Title:

Low but genetically variable male remating ability in a tropical *Drosophila* despite

substantial fitness benefits

Authors:

Andrew D. Saxon, Natalie E. Jones, Eleanor K. O'Brien and Jon R. Bridle[1]

Author affiliations:

[1] School of Biological Sciences, Life Sciences Building, University of Bristol, Bristol. BS8

1TQ. U.K.

Author for correspondence:

Andrew D. Saxon, Office 2A09, Life Sciences Building, 24 Tyndall Avenue, University of

Bristol, Bristol. BS8 1TQ. U.K.

E-mail: andy.saxon@bristol.ac.uk

Telephone: +44 (0)117 3941174

Running title:

Low male remating ability in a tropical *Drosophila*

Word Count (excl. references, tables, figures): 4547

Abstract (249 words)

Mating success is the main source of fitness variation in males, meaning that males should capitalise on all opportunities for mating. Strong selection on male mating success should also reduce genetic variation in male mating traits relative to other traits. We quantified mating latency, mating duration and productivity in males of the tropical fruitfly, *Drosophila birchii*, from 30 isofemale lines collected from across two elevational gradients, when they were given opportunities to mate with up to four females consecutively. Male remating rates were low compared to other *Drosophila* (only 14 – 27% of males achieved a fourth mating), with mean mating durations approximately doubling across successive copulations. However, although successive remating produced progressively fewer offspring, it consistently increased overall male reproductive success, with males that mated four times more than doubling offspring number compared to males mating only once. We also found no reduction in the productivity of sons emerging from later matings, indicating a sustained cumulative fitness benefit to remating. Heritable variation was observed for most traits ($H^2 = 0.035 – 0.292$) except mating latency, but there was no divergence in trait means with elevation. The observed restricted remating ability of male *D. birchii*, despite the clear benefits of remating, may be due to a low encounter rate with females in the field, leading to high investment per gamete (or ejaculate). However, it remains unclear why genetic variation in these traits is high, given we observe no variation in these traits across elevational gradients known to affect local population density.

Keywords: Male remating traits, reproductive success, sperm allocation, heritability, offspring quality

1. Introduction

Male sexual behaviour is determined by a trade-off between current and future mating success (Parker, 1982). Males should invest more in initial copulations if female mating frequency and/or encounter rate is low (Reinhold *et al*, 2002). However, if sperm is costly and male mating rate is high, strategic allocation of resources over successive matings (and successive females) is expected (Byrne and Rice, 2006; Engqvist and Sauer, 2001). It has been observed that males from several taxa partition sperm across consecutive females (Birkhead, 1991; Perez-Staples and Aluja, 2006; Pitnick and Markow, 1994; Rondeau and Sainte-Marie, 2001; Shapiro and Giraldeau, 1996; Svard and Wiklund, 1986). Understanding the evolution of male mating traits therefore demands measurement of the size of fitness benefits that males gain from multiple matings compared to their first mating, and how this varies among males and their sons (Wedell and Tregenza, 1999). Such variation in male life history within and among species determines reproductive skew, sexual conflict and dimorphism, all of which have profound implications for the ecology and evolution of populations.

Sexual activity reduces male lifespan (Partridge and Farquhar, 1981; Van Voorhies, 1992) and incurs significant energy, time and risk-related costs (Bretman *et al*, 2013b; Dowling and Simmons, 2012; Hayward and Gillooly, 2011). These include the cost of searching and competing for females (Dewsbury, 1982), perceiving female signals (Harvanek *et al*, 2017), complex courtship displays (Connolly *et al*, 1969; Cordts and Partridge, 1996) and prolonged copulation duration or extended mate guarding to maximise paternity share (Bretman *et al*, 2013a; Mazzi *et al*, 2009). Sperm production can also be energetically expensive (Dewsbury, 1982; Hayward and Gillooly, 2011; Paukku and Kotiaho, 2005), especially in species that generate large sperm (Pitnick, 1996). Sperm depletion may therefore limit male reproductive success if mating opportunities are frequent (Hines *et al*, 2003; Pitnick and Markow, 1994; Preston *et al*, 2001; Wedell *et al*, 2002). This means that if opportunities for remating are

common in nature, rapid ejaculate replenishment should be under strong directional selection

(Trivers, 1972).

Although larger and more numerous ejaculates generally increase fitness (Simmons, 2001),

sperm quality affects male fitness independently of sperm quantity (Pattarini *et al*, 2006;

Snook, 2005). Declines in sperm quality (e.g. motility, viability) occur over rapid sequential

ejaculations in rabbits (*Oryctolagus cuniculus*), especially as ejaculates become depleted

(Ambriz *et al*, 2002). Variation in sperm quality among males also results in differences in the

quality of offspring (Alavioon *et al*, 2017; Hosken *et al*, 2003; Siva-Jothy, 2000). For example,

in male guppies (*Poecilia reticulata*), declines in sperm competitive ability are associated with

reduced reproductive success of sons (Gasparini *et al*, 2017). Offspring from later matings may

therefore have reduced fitness compared to those from initial matings.

In *Drosophila*, sperm are transferred in a complex seminal fluid comprising many costly

components (Dewsbury, 1982; Perry *et al*, 2013), including accessory gland proteins (Acps)

that can affect female reproductive behaviour (Gillott, 2003). However, producing these

seminal components may also limit the number and size of ejaculates males can produce

(Lefevre and Jonsson, 1962; Linklater *et al*, 2007; Reinhardt *et al*, 2011; Sirot *et al*, 2009;

Wigby *et al*, 2009), and the replenishment of seminal fluids (or maturation of sperm) is

associated with delays before male remating in other insects (Vahed, 2007). Males also adjust

copulation duration and ejaculate quantity in the presence of rival males, presumably associated

with the likelihood of sperm competition (Gage and Baker, 1991). Edvardsson and Canal

(2006) suggest that declines in copulation duration are associated with decreasing ejaculate

transfer, with reductions in mating duration observed in males of several *Drosophila* species

over consecutive matings (Linklater *et al*, 2007; Singh and Singh, 2013; Singh and Singh,

2000).

Selection to prioritise current versus future mating may also vary across environments, and

with population density. In *Drosophila*, variation in ejaculate size correlates with variation in

female fecundity, mating status and age (Lupold *et al*, 2011; Wedell *et al*, 2002), all of which

may vary across environments (Chapman and Partridge, 1996). Ecological conditions also

determine mating patterns (Gromko and Markow, 1993), due to trade-offs between stress

tolerance and reproduction (Marshall and Sinclair, 2010). For example, low resource

availability may reduce male mating rates (Blay and Yuval, 1997) and their ability to produce

sperm (Gage and Cook, 1994). Latitudinal clines in male mating traits indicate that

environmental variation can determine such allocation patterns (Chahal *et al*, 2013).

