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**Homogenous subgroups of atypical meningiomas defined using oncogenic signatures:
basis for a new grading system?**

Abstract

Meningiomas are the most common brain tumor with a prevalence of 3% in the population. Histological grading of meningiomas (1 through 3) has a major role in determining treatment choice and predicting outcome. While largely indolent grade 1 and the highly aggressive grade 3 meningiomas as considered mostly homogenous in clinical behavior, atypical or grade 2 meningiomas have highly diverse biological properties. Our aim was to identify homogenous subgroups of atypical meningiomas with the working hypothesis that these subgroups would share features with grade 1 and grade 3 counterparts. We carried out systems level analysis by gene module discovery using co-expression networks on the transcriptomics of 212 meningiomas. The newly identified subgroups were characterized in terms of recurrence rate and overlapping biological processes in gene ontology. We were able to reclassify 33 of 46 atypical meningiomas (72%) into a benign “grade 1-like” (14/46) and malignant “grade 3-like” (19/46) subgroup based on oncogenic signatures. Recurrence rates of “Grade 1-like” and “grade 3- like” tumors was 0% and 72% respectively. These two new subgroups showed similar recurrence rates and concordant biological processes with the respected grades. Our findings help resolve the heterogeneity/uncertainty around atypical meningioma biology and identify subgroups more homogenous than in prior studies. These results may help reshape prediction, follow-up planning, treatment decisions and recruitment protocols for future and ongoing clinical trials. The findings demonstrate the conceptual advantage of systems biology approaches and underpin the utility of molecular signatures as complements to the current histological grading system.

Keywords: meningioma, transcriptomics, gene expression networks

Introduction

Meningiomas are the most common adult brain tumor, carrying an overall prevalence of 3% in the population. Histopathologic analysis is the mainstay of diagnosis and together with the extent of surgical resection it is a determinant of outcome and treatment planning^{1,2}. According to World Health Organization (WHO) grading, majority of meningiomas (almost 70%) fall into grade 1, of which about two thirds are cured with surgical excision alone² and 15-20 % recur within five years of diagnosis³. Grade 3 meningiomas, by contrast, are rare and aggressive with a five year recurrence rate of approximately 90%². Grade 2 (atypical) meningiomas constitute 20-30% of cases and represent a biological intermediate. Predicting the clinical course for these tumors is particularly challenging given their heterogeneous biology which yields a five-year recurrence rate of approximately 50%⁴. Previous studies have suggested that some grade 2 meningiomas share features with grade 1s while others are more similar to grade 3s based on clinical behavior as well as genetic features such as somatic mutations, copy number variants⁵⁻⁷, methylation status⁸, and genome wide expression profiles^{9,10}. However, most research on gene expression in meningioma focuses on single gene analytics. This is not optimized for the low and additive molecular signals which frequently underlie complex and heterogeneous diseases. Systems biology approaches such as co-expression networks^{11,12}, on the other hand, are able to provide a higher resolution of these complex genetic processes^{11,13-15}. We therefore hypothesized that such methods may reveal subgroups for grade 2 meningiomas based on resemblance to grade 1 or grade 3 counterparts. In this study, we were able to deconvolve most atypical-grade meningiomas into a more indolent “grade 1-like” group and a more aggressive “grade 3-like” group, with

concordant recurrence rates. These findings help further clarify the heterogeneity of this challenging disease, and support a shift towards molecular classification of meningiomas in clinical practice.

The World Health Organization¹⁶ currently recognizes molecular subclasses for some of the most aggressive brain tumors^{17,18}, which have refined our predictions of treatment responses and clinical outcomes. Meningioma, the most prevalent type of adult brain tumor, are nevertheless categorized solely based on their histopathological appearances, a diagnostic heritage dating back almost eighty years¹⁹. Extremes of histological grades (mostly benign grade 1 and malignant grade 3) have relatively homogenous clinical behavior compared to grade 2 (or atypical) variants. There is a lot of uncertainty around the biology, clinical course and treatment response²⁰ of atypical meningiomas. This is reflected by regular revisions in WHO definitions²¹, overlapping molecular signatures with adjacent grades^{7,8} and open questions about the benefits of chemotherapy^{20,22} and adjuvant radiation²³⁻²⁵. Defining subgroups of atypical meningiomas with homogenous biological and clinical properties may be critical to successfully resolving these questions, thereby improving prognostication and treatment for patients.

