

1 **Rapid evolution of spontaneous mutation increases genetic diversity facilitating plant**
2 **population survival**

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7

8 **Abstract**

9 Using a mechanistic eco-evolutionary trait-based neighborhood-model, we quantify the impact
10 of mutations on spatial interactions to better understand the potential effect of niche evolution
11 through mutations on the population dynamics of *Arabidopsis thaliana*. We use 100 twenty-
12 fifth generation mutation accumulation (MA) lines (genotypes) derived from one founder
13 genotype to study mutational effects on neighbor responses in a field experiment. We created
14 individual-based maps (15,000 individuals), including phenotypic variation, to quantify
15 mutational effects within genotypes versus between genotypes on reproduction and survival.
16 At small-scale, survival is enhanced but reproduction is decreased when a genotype is
17 surrounded by different genotypes. At large-scale, seed set is facilitated by different genotypes
18 while the same genotype has either no effect or negative effects. Mutations may provide a
19 mechanism for plants to quickly evolve niches and may drive competition, facilitation and
20 selection with profound consequences for future population and community dynamics.

21 Introduction

22 A complete understanding of population and community dynamics requires linking intraspecific
23 genetic diversity with spatial ecological interactions (Bolnick et al. 2011, Genung et al. 2011).
24 Evolutionary mechanisms -- mutation, drift, gene flow and selection -- are responsible for
25 intraspecific genetic variation contributing to both ecological structure and species diversity
26 (Hart et al. 2016). These community properties impact ecological mechanisms such as
27 competition and facilitation (Whitlock 2014). Competition and facilitation in turn affect
28 population and community dynamics that depend on variation in demographic rates (Chase et
29 al 2002, Solivers et al 2015). Demographic rates are influenced by spatial interactions of genetic
30 variation and this variation may contribute to species coexistence: genotypes can hinder or
31 favor the survival of each other by using similar or different resources with negative or positive
32 consequences for coexisting species (Hart et al. 2016, Hausch et al. 2018). Intraspecific genetic
33 variation contributing to individual trait variation and environmental adaptability may promote
34 species coexistence by both increasing habitat heterogeneity and altering competitive
35 hierarchies (Violle et al. 2012, Ehlers et al. 2016). However, intraspecific genetic variation may
36 hinder species coexistence when intraspecific genetic variation is diminished by competition
37 between individuals (Hart et al 2016). To fully understand the causes of coexistence
38 mechanisms, we need to integrate intraspecific evolutionary mechanisms with intraspecific
39 ecological mechanisms underlying spatial species interactions (Bolnick et al. 2011, Ehlers et al.
40 2016).

41

42 The number of studies exploring eco-evolutionary dynamics and their potential
43 feedbacks on population and community dynamics has dramatically increased over the last two
44 decades (Shefferson and Salguero-Gomez 2015). An important current issue in community and
45 ecosystem genetics research is determining the relevance of intraspecific genetic variation and
46 genetic differentiation (divergence) to ecological and evolutionary processes at the community
47 and ecosystem level (Genung et al. 2011, Pujol et al. 2018). Quantifying intraspecific trait
48 variation defining the fundamental niches of species is an important link between ecology and
49 evolution (Violle and Jiang 2009). Spatial structure is another important component of realistic
50 eco-evolutionary dynamics, such that in spatially structured populations, selection is
51 determined by the interplay between demographic and genetic structures (Lion et al. 2011).
52 Demographic structure describes the spatial distribution of individuals through birth, death and
53 migration resulting in spatial patterns, while genetic structure describes the spatial distribution
54 of genotypes. The amount and spatial pattern of genetic variation may constrain evolution of
55 traits influencing competitive ability of individuals (Wilson 2014). Moreover, competitive
56 abilities depend on species' niches. The strength of competition is determined by how much
57 individual niches overlap with each other (Hutchinson 1957, Holt 2009) and how long
58 coexistence has occurred (Connell 1980).

59
60 Species niche breadths are reflected by individual trait variability and intraspecific and
61 interspecific genetic and spatial interactions driven by environmental conditions. Thus,
62 intraspecific trait variability shaped by intraspecific genetic variation can influence ecological
63 mechanisms driving variation among individual persistence (Lankau 2009). Only when the

64 spatial relationship between traits and demographic parameters is explicitly described can
65 robust hypotheses about the effects of individual variation on competitive outcomes be
66 accurately formulated (Hart et al. 2016). Especially in controlled common garden environments,
67 genetic variation in one species can have predictable and heritable effects on associated
68 communities and ecosystems (Carr and Dudash 1995; Whitham et al. 2003, 2006; Johnson &
69 Stinchcombe 2007; Bailey et al. 2009; Johnson, Vellend & Stinchcombe 2009). Hence, coupling
70 evolutionary genetics with community ecology may advance our understanding of species
71 interactions and population and community dynamics (Baron et al. 2016), especially for
72 populations and communities that suffer environmental change (Bellard et al. 2012).

73
74 An important gap in our understanding of the link between evolutionary and ecological
75 processes is the nearly complete absence of data quantifying how rapidly genetic variation
76 governing within species competitive hierarchies evolves (Hausch et al. 2018). An important
77 source of novel population genetic variation is mutation. Therefore, we aim to investigate the
78 link between evolutionary genetics and spatial ecological interactions of mutation accumulation
79 lines (MA lines) in the model plant organism *Arabidopsis thaliana* (Brassicaceae) to advance our
80 understanding of the genetic origins of population and community dynamics, contributing to a
81 fuller understanding of the maintenance of biodiversity. We take advantage of 25th generation
82 *A. thaliana* mutation accumulation (MA) lines that were planted under field conditions in years
83 2004 and 2005 with spatial records of each individual each year (e.g., Rutter et al. 2010; 2012,
84 Rutter et al. 2018) (Fig. 1).

