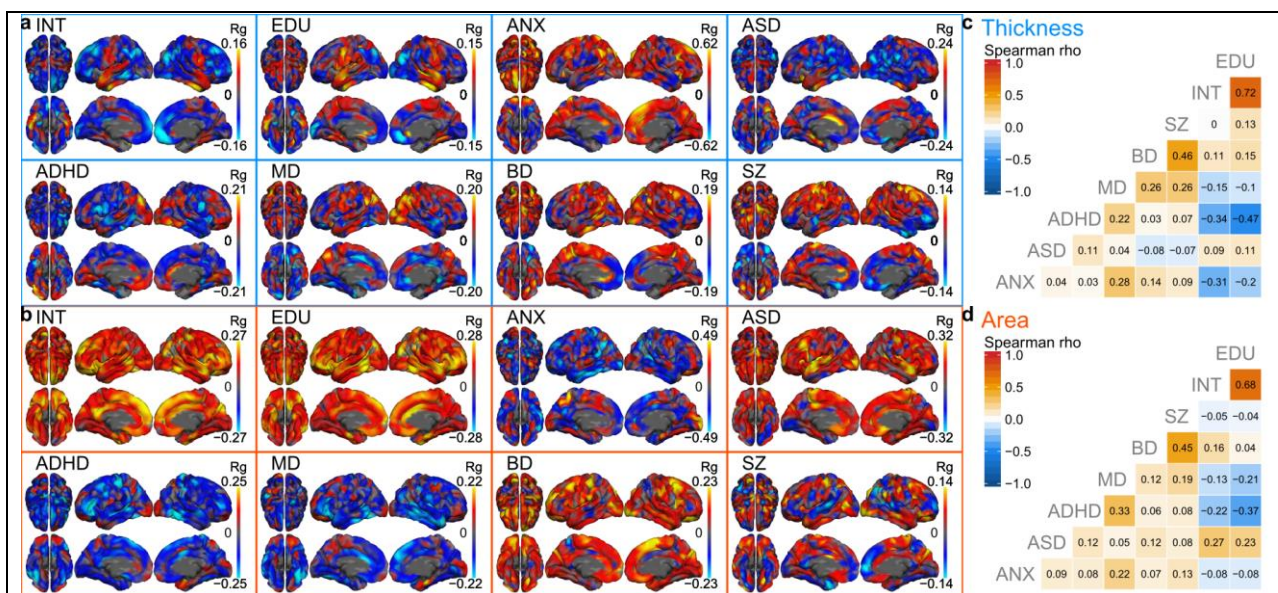
31 We accessed data from the *UK Biobank*¹⁴ and used *Freesurfer*¹⁶ for cortical reconstruction based
32 on T1-weighted magnetic resonance images obtained from 16,612 healthy individuals with
33 European ancestry aged 40 to 70 years (mean: 55.8 years, sd: 7.5 years, 52.1% females). We
34 computed surface maps for cortical thickness and area, registered to *fsaverage4* space (2,562
35 vertices), smoothed using a kernel with full width at half maximum of 15 mm. Next, we
36 performed a genome-wide association study (GWAS) for every vertex using *PLINK*¹⁷, linking
37 single-nucleotide-polymorphism (SNP) data with a given vertex's thickness and area,
38 respectively. Each GWAS accounted for effects of age, age², sex, scanning site and the first four
39 genetic principal components¹⁸ to account for population stratification.

40 We first estimated SNP-heritability of cortical morphology using *LD Score regression*¹⁹ for each
41 vertex (**Suppl. Fig 1**). The spatial correlation between thickness and area heritability maps was
42 moderate ($r=0.28$, $p_{perm}=0.0004$; **Suppl. Fig 1b**), and surface area was significantly more
43 heritable than thickness (**Suppl. Fig 1a**). Both measures showed regional differences, with high
44 heritability of thickness in the postcentral gyrus and Heschl's gyrus (**Suppl. Fig 1c**), and of
45 surface area in the lingual gyrus and the temporal lobe (**Suppl. Fig 1d**). These results from
46 vertex-wise SNP-based analysis largely confirmed earlier reports from twin studies⁹⁻¹³ and from a
47 region-wise SNP-based analysis²⁰, supporting the feasibility of our vertex-wise GWAS approach.

48 We next combined our GWAS results with publicly available summary statistics to compute
49 vertex-wise genetic correlations between brain morphology and cognitive traits and psychiatric
50 disorders. Summary statistics for cognitive phenotypes were obtained from GWAS on
51 intelligence¹ (INT) and educational attainment² (EDU), excluding *23andMe* data. For psychiatric
52 disorders we used summary statistics from analyses on anxiety³ (ANX), autism spectrum
53 disorder⁴ (ASD), attention-deficit-hyperactivity disorder⁵ (ADHD), major depression⁶ (MD,
54 excluding *23andMe* data), bipolar disorder⁷ (BP) and schizophrenia⁸ (SZ). Using *LD Score*
55 *regression*¹⁹, for each vertex we estimated the genetic correlation between each of the phenotypes
56 and thickness and area, respectively. To reduce noise, vertices with a heritability estimate of less
57 than 1.96 times its standard error were excluded from the analysis, in addition to excluding the
58 medial wall, yielding a total of 4550 and 4498 vertices for thickness and area, respectively.

59 **Fig. 1a-b** depict the resulting cortical maps of vertex-wise genetic correlations – hereafter
60 referred to as *cortico-genetic maps*. Each map reflects the overlap between the genetic