Given the relatively consistent biological features of benign grade 1 and malignant grade 3 meningiomas, we used them as genetic “hallmarks” around which to “deconvolve” the heterogeneous atypical class into “grade 1-like” and “grade 3-like” subgroups. Rather than relying on single gene markers, we used co-expression networks, a well-established system-based approach to analyze the gene expression arrays of 212 meningiomas from 6 independent cohorts (Supplementary table S1). The novelty of this systems-based approach is its sensitivity to small and cumulative signals from interacting genes within a biological cascade. This technique has been successful at defining gene expression patterns behind complex phenotypes in Huntington’s disease¹⁴, peripheral nerve regeneration¹⁵ or weight

gain¹¹. Though to our knowledge it has not been used to decipher molecular characteristics of atypical meningiomas.

Results

We firstly established the gene expression profile that differentiates grade 1 from grade 3 meningiomas. Differential gene expression showed 1 up-regulated and 11 down-regulated genes (\log_2 fold change ≥ 1.5 , $p \leq 0.0001$) summarized in Figure 1A and Supplementary Table 2.

We created another signature to distinguish grade 1 from grade 3 meningiomas using gene co-expression networks. This yielded 20 co-expressed gene modules (Figure 1B), of which 13 had expression levels that differed significantly between grades 1 and 3 (Mann-Whitney $p < 0.05$, Figure 1C). A subset of these 13 were also significant between grades 1 and 2 and/or between grades 2 and 3 tumors, suggesting the intermediate biology of atypical meningiomas.

To find a genetic signature that best differentiates grades 1 and 3 tumors, we used two-centroid soft clustering and evaluated the resultant distribution of patients with a balanced sigmoidal cost function. An iterative feature selection approach was conducted using single genes and gene modules which were differentially expressed between grades 1 from 3. Notably, modules consistently yielded better performance (lower cost) than single genes (Figure 2A). The lowest cost was achieved with two modules as inputs; one of which contained 120 genes and is most associated with mRNA splicing while the other consisted of 98 genes and was involved in cell cycle. Using 80% membership probability as a cutoff, we reclassified 33 of 46 atypical meningiomas (72%) into a “grade 1-like” (14/46) and “grade 3-like” (19/46) subgroup of atypical meningiomas. (Figure 2B, Supplementary Figure 2). Recurrence rates were available for only a subset of cases (26/46) and were significantly higher in “grade 3-like” (8/11) compared to “grade 1-like” (0/9) subgroups ($p < 0.005$).

Concordantly, there was no difference in recurrence rates between grade 1 and “grade 1-like” groups nor between grade 3 and “grade 3-like” groups.

Next, we verified the molecular identity of the newly detected subgroups of atypical meningiomas. Using a systematic comparison based on median module expression levels (Figure 3), we found concordance between the biology of our newly identified atypical subtypes with either grade 1 or grade 3 meningiomas (Figure 3A). Differential analysis also suggested that the overall biological separation between the newly described subgroups is similar to the separation between grades 1 and 3. This further lends to the validity of dividing atypical meningiomas into biologically homogenous subgroups which parallel existing grades.

Discussion

The highly heterogeneous clinical behavior of atypical meningiomas suggests that histological criteria do not adequately capture its biology, thus motivating the discovery of molecular markers for better disease resolution. So far, molecular profiling of meningiomas has largely taken a monogenetic approach to marker discovery for aggressive phenotypes⁹. This has been fruitful in identifying recurrence mutations⁷ and transcripts⁹ linked to oncogenic cascades in meningiomas. These approaches however rely on differential gene expression to identify relevant molecular mechanisms and thereby remains limited in its ability to resolve small additive signal often relevant in tumor biology. Bypassing this problem, we applied gene co-expression networks to establish molecular signatures which identified indolent and aggressive subtypes of atypical meningiomas. Furthermore, majority of studies link genetic analysis to histological grade⁷, which does not capture disease biology for atypical meningiomas providing a 50-50 chance of recurrence. Epigenetic studies using conventional clustering have analyzed heterogeneity of meningiomas through all grades⁸ proposing new benign, intermediate and malignant methylation subclasses. “Intermediate” meningiomas are quoted a 20% chance of disease-free survival, more accurate than histology

(50%). Our study focuses on the most heterogeneous group, atypical meningiomas. We were able to separate subgroups with greater homogeneity compared to these preceding studies with 0% and 72,8% recurrence rates for grade 1-like and grade 3-like subgroups respectively. These findings also demonstrate the conceptual advantages of system-based approaches like co-expression networks over conventional techniques like differential gene expression and/or clustering. Our study has limitations: we analyze a single cohort, only a subset of samples have recurrence and follow-up times documented, on the other hand we provide a meta-analysis of 6 independent case-series which minimizes bias and counts towards the robustness of the findings.

Our findings help resolve the heterogeneity of atypical meningiomas by deconvolving into subgroups which are more homogenous than proposed prior studies. These homogenous subgroups may help predict clinical course, thus allowing for customized follow-up planning to manage resource intense investigations such as serial imaging while optimizing patient care. Our results could also help guide recruitment protocols for future and ongoing clinical trials, which are currently limited by the uncertainty of clinical outcomes in atypical meningiomas²⁴. The approach in this study lends to the utility of complex molecular signatures in augmenting histological diagnosis and resolving other heterogeneous and challenging diseases.

