

1 **Strong Patterns of Intraspecific Variation and Local Adaptation in Great Basin Plants**
2 **Revealed Through a Review of 75 Years of Experiments**

3
4 Owen W. Baughman^{1,2}, Alison C. Agneray¹, Matthew L. Forister³, Francis F. Kilkenny⁴, Erin K.
5 Espeland⁵, Rob Fiegner⁶, Matthew E. Horning⁷, Richard C. Johnson⁸, Thomas N. Kaye⁶, Jeffrey
6 E. Ott⁴, J. Bradley St. Clair⁹, Elizabeth A. Leger¹

7
8 ¹Department of Natural Resources and Environmental Science, University of Nevada, 1664 N.
9 Virginia St., Reno, NV 89557 USA; ²Corresponding author, owbaughman@gmail.com, current
10 address: The Nature Conservancy, 67826-A Hwy. 205, Burns, OR 97720 USA, ³Department of
11 Biology, University of Nevada, Reno, 1664 N. Virginia St., Reno, NV 89557 USA; ⁴USDA
12 Forest Service, Rocky Mountain Research Station, 322 E Front St. Suite 401, Boise, ID 83702
13 USA; ⁵Pest Management Research Unit, USDA-Agricultural Research Service Northern Plains
14 Agricultural Laboratory, 1500 N Central Ave., Sidney, MT 59270 USA; ⁶Institute for Applied
15 Ecology, 563 SW Jefferson Ave., Corvallis, OR 97333 USA; ⁷USDA Forest Service Pacific
16 Northwest Region, Deschutes National Forest, 63095 Deschutes Market Rd., Bend, OR 97701;
17 ⁸Washington State University, PO Box 646240, Pullman, WA 99164 USA; ⁹USDA Forest
18 Service, Pacific Northwest Research Station, 3200 SW Jefferson Way, Corvallis OR, 97331

19 **Keywords:** Local adaptation, phenotypic traits, meta-analysis, common garden, reciprocal
20 transplant, natural selection, intraspecific variation, restoration

21
22 **Running head:** Local adaptation in Great Basin plants

23 **Abstract**

24 Variation in natural selection across heterogeneous landscapes often produces 1) among-
25 population differences in phenotypic traits, 2) trait-by-environment associations, and 3) higher
26 fitness of local populations. Using a broad literature review of common garden studies published
27 between 1941 and 2017, we documented the commonness of these three signatures in plants
28 native to North America's Great Basin, an area of extensive restoration and revegetation efforts,
29 and asked which traits and environmental variables were involved. We also asked, independent
30 of geographic distance, whether populations from more similar environments had more similar
31 traits. From 327 experiments testing 121 taxa in 170 studies, we found 95.1% of 305 experiments
32 reported among-population differences, and 81.4% of 161 experiments reported trait-by-
33 environment associations. Locals showed greater survival in 67% of 24 reciprocal experiments
34 that reported survival, and higher fitness in 90% of 10 reciprocal experiments that reported
35 reproductive output. A meta-analysis on a subset of studies found that variation in eight
36 commonly-measured traits was associated with mean annual precipitation and mean annual
37 temperature at the source location, with notably strong relationships for flowering phenology,
38 leaf size, and survival, among others. Although the Great Basin is sometimes perceived as a
39 region of homogeneous ecosystems, our results demonstrate widespread habitat-related
40 population differentiation and local adaptation. Locally-sourced plants likely harbor adaptations
41 at rates and magnitudes that are immediately relevant to restoration success, and our results
42 suggest that certain key traits and environmental variables should be prioritized in future
43 assessments of plants in this region.

44

45 **Introduction**

46 All plant species have limits to the range of conditions in which they can live, and all but
47 the narrowest endemics grow across environments that vary in biotic and abiotic conditions. This
48 natural complexity has significant impacts on individual survival and reproduction, and thus
49 plant evolution (Loveless and Hamrick, 1984; Linhart and Grant, 1996; Ackerly *et al.*, 2000;
50 Reich *et al.*, 2003). As plants are subject to different conditions associated with their local
51 environment, populations of the same species will experience differential selection pressures
52 (Turesson, 1922; Clausen, Keck and Hiesey, 1948; Antonovics and Bradshaw, 1968; Langlet,
53 1971), creating habitat-correlated intraspecific variation. When this intraspecific variation results
54 in populations that are more fit in their home environment than foreign populations, these
55 populations are considered to be locally adapted (Kawecki and Ebert, 2004; Blanquart *et al.*,
56 2013). The existence of local adaptation is well-established across different organisms and
57 ecosystems, although our synthetic knowledge of this important topic rests on surprisingly few
58 reviews of the subject (e.g. Leimu and Fischer, 2008; Hereford, 2009; Oduor, Leimu and van
59 Kleunen, 2016). Here, we focus on a particular region and ask if plant species share patterns of
60 intraspecific variation and local adaptation, and, across taxa, what functional traits and
61 environmental variables are most important for such patterns in this region. The regional focus
62 provides a strong test of expectations generated from more heterogeneous samples, facilitates
63 comparison of the strength of selection among specific traits, and provides an opportunity to link
64 basic evolutionary patterns with applied concerns.

65 The detection of local adaptation ideally involves reciprocal transplant experiments
66 designed to test for a local advantage across environments (Blanquart *et al.*, 2013; Bucharova,
67 Durka, *et al.*, 2017). However, patterns associated with local adaptation (hereafter, signatures)

68 can be detected in non-reciprocal comparisons of different populations of the same species
69 (Endler, 1986). When populations are locally adapted to environmental variables, we expect to
70 see three basic signatures from common garden experiments: 1) differences among populations
71 in fitness-related traits, 2) correlations between these trait values and environmental or other
72 habitat-related variables, and, if reciprocal transplants have been conducted, 3) higher fitness of
73 local over nonlocal populations in the local environment. Although population differences
74 (signature 1) are necessary for local adaptation, they alone are not sufficient evidence due to
75 factors such as genetic drift, high gene flow, and rapid environmental change, among other
76 factors (Kawecki and Ebert, 2004; Blows and Hoffmann, 2005). While fitness differences in
77 reciprocal transplant experiments (signature 3) are the “gold standard” for detecting local
78 adaptation, there are experimental trade-offs between the number of populations sampled and the
79 ability to do fully reciprocal transplants (Blanquart *et al.*, 2013). Thus, correlative approaches
80 (signature 2) are popular alternatives that can sample many more populations to infer local
81 adaptation (e.g. St Clair, Mandel and Vance-Borland, 2005), though spurious correlations, low
82 sample sizes, or high variability in trait values could over- or under-predict the degree of local
83 adaptation in wild populations using this approach. Given these considerations, separately
84 reporting all three signatures can give an overall picture of the likelihood of within-species
85 variation and potential local adaptation in a region, and is the first step towards a better
86 understanding of variation in the strength and consistency of natural selection (Siepielski,
87 Dibattista and Carlson, 2009).

88 The Great Basin Desert of North America is a ~540,000 km² cold desert landscape
89 characterized by hundreds of internally-draining basin and range formations, which create high
90 spatial and environmental heterogeneity and variability (Tisdale and Hironaka, 1981; Comstock

91 and Ehleringer, 1992). While these are the kinds of conditions that would be expected to result in
92 widespread local adaptation, the flora of the Great Basin is poorly represented in the relatively
93 few reviews on the subject (Leimu and Fischer, 2008; Hereford, 2009; Oduor, Leimu and van
94 Kleunen, 2016), and this has resulted in uncertainty as to the prevalence, magnitude, and
95 importance that local adaptation plays in this large and increasingly imperiled region (United
96 States. House of Representatives. Committee on Appropriations., 2014; Jones, Monaco and
97 Rigby, 2015; Chivers *et al.*, 2016). Gaining a better understanding of local adaptation in the
98 Great Basin is important not only because it is a large, relatively intact floristic region in the
99 Western US, but also because this information has direct impacts on conservation and restoration
100 efforts. Large-scale, seed-based restoration has been very common in the Great Basin for many
101 decades (Pilliod, Welty and Toevs, 2017), and trends in large destructive wildfires (Dennison *et*
102 *al.*, 2014) and other disturbances (Rowland, Suring and Michael, 2010; Davies *et al.*, 2011)
103 ensure even higher demand for restoration efforts in the future. Guided by the various national
104 policies and strategies dating from the 1960s (Richards, Chambers and Ross, 1998) to the present
105 National Seed Strategy (Plant Conservation Alliance, 2015) and Integrated Rangeland Fire
106 Management Strategy (USDOI, 2015), a growing majority of these efforts are using native
107 plants. However, few of the widely-available sources of commercially-produced seeds of native
108 species originate from populations within the Great Basin (Jones and Larson, 2005) or have been
109 selected based on their success in restoring Great Basin habitats (Leger and Baughman, 2015).
110 Further, demand for native seed has always exceeded supply (McArthur and Young, 1999;
111 Johnson *et al.*, 2010), which has resulted in the prioritization of seed quantity and uniformity
112 over population suitability and local adaptation (Meyer, 1997; Richards, Chambers and Ross,
113 1998; Leger and Baughman, 2015). Therefore, it is still uncommon for restorationists in this

114 region to prioritize or even have the option to prioritize the use of local populations, despite
115 growing support of the importance of such practices (Basey, Fant and Kramer, 2015; Espeland *et*
116 *al.*, 2017).

117 Though our understanding of the prevalence and scale of local adaptation in the Great
118 Basin is far from complete, there is an abundant literature of peer-reviewed studies on the plants
119 native to this region spanning over 75 years that have directly measured trait variation between
120 populations via laboratory, greenhouse, or field common gardens and reciprocal transplants.
121 Many of these studies have also tested for correlations between intraspecific variation and
122 environmental variables, and some were designed to detect local adaptation. This research
123 includes studies of germination patterns (e.g. McArthur, Meyer and Weber, 1987; Meyer *et al.*,
124 1995), large genecology experiments (e.g. Erickson, Mandel and Sorenson, 2004; Johnson,
125 Leger and Vance-Borland, 2017), and reciprocal transplants (e.g. Evans and Young, 1990;
126 Barnes, 2009), among other types of studies. This rich literature provides an opportunity to
127 summarize local adaptation and its associated patterns, or signatures (defined above), in this
128 region, as well as describe which phenotypic traits have the strongest signatures of local
129 adaptation.

130 Here, we present results of a broad literature review and subsequent meta-analysis using
131 published studies that compared phenotypic traits of multiple populations of native Great Basin
132 species in one or more common environments. Our first objective was to record published
133 instances of the three expected signatures of local adaptation (population variation, trait-by-
134 environment association, and greater local fitness) within grasses, forbs, shrubs, and deciduous
135 trees native to the Great Basin, asking how common these signatures are, as well as which
136 phenotypic traits and environmental variables were most commonly associated with these

137 signatures. We also present results by taxonomic group, lifeform, lifespan, distribution, and
138 mating system. This first objective encompassed all possible studies, including those that did not
139 provide sufficient details for formal meta-analysis, which allowed us to incorporate the broadest
140 range of studies, including older studies that provided minimal quantitative detail. Our second
141 objective was to examine links between the magnitude of trait and environmental divergence
142 (mean annual precipitation and mean annual temperature) among populations across multiple
143 taxa, for the subset of experiments amenable to this approach, asking whether populations from
144 more similar environments were more similar in phenotypic traits. We also used meta-analysis to
145 ask which traits and environmental variables showed the strongest patterns of association.

146 We expected to find widespread evidence of local adaptation and its signatures in the
147 plants of the Great Basin, and we hypothesize that phenological and size-based traits, which
148 show phenotypic variation in response to climate variation in both plants and animals (e.g.
149 Sheridan and Bickford, 2011; Anderson *et al.*, 2012) and have been observed to be under
150 selection in the Great Basin (Leger and Baughman, 2015), would be important indicators of
151 adaptation in this region. We discuss our results both as a contribution to our general
152 understanding of natural selection in plants, and as an example of evolutionary theory applied to
153 the management and restoration of a large geographic region, where active and ongoing
154 management can benefit from information on intraspecific variation and local adaptation.

155 **Methods**

156 *Literature search*

157 We began by using the search engines Google Scholar and Web of Science to search for
158 combinations of key terms (see additional methods in Supporting Information Appendix 1). In
159 order to be included in our review, a study had to meet all these criteria:

- 160 a) Examined a species that is native within the floristic Great Basin
161 b) Examined and compared more than one population of that species
162 c) Measured at least one phenotypic, physiological, phenological, or other
163 potentially fitness-related trait (e.g. survival; hereafter, trait)
164 d) Measured the trait(s) of the populations in at least one common environment
165 (including laboratories, growth chambers, greenhouses, or outside gardens;
166 hereafter, garden).

167 A plant was determined to be native to the Great Basin if the taxa had at least one
168 occurrence with native status within the floristic Great Basin according to occurrence
169 information from the USDA Plants Database (USDA and NRCS, 2018) and/or the U.S Virtual
170 Herbarium Online (Barkworth *et al.*, 2018). A total of 170 studies published between 1941 and
171 July 2017 were encountered that met these criteria.

172 *Categorization and scoring of literature*

173 All studies meeting our criteria were categorized and scored for each signature. The
174 coordinates of all gardens and populations in each study were recorded or, if possible, generated
175 from localities described in the studies (Supporting Information Appendix 1). For each study, we
176 then noted these 15 characteristics: the year published, year(s) of plant material collection,
177 year(s) of experimentation, number of years reported, taxa (genus, species, subspecies), life
178 history traits (taxonomic status, lifeform, geographic range, life span, breeding system),
179 experiment type (laboratory, greenhouse, common garden, reciprocal transplant), number of
180 gardens, number of populations tested, which generation of material was used, and whether or
181 not experimenters attempted to control for maternal effects prior to testing (Supporting
182 Information Appendix 1). Life history traits were compiled for each taxon from the USDA Plants

183 Database as well as from published literature (Supporting Information Appendix 1). Each taxon
184 (subspecies level, if given) was entered separately for studies addressing multiple taxa. In studies
185 where more than one experiment was performed, and the experiments differed in the experiment
186 type (defined above), the identity of the populations being compared, and/or the generation of
187 material used, they were entered as separate experiments. In cases where the list of tested
188 populations was identical among multiple published studies, and these materials came from the
189 same collections, these experiments were entered separately if the garden type or location(s)
190 differed among the studies or if authors separately published different traits from the same
191 gardens, ensuring that no trait was recorded twice for the same set of populations in the same
192 garden. In cases where the list of tested populations did not completely overlap between studies,
193 even if some from each study arose from the same collections, they were entered separately.
194 These methods carefully emphasized the inclusion of the greatest number of relevant
195 experiments and traits without duplication, but nonetheless resulted in some non-independence
196 between some experiments. A total 327 taxa-specific entries (hereafter, experiments) were
197 generated from the 170 published studies (Supporting Information Appendix 2).

198 The first two expected signatures of local adaptation were scored using a Yes/No
199 designation for each experiment which considered all measured phenotypic traits. A score of
200 “Yes”, or, in the absence of supporting statistical evidence, “Authors claim Yes”, was given
201 when at least one measured trait significantly demonstrated the signature for at least two
202 populations, and a score of “No” or “Authors claim No” was given when the signature was not
203 detected between any pair of populations (Supporting Information Appendix 1). In addition, each
204 of the measured and reported traits and environmental variables were scored (hereafter, trait
205 scores) in the same way for each signature. Of the 327 experiments, 305 (93.3%) met the criteria

206 to score for among-population variation (signature 1) and 161 (49.5%) met the criteria to score
207 for trait-by-environment association (signature 2). Pearson's chi-squared tests were used to
208 determine if there were differences in signatures 1 and 2 among plants with different life-history
209 traits, using totals from both "Yes/No" and "Authors Claim Yes/No" results, excluding any life
210 history groups represented by less than 10 experiments.

211 To score whether there was higher fitness of a local population in a common garden
212 (hereafter, signature 3), only experiments in which outdoor reciprocal transplants or common
213 gardens were performed using a local population in at least one garden were considered
214 (Supporting Information Appendix 1). Additionally, the experiment had to measure a fitness-
215 relevant response: survival, reproductive output (number of seeds or flowers, or other
216 reproductive output), a fitness index (a combination of several size and production traits), or total
217 aboveground biomass. Each experiment was assigned a composite score to fully capture
218 variation in the performance of each garden's local population, across multiple gardens as well
219 as through multiple sampling dates (Supporting Information Appendix 1). The five possible
220 composite scores were "Yes for all gardens at all times", "Yes for all gardens at some times",
221 "Yes for some gardens at all times", "Yes for some gardens at some times", and "No for all
222 gardens at all times". These scores refer only to those gardens within each experiment that
223 included their own local population. Of the 326 experiments, 27 (8.3%) were appropriate for this
224 scoring. This scoring provides an estimate of the commonness of higher local fitness, but it is not
225 a measure of the importance of the difference per se. For example, a fitness difference could
226 occur uncommonly, but have a large impact on population trajectories (i.e. large differences in
227 survival after a rare drought event).

228 Our dataset, which had uneven numbers of experiments representing each species,

229 contained the possibility of bias associated with highly-studied taxa influencing patterns more
230 than less-studied taxa. To ask how this affected overall results, we compared tallies of all scores
231 without correcting for multiple experiments per species to tallies using an average score for each
232 species for each signature. To generate these average scores for signature 1 and 2, we totaled all
233 “Yes” and “Authors claim Yes” scores for each species and divided by the total number of scores
234 (all Ys plus all Ns) for that species. For signature 3, all forms of “Yes” (all but “No for all
235 gardens at all times”) were totaled into a Y and divided by the total number of scores. Then, we
236 averaged these per-species scores to re-calculate overall effects in which each species was
237 represented only once, and compared the results of the different averaging methods for each
238 signature.

239 *Quantitative comparison of trait-by-environment associations*

240 As a complement to the survey of author-reported results described above, we conducted
241 a further, quantitative analysis of trait and climate values. Specifically, to examine associations
242 between the differences in trait values and the differences in environmental and geographic
243 distance among population origins, we utilized experiments from which population-specific trait
244 data and geographic coordinates could be extracted or obtained through author contact. Data
245 from laboratory and greenhouse experiments were not considered for this extraction. First, we
246 identified the most commonly measured traits across studies, which were then manually
247 extracted from text, tables, or graphical data (Supporting Information Appendix 1). Next, we
248 extracted trait data from the latest sampling date for which the most populations at the most
249 gardens were represented, and if multiple treatments were used, we only extracted data for the
250 author-defined ‘control’ treatment. However, if no control was defined, we used the treatment
251 that was the most unaltered or representative of the garden environment (e.g. unweeded, or

252 unwatered). For each population/trait combination, we used either author-provided mean values
253 or calculated a mean trait value from available data. Rather than averaging values across gardens,
254 data, data from each garden location within each experiment was extracted separately and
255 considered its own sample. We did this because it is not uncommon for traits to be expressed
256 differently in different common garden locations (e.g. Johnson, Leger and Vance-Borland,
257 2017). Finally, we generated 30-year annual precipitation and mean annual temperature values
258 for each population's location of origin using the ClimateNA v5.10 software package based on
259 methodology described by Wang et al. (2016). These 30-year averages are calculated every 10
260 years (i.e. 1951-1980, 1961-1990, etc.). Because studies took place at many times over the last
261 75 years, we used the most proximate climate normal for each experiment that did not include or
262 surpass the years during which the experiment's populations were collected (Supporting
263 Information Appendix 2).

264 To reduce the likelihood of spurious correlations or false negative results, we limited this
265 dataset to traits measured in at least 5 populations in at least 20 common garden locations (mean
266 locations per trait: 34.4; range: 21-46), resulting in 81 locations (from 56 experiments) that
267 measured at least one of eight frequently-measured phenotypic traits (Table 1). Within each
268 location, we calculated pairwise Euclidean distances for each trait value, climate factor, and
269 geographic distance for every possible pair of populations. Geographic distances were generated
270 using the earth.dist function in fossil package (Vavrek, 2011) in the statistical computing
271 environment R (R Core Team, 2017). Then, partial Mantel tests were used to compare pairwise
272 trait and climate distances for each experiment while controlling for geographic distances, using
273 the vegan package (Oksanen *et al.*, 2018) in R (R Core Team, 2017). We used the metacor.DSL
274 function in the metacor package (Laliberté, 2011) to generate an overall effect size (partial

275 correlation) and upper and lower confidence intervals for each combination of trait and
276 environmental variable. Lastly, to better understand effect sizes for a subset of species, we ran
277 simple linear regression analyses for each location, comparing average trait values and
278 environmental values to generate a slope that estimated trait change per unit change in climate
279 factors. Experiments with R^2 values of 0.2 or less were excluded from this particular analysis,
280 and the median slope across experiments was retained as an estimate of the trait-by-environment
281 relationship. The arbitrary cutoff ($R^2 = 0.2$) for this step was used simply as a way to focus on
282 and report effect sizes from some of the stronger biological relationships that could be of
283 particular interest to managers, restoration practitioners and evolutionary ecologists. Due to
284 limited sample sizes for factors such as lifeform, mating system, geographic distribution, etc., we
285 did not include these factors in any of the quantitative analyses, but present lifeform (shrub,
286 grass, or forb) information for each trait response as additional results in the Supporting
287 Information Appendix 3.

288 **Results**

289 *Summary of reviewed literature*

290 Our literature search revealed 170 published studies that measured trait responses from
291 more than one population in at least one common environment, resulting in 327 separate
292 experiments involving 121 taxa of 104 species of grasses, shrubs, forbs, and deciduous trees
293 (Fig. 1). These experiments represent approximately 3,234 unique populations tested in
294 approximately 208 outdoor garden locations (Fig. 2) and 154 indoor lab or greenhouse
295 experiments. Grasses accounted for 21.0% of the taxa and 40.2% of the experiments, forbs
296 composed 50.8% of the taxa and 30.7% of experiments, shrubs 26.6% of the taxa and 28.5% of
297 experiments, and deciduous trees accounted for only 1.6% of taxa and 0.6% of experiments (Fig.

