

1 **Boundary conditions for early life converge to an organo-**
2 **sulfur metabolism**

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4 Joshua E. Goldford^{1,2}, Hyman Hartman³, Robert Marsland III² and Daniel Segrè^{1,2,4*}

5

6 ¹Bioinformatics Program, Boston University, Boston, MA 02215, USA

7 ²Biological Design Center and Department of Physics, Boston University, Boston, MA 02215,

8 USA

9 ³Earth, Atmosphere and Planetary Science Department, Massachusetts Institute of Technology,

10 Cambridge, MA 02139, USA

11 ⁴Department of Biomedical Engineering, Department of Biology, Boston University, Boston

12 MA, 02215, USA

13 *Corresponding authors

14 Email: dsegre@bu.edu

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27 **Abstract**

28 It has been suggested that a deep memory of early life is hidden in the architecture of metabolic
29 networks, whose reactions could have been catalyzed by small molecules or minerals prior to
30 genetically encoded enzymes (1–6). A major challenge in unraveling these early steps is
31 assessing the plausibility of a connected, thermodynamically consistent proto-metabolism under
32 different geochemical conditions, which are still surrounded by high uncertainty. Here we
33 combine network-based algorithms (9, 10) with physicochemical constraints on chemical
34 reaction networks to systematically show how different combinations of parameters
35 (temperature, pH, redox potential and availability of molecular precursors) could have affected
36 the evolution of a proto-metabolism. Our analysis of possible trajectories indicates that a subset
37 of boundary conditions converges to an organo-sulfur-based proto-metabolic network fueled by a
38 thioester- and redox-driven variant of the reductive TCA cycle, capable of producing lipids and
39 keto acids. Surprisingly, environmental sources of fixed nitrogen and low-potential electron
40 donors seem not to be necessary for the earliest phases of biochemical evolution. We use one of
41 these networks to build a steady-state dynamical metabolic model of a proto-cell, and find that
42 different combinations of carbon sources and electron acceptors can support the continuous
43 production of a minimal ancient “biomass” composed of putative early biopolymers and fatty
44 acids.

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58 The structure of metabolism carries a memory of its evolutionary history that may date back to
59 before the onset of an RNA-based genetic system (1–6). Decoding this ancient evolutionary
60 record could provide important insight into the early stages of life on our planet (2, 5–8), but
61 constitutes a challenging problem. This challenge is due both to the difficulty of interrogating
62 complex biochemical networks under different environmental conditions, and to the uncertainty
63 about these conditions on prebiotic Earth. Estimates of plausible Archean environments that led
64 to the emergence and evolution of living systems vary dramatically (21, 22), ranging from
65 alkaline hydrothermal vents driven by chemical gradients (23) to acidic ocean seawater driven by
66 photochemistry (3, 4). Although geochemical data support the availability of mid-potential
67 electron donors (H₂) (24), sulfur (H₂S) and potentially fixed carbon (25) in ancient environments,
68 several key molecules used in living systems may have been severely limiting, including a
69 source of fixed nitrogen (26, 27) (e.g. ammonia), low-potential electron donors (28, 29) and
70 phosphate (30–32). Rather than assuming a steady supply of these biomolecules, one can
71 critically revisit the notion that these molecules would have been necessary for the emergence of
72 ancient proto-metabolic systems. Indeed, we recently used network-based algorithms to ask
73 whether early metabolism would have required a source of phosphate, and found evidence that
74 thioesters, rather than phosphate, may have endowed ancient metabolism with key energetic and
75 biosynthetic capacity (12). This raises the broader question of whether other molecules and
76 physico-chemical conditions may not be as crucial as previously thought for the emergence of a
77 proto-metabolism. Understanding these dependencies could also reveal whether specific
78 components of metabolism are particularly robust or fragile with respect to these initial
79 conditions.

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81 A computational method that can help address these questions is the network expansion
82 algorithm, which simulates the growth of a biochemical network by iteratively adding to an
83 initial set of compounds the products of reactions enabled by available substrates, until
84 convergence (9, 10). This algorithm, in its application to the study of ancient life (11)(12), relies
85 on three key assumptions: *first*, that chemical reactions successful in early processes were
86 gradually augmented with new pathways, but never replaced; *second*, that, over long time-scales,
87 horizontal gene transfer produced abundant shuffling of biochemical reactions across different

88 organisms (19, 20), suggesting that an ecosystem-level approach to metabolism may be
89 particularly suitable for describing ancient biochemistry; and *third*, that inorganic or small
90 molecular catalysts (12, 13) could catalyze, in a weaker and less specific manner relative to
91 modern enzymes, a large number of metabolic reactions, as confirmed by an increasing body of
92 experimental evidence (14–18).

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94 In this paper, we systematically explore a combinatorial set of molecules and parameters
95 associated with possible early Earth environments, and use an enhanced network expansion
96 algorithm to determine which proto-metabolic networks are thermodynamically reachable under
97 each of these initial conditions. We further use constraint-based flux balance modeling to
98 demonstrate the capacity of some of these networks to sustain flux, in a way that resembles
99 homeostatic growth of present-day cells. Our results suggest that a thioester-driven organic
100 network may have robustly arisen without phosphate, fixed nitrogen or low-potential electron
101 donors. This network, by supporting the biosynthesis of keto acids and fatty acids may have
102 prompted the rise of complex self-sustaining biochemical pathways, marking a key transition
103 towards the origin of life.