Materials and methods

Data collection and pre-processing

This study used open-source data from the repository *Gene Expression Omnibus (GEO)*²⁶. Six studies with primary meningioma transcriptomics were included in the analysis, all of which had WHO grade annotated (Supplementary Table 1). These publicly available gene expression datasets were accessed Gene Expression Omnibus (GEO) under id's: GSE10534, GSE77259, GSE54934, GSE43290, GSE16581 and GSE74385. For each study, the microarray data was backgrounded corrected, quantile normalized, and log-2 transformed using the *Affy*²⁷ and

*Limma*²⁸ R packages for Affymetrix and Illumina platforms, respectively. After removing genes that were not common across these studies, the 6 studies were merged, scaled to a global mean and standard deviation of 0 and 1, respectively²⁹, and batch-corrected using *ComBat*, a well-established empirical Bayes approach³⁰. The resultant data matrix was used during all subsequent analysis.

Differential gene expression analysis

Differential gene expression analysis was used to compare grades 1 and 3 meningiomas. In log₂-transformed space, the fold change (FC) was computed by subtracting the mean expressions of each gene in grade 1 tumors from the corresponding mean expressions in grade 3 tumors. Genes with absolute log₂-transformed FC ≥ 1.5 and $p \leq 0.0001$ were considered significant.

Co-expression networks and module detection

We used the well-established “Weighted Gene Correlation Network Analysis” (WGCNA) to detect “modules” of strongly co-expressed genes¹³. Per these previously described techniques, we first computed an “adjacency matrix” using soft-thresholded Pearson correlation between each gene pair. This was converted into a biologically-inspired topological overlap map (TOM), wherein pairwise gene similarities are derived from comparing their connectivity profiles³¹. Hierarchical clustering converted the TOM into a dendrogram, and a subsequent “dynamic” tree-cut³² served to identify gene modules. These modules were annotated with the *Database for Annotation, Visualization and Integrated Discovery* (DAVID)³³, an open-source bioinformatics resource. Additionally, representative module “meta-genes” for each sample were computed as the first principal component of their constituent genes’ expression values. The utility of this approach was verified in our dataset by demonstrating that higher principle components capture a very small proportion of the overall variance (Supplementary Figure 1A) and showing that neither study batch nor sex cluster along the first principle component (Supplementary Figure 1B and 1C). This eliminates the possibility of batch effect or sex being drivers of our

“meta-gene” values and confounding results. Differences in the expression levels of these “meta-gene” between grades was tested with a Mann-Whitney test, with a $p \leq 0.05$ considered significant.

Feature selection and clustering

In order to better understand the heterogeneity of grade 2 meningiomas, we began by identifying genetic features able to best distinguish grades 1 and 3. Fuzzy C-means (FCM) clustering was applied to the set of all patients in our study and the resultant separation of grades 1 and 3 was established with a sigmoidal cost function that is balanced for differences in the prevalence of both grades:

$$C = \frac{1}{N_1} \sum_{i \in S_1} \left(\frac{1}{1 + e^{-\alpha(P_i - 0.5)}} \right) + \frac{1}{N_3} \sum_{j \in S_3} \left(\frac{1}{1 + e^{-\alpha(0.5 - P_j)}} \right)$$

In the above equation, N_1 and N_3 are the number of grade 1 and grade 3 tumors, respectively; S_1 and S_3 are the sets of grade 1 and grade 3 tumors, respectively; P_k is the FCM clustering-derived probability of patient k being in the grade 3-enriched cluster; and α is a tunable hyperparameter. We used a two centroid model wherein cluster polarity was established by comparing the ratio of grade 3 to grade 1 tumours at both ends of the probability distribution (hard-thresholding at 80% probabilities).

Single genes and module “meta-genes” which were significantly different between grade 1 and grade 3 tumors served as input variables. Backwards elimination and forward selection were used for feature selection with model performance measured using the above cost function. Hyperparameter (α) values of 1, 5, 10, and 100 tested for all models. Once the separation of grades 1 and 3 was optimized, the probability distribution of grade 2 meningiomas within the same output was investigated. Atypical meningiomas with a probability $\geq 80\%$ of being in the grade 1-enriched cluster were defined as “grade 1-like”, and

those with a probability $\geq 80\%$ of being in the grade 3-enriched cluster were defined as “grade 3-like”.