298 1A). Experiments were most commonly conducted in non-reciprocal outdoor common gardens
299 (47.5%) or in the laboratory (31.9%), with fewer conducted in greenhouses (15.3%) or in
300 reciprocal outdoor gardens (5.2%, Fig. 1B). For experiments in outdoor gardens, the median
301 number of gardens per experiment across lifeform ranged from 1 (grasses, shrubs, and trees) to 2
302 (forbs) for non-reciprocal gardens, and from 2 (grasses and forbs) to 4 (shrubs) for reciprocal
303 gardens . Overall, the median number of populations tested in each experiment was 5 (range= 2 -
304 193, IQR = 3 – 11.5, Fig. 1C), and was slightly lower for shrubs (median = 4, range = 2 – 111,
305 IQR = 2 - 8) than grasses (median = 6, range = 2 – 193, IQR = 3 - 12.25), forbs (median = 6,
306 range = 2 – 67, IQR = 3 – 10.25), and trees (median = 7, range = 5 – 9, IQR = 6 – 8).

307 Experiments took place between 1940 and 2015, with collections from native stands
308 occurring between 1938 and 2013 (Fig. 3A). One quarter of the experiments (24.5%) reported
309 only early germination and seedling stages of plants (generally less than 0.5 years), while the
310 remaining experiments (75.5%) reported study periods ranging from 0.5 to 17 years, with an
311 average of 2.1 years (Fig. 3B, C). Average pairwise geographic distance among populations per
312 experiment for the 91% of experiments for which coordinates were available was $351 \text{ km} \pm 20$
313 SE, with a range from 610 m to 2,551 km. Most experiments were conducted on taxa with
314 regional distributions, perennial species, grasses, and outcrossing species; very few annuals,
315 endemic species, or selfing species were represented (Fig. 4). Over half of experiments (58.6%)
316 tested plants grown directly from wild-collected seeds (or the seed of wild collected adults),
317 16.9% tested wild-collected adults, 13% tested materials with mixed generations since collection,
318 6.7% tested 1st or 2nd generation descendants of wild collected seeds, 0.3% tested only cultivars,
319 and 4.3% did not provide enough information to determine.

320 *Among-population variation*

321 Of the 305 experiments appropriate for addressing among-population trait variation
322 (signature 1), 290 (95.1%) experiments reported finding variation among populations in at least
323 one phenotypic trait, with 230 (75.4%) of these 290 reporting significant variation, and 60
324 (19.6%) claiming such variation in the absence of any supporting statistics (Fig. 4A). Only 12
325 (3.9%) experiments reported no such differentiation in any trait after statistically testing for it,
326 and 3 (1%) claimed no such variation without presenting statistical evidence. When categorized
327 by basic life history traits, several differences appeared among groups. Eudicots exceeded
328 monocots (the majority of which were grasses) in the degree of population differentiation ($X_1^2 =$
329 7, $P = 0.0081$), and, similarly, forbs and shrubs had more population differentiation than grasses
330 ($X_2^2 = 8.05$, $P = 0.0143$). There were no significant differences in signature 1 among plants with
331 different geographic distributions, life span, or breeding systems.

332 A total of 1,465 trait scores were recorded from the 305 experiments appropriate for
333 addressing signature 1. Frequently-measured traits (20 or more experiments) that had differences
334 between populations in over 75% of experiments (with or without supporting statistics) were
335 floral structure, vigor, emergence, plant size, number of leaves, plant structure, shoot biomass,
336 leaf structure, and number of inflorescences (Fig. 5).

337 *Trait-by-environment associations*

338 Of the 161 experiments appropriate for testing trait-by-environment associations
339 (signature 2), 131 (81.4%) reported associations for at least one comparison, with 81 (50.3%)
340 supported by statistical tests and 50 (31.1%) supported by claims in the absence of statistics (Fig.
341 4B). Conversely, 13 (8.1%) of experiments reported no such correlations after having
342 statistically tested for it, and 17 (10.6%) reported no such correlations but lacked any supporting
343 statistics. There were no significant differences in the commonness of trait-by-environment

344 associations for taxonomic status, lifeform, geographic distribution, or breeding system, but
345 perennials (both long-lived and short-lived) had more frequent correlations between traits and
346 environment than did annuals or short-lived perennials ($X_3^2 = 8.08$, $P = 0.0444$).

347 A total of 592 trait scores were recorded from the 161 experiments appropriate for
348 addressing signature 2 (Fig. 6A). Frequently-measured traits (20 or more experiments) that were
349 correlated with environmental variables in over 75% of experiments (with or without supporting
350 statistics) were multivariate trait axes, floral structure, and germination date. Every remaining
351 trait that was measured in >15 experiments was correlated with environmental characteristics in
352 over 50% of experiments, and many, including leaf length, survival, flowering date, and leaf
353 structure, were correlated with environmental variables in $\geq 70\%$ of experiments.

354 A total of 426 environmental variable scores were recorded from the 161 experiments
355 appropriate for addressing signature 2 (Fig. 6B). Of the variables most frequently reported as
356 correlated with plant traits, many categorical variables or composite metrics made this list, with
357 seed zones, ecoregions, multivariate environmental axes, and habitat classifications topping the
358 list of important environmental variables (important in > 84% of experiments that reported
359 them). Additionally, derived climate metrics (such as climate continentality, heat/moisture index,
360 potential evapotranspiration, etc.), climate seasonality, and history of invasive species presence
361 were correlated with plant traits in over 75% of studies that reported them.

362 *Higher local performance in a local common garden*

363 The 27 experiments that were suitable for detecting higher fitness of a local population in
364 a local garden (signature 3) generated 39 scores (some experiments measured multiple fitness
365 traits), with 27 scores (69.2%) reporting signature 3 for at least one fitness trait in at least one of
366 the tested gardens during at least one sampling date, and the remaining 12 scores (30.8%) not

367 reporting signature 3 at any point (Fig. 4C). Thirty-two of the 39 scores (82%) were generated
368 from experiments with more than one garden. Survival was the most frequently measured fitness
369 trait in these experiments, reported in 24 of the 27 experiments, followed by reproduction (10),
370 biomass (3), and fitness indices (2). Incidence of the local-does-best pattern was highest in
371 experiments that directly measured reproductive output, with 90% reporting higher values for
372 locals at some point in an experiment, followed by survival (67%), fitness indices that
373 incorporated biomass (50%), and biomass measures (33%). For experiments in which only
374 “some” gardens showed local-does-best patterns (Fig. 4C, hashed bars), the percentage of
375 gardens showing this trend was 40%, 50%, and 40% for reproduction, survival, and biomass
376 traits, respectively (not shown). For experiments in which only “some” sampling dates showed
377 local-does-best patterns (gray bars), the percentage of sampling dates showing this trend was
378 56%, 47%, and 25% for reproduction, survival, and biomass traits, respectively (not shown).

379 *Considering possible biases: highly-studied species and maternal effects*

380 The number of experiments per species in our dataset ranged from 1 (52 species) to 25
381 (*Artemisia tridentata*), with a median of 1 (IQR = 1 – 4). The most highly-represented species
382 were *Artemisia tridentata* (25 experiments), *Elymus elymoides* (24), *Ericameria nauseosa* (17),
383 *Achnatherum hymenoides* (17), *Krascheninnikovia lanata* (13), *Pascopyrum smithii* (11),
384 *Atriplex canescens* (9), *Leymus cinereus* (9), and *Poa secunda*, (8). Results in which scores were
385 averaged for each species (see methods) were similar to uncorrected results: signature 1 was 4%
386 higher when corrected (98% vs. 94%), signature 2 was 1% lower when corrected (79% vs. 80%),
387 and signature 3 was 8% higher when corrected (78% vs. 70%). Thus, uncorrected calculations
388 were used throughout our study.

389 Only 19 experiments (5.8%) used an experimental design that could control for maternal

390 effects (e.g. growing all populations for a generation in a common environment before initiating
391 an experiment). An additional 30 experiments (9.2%) were unclear on this point, and the
392 remaining 278 (85%) experimented directly on populations differing in maternal environment.
393 The incidence of population differences (signature 1) was 100% in the 16 experiments that
394 moderated maternal effects, 95% for the 259 that did not make an attempt, and 97% for the 30
395 which were unclear. Too few of the experiments that attempted to control for maternal effects
396 were appropriate for measuring signature 2 (4 experiments) and signature 3 (1 experiment) to
397 compare incidences of these signatures.

398 *Quantitative comparison of trait-by-environment associations*

399 Overall, we found positive relationships between the magnitude of differences among
400 populations in all eight phenotypic traits and the magnitude of differences between MAT and
401 MAP at the collection locations (Fig. 7). The strongest relationship was observed between
402 differences in flowering time and differences in MAT, and leaf size also showed a strong
403 relationship with MAT. Multiple strong relationships were observed between trait/environment
404 divergence for MAP, with leaf size, survival, shoot mass, inflorescence number, and flowering
405 time all showing strongly positive relationships for grasses, forbs, and shrubs. (Fig. 7, Supporting
406 Information Appendix 3). Regression analyses demonstrated that, for the 15 common garden
407 locations in which strong flowering time and MAT relationships were observed, each degree
408 change in MAT was associated with a median change of 3.5 days (IQR = 1.2 - 5.3) in flowering
409 time. Small sample sizes (few experiments that could be included in the analyses) and challenges
410 with interpreting changes in physical traits across species of various shapes and sizes precluded
411 the presentation of estimates of this nature for the other trait-by-environment relationships.

412 **Discussion**

413 Our results represent the most extensive review of intraspecific variation and local
414 adaptation for plants native to the floristic Great Basin, a region comprised of largely continuous
415 but increasingly imperiled arid and semi-arid plant communities (Davies *et al.*, 2011; Finch *et*
416 *al.*, 2016). Additionally, they represent a significant addition to the noteworthy though relatively
417 small number of reviews investigating this topic in a manner that identifies individual traits and
418 environmental factors involved. We found that Great Basin plant species contain large amounts
419 of intraspecific diversity in a wide range of phenotypic traits, that differences in these phenotypic
420 traits are often associated with the heterogeneous environments of origin, and that differences
421 among populations are commonly relevant to outplanting fitness. The cascading importance of
422 intraspecific variation for the structure, functioning, and biodiversity of communities and
423 ecosystems can be considerable (Bolnick *et al.*, 2011; Bucharova *et al.*, 2016), and may equal or
424 exceed the importance of species diversity (Des Roches *et al.*, 2018). Our quantification of local
425 adaptation and trait-environment associations should serve as encouragement to seriously
426 consider intraspecific diversity in native plant materials used in restoration and conservation in
427 this region throughout the selection, evaluation, and development process (Basey, Fant and
428 Kramer, 2015). The results reported here should also serve as a cautionary note to restoration
429 approaches that focus on only a few specific traits or search for general-purpose genotypes. Our
430 results suggest that, in the absence of species-specific information to the contrary, it is reasonable
431 to assume that local adaptation is present in this region, and that locally-sourced populations
432 would outperform non-local populations a majority of the time.

433 Our investigation encompassed 170 studies published between 1941 and 2017 in which
434 over 3,230 unique populations of 104 native Great Basin plant species were compared in 327
435 experiments, ranging from laboratory germination trials to multiple-year common gardens and

436 reciprocal transplants. The great majority (95%) found differences between populations
437 (signature 1) in the majority of traits measured in a common environment, which indicates that
438 different traits are variable among populations, at both small and large geographic scales.
439 Additionally, a clear majority (81.4%) of experiments found trait-by-collection environment
440 associations (signature 2), suggesting that intraspecific variation is frequently an adaptive
441 outcome of natural selection in heterogeneous environments (Linhart and Grant, 1996; Reich *et*
442 *al.*, 2003). In experiments suitable for detecting local performance advantages (signature 3), local
443 populations had higher performance (measured by differences in reproductive output, survival,
444 and biomass) than nonlocal populations more often than not (69.2%), and this was particularly
445 true when researchers reported traits related to reproductive output (90%). We used a vote-
446 counting method to summarize results for our broadest pool of studies, allowing us to
447 incorporate a wealth of older studies for which quantitative details were not available. Results
448 from a vote-counting approach can sometimes differ from results of meta-analysis, as vote-
449 counting does not incorporate the same level of detail about factors such as study size or effect
450 size (Combs *et al.*, 2011). However, in our study, the overall incidence of “local does best” in the
451 Great Basin is similar to other reviews that have found local adaptation to be commonplace, but
452 not ubiquitous. In a review of local adaptation in plants that compared survival, reproduction,
453 biomass and germination traits in reciprocal transplants, Leimu and Fischer (2008) found that
454 local plants outperformed non-local ones in 71% of 35 published experiments. Similarly,
455 Hereford (2009) quantified local adaptation in 70 published studies (50 of them plants), reporting
456 only survival or reproductive traits, and found evidence of local adaptation in 65-71% of
457 experiments. Our results indicated that the strongest indication of local adaptation came from
458 experiments that directly measured reproductive output, and that using biomass as a fitness proxy

459 may not be an effective way to compare relative performance in the Great Basin. This is
460 consistent with a previous study that demonstrated selection for smaller, rather than larger,
461 individuals in disturbed arid systems (Kulpa and Leger, 2013). Literature reviews conducted
462 across biomes may occlude regionally-important trait differentiation and mask patterns of local
463 adaptation, as we might expect, for example, biomass to be more strongly linked to fitness in
464 regions where light is a contested resource (Espeland, Johnson and Horning, 2017).

465 There are many processes that can reduce or prevent the development of local adaptation,
466 such as the lack of divergent selection between sites, high gene flow, rapid or extreme
467 environmental change, high phenotypic plasticity, and/or low genetic diversity (Sultan and
468 Spencer, 2002; Kawecki and Ebert, 2004; Blows and Hoffmann, 2005). The high incidence of
469 intraspecific variation, much of it habitat-correlated, that we found in the literature confirms that
470 divergent selection by heterogeneous environments is the norm for species native to the Great
471 Basin, presumably outweighing the balancing effects of gene flow and genetic drift. Key
472 environmental factors in the Great Basin such as fire frequency, grazing regimes, resource
473 availability, and climate are certainly being altered to varying degrees by invasive species
474 introductions, changing land uses, and climate change, and it can be argued that such changes
475 could outpace the ability of local populations to remain adapted to their surroundings (Jones and
476 Monaco, 2009; Breed *et al.*, 2013; Havens *et al.*, 2015; Kilkenny, 2015). However, our analysis
477 also demonstrated relatively high instances of trait correlations with relatively recent
478 disturbances such as invasive species introductions. Rapid evolution in response to invasive
479 species (Oduor, 2013) and other anthropogenic changes (Hoffmann and Sgrò, 2011; Franks,
480 Weber and Aitken, 2014) has been documented for many species, indicating that local adaptation
481 can evolve rapidly in some circumstances.

482 Some traits and environmental characteristics stood out as particularly important
483 indicators of local adaptation and its signatures across the studied taxa. For example, in our
484 quantitative comparison of divergence in traits and environments, flowering phenology was
485 strongly affected by MAT, with a median change of 3.5 days in flowering time per degree
486 change in MAT of collection origin. Flowering phenology, along with germination phenology,
487 were also in the top tier of frequently measured traits that showed significant correlations with
488 environmental variables, consistent with other studies that have shown reproductive (Bucharova,
489 Michalski, *et al.*, 2017) and germination (Donohue *et al.*, 2010) phenology to be an important
490 response to environmental variation. Leaf size is also an important adaptive response to
491 differences in temperature globally (Wright *et al.*, 2017), and in concert with this, we saw overall
492 positive responses to MAP and MAT for leaf size in our analyses as well as frequent trait-by-
493 environment associations in the literature. Floral structure, which has important adaptive
494 significance for angiosperms (Harder and Barrett, 2007; Armbruster, 2014), was among the most
495 frequent traits scored for among-population variation and trait-by-environment interactions.
496 Seasonality of precipitation, which varies in this region depending on summer rainfall
497 (Comstock and Ehleringer, 1992), was more predictive of trait variation overall than was mean
498 annual precipitation (signature 2). In our quantitative comparisons, differences in MAP values
499 were important for multiple phenotypic traits, including leaf size, shoot mass, reproductive
500 output, and flowering phenology, in addition to being important for overall plant survival. Larger
501 scale environmental descriptors, such as ecoregions and seed transfer zones, universally
502 demonstrated signature 2, likely because they were developed based on climate/soil/vegetation
503 associations or, in the case of seed transfer zones, developed based on trait-by-environment
504 correlations. As found in other reviews (Geber and Griffen, 2003), physiological traits,

505 phytochemical traits, and root traits were not measured as frequently as other traits, and though
506 these did not show as frequent associations with environmental characteristics as other traits,
507 they are known to vary across environments in some systems (Reich *et al.*, 2003). Additional
508 studies of these traits in the Great Basin would be informative and could reveal different patterns
509 than those observed here.

510 As in any review and analysis of published papers, there are elements of our design that
511 were difficult to control. For example, consistent with other reviews (Gibson *et al.*, 2016), the
512 vast majority of studies involved wild-collected plants or seeds, and thus maternal environment
513 effects almost certainly affected some results (e.g. Bischoff and Müller-Schärer, 2010; Espeland
514 *et al.*, 2016). Additionally, though the majority of populations tested in the literature were from
515 western states, some of the populations compared in the literature were collected from well
516 outside of the Great Basin, which increased the likelihood of observing local adaptation in these
517 species. However, understanding patterns of intraspecific variation across the full range of the
518 species native to the Great Basin is pertinent because it has been common (and for some species,
519 ubiquitous) to utilize sources of native species originating from outside the Great Basin to use for
520 restoration within the Great Basin (Jones and Larson, 2005). Finally, the scores and percentages
521 for each of the signatures used throughout this study are uncorrected for phylogeny, as is our
522 pairwise trait/environment analysis, and calculated such that each experiment is weighed equally.
523 This introduces the possibility for phylogenetic biases, in which closely related taxa represented
524 by many experiments affect the results more than less frequently studied taxa or groups of taxa.
525 Though we did not conduct phylogenetic corrections for relatedness among taxa (Harvey and
526 Pagel, 1991; de Bello *et al.*, 2015), our results were essentially identical for signatures 1-3 when
527 we averaged results across species (scores differed by +3%, -1%, and +8%, respectively),

528 suggesting that our lack of phylogenetic corrections are not unduly affecting our results. We
529 present all species-specific information in Supporting Information Appendix 2 and available
530 datasets section of the electronic supplementary material for further review.

531 Current approaches to seed sourcing in restoration and conservation include genetic (e.g.
532 Williams, Nevill and Krauss, 2014), genecological (e.g. Johnson, Leger and Vance-Borland,
533 2017), local-only (e.g. Erickson *et al.*, 2017), predictive (e.g. Prober *et al.*, 2015), and agronomic
534 (e.g. United States. House of Representatives. Committee on Appropriations., 2014)) strategies,
535 as well as strategies mixing several of these viewpoints (i.e. Rice and Emery, 2003; Rogers and
536 Montalvo, 2004; Breed *et al.*, 2013; Havens *et al.*, 2015; Bucharova *et al.*, 2018). These
537 approaches vary in the degree to which they meet the needs of seed producers and land managers
538 while balancing population differences that stem from adaptive evolution in different
539 environments. The prevalence of local adaptation and its signatures found in our study justify
540 and support incorporating existing best-practices (e.g. Basey, Fant and Kramer, 2015; Espeland
541 *et al.*, 2017) for capturing and preserving important intraspecific variation into seed sourcing and
542 plant production systems. For example, our results demonstrated a strong relationship between
543 flowering time and MAT, so it would be wise to collect materials for research, evaluation, and
544 testing from populations that vary in MAT, to collect seeds at multiple times to fully capture
545 population variation in flowering time, and ensure that seeds are not transferred during
546 restoration among sites that differ strongly in these characteristics. On the production side, best
547 practices for seed harvesting should include methods that avoid inadvertent selection on
548 flowering time, either for reduced variation or for a directional shift away from the wild
549 condition. Similarly, emergence date was correlated with environmental variation in many
550 plants, so testing in common gardens should involve seeding trials in place of or in addition to

551 using transplants, and evaluation trials should guard against inadvertent selection on emergence
552 timing by randomly, rather than systematically, selecting individuals to use in transplant
553 experiments. These examples are not exhaustive, but demonstrate how evidence revealed by this
554 study regarding which traits and environmental factors are generally involved in adaptation in
555 this region can be used to improve approaches to seed sourcing and restoration. Finally, we
556 acknowledge that ours is not the first review and meta-analysis to affirm an abundance of
557 intraspecific variation and local adaptation in plants. However, our focus on the Great Basin is
558 important, because the large and frequent yet commonly unsuccessful restoration efforts
559 occurring in this region have lagged behind those of other regions with respect to recognizing the
560 importance of intraspecific variation and local adaptation on outplanting success.

561 **Conclusions**

562 Reestablishing and maintaining native plant communities in arid regions has proven
563 challenging (Svejcar *et al.*, 2017), and the lack of practical knowledge guiding more appropriate
564 selection of seed sources is a major barrier (Friggens *et al.*, 2012; Gibson *et al.*, 2016). The
565 forestry industry has long adopted the principles of local adaptation in their reforestation
566 guidelines with great success (Matyas, 1996; Johnson *et al.*, 2004; Aitken and Bemmels, 2016),
567 and similar approaches to restoration in the rangelands of the Great Basin may also increase
568 success as our data support similarly high levels of population differentiation within grass, forb
569 and shrub life history groups. Our results, including both a qualitative literature survey and a
570 quantitative meta-analysis, could benefit from future work using additional techniques to explore
571 spatial structure (e.g. Griffith and Peres-Neto, 2006) and the relative importance of geographic
572 distance and environmental variation, especially as additional studies become available in the
573 literature. Nevertheless, our results as they currently stand are in agreement with observations of

574 abundant local adaptation in plant populations world-wide, and further, we identified particular
575 phenotypic traits (flowering and germination phenology, floral structures, leaf size, biomass,
576 survival, and reproductive output), environmental characteristics (MAT, MAP, climate metrics,
577 seasonality), and habitat classifications and site history (seed zones, ecoregions, history of
578 invasive species) that were important predictors of local adaptation in plants native to the Great
579 Basin floristic region. Given the speed and severity with which natural communities are being
580 altered by anthropogenic factors, the application of an evolutionary perspective to restoration
581 ecology is more important than ever. Adjusting seed-selection priorities to account for the
582 existence of locally adapted, intraspecific variation in the Great Basin will promote the
583 maintenance and recovery of resilient, self-sustaining vegetation communities in this region
584 (Meyer, 1997; Lesica and Allendorf, 1999; Rogers and Montalvo, 2004; Broadhurst *et al.*, 2008;
585 Vander Mijnsbrugge, Bischoff and Smith, 2010).