Although selection on male mating traits is likely to vary across environments, mating success

traits should have lower heritabilities than morphological or physiological traits because of

their close association with fitness. Alternatively, low heritability estimates in mating traits

could result from their higher sensitivity to environmental variation (Price and Schluter, 1991).

Understanding genetic variation in mating traits is key to predicting the ecological factors that

determine their evolution across species' ranges, and therefore the range of environmental

conditions where their populations can persist (Nadeau *et al*, 2017).

Studies of male mating traits in *Drosophila* mainly focus on cosmopolitan species that are easy

to rear in the laboratory and occur at high density throughout their ecological range (Gowaty,

2012). Tropical species are less well investigated despite typically having narrower abiotic and

biotic tolerances which should have direct and indirect effects on reproduction (Markow and

O'Grady, 2008), and so provide important information on how ecological and demographic

factors affect he evolution of mating behaviour. In this study, we assayed the mating behaviour

and reproductive success of males of the tropical fly *Drosophila birchii*, to explore the effects

of its previously observed low male remating rate (when used in paternal half-sib breeding

designs), compared to its close relatives *Drosophila serrata* (Sgro and Blows, 2003) and *D.*

bunnanda (Van Homrigh *et al*, 2007). This species is limited to tropical rainforest in north-

eastern Australia and New Guinea and has an elevational range from ~0 – 1100 metres, which

is characterised by considerable variation in population density across elevation (Bridle *et al*,

2009; O'Brien *et al*, 2017). It also displays complex male courtship behaviour (Hoikkala *et al*,

2000) and substantial reductions in male mating success under fluctuating versus constant

thermal regimes (Saxon *et al*, 2018).

We assessed genetic variation across the elevational range of this species (at two locations in

northern Queensland), for the following mating and reproductive traits: (*i*) latency to achieve

copulation, (*ii*) duration of copulation, (*iii*) number of offspring produced with each successive

copulation, and (*iv*) total number of offspring sired, when initially virgin males were presented

with four virgin females sequentially. We then tested how this variation in male mating traits

affects male reproductive success, both in terms of offspring and grand-offspring. Surprisingly,

the remating ability of *D. birchii* was found to be remarkably low across all habitats, despite

(*a*) higher remating rates consistently increasing numbers of offspring and grand-offspring and

(*b*) significant levels of genetic variation in key male mating traits.

2. Materials and Methods

(a) Establishment of isofemale lines

Drosophila birchii isofemale lines (called 'lines' hereafter) were founded from field-mated

females collected from two low or high elevation sites along two gradients (eight sites in total):

Mount Edith (Elevation: ~600 – 1100 m, 17°6'S, 145°38'E) and Mount Lewis (Elevation: ~20

– 900 m, 16°35′S 145°17′E), in Queensland, Australia in 2011. Flies were collected using

banana baited buckets, sampled daily using fine sweep nets and sorted under a microscope

using light CO_2 anaesthesia. Females were placed individually in vials to lay, for 5 – 10 days.

Each line was maintained across 3 – 4 40 ml vials containing 10 ml of *Drosophila* potato food

medium (agar, instant mashed potato powder, raw sugar, inactive yeast, propionic acid, nipagin

supplemented with live yeast) at ~100 individuals per generation for each line. A 'mass-bred'

stock was established by combining 10 male and 10 female flies from each line, as a genetically

mixed background population to provide a source for test females to assess the mating

behaviour and fitness of focal males. Mass-bred stocks were reared in 400 ml bottles with 100

ml of *Drosophila* medium and mixed between generations. All lines and the mass-bred

population were maintained with non-overlapping generations at 19 °C on a 12:12-h light:dark

cycle at 60% relative humidity prior to the experiments. Mount Lewis experiments were

conducted after ~25 laboratory generations, Mount Edith ~50 generations.

(b) Experiment I: Assaying male remating and productivity

Ten lines were used for the Mount Edith assay, with five lines originating from two high

elevation sites and five from two lower elevation sites. Twenty lines were used for the Mount

Lewis assay, with 10 originating from two high and two low sites. Prior to the experiment, line

stocks and the mass-bred population were reared at a constant 25 °C, 12:12 hour light:dark

cycle for two generations to randomise and minimise any transgenerational effects attributable

to maternal condition. The experimental males and background females were reared at minimal

density conditions to minimise larval competition. Five male and five female flies mated and

laid eggs in 10 ml of standard *Drosophila* medium for three days. Parental flies were then

removed and pupation card (75 x 30mm) inserted into the vial.

On eclosion, flies were anaesthetised under CO_2 and sexed using a Leica (MZ9.5) microscope.

30 males for each of 10 lines ($N = 300$) from Mount Edith and 20 males from 20 lines ($N = 400$)

from Mount Lewis were collected, along with mass-bred females, within 12 hours of

emergence to ensure virgin status. All flies were held in single-sex vials for six days, at 25 °C

in low density vials (maximum of 10 flies per vial) with fly food medium *ad libitum,* before

the mating assay commenced.

Mating assay: The assay was conducted at a constant 25 °C and began within an hour of the

daylight period, to coincide with time of peak activity in *Drosophila*. Each male was placed in

a vial with 8 ml of standard fly medium. A virgin female was placed with the focal male and

the start time noted. If mating was initiated, the time was recorded to give time to copulation

(latency), as was the end mating time (duration). Following copulation, the male was moved to

a fresh vial and presented with a new female. This process continued with up to four matings

allowed for each male, with males given up to two hours to mate with each female. If no mating

occurred, then the male was recorded as 'not mated' and the assay concluded for that male.

The assay took place over five days with 60 (for Mount Edith) and 80 (for Mount Lewis) male

flies assayed per day, using equal numbers from each line. The order of males was randomised

within each block to ensure that there was no systematic bias among lines due to diurnal effects.

Blind ID codes were used to avoid observer bias.

Mated females were left in the vials for five days, which ensured that they had laid all fertilised

eggs, given that the lack of spermathecae in *D. birchii* females means that sperm storage is

minimal (R. Snook, personal communication, 2014). The female was then removed and a

pupation card added. At least 20 days after mating, the total productivity (number of offspring)

of each successful mating for each male was recorded.