Analysis of grade 2 subtypes

We first compared the recurrence rates of “grade 1-like” and “grade 3-like” meningiomas, and compared each to the rates of grade 1 and grade 3 tumors (Figure 2B, Supplementary Figure 2). Notably, only 114 of the 212 patients in our cohort have annotated recurrence. To investigate the degree of biological overlap between “grade 1-like” and grade 1 meningiomas, and similarly between “grade 3-like” and grade 3 meningiomas, we used the correlation between their module “meta-gene” expression levels (Figure 3A). In addition, we compared the biological separation between the newly described subtypes of atypical meningiomas to the separation of grades 1 and 3 by correlating their differential module expression levels (Figure 3B).

Data analysis platforms

All computational work relied on the open-source computational platform R³⁴, including packages *WGCNA*¹³, *ppclust*³⁵, *Affy*²⁷, *Limma*²⁸, and *SVA*³⁶.

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Conflict of interest:

Authors declare no conflict of interest.

References

- 1 Goldbrunner R, Minniti G, Preusser M, Jenkinson MD, Sallabanda K, Houdart E *et al*. EANO guidelines for the diagnosis and treatment of meningiomas. *Lancet Oncol* 2016;

- 17: e383–e391.
- 2 Rogers L, Barani I, Chamberlain M, Kaley TJ, McDermott M, Raizer J *et al.* Meningiomas: knowledge base, treatment outcomes, and uncertainties. A RANO review. *J Neurosurg* 2015; **122**: 4–23.
- 3 Sughrue ME, Kane AJ, Shangari G, Rutkowski MJ, McDermott MW, Berger MS *et al.* The relevance of Simpson Grade I and II resection in modern neurosurgical treatment of World Health Organization Grade I meningiomas. *J Neurosurg* 2010; **113**: 1029–1035.
- 4 Aghi MK, Carter BS, Cosgrove GR, Ojemann RG, Amin-Hanjani S, Martuza RL *et al.* Long-term recurrence rates of atypical meningiomas after gross total resection with or without postoperative adjuvant radiation. *Neurosurgery* 2009; **64**: 56–60.
- 5 Clark VE, Erson-Omay EZ, Serin A, Yin J, Cotney J, Özduman K *et al.* Genomic analysis of non-NF2 meningiomas reveals mutations in TRAF7, KLF4, AKT1, and SMO. *Science (80-)* 2013; **339**: 1077–1080.
- 6 Clark VE, Harmancl AS, Bai H, Youngblood MW, Lee TI, Baranoski JF *et al.* Recurrent somatic mutations in POLR2A define a distinct subset of meningiomas. *Nat Genet* 2016; **48**: 1253–1259.
- 7 Harmancl AS, Youngblood MW, Clark VE, CoÅ Kun S, Henegariu O, Duran D *et al.* Integrated genomic analyses of de novo pathways underlying atypical meningiomas. *Nat Commun* 2017; **8**. doi:10.1038/ncomms14433.
- 8 Sahm F, Schrimpf D, Stichel D, Jones DTW, Hielscher T, Schefzyk S *et al.* DNA methylation-based classification and grading system for meningioma: a multicentre, retrospective analysis. *Lancet Oncol* 2017; **18**: 682–694.
- 9 Vasudevan HN, Braunstein SE, Phillips JJ, Pekmezci M, Tomlin BA, Wu A *et al.* Comprehensive Molecular Profiling Identifies FOXM1 as a Key Transcription Factor for Meningioma Proliferation. *Cell Rep* 2018; **22**: 3672–3683.
- 10 Altshuler DM, Durbin RM, Abecasis GR, Bentley DR, Chakravarti A, Clark AG *et al.* An

- integrated map of genetic variation from 1,092 human genomes. *Nature* 2012; **491**: 56–65.
- 11 Ghazalpour A, Doss S, Zhang B, Wang S, Plaisier C, Castellanos R *et al.* Integrating genetic and network analysis to characterize genes related to mouse weight. *PLoS Genet* 2006; **2**: 1182–1192.
 - 12 Horvath S, Zhang B, Carlson M, Lu K V., Zhu S, Felciano RM *et al.* Analysis of oncogenic signaling networks in glioblastoma identifies ASPM as a molecular target. *Proc Natl Acad Sci* 2006; **103**: 17402–17407.
 - 13 Langfelder P, Horvath S. WGCNA: An R package for weighted correlation network analysis. *BMC Bioinformatics* 2008; **9**. doi:10.1186/1471-2105-9-559.
 - 14 Langfelder P, Cattle JP, Chatzopoulou D, Wang N, Gao F, Al-Ramahi I *et al.* Integrated genomics and proteomics define huntingtin CAG length-dependent networks in mice. *Nat Neurosci* 2016; **19**: 623–633.
 - 15 Chandran V, Coppola G, Nawabi H, Omura T, Versano R, Huebner EA *et al.* A Systems-Level Analysis of the Peripheral Nerve Intrinsic Axonal Growth Program. *Neuron* 2016; **89**: 956–970.
 - 16 Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK *et al.* The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. *Acta Neuropathol* 2016; **131**: 803–820.
 - 17 Thompson EM, Hielscher T, Bouffet E, Remke M, Luu B, Gururangan S *et al.* Prognostic value of medulloblastoma extent of resection after accounting for molecular subgroup: a retrospective integrated clinical and molecular analysis. *Lancet Oncol* 2016; **17**: 484–495.
 - 18 Yan H, Parsons DW, Jin G, McLendon R, Rasheed BA, Yuan W *et al.* *IDH1* and *IDH2* Mutations in Gliomas. *N Engl J Med* 2009. doi:10.1056/NEJMoa0808710.
 - 19 Cushing H, Eisenhardt L. *Meningiomas: Their Classification, Regional Behaviour, Life*