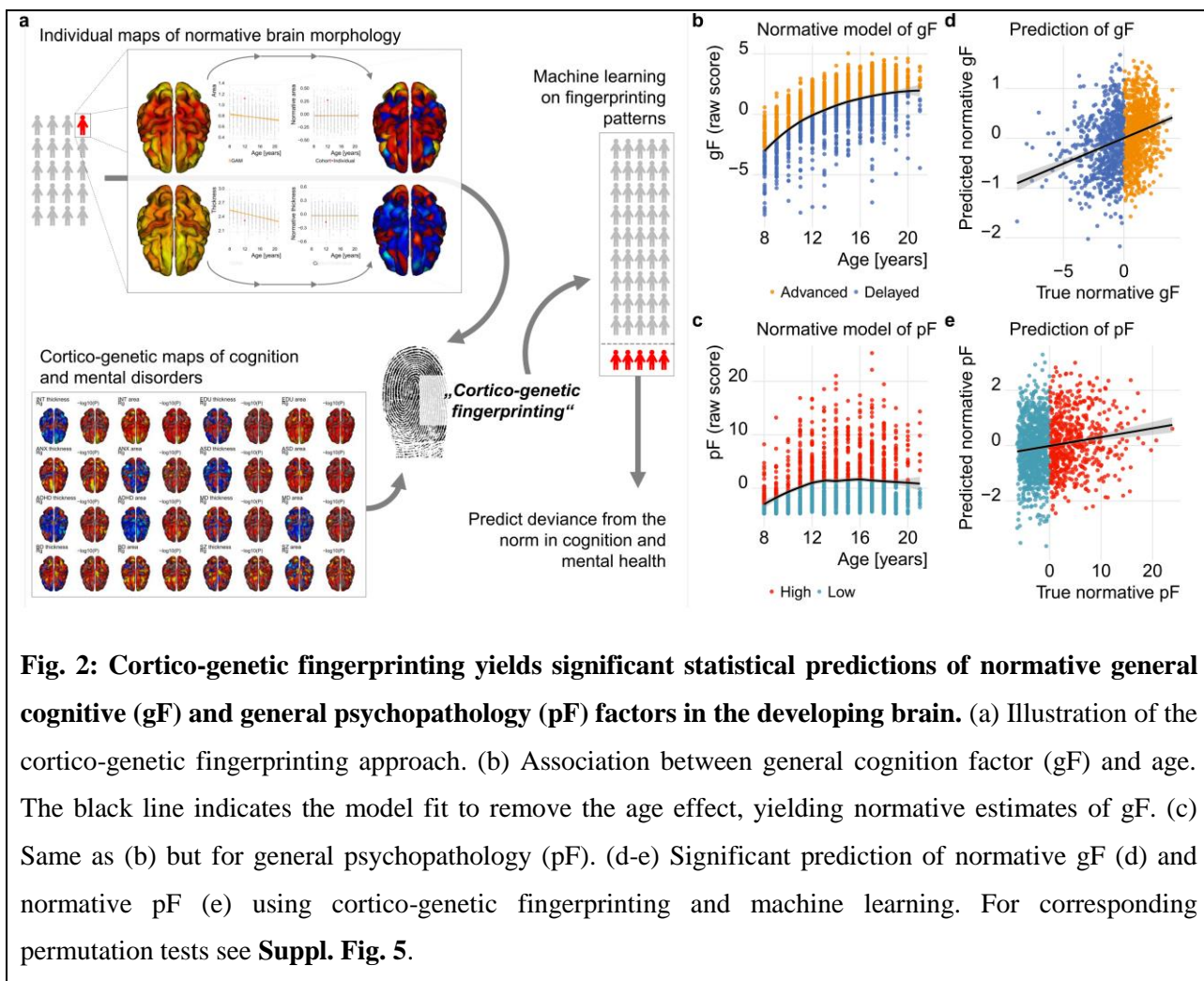
61 architectures of cortical morphology and the given phenotype. For example, area in the right
 62 superior frontal gyrus was positively genetically associated with INT and negatively with MD,
 63 whereas thickness in this area was positively associated with ANX. **Fig. 1c-d** illustrate the
 64 correlation between these cortico-genetic maps, largely in line with the published genetic
 65 relationship between the phenotypes (**Suppl. Fig. 2** for comparison). Importantly, the cortico-
 66 genetic maps were derived from brain imaging data of healthy individuals, thereby reducing the
 67 impact of confounding factors such as comorbid disorders or medication, as might be observed
 68 for case-control brain imaging maps.



69
 70 **Fig. 1: Cortico-genetic maps reflecting the vertex-wise genetic correlations of cortical morphology**
 71 **with cognition and psychiatric disorders.** Genetic correlations (Rg) per phenotype for thickness (a) and
 72 for area (b). The maximum of the scales was individually adjusted to display the 97.5 percentile across all
 73 vertices. Corresponding p-values are depicted in **Suppl. Fig. 3**. (c-d) Pairwise Spearman correlations of
 74 the cortico-genetic maps from (a-b). Corresponding p-values from permutation testing are depicted in
 75 **Suppl. Fig. 4**.

76 Considering the strong evidence of a neurodevelopmental component in the etiology of many
 77 psychiatric disorders^{21,22} and the large amount of maturational brain changes related to individual
 78 adaptation and learning²³, we hypothesized that brain regions associated with the genetic
 79 architecture of psychiatric and cognitive traits in healthy adults are sensitive to normative
 80 deviations during childhood and adolescence. To this end, we tested if the similarity between an
 81 individual's map of brain maturity and the respective cortico-genetic maps allowed us to

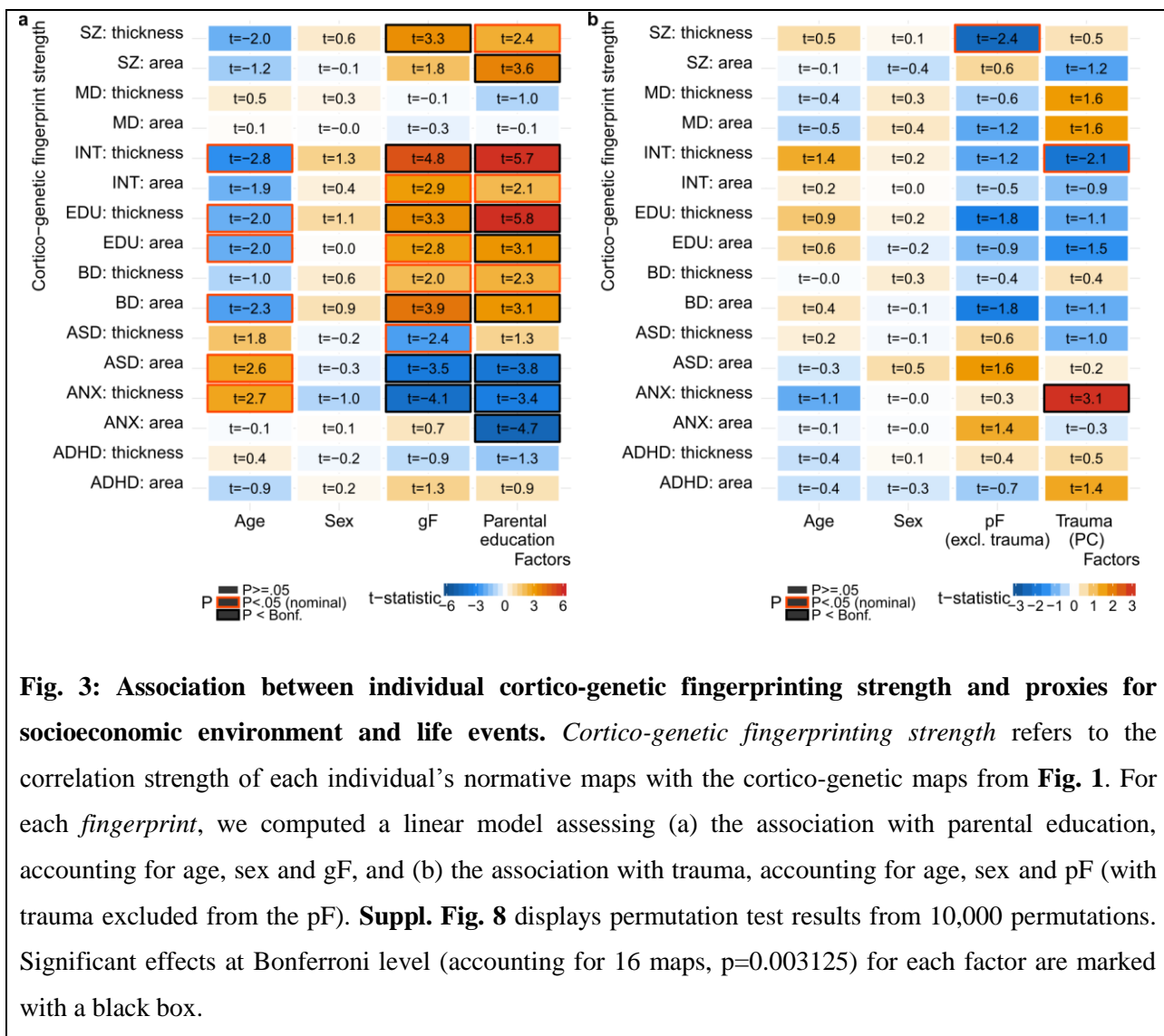
82 statistically predict individual deviations from the developmental norm in the given trait in the
83 *Philadelphia Neurodevelopmental Cohort*¹⁵. Following cortical reconstruction¹⁶, we excluded
84 data due to insufficient quality after manual screening (n=60) and significant or major medical
85 conditions (n=73), yielding a total sample of 1467 individuals aged 8 to 21 years (mean: 14.14
86 years, sd: 3.51 years, 52.9% females).