(c) Experiment II: Assaying the fitness of sons derived from successive paternal matings

Ten virgin males (sons) were collected from each of the first to fourth (1 – 4) paternal matings

of focal males (sires) in Experiment I that had achieved the maximum four matings. Sons were

derived from seven of the ten Mount Edith isofemale lines (N = ~280). Each six-day old male was placed in a single vial of 8 ml of fly medium with a single, virgin mass-bred female of the same age (± 12 hours). The pair were left for three days to mate under the same control conditions as Experiment I. The male was then removed and the female was left to lay for a further five days. The female was then removed and pupation card inserted. After all offspring had emerged, the mean productivity of matings from these sons was assayed with respect to the order of mating and their paternal line.

(d) Statistical analysis

The R (R Core Team, 2016) package *lme4* was used to fit linear mixed models to test for effects of successive matings on male mating traits (latency to mate, duration of mating, number of offspring per mating, total offspring per male). Separate models were fitted for each trait and gradient. All data were untransformed because trait data were normally distributed. Mating number and elevation of origin of line (high or low) were included as fixed factors and focal male nested within isofemale line were specified as random effects. For total offspring per male, only elevation of origin of line was used as a fixed factor. The significance of each factor in the model was determined using likelihood ratio tests to compare the full model with a model where that factor had been removed (Experiment I). *P*-values were corrected for multiple comparisons following the False Discovery Rate method of Benjamini and Hochberg (1995), allowing an FDR of 0.05. Linear mixed models were also run to analyse overall differences in the mean proportion of males per line that attained each successive mating and the mean cumulative number of offspring per line at each mating. Mating number was used as a fixed factor with line as a random effect. For Experiment II, a linear mixed model with mating number as a fixed factor and line as a random effect was used to analyse the productivity of

sons derived from successive paternal mating events when mating with a single, virgin female

from the mass-bred stock.

To estimate broad-sense heritability (H^2) for mating traits, between line variances from the

linear mixed models were used (Experiment I), using within (V_w) and between (V_b) line

variance components derived from models fitted using REML. This was an overall estimate

for each gradient as the small number of lines per elevation precluded estimates for high and

low elevation separately. Inbreeding coefficients (F_t) (with the assumption of full-sibship) for

the number of generations and population size (~100) at which lines had been held at in the

laboratory were calculated using the method of Falconer and Mackay (1996). H^2 was then

calculated following Hoffmann and Parsons (1988)(see SM). The significance of the H^2 was

evaluated from the significance of the between-line variance component in the linear mixed

models.

3. Results

(a) Experiment I: Variation in male mating traits over successive matings

In Experiment I, 76% of males at Mount Edith attained a first mating ($N = 220$ males), with

only 27% reaching a fourth. 19% only mated once. 63% of males at Mount Lewis attained a

first mating ($N = 166$ males), while only 14% attained a fourth mating, with 14% of males

mating only once (Figure 2). Mean cumulative productivity, while significantly increasing

across successive matings, slowed its rate of increase with each copulation (Fig. 3). For Mount

Edith lines, mean productivity per mating (for all males) showed a 64% decrease from mating

1 to mating 4. Focal males that mated only once had mean total offspring of 43.02 (standard

error ± 5.176), while males that mated four times had a mean total cumulative productivity of

145.07 (± 18.305). For Mount Lewis lines, mean productivity per mating declined by 72% from

mating 1 to mating 4. Males that mated only once had mean total offspring of 63.97 ($\pm$ 6.010),

while males reaching a fourth mating had a mean total productivity of 133.31 ($\pm$ 7.041). The

subset of males that reached a fourth mating showed a similar pattern of declining productivity

to the pattern observed for all males (Figure SM1).

Latency to mating did not vary with successive matings for line from either gradient. Between-

line (genetic) variance in this trait was also not observed at either gradient. However, both

mating duration and productivity showed a highly significant change across successive matings

for both gradients. Mating duration increased approximately twofold and productivity more

than halved over the four successive matings (Figure 1; Table 1). Line also showed significant

variance in these traits (with the exception of mating duration in the Mount Lewis lines), but

there was no detectable difference between lines originating from high and low elevation sites

for any of these traits. Broad-sense heritabilities (H^2) for variance components in the Mount

Edith and Mount Lewis (Table 2) lines gave low to moderate estimates. Mating duration gave

low H^2 values (0.035 – 0.126), although the estimate did not significantly differ from zero at

Mount Lewis. H^2 estimates were significantly different from zero for number of offspring

produced per mating, ranging from 0.069 – 0.229. Total productivity per male exhibited

significant heritability at Mount Edith (0.292), but not at Mount Lewis.

(b) Experiment II: Fitness of sons from successive paternal matings

There were no significant differences in the productivity of sons derived from successive

paternal matings in Experiment II (Fig. 4; χ^2 = 0.2675, df = 1, P = ns). There was also no

significant variation in productivity among sons attributable to isofemale line used (Variance

= 6.65 x 10^{-16}, P = ns; Residual = 537.1). Mean number of offspring produced from mating

with a single, virgin female was similar for sons obtained from sire matings 1 to 4: Mating 1 =

73.3 (Standard Error ± 2.59), Mating 2 = 75.4 (± 2.96), Mating 3 = 73.5 (± 3.24), Mating 4 =

76.2 (± 2.98).

4. Discussion

The substantial variation in male mating strategies observed among *Drosophila* species

(Gowaty *et al*, 2003; Singh and Singh, 2013) is likely to correlate with ecological parameters

that relate to the risk of sperm competition, likelihood of repeated mating opportunities, and

the energetic cost of mating relative to resistance to factors such as temperature stress and

pathogen or parasitoid exposure. Here, we demonstrate that males of the rainforest specialist

Drosophila birchii exhibit a much lower remating rate than cosmopolitan species for which

data are available. For example, *D. melanogaster* males will copulate with 10 females per day

given the opportunity (Gromko, 1992; Mossige, 1955), while *D. hydei* can mate up to 10 times

in two hours (Markow, 1985). By contrast, only 27% and 14% of *D. birchii* males, from Mount

Edith and Mount Lewis lines respectively, mated a fourth time within the single day of our

experiments, with only 58% and 49% mating more than once (Fig. 2). This was despite the two

to threefold increase in the total number of offspring produced by males that mated four times

compared to those that mated only once (Fig. 3). In addition, unlike many *Drosophila* species,

female *D. birchii* have no spermathecae and therefore cannot store sperm (R. Snook, personal

communication, 2014). There is also strong last male precedence in paternity for *D. birchii*

when females remate (E.K. O'Brien, unpublished data, 2017). Both of these, coupled with the

sustained increases in productivity where remating occurs, should make remating consistently

beneficial to male fitness, even when other males are around. These observations therefore

make this observation of low male remating rates especially surprising.

