



## 32 **Abstract**

## 33 **Background**

34 The application of polymeric materials in medical industry has grown drastically in the last  
35 two decades due to their various advantages compared to existing materials. The present  
36 research work emphasizes on the sol-gel technique to formulate the polymethyl methyl  
37 acrylate/polystyrene/silica composite membrane.

## 38 **Methods**

39 The characteristic of the composite was investigated through modern state art of  
40 instrumentation.

## 41 **Results**

42 The functional groups attached to the polymer was absorbed by FTIR. The FTIR spectrum  
43 confirm that the blend was mixed thoroughly and the formation of unite intimately between  
44 the polymers. The membranes were observed by SEM for its surface homogeneity which  
45 depends upon the composition of the two blending polymers. The captured SEM images  
46 showed the formation of microcracks on the surface, which was evidently controlled by  
47 varying the constituent polymer ratios. The prepared blend membranes with 2:1 ratio of  
48 PMMA/PS/Si displayed higher water uptake compared to other blended membranes. The  
49 composite membranes had good hydroxyl apatite growth in SBF solution. Furthermore, the *in*  
50 *vitro* cytotoxicity studies carried out by MTT method, using RAW macrophage cells showed  
51 that all the samples exhibited excellent cell viability.

## 52 **Conclusion**

53 The inflammatory response of composite with equal concentration of PMMA-PS were  
54 performed and observed no inflammation in comparison with control and other tested  
55 concentrations.

56 Keywords: Sol-gel, PMMA-PS, Inflammatory response, drug release kinetics, SEM, MTT,  
57 FTIR.

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60

## 61 **Introduction:**

62 Immense research in biomaterials used for hard and soft body tissues replacement and  
63 orthopedic applications were constantly increasing during the past few decades[1–5].  
64 Biomaterials, used for this kind of replacement should be inert, bioactive and biocompatible.  
65 Based on the type of implant needed, the type of material like metals, alloys, ceramics and  
66 polymeric materials can be selected as a suitable biomaterial [6]. Among various kinds of  
67 biomaterials, polymeric biomaterials have gained more importance in recent days due to its  
68 vast advantages. Moreover, polymeric biomaterials are being used as a replacement of  
69 metallic materials (amalgam) due to their added advantage like light weight and tailor made  
70 properties[6]. There are several natural polymers like chitosan, cellulose having bioactivity  
71 and biocompatibility are used as biomaterials for various wide range of biomedical  
72 applications[7, 8]. However, the applications of these natural polymers are limited in terms of  
73 its stability and strength. The synthetic polymer is an alternative to natural polymers which  
74 may enhance the stability, strength and biocompatibility. These tailor-made properties of  
75 synthetic polymers improve its medical and biomedical applications. There is list of synthetic  
76 polymers such as polyamides (PAm), polyethylene (PE), polyether ether ketone (PEEK), poly  
77 methyl methacrylate (PMMA), polysulfone (PSu), polytetrafluoroethylene (PTFE),  
78 polyurethane (PU), and ultra-high molecular weight polyethylene (UHMWPE) used as  
79 biomaterials with inorganic nano bio-materials to induce bioactivity. The bioactivity of these  
80 polymers were induced by functionalizing the polymer or by blending with other polymer  
81 having bioactivity[9].

82 The PMMA is the most successful and investigated material in medical application  
83 such as implant in orthopedic applications due to its good bioactive and biocompatibility  
84 nature when used as bone cements in hard tissue replacements. Despite the many drawbacks  
85 like brittleness, shrinkage and high polymerization exotherm it used as bone cement for  
86 orthopedic applications is still serves a greater advantage. Similarly, polystyrene is one of the  
87 highly researched topic in biomedical applications known for its inert nature and has its  
88 applications in consumer, food packing industries has shown a good bioactivity and enhanced  
89 cell adhesion when modified with silica [10, 11]. It is an important criterion to use porous  
90 bioactive polymer matrix since it is more advantageous considering the ability of  
91 hydroxyapatite to grow within the pores, this phenomenon affixes the formation of interlock  
92 with the adjacent normal bone thereby lifts the fixation of polymer prosthesis. The  
93 application of these polymer matrix as biomaterials greatly influences its biological and

94 mechanical properties in internal bone implants due to the tailor-made nature [12].  
95 Regardless of advantages of PMMA possess a major drawback that is causing inflammation  
96 after the removal of the prosthesis from the implant site [13-14] will be addressed in the  
97 present study.

98 The sol-gel technique has been successfully used for preparation of  
99 PMMA/Polystyrene/Silica nanocomposite membrane. The sol-gel technique is selected due  
100 to various advantages includes, commendable chemical homogeneity, controlled size and  
101 morphology, operates at low temperature [15–17]. The sol-gel composed membranes are  
102 highly embodied matrices, which are used in the fields of biomedical sensors, laser materials  
103 and for sustained drug delivery applications. The present study propose about the bled  
104 formation between PS and PMMA based on the dipole interaction between aromatic groups  
105 of PS with that of the carbonyl group in PMMA. The schematic representation of the same is  
106 given in the Fig. 1.

107

108 **Figure 1** The mechanism of interaction that occurs between PS and PMMA

109

110 Based on the literature survey and to the best of our knowledge we are the first to  
111 report about the combination of PMMA/PS/Si composite material for medical application.  
112 The silica was selected as filler material due to its biocompatibility and ease availability in  
113 nature. There were quite an enormous amount of research that have been conducted with  
114 silica based polymeric materials for biomedical applications such as implants and artificial  
115 skin [18, 19]. Apart from its biocompatibility nature, its incorporation also increase the  
116 strength of polymeric materials and influence the creation of apatite formation thereby  
117 enhancing the bioactivity of composite by interacting with negatively charged silanol and  
118 SBF solution. In the present study, PMMA/PS/Si composite membranes were prepared in  
119 different weight ratios of PMMA and PS (1:1, 1:2, 2:1) with constant weight of silica. The  
120 prepared composite membranes were tested for its bioactivity by immersing in SBF and  
121 cytotoxicity studies by MTT method.

122

## 123 **Materials and Methods**

124 The materials that were used for the study were procured commercially from different  
125 sources. PMMA (Mol. Wt 35,000 Da) was procured from Asian acrylates, Mumbai,  
126 Polystyrene from Sigma Aldrich. Silica and Tetraethyl orthosilicate (TEOS, 99.5% pure)

127 were purchased from Sigma Aldrich. THF, Hydrochloric acid was purchased from Merck.  
128 Different polymer system employed and their corresponding code, weight ratio is shown in  
129 Table 1, in all the case silica concentration was kept constant.

130

### 131 **Preparation of Composite membranes**

132 The polymer silica composite membranes were prepared by weighing different amounts of  
133 PMMA and Polystyrene in separate beakers followed by dissolving in THF solvent. The  
134 polymers solutions were mixed together with constant stirring to form PMMA/PS blends.  
135 The silica particles as mentioned in Table 1 were added to the homogenous polymer solution  
136 and subsequently stirred, ultra-sonicated for 2 hours to get uniform dispersion of silica  
137 particles. In order to enhance the gelation process 0.25 ml of 35% HCl, 5 ml ethanol and 1ml  
138 of tetra ethoxy silane (TEOS) were added to the polymer solution. The solutions were left for  
139 three days stirring to complete the gelation process. The gel obtained was then casted onto  
140 clean-dried petri dishes and left undisturbed for three days at room temperature. After  
141 complete removal of solvent, the membranes were removed from the petri dish for the further  
142 analysis.

143

### 144 **Fourier Transform Infra-Red Spectroscopy (FTIR)**

145 The presence and interaction of different functional groups in polymers were studied using  
146 FTIR. The composite membranes of PMMA/PS prepared were characterized using Perkin  
147 Elmer Spectrum RXI IR spectrophotometer.

148

### 149 **Scanning Electron Microscope (SEM)**

150 The surface morphology and dispersion of silica fillers in the prepared composite membranes  
151 were studied using HITACHI S-3400N Scanning Electron Microscope (SEM). Prior to the  
152 analysis, the samples were dried and their surface was gold sputtered.

153

### 154 **Water absorption**

155 Water absorption property of the polymer composite membranes was carried out as per our  
156 previous studies[20]. The study was carried out by immersing pre-weighed dried membranes  
157 in deionized water on a glass beaker for 24 hours. After 24 hours, wet membranes were  
158 retrieved from the beaker and excess water was blotted gently using a tissue paper. The wet

159 samples were again weighed, and the water absorption percentage was calculated using the  
160 formula.

$$161 \quad \% \text{ Water absorption} = \frac{\text{wt. of wet polymer} - \text{wt. of dry polymer}}{\text{wt. of dry polymer}} \times 100$$

162

## 163 **Bioactivity Study**

164 The bioactivity of the membranes prepared was studied by immersing the membranes in  
165 simulated body fluid (SBF) for 15 days (with and without silica fillers) and analyzing their  
166 surface for the formation of mineral (hydroxycarbonate apatite) layer using SEM (Quanta 200  
167 FEG scanning electron microscope). SBF also known as Kokubo's Solution was prepared  
168 according to the specification given by Kokubo et al [21]. The chemicals required to prepare  
169 SBF were dissolved in deionized water in the specified quantity while the pH was maintained  
170 at 7.25.

171

## 172 **Drug Release Kinetics**

173 The effective drug release kinetics of composite membranes prepared were tested using 5-  
174 Fluorouracil as model drug. The solution was prepared by dissolving 12 mg 5-FU in 1000ml  
175 of deionized water. The composite membranes prepared were weighed and cut into 1cm<sup>2</sup>  
176 followed by immersion in drug solution for 24 h. The increase in weight of sample immersed  
177 after 24 h was measured and compared with initial weight to determine amount of drug  
178 loaded onto the membrane. The amount of drug released from composite was evaluated using  
179 a UV-Visible spectrophotometer (T90+UV/Vis Spectrometer, PG Instruments) at a  
180 wavelength of 293 nm. The drug release kinetics was determined from the drug absorbance  
181 values after immersion in PBS solution and comparing it with the standard values obtained in  
182 the initial composites [22].

183

## 184 **Biocompatibility test**

185 The composites prepared must be biocompatible such that it can be applied *in vivo*. Hence, in  
186 the present study we used MTT assay to quantify the biocompatibility of prepared  
187 composites. The cell viability was determined based on our previous studies [23]. The  
188 biocompatibility analysis was performed using RAW macrophage cell line 264.7 that were  
189 grown on Dulbecco's Modified Eagle's Medium in a 96 well plates. The cells were allowed  
190 to settle after which the samples were added and the viability ratio was analyzed based on

191 absorbance values at 545nm. The obtained values were plotted against the standard control  
192 group and viability ratio were estimated.

193

## 194 ***In vitro* inflammatory studies**

195 We have evaluated the inflammatory response of PMMA-PS composited using RAW 264.7  
196 macrophage cells in vitro. The cells were tested with composite prepared and inflammatory  
197 responses for cytokines TNF- $\alpha$ , IL-6 and IL-1 were checked using qPCR. Briefly, the cells  
198 were treated with the samples and allowed for 6h incubation. The first inflammatory response  
199 was analyzed by extracting the RNA and reverse transcribing the same to form cDNA.  
200 Followed by the analyzing of expression of inflammatory markers using qPCR.