85

86 Here we focus on the neighborhood effects of MA lines on focal plants from the same
87 MA line (within genotypes) or different MA lines (between genotypes) to quantify the evolution
88 of intraspecific genetic variation through the accumulation of spontaneous mutations. We
89 implement trait-based neighborhood models (Nottebrock et al. 2017a, Nottebrock et al. 2017b,
90 Lachmuth et al. 2017) to understand ecological mechanisms of within and between MA lines
91 genetic variation to examine eco-evolutionary dynamics (Lion 2018). Specifically, we ask: 1) Do
92 spontaneous mutations rapidly introduce enough genetic variation among MA lines to influence
93 competition and facilitation of *A. thaliana*? 2) Do spontaneous mutations of MA lines
94 contribute to competition, facilitation and selection promoting differential MA line survival and
95 reproduction at different spatial scales? 3) How may competitive and facilitative effects
96 potentially determine population and community dynamics?

97

98 Methods

99 MA lines are generated from a single nearly homozygous individual founder and cultivated via
100 limited effective population number. In the case of *A. thaliana*, this occurs through single seed
101 descent, resulting in $N_e = 1$. Thus typical MA line cultivation results in an unbiased sample of
102 mutation effects ranging from deleterious to advantageous, although lethal mutations are
103 excluded (Lynch and Walsh 1998). Each MA line accumulates independent spontaneous
104 mutations. After the propagation of a set of MA lines through multiple generations, the genetic
105 differences among the MA lines and between those lines and the founder reflect the input of
106 mutation. Significant MA line effects for multiple traits, including performance and trait
107 measures, were found under both field and greenhouse conditions (Rutter et al. 2010; Roles et
108 al. 2016, Rutter et al. 2018). Each of the MA lines in our experiment is fixed for an average of 20
109 different sequence level mutations, single nucleotide mutations (SNMs) and indels combined
110 (Ossowski et al 2010, Rutter et al. 2012, Weng et al. 2019).

111

112 *MA lines and field experiments*

113 We used survival and seed set data of *A. thaliana* MA lines and the founder as assessed in field
114 experiments in 2004 and 2005 from Rutter et al. (2010, 2012 and 2018) planted in a
115 randomized design (Fig. 1). Rutter et al. (2010, 2012) planted seedlings of 100 MA lines and the
116 founder at the four-leaf stage, approximately two weeks post germination, into a secondary
117 successional field at Blandy Experimental Farm (BEF) in Virginia (39°N, 78°W). Each of the 100
118 MA lines was used to found up to five sublines to minimize biases due to maternal effects

119 introduced by the specific location within the greenhouse. We founded six sublines from each
120 of the six lines representing the premutation founder genotype. In 2003, subline plants were
121 used to generate all seed utilized in all field experiments. In each planting, 7504 individuals
122 were planted, 7000 individuals of 100 MA lines (70 replicates per MA line, 14 replicates per
123 subline) and 504 individuals of the founder (14 replicates per subline). The planting
124 environment corresponds to a spring ephemeral life-history, where plants germinate and
125 complete the life-cycle in the spring. At the time of planting, vegetation was scant but present.
126 By harvest, the *A. thaliana* individuals were dwarfed by naturally occurring vegetation.

127

128 The plot was arranged in 14 spatial blocks with each containing 12 subblocks (Fig. 1B)
129 (total plot area approximately 35 x 25 m). Each block included one seedling from each subline
130 and in total 7504 individuals. We used the spatial information of each individual within the
131 described design and created a raster of all plant individuals with R packages (raster, maps,
132 maptools). We used individual-based maps neighborhood matrices with exact spatial and trait
133 information of each genotype. If all five sublines did not produce enough seedlings to distribute
134 in all blocks, seedlings from other sublines of the same MA line were overrepresented in blocks
135 to maintain the same overall number of plants per MA line. Plants dying within the first 3 days
136 of transplant (about 50 plants) were considered to have died from transplant shock and were
137 replaced with another plant from the same MA line. Plants were censused weekly for survival.
138 Plants were harvested by late May, by which time they had senesced. In 2004 a total of 5915
139 individuals including 394 founders survived. In 2005, a total of 4506 individuals survived

140 including 302 founders. Plants were oven dried and biomass was measured. All fruits produced
141 by each plant were counted and in combination with seed production used as the
142 measurements of seed set. For our analysis, we used measurements of two response variables
143 representing an important part of the life history of *A. thaliana* to calculate direct
144 neighborhood effects on: a) the survival rate of plant individuals and b) individual seed set from
145 plants that survived and produced fruits.

146

147 *Statistical Analyses*

148 We use eco-evolutionary trait-based neighborhood models that include intraspecific genetic
149 variation and phenotypic variation expressed by plant trait biomass to analyze competition and
150 facilitation between individuals of *A. thaliana*. We analyzed plant survival rate and focal seed
151 set measurements from years 2004 and 2005 separately. For each year, we considered all
152 individuals as focal plants in the analysis based on individual-based neighborhood matrices to
153 analyze 1) rate of plant survival to reproduction and 2) seed set of those plants that survived to
154 reproduction. We analyzed neighboring plants to focal plants in a radius of 80 cm (small-scale)
155 or 200 cm (large-scale) of a given focal plant as two spatial scales in the neighborhood analyses
156 (Fig. 1) to quantify selection, competition or facilitation between plants depending on their
157 genotypic and phenotypic variation.

158

159 We used extensions of linear mixed models (package lme4, Bates et al. 2014) in R ver.
160 3.3.3 (www.r-project.org) to conduct neighborhood analyses of focal seed set and survival. We
161 assumed binomial errors for the analyses of plant survival and Poisson errors for analyses of
162 seed set. The mixed models described interactions among plants by including neighborhood
163 indices as explanatory variables at two spatial scales in separate models. Neighborhood indices
164 are spatial density effects of surrounding neighborhood plants that affect focal seed set and
165 survival. For each plant, we used the Euclidian distance between the focal plant and the
166 neighboring plants to compute response effects of intra- (same genotype) or inter-genotypic
167 (different genotype) neighbors in a given radius around focal plants (Nottebrock et al. 2017).
168 Moreover, we used a neighborhood index that accounts for the decline of neighbor effects with

169 distance from the focal plant (Uriarte et al. 2010) and summed the amount of biomass from all
170 individuals in a radius of 80 cm or 200 cm respectively by a Gaussian interaction kernel
171 (Lachmuth et al. 2018, Nottebrock et al. 2017, Damgaard 2004). We used random effects of
172 block and subblock to correct for environmental variation between local heterogeneous
173 conditions. Importantly, we correct for between MA line effects by including MA lines as a
174 random effect. In addition, including a random slope of biomass on each random intercept
175 corrects for the intraspecific phenotypic variation depending on local conditions of plant focal
176 individuals. Moreover, the weighted neighborhood density by plant biomass accounts for
177 environmental variation between neighboring plants. Our model parameters and a detailed
178 model description of subblock and block models can be found in S1 supplementary text
179 (Supplementary Material).