95 **Fig. 2a** details the approach. First, we modeled normative trajectories of brain development in
96 each vertex using generalized additive models²⁴ and removed the statistical relationship with age
97 and sex from all data sets. For each individual, this yielded one normative map for cortical
98 thickness and one for cortical area, where the measures at each vertex reflect its respective
99 deviation from the age- and sex-matched sample norm. Next, we assessed the similarity of each
100 individual's normative thickness and area maps to each of the cortico-genetic maps from **Fig. 1a-**
101 **b**. Following the *connectome fingerprinting* approach²⁵⁻²⁷, we hereafter refer to this as *cortico-*

102 *genetic fingerprinting* outlining that the cortico-genetic maps are used as ‘*fingerprints*’ of
103 cognitive traits and psychiatric disorders that individual patterns of normative development are
104 compared to. Since the direction of effects in studies of brain morphometry may depend on age²⁸,
105 we also fingerprinted using the unsigned statistical maps. For example, whereas SZ is associated
106 with widespread reduced cortical thickness²⁹, the cortico-genetic thickness maps for SZ in **Fig. 1a**
107 show positive genetic correlations in the precentral cortex. Speculatively, this may partly reflect a
108 survivor bias as these maps were generated using data from healthy individuals aged 40 and
109 above with very low life-time probability of a diagnosis. Therefore, in some cases the unsigned
110 effect sizes might be needed to overcome the bias and to obtain predictive value in a
111 neurodevelopmental sample. We thus fingerprinted using both signed genetic correlation maps
112 (**Fig. 1a-b**) and unsigned maps of $-\log_{10}$ transformed p-values (**Suppl. Fig. 3**), yielding a total of
113 eight (phenotypes) by three (area and thickness, and both concatenated) by two (genetic
114 correlation, $-\log_{10}$ transformed p-values) fingerprinting correlations per individual (48 in total).

115 To assess the predictive utility of these fingerprints, we used machine-learning to predict
116 deviations from the developmental norm in cognitive performance and mental health. Using
117 principal component analysis on clinical and cognitive data, we derived a general cognitive score
118 (gF) and a general psychopathology score (pF)³⁰ and first removed effects of age using locally
119 weighted regression (**Fig 2b-c**), yielding normative estimates of gF and pF. In a 10-fold cross-
120 validation framework, we trained a linear machine learning model on 90% of the data to predict
121 normative gF and normative pF using the 48 correlation estimates – the *connectome*
122 *fingerprinting strength* – as features, and iteratively testing on the 10% held-out data. **Fig. 2d-e**
123 illustrates that it was possible to statistically predict normative gF and normative pF using
124 cortico-genetic fingerprinting (both $p_{\text{perm}} < .0001$, **Suppl. Fig. 5**). In models accounting for age and
125 sex, the associations between true and predicted normative values were highly significant both
126 for gF ($r_{\text{partial}}=0.29$; $t=11.71$, $p=2e-16$) and pF ($r_{\text{partial}}=0.14$; $t=5.51$, $p=4e-8$). The machine learning
127 model weights revealed that a range of traits contributed to each prediction (**Suppl. Fig. 6**). These
128 results jointly suggest that individual regional deviations from the norm in youths emerge in
129 those cortical areas that are most strongly associated with the genetic architecture of the
130 respective phenotypes, and that the extent of overlap with those patterns relates to individual
131 differences in cognitive performance and – to a lesser degree - mental health.



132

133 **Fig. 3: Association between individual cortico-genetic fingerprinting strength and proxies for**
 134 **socioeconomic environment and life events.** *Cortico-genetic fingerprinting strength* refers to the
 135 correlation strength of each individual's normative maps with the cortico-genetic maps from **Fig. 1**. For
 136 each *fingerprint*, we computed a linear model assessing (a) the association with parental education,
 137 accounting for age, sex and gF, and (b) the association with trauma, accounting for age, sex and pF (with
 138 trauma excluded from the pF). **Suppl. Fig. 8** displays permutation test results from 10,000 permutations.
 139 Significant effects at Bonferroni level (accounting for 16 maps, $p=0.003125$) for each factor are marked
 140 with a black box.

141 The observed cortico-genetic overlap with individual estimates of brain maturity raises the
 142 question whether and to which degree this relationship is altered by experience. We used parental
 143 education as a proxy for the socioeconomic environment and the first component from a principle
 144 component analysis on trauma questionnaires as a proxy for major negative life events (**Suppl.**
 145 **Fig 7**). Next, we tested for linear associations between these factors and individual cortico-
 146 genetic fingerprinting strength. As depicted in **Fig. 3a**, parental education was positively
 147 associated with fingerprinting strengths on SZ (area), INT (thickness), EDU (thickness and area),
 148 and BD (area), as well as negatively associated with those on ASD (area) and ANX (thickness
 149 and area), each model accounting for age, sex and gF. Trauma exposure was positively associated
 150 with fingerprinting strengths on ANX (thickness), accounting for age, sex and a pF that excluded
 151 trauma items (**Fig. 3b**). In other words, the overlap between an individual's normative cortical

152 morphology and the cortico-genetics maps from **Fig. 1** varied as a function of experience. With
153 caution and under the restriction that these are probabilistic, not deterministic associations, these
154 results indicate that individual developmental patterns are more ANX-like following trauma
155 exposure, more ANX- and ASD-like in individuals from low-educated social environments and
156 more SZ-, BD-, INT- and EDU-like in individuals from high-educated social environments.

157 Taken together, our analysis reveals an intriguing impact of genetic architecture on brain
158 development by illustrating that the similarity between individual patterns of brain maturity and
159 the neurogenetics of cognition and psychopathology is informative for individual normative
160 deviations in cognitive performance and mental health. Nevertheless, despite statistical
161 significance our predictions only explained a proportion of the variance in the data, indicating
162 that other factors have a large part in explaining individual trajectories. Indeed, we identified two
163 environmental factors – proxies of the socioeconomic environment and adverse life events - as
164 significant factors explaining variance in the individual fingerprints. Of note, these factors have
165 substantial genetic components themselves, and future research needs to address to what extent
166 the observed associations with environmental factors can be explained by common genetics.
167 Apart from its utility in pinpointing deviations from the norm in the developing human brain, our
168 cortico-genetic approach may contribute towards the delineation of genetic and environmental
169 factors influencing individual trajectories during sensitive neurodevelopmental phases.

170

171 **Author contributions**

172 T.K. and L.T.W conceived the study; T.K. and D.v.d.M. pre-processed the data. T.K. performed
173 all analyses, with contributions from D.v.d.M and O.F. and with conceptual input from D.A. and
174 L.T.W.; All authors contributed to interpretation of results; T.K. drafted the manuscript and all
175 authors contributed to and approved the final manuscript.

176

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183 sets was provided by grant RC2MH089983 awarded to Raquel Gur and RC2MH089924 awarded
184 to Hakon Hakonarson, and all subjects were recruited through the Center for Applied Genomics
185 at The Children's Hospital in Philadelphia.