201

## 202 **Statistical Analysis**

203 All the experiments were repeated for five times and average from them was used for the  
204 study.

205

## 206 **Results and Discussion**

### 207 **FTIR**

208 The FT-IR Spectra showed the shifting of functional groups towards the lower frequency  
209 from their native frequency due to blending of polymer resulting in the formation of weak  
210 hydrogen bond.

211

212 **Figure 2** FTIR analysis of showing the interaction between PMMA and PS

213

214 The blend containing 1:1 ratio showed characteristic absorption peaks of PMMA  
215 corresponding to asymmetric stretching of CH<sub>3</sub> and C=O were assigned to bands 2951 cm<sup>-1</sup>  
216 and 1736 cm<sup>-1</sup> (Figure 2). The peaks 1482 cm<sup>-1</sup> is a characteristic vibrational band for CH<sub>2</sub>  
217 scissoring, similarly the vibrational band at 1452 cm<sup>-1</sup> corresponding to asymmetric vibration  
218 of CH<sub>3</sub> stretching or due to the PMMA deformation which was further confirmed by OCH<sub>3</sub>  
219 deformation peaks at 1390 cm<sup>-1</sup> (Figure 2). In addition, the absorption peaks at 1600 and 698  
220 cm<sup>-1</sup> in all the blends represents the C-C stretching and ring deformation of polystyrene. The  
221 broadened peaks at 750, 1050 and 1200 were observed for blend of 1:1 which are correspond  
222 to CH<sub>2</sub> twisting, wagging and rocking respectively, whereas the blend of 2:1 and 1:2

223 appeared to be smooth curve in the same region [24]. The IR results confirm that the blend  
224 was strongly mixed and formation of polymer blends in case of equal concentration of  
225 polymer compared to other two concentrations. It was observed that the results were in par  
226 with other reports and confirmed that there was no major shift in the peaks hence the blends  
227 may be due to physical bonding rather than a chemical one [25].

228

## 229 **Water absorption studies**

230

231 **Figure 3:** Percentage water absorption of the prepared composite membranes

232

233 The results of the water absorption studies for the three groups is shown in Figure 3. Figure 3  
234 shows that the composite membrane of PMMA/PS 2:1 has the maximum water absorption  
235 while the membrane of 1:2 has the least water absorption. This variation is attributed to the  
236 hydrophobicity of polystyrene. Hence, it was observed with increase in concentration of  
237 polystyrene decrease in water absorption capacity of the prepared composite membranes. The  
238 swelling capacity of membranes increases with decrease in the dimensional stability, which  
239 in turn directly influence the release of drug from membrane.

240

## 241 **Surface Morphology – SEM Analysis**

242 The SEM images of the composite samples are shown in Figure 4. From the images, it was  
243 clearly seen that in all the samples, the silica fillers are evenly distributed throughout the  
244 polymer matrix.

245

246 **Figure 4.** SEM images of a) PMMA:PS (1:2) b) PMMA:PS (1:1) c) PMMA:PS (2:1)

247

248 The cracks that appear on the surface of the composite membranes are due to capillary  
249 pressure occurring during the solvent evaporation. The presence of these cracks on the  
250 surface will be advantageous for drug delivery and tissue engineering applications. The  
251 presence of these pores and cracks on the surface will help in the ingrowth of the  
252 hydroxyapatite crystals thus enabling good mechanical and chemical bonding of the bone  
253 with the polymer surface.

254

255



## 256 **Tensile strength**

257 The tensile strength of the prepared samples is shown in figure 5. From the figure, it is seen  
258 that the sample of sub group 3 having PMMA: PS ratio of 2:1 has the maximum tensile  
259 strength of 78.3 MPa. With a decrease in the concentration of PMMA in blend membrane, the  
260 tensile strength is also found to decrease gradually due to elastic nature PMMA possess.

261

262 **Figure 5.** Tensile strength of the prepared composites

263

## 264 **Bioactivity study**

265 Bioactivity of the prepared composite membranes was determined by keeping the membranes  
266 immersed in Kokubo solution and observing the growth of the apatite under the scanning  
267 electron microscope. The SEM images are shown in figure 6

268

269 **Figure 6 Shows the apatite growth on the surface of the subgroups 1, 2, 3 of PMMA:PS**  
270 **with silica fillers**

271

272 The SEM images of the three subgroups after immersion in SBF for 15 days at room  
273 temperature is shown in figures 6 that correspond to with and without the silica fillers in  
274 composites. It was clear from figure 6 a, b, c that all the composites incorporated with si  
275 fillers showed excellent bioactivity compared with to the ones that were not incorporated  
276 with si. It was clear from the SEM that the si enhance the bioactivity of membranes and it  
277 was observed that PS as well plays an important role in the bioactivity. It was observed from  
278 the figure 6a, b & c that the composites with equal concentration of PMMA and PS were  
279 highly bioactive and the composite with more wither of PMMA or PS showed comparatively  
280 lower amount of bioactivity. The bioactivity of composites with more concentration of PS  
281 was less bioactive compared to other two composites respectively.

282

## 283 **Drug Release Kinetics**

284 The composites prepared for applying as bone cement was checked for its effective drug  
285 release kinetics as it may serve for multipurpose in enhancing the bone growth with active  
286 release of drug (figure 7). It was observed from drug release kinetics that the drug was  
287 released in a sustained manner for a period of 180 h in PBS at pH 6.8. The initial phase or  
288 burst release was observed in all three cases the models that is in par with other research

289 reports [26-29]. It has to be noted that the drug release in model 1 may be due to fickian  
290 diffusion and took a very long time owing to higher concentration & hydrophilic nature of  
291 PMMA that makes the drug release in a much-sustained fashion. This was not the case with  
292 model 2 & 3, which exhibited a quick release of drug compared to model 1, and this can be  
293 attributed to the complete degradation between the blended membranes since it was not a  
294 chemical bonding between the two polymers. The release kinetics were in par with FTIR  
295 results and confirmed the presence of physical interaction between polymers that might have  
296 resulted in quick release of drug from composites.

297

298 **Figure 7** shows the drug release kinetics of the prepared composite materials with varying  
299 concentration of PMMA: PS

300

301 The release kinetics were fitted onto peppas model to further be confirmative about the type  
302 of release that might have occurred from composite models [30]. The model 1 had a value of  
303  $n \geq 5$  when fitted onto peppas model attributing to the fickian diffusion, the other two  
304 composite did not fit into any other models, which is out of the scope of our study.

305

### 306 ***In vitro* Cytotoxicity by MTT method**

307 The cytotoxicity studies performed showed interesting results. Figure 8 shows the RAW  
308 macrophage 264.7 cells after treating with composite membranes for 48 h. It can be observed  
309 that the cells treated with equal concentration of PMMA: PS showed confluent cells  
310 compared with the other two concentrations. It can be visually observed that amount of viable  
311 cells very lower in case of higher concentration of either PMMA or PS. Figure 8 (b) shows  
312 high density of macrophage cells indicating its compatibility for cell proliferation.

313

314 **Figure 8** Microscopic images of cytotoxicity study. a) Model 1, b) Model 2, c) Model 3

315

316 The results were in par when studied with MTT assay where the composites with equal  
317 concentration of polymers showed an enhanced viability compared to the other two models.  
318 The percentage viability of the cells was calculated after noting the OD values of the control.  
319 The results are displayed in figure 9 in which a graph is plotted by taking the cell count of the  
320 control group as 100 percentage. The viability percentage was in the range of  $93.5 \pm 3$  in case  
321 of equal concentration of polymer whereas it was lower on the other two cases. Among the

322 three groups, the sub group 2 has the best biocompatibility among other subgroups of  
323 PMMA: PS.

324

325 **Figure 9** Graphical representation of the cell viability of the three groups

326

### 327 ***In vitro* Inflammatory Response**

328 The importance of these composites when applied *in vivo* has to be taken into consideration  
329 since most of them cause inflammation when applied *in vivo* or during the application as  
330 prosthesis. Hence, in the present study, we evaluated inflammatory response of these  
331 composites on RAW macrophage cells and its effect was analyzed with inflammatory  
332 specific genes like TNF- $\alpha$ , IL-6 and IL-1 (figure 10). It was observed from the results that  
333 most of composites were not causing a major inflammation in all the three cases and in  
334 particular the expression was least in case of equal concentration of PMMA and PS. It was  
335 also observed that the expression of IL-6 marker gene was at a higher end in case of higher  
336 concentration of PMMA. The results from qPCR confirmed that the concentration has a  
337 direct role in causing inflammation particularly in dental applications that is in par with the  
338 one reported by Spasojevic et al [14] as they. It can be observed that these onflammatory  
339 expression was at lower end with the addition of PS only at a particular concentrations. When  
340 the concentration of PS was increased the inflammatory resposne as well got increased  
341 slightly compared to the equal concentrations. Hence, it is evident that only at equal  
342 concentration of PMMA and PS the composites very good properties. The increase in  
343 concentration of PMMA or PS was not good property as that of the equal concentration.  
344 Therefore, the composites with equal concentration of PMMA-PS can eb suitable for bio  
345 medical aplications.

346

347 **Figure 10** Inflammatory response observed on RAW macrophage cells – *in vitro*

348

### 349 **Conclusion**

350 Polymer silica composites of PMMA/PS were successfully fabricated by sol-gel technique.  
351 SEM and FTIR characterized the blend membranes and confirmed the presence of physical  
352 bonding between them. The SEM images showed that the surface homogeneity of the  
353 membranes was dependent on the concentration and blending between the two constituent

354 polymers. The blend formation between the polymers in the three groups was confirmed by  
355 FTIR. The bioactivity of membranes were analyzed by immersing the samples in SBF and  
356 examining the surface for the formation of apatite layer under the scanning electron  
357 microscope (SEM). Among the three groups, model 1 containing equal concentration of  
358 PMMA/PS blend composite showed the high bioactivity while the group two showed the  
359 most dense apatite formation of Ca-P crystals on its surface. The cytotoxicity studies were  
360 performed to evaluate the biocompatibility of the fabricated composite membranes. The *in*  
361 *vitro* study carried out by MTT method showed the composite with equal concentration of  
362 group II and III samples exhibited excellent cell viability while the group I ,the sample of the  
363 subgroup 3 namely PMMA:PS in the ratio of 2:1 exhibited favorable cytotoxicity value than  
364 the other two subgroups. From this study, it can be concluded that the sol-gel technique is  
365 versatile method to fabricate the polymer composites having immense biomedical  
366 application. Among the polymers investigated, model 1 samples were less suited for load  
367 bearing orthopedic applications but more suited for sustained drug delivery and tissue  
368 engineering application. Model 1 showed very good tensile properties whereas other two  
369 models showed optimum biocompatibility and moderate strength that shall be suitable  
370 orthopedic application.