180

181 Neighborhood matrices of all individuals (individual-based maps) were used to analyze
182 the effect of intra- and inter-genotypic neighbors on survival and focal seed set with spatial
183 interaction kernels of neighborhood (plant biomass) density. By incorporating different
184 genotypes and phenotypic variation, we can quantify how important genetic variation is for
185 neighborhood models and if the phenotypic variation explains spatial interactions between
186 individuals. We assume the consequences of genetic differences to be larger between MA lines
187 than between any MA line with the founder. This is a valid assumption since each MA line
188 differs from the other by approximately $20 + 20 = 40$ mutations, while any two replicates within
189 a MA line will differ by one generation, ≤ 2 mutations. Thus, we simulated line effects from

190 parameters derived from MA lines as random effects with the R package ‘merTools’ and the
191 function ‘plotREsim’ in R 2018.

192

193 We used the trait values of neighbors to calculate trait-based neighborhood indices
194 including plant biomass as a trait (Goldberg & Fleetwood, 1987; Goldberg & Landa, 1991; Cahill
195 et al., 2005). We fitted eco-evolutionary trait-based neighborhood models at two different
196 spatial scales for response variables (survival and seed set) for each of the two and both years.
197 To address our objectives, we first analyzed models with differential effect in which intra-
198 genotypic neighbors (within all MA lines and founder) had a different effect on survival (A1, A2,
199 Table 1) and seed set (B1, B2, Table1) than inter-genotypic neighbors at small-scale (s) and
200 large-scale (l). In addition, we analyzed models with neutral effects on survival (A1, A2, Table 1)
201 and seed set (B1, B2, Table 1) that included total neighbor density without the split between
202 intra-genotypic and inter-genotypic neighbors at small-scale (s) and large-scale (l). To this end,
203 all models were fitted with two separate neighborhood indices that were calculated from intra-
204 and inter-genotypic neighbors. To justify the inclusion of individual plant biomass as trait-values
205 for interacting plants in the model, we used AICc to compare the models with and without the
206 trait-proxy (Burnham and Anderson 2002). We found that all models perform better including
207 biomass as a trait-proxy ($\Delta AICc > 2$). All eco-evolutionary trait-based neighborhood models
208 contained random effects of subblock nested in block at block scale and subblock scale on the
209 intercept, MA line identity on the intercept and the focal trait-value (plant biomass) on the
210 slope. Additionally, because direct environmental variables were not measured during the field
211 experiments, we included in each model the individual’s biomass to correct for environmental

212 conditions for spatial autocorrelation. All variables are scaled and centered to assure
213 comparability between predictor variables. Models of differential and neutral effects for 2004
214 and 2005 (Table 1, A1-A2, B1-B2) are fitted at small-scale (80cm scale) and at large-scale
215 (200cm scale). Hereafter, the 80 cm scale models are referred to as “small-scale” models and
216 the 200 cm scale models are referred to as “large scale” models. Neighborhood indices, intra-
217 and intergenotypic variation and total variation of biomass density are included as inverse
218 density variables ($1/1+\text{density}$). We compared models of differential and neutral effects
219 through likelihood ratio tests (LRTs).

220

221 Results

222 Focal plants surrounded by plants of the same genotypes varied from 0 to 6 individuals at small-
223 scale (sub-block) and 5 to 6 individuals at large-scale (block). After determining that eco-
224 evolutionary trait-based models perform better when including plant biomass as a trait proxy to
225 calculate trait-based neighborhood indices we found that all models including plant biomass
226 perform better than models including only neighborhood density ($\Delta AIC > 2$, Supplementary
227 Material S2).

228

229 *Weighted neighbor effects of biomass density on genotypes for plant survival*

230 At small-scale, survival rate is larger when surrounded by inter-genotypic than intra-genotypic
231 neighbors for both years 2004 and 2005 (model A1 and A2; Table 2 and Fig. 2a). This difference
232 of survival rate between intra- and inter-genotypic plants indicates that inter-genotypic
233 neighbors select for genotypic diversity and show stronger competitive effects on the survival
234 of intra-genotypic neighbors at small-scale. This finding is demonstrated by the superior
235 performance of the differential model relative to the neutral model (year 2004: LRT: $\chi^2_{1DF} =$
236 5.21, $p < 0.05$; year 2005: LRT: $\chi^2_{1DF} = 14.73$, $p < 0.001$). The differential and neutral models of
237 survival at large-scale have only non-significant effects (Table 2). In addition, comparing the AIC
238 we found that models at large-scale perform worse than at small-scale for both 2004 and 2005
239 (Table 2). We therefore will only discuss small-scale effects on survival. The distance kernel
240 (α) at small-scale shows that neighboring plants of different genotypes reduce plant
241 survival by 50% at 53 cm in 2004 and 48 cm in 2005.

242 *Weighted neighbor effects of biomass density on genotypes for seed set*

243 At both small and large scales, the differential model performs better than the neutral model:
244 at small scales (year 2004: LRT: $\chi^2_{1DF} = 52.48$, $p < 0.001$; year 2005: LRT: $\chi^2_{1DF} = 17.37$, $p < 0.001$;
245 respectively) and at large scale (year 2004; LRT: $\chi^2_{1DF} = 679.44$, $p < 0.001$; year 2005: LRT: $\chi^2_{1DF} =$
246 21.17, $p < 0.001$; respectively). The small-scale model of seed set that included weighted
247 biomass densities demonstrates the reduction of seed set is larger when surrounded by inter-
248 genotypic than intra-genotypic neighbors for both years 2004 and 2005 (model B1 and B2;
249 Table 2 and Fig. 2a). The difference between intra- and inter-genotypic plants indicates that
250 inter-genotypic neighbors have a stronger competitive effect on seed set than intra-genotypic
251 neighbors at small-scale. In contrast, at large-scale seed set increases when surrounded by
252 inter-genotypic neighbors but decreases when surrounded by intra-genotypic neighbors for
253 both years 2004 and 2005 (model B1 and B2; Table 2 and Fig. 2b). Comparing the AIC between
254 small and large-scales, models at large-scale for 2004 and 2005 perform better (Table 2).