186

187 **Competing interests**

188 The authors declare no competing financial interests

189

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222

223 **Online methods**

224 *Samples and exclusion criteria*

225 UK Biobank: The UK Biobank is a publicly available resource of imaging, genetics and
226 phenotypic data from an ongoing large-scale cohort study¹⁴. All study procedures were approved
227 by appropriate ethics committees and all study participants gave electronic signed consent. We
228 obtained access with permission no. 27412. No statistical methods were used to pre-determine
229 sample sizes as we analyzed all available data from the 20,000-subject release of brain imaging
230 and corresponding phenotypic and genetics data. Individuals with Caucasian ancestry were
231 identified by the UK Biobank study team using clustering analysis¹⁸ and we followed their
232 selection of individuals in our study. After exclusion of data from individuals with a diagnosed
233 brain disorder or data of insufficient quality (see *pre-processing and quality control*), this yielded
234 a total of 16,612 healthy individuals with Caucasian ancestry. The age range was 40 to 70 years
235 (mean: 55.8 years, sd: 7.5 years, 52.1% females).

236 PNC: The Philadelphia Neurodevelopmental Cohort is a publicly available resource of clinical,
237 cognitive, genetic and neuroimaging data from children and adolescents^{15,31}. Prior to data
238 collection, all study procedures were approved by the institutional review boards of the
239 University of Pennsylvania and the Children's Hospital of Philadelphia, and all participants gave
240 written informed consent. We obtained access with permission no. 8642. No statistical methods
241 were used to predetermine sample sizes as we used all available data, except data with
242 insufficient quality after manual screening (n=60) and data from individuals with significant or
243 major medical conditions (n=73). The final sample comprised 1467 individuals aged 8 to 21
244 years (mean: 14.14 years, sd: 3.51 years, 52.9% females).

245

246 *Image pre-processing and quality control*

247 T1-weighted magnetic resonance images for UK Biobank (MPRAGE, TR 2000 ms, TE 2.01 ms,
248 matrix 208x256x256, resolution 1x1x1 mm) and PNC (MPRAGE, TR 1810 ms, TE 3.51 ms,
249 matrix 192x256x160, resolution 0.9x0.9x1mm) was processed using Freesurfer 5.3¹⁶ (recon-all).
250 In the case of UK Biobank, where manual quality control of 16,612 images was not feasible, we
251 excluded outliers based on global cortical measures. We regressed age, age², sex and scanning
252 site from white surface area and mean cortical thickness of each hemisphere and identified

253 outliers above or below 4 standard deviations of the full population, excluding N=22 individuals.
254 In the case of PNC, we screened all reconstructed images manually, excluding data from N=60
255 children and adolescents that did not adhere to highest data quality standards. Next, for both UK
256 Biobank and PNC, we resampled the surfaces to *fsaverage4* space (2,562 vertices), smoothed
257 using a kernel with full width of half maximum of 15 mm.

258

259 *Vertex-wise genetic analysis in UK Biobank data*

260 Standard quality control procedures were applied to the UK Biobank v3 imputed genetic data,
261 including removal of SNPs with an imputation quality score below 0.5, with a minor allele
262 frequency less than .05, missing in more than 5% of individuals, and failing the Hardy Weinberg
263 equilibrium tests at a $p < 1 \times 10^{-6}$. Genetic principal components were retrieved from the UK
264 Biobank repository and we regressed the first four genetic components, age, age², sex and
265 scanning site from vertex-wise thickness and area maps. Next, we ran one genome-wide
266 association analysis (GWAS) per vertex using *PLINK v1.9*¹⁷, and removed the MHC region from
267 each resulting summary statistic. Using *LD Score regression*¹⁹, we estimated narrow sense
268 heritability. The significance of the correlation between heritability maps of thickness and area
269 was assessed using spin-rotation based permutation testing, which applies random rotations to
270 spherical representations of the cortical surface to generate a null distribution³². Next, we used
271 cross-trait *LD Score regression*^{19,33} to calculate correlations of our vertex-wise GWAS summary
272 statistics with publicly available summary statistics on intelligence¹ (INT), educational
273 attainment² (EDU, excluding *23andMe* data), anxiety³ (ANX, the case-control GWAS), autism
274 spectrum disorder⁴ (ASD), attention-deficit-hyperactivity disorder⁵ (ADHD), major depression⁶
275 (MD, excluding *23andMe* data), bipolar disorder⁷ (BP) and schizophrenia⁸ (SZ). Significance of
276 the correlations between each pair of the resulting cortico-genetic maps was again assessed using
277 spin-rotation based permutation testing³² in addition to correcting the permuted p-values for the
278 number of total correlations (28, Bonferroni correction).

279

280 *Cortico-genetic 'fingerprinting' in PNC data*

281 We utilized generalized additive models²⁴ to remove the statistical relationship with age and sex
282 from vertex-wise thickness and area data, yielding one normative thickness and one normative

283 area map per individual in PNC. Each of these individual subject maps was transformed into a
284 one-column vector and correlated against similar vectors of the cortico-genetic maps of cognition
285 and psychiatric disorders using Spearman correlations. We refer to this approach – in line with
286 the *connectome fingerprinting* literature²⁵⁻²⁷ – as *cortico-genetic fingerprinting*. We fingerprinted
287 against each of the 16 cortico-genetic correlation maps (Rg) from **Fig. 1**, against each of the 16 -
288 log₁₀ transformed cortico-genetic p-value maps (**Suppl. Fig. 3**) and against each of 16 vectors
289 that concatenated thickness and area surface vectors, respectively (8 Rg maps concatenating area
290 and thickness, 8 -log₁₀(P) maps concatenating area and thickness). In sum, this yielded 48
291 Spearman correlation estimates (*'fingerprinting strength'*) per subject. To assess the predictive
292 utility of the fingerprinting strengths, we used those 48 correlations as features in machine
293 learning based prediction of normative estimates of general psychopathology (pF) and general
294 cognition (gF), respectively. PF and gF were obtained from a PCA following previous protocols³⁰
295 from the full PNC sample (9490 individuals) and the respective scores extracted for those
296 individuals with imaging data available. Dependencies with age were removed using locally
297 weighted regression to account for non-linear effects. Machine learning was performed in a 10-
298 fold cross-validation framework using *slm* from the *care* package³⁴ in R statistics and normative
299 estimates of pF and gF were predicted. Significance of the predictions was assessed using
300 permutation testing, repeating 10,000 runs of a full 10-fold cross-validation loop using a different
301 random permutation of the response variable in each run. Feature weights were assessed using
302 *CAR scores* and translated to -log₁₀ transformed p-values³⁴. Finally, to assess environmental
303 impact on cortico-genetic fingerprinting strength, we computed parental education as the mean of
304 maternal and paternal education, and a principal component analysis across various trauma
305 questions (**Suppl. Fig. 7**) yielding a general trauma score (the first factor). For each cortico-
306 genetic map we tested for linear associations between individual fingerprinting strength and
307 parental education accounting for age, sex and gF. Likewise, for each map we tested for linear
308 associations with trauma, accounting for age, sex and pF (the pF was recomputed for this analysis
309 to exclude trauma items). Significance of the linear associations was assessed using permutation
310 testing, permuting the fingerprinting strength 10,000 times and each time recomputing the models
311 on the permuted data. In addition, resulting P-values were corrected for multiple comparison
312 using Bonferroni correction (p=.003125, 16 tests).