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105 We first sought to systematically characterize the effect of various geochemical scenarios
106 on the possible structure of ancient metabolism. Building on prior work (11, 12), we constructed
107 a model of ancient biosphere-level metabolism based on the KEGG database (33). We first
108 modified the network (as described in Methods) to account for previously proposed primitive
109 thioester-coupling and redox reactions (12). For each possible set of environmental parameters
110 (including temperature, pH and redox potential) we computed the thermodynamic feasibility of
111 each reaction, and removed infeasible reactions (see Methods). This allowed us to implement a
112 thermodynamically-constrained network expansion algorithm (12), which iteratively adds
113 metabolites and thermodynamically-feasible reactions to a network until convergence (see
114 Methods). We performed thermodynamically-constrained network expansion (see Methods and
115 Fig. 1A) for $n=672$ different geochemical scenarios, systematically varying pH, temperature,
116 redox potential of primitive redox systems, and the availability of key biomolecules including
117 thiols (that subsequently form thioesters), fixed carbon (formate/acetate) and fixed nitrogen
118 (ammonia) (Methods, Fig. 1).

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120 Of the 672 different simulated geochemical scenarios, we found that 288 (43%) expanded
121 to networks containing over 100 metabolites (Fig. 1B). A logistic regression classifier that uses
122 geochemical parameters as predictors (see Methods and Fig. 1C) allowed us to quantify the
123 importance of each environmental parameter in determining whether the expanded network
124 would reach such a large size. Surprisingly, removing the variable associated with
125 presence/absence of ammonia did not affect predictive power of the classifier, suggesting that a
126 source of fixed nitrogen is not an important determinant of the expansion. Consistent with the
127 relevance of this result to ancient metabolism, we found that the enzymes that catalyze reactions
128 in the expanded networks before the addition of ammonia were depleted in nitrogen-containing
129 coenzymes (see Fig. S1C-D, one-tailed Wilcoxon sign rank test: $P < 10^{-24}$) and in active site
130 amino acids with nitrogeneous side chains (see Fig. SE-F, one-tailed Wilcoxon sign rank test: P
131 $< 10^{-24}$) relative to enzymes added after the addition of ammonia (see Supplemental Text). These
132 results suggest that ammonia may have not been essential for the initial expansion of
133 metabolism, and point to a thioester-coupled organo-sulfur metabolic network (Fig. 1) as a core
134 network that deserves further attention.

135 Beyond the dispensability of nitrogen, the simulations described above revealed a number
136 of relationships between plausible geochemical scenarios and the structure and size of our
137 simulated proto-metabolic networks. First, expansion beyond 100 metabolites was feasible in
138 the absence of a source of fixed carbon, but only when thiols were provided in the seed set,
139 highlighting the importance for thioester-coupling for ancient carbon fixation pathways (12, 28,
140 29). The presence of thiols enabled the production of key biomolecules, including fatty acids and
141 branched chain keto acids (see Fig S2). Second, we explored the effect of the primitive redox
142 system by systematically varying the reduction potential of the electron donor in the seed set (see
143 Methods, Fig. 2A). Unexpectedly, we found that as we increased the fixed potential of the
144 electron donor, expansion to a large network was feasible over a broad range of reduction
145 potentials (between -150 and 50 mV). Only upon reaching 50mV the expanded network
146 collapsed to a much smaller solution, suggesting that the generation of low-potential electron
147 donors from H₂ may not have been a necessary condition for the early expansion of a proto-
148 metabolism (Fig. 3A). Thus, less stringent constraints, e.g. the presence of mid-potential redox
149 couples and thioester-forming thiols could have enabled the emergence of an autotrophic proto-

150 metabolic network capable of producing key biomolecules. Notably, both functions can be
151 potentially carried out by disulfides.

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153 Analysis of the expanded networks without nitrogen revealed that a large number of
154 different initial conditions converged to similar expanded organo-sulfur proto-metabolic
155 networks, spanning variants of key pathways in central carbon metabolism (Fig. 3B). For the
156 majority of simulations, variants of modern heterotrophic carbon assimilation pathways,
157 including the glyoxylate cycle and TCA cycle, were highly represented in the network (Fig. 3B).
158 Several carbon fixation pathways were highly represented in the simulated networks as well: in
159 over half of the networks that expanded beyond 100 metabolites, we found 92 % (12/13) of the
160 compounds (or generalized derivatives) that participate in the reductive tricarboxylic acid
161 (rTCA) cycle, with the exception of phosphoenolpyruvate. We also found that under several
162 geochemical conditions, all intermediates were producible for three carbon fixation pathways,
163 including the 3-hydroxypropionate bi-cycle, the hydroxypropionate-hydroxybutylate cycle, and
164 the dicarboxylate-hydroxybutyrate cycle (Fig. 3A). At-most, only 3 of 9 metabolites used in the
165 Wood-Ljungdahl (WL) pathway were observed, due to the lack of nitrogen-containing pterins in
166 the network. This does not necessarily rule out the primordial importance of the WL-pathway, as
167 its early variants could have been radically different than today's WL-pathway, relying on native
168 metals to facilitate reduction of CO₂ to acetate (25, 29). In addition to observing a large number
169 of metabolites used in carbon fixation pathways, we found that a large fraction of the β -oxidation
170 pathway was represented in our networks, which may have supported the production of fatty
171 acids in ancient living systems by operating in the reverse direction. Lastly, we observed that the
172 majority of intermediates involved in the production of branched-chain amino acids were also
173 producible in the expanded networks.