- History, and Surgical End Results*. Springfield, IL: Charles C. Thomas, 1938.
- 20 Moazzam AA, Wagle N, Zada G. Recent developments in chemotherapy for meningiomas: a review. *Neurosurg Focus* 2013; **35**: E18.
- 21 Pearson BE, Markert JM, Fisher WS, Guthrie BL, Fiveash JB, Palmer CA *et al*. Hitting a moving target: evolution of a treatment paradigm for atypical meningiomas amid changing diagnostic criteria. *Neurosurg Focus* 2008; **24**: E3.
- 22 Norden AD, Raizer JJ, Abrey LE, Lamborn KR, Lassman AB, Chang SM *et al*. Phase II trials of erlotinib or gefitinib in patients with recurrent meningioma. *J Neurooncol* 2010; **96**: 211–217.
- 23 Kaur G, Sayegh ET, Larson A, Bloch O, Madden M, Sun MZ *et al*. Adjuvant radiotherapy for atypical and malignant meningiomas: A systematic review. *Neuro. Oncol.* 2014; **16**: 628–636.
- 24 Jenkinson MD, Javadpour M, Haylock BJ, Young B, Gillard H, Vinten J *et al*. The ROAM/EORTC-1308 trial: Radiation versus Observation following surgical resection of Atypical Meningioma: Study protocol for a randomised controlled trial. *Trials* 2015; **16**. doi:10.1186/s13063-015-1040-3.
- 25 Jo K, Park HJ, Nam DH, Lee J Il, Kong DS, Park K *et al*. Treatment of atypical meningioma. *J Clin Neurosci* 2010; **17**: 1362–1366.
- 26 Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M *et al*. NCBI GEO: archive for functional genomics data sets — update. 2018; **41**: 991–995.
- 27 Gautier L, Cope L, Bolstad BM, Irizarry RA. affy — analysis of Affymetrix GeneChip data at the probe level. 2018; **20**: 307–315.
- 28 Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W *et al*. limma powers differential expression analyses for RNA-sequencing and microarray studies. 2015; **43**. doi:10.1093/nar/gkv007.
- 29 Hughey JJ, Butte AJ. Robust meta-analysis of gene expression using the elastic net.

- Nucleic Acids Res* 2015; **43**: 1–11.
- 30 Chen C, Grennan K, Badner J, Zhang D, Gershon E, Jin L *et al*. Removing Batch Effects in Analysis of Expression Microarray Data: An Evaluation of Six Batch Adjustment Methods. 2011; **6**. doi:10.1371/journal.pone.0017238.
- 31 Zhang B, Horvath S. A General Framework for Weighted Gene Co-expression Network Analysis. 2005.
- 32 Methods T, Clustering H, Author D, Langfelder P, Zhang B, Horvath S *et al*. Package ‘dynamicTreeCut’. 2016; : 1–14.
- 33 Huang DW, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 2009; **4**: 44–57.
- 34 R Development Core Team. R: A Language and Environment for Statistical Computing. *R Found Stat Comput Vienna Austria* 2016; **0**: {ISBN} 3-900051-07-0.
- 35 Package T, Probabilistic T, Analysis PC, Ce- C, C- PF, Algo- PC. Package ‘ppclust’. 2018 doi:10.1109/91.227387>.
- 36 Leek AJT, Johnson WE, Parker HS, Fertig EJ, Jaffe AE, Storey JD *et al*. Package ‘sva’. 2018.