313

314 **Data availability**

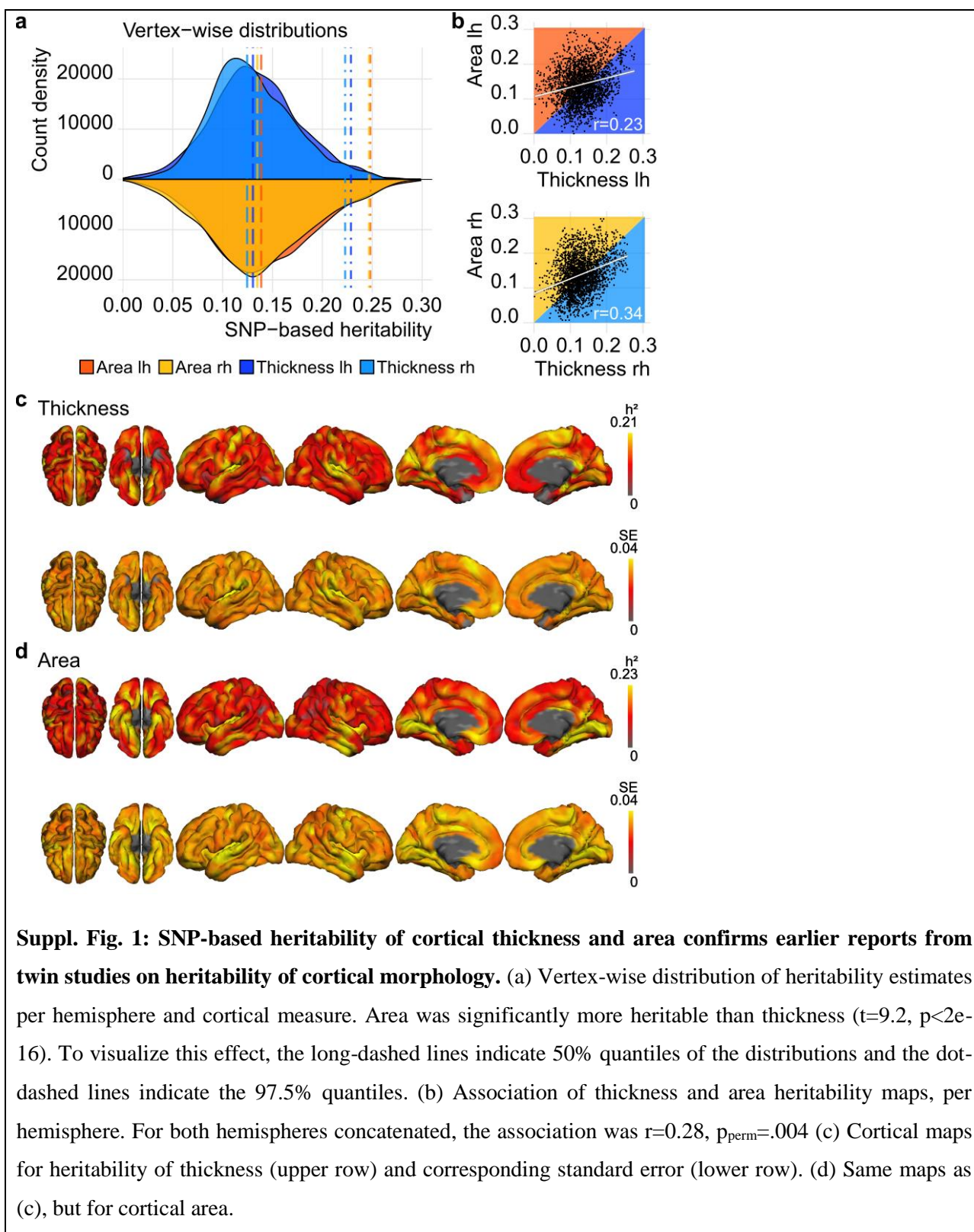
315 The data incorporated in this work are available as part of the publicly available UK Biobank
316 (<https://www.ukbiobank.ac.uk/>) and Philadelphia Neurodevelopmental Cohort (PNC,
317 https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000607.v2.p2).

318

319 **Code availability**

320 Scripts are available upon request from the first author (tobias.kaufmann@medisin.uio.no).

321 **Supplementary figures**



322

323 **Suppl. Fig. 1: SNP-based heritability of cortical thickness and area confirms earlier reports from**

324 **twin studies on heritability of cortical morphology.** (a) Vertex-wise distribution of heritability estimates

325 per hemisphere and cortical measure. Area was significantly more heritable than thickness ($t=9.2$, $p<2e-$

326 16). To visualize this effect, the long-dashed lines indicate 50% quantiles of the distributions and the dot-

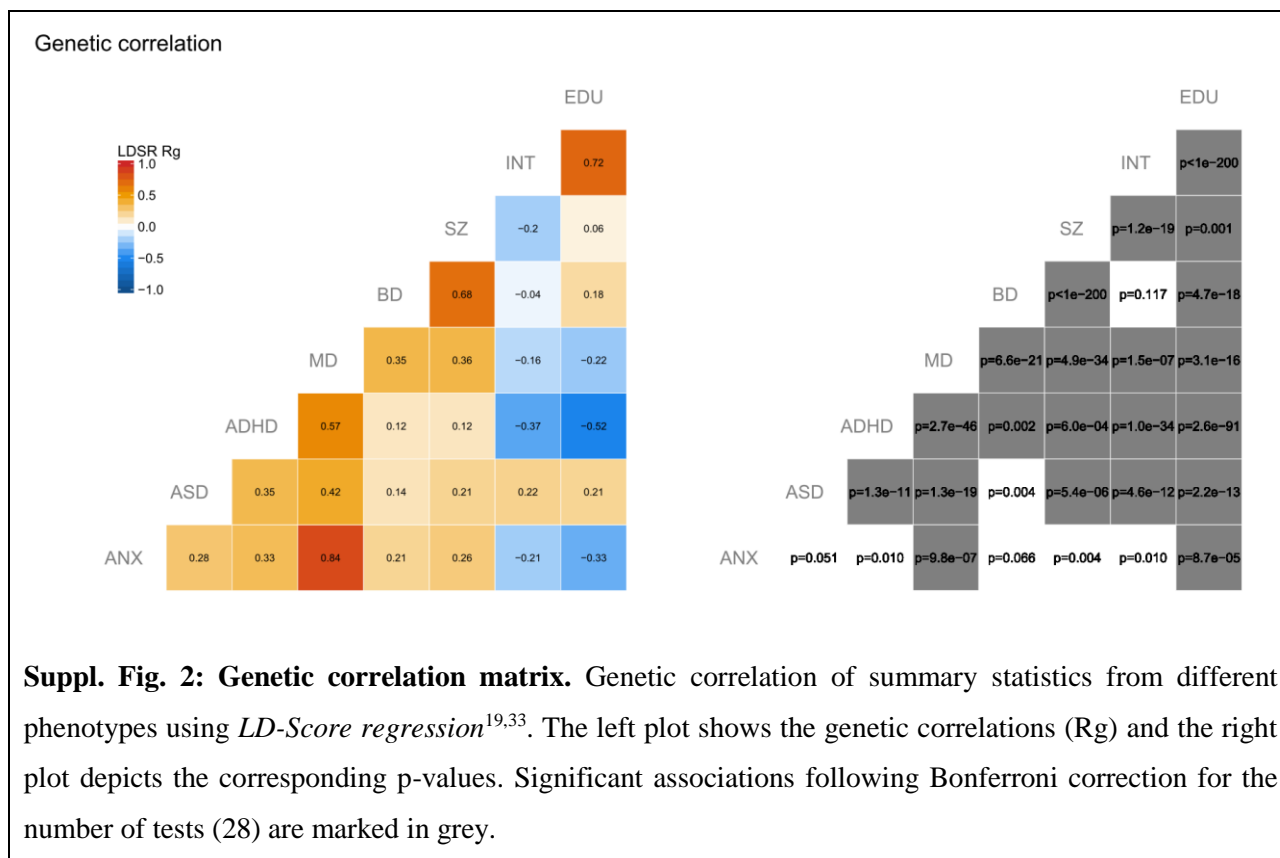
327 dashed lines indicate the 97.5% quantiles. (b) Association of thickness and area heritability maps, per