586 **Acknowledgements**

587 We would like to acknowledge Vicki Thill and Sage Ellis for many hours of extracting
588 coordinates and trait data, as well as Susan Meyer, Andrea Kramer, Tom Jones, Vicky Erickson,
589 Allan Stevens, Dan Atwater, Clinton Shock, Jessica Irwin, Huixuan Liao, and David Solance
590 Smith for providing information and data not available in their publications. We also thank
591 several anonymous reviewers for their insightful suggestions that greatly improved this work.

592 **Dedication**

593 We would like to dedicate this paper to the memory of our co-author Dr. Erin K.
594 Espeland, friend and collaborator to all of us, who worked on this manuscript. Erin's light and
595 life will never be forgotten by those who knew her, and we want to recognize her creative
596 contributions to the field of plant ecology, including this effort. Erin is dearly missed.

597 **Funding**

598 This project was funded by a grant from the United States Department of the Interior
599 Great Basin Landscape Conservation Cooperative (2016-Kilkenny/Leger), which provided
600 support for EAL and OWB. Additionally, ACA was supported by a grant from the United States
601 Department of Agriculture National Institute of Food and Agriculture; EE was supported by
602 United States congressional appropriation 3032-21220-002-00-D; FFK and JEO were supported
603 by the Great Basin Native Plant Project, the United States Department of the Interior Bureau of
604 Land Management, and the United States Department of Agriculture Forest Service; JBS and
605 MEH were supported by the United States Department of Agriculture Forest Service; MLF was
606 supported by a Trevor James McMinn fellowship; RCJ was supported by the Great Basin Native
607 Plant Project; RF was supported by the Institute for Applied Ecology; and TMK was supported
608 by the United States Department of the Interior Bureau of Land Management and the Institute for
609 Applied Ecology

610 **Data accessibility**

611 Raw datasets and statistical code supporting this study (Baughman *et al.*, In Review) have been
612 deposited at Dryad, [DOI: TBD]

613 **Authors' contributions**

614 EAL, OWB, FFK, EKE, RF, TNK, and JBS conceived and designed the study; OWB
615 conducted the literature search; OWB, ACA, FFK, JEO, RCJ, and JBS categorized, compiled
616 and extracted data; OWB, EAL, FFK, ACA and MLF analyzed data; OWB, EAL, and ACA
617 drafted the manuscript; all authors critically revised the manuscript for important intellectual
618 content and approved of the version to be published.

619

620 **References**

- 621
- 622 Ackerly, D. D. *et al.* (2000) ‘The Evolution of Plant Ecophysiological Traits : Recent Advances
623 and Future Directions’, *AIBS Bulletin*, pp. 979–995. doi: 10.1641/0006-
624 3568(2000)050[0979:teopet]2.0.co;2.
- 625 Aitken, S. N. and Bemmels, J. B. (2016) ‘Time to get moving: Assisted gene flow of forest
626 trees’, *Evolutionary Applications*, pp. 271–290. doi: 10.1111/eva.12293.
- 627 Anderson, J. T. *et al.* (2012) ‘Phenotypic plasticity and adaptive evolution contribute to
628 advancing flowering phenology in response to climate change’, *Proceedings of the Royal Society*
629 *of London B: Biological Sciences*. doi: 10.1098/rspb.2012.1051.
- 630 Antonovics, J. and Bradshaw, A. (1968) ‘Evolution in closely adjacent plant populations’,
631 *Heredity*, 23(4), pp. 507–524. doi: 10.1038/hdy.1970.36.
- 632 Armbruster, W. (2014) ‘Floral specialization and angiosperm diversity: phenotypic divergence,
633 fitness trade-offs and realized pollination accuracy’, *AoB Plants*, 6.
- 634 Barkworth, M. *et al.* (2018) ‘US Virtual Herbarium’. Available at: <http://usvhproject.org/#/>.
- 635 Barnes, M. (2009) ‘The effect of plant source location on restoration success: a reciprocal
636 transplant experiment with winterfat (*Krascheninnikovia lanata*)’, *Doctoral Dissertation. The*
637 *University of New Mexico*. Available at: http://digitalrepository.unm.edu/biol_etds/4 (Accessed:
638 8 January 2018).
- 639 Basey, A. C., Fant, J. B. and Kramer, A. T. (2015) ‘Producing native plant materials for
640 restoration: 10 rules to collect and maintain genetic diversity’, *Native Plants Journal*, 16(1), pp.
641 37–53. doi: 10.3368/npj.16.1.37.
- 642 Baughman, O. *et al.* (In Review) ‘Data from: Strong Patterns of Intraspecific Variation and
643 Local Adaptation in Great Basin Plants Revealed Through 75 Years of Experiments’, *Dryad*
644 *Digital Repository*. doi: TBD.
- 645 de Bello, F. *et al.* (2015) ‘On the need for phylogenetic “corrections” in functional trait-based
646 approaches’, *Folia Geobotanica*, 50, pp. 349–357. doi: 10.1007/s12224-015-9228-6.

- 647 Bischoff, A. and Müller-Schärer, H. (2010) ‘Testing population differentiation in plant species -
648 How important are environmental maternal effects’, *Oikos*, 119(3), pp. 445–454. doi:
649 10.1111/j.1600-0706.2009.17776.x.
- 650 Blanquart, F. *et al.* (2013) ‘A practical guide to measuring local adaptation’, *Ecology Letters*, pp.
651 1195–1205. doi: 10.1111/ele.12150.
- 652 Blows, M. W. and Hoffmann, A. A. (2005) ‘A reassessment of genetic limits to evolutionary
653 change’, *Ecology*, pp. 1371–1384. doi: 10.1890/04-1209.
- 654 Bolnick, D. I. *et al.* (2011) ‘Why intraspecific trait variation matters in community ecology’,
655 *Trends in Ecology and Evolution*. doi: 10.1016/j.tree.2011.01.009.
- 656 Breed, M. F. *et al.* (2013) ‘Which provenance and where? Seed sourcing strategies for
657 revegetation in a changing environment’, *Conservation Genetics*, 14(1), pp. 1–10. doi:
658 10.1007/s10592-012-0425-z.
- 659 Broadhurst, L. M. *et al.* (2008) ‘Seed supply for broadscale restoration: Maximizing evolutionary
660 potential’, *Evolutionary Applications*, 1(4), pp. 587–597. doi: 10.1111/j.1752-
661 4571.2008.00045.x.
- 662 Bucharova, A. *et al.* (2016) ‘Plant ecotype affects interacting organisms across multiple trophic
663 levels’, *Basic and Applied Ecology*. doi: 10.1016/j.baae.2016.09.001.
- 664 Bucharova, A., Durka, W., *et al.* (2017) ‘Are local plants the best for ecosystem restoration? It
665 depends on how you analyze the data’, *Ecology and Evolution*, 7(24), pp. 10683–10689. doi:
666 10.1002/ece3.3585.
- 667 Bucharova, A., Michalski, S., *et al.* (2017) ‘Genetic differentiation and regional adaptation
668 among seed origins used for grassland restoration: lessons from a multispecies transplant
669 experiment’, *Journal of Applied Ecology*, 54(1), pp. 127–136. doi: 10.1111/1365-2664.12645.
- 670 Bucharova, A. *et al.* (2018) ‘Mix and match: regional admixture provenancing strikes a balance
671 among different seed-sourcing strategies for ecological restoration’, *Conservation Genetics*. doi:
672 10.1007/s10592-018-1067-6.

- 673 Chivers, I. H. *et al.* (2016) ‘The merits of artificial selection for the development of restoration-
674 ready plant materials of native perennial grasses’, *Restoration Ecology*, 24(2), pp. 174–183. doi:
675 10.1111/rec.12323.
- 676 Clausen, J., Keck, D. and Hiesey, W. (1948) ‘Experimental studies on the nature of species. III.
677 Environmental responses of climatic races of *Achillea*’, *Carnegie Institution of Washington*, 581,
678 pp. 1–129.
- 679 Combs, J. G. *et al.* (2011) ‘Assessing Cumulative Evidence within “Macro” Research: Why
680 Meta-Analysis Should be Preferred Over Vote Counting’, *Journal of Management Studies*. doi:
681 10.1111/j.1467-6486.2009.00899.x.
- 682 Comstock, J. P. and Ehleringer, J. R. (1992) ‘Correlating genetic variation in carbon isotopic
683 composition with complex climatic gradients.’, *Proceedings of the National Academy of*
684 *Sciences*, 89(16), pp. 7747–7751. doi: 10.1073/pnas.89.16.7747.
- 685 Davies, K. W. *et al.* (2011) ‘Saving the sagebrush sea: An ecosystem conservation plan for big
686 sagebrush plant communities’, *Biological Conservation*, 144(11), pp. 2573–2584. doi:
687 10.1016/j.biocon.2011.07.016.
- 688 Dennison, P. E. *et al.* (2014) ‘Large wildfire trends in the western United States, 1984–2011’,
689 *Geophysical Research Letters*, 41(8), pp. 2928–2933. doi: 10.1002/2014GL059576.
- 690 Donohue, K. *et al.* (2010) ‘Germination, postgermination adaptation, and species ecological
691 ranges’, *Annual Review of Ecology, Evolution, and Systematics*, 41, pp. 293–319.
- 692 Endler, J. A. (1986) *Natural selection in the wild*, *Monographs in Population Biology*. doi:
693 10.2307/302397.
- 694 Erickson, T. E. *et al.* (2017) ‘Benefits of adopting seed-based technologies for rehabilitation in
695 the mining sector: a Pilbara perspective’, *Australian Journal of Botany*. doi: 10.1071/BT17154.
- 696 Erickson, V. J., Mandel, N. L. and Sorenson, F. C. (2004) ‘Landscape patterns of phenotypic
697 variation and population structuring in a selfing grass, *Elymus glaucus* (blue wildrye)’, *Canadian*
698 *Journal of Botany*, 82(12), pp. 1776–1789. doi: 10.1139/B04-141.

- 699 Espeland, E. K. *et al.* (2016) ‘Perennial grass cultivars grown at different production farms
700 respond differently to storage and planting environments’, *Crop Science*, 56, pp. 249–258. doi:
701 10.2135/cropsci2015.05.0318.
- 702 Espeland, E. K. *et al.* (2017) ‘Evolution of plant materials for ecological restoration: Insights
703 from the applied and basic literature’, *Journal of Applied Ecology*, 54, pp. 102–115. doi:
704 10.1111/1365-2664.12739.
- 705 Espeland, E. K., Johnson, R. C. and Horning, M. E. (2017) ‘Plasticity in native perennial grass
706 populations: Implications for restoration’, *Evolutionary Applications*, 00, pp. 1–10. doi:
707 10.1111/eva.12560.
- 708 Evans, R. A. and Young, J. A. (1990) ‘Survival and Growth of Big Sagebrush (*Artemisia*
709 *tridentata*) Plants in Reciprocal Gardens’, *Weed Science*, 38(3), pp. 215–219. doi:
710 10.2307/4045014.
- 711 Finch, D. M. *et al.* (2016) ‘Conservation and Restoration of Sagebrush Ecosystems and Sage-
712 Grouse: An Assessment of USDA Forest Service Science’, *General technical report*, RMRS-
713 GTR-3(US Department of Agriculture, Forest Service, Rocky Mountain Research Station).
714 Available at: https://www.fs.fed.us/rm/pubs/rmrs_gtr348.pdf.
- 715 Franks, S. J., Weber, J. J. and Aitken, S. N. (2014) ‘Evolutionary and plastic responses to climate
716 change in terrestrial plant populations’, *Evolutionary Applications*, 7(1), pp. 123–139. doi:
717 10.1111/eva.12112.
- 718 Friggens, M. M. *et al.* (2012) ‘Decision support: Vulnerability, conservation, and restoration
719 (Chapter 8)’, *USDA Forest Service, Rocky Mountain Research Station - General Technical*
720 *Report*, (RMRS-GTR-285), pp. 116–139. Available at:
721 [http://www.scopus.com/inward/record.url?eid=2-s2.0-](http://www.scopus.com/inward/record.url?eid=2-s2.0-84865151549&partnerID=40&md5=7e0d86d3567ac2ed6c834c0d831cbdca)
722 [84865151549&partnerID=40&md5=7e0d86d3567ac2ed6c834c0d831cbdca](http://www.scopus.com/inward/record.url?eid=2-s2.0-84865151549&partnerID=40&md5=7e0d86d3567ac2ed6c834c0d831cbdca).
- 723 Geber, M. A. and Griffen, L. R. (2003) ‘Inheritance and Natural Selection on Functional Traits’,
724 *International Journal of Plant Sciences*, 164(S3), pp. S21–S42. doi: 10.1086/368233.
- 725 Gibson, A. L. *et al.* (2016) ‘Can local adaptation research in plants inform selection of native

- 726 plant materials? An analysis of experimental methodologies’, *Evolutionary Applications*, 9(10),
727 pp. 1219–1228. doi: 10.1111/eva.12379.
- 728 Griffith, D. A. and Peres-Neto, P. R. (2006) ‘Spatial modeling in ecology: The flexibility of
729 eigenfunction spatial analyses’, *Ecology*. doi: 10.1890/0012-
730 9658(2006)87[2603:SMIETF]2.0.CO;2.
- 731 Harder, L. D. and Barrett, S. C. H. (2007) *Ecology and evolution of flowers*. New York: Oxford
732 University Press.
- 733 Harvey, P. H. and Pagel, M. D. (1991) ‘The comparative method in evolutionary biology’,
734 *Oxford Series in Ecology and Evolution*, p. 239. doi: 10.1016/0169-5347(92)90117-T.
- 735 Havens, K. *et al.* (2015) ‘Seed Sourcing for Restoration in an Era of Climate Change’, *Natural*
736 *Areas Journal*, 35(1), pp. 122–133. doi: 10.3375/043.035.0116.
- 737 Hereford, J. (2009) ‘A Quantitative Survey of Local Adaptation and Fitness Trade- Offs’, *The*
738 *American Naturalist*, 173(5), pp. 579–588. doi: 10.1086/597611.
- 739 Hoffmann, A. A. and Sgrò, C. M. (2011) ‘Climate change and evolutionary adaptation.’, *Nature*,
740 470(7335), pp. 479–485. doi: 10.1038/nature09670.
- 741 Johnson, G. *et al.* (2004) ‘Pacific Northwest Forest Tree Seed Zones: A Template for Native
742 Plants?’, *Native Plants Journal*, 5, pp. 131–140. doi: 10.2979/NPJ.2004.5.2.131.
- 743 Johnson, R. *et al.* (2010) ‘What are the best seed sources for ecosystem restoration on BLM and
744 USFS lands?’, *Native Plants Journal*, 11(2), pp. 117–131. doi: 10.2979/NPJ.2010.11.2.117.
- 745 Johnson, R. C., Leger, E. A. and Vance-Borland, K. (2017) ‘Genecology of Thurber’s
746 Needlegrass (*Achnatherum thurberianum* [Piper] Barkworth) in the Western United States’,
747 *Rangeland Ecology & Management*. the Society for Range Management, 70(4), pp. 509–517.
748 doi: 10.1016/j.rama.2017.01.004.
- 749 Jones, T. A. and Larson, S. R. (2005) ‘Status and use of important native grasses adapted to
750 sagebrush communities’, in *Shaw, Nancy L.; Pellant, Mike; Monsen, Stephen B., comps. Sage-*
751 *grouse habitat restoration symposium proceedings; 2001 June 4-7, Boise, ID. Proc. RMRS-P-38.*

- 752 Fort Collins, CO: U.S. Department of Agriculture, Forest Service, Rocky Mountain Research
753 Station., pp. 49–55.
- 754 Jones, T. A. and Monaco, T. A. (2009) ‘A role for assisted evolution in designing native plant
755 materials for domesticated landscapes’, *Frontiers in Ecology and the Environment*, 7(10), pp.
756 541–547. doi: 10.1890/080028.
- 757 Jones, T. A., Monaco, T. A. and Rigby, C. W. (2015) ‘The potential of novel native plant
758 materials for the restoration of novel ecosystems’, *Elementa: Science of the Anthropocene*, 3.
759 doi: 10.12952/journal.elementa.000047.
- 760 Kawecki, T. and Ebert, D. (2004) ‘Conceptual issues in local adaptation’, *Ecology Letters*, 7(12),
761 pp. 1225–1241.
- 762 Kilkenny, F. F. (2015) ‘Genecological Approaches to Predicting the Effects of Climate Change
763 on Plant Populations’, *Natural Areas Journal*, 35(1), pp. 152–164. doi: 10.3375/043.035.0110.
- 764 Kramer, A. T., Larkin, D. J. and Fant, J. B. (2015a) ‘Assessing potential seed transfer zones for
765 five forb species from the Great Basin Floristic Region, USA’, *Natural Areas Journal*, 35(1).
766 doi: 10.3375/043.035.0119.
- 767 Kramer, A. T., Larkin, D. J. and Fant, J. B. (2015b) ‘Assessing Potential Seed Transfer Zones for
768 Five Forb Species from the Great Basin Floristic Region, USA’, *Natural Areas Journal*, 35(1),
769 pp. 174–188. doi: 10.3375/043.035.0119.
- 770 Kulpa, S. M. and Leger, E. A. (2013) ‘Strong natural selection during plant restoration favors an
771 unexpected suite of plant traits’, *Evolutionary Applications*, 6(3), pp. 510–523. doi:
772 10.1111/eva.12038.
- 773 Laliberté, E. (2011) ‘metacor: Meta-analysis of correlation coefficients’, *R package 1.0-2*.
774 Available at: <https://cran.r-project.org/package=metacor>.
- 775 Langlet, O. (1971) ‘Two Hundred Years Genecology’, *Taxon*. International Association for Plant
776 Taxonomy (IAPT), 20(5/6), pp. 653–721. doi: 10.2307/1218596.
- 777 Larsen, E. C. (1947) ‘Photoperiodic Responses of Geographical Strains of *Andropogon*

- 778 scoparius', *Botanical Gazette*, 109(2), pp. 132–149. doi: 10.1086/335463.
- 779 Leger, E. A. and Baughman, O. W. (2015) 'What seeds to plant in the Great Basin? Comparing
780 traits prioritized in native plant cultivars and releases with those that promote survival in the
781 field', *Natural Areas Journal*, 35(1), pp. 54–68. doi: 10.3375/043.035.0108.
- 782 Leimu, R. and Fischer, M. (2008) 'A meta-analysis of local adaptation in plants', *PLoS ONE*,
783 3(12), p. e4010. doi: 10.1371/journal.pone.0004010.
- 784 Lesica, P. and Allendorf, F. W. (1999) 'Ecological Genetics and the Restoration of Plant
785 Communities: Mix or Match?', *Restoration Ecology*, 7(1), pp. 42–50. doi: 10.1046/j.1526-
786 100X.1999.07105.x.
- 787 Linhart, Y. B. and Grant, M. C. (1996) 'Evolutionary Significance of Local Genetic
788 Differentiation in Plants', *Annual Review of Ecology and Systematics*, 27(1), pp. 237–277. doi:
789 10.1146/annurev.ecolsys.27.1.237.
- 790 Loveless, M. D. and Hamrick, J. L. (1984) 'Ecological Determinants of Genetic Structure in
791 Plant Populations', *Annual Review of Ecology and Systematics*, 15(1), pp. 65–95. doi:
792 10.1146/annurev.es.15.110184.000433.
- 793 Matyas, C. (1996) 'Climatic adaptation of trees: rediscovering provenance tests', *Euphytica*, Jan
794 1(92 (1-2)), pp. 45–54.
- 795 McArthur, E. D., Meyer, S. E. and Weber, D. J. (1987) 'Germination rate at low temperature:
796 Rubber rabbitbrush population differences', *Journal of Range Management*, 40(6), pp. 530–533.
797 doi: 10.2307/3898874.
- 798 McArthur, E. D. and Young, S. A. (1999) 'Development of native seed supplies to support
799 restoration of pinyon-juniper sites', in *S.B. Monsen and R. Stevens, compilers, Proceedings:
800 ecology and management of pinyon-juniper communities within the Interior West. Proc. RMRS-
801 P-9*. USDA Forest Service, Rocky Mountain Research Station, Ogden, UT, pp. 327–330.
- 802 Meyer, S. E. *et al.* (1995) 'Germination ecophysiology of *Leymus cinereus* (Poaceae)',
803 *International Journal of Plant Sciences*, 156(2), pp. 206–215. doi: 10.1086/297242.