371

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380

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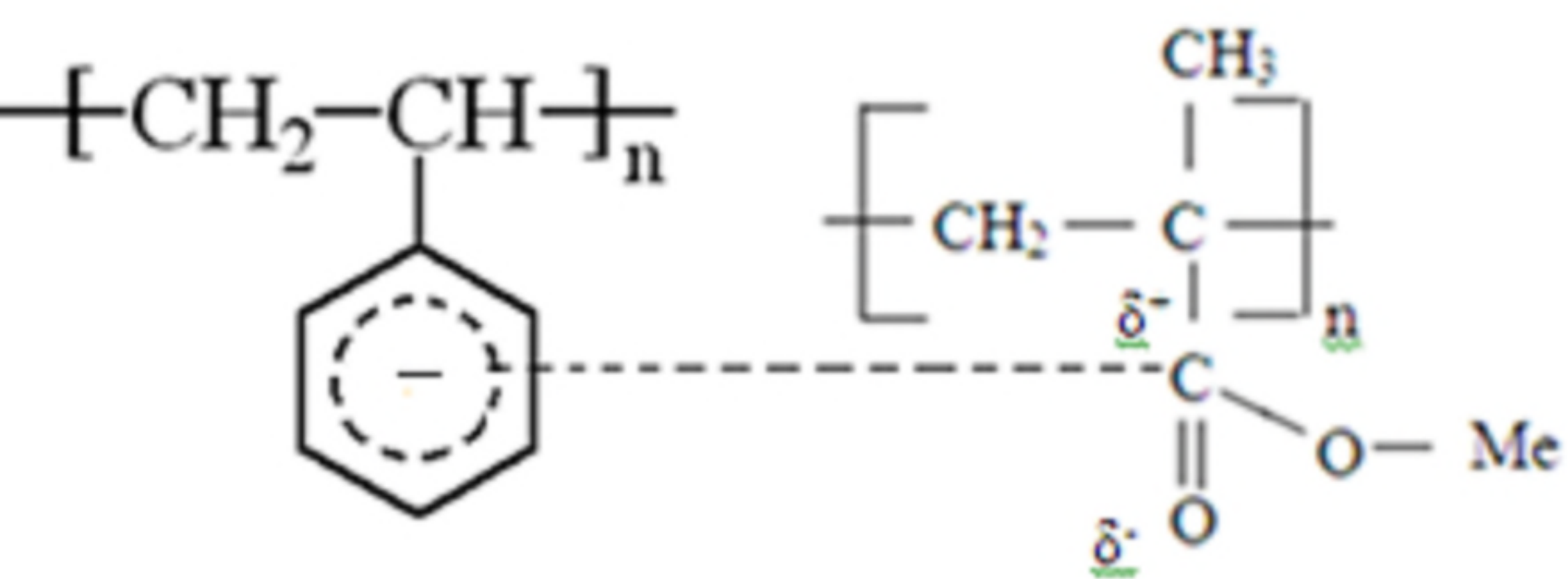