Drosophila birchii are often found at low densities in their rainforest habitat (Bridle *et al*, 2009;

O'Brien *et al*, 2017), compared to other tropical drosophilids. This means conspecific

interactions (especially the female encounter rate for males) may be relatively rare, which is

likely to shape resource allocation across matings (Aspi and Hoffmann, 1998; Shelly and

Bailey, 1992; Willis *et al*, 2011). In particular, reduced risk of sperm competition may favour

reductions in sperm quantity (Bjork *et al*, 2007), in favour of increased investment per gamete

(Snook, 2005) and reduced allocation across multiple matings (Ingleby *et al*, 2010). Reductions

in production of seminal fluid proteins also occur in less sexually competitive environments

(Wigby *et al*, 2016).

Ejaculate components such as sperm (Wedell *et al*, 2002) or seminal fluids (Wigby *et al*, 2009)

can limit male remating. Spermiogenesis in male *Drosophila melanogaster* takes five days (at

25 °C) (Fabian and Brill, 2012) and mated males with depleted seminal fluid proteins require

three days of sexual inactivity before transferring equivalent quantities again (Sirot *et al*, 2009).

Although sperm size in *D. birchii* is not known, long flagella and long ventral receptacles in

females (R. Snook, personal communication, 2014) suggest that male investment in sperm is

high (Markow, 2015). If energetic costs of mating are high for male *D. birchii*, successive

mating events may increase male choosiness, as the relative cost of mating rises with male

resource depletion (Byrne and Rice, 2006). In this study however, no variation was found in

latency to copulation across matings (Fig. 1), suggesting no increase in male choosiness (i.e.

inclination to remate) (Engqvist and Sauer, 2001) or reduction in male attractiveness to females

(Taylor *et al*, 2007).

Multiple mating in *D. birchii* was associated with a reduction in the per mating number of

offspring produced in later mating events (Fig. 1). This is also observed in *D. melanogaster*

(Lefevre and Jonsson, 1962), despite substantially lower absolute productivity per mating in

D. birchii. Importantly, these patterns were not an artefact of the non-random subset of males

for which estimates of fourth matings are possible (i.e. those males that successfully copulate four times) because a comparable reduction in productivity across matings was seen for this subset of males too (Fig. SM1). This suggests rapid ejaculate depletion even in males that successfully mated four times, and contrasts with studies in tropical tephritid flies that found males do not deplete sperm stores, allocating similar quantities over three consecutive matings (Perez-Staples and Aluja, 2006).

A consistent increase in mean mating duration was observed with each successive copulation, from ~3 to 6 minutes over four matings (Fig. 1). Extending mating durations can increase male fitness by increasing paternity (Bretman *et al*, 2013a; Mazzi *et al*, 2009). By contrast, in several other *Drosophila* species copulation durations decline over consecutive matings (Linklater *et al*, 2007; Singh and Singh, 2013; Singh and Singh, 2000), suggesting that duration decreases as male ejaculate becomes depleted (Edvardsson and Canal, 2006). In these experiments however, declining productivity with increasing duration indicates that extended copulation duration is not associated with increased sperm transfer. Gilchrist and Partridge (2000) found that sperm transfer occurs rapidly after the initiation of mating in *D. melanogaster* and that extended copulations delay female remating by transmitting seminal fluids that boost fecundity and reduce receptivity in females. Similarly, increases in mating duration with repeated male copulations in the housefly (*Musca domestica*), were likely to be due to the depletion of such accessory secretions, with males only terminating copulations when the levels transferred were sufficient to stimulate an inhibitory response to further mating in females (Leopold *et al*, 1971). While mating duration is largely controlled by males in several *Drosophila* (MacBean and Parsons, 1967; Parsons and Kaul, 1966; Patty, 1975; Singh and Singh, 2000), studies with *D. birchii* have shown that females can dislodge males (Hoikkala and Crossley, 2000). Similarly, females have an influence on copulation duration in *D. mojavensis* (Krebs, 1991), *D. elegans* (Hirai *et al*, 1999) and *D. montana* (Mazzi *et al*, 2009). Further research is necessary to clarify

female fitness effects of prolonging copulations. Alternatively, prolonging copulation might

represent a mate guarding strategy by males, particularly as sperm becomes limited (Simmons,

2001). However, the relatively small absolute increases in mean copulation duration in this

study make such hypothesised mate guarding unlikely.

Males were assayed under standardised (constant 25 °C) laboratory conditions. However, male

allocation strategies are likely to show adaptive responses to the more variable conditions

experienced in space and time by natural populations (Wedell *et al*, 2002; Wigby *et al*, 2016).

Pitnick and Markow (1994) propose that submaximal male ejaculate allocation over successive

matings may act as bet-hedging for male *Drosophila*, particularly where environments are

stressful. Variation in abiotic conditions also affects insect mating duration (Horton *et al*,

2002), sperm allocation and remating rate (Katsuki and Miyatake, 2009) within genotypes.

Latitudinal variation in male remating traits within two cosmopolitan *Drosophila* species has

also been observed (Chahal *et al*, 2013; Singh and Singh, 2000). Bouletreaumerle *et al* (1982)

found that female fecundity was lower in tropical versus temperate populations of *D.*

melanogaster, suggesting that males of tropical species may also invest less in sperm or seminal

components associated with the environments they inhabit.

Despite this evidence for adaptive divergence in male trait in response to the local environment,

we found no evidence for genetic divergence in any mating trait across either elevational

gradient (Table 1), even though high and low elevation thermal environments (~7 °C difference

in mean temperature) characterise the cold and warm ecological limits of this species.

However, there was significant genetic variation between isofemale lines in number of

offspring (productivity) over successive matings at both gradients, with between line variance

representing ~17% of total variance in Mount Edith lines and ~4% at Mount Lewis. Similar

among-line variation was observed for mating duration at Mount Edith although not at Mount

Lewis. Broad-sense heritability (H^2) estimates varied substantially between gradients with

Mount Edith showing consistently lower levels of genetic variance. For instance, offspring per

mating gave values of 0.229 at Mount Edith compared to 0.069 at Mount Lewis. However,

latency to mate showed no between-line variation and mating duration was only significantly

heritable at Mount Edith.

Despite the restricted distribution of *D. birchii*, and the strong effects of these traits on fitness,

heritabilities are remarkably high, at least at Mount Edith, compared to the wide-range of

estimates obtained in *D. melanogaster*, for male courtship traits (0.033 – 0.094) (Gaertner *et*

al, 2015), mating latency (0.01), duration (0.007) (Gromko, 1987) and mating success (0.25)

(Tucic *et al*, 1988). Taylor *et al* (2013) observed no significant heritable variation in male

mating latency and duration. Maintenance of genetic variation in such key fitness traits may

depend on potential trade-offs between investment in sperm/seminal fluids and resistance to

environmental stress. For example, thermal stress can have a substantial detrimental impact on

male reproduction in ectotherms, including *D. birchii*, possibly due to the need to divert

resources away from sperm production to upregulate physiological stress responses (Sales *et*

al, 2018; Saxon *et al*, 2018).