- 804 Meyer, S. E. (1997) ‘Genecological Considerations in Grassland Restoration Using Wild-
805 Collected Seed Sources’, in *Proceesings of the 18th International Grassland Congress*,
806 *Winnipeg and Saskatoon, Canada*, pp. 8–17. Available at:
807 <http://www.internationalgrasslands.org/files/igc/publications/1997/iii-299.pdf> (Accessed: 8
808 January 2018).
- 809 Meyer, S. E. and Monsen, S. B. (1991) ‘Habitat-correlated variation in mountain big sagebrush
810 (*Artemisia tridentata* ssp. *vaseyana*) seed germination patterns’, *Ecology*, 72(2), pp. 739–742.
811 doi: 10.2307/2937214.
- 812 Vander Mijnsbrugge, K., Bischoff, A. and Smith, B. (2010) ‘A question of origin: Where and
813 how to collect seed for ecological restoration’, *Basic and Applied Ecology*, 11(4), pp. 300–311.
814 doi: 10.1016/J.BAAE.2009.09.002.
- 815 Oduor, A. M. O. (2013) ‘Evolutionary responses of native plant species to invasive plants: a
816 review’, *New Phytologist*, 200(4), pp. 986–992. doi: Doi 10.1111/Nph.12429.
- 817 Oduor, A. M. O., Leimu, R. and van Kleunen, M. (2016) ‘Invasive plant species are locally
818 adapted just as frequently and at least as strongly as native plant species’, *Journal of Ecology*.
819 doi: 10.1111/1365-2745.12578.
- 820 Oksanen, J. *et al.* (2018) ‘Vegan: community ecology package’, *R package 2.4-6*. doi:
821 10.4135/9781412971874.n145.
- 822 Pilliod, D. S., Welty, J. L. and Toevs, G. R. (2017) ‘Seventy-Five Years of Vegetation
823 Treatments on Public Rangelands in the Great Basin of North America’, *Rangelands*, 39(1), pp.
824 1–9. doi: 10.1016/j.rala.2016.12.001.
- 825 Plant Conservation Alliance, P. (2015) ‘National Seed Strategy for Rehabilitation and
826 Restoration 2015-2020’. Bureau of Land Management. Available at:
827 [https://www.blm.gov/programs/natural-resources/native-plant-communities/national-seed-](https://www.blm.gov/programs/natural-resources/native-plant-communities/national-seed-strategy)
828 [strategy](https://www.blm.gov/programs/natural-resources/native-plant-communities/national-seed-strategy).
- 829 Prober, S. M. *et al.* (2015) ‘Climate-adjusted provenancing: a strategy for climate-resilient
830 ecological restoration’, *Frontiers in Ecology and Evolution*, 3, p. 65. doi:

- 831 10.3389/fevo.2015.00065.
- 832 R Core Team (2017) ‘R: A Language and Environment for Statistical Computing’, *R Foundation*
833 *for Statistical Computing*. Vienna. Available at: <https://www.r-project.org/>.
- 834 Reich, P. B. *et al.* (2003) ‘The evolution of plant functional variation: traits, spectra, and
835 strategies’, *International Journal of Plant Sciences*, 164(S3), pp. S143–S164. doi:
836 10.1086/374368.
- 837 Rice, K. J. and Emery, N. C. (2003) ‘Managing microevolution: Restoration in the face of global
838 change’, *Frontiers in Ecology and the Environment*, 1(9), pp. 469–478. doi: 10.1890/1540-
839 9295(2003)001[0469:MMRITF]2.0.CO;2.
- 840 Richards, R. T., Chambers, J. C. and Ross, C. (1998) ‘Use of native plants on federal lands:
841 policy and practice’, *Journal of Range Management*, 51(6), pp. 625–632. doi: 10.2307/4003603.
- 842 Des Roches, S. *et al.* (2018) ‘The ecological importance of intraspecific variation’, *Nature*
843 *Ecology & Evolution*, 2(1), pp. 57–64. doi: 10.1038/s41559-017-0402-5.
- 844 Rogers, D. L. and Montalvo, A. M. (2004) ‘Genetically appropriate choices for plant materials to
845 maintain biological diversity’, in *University of California. Report to the USDA Forest Service,*
846 *Rocky Mountain Region, Lakewood, CO*. doi: Genetic Considerations in Ecological Restoration.
- 847 Rowland, M. M., Suring, L. H. and Michael, J. (2010) ‘Assessment of habitat threats to
848 shrublands in the Great Basin: a case study’, *Advances in threat assessment and their application*
849 *to forest and rangeland management. General Technical Report. PNW-GTR-802*, pp. 673–685.
850 Available at: <http://www.northern-ecologic.com/publications/54.pdf>.
- 851 Sheridan, J. A. and Bickford, D. (2011) ‘Shrinking body size as an ecological response to climate
852 change’, *Nature Climate Change*, 1(8), pp. 401–406. doi: 10.1038/nclimate1259.
- 853 Siepielski, A. M., Dibattista, J. D. and Carlson, S. M. (2009) ‘It’s about time: The temporal
854 dynamics of phenotypic selection in the wild’, *Ecology Letters*, pp. 1261–1276. doi:
855 10.1111/j.1461-0248.2009.01381.x.
- 856 St Clair, J. B., Mandel, N. L. and Vance-Borland, K. W. (2005) ‘Genecology of Douglas fir in

- 857 Western Oregon and Washington’, *Annals of Botany*, 96(7), pp. 1199–1214. doi:
858 10.1093/aob/mci278.
- 859 Sultan, S. E. and Spencer, H. G. (2002) ‘Metapopulation structure favors plasticity over local
860 adaptation’, *The American Naturalist*, 160(2), pp. 271–283. doi: 10.1086/341015.
- 861 Svejcar, T. *et al.* (2017) ‘Challenges and limitations to native species restoration in the Great
862 Basin, USA’, *Plant Ecology*, 218(1), pp. 81–94. doi: 10.1007/s11258-016-0648-z.
- 863 Tisdale, E. W. and Hironaka, M. (1981) ‘The Sagebrush-Grass Region: A Review of the
864 Ecological Literature’, *University of Idaho Forest, Wildlife, and Range Experiment Station*.
865 Available at: <http://digital.lib.uidaho.edu/cdm/ref/collection/fwres/id/169> (Accessed: 5 January
866 2018).
- 867 Turesson, G. (1922) ‘The genotypical response of the plant species to the habitat’, *Hereditas*,
868 3(3), pp. 211–350. doi: 10.1111/j.1601-5223.1922.tb02734.x.
- 869 United States. House of Representatives. Committee on Appropriations. (2014) ‘American Seed
870 Trade Association Statement by Mark Mustoe, Co-Owner and Manager of Clearwater Seed
871 Regarding Efficient Native Seed Use by the Bureau of Land Management’. Alexandria, VA.
872 Available at: [http://docs.house.gov/meetings/AP/AP06/20140410/101762/HHRG-113-AP06-
873 Wstate-MustoeM-20140410.pdf](http://docs.house.gov/meetings/AP/AP06/20140410/101762/HHRG-113-AP06-Wstate-MustoeM-20140410.pdf) (Accessed: 4 January 2018).
- 874 USDA and NRCS (2018) *The PLANTS Database, National Plant Data Team, Greensboro, NC*
875 *27401-4901 USA*. Available at: <http://plants.usda.gov> (Accessed: 8 January 2018).
- 876 USDOJ (2015) *SO-3336: An Integrated Rangeland Fire Management Strategy. Final Report to*
877 *the Secretary of the Interior*. Available at:
878 [https://www.forestsandrangelands.gov/rangeland/documents/IntegratedRangelandFireManageme
879 ntStrategy_FinalReportMay2015.pdf](https://www.forestsandrangelands.gov/rangeland/documents/IntegratedRangelandFireManagementStrategy_FinalReportMay2015.pdf). [Accessed January 26, 2018].
- 880 Vavrek, M. J. (2011) ‘Fossil: palaeoecological and palaeogeographical analysis tools’,
881 *Palaeontologia Electronica*, 14(1), p. 16.
- 882 Wang, T. *et al.* (2016) ‘Locally downscaled and spatially customizable climate data for historical
883 and future periods for North America’, *PLoS ONE*, 11(6), p. e0156720. doi:

884 10.1371/journal.pone.0156720.

885 Ward, R. T. (1969) 'Ecotypic variation in *Deschampsia caespitosa* (L.) Beauv. from Colorado',
886 *Ecology*, 50(3), pp. 519–522. doi: 10.2307/1933914.

887 Williams, A. V., Nevill, P. G. and Krauss, S. L. (2014) 'Next generation restoration genetics:
888 Applications and opportunities', *Trends in Plant Science*, 19(8), pp. 529–537. doi:
889 10.1016/j.tplants.2014.03.011.

890 Wright, I. J. *et al.* (2017) 'Global climatic drivers of leaf size', *Science*, 357(6354), pp. 917–921.
891 doi: 10.1126/science.aa14760.

892

893

894 Table 1. Traits measured in outdoor common gardens or reciprocal transplants for at least 5
895 populations in at least 20 common garden locations, with data available from text, tables, author
896 contact, or extraction from figures. Note that in some cases, multiple highly similar measures
897 were grouped, as indicated in footnotes.

898

Trait	Units	Locations
date – flowering ¹	# days	34
size – floral ²	cm	22
height - plant	cm	46
size – leaf ³	cm	30
mass – shoots ⁴	g	43
number - inflorescence ⁵	#	36
number – seeds ⁶	#	21
survival	%	43

¹Flowering date or any other floral phenology

²Any size measurement of a floral structure

³Most frequently, leaf length; occasionally leaf width

⁴ Any measure of aboveground biomass

⁵ Counts of flowers or flowering structures

⁶ Most frequently seed number, but also seed yield in mass and/or seed yield rating/rank

899

900

901 **Figure captions**

902 Figure 1. Summary of reviewed literature that compared traits among at least two populations in
903 at least one common environment, by lifeform. Total counts of published studies, species, taxa,
904 and taxa-specific experiments (A); types of experiments (B); means and standard errors of
905 duration of the experiments that measured more than germination traits (C); total counts of
906 experiments that measured only germination traits, (D); means and standard errors of number of
907 populations tested in each experiment (E), and garden sites per experiment for outdoor reciprocal
908 transplant and common garden experiments (F).

909

910 Figure 2. Map of 129 different outdoor common garden locations (A) and 2953 unique
911 population collection sites (B) for the 80% of outdoor gardens and 91% of experiments for which
912 coordinates could be obtained or generated, from 170 studies reviewed. The size of the marker in
913 panel A represents the number of experiments in which each specific garden location was used,
914 with larger symbols indicating garden locations used in more experiments. Although all species
915 represented are native to the floristic Great Basin (white outline), many populations were
916 collected and tested outside this region.

917

918 Figure 3. Summary of the years in which the collections of each experiment were made (A, left),
919 the year each experiment was performed (A, right), and the average geographic distance among
920 population collections sites in each experiment. The percent of 327 experiments that reported this
921 information were 99% and 88% (respectively) for panel A, and 80% for panel B. Collection year
922 and experiment year represent the average for each experiment, as it was common for materials
923 to be collected and tested over multiple years for each experiment. Geographic distance is the

924 mean pairwise distance among populations in each experiment; note the noncontinuous vertical
925 axis.

926

927 Figure 4. Summary of among-population variation (A, signature 1) and trait-by-environment
928 associations (B, signature 2) for any measured trait, grouped by five life history traits. Summary
929 of local advantage (C, signature 3) for reproductive traits, survival traits, fitness indices, or
930 biomass. Data compiled from 327 experiments from 170 published studies on Great Basin plants
931 (see Supporting Information Appendix 2 and available datasets in electronic supplementary
932 material). For signatures 1 and 2, “Yes” and “No” represent statistical comparisons, while
933 “Authors claim “Yes”” and “Authors claim “No”” represent textual, claim-based results where
934 supporting statistics were not reported (common in older studies). For signature 3, most
935 experiments had multiple gardens, and many evaluated performance at multiple sampling dates,
936 leading to 5 different scores. These scores, from “All gardens, all times” to “No gardens at any
937 time” represent a gradient of incidence and frequency of this signature (see methods). For all
938 panels, numbers in parentheses, (x), indicate the number of experiments scored in a given
939 category, and the dashed gray lines indicate 50%.

940

941 Figure 5. Summary of 1,465 trait scores from the 305 experiments appropriate for detecting
942 signature 1 (differences between populations). Scores of “Yes” and “No” were supported by
943 statistical comparisons, while the “Authors claim...” scores represent textual, claim-based results
944 where supporting statistics were not reported (common in older studies). Numbers in
945 parentheses, (x), indicate the total experiments that measured each trait or reported each factor,
946 and dashed gray line indicates 50%.

947

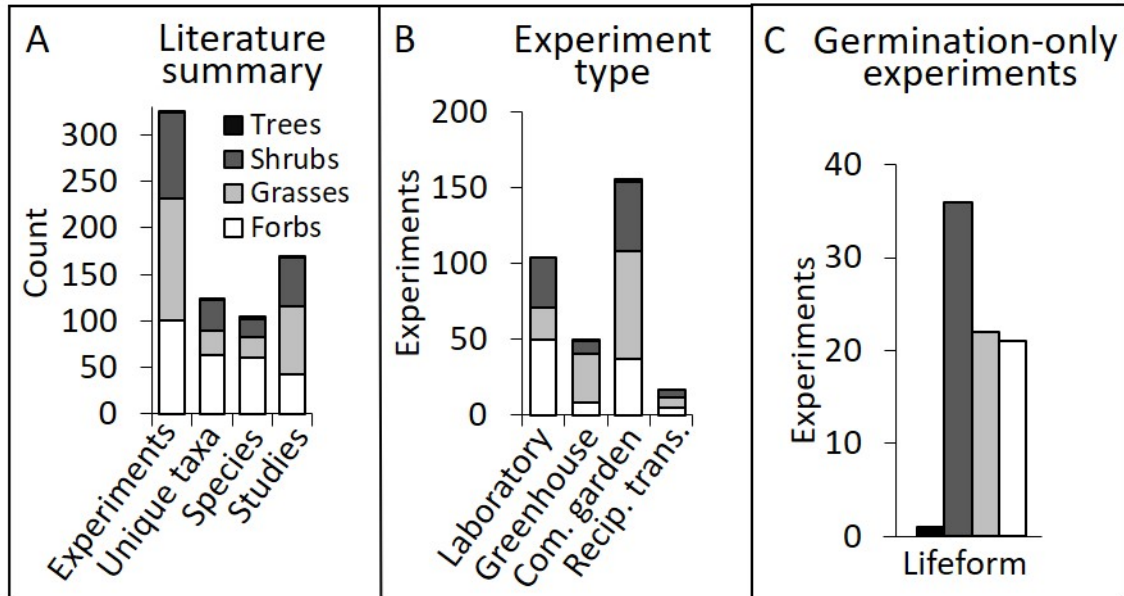
948 Figure 6. Summary of scores for associations between 592 traits (A) and 426 environmental
949 factors (B) from the 161 experiments appropriate for detecting signature 2 (trait-by-environment
950 association), expressed by trait/factors, and an example from the literature (C, redrawn with
951 permission from (Meyer and Monsen, 1991)) in which date of germination for mountain big
952 sagebrush is correlated with a measure of monthly temperature (treatment: 2-week chill). Scores
953 of “Yes” and “No” were supported by statistical comparisons, while the “Authors claim...”
954 scores represent textual, claim-based results where supporting statistics were not reported
955 (common in older studies). For panels A and B, numbers in parentheses, (x), indicate the total
956 experiments that measured each trait or reported each factor, and the dashed gray lines indicate
957 50%.

958

959 Figure 7. Results of comparisons of pairwise trait and environmental distances for eight
960 frequently measured phenotypic traits and (A) the mean annual precipitation (MAP) or (B) mean
961 annual temperature (MAT) at the original collection location. Values are effect sizes and 95%
962 confidence intervals for each trait, averaged across all experiments for which data were available
963 (number of experiments in parentheses). Examples of the two strongest relationships are shown
964 for leaf size and MAP (C), where each line shows the correlation coefficient and confidence
965 intervals for an individual experiment, for which we calculated the relationship between
966 differences in percent survival and difference MAP at location of origin. Color indicates
967 functional groups: Green = grasses, blue = shrubs, orange = forbs. Examples are shown for the
968 two highest effect sizes: D), experiment 297A, (Kramer, Larkin and Fant, 2015), *Penstemon*
969 *deustus* and E), experiment 297A, (Kramer, Larkin and Fant, 2015), *Eriogonum microthecum*.

970 Similarly, flowering time and MAT (F) is shown, with examples of G) experiment 271A,
971 (Larsen, 1947), *Schizachyrium scoparium*, and H) experiment 245A, (Ward, 1969), *Deschampsia*
972 *caespitosa*. Full results for each trait/environment relationship are shown as additional results in
973 Supporting Information Appendix 3.
974
975

976 Figure 1.

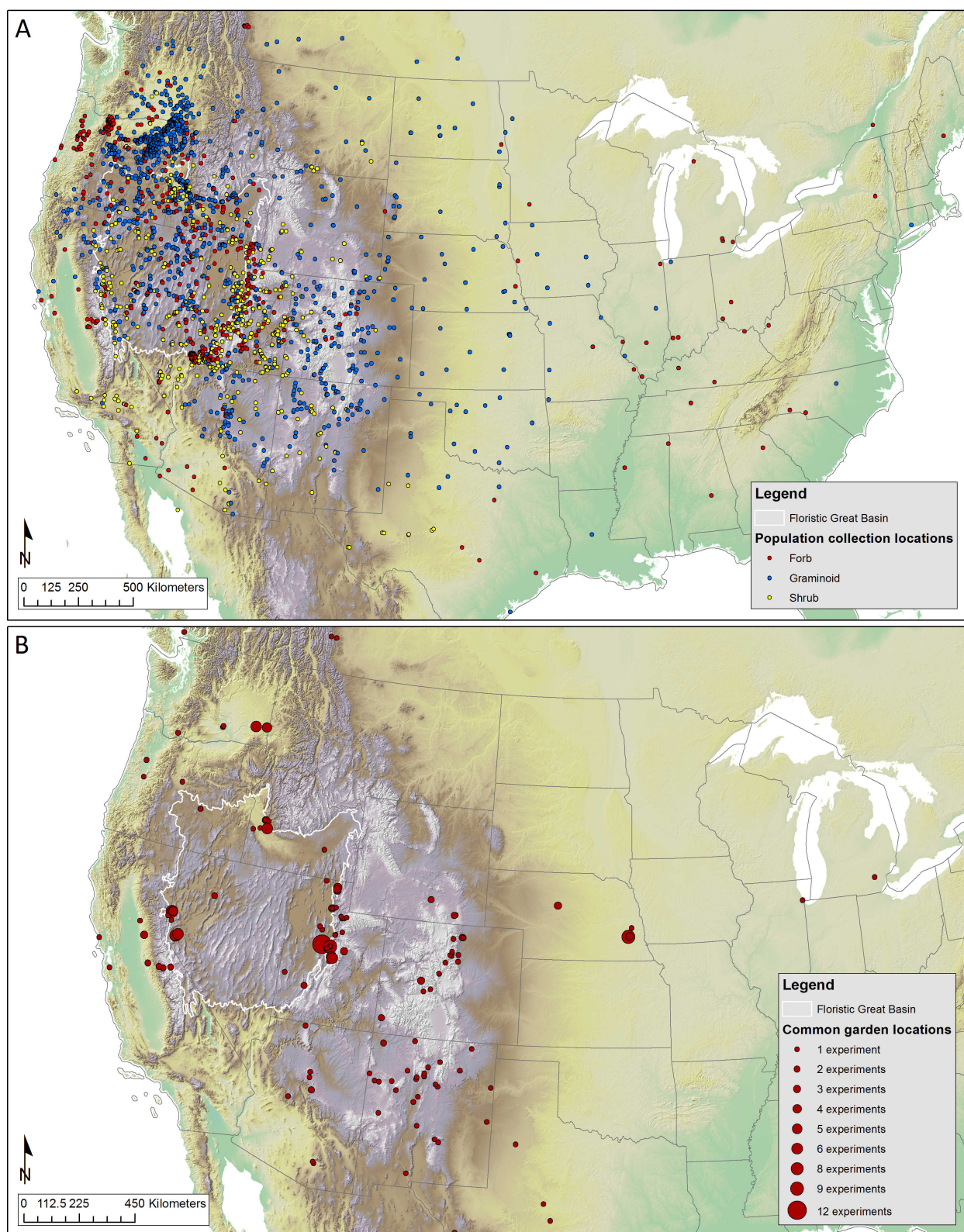


977

978

979

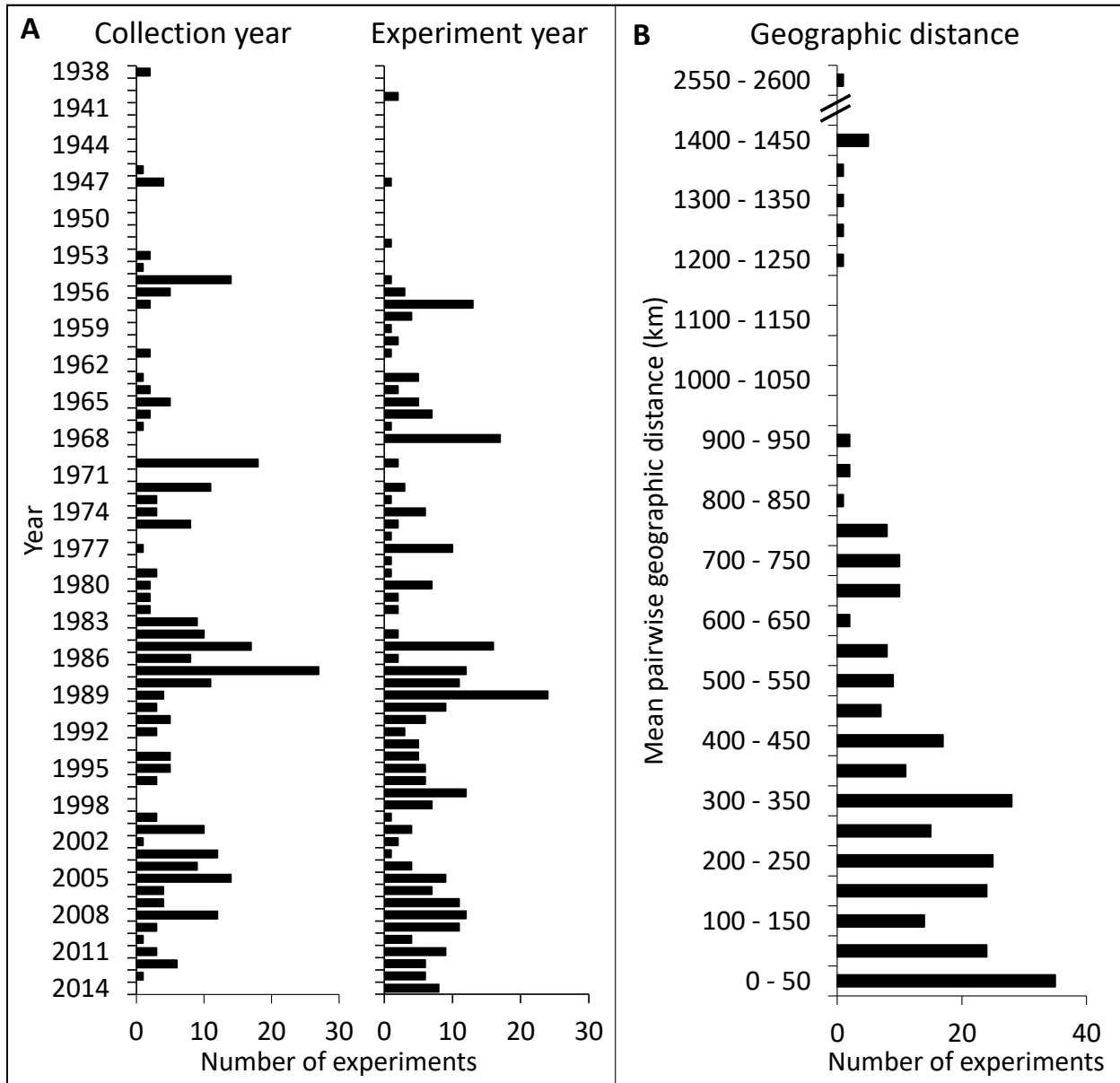
980 Figure 2.