255

256 *Genotype effects on plant survival and seed set*

257 The distance kernel at small-scale in 2004 indicates a reduction of seed set (competition)
258 whereas at large-scale are consistent with an increase of seed set (facilitation). Neighboring
259 plants from all genotypes of *A. thaliana* at small-scale reduce 50% of focal seed set at 23 cm
260 and at large-scale facilitate 50% of focal seed set at 86 cm. The estimation of genotype effects
261 (MA lines and founder) simulated as conditional means and expressed as odds ratios show
262 hierarchical orders of MA line and founder competitive effects at small-scale (Fig. 3). We found

263 no significance of simulated competitive effects of all MA lines and founder calculated from
264 survival and seed set models at small-scale (ANOVA, $p > 0.1$, Fig. 4a). In addition, we found no
265 significant correlation between competitive effects of MA lines and founder at large-scale. In
266 addition, we did not find selective effects of MA lines and founder on the survival at small-scale
267 (ANOVA, $p > 0.1$, Fig. 4b). However, we found strong correlation between competitive effects
268 of MA lines and founder at small-scale and large-scale indicating a genetic trade-off (ANOVA,
269 $F=460.07$, $P < 0.0001$, Fig. 4c).

270

271 Discussion

272 We are able to quantify individual spatial interactions between and within genotypes (MA lines
273 and founder) using a mechanistic eco-evolutionary trait-based neighborhood model. We
274 demonstrate intraspecific genetic variation due to spontaneous mutations can shape
275 competitive abilities of genetically different individuals of *A. thaliana*. Notably, these
276 differences arose in just 25 generations of mutation and in the absence of natural selection.
277 Many of the mutations differentiating the MA lines also appear in nature, contributing to *A.*
278 *thaliana* genetic polymorphism (Weng et al., 2019). Plant survival is higher when surrounded by
279 inter-genotypic neighbors supporting genetic diversity. Effects on focal seed sets were reversed
280 between small and large scales; inter-genotypic neighbors have stronger negative (competitive)
281 or positive (facilitative) effects than intra-genotypic neighbors on focal seed sets at small-scale
282 or large-scale, respectively. Moreover, at small-scale, competitive effects of different MA lines
283 have similar impacts on survival and seed set. At large-scale, the intra-genotypic effect on
284 survival turns into less competitive effects on seed set. In contrast, different genotypes show
285 stronger facilitative effects within the population at large-scale, maybe because plants are more
286 competitive against other species. Below we discuss how these scale effects provide insight on
287 community assembly and potential coexistence.

288

289 *Selection, Competition and Facilitation*

290 Because of stronger statistical signals of effect sizes between genotypes than within genotypes,
291 niche variation may result in niche expansion or reduction (Ehlers et al. 2016). However, these

292 competitive effects only occur between plants that survive to reproduction and affect seed-set.
293 The neighbor effect on plant survival suggests kin recognition despite our randomized field
294 experiment that likely reduced the spatial autocorrelation of genotypes (Hamilton 1964,
295 Kubisch et al. 2013). Seed-dispersal is limited in space because the *A. thaliana* fruit do not
296 dehiscently explode as they do in related species (Hofhuis and Hay 2017), although a seed bank
297 ensures some dispersal through time (Falahati-Anbaren et al. 2014). The neighbor effect
298 increasing genetic diversity of *A. thaliana* populations can have a positive effect on the
299 coexistence of competing species (Vellend 2006). If we assume that the increase of genetic
300 diversity due to spontaneous mutations of *A. thaliana* affects the extent of individual plant
301 niches, at large-scale the inclusion of other plant species should result in stronger competitive
302 effects of *A. thaliana* with other plant species (Hausch et al. 2018). We demonstrate that *A.*
303 *thaliana* individuals increase their competitive abilities with an increase of genetic diversity,
304 perhaps due to the expansion of their individual niches due to more variation in plant traits.

305

306 Antagonistic interactions between genotypes on different spatial scales may result in
307 genetic trade-offs when mutations are advantageous or deleterious, which over many
308 generations could provide a strong stabilizing force maintaining both species and genetic
309 diversity in this system and promote coexistence (Lankau 2008). Thus, spontaneous mutation
310 could also provide an additional evolutionarily stabilizing effect on community dynamics. Our
311 study demonstrates the potential for a fundamental evolutionary process, mutation, to have
312 profound consequences for community structure. When *A. thaliana* is rare, selection would
313 favor genotypes that compete well and enhance the population's survival relative to that of its

314 interspecific competitor (Lankau 2009). In contrast, when *A. thaliana* is common and the
315 interspecific competitor is rare, selection would favor *A. thaliana* genotypes that are good
316 intraspecific competitors. This trade-off may result in a decrease in the interspecific competitive
317 ability of *A. thaliana*, effectively increasing the competitor's fitness relative to that of *A.*
318 *thaliana* (Lankau 2009, Lankau and Strauss 2007). However, the genetic trade-off between
319 intra-and interspecific competitive abilities due to mutation remains unknown because our
320 dataset only includes performance of *A. thaliana* individuals without quantifying the
321 surrounding intraspecific environment (Chesson 2000, Adler et al. 2007).

322

323 *Competitive Hierarchies of MA lines and Founder lines*

324 Our result show scale dependent competition and facilitation (Nottebrock et al. 2017b)
325 providing further evidence that in a relative short evolutionary time scale spontaneous
326 mutations may change the competitive hierarchies between founders and specific MA lines due
327 to advantageous or deleterious mutations (Rutter et al. 2010). In contrast to Masclaux et al.
328 2010, we found differential responses to similar genotypes vs different genotypes in *A. thaliana*
329 depending on the neighbor effect on the plant survival or seed set. However, the interaction
330 between similar genotypes and different genotypes seems to depend on the strength of the
331 competitive abilities of the accessions (this study). Comparisons between MA lines and founder
332 demonstrate that competitive hierarchies follow different orders and competitive effects occur
333 at different life-history stages at different spatial scales. Eco-evolutionary processes might
334 reflect spatial selection for diversified genotypes because of niche evolution and individuals of
335 different genotypes that survived to reproduction have stronger competitive abilities (Ehlers et

336 al. 2016). Moreover, the hierarchical order of competitive effects between MA lines and
337 founders shows strong variation.