328 hemisphere. For both hemispheres concatenated, the association was $r=0.28$, $p_{\text{perm}}=.004$ (c) Cortical maps

329 for heritability of thickness (upper row) and corresponding standard error (lower row). (d) Same maps as

330 (c), but for cortical area.

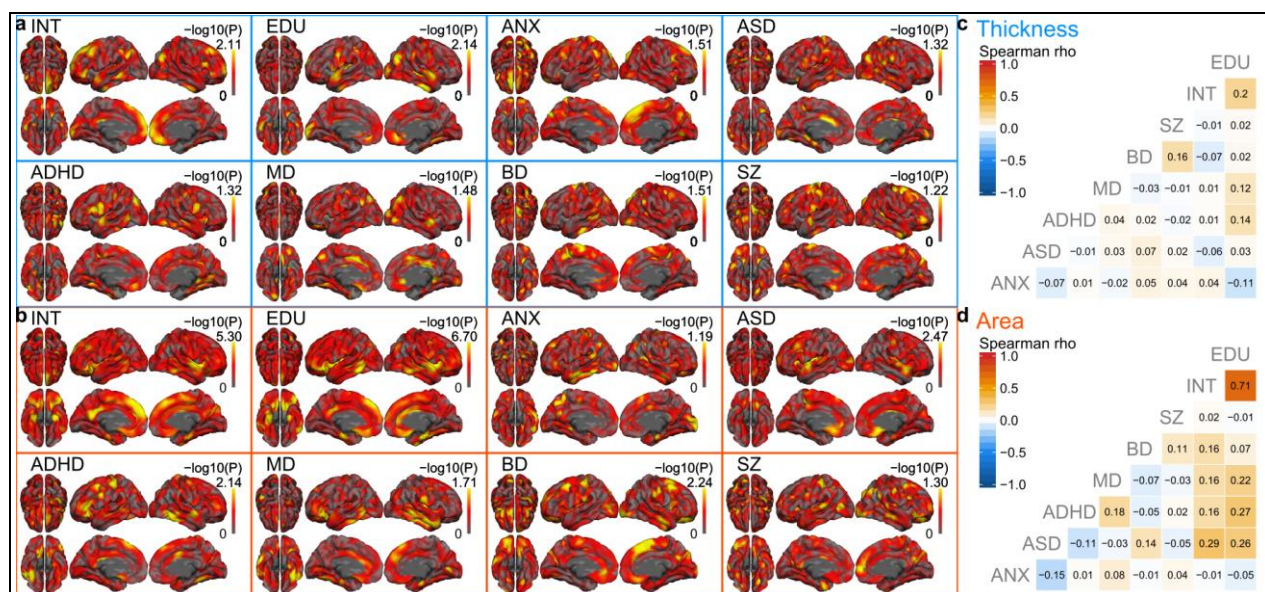
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333 **Suppl. Fig. 2: Genetic correlation matrix.** Genetic correlation of summary statistics from different
 334 phenotypes using *LD-Score regression*^{19,33}. The left plot shows the genetic correlations (R_g) and the right
 335 plot depicts the corresponding p-values. Significant associations following Bonferroni correction for the
 336 number of tests (28) are marked in grey.

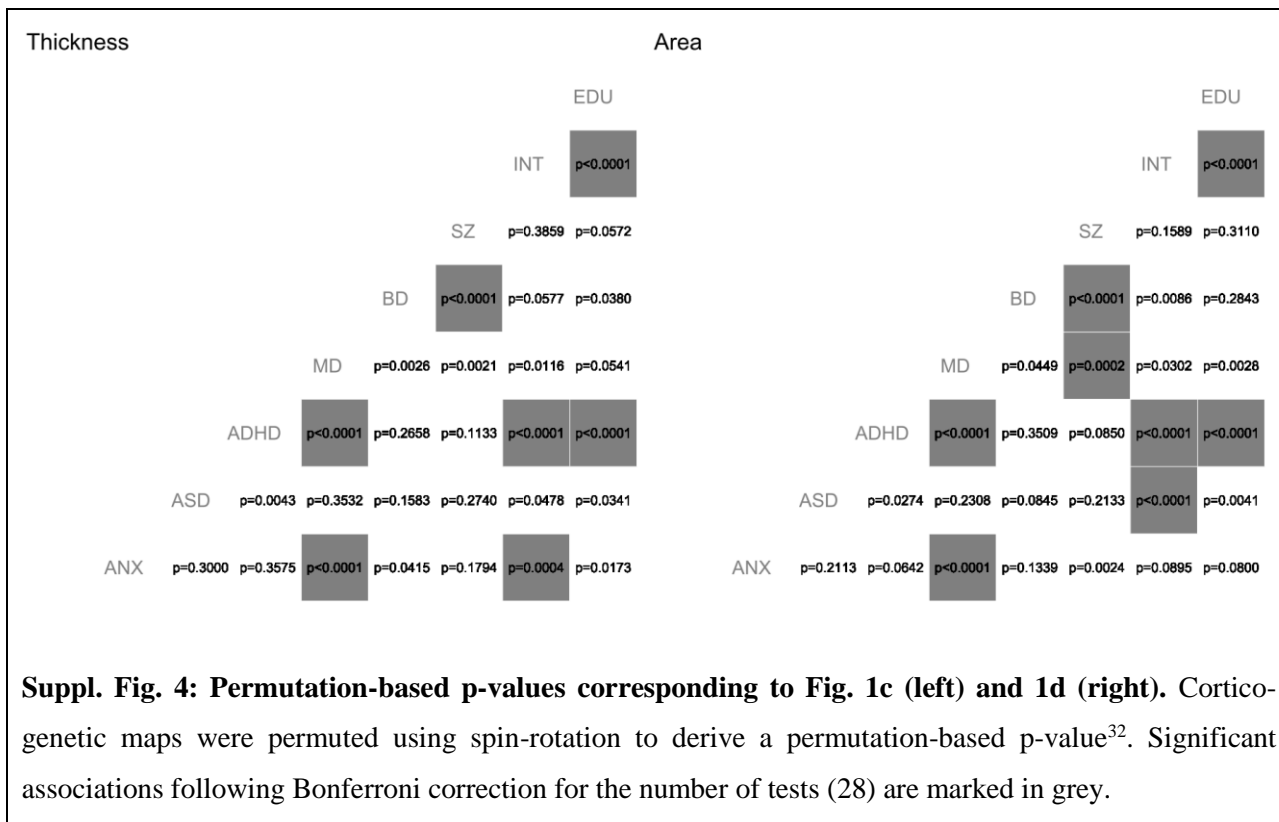
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339 **Suppl. Fig. 3: $-\log_{10}$ transformed p-values of cortico-genetic maps for cognition and psychiatric**
 340 **disorders.** Corresponding with **Fig. 1** which displays the genetic correlations (R_g), the figures display the
 341 $-\log_{10}$ transformed p-values from vertex-wise *LD-score regression*^{19,33} for (a) thickness and (b) area. (c-d)
 342 Pairwise spearman correlations of each $-\log_{10}$ transformed p-value map.