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175 A more detailed analysis of the convergent organo-sulfur proto-metabolic network
176 reveals new possible ancestral metabolic pathways that involve previously unexplored
177 combinations of reactions and metabolites. Fig. 3B shows a variant of the (r)TCA cycle that is a
178 component of these expanded networks, and that may have served as the core organo-sulfur
179 network fueling ancient living systems. Rather than using ATP-dependent reactions found in
180 extant species (e.g. Succinyl-CoA synthetase and ATP citrate lyase), these reactions are

181 substituted with non-ATP-dependent reaction mechanisms. For instance, the production of a
182 succinyl-thioester in the extant rTCA cycle relies on Succinyl-CoA synthetase, performing the
183 following reaction: $\text{ATP} + \text{Succinate} + \text{CoA} \rightarrow \text{Succinyl-CoA} + \text{ADP} + \text{Pi}$. However, in the
184 network presented in Fig. 3B, malyl-thioester, producible through alternative reactions, donates a
185 thiol to succinate, subsequently forming a succinyl-thioester. This (r)TCA cycle analogue is able
186 to produce eight keto acids normally serving as key intermediates and precursors to common
187 amino acids in central carbon metabolism (glyoxylate, pyruvate, oxaloacetate, 2-oxoglutarate
188 and hydroxypyruvate), as well as a few branched-chain keto acids. Additionally, long-chain
189 fatty acids like palmitate are producible in this network, driven by thioester and redox-coupling
190 rather than ATP, like in extant fatty acid biosynthesis. Thus, despite the simplicity of seed
191 compounds, several small molecular weight keto acids and fatty acids may have been producible
192 in an organo-sulfur proto-metabolism.

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194 So far, we have focused only on the topology and global thermodynamic feasibility of
195 putative ancient metabolic networks. Inspired by recent studies on the molecular budget of
196 present-day cells, we decided to further explore whether proto-metabolic networks could support
197 steady state fluxes, and fuel primitive proto-cells with internal energy sources (e.g. thioesters),
198 redox gradients, and primitive biopolymer capable of catalysis and compartmentalization. Flux
199 balance analysis (FBA), originally developed for the study of microbial metabolism, enables the
200 prediction of systems-level properties of metabolic networks at steady-state (34). Fundamentally,
201 FBA computes possible reaction rates in a network constrained by mass and energy balance,
202 usually under the assumption that a specific composition of biomolecules is efficiently produced
203 during a homeostatic growth process. In microbial metabolism, FBA is used to simulate the
204 production of cellular biomass (e.g. protein, lipids, and nucleic acids) at fixed proportions, which
205 are derived from known composition of extant cells. We realized that the same approach could
206 help test the sustainability of a proto-metabolic biochemical system, provided that we could
207 develop a plausible hypothesis for the “biomass composition” of ancient proto-cells. As a
208 starting point, we recalled Christian de Duve’s suggestion that the thioester-driven
209 polymerization of monomers producible from ancient proto-metabolism may have led to
210 "catalytic multimers," which could have served as catalysts for ancient biochemical reactions (4).
211 Under nitrogen limited conditions, keto acids producible from proto-metabolism (see Fig. 3B)

212 could have been reduced to α -hydroxy acids, and polymerized into polyesters using thioesters as
213 a condensing agent (see Fig. S3). Recent work has suggested that polymers of α -hydroxy acids
214 may have been stably produced in geochemical environments (35), and that these molecules
215 could have served as primitive catalysts (36). These results all point to the intriguing possibility
216 that the thioester-driven polymerization of α -hydroxy acids (producible from keto acid
217 precursors of common amino acids) generated the first metabolically sustainable cache of ancient
218 catalysts, leading to a collectively autocatalytic protocellular system. We employed FBA to
219 specifically test the feasibility of such a system. Using an expanded metabolic network as a
220 scaffold for network reconstruction (Fig. 3B), we constructed a constraint-based model of an
221 ancient proto-cell using a biomass composition consisting of fatty acids (for proto-cellular
222 membranes), "catalytic multimers" derived from eight keto acids (Fig. 4B), and redox and
223 thioester-based free energy sources (Methods, Fig. 4A). We used thermodynamic metabolic flux
224 analysis (TMFA), a variant of FBA that explicitly considers thermodynamic constraints (37) (see
225 Methods), to determine whether homeostatic growth of the whole system was achievable. We
226 found that growth of the proto-cell metabolic model is indeed feasible under a wide variety of
227 assumptions regarding macromolecular compositions and input molecules (Fig. 4B). Notably,
228 growth is achievable in simple chemoautotrophic conditions with either H_2 or H_2S , but not
229 $Fe(II)$, as electron donors (Fig. 4B). In this model, thiols and thioesters are not supplied as food
230 sources, but rather are recycled during steady-state growth of the proto-cell. This reflects the
231 possibility that thiols could have been initially supplied abiotically, followed by the rapid
232 takeover of biotic production of mercaptopyruvate, a keto acid that could have been incorporated
233 into primitive multimers.