981

982

983 Figure 3.

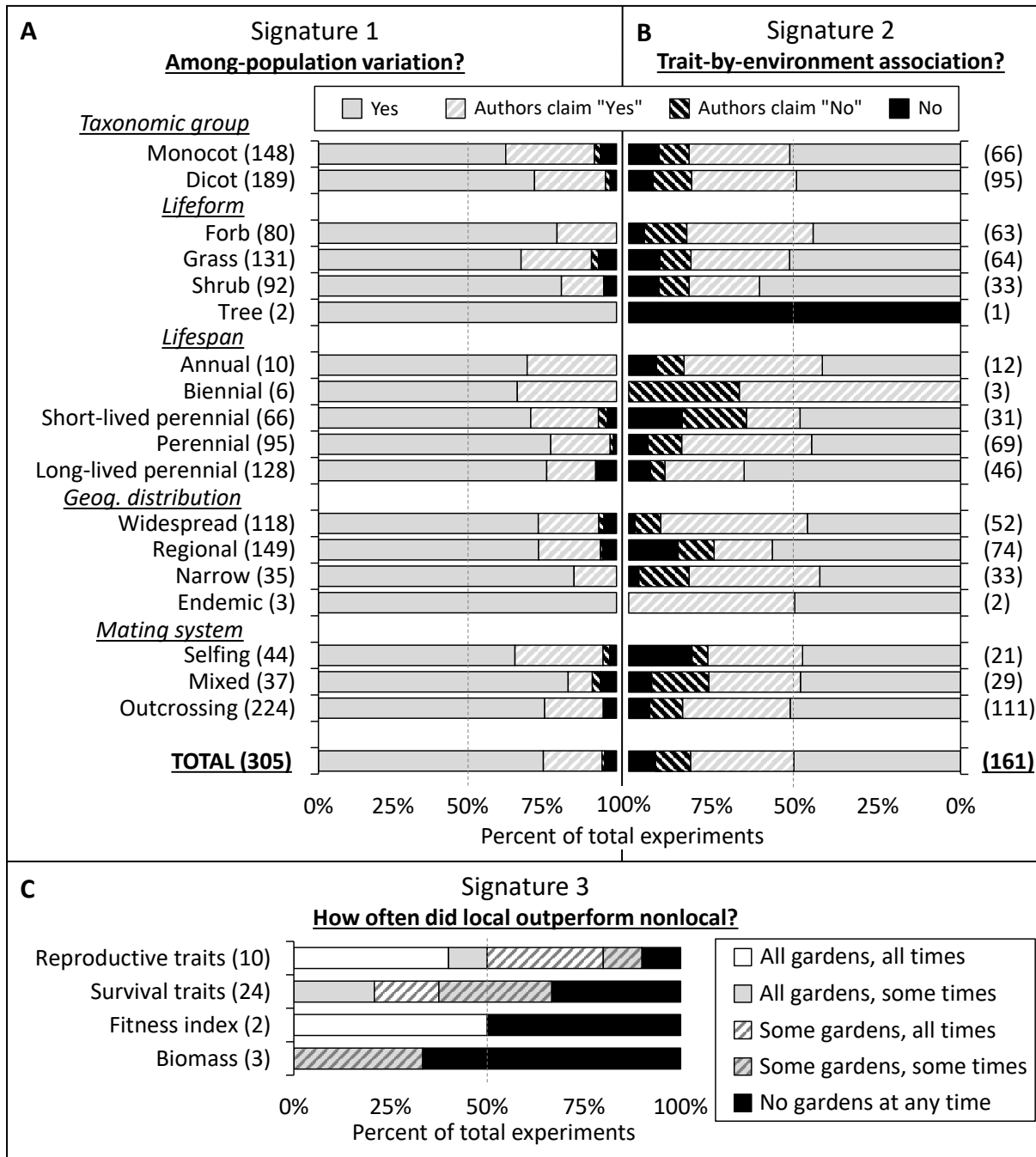


984

985

986 Figure 4.

987

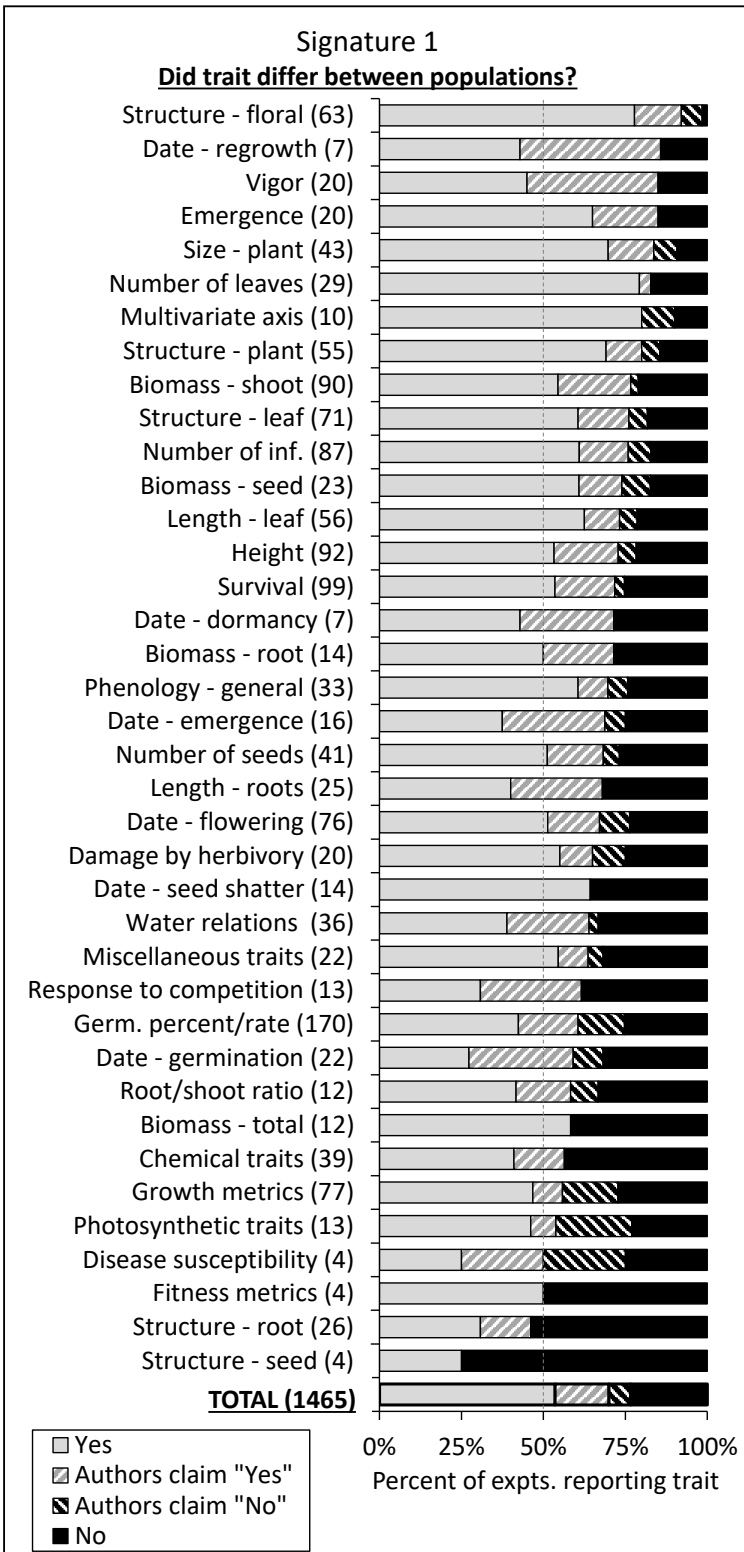


988

989

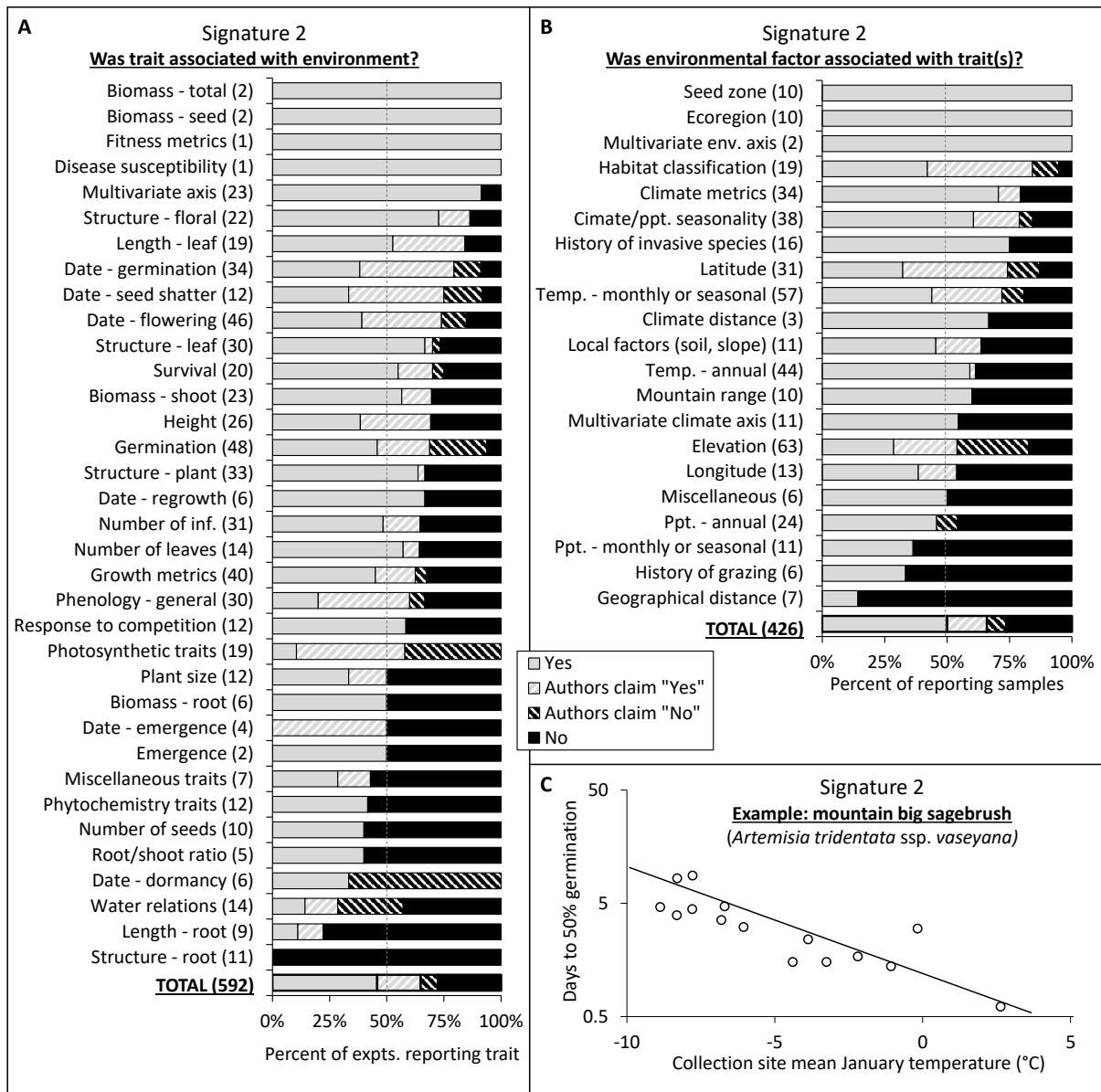
990

991 Figure 5.



992

993 Figure 6.



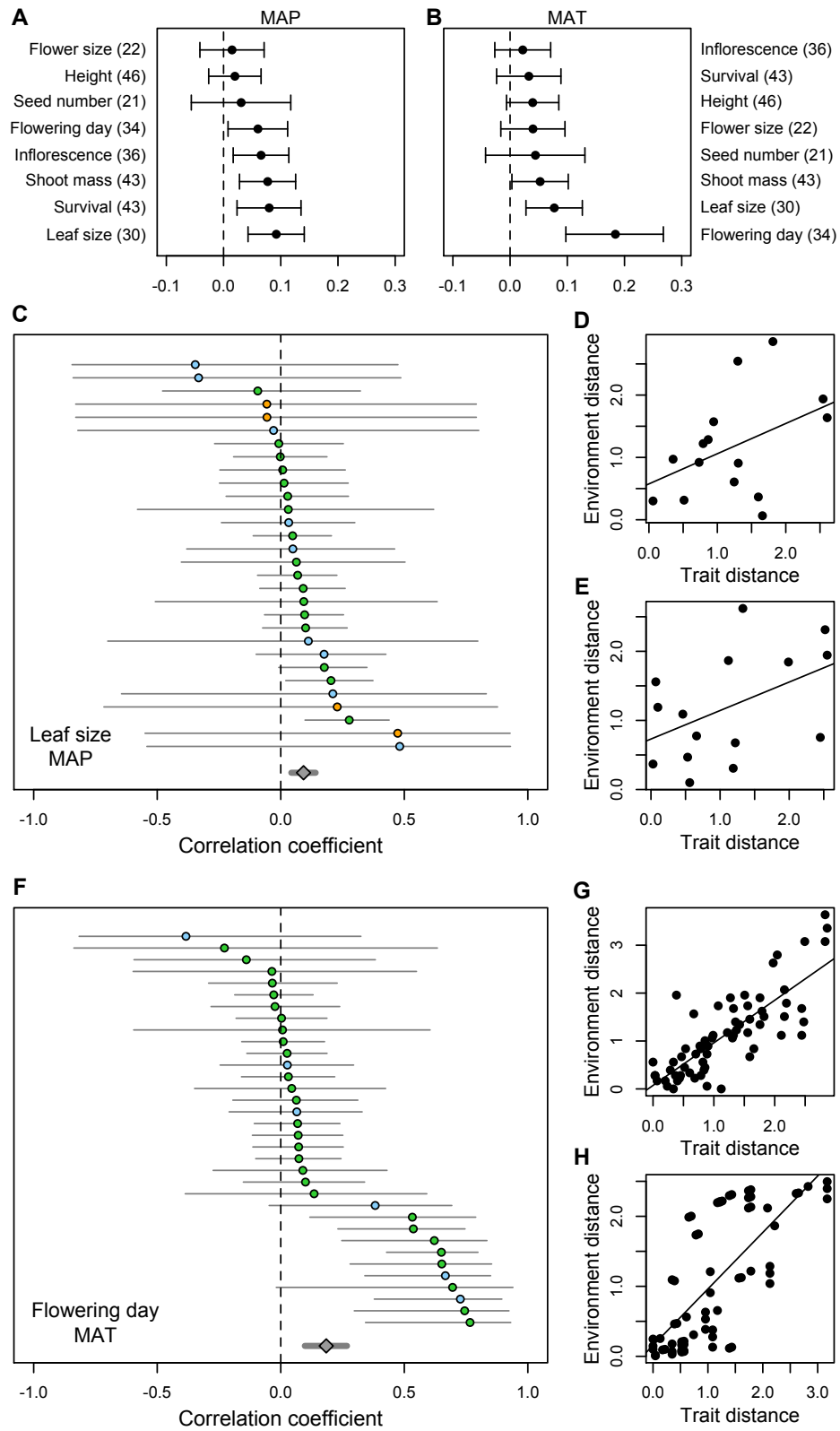
994

995

996

997

998 Figure 7.



1000 **Appendix 1. Additional Methods**

1001

1002 *Literature search*

1003 Terms used to search the literature included ‘plant’, ‘Great Basin’, ‘Intermountain West’,
1004 ‘western United States’, ‘local adaptation’, ‘ecotypic variation’, ‘phenotypic variation’, ‘genetic
1005 variation’, ‘habitat-correlated variation’, ‘genecology’, ‘intraspecific variation’, ‘ecotype’, ‘seed zones’,
1006 ‘common garden’, ‘reciprocal garden’, and ‘transplant garden’, as well as combinations of these terms.
1007 Literature was obtained primarily using the World Wide Web as well as databases such as Google
1008 Scholar, Web of Science, Academic Search Premier, JSTOR, Science Direct, and Wiley Online Library.
1009 When digital copies were not available, they were obtained from academic libraries. The citations within
1010 the resulting literature were also mined for additional literature that our first search had missed.

1011

1012 *Geographic range categorization*

1013 Four categories of geographic range were assigned from distributions in the USDA Plants
1014 Database (<https://plants.sc.egov.usda.gov>), as follows. Widespread: found in majority of United States
1015 (e.g. *Elymus elymoides* (Raf.) Swezey); Regional: common in the floristic Great Basin but not found
1016 throughout the United States (e.g. *Atriplex confertifolia* (Torr. & Frém.) S. Watson); Narrow: limited to
1017 specific, well-defined habitats within the Great Basin (e.g. *Penstemon confusus* M.E. Jones); Endemic:
1018 restricted to several counties (e.g. *Allium passeyi* N.H. Holmgren & A.H. Holmgren).

1019

1020 *Geographic coordinate generation*

1021 Geographic coordinates and elevations for gardens and populations were recorded verbatim from
1022 studies that contained precise coordinates, or were generated manually using Google Earth Pro (Google
1023 Inc., 2018) with assistance from the Geographic Names Information System (US Geological Survey,
1024 2018) when vague coordinates or textual localities were given. All coordinates were converted to decimal
1025 degrees (WGS 84) and elevations were recorded in meters. Uncertainties in manually generated

1026 coordinates were recorded in a measure of accuracy, either ‘high’ (confident to within a ~2 mile radius),
1027 ‘fair’ (confident to within a ~5 mile radius), or ‘low’ (confident to within a ~15 mile radius). Numeric
1028 coordinates given in the studies were assumed to be accurate to within one mile. If elevations were given
1029 for populations or gardens with vague localities, we utilized this information to increase the confidence of
1030 our generated location. Coordinates were not generated for localities that were exceptionally vague or
1031 studies which did not include localities. If a study utilized a named release or cultivar, the location of
1032 origin was determined by locating the original published release notice, if available. Cultivars bred using
1033 populations from multiple locations were not assigned origin coordinates.

1034

1035 *Scoring experiments for each signature of local adaptation*

1036 For among-population variation (signature 1), a score of ‘Yes’ was given when at least one
1037 measured trait was reported to differ significantly between at least two populations, and a score of ‘No’
1038 was given when differences in any phenotypic trait were not detected between any pair of populations.
1039 For trait-by-environment association (signature 2), a score of ‘Yes’ was given when authors reported a
1040 significant association between at least one trait and one measure of the environment of origin, and a
1041 score of ‘No’ was given when the author tested for but found no such relationship. In addition to a score
1042 for each experiment, each of the measured and reported traits and environmental variables were scored
1043 (hereafter, trait scores) in a manner that indicated which traits did or did not vary between populations, as
1044 well as which traits and environmental variables were or were not correlated with each other (see
1045 available datasets in electronic supplementary material). Some experiments met the criteria for both
1046 signatures while others met only one or the other. In several studies, especially older studies or studies
1047 whose analyses did not include among-population comparisons, the significance of variation and/or
1048 correlation needed for scoring signatures 1 and 2 could not be determined because the authors provided
1049 results without statistical analyses. In these cases, results were scored as ‘Authors Claim Yes’ or ‘Authors
1050 Claim No’, and the scoring was done as described above, taking authors at their word in the absence of
1051 published statistical evidence.

1052 To score whether there was higher fitness of a local population in a common garden (signature 3),
1053 only experiments in which outdoor reciprocal transplants or common gardens were performed using a
1054 local population (identified as such by the author, or clearly collected from the common garden site) in at
1055 least one garden were considered. Additionally, the experiment had to measure survival, reproductive
1056 output (number of seeds or flowers, or other reproductive output), a fitness index (a combination of
1057 several size and production traits), or total aboveground biomass. Each experiment was given a composite
1058 score to fully capture variation in the performance of the local population across gardens (spatial), as well
1059 as through different sampling dates (temporal). For the spatial component, ‘Yes for all gardens’ indicates
1060 the highest values in each garden belonged to that garden’s local population, ‘Yes for some gardens’
1061 indicates the highest value in at least one but not all of the gardens belonged to each garden’s local
1062 population, and ‘No for all gardens’ if the highest value never belonged to a garden’s local population.
1063 For the temporal component, the experiment was scored as ‘Always’ if the local population had the
1064 highest value at all sampling dates, or ‘Sometimes’ if the local population had the highest value at one but
1065 not all of the sampling dates. For “some” and “sometimes” scores, we calculated the number of
1066 observations of higher fitness of local populations per garden and per time measured to understand what
1067 proportion of gardens and sampling dates showed higher local fitness. This provides an estimate of the
1068 frequency of higher local fitness, but it is not a measure of the importance of the difference per se. For
1069 example, a fitness difference could occur at a low frequency, but have a large impact on population
1070 trajectories (i.e. large differences in survival after a rare drought event).

1071

1072 *Determining whether maternal effects were controlled*

1073 Experiments which tested populations that had all shared one or more generations in the same
1074 location prior to testing were considered to have attempted to control for maternal effects. We determined
1075 the number of generation in common by carefully reading the methods for mentions of the populations’
1076 lineages prior to testing. Some experiments supplied the original location of material collection but
1077 indicated that all materials were collected from areas such as ‘evaluation plots’, ‘seed fields’, ‘uniform

1078 gardens', or 'increase fields', indicating that at least one generation was shared among all populations,
1079 and therefore an attempt had been made to control maternal effects (intentional or not). Some complex
1080 studies had to be split into multiple experiments because they used different generations of the same
1081 populations in different tests. For example, a study which collected wild adults from their native habitats
1082 and grew them in a common garden for the duration of the experiment before measuring traits of the
1083 plants as well as traits of the seeds they produced were split into two experiments, one containing the
1084 traits of the adult plants (which did not attempt to control for maternal effects, because the progenitors of
1085 the measured material did not share a common location), and one for the seed traits (which did attempt to
1086 control for maternal effects, because the progenitors of the measured material did share a common
1087 location).