Figure legends

Figure 1: Gene expression signatures associated with meningioma grade. A: Differential gene expression between grades 3 and 1 meningiomas reveal 1 upregulated and 11 downregulated genes in grade 3 tumours ($|\log_2(\text{fold change})| \geq 1.5$, $p < 0.0001$). B: Dendrogram of genes based on the topological overlap map, with the 20 gene modules represented as colors in the bar below. Grey represents unclassified genes. C: Plot of median module expression differences between grade (m) and grade (n). Only modules with significantly different expression between grades 1 and 3 are included (Mann Whitney $p < 0.05$). Red indicates modules which are upregulated in grade (m), and darker shades indicate larger effect sizes. Notably, 11/13

modules are significantly different between grades 1 and 2 while 2/13 are also significantly different between grades 2 and 3. * $p < 0.05$ (Mann-Whitney).

Figure 2: Optimized soft clustering reveals two subgroups of atypical meningiomas. A: Cost of multiple input configurations: "salmon" (S) and "lightcyan" (LC) modules in blue, "salmon" module alone and optimized differentially expressed genes in grey. Top inset depicts shape of sigmoid function with varied alphas. B: Summary graph of fuzzy C-means clustering best performing inputs (S + LC). The x-axis represents the probability of being in the grade-3 enriched cluster and y-axis represents the proportion of patients in each bin of 10%. Line graph component represents normalized frequency distribution of each histological grade (green = grade 1, black = grade 2, red = grade 3). Top jitter plot represents individual patients. Dark green and red bars above represent the 20 and 80% thresholding into grade 1-like and grade 3-like subgroups of atypical meningiomas. Recurrence rates are plotted on the right by grade (green, black, red) and subgroup ("grade 1-like" and "grade 3-like"). *Chi-square $p < 0.05$.

Figure 3: Molecular identity of newly described atypical meningioma subgroups. A: Scatter plot of median module expression. Larger circles indicate Mann Whitney $p < 0.05$. Colors correspond to previously identified modules in Figure 1B. ρ =Pearson coefficient, * $p < 0.05$. Note the positive correlation between the modules of grade 1 and "grade 1-like" and grade 3 and "grade 3-like" subtypes. B: Scatter plots of genetic separation between atypical subtypes as histological grades. The x-axis represents the difference in median module expression between grades 3 and 1, while the y axis represents the difference in median module expression between "grade 3-like" and "grade 1-like". Large circles represent modules which are significantly different in both comparisons and empty circles indicate modules which are not significantly different in either. Of the remainder, 5/6 are significantly different between

grades 3 and 1 only and 1/6 is significantly different between "grade 3-like" and "grade 1-like" (#).