Sexual conflict, 'good genes' and 'sexy sons' models of sexual selection all predict that males

pass on genetic benefits to offspring (Kokko, 2001; Taylor *et al*, 2013; Wedell and Tregenza,

1999). However, if decreasing ejaculate quantity over successive matings in males is associated

with declines in sperm quality, this may affect the fitness of offspring from later matings,

making it easier to explain such high levels of variation in male mating strategies, even when

they increase the number of first-generation offspring produced. However, Experiment II

demonstrated that the paternal (sire) mating sequence had no significant effect on the

reproductive success of sons from later matings compared to those from first mating, when

they were mated to a single female (Fig. 4). This means that no deterioration in sperm/ejaculate

quality is observed at least until after males' fourth mating, which is associated with a 237%

relative increase in mean total productivity at Mount Edith and 111% increase at Mount Lewis

compared to a male that mates only once. Such variation in male remating rate within *D. birchii*

populations therefore remains unexpected, given the apparent unequivocal fitness advantage it

provides.

Acknowledgements: We are grateful to Rhonda Snook for work assaying *D. birchii* sperm

morphology and female reproductive tracts and to Tom Tregenza for helpful comments on a

draft of the manuscript. This work was funded by a University of Bristol PhD scholarship for

ADS and a Natural Environment Research Council standard grant (no. NE/G007039/1) to JRB.

Conflict of Interest: The authors declare no conflict of interest.

Data Archiving: All data referred to in this manuscript will be archived in Dryad.

References

Alavioon G, Hotzy C, Nakhro K, Rudolf S, Scofield DG, Zajitschek S *et al* (2017). Haploid
selection within a single ejaculate increases offspring fitness. *Proceedings of the National*
Academy of Sciences of the United States of America **114**(30)**:** 8053-8058.

Ambriz D, Rosales AM, Sotelo R, Mora JA, Rosado A, Garcia AR (2002). Changes in the
quality of rabbit semen in 14 consecutive ejaculates obtained every 15 minutes. *Archives of*
Andrology **48**(5)**:** 389-395.

Aspi J, Hoffmann AA (1998). Female encounter rates and fighting costs of males are
associated with lek size in *Drosophila mycetophaga*. *Behavioral Ecology and Sociobiology*
42(3)**:** 163-169.

Benjamini Y, Hochberg Y (1995). Controlling the false discovery rate - a practical and
powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B-*
Methodological **57**(1)**:** 289-300.

418
419 Birkhead TR (1991). Sperm depletion in the Bengalese Finch, *Lonchura striata. Behavioral*
420 *Ecology* **2**(4)**:** 267-275.

421
422 Bjork A, Dallai R, Pitnick S (2007). Adaptive modulation of sperm production rate in
423 *Drosophila bifurca*, a species with giant sperm. *Biology Letters* **3**(5)**:** 517-519.

424
425 Blay S, Yuval B (1997). Nutritional correlates of reproductive success of male Mediterranean
426 fruit flies (Diptera: Tephritidae). *Animal Behaviour* **54:** 59-66.

427
428 Bouletreaumerle J, Allemand R, Cohet Y, David JR (1982). Reproductive strategy in
429 *Drosophila melanogaster* - significance of a genetic divergence between temperate and
430 tropical populations. *Oecologia* **53**(3)**:** 323-329.

431
432 Bretman A, Westmancoat JD, Chapman T (2013a). Male control of mating duration
433 following exposure to rivals in fruitflies. *Journal of Insect Physiology* **59**(8)**:** 824-827.

434
435 Bretman A, Westmancoat JD, Gage MJG, Chapman T (2013b). Costs and benefits of lifetime
436 exposure to mating rivals in male *Drosophila melanogaster. Evolution* **67**(8)**:** 2413-2422.

437
438 Bridle JR, Gavaz S, Kennington WJ (2009). Testing limits to adaptation along altitudinal
439 gradients in rainforest *Drosophila. Proceedings of the Royal Society B-Biological Sciences*
440 **276**(1661)**:** 1507-1515.

441
442 Byrne PG, Rice WR (2006). Evidence for adaptive male mate choice in the fruit fly
443 *Drosophila melanogaster. Proceedings of the Royal Society B-Biological Sciences*
444 **273**(1589)**:** 917-922.

445
446 Chahal J, Dev K, Kataria SK, Parkash R (2013). Opposite latitudinal clines for first mating
447 and second mating (remating) in males of *Drosophila melanogaster. Ethology* **119**(11)**:** 926-
448 936.

449
450 Chapman T, Partridge L (1996). Female fitness in Drosophila melanogaster: An interaction
451 between the effect of nutrition and of encounter rate with males. *Proceedings of the Royal*
452 *Society B-Biological Sciences* **263**(1371)**:** 755-759.

453
454 Connolly K, Burnet B, Sewell D (1969). Selective mating and eye pigmentation - an analysis
455 of visual component in courtship behavior of *Drosophila melanogaster. Evolution* **23**(4)**:**
456 548-&.

458   Cordts R, Partridge L (1996). Courtship reduces longevity of male *Drosophila melanogaster*.
459   *Animal Behaviour* **52**: 269-278.

460
461   Dewsbury DA (1982). Ejaculate cost and male choice. *American Naturalist* **119**(5): 601-610.

462
463   Dowling DK, Simmons LW (2012). Ejaculate economics: Testing the effects of male sexual
464   history on the trade-off between sperm and immune function in Australian crickets. *PLoS*
465   *One* **7**(1): e30172.

466
467   Edvardsson M, Canal D (2006). The effects of copulation duration in the bruchid beetle
468   *Callosobruchus maculatus*. *Behavioral Ecology* **17**(3): 430-434.

469
470   Engqvist L, Sauer KP (2001). Strategic male mating effort and cryptic male choice in a
471   scorpionfly. *Proceedings of the Royal Society B-Biological Sciences* **268**(1468): 729-735.

472
473   Fabian L, Brill JA (2012). *Drosophila* spermiogenesis: Big things come from little packages.
474   *Spermatogenesis* **2**(3): 197-212.

475
476   Falconer DS, Mackay TFC (1996). *Introduction to Quantitative Genetics*, 4th edn. Pearson
477   Education Ltd: Harlow, UK.

