Learning of pathogenic bacteria in adult *C. elegans* bidirectionally regulates pathogen

response in the progeny

Ana Pereira[1] and Yun Zhang[1,*]

1. Department of Organismic and Evolutionary Biology, Center for Brain Science, Harvard

University, Cambridge, MA 02138, USA

*: correspondence is addressed to: yzhang@oeb.harvard.edu

Parental experience can generate adaptive changes in the behavioral and physiological traits of the offspring[1-3]. However, the biological properties of this intergenerational regulation and the underlying molecular and cellular mechanism are not well understood. Here, we show that the experience of learning to avoid pathogenic bacteria in *C. elegans* alters the behavioral response to the pathogen in the progeny through the endogenous RNA interference (RNAi) pathway. We previously show that the adult *C. elegans* learns to avoid the smell of pathogenic bacteria, such as the *Pseudomonas aeruginosa* strain PA14, after feeding on the pathogen for a few hours[4,5]. Here, we report that this learning experience can bidirectionally regulate the olfactory response to PA14 in the progeny that are never directly exposed to the pathogen. The olfactory preference for PA14 in these progeny is linearly correlated with the learned avoidance of PA14 in their mothers. If the mothers show strong learning of PA14, their progeny avoid PA14; intriguingly, if the mothers show weak learning of PA14, the progeny prefer PA14, suggesting that the PA14-trained mothers transmit both the negative and positive information of PA14 to their progeny. The intergenerational behavioral effect results from an altered behavioral decision regulated by an olfactory sensorimotor neural circuit. Learning to avoid the pathogen also influences the development of the progeny, which is regulated independently from the behavioral change. Animals mutated for the RRF-3/RNA-directed RNA polymerase, a master regulator for the synthesis of the small interfering RNAs that are maternally inherited or in the soma[6,7], display the normal naive and learned response to PA14 but are defective in regulating the olfactory response to PA14 in their progeny. Our results characterize an intergenerational effect that allows the progeny to rapidly adapt to an environmental condition that is critical for survival.

In many organisms, the parental experience generates adaptive changes in the behavior and the physiology of the progeny[1-3]. However, whether the parental interaction with the environment regulates the same behavioral modalities in the parents and the progeny, indicating the transmission of specific sensory information, is not clear. In addition, the signaling pathways underlying the transmission of the parental information to the progeny are poorly understood. Olfaction plays critical roles in animal behaviors that are essential for survival, such as searching for food and avoiding dangers. While genetically encoded[8,9], the olfactory system is highly adaptable. Experience can profoundly shape the meaning of an odorant to an animal[10,11]. *Caenorhabditis elegans* feeds on bacteria and is often attracted by the smell of the bacteria in its habitat[4,12,13]. Meanwhile, some bacteria are pathogenic. Ingesting the pathogenic

bacteria makes the worm ill through intestinal infection[14]. Previous studies show that after briefly

feeding on certain pathogenic bacteria, such as the *Pseudomonas aeruginosa* strain PA14,

adult *C. elegans* reduces the preference for the smell of the pathogen, even if it is initially

attractive[4,5]. This form of learning is contingent on the pathogenesis of the training bacteria[4,5]

and resembles the Garcia effect, a robust form of conditioned aversion that allows animals to

learn to avoid the smell or taste of a food that makes them ill[15].

Although the intestinal infection caused by ingesting PA14 results in a slow death of the worm

over a course of few days[14], the bacteria in the *Pseudomonas* genus are abundant in the natural

habit of *C. elegans* and serve as common food sources for the worm[13]. Therefore, the

pathogenic *Pseudomonas* PA14 contains the information of food and danger, both of which are

critical for the survival of the worm. We asked whether learning to avoid PA14 regulated the

olfactory response to the smell of PA14 in the offspring. We cultivated the worms under the

standard conditions on the benign bacteria strain *Escherichia Coli* OP50 until the adult stage[16].

We then trained the adult hermaphrodite worms (F0s) by feeding them on PA14 for 8 hours

(Figure 1A). We quantified the preference between the smell of OP50 and the smell of PA14 in

the trained and the naive F0 worms using an automated assay[5]. In this assay, a positive choice

index indicates a preference for PA14 and a negative choice index indicates a preference for

OP50 (Methods)[5]. After randomly sampling the preference in F0s, we harvested the embryos

(F1s) from the trained and the naive F0s with a bleach solution that dissolved the adult bodies

and the associated bacteria. We cultivated the F1 worms on the benign bacteria *E. Coli* OP50

under the standard conditions. Thus, the F1 progeny of the PA14-trained F0s and the F1

progeny of the naive F0s were cultivated under the same condition and neither was directly

exposed to PA14. We quantified the preference of F1s between the smell of OP50 and the

smell of PA14 after they reached the adult stage. We measured the olfactory preference of F1

worms using a two-choice assay, in which a worm chooses between a drop of supernatant of an

OP50 bacteria culture and a drop of supernatant of an PA14 bacteria culture by navigating

towards the drop (Figure 1A). We recorded the behavior of the F1 worms during the two-choice

assay and analyzed their chemotactic movement and their choices. A positive choice index in

the two-choice assay indicates a preference for PA14 and a negative choice index indicates a

preference for OP50 (Figure 1A and Methods). While the automated assay allows us to rapidly

measure the olfactory preference in F0s before handling the F1 embryos, the two-choice assay

allows us to examine the olfactory behavior in F1 worms in detail (Methods).