338

339 *Eco-evolutionary dynamics*

340 Populations can adapt evolutionarily to their environment on a time scale equivalent to that of
341 ecological processes and affect present day species interactions, i.e., coexistence (Fussmann et
342 al. 2007). Yet, sustained and rapid climate change could deplete genetic variance faster than it
343 can be replenished by mutation (Fournier-Level et al. 2016). In our study, we show that only 25
344 generations are enough to influence present day plant interactions of *A. thaliana* in the
345 presence of other plant species. Possible effects of mutations on trait mechanisms might reflect
346 the increased seed set of *A. thaliana* at large-scale, because of the presence of another plant
347 species allowing the establishment and growth of *A. thaliana* (Ehlers et al. 2016). Many traits
348 can influence competition and facilitation between individuals of different genotypes (e.g.
349 Chapin et al. 1993; Caradus and Woodfield 1998; Hausmann et al. 2005). Intraspecific
350 competition between *A. thaliana* individuals tends to be higher than between interspecific
351 competitors (Lankau and Strauss 2007, Van Dam and Baldwin 1998; Tiffin 2002, Strauss and
352 Irwin 2004). Such spatial mechanisms might be described by the scale difference of competition
353 at small-scale and facilitation at large-scale between different *A. thaliana* genotypes.
354 Competition and facilitation are therefore dependent on the extension or reduction of
355 individual niches, which is based on the ability to advance or distract traits due to spontaneous
356 mutations. However, we are not able to directly link our results to benefactor or antagonistic
357 plants, because we focused in our experiment on *A. thaliana* densities and fitness components.

358 Nevertheless, genotypic-specific interactions are well studied. A number of studies of *A.*
359 *thaliana* MA lines demonstrate mutation effects on plant traits such as leaf weight, flowering
360 time, trichome density, number of leaves at bolting, duration of vegetative and flowering
361 period (Camara and Pigliucci 1999, Rutter et al. 2010, Stearns and Fenster 2016). It is
362 conceivable that the increase in variance of these traits through spontaneous mutations could
363 contribute to the evolution of competitive hierarchies.

364

365 *Consequences for population and community dynamics*

366 Competition between genetically different individuals of *A. thaliana* at small-scale and
367 facilitation at large-scale has important implications for population structure (Cahill et al. 2005).
368 Plants may cooperate by competing less, or act selfishly by competing more (Dudley et al.
369 2013). However, kin recognition might be the result of kin selection when occurring in a
370 heterogeneous environment with various genotypes (Hamilton 1964). Especially, at large-scale,
371 we show that different genotypes facilitate seed set that stabilize the population dynamics by
372 increasing the performance of neighboring plant individuals (Latzel et al. 2013, Castellanos et al.
373 2014). Thus, selection for diversified genotypes might reveal a coexistence mechanism, because
374 mutations at a short time scale (25 generations) are able to alter the competitive hierarchies of
375 *A. thaliana* individuals of different genotypes. These genotypes might expand or reduce their
376 niches and can therefore act more competitive against strangers (Ehlers et al 2016). Moreover,
377 variation in competitive ability among genotypes due to mutation can lead to intransitive
378 competitive hierarchies at a small-scale, and allow coexistence of competitors at large-scale
379 when there is no single dominant competitor (Vellend and Geber 2005, Taylor and Aarssen

380 1990, Laure et al. 2017). This might also reflect the genetic trade-off between competitive
381 abilities of MA lines of small-scale competition and of large-scale facilitation (Timan 2004). Low
382 competition between *A. thaliana* individuals at small-scale transmits into low facilitation
383 between different species at larger scale. In turn, strong competition within *A. thaliana* at
384 small-scale results in weaker competition between different species at large-scale. Thus, the
385 constant input of mutation to genetic variation of competitive hierarchies suggests that genetic
386 variation is likely never a limiting factor in the evolution of those traits that influence plant
387 community dynamics.

388

389 *Implementation of eco-evolutionary trait-based model for population and community ecology*

390 In ecology, two primary contrasting models are crucial to population and community dynamics
391 for understanding the maintenance of biodiversity: the niche and neutral theories of ecology
392 (Fisher and Mehta 2015). The niche theory claims that niches are determined by the difference
393 of abiotic or biotic ecological processes e.g. competition or facilitation (Hutchinson 1957, Bruno
394 et al. 2003, Colwell and Rangel 2009) whereas the neutral theory predicts that ecological
395 processes are neutral and are solely modified by stochasticity (Hubbel 2005). Although it is still
396 under debate which model predicts the dynamic properties of communities and determines the
397 maintenance of biodiversity (Violle et al. 2017), both models miss the importance of
398 evolutionary mechanisms and individual variation influencing population and community
399 dynamics (Violle et al. 2012). The difference of competition and facilitation between or within
400 genotypes demonstrates that evolutionary mechanisms that shape individual variation could

401 change hierarchies of intra-specific interactions in natural plant populations over a short
402 evolutionary time scale. Our results are concordant with the niche theory stressing the
403 changing of competitive hierarchies among phenotypes in a given environment (Levine and
404 HilleRisLambers 2009, Nottebrock et al. 2017). Moreover, selection and trait evolution favor
405 population survival of *A. thaliana* in the wild, which stresses the importance of combining
406 spatial ecological and evolutionary mechanisms for our understanding of population and
407 community dynamics.