478
479   Gaertner BE, Ruedi EA, McCoy LJ, Moore JM, Wolfner MF, Mackay TFC (2015). Heritable
480   variation in courtship patterns in *Drosophila melanogaster*. *G3-Genes Genomes Genetics*
481   **5**(4): 531-539.

482
483   Gage MJG, Baker RR (1991). Ejaculate size varies with sociosexual situation in an insect.
484   *Ecological Entomology* **16**(3): 331-337.

485
486   Gage MJG, Cook PA (1994). Sperm size or numbers : Effects of nutritional stress upon
487   eupyrene and apyrene sperm production strategies on the moth *Plodia interpunctella*
488   (Lepidoptera: Pyralidae). *Functional Ecology* **8**(5): 594-599.

489
490   Gasparini C, Dosselli R, Evans JP (2017). Sperm storage by males causes changes in sperm
491   phenotype and influences the reproductive fitness of males and their sons. *Evolution Letters*
492   **1**(1): 16-25.

493
494   Gilchrist AS, Partridge L (2000). Why it is difficult to model sperm displacement in
495   *Drosophila melanogaster*: The relation between sperm transfer and copulation duration.
496   *Evolution* **54**(2): 534-542.

497
498   Gillott C (2003). Male accessory gland secretions: Modulators of female reproductive
499   physiology and behavior. *Annual Review of Entomology* **48**: 163-184.

501   Gowaty PA (2012). The evolution of multiple mating Costs and benefits of polyandry to
502   females and of polygyny to males. *Fly* **6**(1)**:** 3-11.

503
504   Gowaty PA, Steinichen R, Anderson WW (2003). Indiscriminate females and choosy males:
505   Within- and between-species variation in *Drosophila*. *Evolution* **57**(9)**:** 2037-2045.

506
507   Gromko MH (1987). Genetic constraint on the evolution of courtship behavior in *Drosophila*
508   *melanogaster*. *Heredity* **58:** 435-441.

509
510   Gromko MH (1992). Genetic correlation of male and female mating frequency - evidence
511   from *Drosophila melanogaster*. *Animal Behaviour* **43**(1)**:** 176-177.

512
513   Gromko MH, Markow TA (1993). Courtship and remating in field populations of
514   *Drosophila*. *Animal Behaviour* **45**(2)**:** 253-262.

515
516   Harvanek ZM, Lyu Y, Gendron CM, Johnson JC, Kondo S, Promislow DEL *et al* (2017).
517   Perceptive costs of reproduction drive ageing and physiology in male *Drosophila*. *Nature*
518   *Ecology & Evolution* **1**(6)**:** 152.

519
520   Hayward A, Gillooly JF (2011). The cost of sex: Quantifying energetic investment in gamete
521   production by males and females. *PLoS One* **6**(1)**:** e16557.

522
523   Hines AH, Jivoff PR, Bushmann PJ, van Montfrans J, Reed SA, Wolcott DL *et al* (2003).
524   Evidence for sperm limitation in the blue crab, Callinectes sapidus. *Bulletin of Marine*
525   *Science* **72**(2)**:** 287-310.

526
527   Hirai Y, Sasaki H, Kimura MT (1999). Copulation duration and its genetic control in
528   *Drosophila elegans*. *Zoological Science* **16**(2)**:** 211-214.

529
530   Hoffmann AA, Parsons PA (1988). The analysis of quantitative variation in natural
531   populations with isofemale strains. *Genetics Selection Evolution* **20**(1)**:** 87-98.

532
533   Hoikkala A, Crossley S (2000). Copulatory courtship in *Drosophila*: Behavior and songs of
534   *D. birchii* and *D. serrata*. *Journal of Insect Behavior* **13**(1)**:** 71-86.

535
536   Hoikkala A, Crossley S, Castillo-Melendez C (2000). Copulatory courtship in *Drosophila*
537   *birchii* and *D. serrata* - species recognition and sexual selection. *Journal of Insect Behavior*
538   **13**(3)**:** 361-373.

539
Horton DR, Lewis TM, Hinojosa T (2002). Copulation duration in three species of
Anthocoris (Heteroptera : Anthocoridae) at different temperatures and effects on insemination
and ovarian development. *Pan-Pacific Entomologist* **78**(1)**:** 43-55.

Hosken DJ, Garner TWJ, Tregenza T, Wedell N, Ward PI (2003). Superior sperm
competitors sire higher-quality young. *Proceedings of the Royal Society B-Biological*
Sciences **270**(1527)**:** 1933-1938.

Ingleby FC, Lewis Z, Wedell N (2010). Level of sperm competition promotes evolution of
male ejaculate allocation patterns in a moth. *Animal Behaviour* **80**(1)**:** 37-43.

Katsuki M, Miyatake T (2009). Effects of temperature on mating duration, sperm transfer and
remating frequency in *Callosobruchus chinensis*. *Journal of Insect Physiology* **55**(2)**:** 112-
115.

Kokko H (2001). Fisherian and "good genes" benefits of mate choice: how (not) to
distinguish between them. *Ecology Letters* **4**(4)**:** 322-326.

Krebs RA (1991). Function and genetics of long versus short copulations in the cactophilic
fruit-fly, *Drosophila mojavensis* (Diptera, Drosophilidae). *Journal of Insect Behavior* **4**(2)**:**
221-233.

Lefevre G, Jonsson UB (1962). Sperm transfer, storage, displacement, and utilization in
Drosophila melanogaster. *Genetics* **47**(12)**:** 1719-1736.

Leopold RA, Terranova AC, Swilley EM (1971). Mating refusal in *Musca domestica* -
Effects of repeated mating and decerebration upon frequency and duration of copulation.
Journal of Experimental Zoology **176**(3)**:** 353-359.

Linklater JR, Wertheim B, Wigby S, Chapman T (2007). Ejaculate depletion patterns evolve
in response to experimental manipulation of sex ratio in Drosophila melanogaster. *Evolution*
61(8)**:** 2027-2034.

Lupold S, Manier MK, Ala-Honkola O, Belote JM, Pitnick S (2011). Male *Drosophila*
melanogaster adjust ejaculate size based on female mating status, fecundity, and age.
Behavioral Ecology **22**(1)**:** 184-191.

MacBean IT, Parsons PA (1967). Directional selection for duration of copulation in
Drosophila melanogaster. *Genetics* **56**(2)**:** 233-239.

Markow TA (1985). A comparative investigation of the mating system of *Drosophila hydei*.
Animal Behaviour **33**(Aug)**:** 775-781.