We found that consistent with our previous findings[4,5], training the F0 adult worms with PA14 for

8 hours strongly reduced the preference for PA14 (Extended Figure 1). This experience-

dependent change is indicated by a learning index (LI), which is the difference between the

choice index (CI) of the naive F0s and the choice index of the trained F0s in the automated

assay (Figure 1B and Methods). A positive LI indicates learned avoidance of PA14 (Methods).

Next, we examined whether the training experience of the F0s generated any difference in the

olfactory preference of their F1 progenies. Thus, we measured the difference between the

choice index of the F1 progeny of the naive F0 worms and the choice index of the F1 progeny of

the trained F0 worms, and defined this difference as the parental-experience dependent

learning index (P_LI) (Figure 1B and Methods). A positive P_LI indicates increased avoidance

of PA14 in F1 induced by the parental experience with PA14 (Methods). We found that P_LI in

F1s were significantly correlated with the LI in their F0 mothers (Figure 1B). On one end of this

spectrum, the more the trained F0s learned to reduce their preference for the PA14 smell, the

more their F1 progeny avoid the PA14 smell in comparison with the F1s of the naive F0s that

were tested in parallel (Figure 1B), indicating that a strong aversive learning of PA14 in the F0s

generates the avoidance of the PA14 smell in F1. Surprisingly, on the other end of the

spectrum, we found that for F0s that only weakly learned to reduce their preference for the

PA14 smell, the less the trained F0s learned to suppress their preference for PA14, the more

their F1s preferred the PA14 smell in comparison with the F1s of the naive F0s tested in parallel

(Figure 1B), suggesting that the F0s that weakly learned transmit the beneficial information of

PA14. These results show that the maternal learning experience with PA14 bidirectionally

regulates the olfactory response to PA14 in the progeny.

Next, we further examined the bidirectional effect of the F0's learning experience on the

olfactory choice of F1s. We separated all the experiments into two groups, the experiments in

which F0s displayed a high level of learning of PA14 and the experiments in which F0s

displayed a low level of learning of PA14 (Methods). We found that the F1 progeny of the F0s

with strong learning avoided PA14 in the two-choice assay, indicated by their negative choice

indexes (P_CI, Figure 1C). In contrast, the F1 progeny of the F0s with weak learning preferred

PA14 in the two-choice assay, indicated by their positive choice indexes (P_CI, Figure 1C). The

extent of the F0 mothers' learning significantly regulated the preference in their F1 progeny.

This regulatory effect is also evident in a significant correlation between the learning indexes of

F0s (LIs) and the choice indexes of the F1s (P_CIs) of the trained F0s (Figure 1E). In

comparison, the choice indexes of the F1s (P_CIs) of the naive F0 mothers in the experiments

where training generated strong aversive learning were similar to the P_CIs of the F1s of the

naive F0 mothers in the experiments where training generated weak aversive learning (Figure

1D). Consistently, there is no correlation between the choice indexes of the F1s (P_CI) of the

naive F0s with the learning indexes of the F0 generation (LIs) (Figure 1F). Thus, it is

conceivable that the parental experience with PA14 signals two types of information, aversive

information such as pathogenesis and appetitive information such as food, and the weight of

these information transmitted to the progeny depends on the degree of the parental learning.

To examine whether the difference in the olfactory response to PA14 in the F1 generation

passed onto the F2s, we isolated the F2 worms from the F1 mothers and cultivated F2s under

the standard conditions on *E. coli* OP50. Similarly, we quantified the parental-experience

dependent learning index (P_LI) in F2s, which was defined as the difference between the choice

index of the F2s of the naive F0 worms and the choice index of the F2s of the trained F0 worms.

We found that the P_LIs of the F2s did not depend on the P_LIs of their F1 mothers (Figure

1G). These results together indicate that the parental experience with PA14 only regulates the

olfactory response of their first-generation progeny, leaving the flexibility for F2s to respond to

further changes in the environmental conditions.

To further characterize the parental-experience dependent olfactory learning in F1s, we asked

what behavioral changes gave rise to their altered olfactory choice. We analyzed the movement

of the F1s when they navigated toward OP50 or PA14 during the two-choice assay (Figure 2A).

We measured the navigation index, which is defined as the ratio of the radial speed towards the

target and the actual traveling speed to quantify the efficiency of the chemotactic movement

towards an odorant source[17](Figure 2B). We divided the assay plate in sections along the axis

of the two drops of the bacteria cultures that were 2cm apart and quantified the navigation index

for each section in each worm. We analyzed the median value of the navigation indexes for all

the worms in each section as a function of the distance away from the target. We found that the

navigation index of the F1s of the trained F0s increased when the animals approached the

target, indicated by a significant correlation between the navigation index and the distance from

the target regardless of whether the worms chose OP50 or PA14 (Figure 2B); in contrast, the

navigation index of the F1 worms of the naive F0s did not correlate with the distance to the

target (Figure 2B). We also measured the locomotory speed, the distance that the worms

travelled and the time spent on travelling before reaching the target, the rate of reorienting

movements, the reversals and the large body bends, and did not find any difference between

the F1 worms of the trained F0s and the F1 worms of the naive ones (Figure 2C, 2D and

Extended Figure 2A-2C). These results identify different strategies for chemotactic movements

used by the F1 progeny of the trained mothers versus the F1 progeny of the naive mothers.