408

409 Conclusion

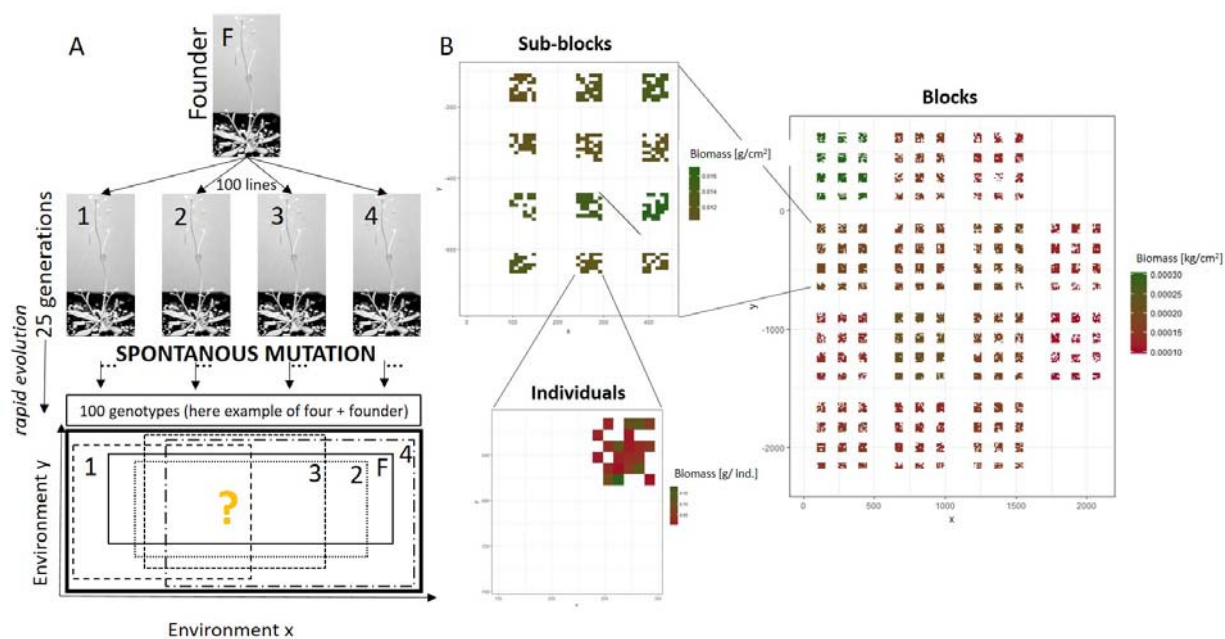
410 Rapid evolution of *A. thaliana* due to spontaneous mutation alone has profound consequences
411 for population and community dynamics. Competition of *A. thaliana* individuals is genotype-
412 dependent and *A. thaliana* is not a single weak competitor among co-occurring plant species
413 (Ehlers et al. 2016, Soliveres et al 2017). Moreover, the discovery of the underlying eco-
414 evolutionary nature of competition in *A. thaliana* supports a shift from species-based to
415 individual-based community ecology. This would lead to a more predictive ecological theory
416 (Violle et al 2017). We show that including the intraspecific genetic variation and phenotypic
417 variation as separate aspects to a trait-based neighborhood model increases the predictive
418 power to understand population and community dynamics. In particular, non-neutral
419 intraspecific processes may determine species coexistence, because genetic diversity is
420 promoted by having stronger competitive abilities at small-scale and stabilizing population
421 survival at large-scale (Clark et al. 2010). Additionally, our study demonstrates higher genetic
422 diversity increases population survival due to rapid evolution with implications to forecast the
423 fate of species and functional diversity in response to environmental changes (Violle et al 2012).
424 Depending on the time of environmental changes, species may adapt to environmental change
425 by shifting their fundamental niches (Clark 2010). The combination of intraspecific genotypic
426 variation and spatial interactions might advance our understanding of community dynamics,
427 especially of rapid evolution (Koch et al. 2014, Turcotte and Levine 2016). Incorporating genetic
428 variation and the eco-evolutionary process for determining standing levels of genetic variation
429 will provide a better understanding of species interactions underlying the maintenance of
430 biodiversity.

431

432 Keywords (10 for indexing purpose): trait-based community ecology, spontaneous mutations,
433 competition, facilitation, niche evolution, intraspecific genetic variation, intraspecific
434 phenotypic variation, *Arabidopsis thaliana*, mutation accumulation lines, eco-evolutionary
435 dynamics.

436

437 Figures and Tables



438
439 Figure 1. (A) 100 Mutation accumulation lines (here example of four MA-lines) of *Arabidopsis*
440 *thaliana* derived from a single founder (Columbia) after 25 generations with a mutation rate of
441 ~ 0.7 per generation (Weng et al. 2019). Each MA line has a unique genotype that differs from
442 the founder. Fundamental niche of individual genotypes (different dashed and dotted lines) and
443 founder genotype (black line) are distributed within the hypothetical fundamental niche of a
444 population of *A. thaliana* (bold black line). Axes represent niche axis (resource-use,
445 environmental factors, etc.). (B) Planting design of *A. thaliana* in a natural field experiment
446 showing spatial biomass variation of individual survivors in 2004.

447

	Year 2004	Year 2005	Model
Individual survival	A1 (s,l)	A2 (s,l)	Differential
	A1 (s,l)	A2 (s,l)	Neutral
Seed set	B1 (s,l)	B2 (s,l)	Differential
	B1 (s,l)	B2 (s,l)	Neutral