Fig1



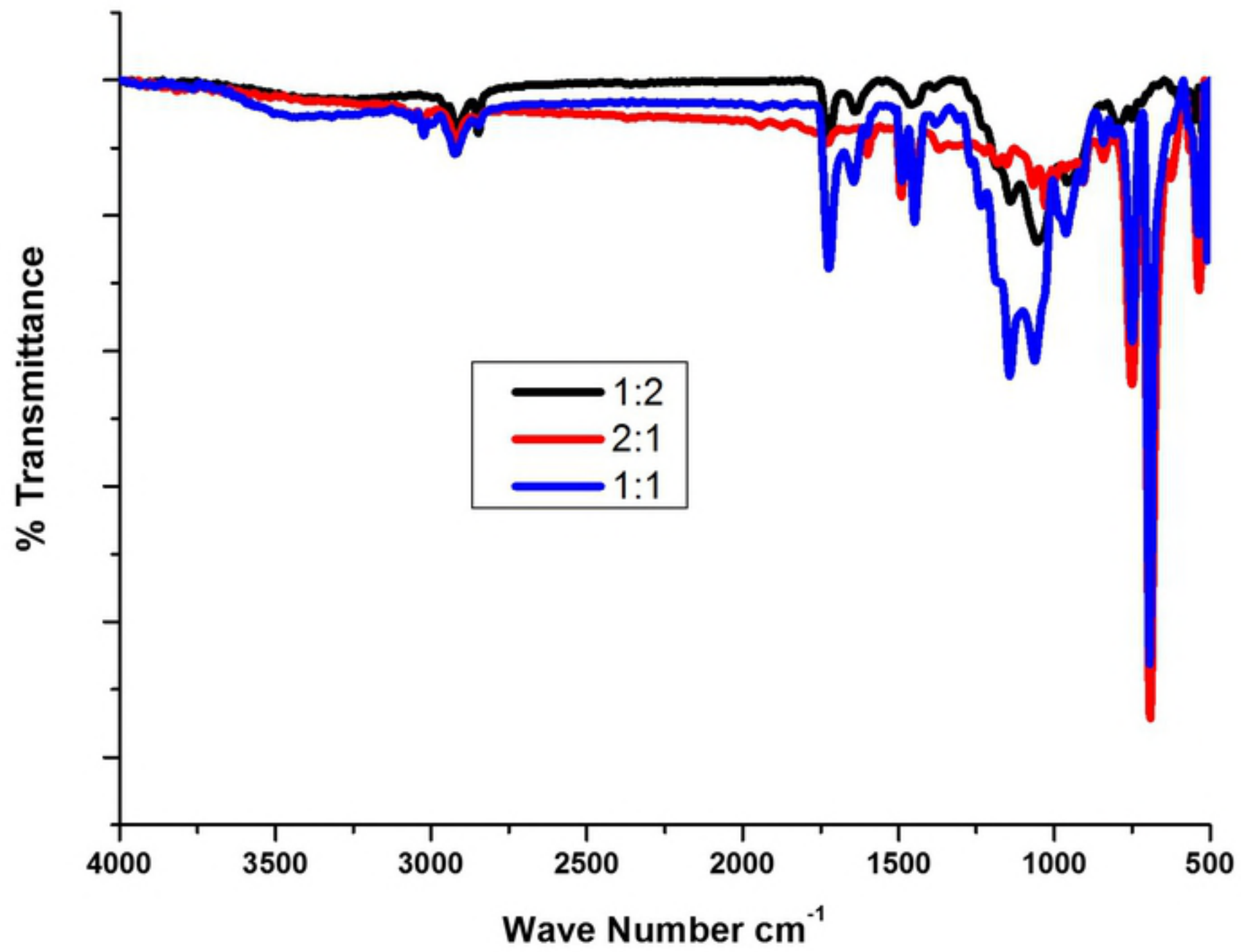


Fig2

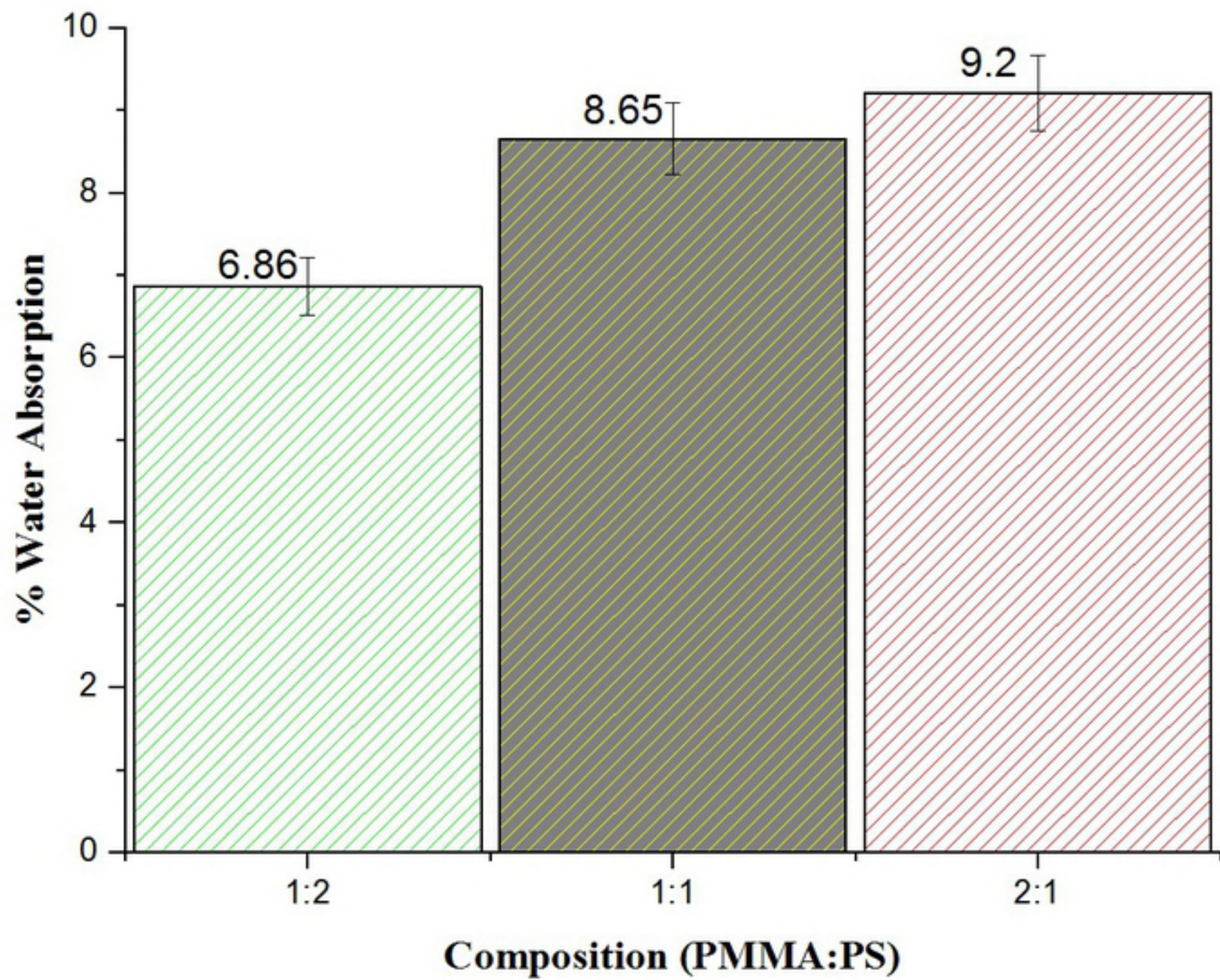


Fig3



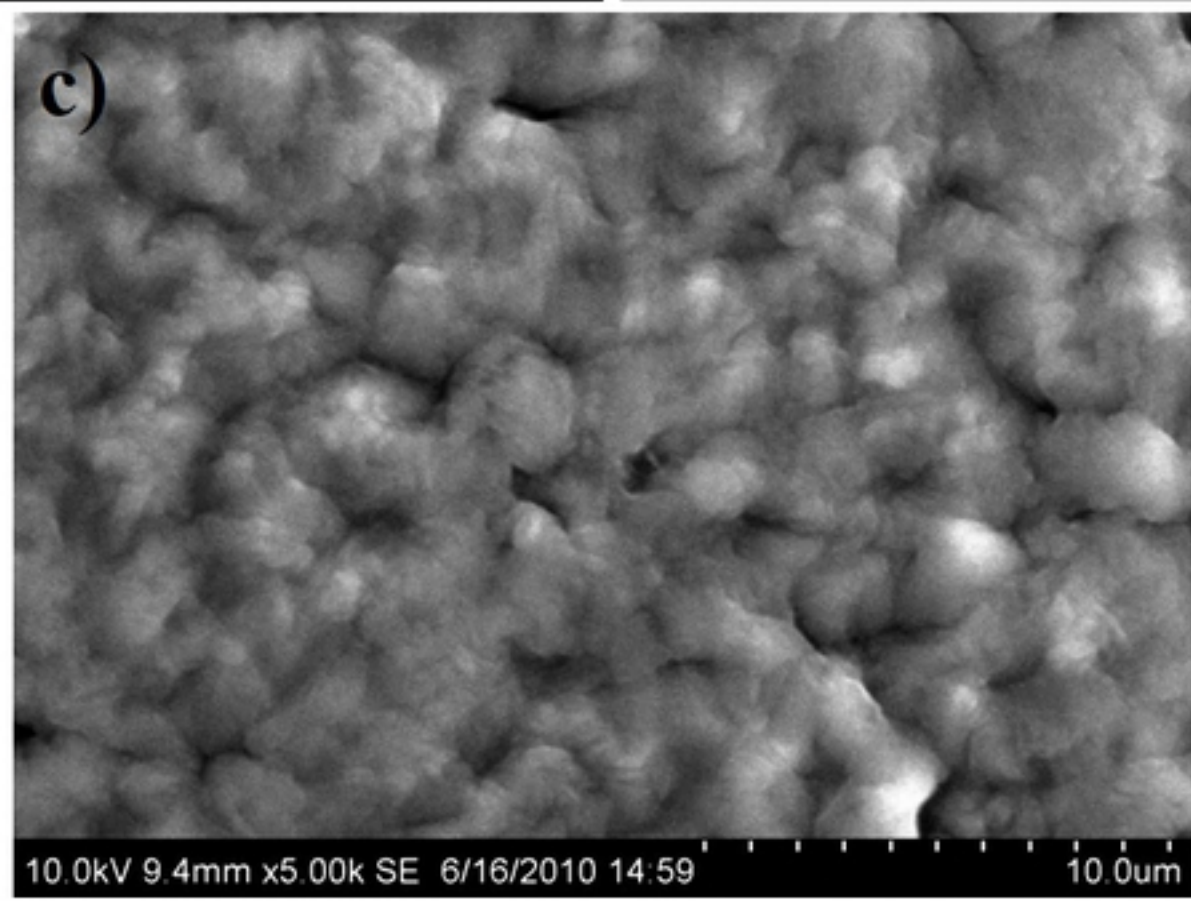
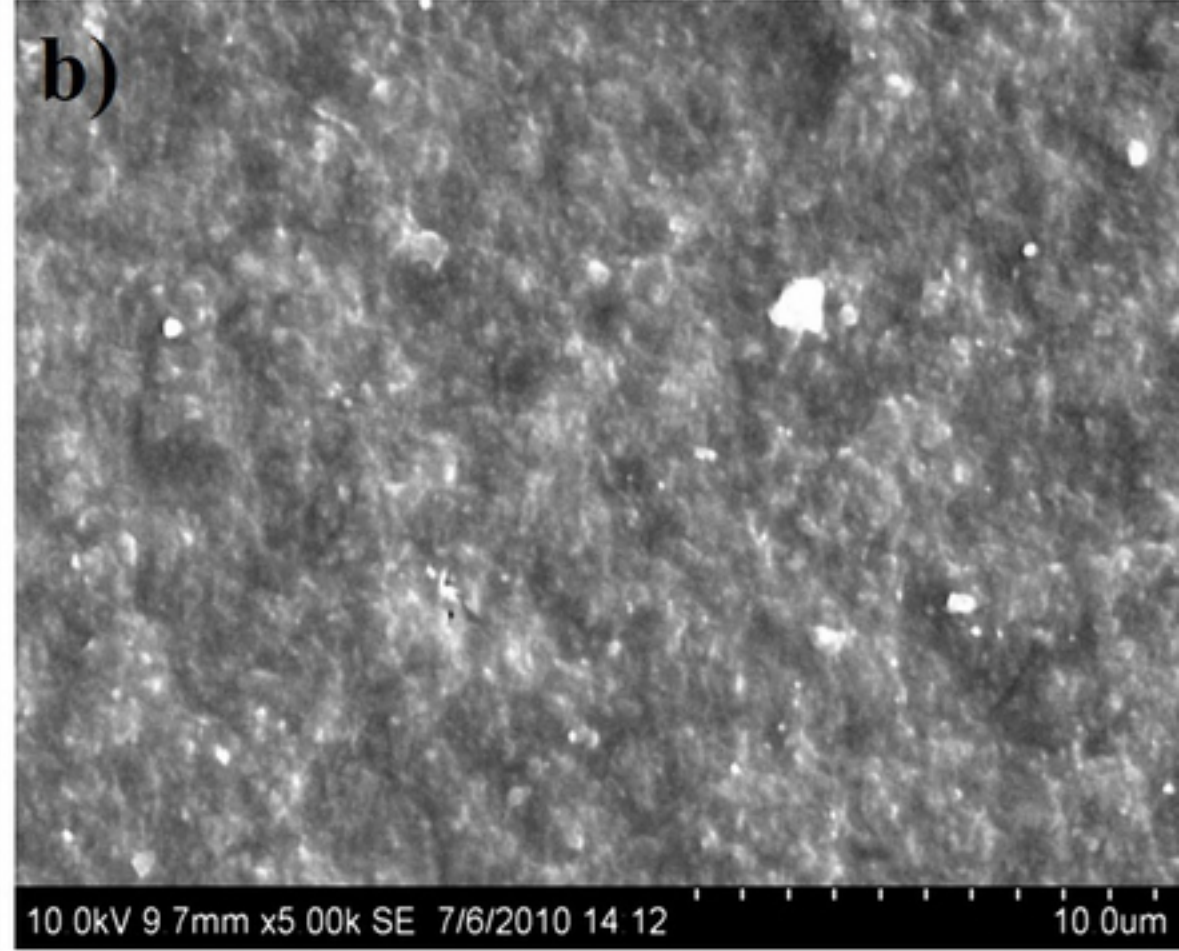
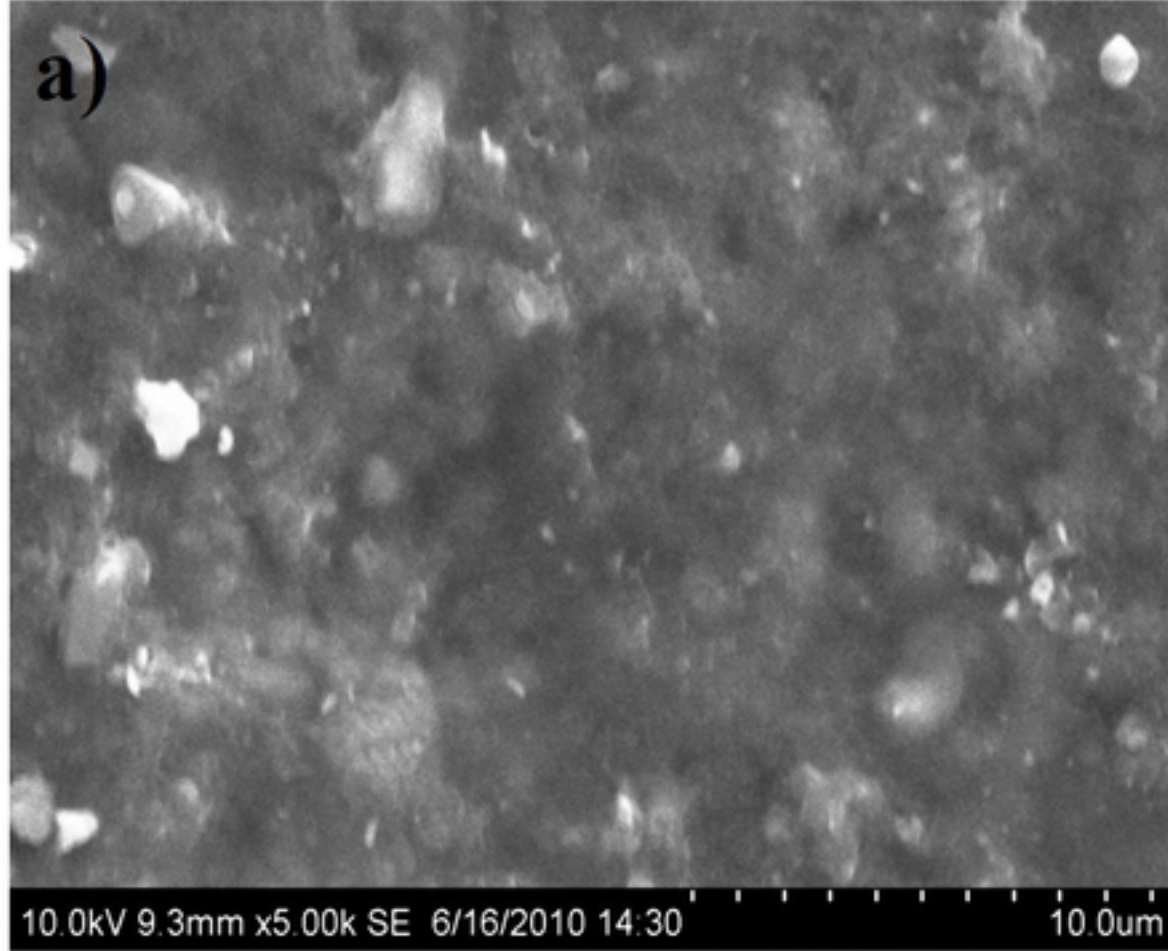