Previous studies show that the sensorimotor integration of a cholinergic neural circuit regulates

the efficiency of chemotactic movements towards odorants, indicated by the navigation index[17].

These results together identify the neuronal substrates for the parental-experience dependent

olfactory plasticity in the F1s.

In addition, we asked whether the parental experience with PA14 regulated other physiological

traits of the F1s. We first examined the immune resistance to PA14 in F1s using a slow-killing

assay that measures the survival rate over time in worms cultivated on a lawn of PA14[14]. We

used the *nsy-1(k397)* animals as a control, because *nsy-1(k397)* is mutated for the worm

homolog of MAPKKK and displays significantly compromised resistance to the infection of

PA14[18]. We found that while the *nsy-1(k397)* animals showed a much reduced survival rate, the

F1s of the PA14-trained F0s and the F1s of the naive F0s showed similar survival rates (Figure

2E), indicating that training the adult worms with PA14 do not alter the resistance to PA14 in

their progeny. However, we noticed that the F1s of the trained mothers were larger in the body

size than the F1s of the naive F0 mothers (Figure 2F). To characterize this difference, we

examined the F1 embryos inside the F0s. While the trained F0 mothers and the naive F0

mothers contained the same amount of embryos in their bodies on average (trained F0s: 12 ±

0.8 eggs; naive F0s, 11 ± 0.7 eggs; n = 7 adults each, mean ± sem, two-tailed Student's t-test, p

= 0.3), the F1 embryos inside the trained F0s were more advanced in the developmental stage

than the F1 embryos inside the naive F0s (Extended Figure 2D). In addition, the F1s of the

trained F0 mothers entered the L4-larval stage, a developmental stage with distinct anatomical

features[19], sooner than the F1s of the naive F0 mothers (Figure 2G). Thus, the parental

experience with PA14 influences the development of the offspring. Notably, there is no

correlation between the body size of the F1 progeny of the trained mothers with either the

learning index of the mothers (LI) or the olfactory choice of the F1s (P_CI) (Figure 2H and 2I),

indicating that the parental-experience dependent changes in the olfactory choice of the F1s

and in the development of the F1s are independently regulated.

Next, we sought the signaling mechanisms whereby the information of the parental experience

is transmitted to the progeny. In *C. elegans,* the endogenous RNA interference pathway can

respond to environmental conditions to generate small interfering RNAs (endo-siRNAs) that

transmit the parental experience cross generations and to regulate the physiology of the

offspring[20]. We tested the mutant animals that contained a deletion in *rrf-3,* which encodes a

RNA-directed RNA Polymerase(RdRP)[21] that is needed for the biogenesis of endo-siRNAs that

are maternally inherited or found in the soma[6,7]. We found that the *rrf-3(pk1426)* mutant animals

display normal naive and learned olfactory preference for PA14 in the F0 generation (Figure

3A), but are significantly defective in the parental-experience dependent learning in F1s (Figure

3B). While the parental-experience dependent learning index (P_LI) in F1s significantly

correlated with the learning index (LI) in F0s in wild-type animals, this correlation was

completely abolished in the *rrf-3(pk1426)* mutants (Figure 3B). Thus, the pathway of the endo-

siRNAs regulates the transmission of the parental experience to the progeny.

We were intrigued by the bidirectional effect of the parental experience with PA14 on the F1's

response to PA14. Previous studies show that although adult worms reduce their preference for

the smell of PA14 after training, they still prefer the PA14 smell more than non-food smells[5],

indicating that PA14 represents as a less preferred food source to the trained F0s. Thus, we

tested the possibility that training F0s with a shorter duration, which reduces the infection of

PA14 to the F0s, induces a preference for PA14 in the F1s. Interestingly, we found that

shortening the training of adult F0s to 4 hours generated aversive learning of PA14 in F0s

(Figure 3C) and induced a significant increase in the preference for PA14 in the F1s of the

trained F0s in comparison with the F1s of the naive F0s (Figure 3D). Furthermore, we found that

although the *rrf-3(pk1426)* F0s were normal in learning of PA14 after 4-hour training (Figure

3C), their F1s were defective in the induced preference for PA14, indicated by the similar

preference displayed by the *rrf-3(pk1426)* F1s of the trained and the naive mothers (Figure 3D).

These results further demonstrate that the parental experience can generate the appetitive and

the aversive effects on the olfactory response of PA14 in the progeny and that both of the

intergenerational effects are regulated by the endogenous RNAi pathway.