448

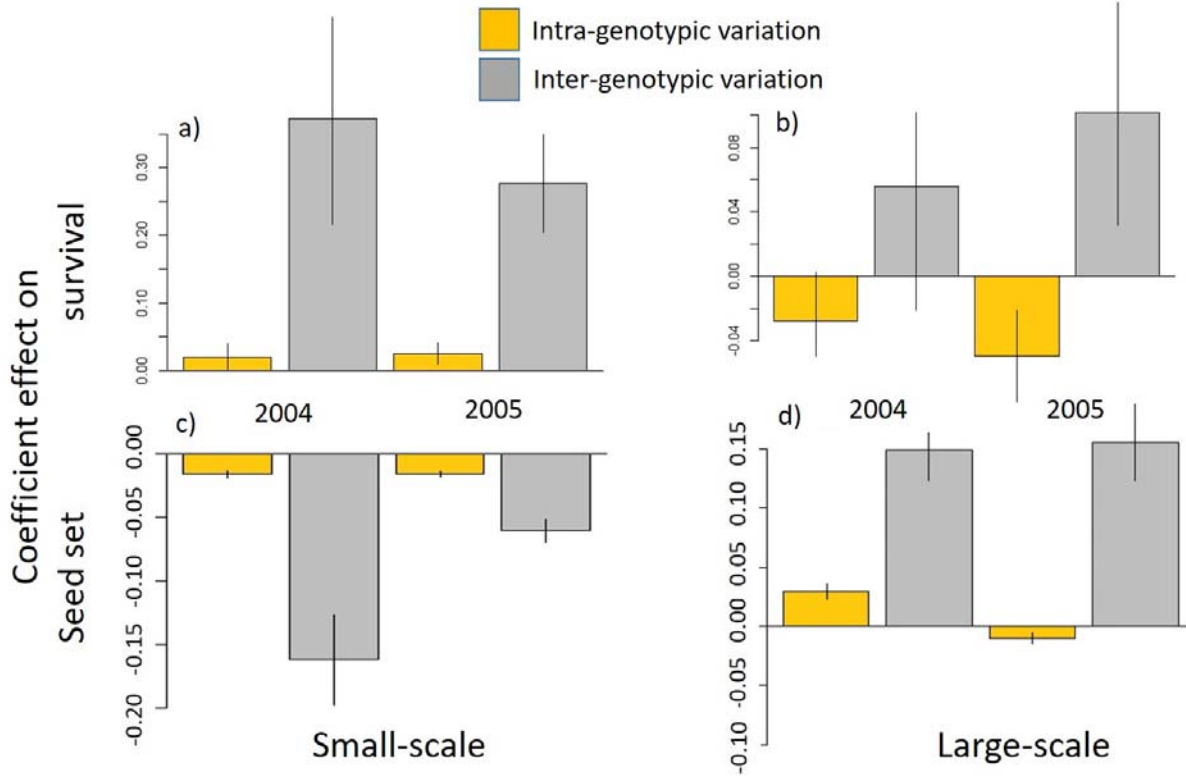
449 Table 1. Table of all different eco-evolutionary trait-based neighborhood models including
450 intraspecific variation to test our three objectives of density effects on survival and seed set of
451 *A. thaliana*. Differential models split the neighbor responses into within genotypes and
452 between genotypes. Neutral models describe the overall neighbor response without
453 differentiating between genotypes. All models include random effects of plant biomass on the
454 slope, subblock or block and MA-line on the intercept (see Fig. 1 for design), and respectively
455 depending on small (s) or large-scale (l) (for more details see method section).

Survival model							
(A1) 2004		(s) Small scale (80cm)		AIC	(l) Larger scale (200cm)		AIC
		<i>Intra</i>	<i>Inter</i>		<i>Intra</i>	<i>Inter</i>	
Weighted density							
MA	neutral	-0.02 ^{ns}		7551.7	0.004 ^{ns}		7644.5
Lines	differential	-0.02 ^{ns}	-0.37**	7548.6	0.15 ^{ns}	-0.30 ^{ns}	7644.2
(A2) 2005							
		(s) Small scale (80cm)		AIC	(l) Larger scale (200cm)		AIC
		<i>Intra</i>	<i>Inter</i>		<i>Intra</i>	<i>Inter</i>	
Weighted density							
MA	neutral	0.03 ^{ns}		9637.3	0.03 ^{ns}		9872.9
lines	differential	-0.02 ^{ns}	-0.28***	9626.1	0.05 ^{ns}	-0.10 ^{ns}	9873.3
Seed set model							
(B1) 2004		(s) Small scale (80cm)		AIC	(l) Larger scale (200cm)		AIC
		<i>Intra</i>	<i>Inter</i>		<i>Intra</i>	<i>Inter</i>	
Weighted density							
MA	neutral	-0.02***		52982	0.03		52944
lines	differential	-0.02**	-0.07***	52948	0.03***	0.15***	52939

(B2) 2005		(s) Small scale (80cm)		AIC	(l) Larger scale (200cm)		AIC
		<i>Intra</i>	<i>Inter</i>		<i>Intra</i>	<i>Inter</i>	
Weighted density							
MA	neutral	-0.008*		49103	-0.008*		49049
lines	differential	-0.009**	-0.09***	49095	-0.01*	0.16***	49026

456

457 Table 2. The coefficient and AIC from eco-evolutionary trait-based neighborhood models of the
458 *Arabidopsis thaliana* field experiment in the years 2004 and 2005. Models include intraspecific
459 genetic variation defined as intra-genotypic (within genotypes) and inter-genotypic (between
460 genotypes) variation. Models are shown at two different spatial scales: weighted subblock
461 density (80cm radius) and weighted block density (200cm radius). All scales include variables of
462 neutral effects (absence of intra- and inter-genotypic variation (total sum of all neighboring
463 genotypes)) and differential effects (presence of intra- and inter-genotypic variation (split of
464 neighboring genotypes in intra- and inter-genotypic genotypes). Thus, neighbor identity is split
465 into intra- and inter-genotypes as differential effects or are combined as neutral effects, which
466 includes all neighbors. P-value levels <0.05*, <0.005**, <0.001*** and >0.05^{ns}.



467

468 Figure 2. The barplots show standardized coefficient effect sizes and associated standard errors

469 of eco-evolutionary trait-based neighborhood models at small-scale (a, c) and large-scale (b, d)

470 for all MA-lines for the survival model (a, b) and the seed set model (c, d). Intra-genotypic

471 effects (yellow) and inter-genotypic effects (grey) are presented for 2004 and 2005. Coefficient