Markow TA (2015). *Drosophila* reproduction: Molecules meet morphology. *Proceedings of*
the National Academy of Sciences of the United States of America **112**(27)**:** 8168-8169.

Markow TA, O'Grady P (2008). Reproductive ecology of Drosophila. *Functional Ecology*
22(5)**:** 747-759.

Marshall KE, Sinclair BJ (2010). Repeated stress exposure results in a survival-reproduction
trade-off in *Drosophila melanogaster*. *Proceedings of the Royal Society B-Biological*
Sciences **277**(1683)**:** 963-969.

Mazzi D, Kesaniemi J, Hoikkala A, Klappert K (2009). Sexual conflict over the duration of
copulation in *Drosophila montana*: Why is longer better? *Bmc Evolutionary Biology* **9**.

Mossige JC (1955). Sperm utilization and brood patterns in *Drosophila melanogaster*.
American Naturalist **89**(845)**:** 123-127.

Nadeau CP, Urban MC, Bridle JR (2017). Climates Past, Present, and Yet-to-Come Shape
Climate Change Vulnerabilities. *Trends in Ecology & Evolution* **32**(10)**:** 786-800.

O'Brien EK, Higgie M, Reynolds A, Hoffmann AA, Bridle JR (2017). Testing for local
adaptation and evolutionary potential along altitudinal gradients in rainforest *Drosophila*:
beyond laboratory estimates. *Global Change Biology* **23**(5)**:** 1847-1860.

Parker GA (1982). Why are there so many tiny sperm - Sperm competition and the
maintenance of 2 sexes. *Journal of Theoretical Biology* **96**(2)**:** 281-294.

Parsons PA, Kaul D (1966). Mating speed and duration of copulation in Drosophila
pseudoobscura. *Heredity* **21**(2)**:** 219-225.

Partridge L, Farquhar M (1981). Sexual activity reduces lifespan of male fruitflies. *Nature*
294(5841)**:** 580-582.

Pattarini JA, Starmer WT, Bjork A, Pitnick S (2006). Mechanisms underlying the sperm
quality advantage in *Drosophila melanogaster*. *Evolution* **60**(10)**:** 2064-2080.

Patty RA (1975). Investigation of genetic factors influencing duration of copulation in
"eastern" and "western" Drosophila athabasca. *Animal Behaviour* **23**(2)**:** 344-348.

Paukku S, Kotiaho JS (2005). Cost of reproduction in Callosobruchus maculatus: effects of
mating on male longevity and the effect of male mating status on female longevity. *Journal*
of Insect Physiology **51**(11)**:** 1220-1226.

Perez-Staples D, Aluja M (2006). Sperm allocation and cost of mating in a tropical tephritid
fruit fly. *Journal of Insect Physiology* **52**(8)**:** 839-845.

Perry JC, Sirot L, Wigby S (2013). The seminal symphony: How to compose an ejaculate.
Trends in Ecology & Evolution **28**(7)**:** 414-422.

Pitnick S (1996). Investment in testes and the cost of making long sperm in *Drosophila*.
American Naturalist **148**(1)**:** 57-80.

Pitnick S, Markow TA (1994). Male gametic strategies - Sperm size, testes size, and the
allocation of ejaculate among successive mates by the sperm-limited fly *Drosophila pachea*
and its relatives. *American Naturalist* **143**(5)**:** 785-819.

Preston BT, Stevenson IR, Pemberton JM, Wilson K (2001). Dominant rams lose out by
sperm depletion - A waning success in siring counters a ram's high score in competition for
ewes. *Nature* **409**(6821)**:** 681-682.

Price T, Schluter D (1991). On the low heritability of life-history traits. *Evolution* **45**(4)**:** 853-
861.

R Core Team. (2016). R Foundation for Statistical Computing.: Vienna, Austria.

Reinhardt K, Naylor R, Siva-Jothy MT (2011). Male mating rate is constrained by seminal
fluid availability in bedbugs, *Cimex lectularius*. *PLoS One* **6**(7)**:** e22082.

Reinhold K, Kurtz J, Engqvist L (2002). Cryptic male choice: sperm allocation strategies
when female quality varies. *Journal of Evolutionary Biology* **15**(2)**:** 201-209.

Rondeau A, Sainte-Marie B (2001). Variable mate-guarding time and sperm allocation by
male snow crabs (Chionoecetes opilio) in response to sexual competition, and their impact on
the mating success of females. *Biological Bulletin* **201**(2)**:** 204-217.

Sales K, Vasudeva R, Dickinson ME, Godwin JL, Lumley AJ, Michalczyk L *et al* (2018).
Experimental heatwaves compromise sperm function and cause transgenerational damage in
a model insect. *Nature Communications* **9**.

Saxon AD, O'Brien EK, Bridle JR (2018). Temperature fluctuations during development
reduce male fitness and may limit adaptive potential in tropical rainforest Drosophila.
Journal of Evolutionary Biology **31**(3)**:** 405-415.

Sgro CM, Blows MW (2003). Evolution of additive and nonadditive genetic variance in
development time along a cline in Drosophila serrata. *Evolution* **57**(8)**:** 1846-1851.

Shapiro DY, Giraldeau LA (1996). Mating tactics in external fertilizers when sperm is
limited. *Behavioral Ecology* **7**(1)**:** 19-23.

Shelly TE, Bailey WJ (1992). Experimental manipulation of mate choice by male katydids:
The effect of female encounter rate. *Behavioral Ecology and Sociobiology* **30**(3-4)**:** 277-282.

673
674 Simmons LW (2001). *Sperm Competition and Its Evolutionary Consequences in the Insects*.
675 Princeton University Press: USA.

676
677 Singh A, Singh BN (2013). Studies on remating behaviour in the *Drosophila bipectinata*
678 species complex: Intra- and interspecific variations. *Behavioural Processes* **96:** 79-87.

679
680 Singh SR, Singh BN (2000). Male remating in *Drosophila ananassae*: Evidence for
681 interstrain variation in remating time and shorter duration of copulation during second
682 mating. *Zoological Science* **17**(3)**:** 389-393.

683
684 Sirot LK, Buehner NA, Fiumera AC, Wolfner MF (2009). Seminal fluid protein depletion
685 and replenishment in the fruit fly, *Drosophila melanogaster*: An ELISA-based method for
686 tracking individual ejaculates. *Behavioral Ecology and Sociobiology* **63**(10)**:** 1505-1513.

687
688 Siva-Jothy MT (2000). The young sperm gambit. *Ecology Letters* **3**(3)**:** 172-174.

689
690 Snook RR (2005). Sperm in competition: Not playing by the numbers. *Trends in Ecology &*
691 *Evolution* **20**(1)**:** 46-53.