Previously, we show that the adult *C. elegans* learns to avoid the smell of pathogenic bacteria,

such as the *Pseudomonas aeruginosa* strain PA14, after feeding on the pathogen for 4-8

hours[4,5]. Meanwhile, it is shown that exposing the L1-stage larvae worms to PA14 generates the

avoidance of PA14 when the animals reach the adult stage and the imprinting of PA14 requires

12-hour training of the L1 larvae[22]. In this study we identify a new form of adaptive behavior

where briefly training the adult worms with PA14 alters the olfactory response to PA14 in their

progeny. Together, these results reveal versatile behavioral responses that *C. elegans* uses to

interact with pathogenic bacteria that the worm frequently encounters in its habitat.

Unexpectedly, the experience with PA14 in the adults can regulate the olfactory response of

their progeny bidirectionally. While the strong learning of the adult mothers leads to the

avoidance of PA14 in the progeny, weak learning of the mothers induces attractive response to

PA14 in their progeny. We found that training the mothers for 4 hours, the minimal amount of

training needed to induce significant aversive learning of PA14 in the adults[5], significantly

increases the progeny's preference for the smell of PA14, suggesting the mother-to-offspring

transmission of beneficial information of PA14. The smell of PA14 is attractive to the worms that

never ingest PA14; even after training, the PA14-trained adult worms that display a reduced

preference for the PA14 smell still prefer the smell of PA14 more than the non-food odorants[5].

The bacteria of the *Pseudomonas* genus are highly abundant in the habits of *C. elegans* with

many isolates being beneficial for the development and the reproduction of the worm[13]. Thus, it

is conceivable that the parental experience transmits information of food and pathogenesis of

PA14 to the progeny and the resulting behavioral effect is regulated by the extent of the

experience.

In many animals, the parental experience with the environmental conditions, especially those

associated with food and danger, regulates the development, physiology and behavior of the

offspring and the effect can last for various amounts of time[1-3,23-29]. In *C. elegans,* the insulin-like

signaling acts in the soma or the germline of the mothers to regulate the development and the

stress responses in response to the parental experience with food deprivation or osmotic

stresses, respectively[26,27]. However, in most cases, especially when the parental experience

generates adaptive changes in the same behavioral modality in both the mothers and the

progeny, the signaling mechanism underlying the transmission of the parental experience is

largely unknown. In *C. elegans,* it is shown that the pathways of the endogenous small

interfering RNAs (endo-siRNAs) can regulate the gene expression and the function of the

nervous system in response to the experience within the same generation[30,31]. In addition, it is

shown that the production of endo-siRNAs is regulated by prolonged starvation and the altered

siRNA profile persists for several generations[20]. In this study, we show that the RRF-3/RdRP[21],

which is critically required for the biogenesis of endo-siRNAs that are produced in the soma or

maternally inherited[6,7], specifically regulates the parental-experience dependent changes in the

olfactory response to the pathogenic bacteria PA14, revealing a new function of the endo-

siRNAs in regulating intergenerational adaptive behavioral responses.

Methods

Strains

C. elegans hermaphrodites were used in this study and cultivated using the standard

conditions[16]. The strains used in this study include: N2 (Bristol), ZC2834 *rrf-3(pk1426)* II and

CX4998 kyIs 140 I; *nsy-1(k397)* II.

The aversive olfactory training with PA14 in F0s and the cultivation of the F1s

The aversive olfactory training was preformed mainly as previously described[4] with minor

modifications. To prepare the training plates, individual OP50 colonies or PA14 colonies were

used to inoculate 50 mL nematode growth medium (NGM, 2.5g/L Bacto Peptone, 3.0g/L NaCl,

1mM CaCl2, 1mM MgSO4, 25mM KPO4 pH6.0), which were cultivated at 27ºc for overnight.

800 mL (for 4h training protocol) or 400 mL (for 8h training protocol) OP50 or PA14 culture was

spread onto each NGM plate, which was incubated at 27ºc for 2 days to prepare the naive

control and the training plates, respectively. To preform training, the first-day adult

hermaphrodites cultivated under the standard conditions with OP50 as food were transferred to

the control or the training plate, respectively, and kept at 20ºc for 4 or 8 hours as specifically

described. By the end of the training, some of the naive control and the trained F0 worms were

randomly picked to measure the olfactory choice and learning in the automated assay and the

rest of the F0s worms were collected from the plates with S-basal medium and passed through

a cell strainer 40 micrometer nylon filter (Falcon) to remove laid eggs. Worms were then treated

with a bleach solution to isolate the F1 embryos, which were hatched and cultivated under the

standard conditions until the adult stage.