343



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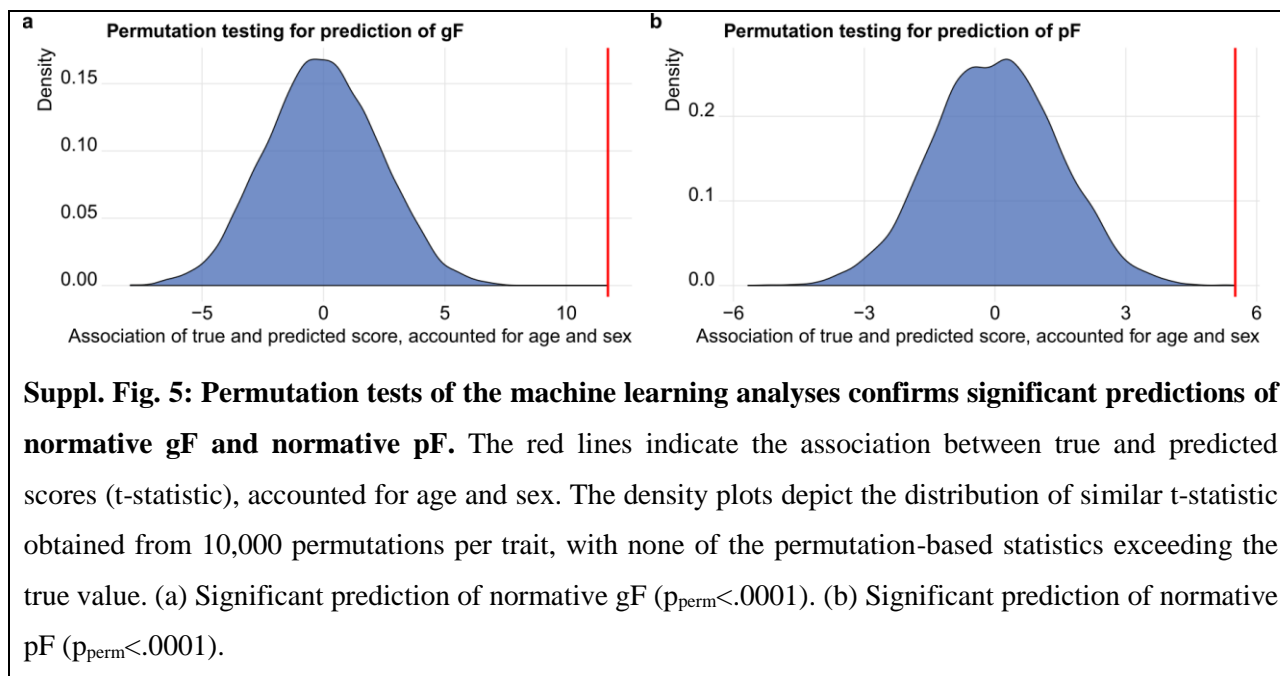
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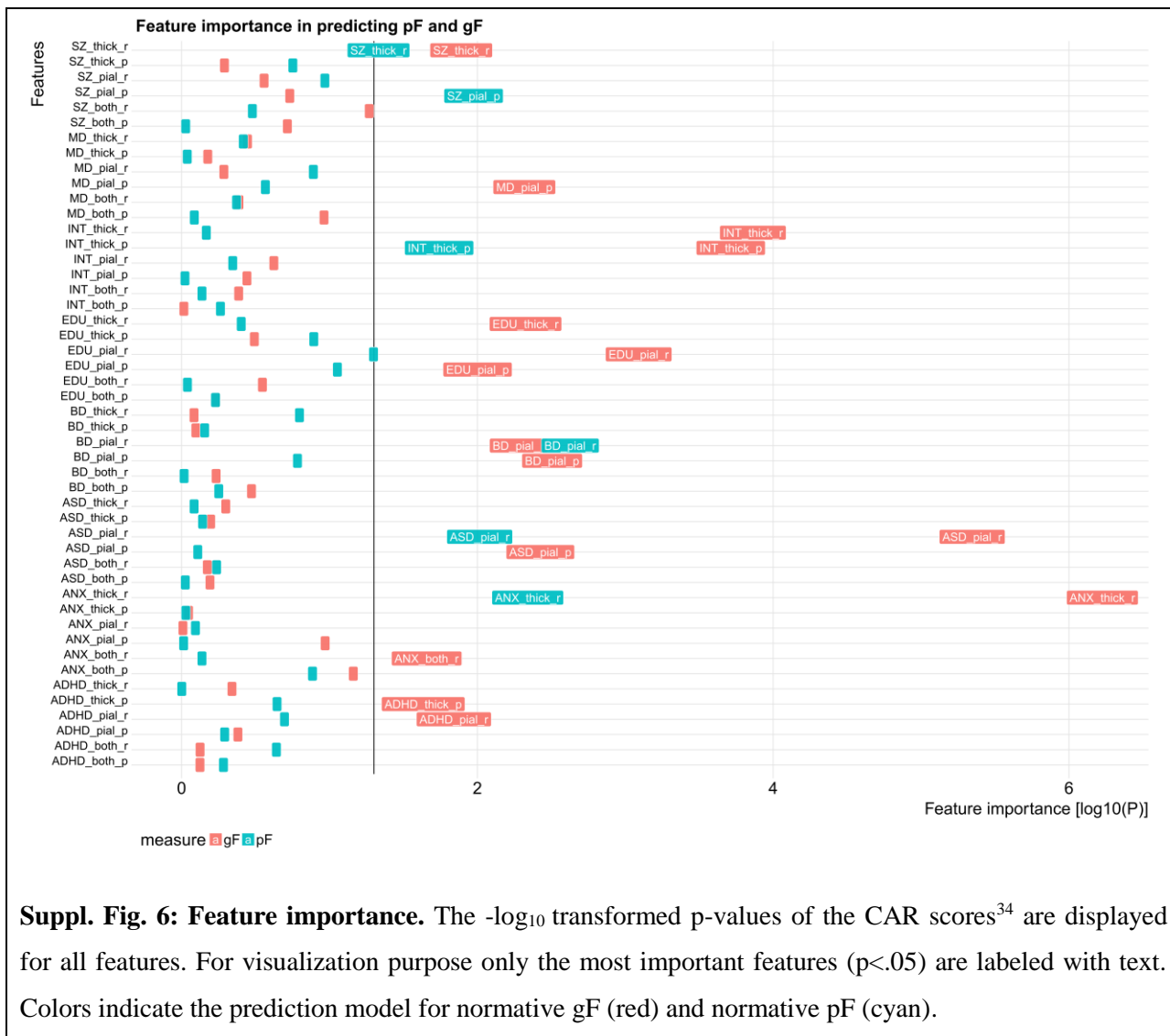
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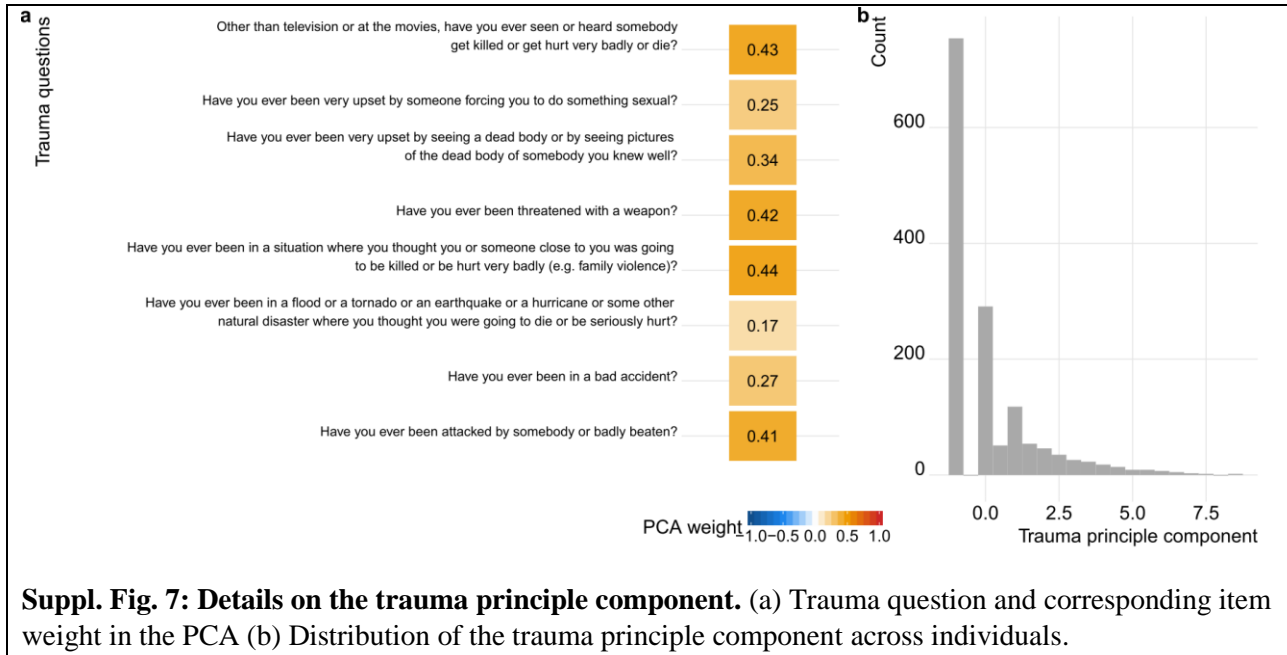
Suppl. Fig. 4: Permutation-based p-values corresponding to Fig. 1c (left) and 1d (right). Cortico-genetic maps were permuted using spin-rotation to derive a permutation-based p-value³². Significant associations following Bonferroni correction for the number of tests (28) are marked in grey.