Fig4



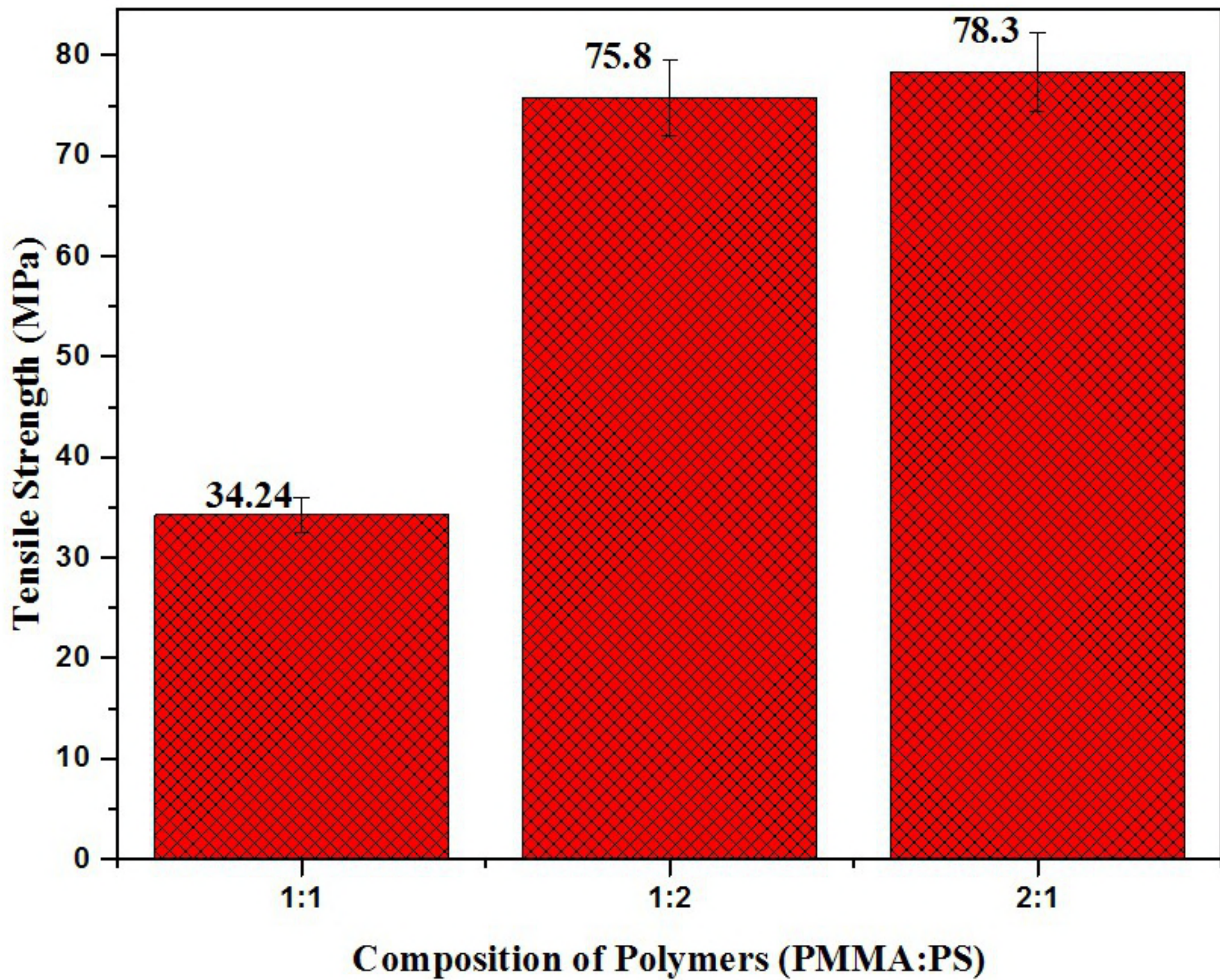


Fig5



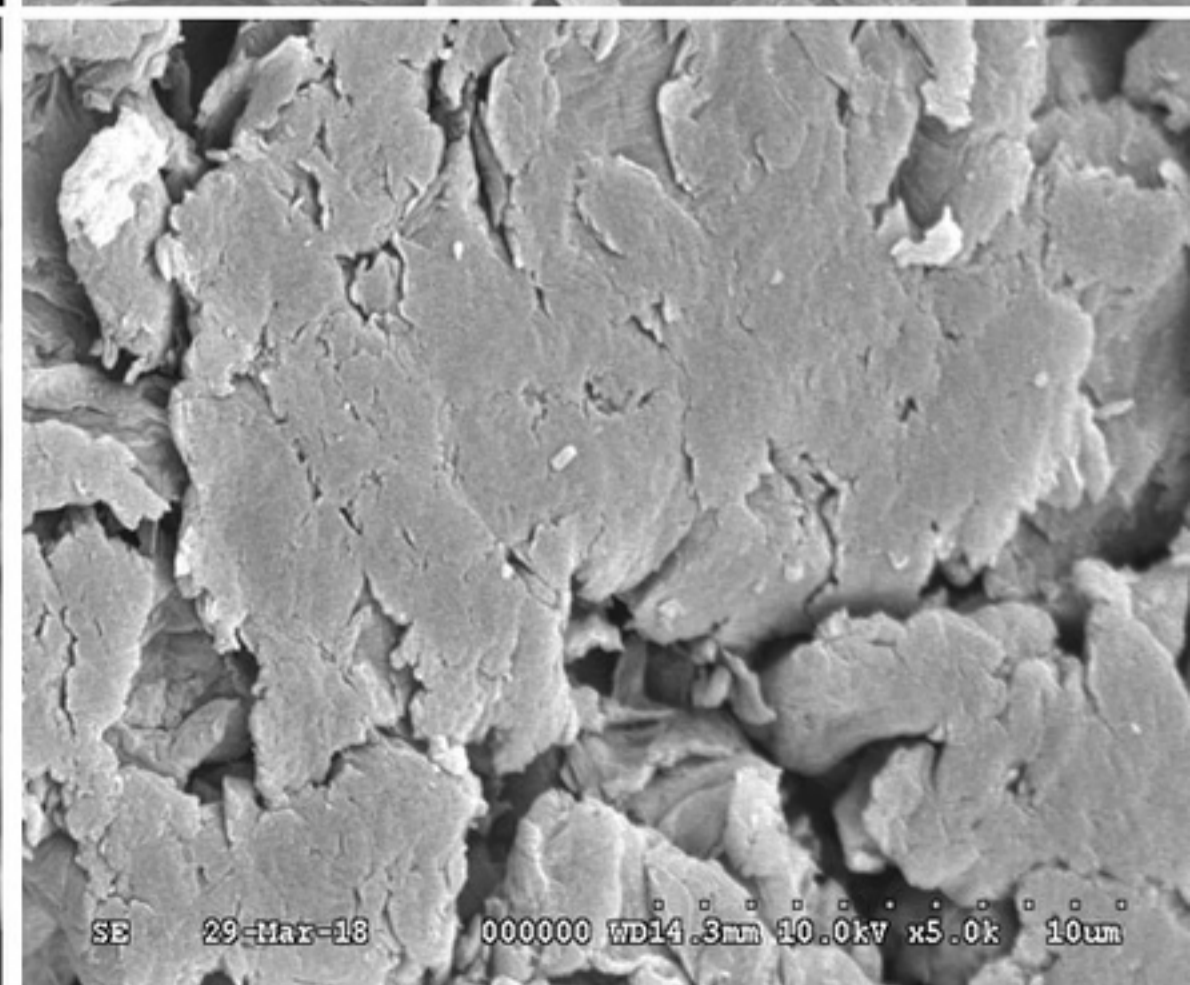
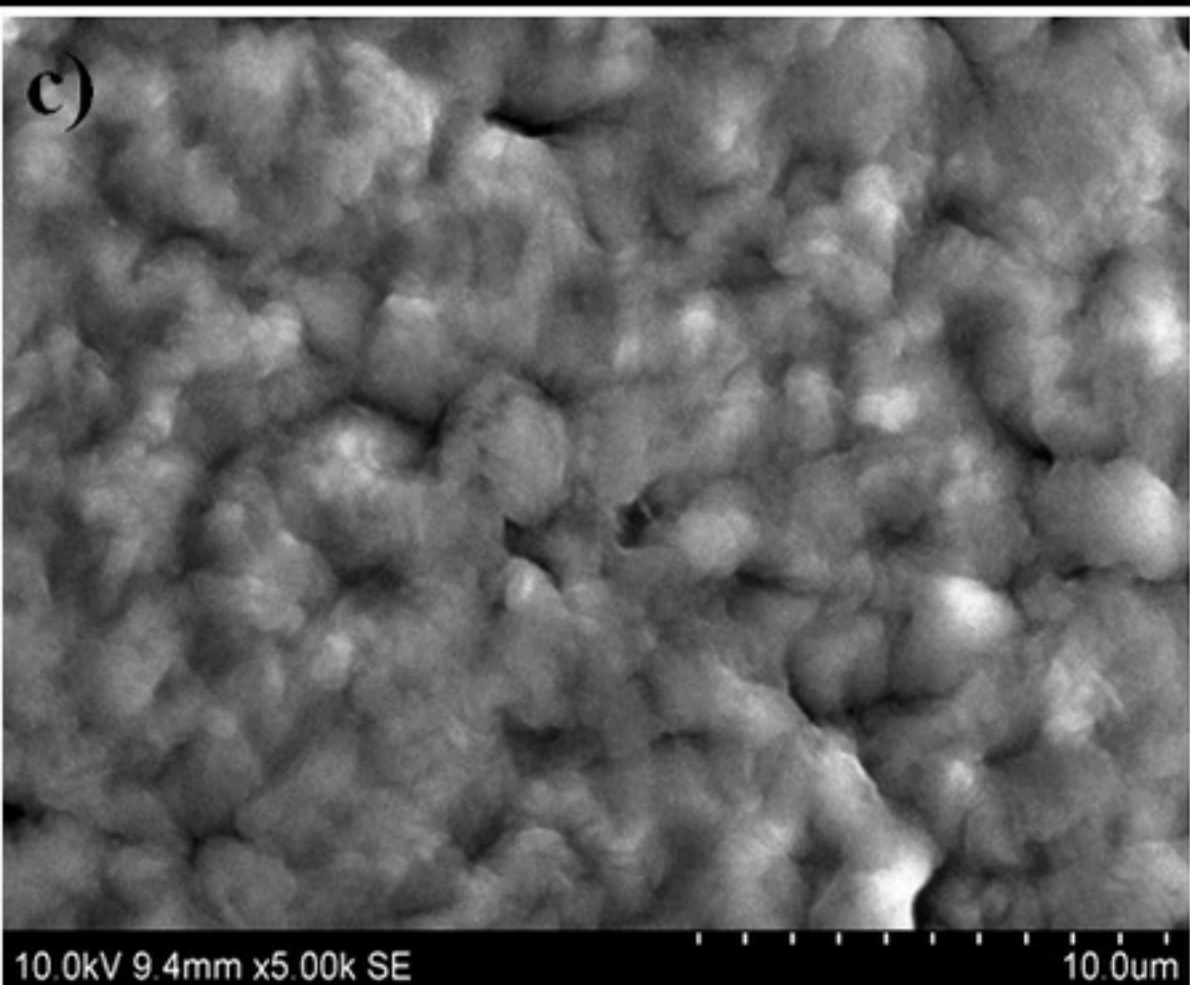