1088

1089 *Extraction for quantitative comparison of trait-by-environment association*

1090 To examine links between the variation in trait values and the variation in environmental and
1091 geographic distance among the population's origins, we utilized experiments from which population-
1092 specific trait data as well as geographic coordinates for at least one garden and at least two populations
1093 could be extracted or obtained through author contact. Data from laboratory and greenhouse experiments
1094 were not considered for this extraction, because the great majority of these experiments were not designed
1095 to completely simulate natural growing conditions. Excluding these experiments reduced our pool from
1096 325 to 161. Next, a list of priority fitness traits were developed (Table S1-1) based on traits that were
1097 most commonly measured and potentially associated with plant fitness in the Great Basin (Bower, Clair,
1098 and Erickson, 2014; Leger and Baughman, 2015). Any experiment that did not measure at least one
1099 priority trait was omitted from next steps, and this further reduced our pool from 161 to 153.

1100 Table S1-1. Priority traits targeted in the extraction for the dataset used in the quantitative comparison,
 1101 and the preferred units. Note that for several traits, several highly similar measures were included, as
 1102 indicated in footnotes.

Trait	units	Trait	units
survival	%	number - inflorescence ⁴	#
emergence	%	number – seeds ⁵	#
germination	%	number – leaves	#
height - plant	cm	date – germination	# days
length – root	cm	date – regrowth/greenup	# days
length – leaf ¹	cm	date – emergence	# days
dimensions – floral ²	cm	date – flowering ⁶	# days
mass – roots	g	date – seed shatter	# days
mass – shoots ³	g	date – senescence	# days
mass – seed	g/seed		

¹If unmeasured, then leaf width was recorded, if available

²Any measure of a floral structure

³Any measure of aboveground biomass

⁴Any kind of count of flowers or flowering structures was recorded

⁵If no direct count was available, any measure of seed yield was recorded, including total seed yield in weight and/or seed yield rating/rank

⁶If unmeasured, any other floral phenology was considered

1103 The remaining studies were then examined for textual, tabular, or visual data that could be
 1104 extracted as mean values of priority traits for each population in each garden. Extracted values for were
 1105 recorded verbatim from tables and throughout the text where possible, and from figures using
 1106 WebPlotDigitizer (Rohatgi, 2017) when needed. Means for at least two populations in at least one garden
 1107 were required for extraction. If exact matches to certain priority traits were not reported in the studies,
 1108 similar measures that were likely to be strongly correlated to the given trait could be recorded as
 1109 surrogates if available, and a note was made (footnotes, Table S1-1). We extracted the latest date for
 1110 which the most populations at the most gardens were represented if studies presented data for multiple
 1111 dates throughout the experiment. In some cases, experiments were conducted with multiple treatments in
 1112 which growing conditions were altered to address study questions. In these cases, we only extracted data
 1113 for the author-defined ‘control’ treatment. However, if no control was defined, we used the treatment that
 1114 was the most unaltered or representative of the garden environment (e.g. unweeded, or unwatered).

1115 **Appendix 2. Summary of literature and available datasets**

1116 The data collected and generated by this study (Baughman *et al.*, 2019), as well as the list of
1117 publications that were involved in each part of this study, are provided so that additional questions may be
1118 addressed and for other applications. We encourage such additional analyses.

1119 *Summary of literature*

1120 Appendix 2 Table 1. Summary, by species, of the literature included in this study, including
1121 lifeform (F = forb, G = grass, S = shrub, T = tree), counts of studies, experiments, unique populations, and
1122 experiments by type (LAB = laboratory, GH = greenhouse, CG = outdoor common garden, RT = outdoor
1123 reciprocal transplant), the incidence of each signature of local adaptation (1 = differences among
1124 populations, 2 = trait/environment correlations, 3 = higher performance of local than nonlocal population
1125 in local's environment), counts of experiments used in the quantitative comparison of trait-by-
1126 environment associations (QC), and a list of traits used in the QC. See footnotes for additional
1127 information.

Species	Lifeform	# Studies	# Expts.	# Pops ¹	# LAB expts.	# GH expts.	# CG expts.	# RT expts.	Signature 1 ²	Signature 2 ²	Signature 3 ³	# Expts. in QC	Traits in QC ⁴
<i>Achnatherum thurberianum</i>	G	2	2	68	-	1	1	-	2Y/ON	2Y/ON	-	1	1-7
<i>Allium acuminatum</i>	F	2	2	56	1	-	1	-	2Y/ON	2Y/ON	-	1	1,4-8
<i>Allium brandegeei</i>	F	1	1	3	1	-	-	-	1Y/ON	-	-	-	-
<i>Allium passeyi</i>	F	1	1	3	1	-	-	-	1Y/ON	-	-	-	-
<i>Amelanchier utahensis</i>	S	2	4	18*	-	-	4	-	4Y/ON	3Y/ON	-	-	-
<i>Andropogon scoparius</i>	G	5	6	55*	-	2	3	1	6Y/ON	4Y/ON	-	-	-
<i>Artemisia tridentata</i>	S	24	25	172*	7	-	14	4	22Y/3N	8Y/ON	2Y/5N	-	-
<i>Astragalus filipes</i>	F	1	1	67	-	-	1	-	1Y/ON	1Y/ON	-	1	1-4,8
<i>Atriplex canescens</i>	S	9	9	79	4	-	5	-	9Y/ON	3Y/ON	1Y/2N	2	1,2,8
<i>Atriplex confertifolia</i>	S	2	3	126	2	-	1	-	3Y/ON	3Y/ON	-	-	-
<i>Bouteloua gracilis</i>	G	5	6	66*	-	1	4	1	6Y/ON	4Y/ON	-	3	5
<i>Bromus carinatus</i>	G	1	1	193	-	-	1	-	1Y/ON	1Y/ON	-	1	1-6
<i>Carex aquatilis</i>	G	1	1	5	-	-	-	1	1Y/ON	1Y/ON	2Y/ON	-	-
<i>Cercocarpus montanus</i>	T	1	1	9	-	-	1	-	1Y/ON	0Y/1N	-	-	-
<i>Chaenactis douglasii</i>	F	1	1	15	-	-	1	-	1Y/ON	1Y/ON	-	1	1,2,4,8
<i>Chrysothamnus viscidiflorus</i>	S	4	4	8	-	-	4	-	2Y/2N	-	-	-	-
<i>Cistanthe umbellata</i>	F	1	2	5*	2	-	-	-	-	2Y/ON	-	-	-
<i>Claytonia perfoliata</i>	F	1	2	2*	-	-	-	2	2Y/ON	-	3Y/ON	-	-
<i>Cleome lutea lutea</i>	F	1	1	9	-	1	-	-	1Y/ON	1Y/ON	-	-	-
<i>Coleogyne ramosissima</i>	S	2	2	53	1	-	1	-	2Y/ON	2Y/ON	-	1	2,3
<i>Cryptantha circumscissa</i>	F	1	2	4*	2	-	-	-	-	1Y/1N	-	-	-
<i>Dalea ornata</i>	F	1	1	22	-	-	1	-	1Y/ON	1Y/ON	-	1	1,3-5
<i>Dalea searlsiae</i>	F	1	1	20	-	-	1	-	1Y/ON	1Y/ON	-	1	1,3,4
<i>Delphinium nelsonii</i>	F	1	1	3	-	-	-	1	1Y/ON	1Y/ON	2Y/ON	-	-
<i>Deschampsia cespitosa</i>	G	2	2	22*	-	-	2	-	2Y/ON	2Y/ON	0Y/1N	-	1,2,5,6
<i>Elymus canadensis</i>	G	3	4	30*	-	1	2	1	4Y/ON	2Y/ON	-	3	5
<i>Elymus elymoides</i>	G	19	24	170*	5	11	8	-	21Y/1N	6Y/4N	2Y/1N	3	1,3,5
<i>Elymus glaucus</i>	G	3	4	158*	1	1	1	1	4Y/ON	1Y/ON	2Y/ON	2	1,5-8
<i>Elymus multisetus</i>	G	7	7	36*	2	4	1	-	6Y/ON	3Y/1N	1Y/ON	1	1,5
<i>Elymus trachycaulus</i>	G	2	3	5	1	1	1	-	2Y/1N	-	-	-	-
<i>Encelia farinosa</i>	S	1	1	2	-	1	-	-	1Y/ON	1Y/ON	-	-	-
<i>Ephedra nevadensis</i>	S	1	1	2	-	-	1	-	1Y/ON	-	1Y/ON	-	-
<i>Epilobium densiflorum</i>	F	1	1	22	-	-	1	-	1Y/ON	1Y/ON	-	1	1,3,5-7
<i>Ericameria nauseosa</i>	S	16	17	64*	9	-	8	-	15Y/2N	3Y/ON	0Y/1N	-	-
<i>Eriogonum microthecum</i>	S	1	1	6	-	-	1	-	1Y/ON	1Y/ON	-	1	1-4,6,8
<i>Eriogonum ovalifolium</i>	S	1	1	6	-	-	1	-	1Y/ON	1Y/ON	-	1	1-4,6,8
<i>Eriogonum umbellatum</i>	S	1	1	5	-	-	1	-	1Y/ON	0Y/1N	-	-	-
<i>Erysimum capitatum</i>	F	2	4	9*	2	-	2	-	2Y/ON	1Y/1N	1Y/ON	-	-
<i>Festuca idahoensis</i>	G	5	6	43*	2	-	4	-	6Y/ON	2Y/1N	-	3	1,3
<i>Grayia spinosa</i>	S	2	2	7	1	-	1	-	2Y/ON	-	-	-	-
<i>Gutierrezia sarothrae</i>	S	3	4	6*	-	3	1	-	4Y/ON	0Y/1N	-	-	-
<i>Helianthus anomalus</i>	F	1	1	8	-	1	-	-	1Y/ON	1Y/ON	-	-	-
<i>Hesperostipa comata</i>	G	5	5	10*	-	2	3	-	3Y/2N	-	1Y/ON	-	-

<i>Holodiscus discolor</i>	S	1	1	39	-	-	1	-	1Y/0N	1Y/0N	-	-	-
<i>Koeleria macrantha</i>	G	4	5	29*	1	2	2	-	4Y/1N	-	-	2	1,5,6
<i>Krascheninnikovia lanata</i>	S	9	13	26*	7	5	-	1	13Y/0N	2Y/0N	3Y/0N	1	2,7
<i>Leptosiphon nuttallii</i>	F	1	1	3	1	-	-	-	-	1Y/0N	-	-	-
<i>Leymus cinereus</i>	G	6	9	141*	3	1	4	1	8Y/1N	3Y/0N	1Y/0N	3	1-7
<i>Linum lewisii</i>	F	3	6	22*	1	1	4	-	6Y/0N	2Y/0N	-	2	2,4,8
<i>Lomatium dissectum</i>	F	1	1	3	-	-	1	-	1Y/0N	-	-	-	-
<i>Lotus utahensis</i>	F	1	1	14	-	-	1	-	1Y/0N	0Y/1N	-	-	-
<i>Lupinus latifolius</i>	F	1	1	53	-	-	1	-	1Y/0N	1Y/0N	-	-	-
<i>Machaeranthera canescens</i>	F	1	1	9	-	-	1	-	1Y/0N	-	-	1	1,2,8
<i>Mimulus cardinalis</i>	F	1	1	6	-	-	1	-	1Y/0N	-	-	1	-
<i>Mimulus lewisii</i>	F	1	1	6	-	-	1	-	1Y/0N	-	-	1	-
<i>Nicotiana attenuata</i>	F	1	1	43	-	1	-	-	1Y/0N	0Y/1N	-	-	-
<i>Oxyria digyna</i>	F	1	2	11	2	-	-	-	-	0Y/2N	-	-	-
<i>Pascopyrum smithii</i>	G	5	11	42*	-	2	9	-	10Y/1N	1Y/0N	0Y/1N	5	1,3,4,6,8
<i>Penstemon acuminatus</i>	F	1	1	2	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon comarrhenus</i>	F	2	2	4*	2	-	-	-	2Y/0N	Y/1N	-	-	-
<i>Penstemon confusus</i>	F	1	1	3	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon cyananthus</i>	F	1	1	10	1	-	-	-	1Y/0N	-	-	-	-
<i>Penstemon cyanocaulis</i>	F	1	1	3	1	-	-	-	1Y/0N	-	-	-	-
<i>Penstemon deustus</i>	F	3	4	12*	2	-	2	-	4Y/0N	2Y/0N	-	1	2-4,6-8
<i>Penstemon eatonii</i>	F	2	3	13*	2	-	1	-	3Y/0N	2Y/0N	-	-	-
<i>Penstemon fruticosus</i>	F	1	1	3	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon humilis</i>	F	1	1	6	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon leiophyllus</i>	F	1	1	4	1	-	-	-	1Y/0N	-	-	-	-
<i>Penstemon leonardii</i>	F	1	1	3	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon linarioides</i>	F	1	1	2	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon newberryi</i>	F	1	2	2*	2	-	-	-	-	1Y/1N	-	-	-
<i>Penstemon pachyphyllus</i>	F	3	4	14*	2	-	2	-	4Y/0N	4Y/0N	-	1	2-4,6-8
<i>Penstemon palmeri</i>	F	1	2	13	2	-	-	-	2Y/0N	0Y/1N	-	-	-
<i>Penstemon petiolatus</i>	S	1	1	2	1	-	-	-	1Y/0N	0Y/1N	-	-	-
<i>Penstemon rostriflorus</i>	F	3	5	16*	3	-	2	-	5Y/0N	4Y/1N	-	1	2-4,6-8
<i>Penstemon rydbergii</i>	F	1	1	2	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon subglaber</i>	F	1	1	3	1	-	-	-	1Y/0N	-	-	-	-
<i>Penstemon utahensis</i>	F	1	1	5	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Penstemon watsonii</i>	F	1	1	4	1	-	-	-	1Y/0N	1Y/0N	-	-	-
<i>Plantago ovata</i>	F	1	2	12*	1	-	1	-	2Y/0N	2Y/0N	-	-	-
<i>Poa secunda</i>	G	7	8	158*	2	2	3	1	8Y/0N	3Y/0N	2Y/2N	2	1-7
<i>Polemonium viscosum</i>	F	1	2	2	-	1	-	1	2Y/0N	1Y/0N	2Y/0N	-	-
<i>Polygonum viviparum</i>	F	1	1	3	1	-	-	-	1Y/0N	-	-	-	-
<i>Populus angustifolia</i>	T	1	1	5	-	1	-	-	1Y/0N	-	-	-	-
<i>Potentilla pulcherrima</i>	F	1	1	15	-	-	1	-	1Y/0N	1Y/0N	-	-	-
<i>Pseudoroegneria spicata</i>	G	6	7	138	1	1	5	-	6Y/1N	1Y/0N	1Y/0N	3	1,3-7
<i>Purshia tridentata</i>	S	2	2	24	1	-	1	-	1Y/0N	0Y/2N	-	-	-
<i>Ranunculus flammula</i>	F	1	1	25	-	-	1	-	1Y/0N	1Y/0N	-	-	-
<i>Saxifraga oregana</i>	F	1	1	10	-	-	1	-	1Y/0N	1Y/0N	-	-	-
<i>Solidago gigantea</i>	F	1	1	7	-	1	-	-	1Y/0N	0Y/1N	-	-	-
<i>Solidago velutina</i>	F	1	1	6	-	-	1	-	1Y/0N	1Y/0N	-	-	-

<i>Sphaeralcea coccinea</i>	F	1	2	2*	-	-	2	-	2Y/0N	-	-	-	-
<i>Sphaeralcea grossulariifolia</i>	F	1	1	3	-	-	1	-	1Y/0N	-	-	-	-
<i>Sphaeralcea munroana</i>	F	1	2	49*	-	-	2	-	2Y/0N	-	-	-	-
<i>Sphaeralcea parvifolia</i>	F	1	1	7	-	-	1	-	1Y/0N	-	-	-	-
<i>Sporobolus airoides</i>	G	4	4	10	1	1	2	-	3Y/1N	3Y/0N	-	-	-
<i>Sporobolus cryptandrus</i>	G	3	4	30*	1	-	3	-	4Y/0N	4Y/0N	-	1	1,3,5
<i>Stellaria longipes</i>	F	1	2	7*	1	-	-	1	2Y/0N	-	-	-	-
<i>Stephanomeria minor</i>	F	1	2	3*	2	-	-	-	-	1Y/1N	-	-	-
<i>Stipa hymenoides</i>	G	15	17	397*	3	-	13	1	16Y/1N	3Y/4N	0Y/1N	2	1,3-8
<i>Symphoricarpos oreophilus</i>	S	1	1	25	-	-	1	-	1Y/0N	0Y/1N	-	-	-
<i>Typha latifolia</i>	F	1	1	2	1	-	-	-	1Y/0N	-	-	-	-
<i>Xanthium strumarium</i>	F	1	2	49*	1	-	1	-	2Y/0N	2Y/0N	-	1	5

¹Numbers followed by a * indicates a reduced count that accounts for the same populations used in multiple experiments

²Count of experiments; Y = "Yes" and "Authors claim Yes" scores, N = "No" and "Authors claim No" scores.

³Count of experiments; Y = Local did best in at least one garden during at least one sampling date, N = Local never did best

⁴Indicates which traits were used for this species in the quantitative comparison for trait-by-environment associations; 1 = height, 2 = survival, 3 = shoot mass, 4 = number of inflorescences, 5 = flowering day, 6 = leaf size, 7 = floral size, 8 = seed number.

1130

1131 Available datasets

1132 Data have been uploaded to Dryad at DOI: TBD (Baughman *et al.*, In Review). Several datasets
 1133 are available. The “Summary and signature scores” dataset includes all of the studies and experiments and
 1134 summarizes literature categorization as well as scores and associated information for each of the
 1135 signatures of local adaptation. The “Trait scores” dataset includes basic study categorization as well as
 1136 information that indicated which phenotypic traits (for signatures 1 and 2) and environmental variables
 1137 (for signature 2) were involved in each of the signatures of local adaptation. The “Quantitative
 1138 comparison” dataset includes all of the data used to conduct the quantitative comparison of trait-by-
 1139 environment associations, and lists population-specific mean values for our priority traits for all studies
 1140 for which such data were available, the latitude and longitude of each population origin, and extensive
 1141 climate information for each origin generated with the ClimateNA v5.10 software package based on
 1142 methodology described by Wang *et al.* (2016). The “Location data” dataset lists all outdoor gardens and
 1143 population origin coordinates and elevations for which authors gave this information, as well as those for

1144 which we could confidently generate it. For descriptions of each column in each of these datasets, refer to
1145 the “Data Dictionary” file.

1146

1147 *Bibliography of reviewed literature*

1148 A list of all the literature used in any of the datasets is provided below. Following each citation is
1149 a set of codes in brackets indicating which parts of our study the publication was used in. Codes S1, S2,
1150 and S3 indicate that at least one of the “experiments” in the given publication was used to generate a
1151 score for signatures 1, 2, and 3, and code QC indicates the publication (or the data summarized in it, even
1152 if not available from the publication itself) was used in analyses for the quantitative comparison of trait-
1153 by-environment associations. Note that some published studies were scored as multiple experiments for
1154 multiple species.