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350 **Suppl. Fig. 5: Permutation tests of the machine learning analyses confirms significant predictions of**
351 **normative gF and normative pF.** The red lines indicate the association between true and predicted
352 scores (t-statistic), accounted for age and sex. The density plots depict the distribution of similar t-statistic
353 obtained from 10,000 permutations per trait, with none of the permutation-based statistics exceeding the
354 true value. (a) Significant prediction of normative gF ($p_{\text{perm}} < .0001$). (b) Significant prediction of normative
355 pF ($p_{\text{perm}} < .0001$).

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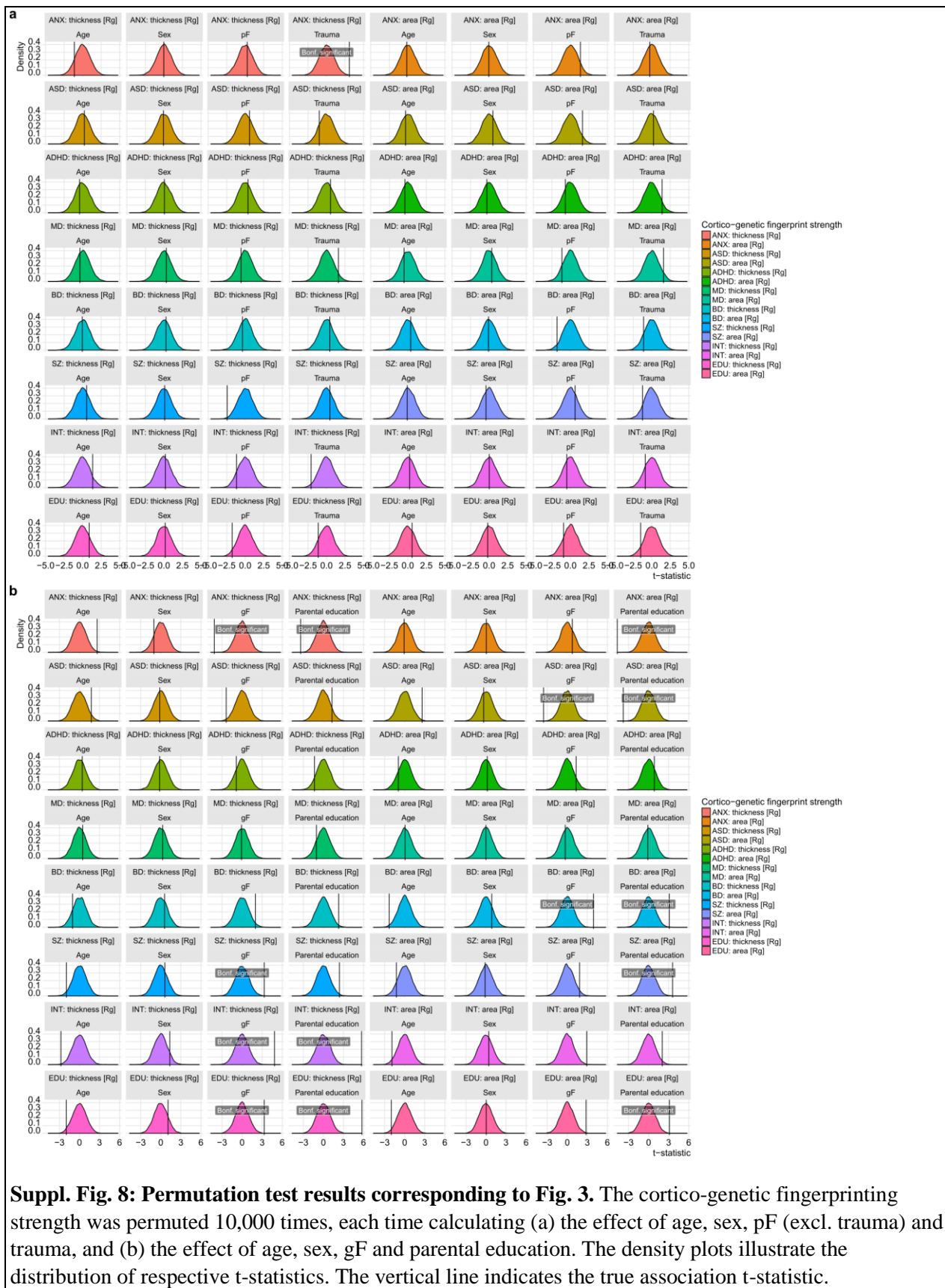


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