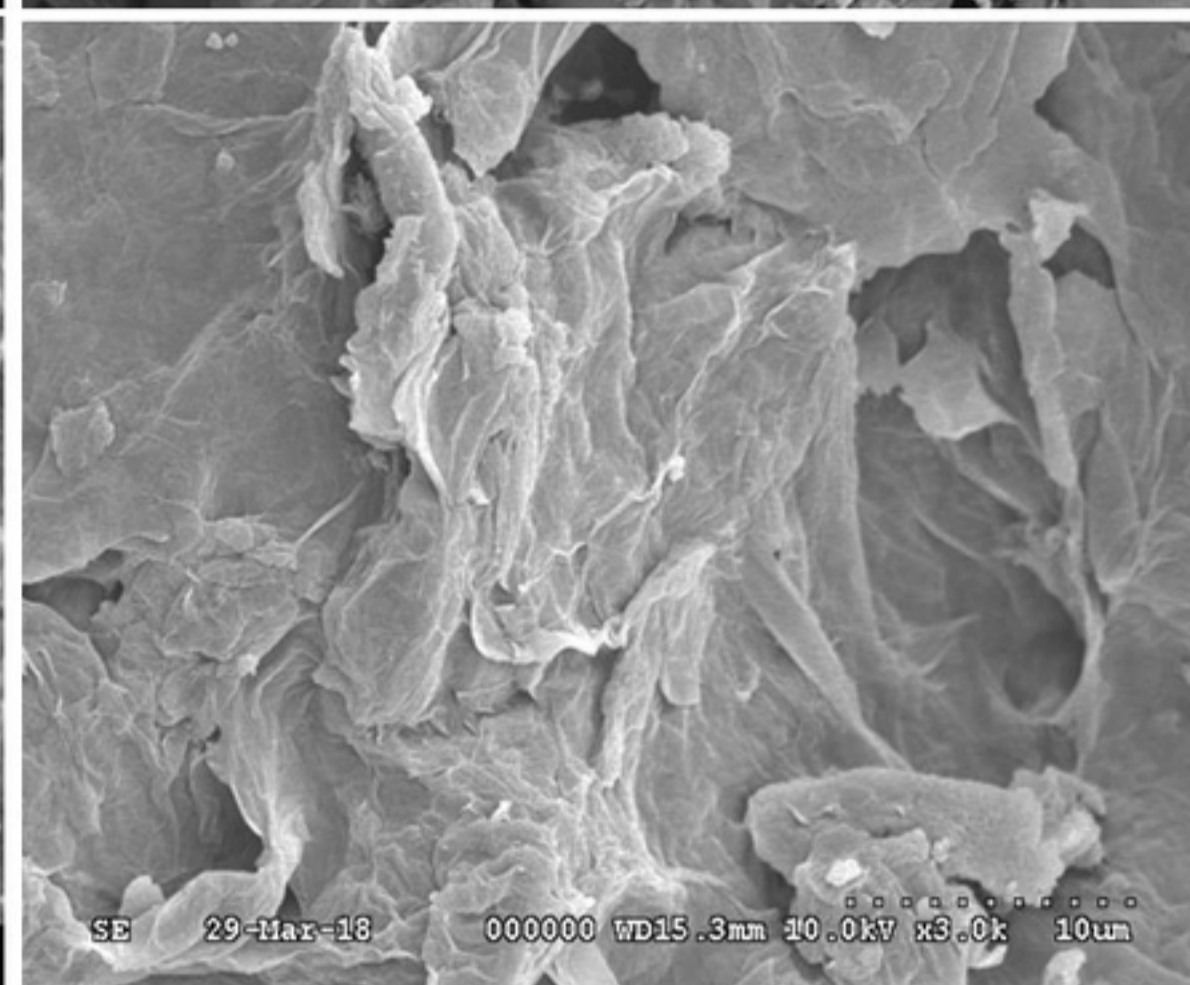
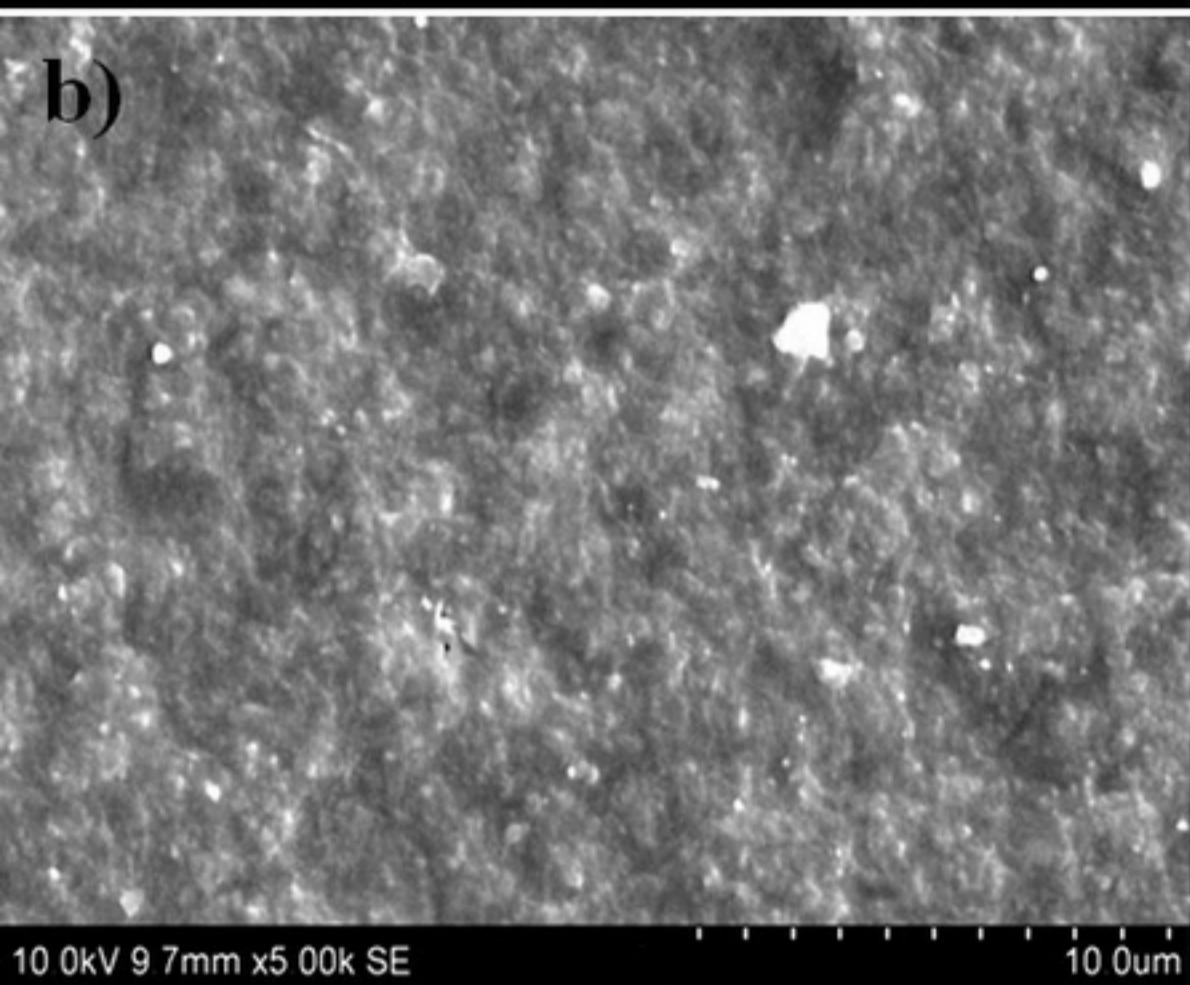
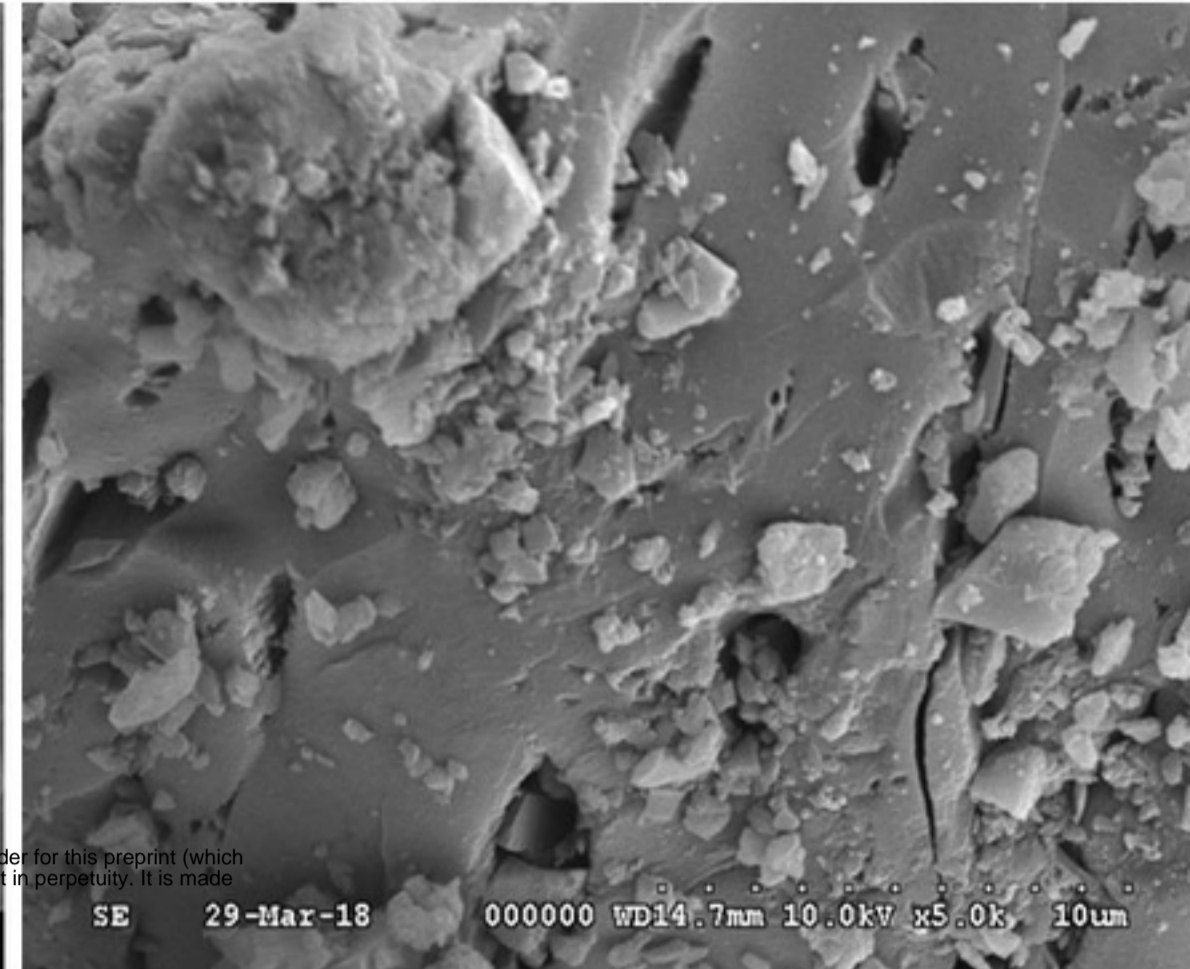
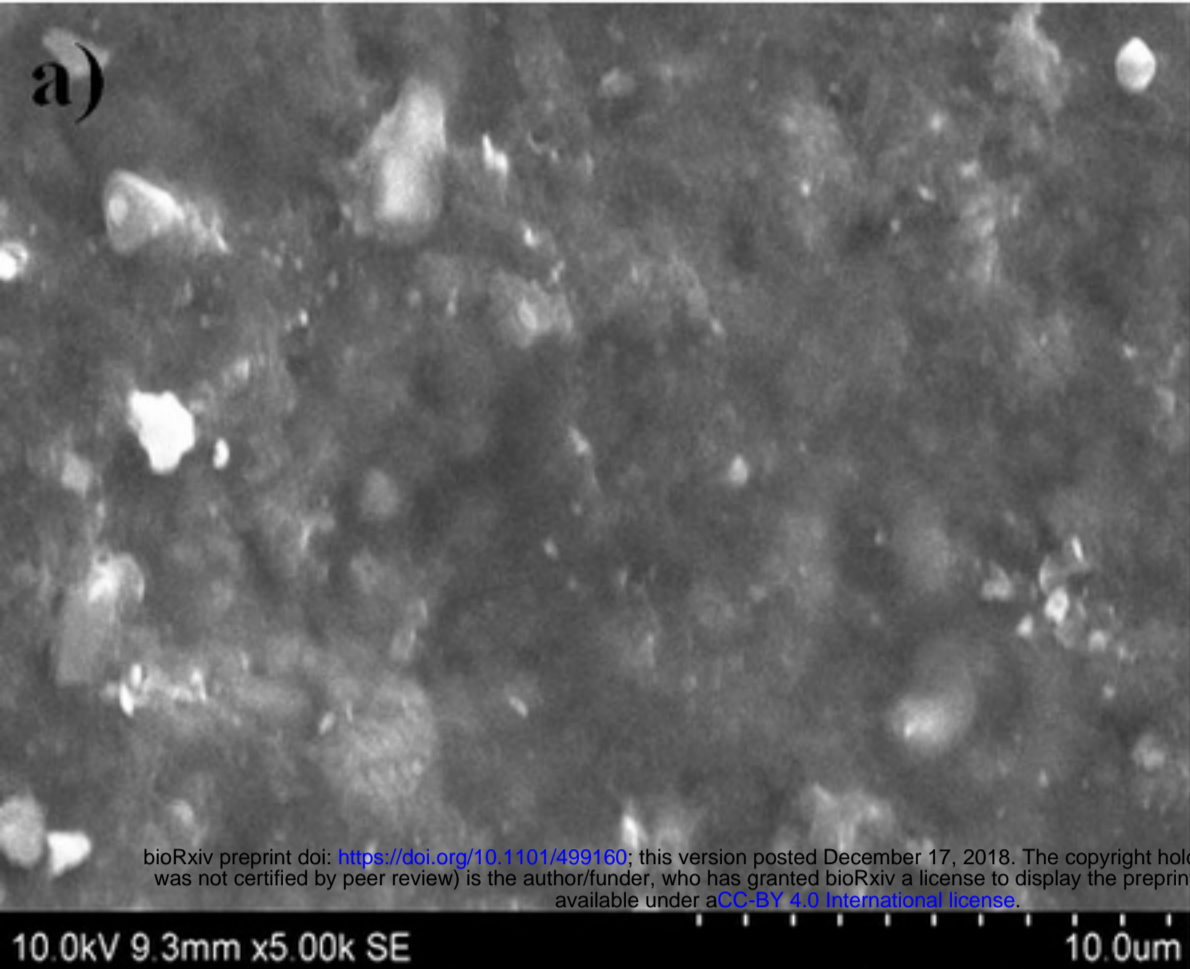