Olfactory preference assay using the automated olfactory assay

The automated assay that quantifies the olfactory preference in individual animals were

conducted as previously described[5]. Briefly, by the end of the training, the naive and trained F0s

were washed with buffer and individually placed in the droplets of 2 μL NMG buffer (1mM

CaCl2, 1mM MgSO4, 25mM KPO4 pH6.0) in an enclosed chamber and subjected to alternating

airstreams that were saturated with the smell of OP50 or PA14 by blowing clean air through

freshly generated bacterial cultures. Each olfactory stimulation lasted for 30 second and each

assay contains 12 cycles of stimulation. The locomotion of the worms is video recorded and the

large body bends of the tested worms were identified with machine-vision softwares. Because

large body bends are followed by reorientation, a higher rate of the large body bends indicates a

lower preference to the tested airstream[5,32]. The choice index (CI) is defined as the body bends

evoked by OP50 smell minus the body bends evoked by PA14 smell and normalized by the total

body bends [CI = (bends evoked by OP50 - bends evoked by PA14)/(bends evoked by OP50 +

bends evoked by PA14)] and the learning index (LI) is defined as the CI of the naive worms

minus CI of the trained worms (LI = CI of naive worms – CI of trained worms). A positive LI

indicates learned avoidance of PA14.

Olfactory preference assay using the two-choice assay

The two-choice assay is similar to the one described in Zhang et al., 2005, except for several

modifications. To measure the olfactory preference for bacteria, a drop of 5 μL supernatant of

OP50 culture and a drop of 5 μL PA14 culture were put 2cm apart on a 6 cm NMG plate. In

each assay, one worm was placed on a plate equidistant to the two drops of the bacteria culture

supernatant and allowed to crawl to the preferred stimulus. The movement and the choice of

each worm were recorded and later analyzed. The worms that did not make choice after 10

minutes are counted in the total number. The parental-experience dependent choice index

(P_CI) in the two-choice assay is defined as the number of worms that choose PA14 minus the

number of worms that choose OP50 normalized by the total number of worms tested [P_CI =

(number of worms that choose PA14 - number of worms that choose OP50)/total number of

worms] and the parental-experience dependent learning index (P_LI) for the two-choice assay is

defined as the P_CI of F1s of naive F0s minus P_CI of F1s of trained F0s (P_LI = P_CI of F1s

of naive F0s – P_CI of F1s of trained F0s). A positive P_LI indicates increased avoidance of

PA14 in F1 induced by the parental experience with PA14.

Slow killing assay

The slow killing assay was performed essentially as previously described[14]. 200 μL freshly

prepared PA14 culture incubated at 27ºc in the Luria-Bertani (LB) medium overnight were

spread into a 4cm diameter circle on a 6cm NMG plate and incubated at 37ºc for 24 hours and

then left at room temperature for another 24 hours before the assay. 20 F1 young adult

hermaphrodites cultivated under the standard conditions (Figure 1A) were transferred onto each

slow killing plate and kept at 20ºc. The dead and the alive worms were counted at the specific

time points as shown in Figure 2 over a course of 50 hours.

The analysis of parental-experience dependent learning index

To separate the experiments in which F0s exhibited high learning indexes from the experiments in which F0s exhibited low learning indexes, we tested for a significant linear correlation between the learning index (LI) of F0s and the parental-experience dependent learning index of F1s (P_LI). We found that these 2 variables are linearly correlated as shown in Figure 1B. We then used the point where the line of linear regression intersects with the x-axis (LI of F0) to separate the experiments into two groups. The experiments in which the LI of F0 is larger than the point of intersection are included in the group in which F0s exhibited a high level of PA14 learning; conversely, the experiments in which the LI of F0 is smaller than the point of intersection are included in the group in which F0s exhibited a low level of PA14 learning.

Quantification of the body size and the chemotactic movements

The body size and the parameters of the chemotactic movements in the two-choice assays were quantified by analyzing the recorded worms with the Wormlab tracker (https://www.mbfbioscience.com/wormlab).

Statistics

The statistical methods, sample size and the number of the replicates are indicated in the legend of each figure.

Data Availability

All the data are available upon request. Figure 1-3 and Extended Figure 1 and 2 are generated based on raw data.

Code availability

The method used for the automated assay has been previously published[5] and the code is freely available upon request. The software used to analyze F1 chemotaxis is available at https://www.mbfbioscience.com/wormlab.

Acknowledgements

We thank *Caenorhabditis Genetics Center*, which is funded by National Institutes of Health - Office of Research Infrastructure Programs (P40 OD010440), for strains. Y.Z. is funded by National Institutes of Health.

Author contributions

A.P. and Y.Z. designed the experiments, interpreted the results and wrote the manuscript. A.P.

performed the experiments and analyzed the data.

Competing Interests

The authors declare no competing interests.