234
235 While most efforts to reconstruct ancient biochemistry have traditionally relied on
236 building qualitative models of small pathways (2, 4, 5, 7, 38), we found that quantitative
237 modeling of larger networks can provide substantial new insight into the origin of life. By
238 computationally mapping geochemical scenarios to plausible ancient proto-metabolic structures,
239 we estimated which portions of extant biochemistry may have been very sensitive or very robust
240 to initial geochemical conditions. Our approach reveals that, contrary to expectations (8, 28, 39),
241 environmental sources of fixed nitrogen and low-potential electron donors may have not been
242 necessary for early biochemical evolution, and a substantial degree of complexity may have

243 emerged prior to incorporation of nitrogen into the biosphere (3). The key catalytic role played
244 by nitrogen in the active sites of modern enzymes may have been preceded by positively charged
245 surfaces or metal ions (18, 25), which could have been replaced by amino/keto acids with
246 nitrogen side chains once nitrogen became incorporated into proto-metabolism. Our simulations
247 also cast doubts on the essential role of a low-potential electron donor in early life (8, 28, 29),
248 consistent with the proposal that low-potential electron donors may not be necessary for
249 acetogenesis (40), and with the possibility that energy conservation via electron bifurcation
250 might not have been necessary in primordial metabolism. The independence of our inferred
251 ancestral networks of low-potential electron donors and ATP, both key substrates for nitrogen
252 fixation (41), suggests that nitrogen fixation may have evolved later throughout the history of life
253 (42–44). A striking feature of our analysis is the convergence of multiple geochemical scenarios
254 towards a core organo-sulfur proto-metabolic network capable of producing various keto acids
255 and fatty acids (Fig. 3B). This feature provides a window into how thioester-driven
256 polymerization of α -hydroxy acid monomers (derived from producible keto acids) could have
257 added primitive macromolecular organic catalysts (4) to initial inorganic minerals or metal ion
258 catalysts (18, 25). Further tests of this hypothesis could be pursued by measuring the capacity of
259 these polymers to catalyze key reactions in the network, and by exploring whether these organic
260 compounds are produced in living systems today via mechanisms similar to polyketide or non-
261 ribosomal peptide synthesis. Finally, our constraint-based models of this core organo-sulfur
262 proto-metabolism provide a first example of how network expansion-based predictions can be
263 translated into dynamical models, whose capacity to estimate sustainable collective growth can
264 drive the search for specific self-reproducing chemical networks and metabolically-driven
265 artificial protocells.

266

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272 Contributions

273 J.E.G., H.H. and D.S. designed the research. J.E.G. wrote code, ran simulations and performed
274 analysis. R.M. contributed to the non-equilibrium steady-state modeling. J.G. and D.S. wrote the
275 manuscript. All authors read and approved the final manuscript.

276 Competing Financial Interests

277 The authors declare no competing financial interests.

278 Corresponding Authors

279 Correspondence to: dsegre@bu.edu.

280 References

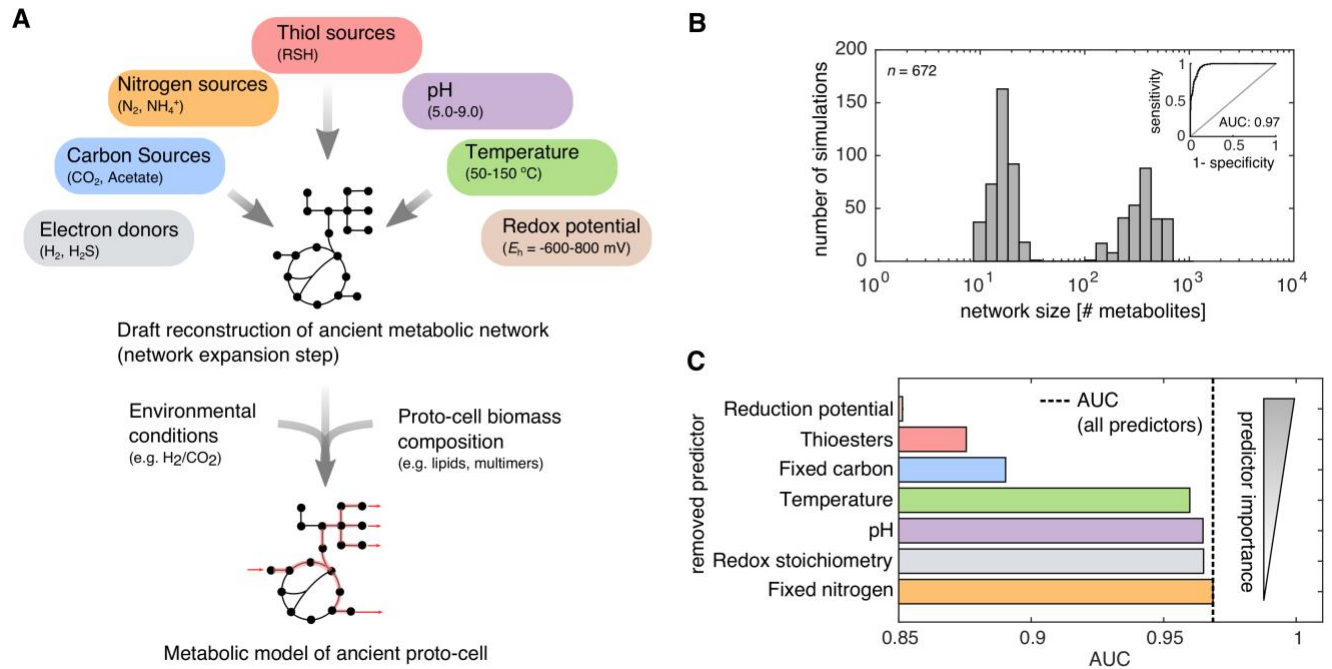
- 281 1. Eck, R. V., and Dayhoff, M. O. (1966) Evolution of the structure of ferredoxin based on
282 living relics of primitive amino Acid sequences. *Science*. **152**, 363–366
- 283 2. Hartman, H. (1975) Speculations on the origin and evolution of metabolism. *J. Mol. Evol.*
284 **4**, 359–370
- 285 3. Hartman, H. (1992) Conjectures and reveries. *Photosynth. Res.* **33**, 171–6
- 286 4. de Duve, C. (1991) *Blueprint for a cell: the nature and origin of life*, Neil Patterson
287 Publishers, Carolina Biological Supply Company, Burlington, N.C.
- 288 5. Morowitz, H. J., Kostelnik, J. D., Yang, J., and Cody, G. D. (2000) The origin of
289 intermediary metabolism. *Proc. Natl. Acad. Sci. U. S. A.* **97**, 7704–8
- 290 6. Smith, E., and Morowitz, H. J. (2016) *The Origin and Nature of Life On Earth*, 1st Ed.,
291 Cambridge University Press, Cambridge, United Kingdom, 10.1017/CBO9781316348772
- 292 7. Smith, E., and Morowitz, H. J. (2004) Universality in intermediary metabolism. *Proc.*
293 *Natl. Acad. Sci. U. S. A.* **101**, 13168–73
- 294 8. Sousa, F. L., Thiergart, T., Landan, G., Nelson-Sathi, S., Pereira, I. a C., Allen, J. F., Lane,
295 N., and Martin, W. F. (2013) Early bioenergetic evolution. *Philos. Trans. R. Soc. Lond. B.*
296 *Biol. Sci.* **368**, 20130088
- 297 9. Ebenhöh, O., Handorf, T., and Heinrich, R. (2004) Structural analysis of expanding
298 metabolic networks. *Genome Inform.* **15**, 35–45