Fig6



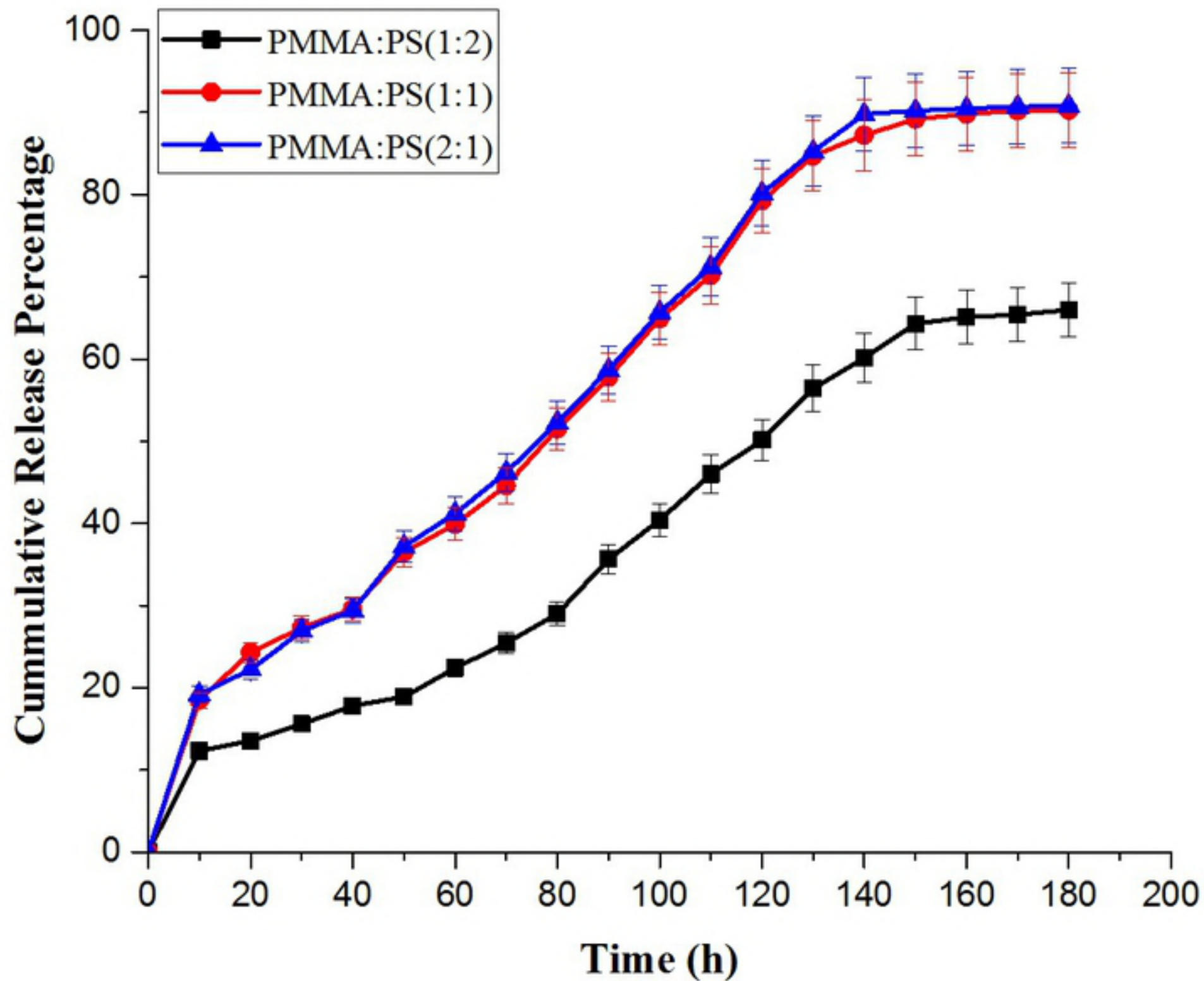


Fig7

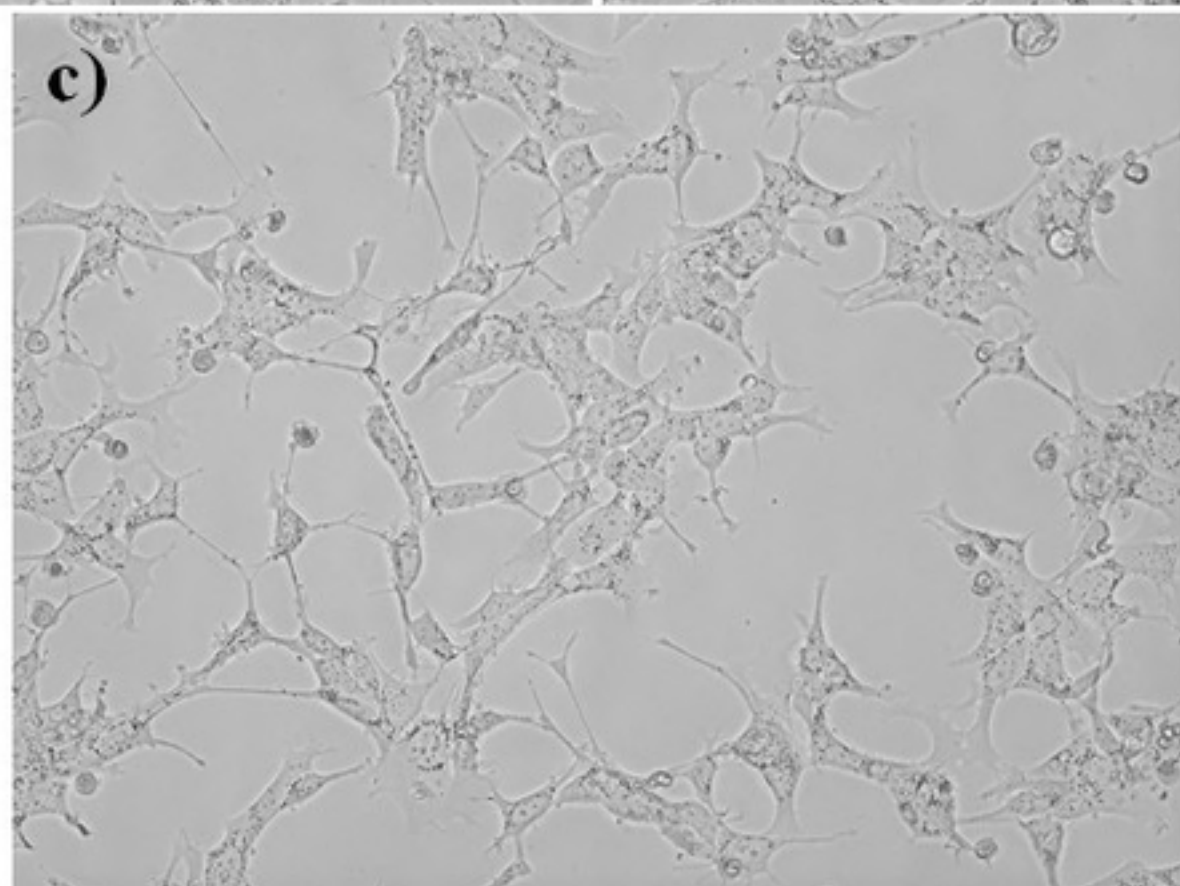
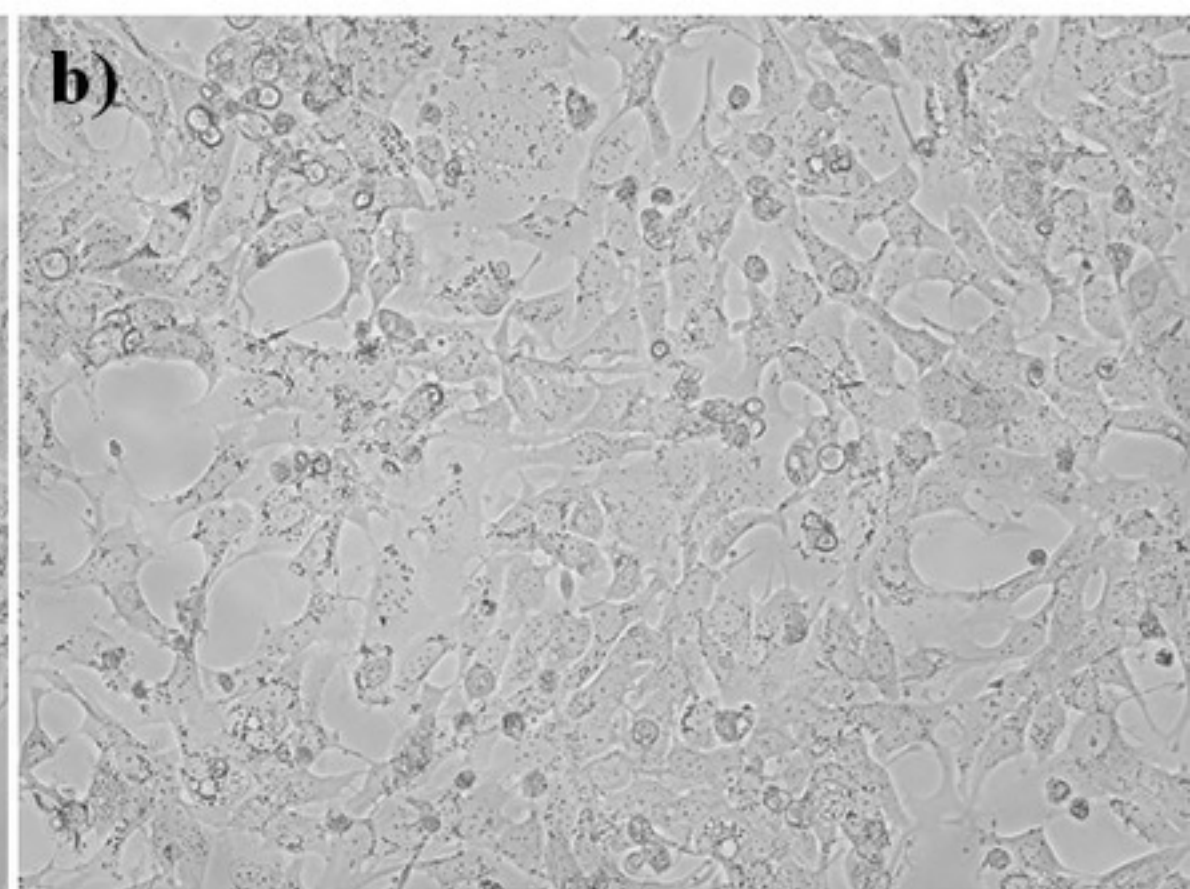
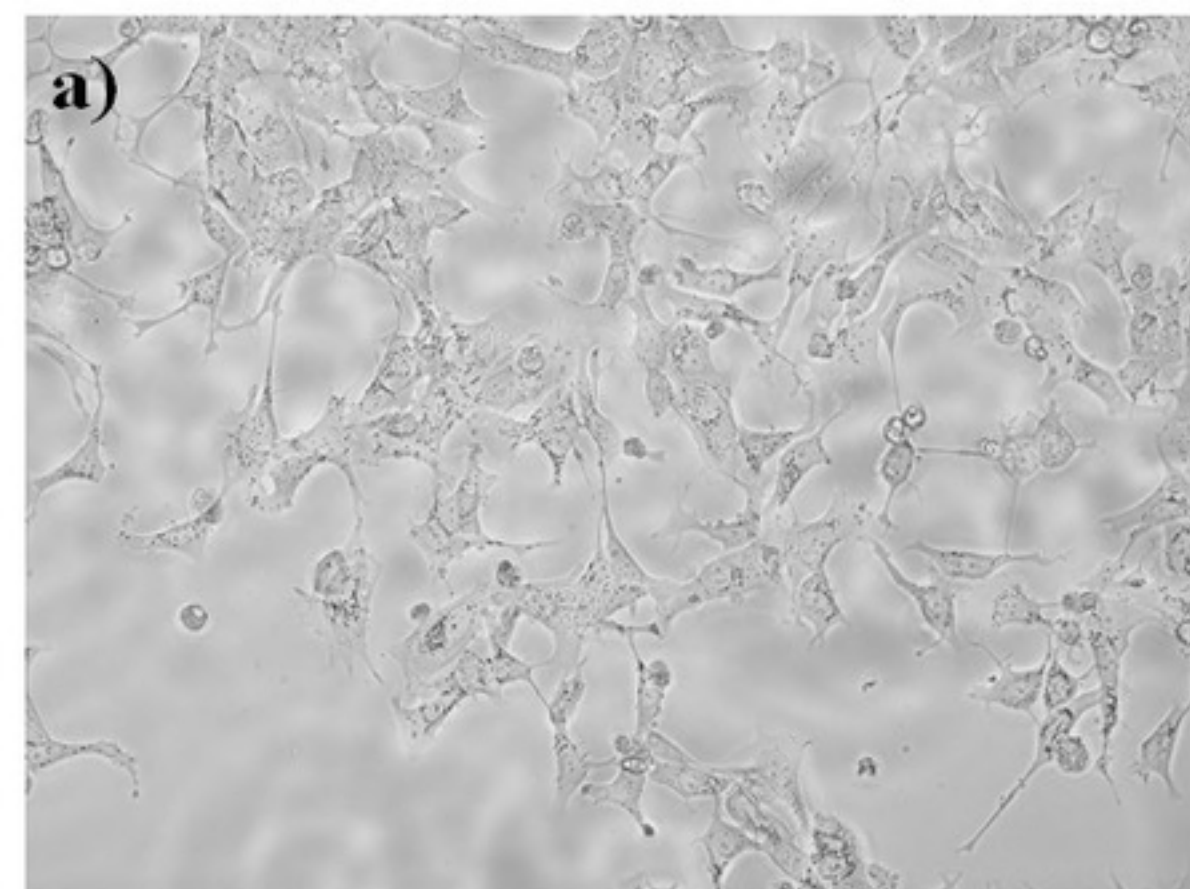