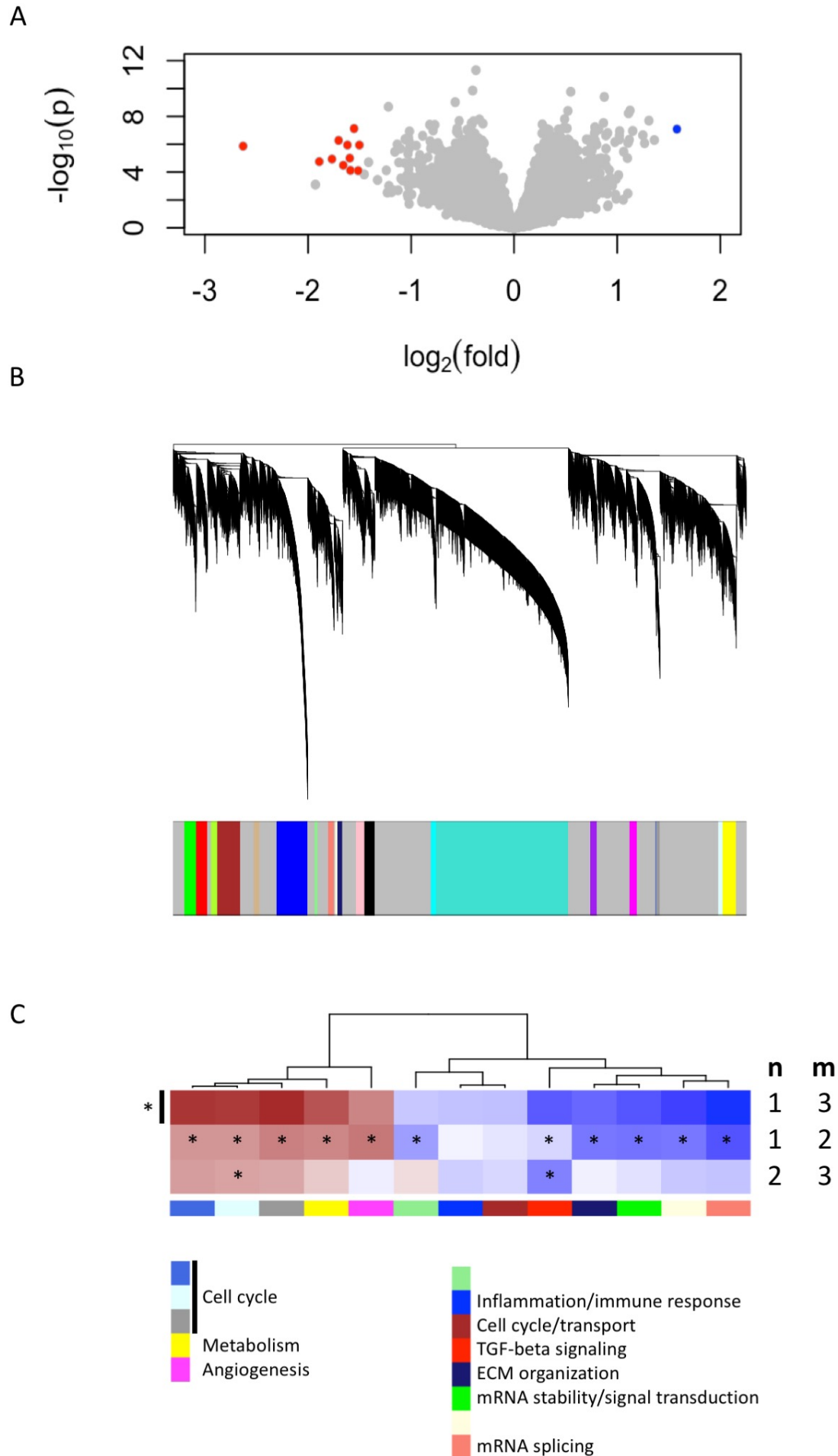
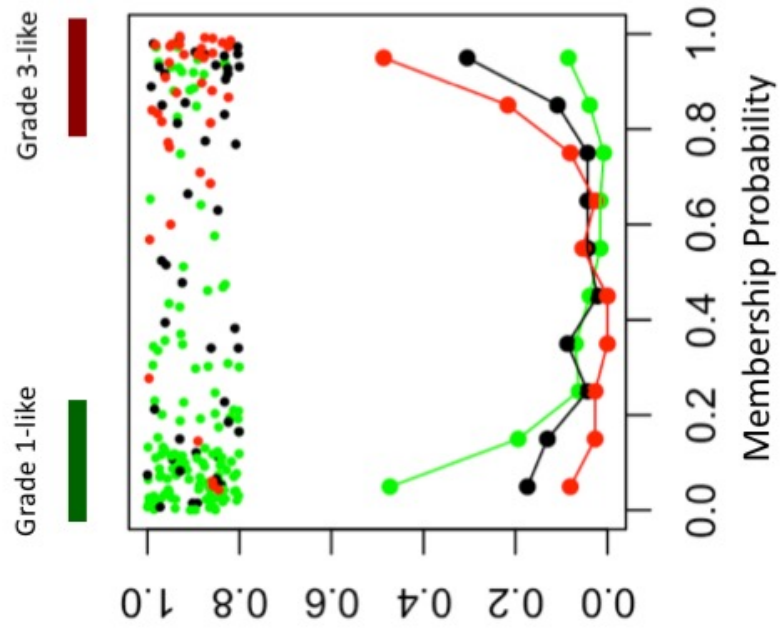


Figure 1

B



A

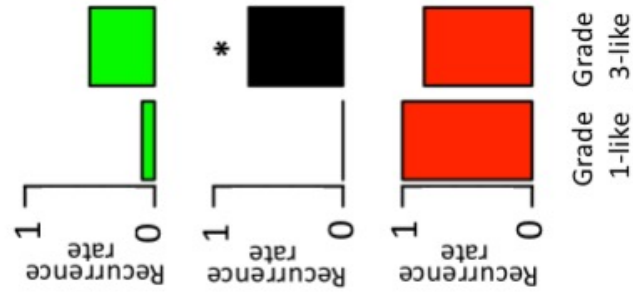
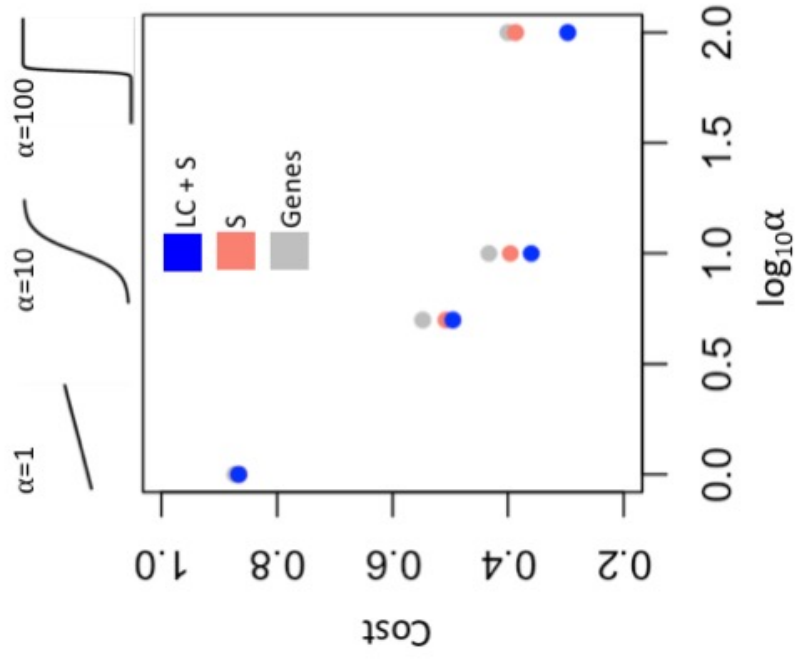