- 299 10. Handorf, T., Ebenhöf, O., and Heinrich, R. (2005) Expanding metabolic networks: scopes
300 of compounds, robustness, and evolution. *J. Mol. Evol.* **61**, 498–512
- 301 11. Raymond, J., and Segrè, D. (2006) The effect of oxygen on biochemical networks and the
302 evolution of complex life. *Science.* **311**, 1764–7
- 303 12. Goldford, J. E., Hartman, H., Smith, T. F., and Segrè, D. (2017) Remnants of an Ancient
304 Metabolism without Phosphate. *Cell.* **168**, 1126–1134.e9
- 305 13. Goldford, J. E., and Segrè, D. (2018) Modern views of ancient metabolic networks. *Curr.*
306 *Opin. Syst. Biol.* **8**, 117–124
- 307 14. Keller, M. A., Zylstra, A., Castro, C., Turchyn, A. V., Griffin, J. L., and Ralser, M. (2016)
308 Conditional iron and pH-dependent activity of a non-enzymatic glycolysis and pentose
309 phosphate pathway. *Sci. Adv.* **2**, e1501235
- 310 15. Keller, M. A., Turchyn, A. V., and Ralser, M. (2014) Non-enzymatic glycolysis and
311 pentose phosphate pathway-like reactions in a plausible Archean ocean. *Mol. Syst. Biol.*
312 **10**, 725
- 313 16. Messner, C. B., Driscoll, P. C., Piedrafita, G., De Volder, M. F. L., and Ralser, M. (2017)
314 Nonenzymatic gluconeogenesis-like formation of fructose 1,6-bisphosphate in ice. *Proc.*
315 *Natl. Acad. Sci.* **114**, 7403–7407
- 316 17. Keller, M. A., Piedrafita, G., and Ralser, M. (2015) The widespread role of non-enzymatic
317 reactions in cellular metabolism. *Curr. Opin. Biotechnol.* **34**, 153–161
- 318 18. Muchowska, K. B., Varma, S. J., Chevallot-Beroux, E., Lethuillier-Karl, L., Li, G., and
319 Moran, J. (2017) Metals promote sequences of the reverse Krebs cycle. *Nat. Ecol. Evol.* **1**,
320 1716–1721
- 321 19. Vetsigian, K., Woese, C., and Goldenfeld, N. (2006) Collective evolution and the genetic
322 code. *Proc. Natl. Acad. Sci. U. S. A.* **103**, 10696–701
- 323 20. Ochman, H., Lawrence, J. G., and Groisman, E. A. (2000) Lateral gene transfer and the
324 nature of bacterial innovation. *Nature.* **405**, 299–304
- 325 21. Lazcano, A., and Miller, S. L. (1996) The origin and early evolution of life: prebiotic
326 chemistry, the pre-RNA world, and time. *Cell.* **85**, 793–8
- 327 22. Deamer, D., and Weber, A. L. (2010) Bioenergetics and life’s origins. *Cold Spring Harb.*
328 *Perspect. Biol.* **2**, a004929
- 329 23. Martin, W., Baross, J., Kelley, D., and Russell, M. J. (2008) Hydrothermal vents and the
330 origin of life. *Nat. Rev. Microbiol.* **6**, 805–14
- 331 24. Russell, M. J., Hall, A. J., and Martin, W. (2010) Serpentinization as a source of energy at
332 the origin of life. *Geobiology.* **8**, 355–71
- 333 25. Varma, S. J., Muchowska, K. B., Chatelain, P., and Moran, J. (2018) Native iron reduces
334 CO₂ to intermediates and end-products of the acetyl-CoA pathway. *Nat. Ecol. Evol.*