References

1 Gluckman, P. D. & Hanson, M. A. Living with the past: evolution, development, and
patterns of disease. *Science* **305**, 1733-1736, doi:10.1126/science.1095292 (2004).
2 Mousseau, T. A. & Fox, C. W. The adaptive significance of maternal effects. *Trends Ecol*
Evol **13**, 403-407 (1998).
3 Galloway, L. F. & Etterson, J. R. Transgenerational plasticity is adaptive in the wild.
Science **318**, 1134-1136, doi:10.1126/science.1148766 (2007).
4 Zhang, Y., Lu, H. & Bargmann, C. I. Pathogenic bacteria induce aversive olfactory
learning in Caenorhabditis elegans. *Nature* **438**, 179-184, doi:10.1038/nature04216
(2005).
5 Ha, H. I. *et al.* Functional organization of a neural network for aversive olfactory learning
in Caenorhabditis elegans. *Neuron* **68**, 1173-1186, doi:10.1016/j.neuron.2010.11.025
(2010).
6 Han, T. *et al.* 26G endo-siRNAs regulate spermatogenic and zygotic gene expression in
Caenorhabditis elegans. *Proc Natl Acad Sci U S A* **106**, 18674-18679,
doi:10.1073/pnas.0906378106 (2009).
7 Gent, J. I. *et al.* Distinct phases of siRNA synthesis in an endogenous RNAi pathway in
C. elegans soma. *Mol Cell* **37**, 679-689, doi:10.1016/j.molcel.2010.01.012 (2010).
8 Su, C. Y., Menuz, K. & Carlson, J. R. Olfactory perception: receptors, cells, and circuits.
Cell **139**, 45-59, doi:10.1016/j.cell.2009.09.015 (2009).
9 Keller, A. & Vosshall, L. B. Better smelling through genetics: mammalian odor
perception. *Curr Opin Neurobiol* **18**, 364-369, doi:10.1016/j.conb.2008.09.020 (2008).
10 Keene, A. C. & Waddell, S. Drosophila olfactory memory: single genes to complex
neural circuits. *Nat Rev Neurosci* **8**, 341-354, doi:10.1038/nrn2098 (2007).
11 McIntyre, J. C., Thiebaud, N., McGann, J. P., Komiyama, T. & Rothermel, M.
Neuromodulation in Chemosensory Pathways. *Chem Senses* **42**, 375-379,
doi:10.1093/chemse/bjx014 (2017).
12 Bargmann, C. I., Hartwieg, E. & Horvitz, H. R. Odorant-selective genes and neurons
mediate olfaction in C. elegans. *Cell* **74**, 515-527 (1993).
13 Samuel, B. S., Rowedder, H., Braendle, C., Felix, M. A. & Ruvkun, G. Caenorhabditis
elegans responses to bacteria from its natural habitats. *Proc Natl Acad Sci U S A* **113**,
E3941-3949, doi:10.1073/pnas.1607183113 (2016).
14 Tan, M. W., Mahajan-Miklos, S. & Ausubel, F. M. Killing of Caenorhabditis elegans by
Pseudomonas aeruginosa used to model mammalian bacterial pathogenesis. *Proc Natl*
Acad Sci U S A **96**, 715-720 (1999).
15 Garcia, J., Hankins, W. G. & Rusiniak, K. W. Behavioral regulation of the milieu interne
in man and rat. *Science* **185**, 824-831 (1974).
16 Brenner, S. The genetics of Caenorhabditis elegans. *Genetics* **77**, 71-94 (1974).
17 Liu, H. *et al.* Cholinergic Sensorimotor Integration Regulates Olfactory Steering. *Neuron*
97, 390-405 e393, doi:10.1016/j.neuron.2017.12.003 (2018).

18 Kim, D. H. *et al.* A conserved p38 MAP kinase pathway in Caenorhabditis elegans innate
immunity. *Science* **297**, 623-626, doi:10.1126/science.1073759 (2002).
19 Yochem, J. K. Nomarski images for learning the anatomy, with tips for mosaic analysis.
WormBook, 1-47, doi:10.1895/wormbook.1.100.1 (2006).
20 Rechavi, O. *et al.* Starvation-induced transgenerational inheritance of small RNAs in C.
elegans. *Cell* **158**, 277-287, doi:10.1016/j.cell.2014.06.020 (2014).
21 Simmer, F. *et al.* Loss of the putative RNA-directed RNA polymerase RRF-3 makes C.
elegans hypersensitive to RNAi. *Curr Biol* **12**, 1317-1319 (2002).
22 Jin, X., Pokala, N. & Bargmann, C. I. Distinct Circuits for the Formation and Retrieval of
an Imprinted Olfactory Memory. *Cell* **164**, 632-643, doi:10.1016/j.cell.2016.01.007
(2016).
23 Dantzer, B. *et al.* Density triggers maternal hormones that increase adaptive offspring
growth in a wild mammal. *Science* **340**, 1215-1217, doi:10.1126/science.1235765
(2013).
24 Dias, B. G. & Ressler, K. J. Parental olfactory experience influences behavior and neural
structure in subsequent generations. *Nat Neurosci* **17**, 89-96, doi:10.1038/nn.3594
(2014).
25 Ost, A. *et al.* Paternal diet defines offspring chromatin state and intergenerational
obesity. *Cell* **159**, 1352-1364, doi:10.1016/j.cell.2014.11.005 (2014).
26 Hibshman, J. D., Hung, A. & Baugh, L. R. Maternal Diet and Insulin-Like Signaling
Control Intergenerational Plasticity of Progeny Size and Starvation Resistance. *PLoS*
Genet **12**, e1006396, doi:10.1371/journal.pgen.1006396 (2016).
27 Burton, N. O. *et al.* Insulin-like signalling to the maternal germline controls progeny
response to osmotic stress. *Nat Cell Biol* **19**, 252-257, doi:10.1038/ncb3470 (2017).
28 Bestion, E., Teyssier, A., Aubret, F., Clobert, J. & Cote, J. Maternal exposure to predator
scents: offspring phenotypic adjustment and dispersal. *Proc Biol Sci* **281**,
doi:10.1098/rspb.2014.0701 (2014).
29 Harvey, S. C. & Orbidans, H. E. All eggs are not equal: the maternal environment affects
progeny reproduction and developmental fate in Caenorhabditis elegans. *PLoS One* **6**,
e25840, doi:10.1371/journal.pone.0025840 (2011).
30 Juang, B. T. *et al.* Endogenous nuclear RNAi mediates behavioral adaptation to odor.
Cell **154**, 1010-1022, doi:10.1016/j.cell.2013.08.006 (2013).
31 Sims, J. R. *et al.* Developmental programming modulates olfactory behavior in C.
elegans via endogenous RNAi pathways. *Elife* **5**, doi:10.7554/eLife.11642 (2016).
32 Pierce-Shimomura, J. T., Morse, T. M. & Lockery, S. R. The fundamental role of
pirouettes in Caenorhabditis elegans chemotaxis. *J Neurosci* **19**, 9557-9569 (1999).
432
433