692
693 Svard L, Wiklund C (1986). Different ejaculate delivery strategies in 1st versus subsequent
694 matings in the Swallowtail butterfly *Papilio machaon L. Behavioral Ecology and*
695 *Sociobiology* **18**(5)**:** 325-330.

696
697 Taylor ML, Evans JP, Garcia-Gonzalez F (2013). No evidence for heritability of male mating
698 latency or copulation duration across social environments in *Drosophila melanogaster*. *PLoS*
699 *One* **8**(10)**:** e77347.

700
701 Taylor ML, Wedell N, Hosken DJ (2007). The heritability of attractiveness. *Current Biology*
702 **17**(22)**:** R959-R960.

703
704 Trivers RL (1972). Parental investment and sexual selection. In: Campbell B (ed) *Sexual*
705 *selection and the descent of man 1871-1971*. Aldine: Chicago, pp 136-179.

706
707 Tucic N, Cvetkovic D, Milanovic D (1988). The genetic variation and covariation among
708 fitness components in *Drosophila melanogaster* females and males. *Heredity* **60:** 55-60.

709
710 Vahed K (2007). Comparative evidence for a cost to males of manipulating females in
711 bushcrickets. *Behavioral Ecology* **18**(3)**:** 499-506.

713 Van Homrigh A, Higgie M, McGuigan K, Blows MW (2007). The depletion of genetic
714 variance by sexual selection. *Current Biology* **17**(6)**:** 528-532.

715
716 Van Voorhies WA (1992). Production of sperm reduces nematode lifespan. *Nature*
717 **360**(6403)**:** 456-458.

718
719 Wedell N, Gage MJG, Parker GA (2002). Sperm competition, male prudence and sperm-
720 limited females. *Trends in Ecology & Evolution* **17**(7)**:** 313-320.

721
722 Wedell N, Tregenza T (1999). Successful fathers sire successful sons. *Evolution* **53**(2)**:** 620-
723 625.

724
725 Wigby S, Perry JC, Kim YH, Sirot LK (2016). Developmental environment mediates male
726 seminal protein investment in *Drosophila melanogaster*. *Functional Ecology* **30**(3)**:** 410-419.

727
728 Wigby S, Sirot LK, Linklater JR, Buehner N, Calboli FC, Bretman A *et al* (2009). Seminal
729 fluid protein allocation and male reproductive success. *Current Biology* **19**(9)**:** 751-757.

730
731 Willis PM, Ryan MJ, Rosenthal GG (2011). Encounter rates with conspecific males influence
732 female mate choice in a naturally hybridizing fish. *Behavioral Ecology* **22**(6)**:** 1234-1240.

Table 1. Linear mixed effects analyses for male mating traits in Mount Edith and Mount Lewis lines. Mating (1 – 4) and elevation of origin of isofemale line are fixed effects, with nested random effects of isofemale line and focal male. *P*-values were obtained by likelihood ratio tests of the full model with the effect against the model with the effect excluded (*ns*= not significant, *adjusted P*≤ 0.05, **P* ≤ 0.01, ***P* ≤ 0.001).

Mating trait	Gradient	Fixed effect				Random effects		
		Predictor	χ^2	df	P	Variance component	Variance	P
Latency	Mt. Edith	Mating	0.5111	3	ns	Isofemale line	28.61	ns
						Male	9.803×10^{-13}	ns
		Elevation	2.097	1	ns	Residual	1162	-
							-	
	Mt. Lewis	Mating	4.0038	3	ns	Isofemale line	10.15	ns
						Male	39.95	ns
		Elevation	0.7601	1	ns	Residual	1031.17	-
							-	
Duration	Mt. Edith	Mating	184.84	3	***	Isofemale line	0.08798	*
						Male	0.04592	ns
		Elevation	0.653	1	ns	Residual	3.00321	-
							-	
	Mt. Lewis	Mating	64.142	3	***	Isofemale line	0.7751	ns
						Male	6.507×10^{-14}	ns
		Elevation	0.5598	1	ns	Residual	8.535	-
							-	
Offspring per mating	Mt. Edith	Mating	76.697	3	***	Isofemale line	221.5	***
						Male	106.3	ns
		Elevation	1.9363	1	ns	Residual	958.3	-
							-	
	Mt. Lewis	Mating	18.959	3	***	Isofemale line	48.47	*
						Male	184.27	*
		Elevation	0.051	1	ns	Residual	1012.86	-
							-	
Total Offspring	Mt. Edith	Elevation	1.8045	1	ns	Isofemale Line	1831	***
						Residual	5809	-
	Mt Lewis	Elevation	0.094	1	ns	Isofemale Line	365.1	ns
						Residual	3148.5	-

759 **Table 2.** Broad-sense heritability (H^2) estimates for mating traits for Mount Edith and Mount
760 Lewis lines. Calculated from between (V_b) and within (V_w) line variance components and
761 inbreeding coefficient (F_t) for number of laboratory generations. Significant H^2 estimates are
762 in **bold.**

Mating trait	*Gradient*	V_b	V_w	F_t	H^2
Latency	Mt. Edith	28.61	1162	0.4104	0.029
	Mt. Lewis	10.15	1031.17	0.3317	0.015
Duration	Mt. Edith	0.08798	3.00321	0.4104	**0.035** *
	Mt. Lewis	0.7751	8.535	0.3317	0.126
Offspring per mating	Mt. Edith	221.5	958.3	0.4104	**0.229** ***
	Mt. Lewis	48.47	1012.86	0.3317	**0.069** *
Total offspring	Mt. Edith	1831	5809	0.4104	**0.292** ***
	Mt. Lewis	365.1	3148.5	0.3317	0.157

Figure 1. *Increased mating duration and reduced productivity with successive matings*:
Mean mating latency, mean mating duration and mean number of offspring (productivity) for
all isofemale lines for each successive male mating (1 – 4). Data from Mount Edith lines are
shown in light grey (left), Mount Lewis lines shown in dark grey (right). Error bars indicate
95% confidence intervals.

772
Figure 2. *Mean proportion (%) of males per line reaching each mating*. Mount Edith (left) and
Mount Lewis (right) with 95% confidence intervals.

Figure 3. *Cumulative number of offspring produced per line with successive matings*. Mount
Edith (left) and Mount Lewis (right) with 95% confidence intervals.

Figure 4. *No decline in reproductive success of male offspring from later matings*. Number
of (grand) offspring produced by sons derived from successive paternal matings (1 – 4) when
mating with a single, virgin female (with 95% confidence intervals) for Mount Edith lines.