- 335 10.1038/s41559-018-0542-2
- 336 26. Dörr, M., Kässbohrer, J., Grunert, R., Kreisel, G., Brand, W. A., Werner, R. A., Geilmann,
337 H., Apfel, C., Robl, C., and Weigand, W. (2003) A possible prebiotic formation of
338 ammonia from dinitrogen on iron sulfide surfaces. *Angew. Chem. Int. Ed. Engl.* **42**, 1540–
339 3
- 340 27. Navarro-González, R., McKay, C. P., and Mvondo, D. N. (2001) A possible nitrogen
341 crisis for Archaean life due to reduced nitrogen fixation by lightning. *Nature.* **412**, 61–64
- 342 28. Martin, W. F., and Thauer, R. K. (2017) Energy in Ancient Metabolism. *Cell.* **168**, 953–
343 955
- 344 29. Sousa, F. L., Preiner, M., and Martin, W. F. (2018) Native metals, electron bifurcation,
345 and CO₂ reduction in early biochemical evolution. *Curr. Opin. Microbiol.* **43**, 77–83
- 346 30. Halmann, M. (1974) Evolution and Ecology of Phosphorus Metabolism. in *The Origin of*
347 *Life and Evolutionary Biochemistry* (Dose, K., Fox, S. W., Deborin, G. A., and
348 Pavlovskaya, T. E. eds), pp. 169–182, Springer US
- 349 31. Schwartz, A. W. (2006) Phosphorus in prebiotic chemistry. *Philos. Trans. R. Soc. Lond.*
350 *B. Biol. Sci.* **361**, 1743–9
- 351 32. Keefe, A. D., and Miller, S. L. (1995) Are polyphosphates or phosphate esters prebiotic
352 reagents? *J. Mol. Evol.* **41**, 693–702
- 353 33. Kanehisa, M., and Goto, S. (2000) KEGG: kyoto encyclopedia of genes and genomes.
354 *Nucleic Acids Res.* **28**, 27–30
- 355 34. Orth, J. D., Thiele, I., and Palsson, B. O. (2010) What is flux balance analysis? *Nat*
356 *Biotech.* **28**, 245–248
- 357 35. Chandru, K., Guttenberg, N., Giri, C., Hongo, Y., Butch, C., Mamajanov, I., and Cleaves,
358 H. J. (2018) Simple prebiotic synthesis of high diversity dynamic combinatorial polyester
359 libraries. *Commun. Chem.* **1**, 30
- 360 36. Forsythe, J. G., Yu, S. S., Mamajanov, I., Grover, M. A., Krishnamurthy, R., Fernández,
361 F. M., and Hud, N. V. (2015) Ester-Mediated Amide Bond Formation Driven by Wet-Dry
362 Cycles: A Possible Path to Polypeptides on the Prebiotic Earth. *Angew. Chemie - Int. Ed.*
363 **54**, 9871–9875
- 364 37. Henry, C. S., Broadbelt, L. J., and Hatzimanikatis, V. (2007) Thermodynamics-based
365 metabolic flux analysis. *Biophys. J.* **92**, 1792–1805
- 366 38. Wächtershäuser, G. (1990) Evolution of the first metabolic cycles. *Proc. Natl. Acad. Sci.*
367 **87**, 200–204
- 368 39. Martin, W., and Russell, M. J. (2007) On the origin of biochemistry at an alkaline
369 hydrothermal vent. *Philos. Trans. R. Soc. London B Biol. Sci.* **362**, 1887–1926
- 370 40. Bar-Even, A. (2013) Does acetogenesis really require especially low reduction potential?

- 371 *Biochim. Biophys. Acta.* **1827**, 395–400
- 372 41. Duval, S., Danyal, K., Shaw, S., Lytle, A. K., Dean, D. R., Hoffman, B. M., Antony, E.,
373 and Seefeldt, L. C. (2013) Electron transfer precedes ATP hydrolysis during nitrogenase
374 catalysis. *Proc. Natl. Acad. Sci.* **110**, 16414–16419
- 375 42. Weiss, M. C., Sousa, F. L., Mrnjavac, N., Neukirchen, S., Roettger, M., Nelson-sathi, S.,
376 and Martin, W. F. (2016) The physiology and habitat of the last universal common
377 ancestor. *Nat. Microbiol.* **1**, 1–8
- 378 43. Gogarten, J. P., and Deamer, D. (2016) Is LUCA a thermophilic progenote? *Nat.*
379 *Microbiol.* **1**, 16229
- 380 44. Weiss, M. C., Neukirchen, S., Roettger, M., Mrnjavac, N., Nelson-Sathi, S., Martin, W.
381 F., and Sousa, F. L. (2016) Reply to “Is LUCA a thermophilic progenote?” *Nat.*
382 *Microbiol.* **1**, 16230
- 383 45. Mall, A., Sobotta, J., Huber, C., Tschirner, C., Kowarschik, S., Bačnik, K., Mergelsberg,
384 M., Boll, M., Hügler, M., Eisenreich, W., and Berg, I. A. (2018) Reversibility of citrate
385 synthase allows autotrophic growth of a thermophilic bacterium. *Science (80-.).* **359**,
386 563–567
- 387 46. Nunoura, T., Chikaraishi, Y., Izaki, R., Suwa, T., Sato, T., Harada, T., Mori, K., Kato, Y.,
388 Miyazaki, M., Shimamura, S., Yanagawa, K., Shuto, A., Ohkouchi, N., Fujita, N., Takaki,
389 Y., Atomi, H., and Takai, K. (2018) A primordial and reversible TCA cycle in a
390 facultatively chemolithoautotrophic thermophile. *Science (80-.).* **359**, 559–563
- 391