1155 Angert, A. L., and D. W. Schemske. 2005. “The Evolution of Species’ Distributions: Reciprocal
1156 Transplants across the Elevation Ranges of *Mimulus Cardinalis* and *M. Lewisii*.” *Evolution* 59
1157 (8):1671–84. <https://doi.org/10.1554/05-107.1>. [S1]

1158 Atwater, Daniel Z., and Ragan M. Callaway. 2015. “Testing the Mechanisms of Diversity-
1159 Dependent Overyielding in a Grass Species.” *Ecology* 96 (12):3332–42.
1160 <https://doi.org/10.1890/15-0889.1>. [S1, QC]

1161 Barker, Jr, and Cm McKell. 1986. “Differences in Big Sagebrush (*Artemisia Tridentata*) Plant
1162 Stature along Soil-Water Gradients: Genetic Components.” *Journal of Range Management* 39
1163 (2):147–51. <http://www.jstor.org/stable/10.2307/3899288>. [S1, QC]

1164 Barnes, Melanie G, and Diane L Marshall. 2009. “The Effect of Plant Source Location on
1165 Restoration Success: A Reciprocal Transplant Experiment with Winterfat (*Krascheninnikovia*
1166 *Lanata*).” *Ecology*. <https://doi.org/3390801>. [S1, S2, S3, QC]

1167 Baughman, Owen W., Susan E. Meyer, Zachary T. Aanderud, and Elizabeth A. Leger. 2016.
1168 “Cheatgrass Die-Offs as an Opportunity for Restoration in the Great Basin, USA: Will Local
1169 or Commercial Native Plants Succeed Where Exotic Invaders Fail?” *Journal of Arid*
1170 *Environments* 124:193–204. <https://doi.org/10.1016/j.jaridenv.2015.08.011>. [S1, S3, QC]

1171 Beckstead, Julie, Susan E. Meyer, and Phil S. Allen. n.d. “Effects of Afterripening on Cheatgrass
1172 (*Bromus Tectorum*) and Squirreltail (*Elymus Elymoides*) Germination.” In: Roundy, Bruce A.;
1173 McArthur, E. Durant; Haley, Jennifer S.; Mann, David K., Comps. 1995. Proceedings:
1174 Wildland Shrub and Arid Land Restoration Symposium; 1993 October 19-21; Las Vegas, NV.
1175 Gen. Tech. Rep. INT-GTR-315. Ogden, UT: U.S.D.A. [S1, S2]

- 1176 Bergum, Karin E., Ann L. Hild, and Brian A. Mealor. 2010. "Phenotypes of Two Generations of
1177 Sporobolus Airoides Seedlings Derived from Acroptilon Repens-Invaded and Non-Invaded
1178 Grass Populations." *Restoration Ecology* 20 (2):227–33. [https://doi.org/10.1111/j.1526-](https://doi.org/10.1111/j.1526-100X.2010.00754.x)
1179 [100X.2010.00754.x](https://doi.org/10.1111/j.1526-100X.2010.00754.x). [S1, S2]
- 1180 Bhattarai, Kishor, B. Shaun Bushman, Douglas A. Johnson, and John G. Carman. 2010. "Phenotypic
1181 and Genetic Characterization of Western Prairie Clover Collections from the Western United
1182 States." *Rangeland Ecology and Management* 63 (6):696–706. [https://doi.org/10.2111/REM-](https://doi.org/10.2111/REM-D-10-00008.1)
1183 [D-10-00008.1](https://doi.org/10.2111/REM-D-10-00008.1). [S1, S2, QC]
- 1184 Bhattarai, Kishor, Douglas A. Johnson, Thomas A. Jones, Kevin J. Connors, and Dale R. Gardner.
1185 2008. "Physiological and Morphological Characterization of Basalt Milkvetch (*Astragalus*
1186 *Filipes*): Basis for Plant Improvement." *Rangeland Ecology and Management* 61 (4):444–55.
1187 <https://doi.org/10.2111/08-011.1>. [S1, S2, QC]
- 1188 Bhattarai, Kishor, B. Shaun Bushman, Douglas A. Johnson, and John G. Carman. 2011. "Searls
1189 Prairie Clover (*Dalea Searlsiae*) for Rangeland Revegetation: Phenotypic and Genetic
1190 Evaluations." *Crop Science* 51 (2):716–27. <https://doi.org/10.2135/cropsci2010.07.0387>. [S1,
1191 S2, QC]
- 1192 Bleak, A.T., and Neil C. Frischknecht. 1965. "Problems in Artificial and Natural Revegetation of the
1193 Arid Shadscale Vegetation Zone of Utah and Nevada." *Journal of Range Management*, 59–65.
1194 [S1]
- 1195 Bohmont, B.L., and Robert Lang. 1957. "Some Variations in Morphological Characteristics and
1196 Palatability among Geographic Strains of Indian Ricegrass." *Journal of Range Management*,
1197 127–31.[S1]
- 1198 Booth, D. Terrance. 1992. "Seedbed Ecology of Winterfat: Imbibition Temperature Affects Post-
1199 Germination Growth." *Journal of Range Management* 45 (2):159–64. [S1]
- 1200 Booth, D. Terrance, Charles G. Howard, and Charles E. Mowry. 1980. "'Nezpar' Indian Ricegrass:
1201 Description, Justification for Release, and Recommendations for Use." *Rangelands Archives* 2
1202 (2):53–54. [S1]
- 1203 Brabec, Martha M., Matthew J. Germino, Douglas J. Shinneman, David S. Pilliod, Susan K.
1204 McIlroy, and Robert S. Arkle. 2015. "Challenges of Establishing Big Sagebrush (*Artemisia*
1205 *Tridentata*) in Rangeland Restoration: Effects of Herbicide, Mowing, Whole-Community
1206 Seeding, and Sagebrush Seed Sources." *Rangeland Ecology and Management* 68 (5):432–35.
1207 <https://doi.org/10.1016/j.rama.2015.07.001>. [S1, S3, QC]
- 1208 Brouillette, Larry C., Chase M. Mason, Rebecca Y. Shirk, and Lisa A. Donovan. 2014. "Adaptive
1209 Differentiation of Traits Related to Resource Use in a Desert Annual along a Resource
1210 Gradient." *New Phytologist* 201 (4):1316–27. <https://doi.org/10.1111/nph.12628>. [S1, S2]
- 1211 Butterfield, Bradley J., and Troy E. Wood. 2015. "Local Climate and Cultivation, but Not Ploidy,
1212 Predict Functional Trait Variation in *Bouteloua Gracilis* (Poaceae)." *Plant Ecology* 216
1213 (10):1341–49. <https://doi.org/10.1007/s11258-015-0510-8>. [S1, S2]
- 1214 Chabot, Brian F., and W. D. Billings. 1972. "Origins and Ecology of the Sierran Alpine Flora and
1215 Vegetation." *Ecological Monographs* 42 (2):163–99. <https://doi.org/10.2307/1942262>. [S2]

- 1216 Chaney, Lindsay, Bryce A. Richardson, and Matthew J. Germino. 2017. "Climate Drives Adaptive
1217 Genetic Responses Associated with Survival in Big Sagebrush (*Artemisia Tridentata*)."
1218 *Evolutionary Applications* 10 (4):313–22. <https://doi.org/10.1111/eva.12440>. [S1, S2, QC]
- 1219 Chapin, F. S. III, and C. M. Chapin. 1981. "Ecotypic Differentiation of Growth Processes in *Carex*
1220 *Aquaticilis* along a Latitudinal and Local Gradients." *Ecology* 62 (4):1000–1009.
1221 <https://doi.org/10.2307/1936999>. [S1, S2, S3, QC]
- 1222 Clark, Lesley D., and Neil E. West. 1971. "Further Studies of *Eurotia Lanata* Germination in
1223 Relation to Salinity." *The Southwestern Naturalist*, 371–75. [S1]
- 1224 Clary, Warren P. 1975. "Ecotypic Adaptation in *Sitanion Hystrix*." *Ecology* 56 (6):1407–15. [S1,
1225 S2, QC]
- 1226 Clary, Warren P. 1979. "Variation in Leaf Anatomy and CO₂ Assimilation in *Sitanion Hystrix*
1227 Ecotypes." *The Great Basin Naturalist*, 427–32. [S1, S2]
- 1228 Clauss, M. J., and D. L. Venable. 2000. "Seed Germination in Desert Annuals: An Empirical Test of
1229 Adaptive Bet Hedging." *The American Naturalist* 155 (2):168–86.
1230 <https://doi.org/10.1086/303314>. [S1, S2, QC]
- 1231 Cook, Stanton A, and Michael P Johnson. 1968. "Adaptation to Heterogeneous Environments. I.
1232 Variation in Heterophylly in *Ranunculus Flammula* L." *Evolution* 22 (3):496–516.
1233 <https://doi.org/10.1111/j.1558-5646.1968.tb03988.x>. [S1, S2]
- 1234 Dewey, Douglas R. 1960. "Salt Tolerance of Twenty-Five Strains of *Agropyron*." *Agronomy*
1235 *Journal* 52 (11):631–35. <https://doi.org/10.2134/agronj1960.00021962005200110006x>. [S1,
1236 QC]
- 1237 Doede, David L. 2005. "Genetic Variation in Broadleaf Lupine (*Lupinus Latifolius*) on the Mt Hood
1238 National Forest and Implications for Seed Collection and Deployment." *Native Plants Journal*
1239 6 (1):36–48. <https://doi.org/10.1353/npj.2005.0018>. [S1, S2]
- 1240 Doescher, P.S. 1983. "Phyto-Edaphic Relationships and Ecotypic Development of *Festuca*
1241 *Idahoensis* in Eastern Oregon Habitat Types of *Artemisia Tridentata*." Ph.D. Dissertation,
1242 Oregon State University, Corvallis. [S1, S2, QC]
- 1243 Emery, R. J. N., C. C. Chinnappa, and J. G. Chmielewski. 1994. "Specialization, Plant Strategies,
1244 and Phenotypic Plasticity in Populations of *Stellaria Longipes* Along an Elevational Gradient."
1245 *International Journal of Plant Sciences* 155 (2):203–19. <https://doi.org/10.2307/2995565>. [S1,
1246 QC]
- 1247 Erickson, Vicky J, Nancy L Mandel, and Frank C Sorenson. 2004. "Landscape Patterns of
1248 Phenotypic Variation and Population Structuring in a Selfing Grass, *Elymus Glaucus* (Blue
1249 Wildrye)." *Canadian Journal of Botany* 82:1776–89. <https://doi.org/10.1139/B04-141>. [S1, S2,
1250 QC]
- 1251 Evans, Raymond A, and James A Young. 1990. "Survival and Growth of Big Sagebrush (*Artemisia*
1252 *Tridentata*) Plants in Reciprocal Gardens." *Weed Science* 38 (3):215–19.
1253 <https://doi.org/10.2307/4045014>. [S1, S3, QC]
- 1254 Ferguson, Scot D., Elizabeth A. Leger, Jun Li, and Robert S. Nowak. 2015. "Natural Selection

- 1255 Favors Root Investment in Native Grasses during Restoration of Invaded Fields.” *Journal of*
1256 *Arid Environments* 116:11–17. <https://doi.org/10.1016/j.jaridenv.2015.01.009>. [S1]
- 1257 Ferrero-Serrano, Ángel, Ann L. Hild, and Brian A. Mealor. 2011. “Can Invasive Species Enhance
1258 Competitive Ability and Restoration Potential in Native Grass Populations?” *Restoration*
1259 *Ecology* 19 (4):545–51. <https://doi.org/10.1111/j.1526-100X.2009.00611.x>. [S1, S2]
- 1260 Fisk, Matthew R. 2016. “Dynamics of Cold Hardiness Accumulation and Loss in the Great Basin
1261 Native Species *Eriogonum Umbellatum*.” Ph.D. Dissertation, University of Idaho, Boise. [S1,
1262 S2]
- 1263 Fitzsimmons, Kevin, Cynthia Lovely, and Edward Glenn. 1998. “Growth Differences among
1264 Widely Separated Geographic Accessions of Fourwing Saltbush (*Atriplex Canescens*) in the
1265 Great Basin Desert, New Mexico, USA.” *Arid Soil Research and Rehabilitation* 12 (2):87–94.
1266 <https://doi.org/10.1080/15324989809381501>. [S1, S2, QC]
- 1267 Fonseca, Carolina, Erin Espeland, and James W. Baxter. 2014. “Patterns of Population
1268 Differentiation in Early Traits of Development in *Elymus Glaucus*: Implications for
1269 Restoration.” *Ecological Restoration* 32 (4):388–95. <https://doi.org/10.3368/er.32.4.388>. [S1]
- 1270 Galen, Candace, Joel S Shore, and Hudson Deyoe. 1991. “Ecotypic Divergence in Alpine
1271 *Polemonium Viscosum*: Genetic Structure, Quantitative Variation, and Local Adaptation.”
1272 *Evolution* 45 (455):1218–28. <https://doi.org/10.2307/2409729>. [S1, S2, S3, QC]
- 1273 Goergen, Erin M., Elizabeth A. Leger, and Erin K. Espeland. 2011. “Native Perennial Grasses Show
1274 Evolutionary Response to *Bromus Tectorum* (Cheatgrass) Invasion.” *PLoS ONE* 6 (3).
1275 <https://doi.org/10.1371/journal.pone.0018145>. [S1, S2]
- 1276 Hall, J.W., D.G. Stout, and B. Brooke. 1990. “Effect of Seed Source on Growth of Giant Wildrye
1277 (*Elymus Cinereus*) at Two Elevations in Interior British Columbia.” *Canadian Journal of Plant*
1278 *Science* 70 (2):551–54. [S1, S3, QC]
- 1279 Hardegree, Stuart P., Thomas A. Jones, Frederick B. Pierson, Patrick E. Clark, and Gerald N.
1280 Flerchinger. 2008. “Dynamic Variability in Thermal-Germination Response of Squirreltail
1281 (*Elymus Elymoides* and *Elymus Multisetus*).” *Environmental and Experimental Botany* 62
1282 (2):120–28. <https://doi.org/10.1016/j.envexpbot.2007.07.010>. [S2]
- 1283 Harmon, Dan, and Charlie D. Clements. 2016. “Characteristics That Determine a Successful
1284 Squirreltail (*Elymus Elymoides*).” In Poster Session Presented at the Society for Range
1285 Management, Corpus Christi, TX. [S1]
- 1286 Hergert, Holden J., Brian A. Mealor, and Andrew R. Kniss. 2015. “Inter-and Intraspecific Variation
1287 in Native Restoration Plants for Herbicide Tolerance.” *Ecological Restoration* 33 (1):74–81.
1288 <https://doi.org/10.3368/er.33.1.74>. [S1]
- 1289 Hild, A L, J M Muscha, and N L Shaw. 2007. “Emergence and Growth of Four Winterfat
1290 Accessions in the Presence of the Exotic Annual Cheatgrass.” *Proceedings: Shrubland*
1291 *Dynamics-Fire and Water*; 2004 August 10-12; Lubbock, TX., no. 47:0–147. [S1]
- 1292 Hintz, Lisa, M.M. Eshelman, A. Foxx, T.E. Wood, and A. Kramer. 2016. “Population
1293 Differentiation in Early Life History Traits of *Cleome Lutea* Var. *Lutea* in the Intermountain
1294 West.” *Western North American Naturalist* 76 (1):6–17. [S1, S2]

- 1295 Horning, Matthew E., Theresa R. McGovern, Dale C. Darris, Nancy L. Mandel, and Randy Johnson.
1296 2010. "Genecology of *Holodiscus Discolor* (Rosaceae) in the Pacific Northwest, U.S.A."
1297 *Restoration Ecology* 18 (2):235–43. <https://doi.org/10.1111/j.1526-100X.2008.00441.x>. [S1,
1298 S2]
- 1299 Humphrey, L. David, and Eugene W. Schupp. 2002. "Seedling Survival from Locally and
1300 Commercially Obtained Seeds on Two Semiarid Sites." *Restoration Ecology* 10 (1):88–95.
1301 <https://doi.org/10.1046/j.1526-100X.2002.10109.x>. [S1, S3, QC]
- 1302 Jaindl, Raymond G., Paul Doescher, Richard F. Miller, and Lee E. Eddleman. 1994. "Persistence of
1303 Idaho Fescue on Degraded Rangelands: Adaptation to Defoliation or Tolerance." *Journal of*
1304 *Range Management* 47 (1):54. <https://doi.org/10.2307/4002841>. [S1, S2, QC]
- 1305 Johnson, R C, V J Erickson, N L Mandel, J Bradley St Clair, and K W Vance-Borland. 2010.
1306 "Mapping Genetic Variation and Seed Zones for *Bromus Carinatus* in the Blue Mountains of
1307 Eastern Oregon, USA." *Botany* 88 (8):725–36. <https://doi.org/10.1139/B10-047>. [S1, S2, QC]
- 1308 Johnson, R C, B C Hellier, and K W Vance-Borland. 2013. "Genecology and Seed Zones for
1309 Tapertip Onion in the US Great Basin." *Botany-Botanique* 91 (10):686–94. [https://doi.org/DOI](https://doi.org/DOI.10.1139/cjb-2013-0046)
1310 [10.1139/cjb-2013-0046](https://doi.org/DOI.10.1139/cjb-2013-0046). [S1, S2, QC]
- 1311 Johnson, R. C., M. J. Cashman, and K. Vance-Borland. 2012. "Genecology and Seed Zones for
1312 Indian Ricegrass Collected in the Southwestern United States." *Rangeland Ecology and*
1313 *Management* 65 (5):523–32. <https://doi.org/10.2111/REM-D-11-00165.1>. [S1, S2, QC]
- 1314 Johnson, R. C., E. A. Leger, and Ken Vance-Borland. 2017. "Genecology of Thurber's Needlegrass
1315 (*Achnatherum Thurberianum* [Piper] Barkworth) in the Western United States." *Rangeland*
1316 *Ecology and Management* 70 (4):509–17. <https://doi.org/10.1016/j.rama.2017.01.004>. [S1, S2,
1317 QC]
- 1318 Johnson, R. C., and Ken Vance-Borland. 2016. "Linking Genetic Variation in Adaptive Plant Traits
1319 to Climate in Tetraploid and Octoploid Basin Wildrye [*Leymus Cinereus* (Scribn. & Merr.) A.
1320 Love] in the Western U.S." *PLoS ONE* 11 (2). <https://doi.org/10.1371/journal.pone.0148982>.
1321 [S1, S2, QC]
- 1322 Johnson, Richard C., Matthew E. Horning, Erin K. Espeland, and Ken Vance-Borland. 2015.
1323 "Relating Adaptive Genetic Traits to Climate for Sandberg Bluegrass from the Intermountain
1324 Western United States." *Evolutionary Applications* 8 (2):172–84.
1325 <https://doi.org/10.1111/eva.12240>. [S1, S2, QC]
- 1326 Jones, T A, D C Nielson, J T Arredondo, and M G Redinbaugh. 2003. "Characterization of
1327 Diversity among 3 Squirreltail Taxa." *Journal of Range Management* 56 (5):474–82.
1328 <https://doi.org/Doi.10.2307/4003839>. [S1, QC]
- 1329 Jones, T.A. 2004. "Registration of Ribstone Indian Ricegrass Germplasm." *Crop Science* 44
1330 (3):1031–33. [S1]
- 1331 Jones, T.A., D.C. Nielson, S.K. Caicco, G.A. Fenchel, and S. A. Young. 2005. "Registration of Star
1332 Lake Indian Ricegrass Germplasm." *Crop Science* 45 (4):1666–67. [S1]
- 1333 Jones, Thomas A., S.R. Winslow, S.D. Parr, and K.L. Memmott. 2010. "Notice of Release of White
1334 River Germplasm Indian Ricegrass." *Native Plants Journal* 11 (2):133–36. [S1]

- 1335 Kardol, P., J. R. De Long, and D. A. Wardle. 2014. "Local Plant Adaptation across a Subarctic
1336 Elevational Gradient." *Royal Society Open Science* 1 (3):140141–140141.
1337 <https://doi.org/10.1098/rsos.140141>. [S1]
- 1338 Kim, Eunsuk, and Kathleen Donohue. 2013. "Local Adaptation and Plasticity of *Erysimum*
1339 *Capitatum* to Altitude: Its Implications for Responses to Climate Change." *Journal of Ecology*
1340 101 (3):796–805. <https://doi.org/10.1111/1365-2745.12077>. [S1, S3, QC]
- 1341 Kitchen, Stanley G. n.d. "Return of the Native: A Look at Select Accessions of North American
1342 Lewis Flax." In: Roundy, Bruce A.; McArthur, E. Durant; Haley, Jennifer S.; Mann, David K.,
1343 Comps. 1995. *Proceedings: Wildland Shrub and Arid Land Restoration Symposium; 1993*
1344 *October 19-21; Las Vegas, NV. Gen. Tech. Rep. INT-GTR-315. Ogden, UT: U.S.D.A.* [S1]
- 1345 Kitchen, Stanley G., and Loren St. John. 1996. "Release Documentation for Maple Grove Lewis
1346 Flax." USDA NRCS Aberdeen Plant Materials Center, Aberdeen, Idaho.
1347 https://www.nrcs.usda.gov/Internet/FSE_PLANTMATERIALS/publications/idpmcrn5639.pdf.
1348 [S1, S2, QC]
- 1349 Kramer, Andrea T. 2009. "Ecological Genetics of *Penstemon* in the Great Basin, USA." Ph.D.
1350 Dissertation, University of Illinois at Chicago. [S1, S2]
- 1351 Kramer, Andrea T., Daniel J. Larkin, and Jeremie B. Fant. 2015. "Assessing Potential Seed Transfer
1352 Zones for Five Forb Species from the Great Basin Floristic Region, USA." *Natural Areas*
1353 *Journal* 35 (1):174–88. [S1, S2, QC]
- 1354 Larsen, Eugene C. 1947. "Photoperiodic Responses of Geographical Strains of *Andropogon*
1355 *Scoparius*." *Botanical Gazette* 109 (2):132–49. [S1, S2, QC]
- 1356 Leger, Elizabeth A. 2008. "The Adaptive Value of Remnant Native Plants in Invaded Communities:
1357 An Example from the Great Basin." *Ecological Applications* 18 (5):1226–35. [S1, S2]
- 1358 Li, Dapeng, Ian T. Baldwin, and Emmanuel Gaquerel. 2015. "Navigating Natural Variation in
1359 Herbivory-Induced Secondary Metabolism in Coyote Tobacco Populations Using MS/MS
1360 Structural Analysis." *Proceedings of the National Academy of Sciences* 112 (30):E4147–55.
1361 [S1, S2]
- 1362 Liao, Huixuan, Priscila C.S. Gurgel, Robert W. Pal, David Hooper, and Ragan M. Callaway. 2016.
1363 "Solidago Gigantea Plants from Nonnative Ranges Compensate More in Response to Damage
1364 than Plants from the Native Range." *Ecology* 97 (9):2355–63.
1365 <https://doi.org/10.1002/ecy.1481>. [S1, S2]
- 1366 Link, Steven O., Jeffrey L. Smith, Jonathan J. Halvorson, and Harvey Bolton. 2003. "A Reciprocal
1367 Transplant Experiment within a Climatic Gradient in a Semiarid Shrub-Steppe Ecosystem:
1368 Effects on Bunchgrass Growth and Reproduction, Soil Carbon, and Soil Nitrogen." *Global*
1369 *Change Biology* 9 (7):1097–1105. <https://doi.org/10.1046/j.1365-2486.2003.00647.x>. [S1, S3,
1370 QC]
- 1371 Love, Stephen L, Robert R Tripepi, and Thomas Salaiz. 2014. "Influence of Harvest Timing and
1372 Storage Interval on Rabbitbrush Seed Germination, Emergence, and Viability." *Native Plants*
1373 *Journal* (University of Wisconsin Press) 15 (2):98–108. <https://doi.org/10.1353/npj.2014.0017>.
1374 [S1]