Figure 1

Figure 1. Training with PA14 bidirectionally regulates the olfactory response to PA14 in the progeny.

(A) Schematics for the parental-experience dependent olfactory learning.

(B) Parental-experience dependent olfactory learning indexes (P_LI) in F1s are linearly correlated with the aversive olfactory learning indexes (LI) of the F0 mothers.
Choice index of F0 (CI) = (bends evoked by OP50 - bends evoked by PA14)/(bends evoked by OP50 + bends evoked by PA14);
LI = CI of naive F0s – CI of trained F0s;
Choice index of F1 (P_CI) = (number of worms that choose PA14 - number of worms that choose OP50)/total number of worms.
P_LI = P_CI of F1s of naive F0s – P_CI of F1s of trained F0s;
n = 12 independent experiments that contained 12 independent learning assays for F0s (274 animals) and 12 independent two-choice assays where 186 F1 worms were individually tested, p value denotes the significance of the linear correlation coefficient.

(C, D) The F1s of the trained F0 mothers in experiments in which F0s exhibited a high level of aversive learning of PA14 avoided PA14, indicated by a negative choice index (P_CI), and the F1s of the trained F0 mothers in experiments in which F0s exhibited a low level of aversive learning of PA14 preferred PA14, indicated by a positive choice index (P_CI) **(C)**; in contrast, the F1s of the naive F0s tested in the same sets of experiments as shown in **C**

where F0s exhibited a high level or a low level of learning had similar choice indexes (P_CI)
(D). n = 6 experiments for high F0 learning and 6 experiments for low F0 learning, where 95
(high F0 learning) and 91 (low F0 learning) F1 worms individually tested in the two-choice
assay for each group; Student's t- test (data are normally distributed), ** p = 0.004 for **C** and
p = 0.72 for **D**; Mean ± SEM.
(E, F) The choice indexes of the F1s (P_CI) of the trained F0s are inversely correlated with the
learning indexes (LI) of their trained F0 mothers **(E)**; in contrast, the choice indexes of the
F1s (P_CI) of the naive F0s have no correlation with the learning indexes (LI) of their naive
F0 mothers **(F).** A positive or a negative P_CI respectively indicates a preference for PA14
or OP50, n = 12 learning assays for F0s (n = 274 animals) and n = 12 two-choice assays
where 186 F1 worms were individually tested, P values denote the significance of the linear
correlation coefficients.
(G) Parental-experience dependent olfactory learning indexes (P-LI) in F2s is not significantly
correlated with the P-LI of the F1s.
P-LI = choice index of F1s (or F2) of naive F0s – choice index of F1s (or F2) of trained F0s;
choice index of F1 (or F2) = (number of worms that choose PA14 - number of worms that
choose OP50)/total number of animals. n = 6 independent experiments that included 91 F1
worms and 88 F2 worms individually tested in the two-choice assays, P values denote the
significance of the linear correlation coefficient.
474

Figure 2

Figure 2. The parental experience with PA14 regulates the olfactory behavior and the development of the F1 progeny.

(A) Schematics showing the trajectory of a worm in the two-choice assay.

(B) The navigation indexes in the F1s of the trained F0s inversely and linearly correlate with the distance between the navigating F1s and the target drop of bacteria culture, regardless of the chosen bacteria; in contrast, the navigation indexes in the F1s of the naive F0 mothers do not have the similar correlations. The median value of the navigation indexes for each section was used to analyze linear regression; n = 40 F1s of naive F0 (OPchoice), n = 47 F1s of naive F0 (PAchoice), n = 42 F1s of trained F0 (OPchoice), n = 47 F1s of trained F0 (PAchoice). Data was not analyzed for worms that didn't made a choice at the end of the trial. Data is missing for 7 worms due to problems during recordings.

(C, D) The F1s of the trained F0s and the F1s of the naive F0s do not differ in the locomotory

speeds during the chemotactic movement towards bacteria cultures **(C)** and traveled similar

distances before reaching the target **(D)**. The box plots show the median, the first and the

third quantile,1.5 IQR and the outliers; the number of worms for each group is the same as

in **B**, Wilcoxon rank-sum test with correction for multiple comparison (the data is not

normally distributed).