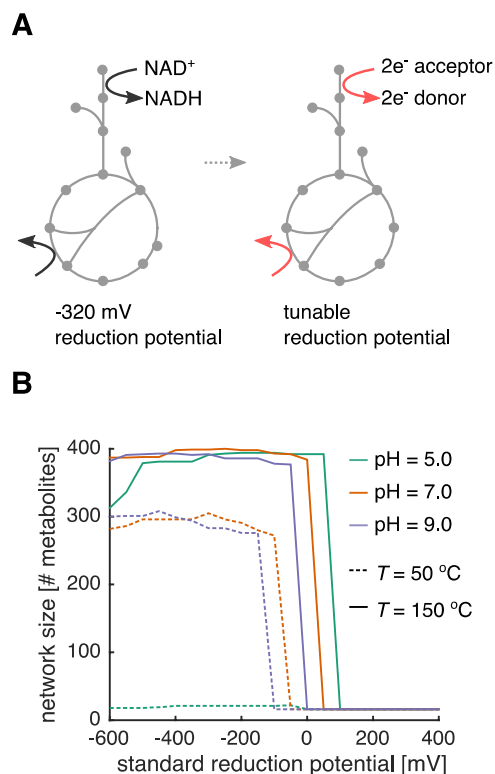
392 **Figures**



393

394 **Figure 1: Nitrogen is not essential for the initial expansion of metabolism.** (A) A
 395 thermodynamically-constrained network expansion algorithm was used to simulate the early
 396 expansion of metabolism under 672 scenarios, systematically varying the availability of
 397 reductants in the environment, pH, carbon sources, the presence of thiols, temperature and the
 398 availability of ammonia. (B) A histogram of network sizes (*x*-axis, number of metabolites)
 399 revealed that 43 % (288/672) of the scenarios resulted a bimodal distribution, where expansion
 400 occurred beyond 100 metabolites. (inset) A logistic regression classifier was constructed to
 401 predict whether a geochemical scenario resulted in a network that exceeded 100 metabolites, and
 402 a receiver operating curve (ROC) was plotted. The trained classifier resulted in an area under the
 403 curve (AUC) of 0.97 and leave-one out cross-validation accuracy of 0.89. (C) Models were
 404 trained without information on specific geochemical variables (*y*-axis), and the ensuing AUC
 405 was plotted as a bar-chart (*x*-axis), revealing that knowledge of the availability of fixed nitrogen
 406 offers no information on whether networks expanded.

407



408 **Figure 2: Reduction potential of nicotinamide and flavin substitutes influences network**
409 **expansion.** (A) Redox coenzymes (NAD, NADP, and FAD) were substituted with an arbitrary
410 electron donor/acceptor at a fixed reduction potential. (B) We performed thermodynamic
411 network expansion in acidic (pH 5), neutral (pH 7) and alkaline (pH 9) conditions at two
412 temperatures ($T=50$ and 150 °C), using a two-electron redox couple at a fixed potential (x -axis)
413 as a substitute for NAD(P)/FAD coupling in extant metabolic reactions (see Methods). We
414 plotted the final network size across all pH and temperatures with no fixed carbon sources (e.g.
415 only CO_2) and thiols. Notably, for these simulations, we used a base seed set of: H_2 , H_2S , H_2O ,
416 HCO_3^- , H^+ and CO_2 .