Figure 2

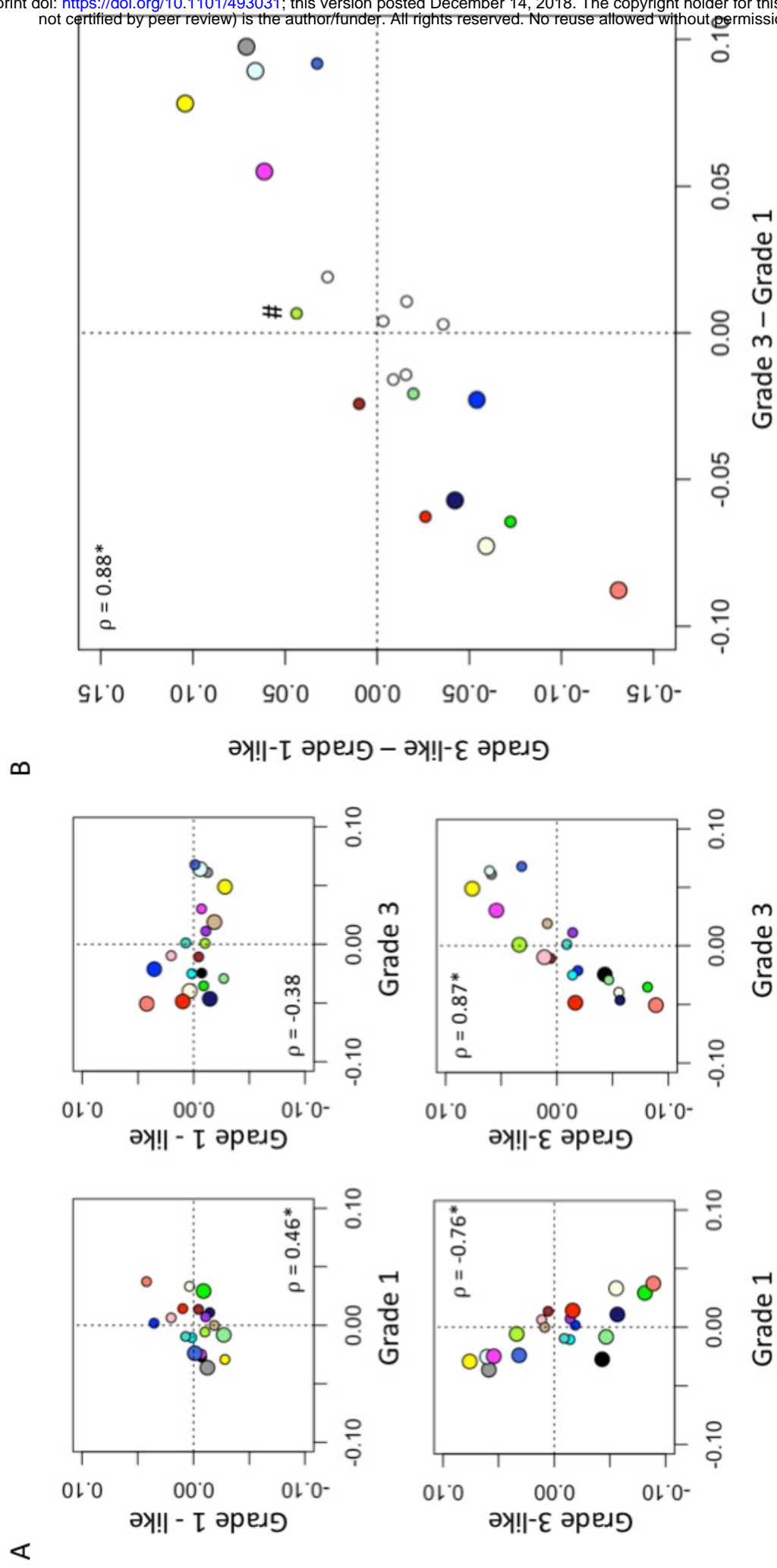


Figure 3