Fig8

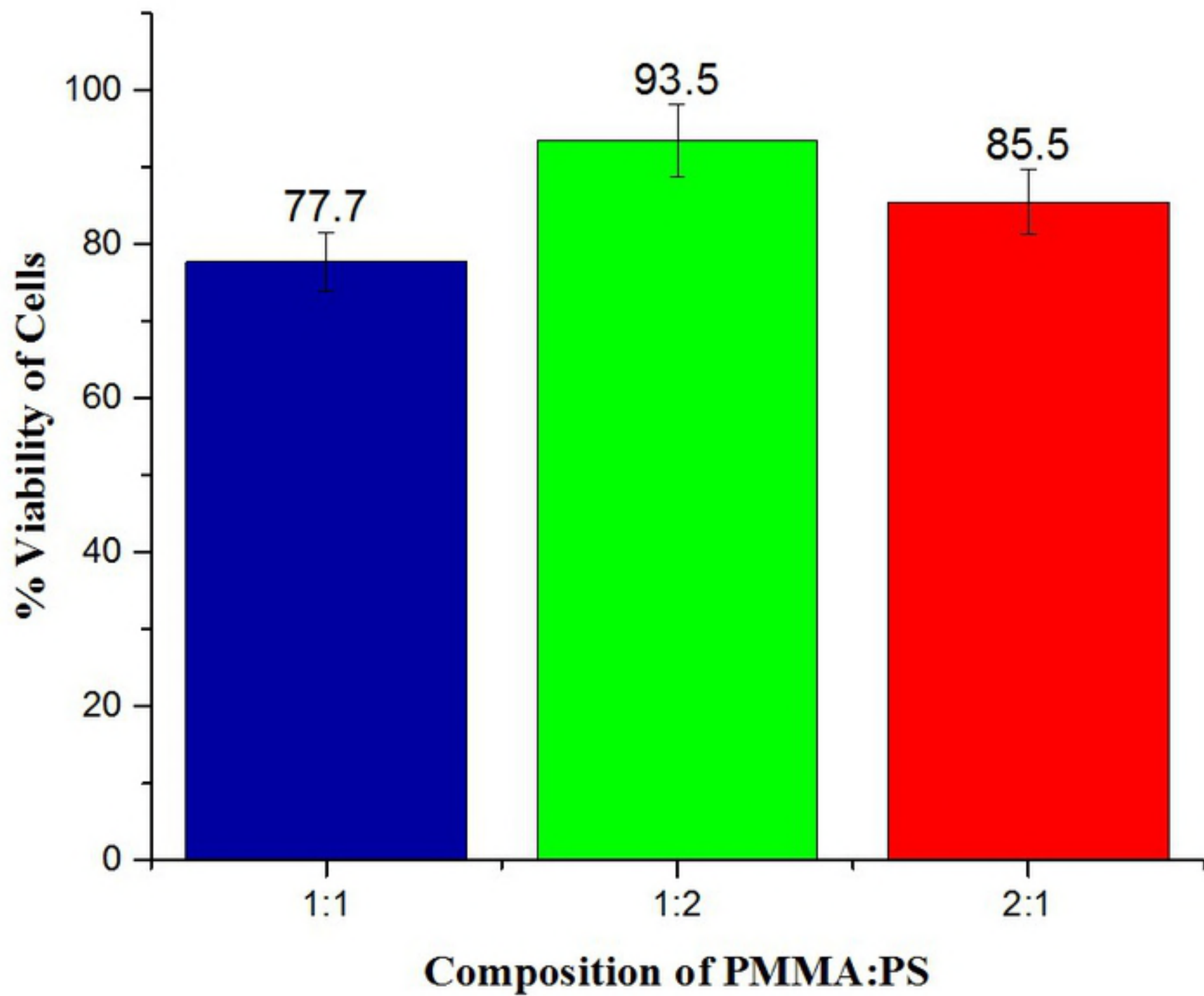


Fig9



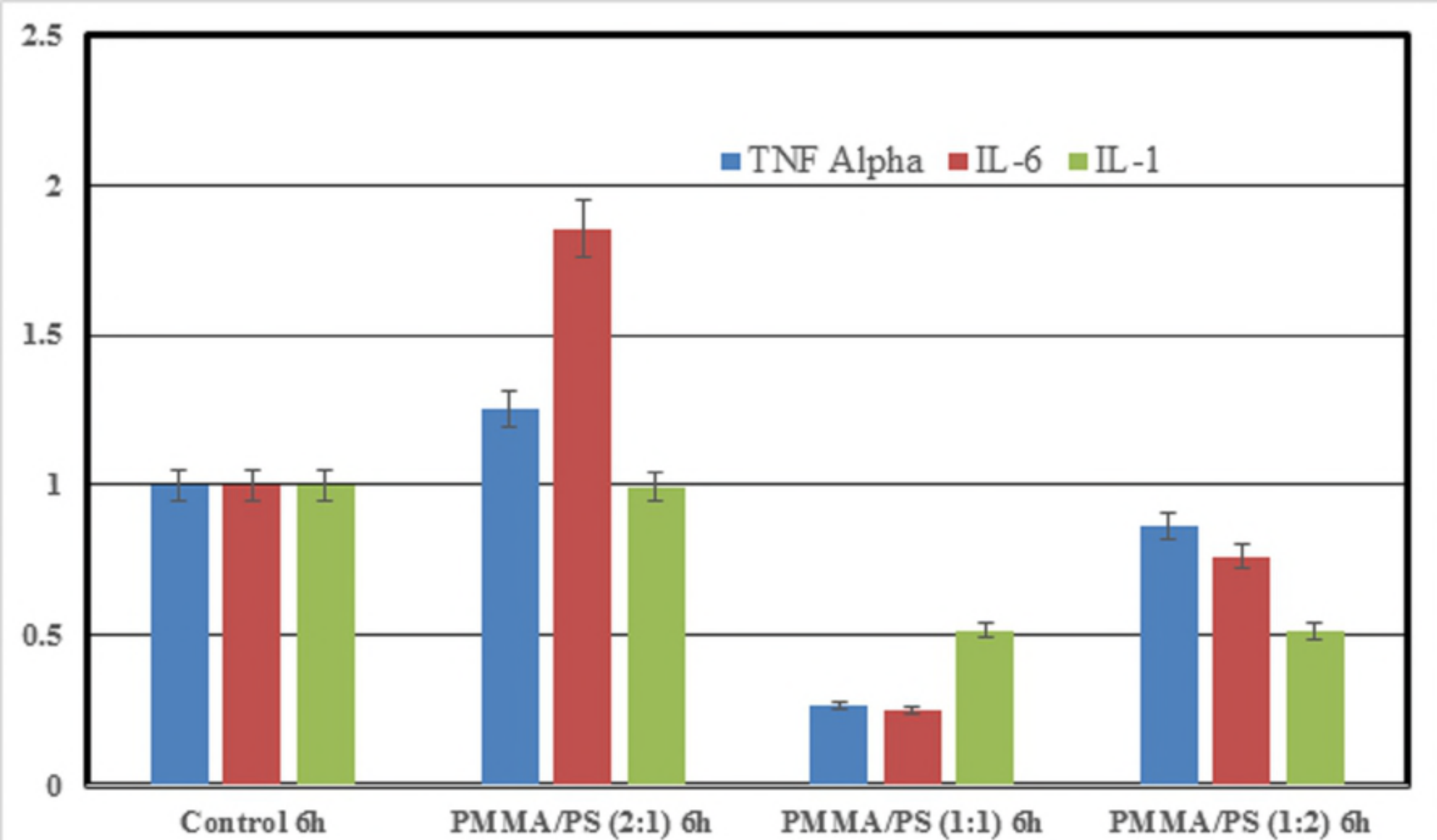


Fig10