(E) In the slow killing assay, the wild-type (WT) F1s of the trained F0s and the wild-type F1s of

the naive F0s did not differ in their survival rates (p = 0.104), while the *nsy-1* mutant animals

were significantly weaker in resisting PA14 infection (p < 0.0001). Two independent

experiments generated the same conclusion and one experiment containing two replicates

(n = 20 animal each plate) was presented, Mantel-Cox log rank test.

(F, G) The F1s of the trained F0s are larger in the body size than the F1s of the naive F0s [**F,**

the box plots show the median, the first and the third quantile, 1.5 IQR and outliers; n = 87

F1s of naive F0 and n = 89 F1s of trained F0, Wilcoxon rank-sum test with correction for

multiple comparison (the data is not normally distributed)] and contain a higher percentage

of the L4-stage larvae after developing from the embryos extracted from F0s for the same

amount of time (**G,** n = 5 experiments with 10 animals for each group in each experiment,

Mantel-Cox log rank test).

(H, I) The body size of the F1s of the trained F0s do not correlate with the learning index (LI) of

their F0 mothers (**H**) or their own choice indexes (P_CI) (**I**). n = 12 experiments in which the

learning index of F0s and the choice index of F1s are measured and n = 89 F1 worms were

measured for the body size.

512
513

Figure 3

514
515
Figure 3. The RNA dependent RNA polymerase RRF-3 regulates the parental experience-
dependent olfactory learning of PA14.
(A, B) Training with PA14 for 8 hours significantly decreased the preference for the smell of
PA14, which is indicated by a positive choice index, in the F0 wild-type (WT) and the F0 *rrf-*
3(pk1426) mutant animals (**A**, n = 6 learning assays for wild type (143 animals) and the *rrf-3*
mutant (144 animals); two-tailed Student's t-test, *** p < 0.001 and * p < 0.05, Mean ± SEM);
however, while wild-type animals displayed a strong correlation between the parental-
experience dependent learning index (P_LI) of F1s with the learning index (LI) of F0s, the
mutation in the *rrf-3(pk1426)* animals abolished the correlation [**B**, n = 6 learning assays for
F0 (143 WT animals and the 144 *rrf-3* animals) and n = 6 two-choice assays where 88 WT
F1s and 92 *rrf-3* F1s were individually tested; P values denote the significance of the linear
correlation coefficients].
(C, D) Training with PA14 for 4 hours induced similar aversive learning of PA14 in the F0 wild-
type (WT) and the F0 *rrf-3(pk1426)* [**C**, n = 3 learning assays for WT (36 animals) and *rrf-3*
(36 animals), p = 0.71]; however, while the wild-type F1 of the trained F0s displayed a
significantly higher preference for PA14 than the wild-type F1s of the naive F0s in the two-
choice assay, the *rrf-3(pk1426)* F1s of the trained F0s and the *rrf-3(pk1426)* F1s of the naive
F0s displayed similar preference (**D**, n = 4 two-choice assays where 48 WT F1s and 51 *rrf-3*
F1s were individually tested, * p = 0.0286 for WT and p = 0.91 for *rrf-3*). Wilcoxon rank-sum
test because the data in **C** and **D** are not normally distributed, the box plots show the
median, the first and the third quantile, 1.5 IQR.
537

Extended Figure 1

Extended Figure 1. Training with PA14 for 8 hours strongly reduced the preference for
the smell of PA14. The preference of the naive and trained F0 worms were measured in the
automated assay. Choice index of F0 (CI) = (bends evoked by OP50 - bends evoked by
PA14)/(bends evoked by OP50 + bends evoked by PA14); therefore, a positive choice index
indicates a preference for the PA14 smell. n = 12 experiments (274 animals), two-tailed
Student's t-test, Mean ± SEM, *** p < 0.001.
546

Extended Figure 2

■ F1 of naive F0 (OPchoice)　　■ F1 of naive F0 (PAchoice)　　■ F1 of trained F0 (OPchoice)　　■ F1 of trained F0 (PAchoice)

549
550

Extended Figure 2. The parental experience with PA14 regulates the olfactory behavior and the development of the F1 progeny.

(A, B, C) The F1s of the trained F0s and the F1s of the naive F0s do not differ in the rate of reversals (**A**) or omega bends (**B,** the large body bends resembling the Greek letter Ω that lead to reorientation) during chemotactic movements towards a drop of bacteria culture in the two-choice assay, or in the time needed to reach the target **(C)**. The box plots show the median, the first and the third quantile, 1.5 IQR and outliers; the number of worms for each group is the same as stated for Figure **2B**, Wilcoxon rank-sum test with correction for multiple comparison was used because the data is not normally distributed.

(D) The eggs inside the bodies of the trained F0s are older than the eggs inside the naive F0s. The number of cells in each egg was counted. The box plots show the median, the first and the third quantile, 1.5 IQR and outliers; all the eggs in n = 7 worms per treatment were counted, Wilcoxon rank-sum test with correction for multiple comparison was used because the data is not normally distributed, ** $p < 0.01$.

565