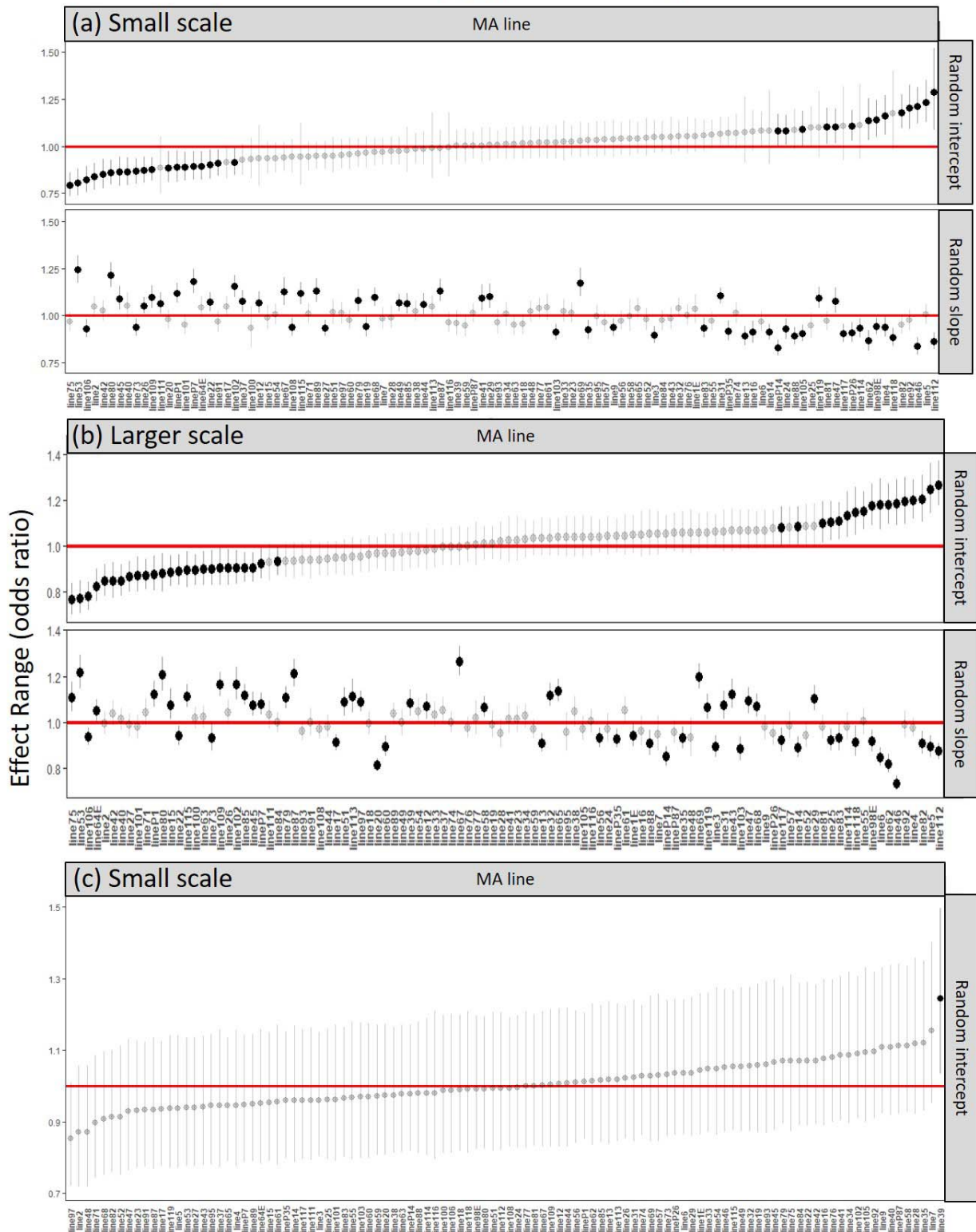
472 effect sizes indicate reduction or an increase of seed set depending on biomass density of intra-

473 or inter-genotypic neighbors. In addition, coefficient effect sizes indicate a reduction of survival

474 of intra or inter-genotypic neighbors.

475

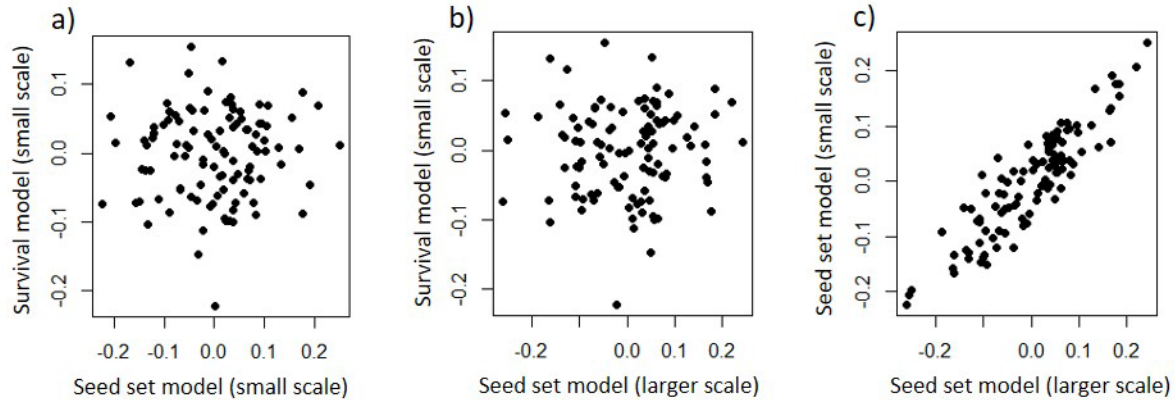
MA line effect sizes of seed set (a), (b) and survival (c) model



477 Figure 3. The effect range on seed set (a), (b) and survival (c) computed from eco-evolutionary
478 trait-based neighborhood models shown as odds ratios for year 2004 (for 2005 see
479 Supplementary Material). (a) Shows the hierarchical competitive order of MA lines and founder
480 for the seed set model including range effects of each MA line and founder as random
481 intercepts and the trait proxy biomass as random slopes at small-scale and (b) at large-scale. (c)
482 Shows the hierarchical selective order of MA lines and founder for the survival model including
483 range effects of each MA line and founder as random intercepts at small-scale.

484

Correlation between MA line effect sizes of significant seed set and survival model



485

486 Figure 4. Correlation of the simulated conditional mean represented as odds ratio of same
487 genotypes (MA lines) reducing or increasing the effect sizes of the seed set model and of the
488 survival model at small-scale or large-scale for 2004, respectively. For year 2005 see
489 supplementary material. Each point represents different genotypes of *A. thaliana*. Negative
490 values means lower effect sizes on the survival or seed set and positive values means increasing
491 the effect sizes on the survival or seed set. a) Relationship between survival and seed set
492 model. b) Relationship between survival and seed set model. c) Relationship between small-
493 scale survival and large-scale seed set model. Since we are plotting effect sizes, the positive
494 correlation in panel c reflects a trade-off between competition and facilitation (see results).

495

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