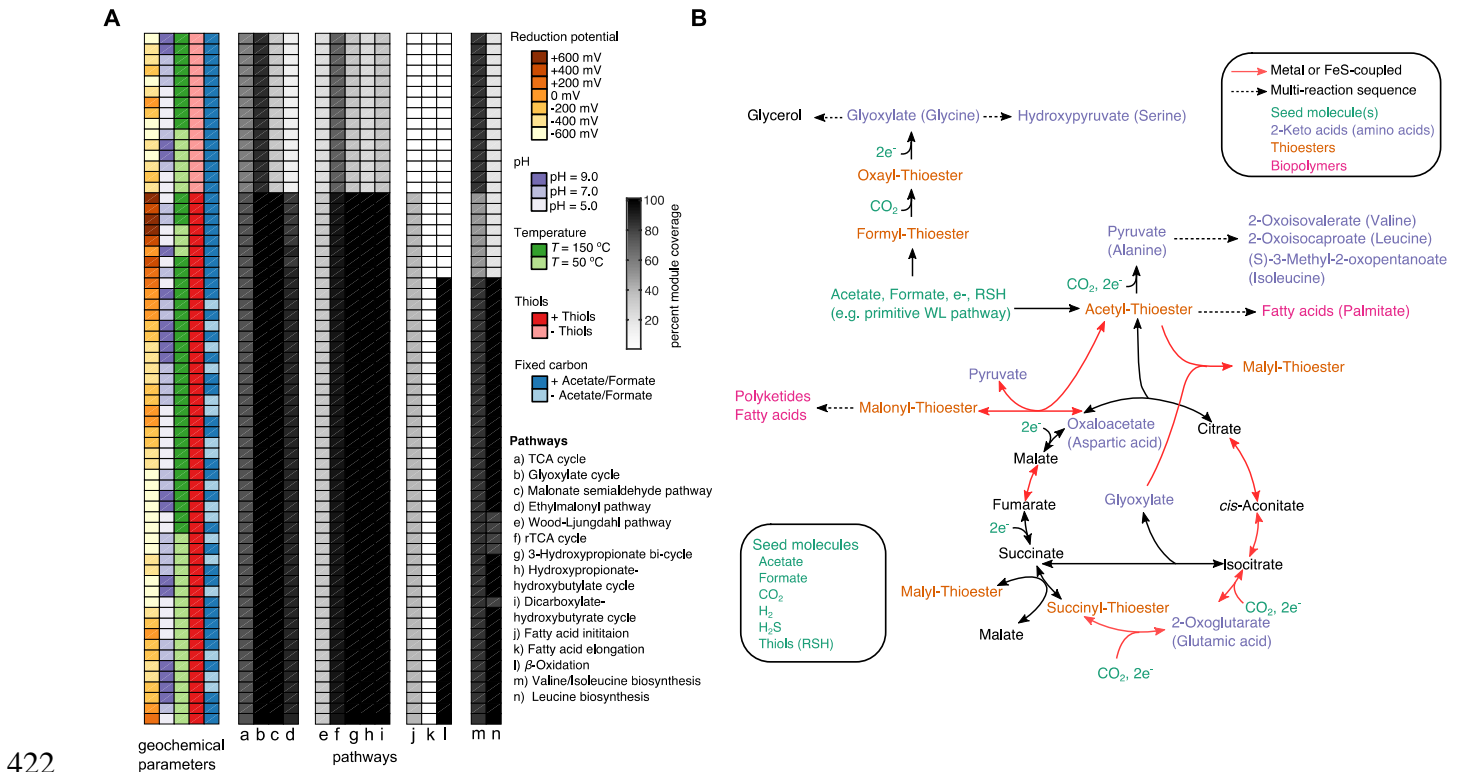
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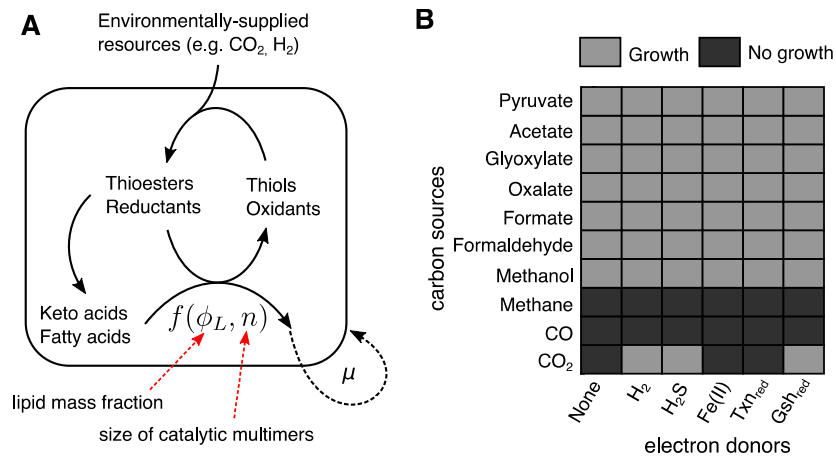


422

423 **Figure 3: Systematic exploration of prebiotic scenarios reveals a core organo-sulfur**
 424 **network.** (A) A thermodynamically constrained network expansion algorithm was used to
 425 simulate the early expansion of proto-metabolism under various scenarios, including the
 426 availability of reductants in the environment, pH, temperature, and the availability of fixed
 427 carbon sources and thiols. The proportion of molecules selected KEGG modules involved carbon
 428 metabolism are plotted as a heatmap to the right of the parameters. (B) A representation of the
 429 core network producible from a prebiotically plausible seed set without nitrogen or phosphate
 430 (bottom left box). Acetyl-thioesters are first produced, potentially from a primitive Wood-
 431 Ljungdahl pathway (8, 25) from acetate and thiols provided as seed molecules (green). Acetyl-
 432 thioesters enable the production of all intermediates in the reductive tricarboxylic acid (rTCA)
 433 cycle, with the exception of phosphoenolpyruvate. ATP-dependent reactions in the rTCA cycle
 434 may have been substituted with a primitive malate synthase and transthioesterification of
 435 succinate as well as the recently discovered reversible citrate synthase (45, 46). The keto acid
 436 precursors for 8 common amino acids (A,D,E,G,I,L,S,V) are highlighted in purple, while routes
 437 to thioester-mediated polymerization of fatty acids and polyketides are highlighted in pink.

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439



440

441 **Figure 4: Constraint-based modeling of plausible ancient proto-cells.** (A) We constructed a
 442 metabolic model of a plausible ancient proto-cell and used thermodynamic metabolic flux
 443 analysis (37) to simulate the maximum growth yields, steady-state fluxes, and metabolite
 444 concentrations under a variety of environmental conditions. The metabolic model was
 445 constructed using internally-generated reductants and thioesters that fueled biomass formation.
 446 The biomass composition was specified as variable fractions of fatty acids, and polymerized
 447 hydroxyacids from keto-acid precursors (see Methods). In this model, the internal redox
 448 coenzyme was assumed to be disulfide/dithiol at a standard reduction potential of -220 mV, and
 449 the production of biomass was fueled by the hydrolysis of acetyl-thioesters. We parameterized
 450 the biomass composition using a two-parameter model (see Methods), with the mass fraction of
 451 lipids in the proto-cell set to $\phi_L = 0.2$, and the average size of a catalytic multimer, $n = 10$. (B)
 452 We simulated growth on a variety of simple carbon sources (y-axis) and electron donors (x-axis),
 453 and show environments supporting non-zero growth (light grey) or no growth (dark grey).
 454 Interestingly, H₂, H₂S and glutathione were the only reductants capable of supporting fully
 455 autotrophic growth on CO₂. Furthermore, CO and Methane could not support growth in this
 456 model, while other one-carbon sources like methanol, formate and formaldehyde could support
 457 biomass growth.