- 1375 Mann, Rebecca K. 2016. "Intraspecific Variation in the Response of *Elymus Elymoides* to
1376 Competition from *Bromus Tectorum*." Ph.D. Dissertation, Utah State University, Logan. [S1,
1377 S2]
- 1378 McArthur, E. Durant, Susan E. Meyer, and Darrel J. Weber. 1987. "Germination Rate at Low
1379 Temperature: Rubber Rabbitbrush Population Differences." *Journal of Range Management*,
1380 530–33. [S1, S2]
- 1381 McArthur, E. Durant, Richard Stevens, and A. Clyde Blauer. 1983. "Growth Performance
1382 Comparisons among 18 Accessions of Fourwing Saltbush [*Atriplex Canescens*] at Two Sites in
1383 Central Utah." *Journal of Range Management*, 78–81. [S1, S3, QC]
- 1384 McArthur, E. Durant, and Bruce L. Welch. 1982. "Growth Rate Differences among Big Sagebrush
1385 [*Artemisia Tridentata*] Accessions and Subspecies." *Journal of Range Management* 35
1386 (3):396–401. <https://doi.org/10.2307/3898327>. [S1, QC]
- 1387 McIntyre, Patrick J., and Sharon Y. Strauss. 2014. "Phenotypic and Transgenerational Plasticity
1388 Promote Local Adaptation to Sun and Shade Environments." *Evolutionary Ecology* 28
1389 (2):229–46. <https://doi.org/10.1007/s10682-013-9670-y>. [S1, S3, QC]
- 1390 McMillan, Calvin. 1957. "Nature of the Plant Community. III. Flowering Behavior within Two
1391 Grassland Communities under Reciprocal Transplanting." *American Journal of Botany*, 144–
1392 53. [S1, QC]
- 1393 McMillan, Calvin. 1959a. "Nature of the Plant Community. V. Variation within the True Prairie
1394 Community-Type." *American Journal of Botany*, 418–24. [S1, QC]
- 1395 McMillan, Calvin. 1959b. "The Role of Ecotypic Variation in the Distribution of the Central
1396 Grassland of North America." *Ecological Monographs* 29 (4):286–308. <https://doi.org/Genetic>
1397 Considerations in Ecological Restoration. [S1, S2, QC]
- 1398 McNaughton, S.J. 1966. "Thermal Inactivation Properties of Enzymes from *Typha Latifolia* L.
1399 Ecotypes." *Plant Physiology* 41:1736–38. <https://doi.org/10.2307/4260909>. [S1]
- 1400 Meador, Brian A., and Ann L. Hild. 2007. "Post-Invasion Evolution of Native Plant Populations: A
1401 Test of Biological Resilience." *Oikos* 116 (9):1493–1500. [https://doi.org/10.1111/j.2007.0030-
1402 \[1299.15781.x\]\(https://doi.org/10.1111/j.2007.0030-1299.15781.x\)](https://doi.org/10.1111/j.2007.0030-). [S1, S2]
- 1403 Messina, Frank J., Susan L. Durham, James H. Richards, and E. Durant McArthur. 2002. "Trade-off
1404 between Plant Growth and Defense? A Comparison of Sagebrush Populations." *Oecologia* 131
1405 (1):43–51. <https://doi.org/10.1007/s00442-001-0859-3>. [S1, QC]
- 1406 Meyer, S. E., and S. G. Kitchen. 1994. "Life History Variation in Blue Flax (*Linum Perenne*:
1407 Linaceae): Seed Germination Phenology." *American Journal of Botany* 81 (5):528–35.
1408 <https://doi.org/10.2307/2445726>. [S1, S2, QC]
- 1409 Meyer, S. E., S. G. Kitchen, and S. L. Carlson. 1995. "Seed Germination Timing Patterns in
1410 Intermountain Penstemon (*Scrophulariaceae*)." *American Journal of Botany*.
1411 <https://doi.org/10.2307/2445584>. [S1, S2]
- 1412 Meyer, S. E., E. D. McArthur, and G. L. Jorgensen. 1989. "Variation in Germination Response to
1413 Temperature in Rubber Rabbitbrush (*Chrysothamnus Nauseosus*: Asteraceae) and Its

- 1414 Ecological Implications.” *American Journal of Botany*. <https://doi.org/10.2307/2444519>. [S1]
- 1415 Meyer, S. E., and S. B. Monsen. 1992. “Big Sagebrush Germination Patterns: Subspecies and
1416 Population Differences.” *Journal of Range Management* 45 (1):87–93.
1417 <https://doi.org/10.2307/4002533>. [S1, S2, QC]
- 1418 Meyer, SE, J Beckstead, PS Allen, and H Pullman. 1995. “Germination Ecophysiology of *Leymus*
1419 *Cinereus* (Poaceae).” *International Journal of Plant Sciences* 156 (2):206–15.
1420 <https://doi.org/10.1086/297242>. [S1, S2, QC]
- 1421 Meyer, Susan E. 1992. “Habitat Correlated Variation in Firecracker Penstemon (*Penstemon Eatonii*
1422 Gray: Scrophulariaceae) Seed Germination Response.” *Bulletin of the Torrey Botanical Club*
1423 119 (3):268–79. <https://doi.org/10.2307/2996758>. [S1, S2]
- 1424 Meyer, Susan E. 1997. “Ecological Correlates of Achene Mass Variation in *Chrysothamnus*
1425 *Nauseosus* (Asteraceae).” *American Journal of Botany* 84 (4):471–77.
1426 <https://doi.org/10.2307/2446023>. [S1, S2, QC]
- 1427 Meyer, Susan E., and Stephanie L. Carlson. n.d. “Seed Germination Biology of Intermountain
1428 Populations of Fourwing Saltbush (*Atriplex Canescens*: Chenopodiaceae).” In: Sosebee,
1429 Ronald E.; Wester, David B.; Britton, Carlton M.; McArthur, E. Durant; Kitchen, Stanley G.,
1430 Comps. 2007. *Proceedings: Shrubland Dynamics—fire and Water; 2004 August 10-12;*
1431 *Lubbock, TX. Proceedings RMRS-P-47. Fort Collins, CO: U.S.D.A.* [S1, S2]
- 1432 Meyer, Susan E., Stephanie L. Carlson, and Susan C. Garvin. 1998. “Seed Germination Regulation
1433 and Field Seed Bank Carryover in Shadscale (*Atriplex Confertifolia*: Chenopodiaceae).”
1434 *Journal of Arid Environments* 38 (2):255–67. <https://doi.org/10.1006/jare.1997.0321>. [S1, S2,
1435 QC]
- 1436 Meyer, Susan E., and Stephen B. Monsen. 1991. “Habitat-Correlated Variation in Mountain Big
1437 Sagebrush (*Artemisia Tridentata* Ssp. *Vaseyana*) Seed Germination Patterns.” *Ecology* 72
1438 (2):739–42. [S1, S2]
- 1439 Meyer, Susan E., Stephen B. Monsen, and E. Durant McArthur. 1990. “Germination Response of
1440 *Artemisia Tridentata* (Asteraceae) to Light and Chill: Patterns of Between-Population
1441 Variation.” *Botanical Gazette*. <https://doi.org/10.1086/337817>. [S1, S2]
- 1442 Meyer, Susan E, and Stanley G Kitchen. 1994. “Habitat-Correlated Variation in Seed Germination
1443 Response to Chilling in *Penstemon Section Glabri* (Scrophulariaceae).” *American Midland*
1444 *Naturalist* 132 (2):349–65. <https://doi.org/10.2307/2426591>. [S1]
- 1445 Miller, Roy V. 1967. “Ecotypic Variation in *Andropogon Scoparius* and *Bouteloua Gracilis*.” Ph.D.
1446 Dissertation, Colorado State University, Fort Collins. [S1, S2]
- 1447 Miller, Stephanie A., Amy Bartow, Melanie Gisler, Kimiora Ward, Amy S. Young, and Thomas N.
1448 Kaye. 2011. “Can an Ecoregion Serve as a Seed Transfer Zone? Evidence from a Common
1449 Garden Study with Five Native Species.” *Restoration Ecology* 19 (201):268–76.
1450 <https://doi.org/10.1111/j.1526-100X.2010.00702.x>. [S1, S2, QC]
- 1451 Monaco, T. A., S. B. Monsen, B. N. Smith, and L. D. Hansen. 2005. “Temperature-Dependent
1452 Physiology of *Poa Secunda*, a Cool Season Grass Native to the Great Basin, United States.”
1453 *Russian Journal of Plant Physiology* 52 (5):653–58. <https://doi.org/10.1007/s11183-005-0096->

- 1454 [4](#). [S1]
- 1455 Monson, R K, S D Smith, J L Gehring, W D Bowman, S R Szarek, British Ecological Society, and
1456 Functional Ecology. 1992. “Physiological Differentiation within an *Encelia Farinosa*
1457 Population along a Short Topographic Gradient in the Sonoran Desert.” *Functional Ecology* 6
1458 (6):751–59. [S1, S2]
- 1459 Moyer, J.L., and R. L. Lang. 1976. “Variable Germination Response to Temperature for Different
1460 Sources of Winterfat Seed.” *Journal of Range Management* 29:320–21. [S1]
- 1461 Mummey, Daniel L., M.E. Herget, K.M. Hufford, and L. Shreading. 2016. “Germination Timing
1462 and Seedling Growth of *Poa Secunda* and the Invasive Grass, *Bromus Tectorum*, in Response
1463 to Temperature: Evaluating Biotypes for Seedling Traits That Improve Establishment.”
1464 *Ecological Restoration* 34 (3):200–208. [S1, S2]
- 1465 Munda, B.D., S.M. Lambert, and J.C. Garrison. 1990. “Registration of ‘Santa Rita’ Fourwing
1466 Saltbush.” *Crop Science* 30 (6). [S1]
- 1467 Nasri, Mohamed, and Paul S. Doescher. 1995. “Effect of Temperature on Growth of Cheatgrass and
1468 Idaho Fescue.” *Journal of Range Management*, 406–9. [S1]
- 1469 Orodho, A B, R L Cuany, and M J Trlica. 1998. “Previous Grazing or Clipping Affects Seed of
1470 Indian Ricegrass.” *Journal of Range Management* 51 (1):37–41. <https://doi.org/none>. [S1, S2]
- 1471 Orodho, Apollo B., and M J Trlica. 1990. “Clipping and Long-Term Grazing Effects on Biomass
1472 and Carbohydrate Reserves of Indian Ricegrass.” *Journal of Range Management*, 52–57. [S1,
1473 S2]
- 1474 Par, Steve, and Marti Walsh. 2008. Notice of Release of Long Ridge Germplasm Utah Serviceberry.
1475 Upper Colorado Environmental Plant Center, USDA NRCS, Colorado State Agricultural
1476 Experiment Station.
1477 https://www.nrcs.usda.gov/Internet/FSE_PLANTMATERIALS/publications/copmcrn8043.pdf
1478 [S1]
- 1479 Parsons, Matthew C., Thomas A. Jones, Steven R. Larson, Ivan W. Mott, and Thomas A. Monaco.
1480 2011. “Ecotypic Variation in *Elymus Elymoides* Subsp. *Brevifolius* in the Northern
1481 Intermountain West.” *Rangeland Ecology and Management* 64 (6):649–58.
1482 <https://doi.org/10.2111/REM-D-09-00143.1>. [S1]
- 1483 Parsons, Matthew C., Thomas A. Jones, and Thomas A. Monaco. 2011. “Genetic Variation for
1484 Adaptive Traits in Bottlebrush Squirreltail in the Northern Intermountain West, United States.”
1485 *Restoration Ecology* 19 (4):460–69. <https://doi.org/10.1111/j.1526-100X.2010.00705.x>. [S1,
1486 S2]
- 1487 Percy, R W, and R T Ward. 1972. “Phenology and Growth of Rocky Mountain Populations of
1488 *Deschampsia Caespitosa* at Three Elevations in Colorado.” *Ecology* 53:1171–78.
1489 <https://doi.org/10.2307/1935431>. [S1, S2, S3, QC]
- 1490 Pendleton, B. K., and S. E. Meyer. 2004. “Habitat-Related Variation in Blackbrush (*Coleogyne*
1491 *Ramosissima*: Rosaceae) Seed Germination Response.” *Journal of Arid Environments* 59
1492 (2):229–43. <https://doi.org/10.1016/j.jaridenv.2003.12.009>. [S1, S2]

- 1493 Petersen, J L, D N Ueckert, R L Potter, and J E Huston. 1987. "Ecotypic Variation in Selected
1494 Fourwing Saltbush Populations in Western Texas USA." *Journal of Range Management* 40
1495 (4):361–66. <https://doi.org/10.2307/361>. [S1, S3, QC]
- 1496 Phillips, Nathan C., Daniel T. Drost, William A. Varga, Leila M. Shultz, and Susan E. Meyer. 2010.
1497 "Germination Characteristics along Altitudinal Gradients in Three Intermountain Allium
1498 spp.(Amaryllidaceae)." *Seed Technology* 32 (1):15–25. <http://www.jstor.org/stable/23433619>.
1499 [S1, S2]
- 1500 Polley, H. W., and J. K. Detling. 1988. "Herbivory Tolerance of *Agropyron Smithii* Populations
1501 with Different Grazing Histories." *Oecologia* 77 (2):261–67.
1502 <https://doi.org/10.1007/BF00379196>. [S1, S2]
- 1503 Potter, R.L., D.N. Ueckert, and J L Petersen. 1986. "Germination of Fourwing Saltbush Seeds:
1504 Interaction of Temperature, Osmotic Potential, and pH." *Journal of Range Management*, 43–
1505 46. [S1]
- 1506 Quinn, James A., and Richard T Ward. 1969. "Ecological Differentiation in Sand Dropseed
1507 (*Sporobolus Cryptandrus*)." *Ecological Monographs* 39 (1):61–78. [S1, S2, QC]
- 1508 Quinn, James A, and Jeffrey D Wetherington. 2002. "Genetic Variability and Phenotypic Plasticity
1509 in Flowering Phenology in Populations of Two Grasses." *Journal of the Torrey Botanical
1510 Society* 129 (2):96–106. <https://doi.org/10.2307/3088723>. [S1, S2]
- 1511 Ray, Peter M., and William E. Alexander. 1966. "Photoperiodic Adaptation to Latitude in *Xanthium
1512 Strumarium*." *American Journal of Botany*, 806–16. [S1, S2, QC]
- 1513 Rice, Kevin J., and Eric E. Knapp. 2008. "Effects of Competition and Life History Stage on the
1514 Expression of Local Adaptation in Two Native Bunchgrasses." *Restoration Ecology* 16 (1):12–
1515 23. <https://doi.org/10.1111/j.1526-100X.2007.00257.x>. [S1, S3, QC]
- 1516 Richardson, Bryce A., Stanley G. Kitchen, Rosemary L. Pendleton, Burton K. Pendleton, Matthew
1517 J. Germino, Gerald E. Rehfeldt, and Susan E. Meyer. 2014. "Adaptive Responses Reveal
1518 Contemporary and Future Ecotypes in a Desert Shrub." *Ecological Applications* 24 (2):413–
1519 27. <https://doi.org/10.1890/13-0587.1>. [S1, S2]
- 1520 Richardson, Bryce A., Hector G. Ortiz, Stephanie L. Carlson, Deidre M. Jaeger, Nancy L. Shaw,
1521 and D. P.C. Peters. 2015. "Genetic and Environmental Effects on Seed Weight in Subspecies
1522 of Big Sagebrush: Applications for Restoration." *Ecosphere* 6 (10).
1523 <https://doi.org/10.1890/ES15-00249.1>. [S1, QC]
- 1524 Robertson, Phillip A. 1976. "Photosynthetic and Respiratory Responses of Natural Populations of
1525 *Koeleria Cristata* Grown in Three Environmental Regimes." *Botanical Gazette* 137 (1):94–98.
1526 [S1]
- 1527 Robertson, Phillip A., and Richard T Ward. 1970. "Ecotypic Differentiation in *Koeleria Cristata* (L.)
1528 Pers. from Colorado and Related Area." *Ecology* 51 (6):1083–87. [S1, S2, QC]
- 1529 Rogler, George A. 1960. "Relation of Seed Dormancy of Indian Ricegrass (*Oryzopsis Hymenoides*
1530 (Roem. & Schult) Ricker.) to Age and Treatment 1." *Agronomy Journal* 52 (8):470–73. [S1]
- 1531 Rowe, Courtney L J, and Elizabeth A. Leger. 2011. "Competitive Seedlings and Inherited Traits: A

- 1532 Test of Rapid Evolution of *Elymus Multisetus* (Big Squirreltail) in Response to Cheatgrass
1533 Invasion.” *Evolutionary Applications* 4 (3):485–98. [https://doi.org/10.1111/j.1752-](https://doi.org/10.1111/j.1752-4571.2010.00162.x)
1534 [4571.2010.00162.x](https://doi.org/10.1111/j.1752-4571.2010.00162.x). [S1, S2]
- 1535 Rowe, Courtney L. J., and Elizabeth A. Leger. 2012. “Seed Source Affects Establishment of *Elymus*
1536 *Multisetus* in Postfire Revegetation in the Great Basin.” *Western North American Naturalist* 72
1537 (4):543–53. <https://doi.org/10.3398/064.072.0410>. [S1, S3]
- 1538 Rumbaugh, M.D., and B.M. Pendery. 1993. “Registration of ARS-2892 Munroe Globemallow
1539 Germplasm.” *Crop Science* 33 (5). [S1]
- 1540 Rumbaugh, M.D., B.M. Pendery, H.F. Mayland, and G.E. Shewmaker. 1993. “Registration of ARS-
1541 2936 Scarlet Globemallow Germplasm.” *Crop Science* 33:1106–8. [S1]
- 1542 Sanderson, S. C., H. C. Stutz, and E. D. McArthur. 1990. “Geographic Differentiation in *Atriplex*
1543 *Confertifolia*.” *American Journal of Botany*. [S1, S2]
- 1544 Schellenberg, M.P. 2003. “Germination Temperature Response of Two Ecotypes of Winterfat
1545 [*Kraschennikovia Lanata* (Pursh) Guldenstaedt].” *Canadian Journal of Plant Science* 83
1546 (1):65–68. [S1]
- 1547 Shaw, Nancy L., Marshall R. Haferkamp, and Emerenciana G. Hurd. 1994. “Germination and
1548 Seedling Establishment of Spiny Hopsage in Response to Planting Date and Seedbed
1549 Environment.” *Journal of Range Management* 47 (2):165–74. <https://doi.org/10.2307/4002827>.
1550 [S1, QC]
- 1551 Shock, Clinton C., Erik B. Feibert, A. Rivera, Lamont D. Saunders, Nancy Shaw, and Francis F.
1552 Kilkenny. 2016. “Irrigation Requirements for Seed Production of Five *Lomatium* Species in a
1553 Semiarid Environment.” *HortScience* 51 (10):1270–77. [S1]
- 1554 Slauson, William L., and Richard T Ward. 1986. “Ecogenetic Patterns of Four Shrub Species in
1555 Semi-Arid Communities of Northwest Colorado.” *The Southwestern Naturalist*, 319–29. [S1,
1556 S2]
- 1557 Smith, David Solance, Matthew K. Lau, Ryan Jacobs, Jenna A. Monroy, Stephen M. Shuster, and
1558 Thomas G. Whitham. 2015. “Rapid Plant Evolution in the Presence of an Introduced Species
1559 Alters Community Composition.” *Oecologia* 179 (2):563–72. [https://doi.org/10.1007/s00442-](https://doi.org/10.1007/s00442-015-3362-y)
1560 [015-3362-y](https://doi.org/10.1007/s00442-015-3362-y). [S1, S2]
- 1561 Smith, David Solance, Jennifer A. Schweitzer, Philip Turk, Joseph K. Bailey, Stephen C. Hart,
1562 Stephen M. Shuster, and Thomas G. Whitham. 2012. “Soil-Mediated Local Adaptation Alters
1563 Seedling Survival and Performance.” *Plant and Soil* 352 (1–2):243–51.
1564 <https://doi.org/10.1007/s11104-011-0992-7>. [S1]
- 1565 Springfield, H.W. 1968. “Germination of Winterfat Seeds under Different Moisture Stresses and
1566 Temperatures.” *Journal of Range Management*, 314–16. [S1]
- 1567 Springfield, H.W. 1966. “Germination of Fourwing Saltbush Seeds at Different Levels of Moisture
1568 Stress.” *Agronomy Journal* 58 (2):149–50. [S1]
- 1569 Springfield, H.W. 1964. “Some Factors Affecting Germination of Fourwing Saltbush.” Rocky
1570 Mountain Forest and Range Experiment Station, Forest Service, US Department of Agriculture

- 1571 25. [S1, S2]
- 1572 St. Clair, John Bradley, Francis F. Kilkenny, Richard C. Johnson, Nancy L. Shaw, and George
1573 Weaver. 2013. "Genetic Variation in Adaptive Traits and Seed Transfer Zones for
1574 *Pseudoroegneria Spicata* (Bluebunch Wheatgrass) in the Northwestern United States."
1575 *Evolutionary Applications* 6 (6):933–48. <https://doi.org/10.1111/eva.12077>. [S1, S2, QC]
- 1576 Staub, Jack E, Matthew D Robbins, Yingmei Ma, and Paul G Johnson. 2014. "Phenotypic and
1577 Genotypic Analysis of a US Native Fine-Leaved *Festuca* Population Reveals Its Potential Use
1578 for Low-Input Urban Landscapes." *Journal of the American Society for Horticultural Science*
1579 139 (6):706–15. [S1, QC]
- 1580 Stettler, Jason M., Douglas A. Johnson, B. Shaun Bushman, Kevin J. Connors, Thomas A. Jones,
1581 Jennifer W. Macadam, and David J. Hole. 2017. "Utah Lotus: North American Legume for
1582 Rangeland Revegetation in the Southern Great Basin and Colorado Plateau." *Rangeland*
1583 *Ecology and Management* 70 (6):691–99. <https://doi.org/10.1016/j.rama.2017.06.002>. [S1, S2]
- 1584 Stevens, Allan R, Val Jo Anderson, and Rachel Fugal. 2014. "Competition of Squirreltail with
1585 Cheatgrass at Three Nitrogen Levels *." *American Journal of Plant Sciences* 5:990–96. [S1,
1586 QC]
- 1587 Stinson, Kristina A. 2004. "Natural Selection Favors Rapid Reproductive Phenology in *Potentilla*
1588 *Pulcherrima* (Rosaceae) at Opposite Ends of a Subalpine Snowmelt Gradient." *American*
1589 *Journal of Botany* 91 (4):531–39. <https://doi.org/10.3732/ajb.91.4.531>. [S1, S2]
- 1590 Tilley, Derek J. 2015. "Notice of Release of Amethyst Germplasm Hoary Tansyaster: Selected Class
1591 of Natural Germplasm." *Native Plants Journal* 16 (1):54–60. [S1, QC]
- 1592 Tilley, Derek J. 2015. Douglas' Dustymaiden Initial Evaluation Planting Final Study Report. USDA
1593 NRCS Aberdeen Plant Materials Center.
1594 https://www.nrcs.usda.gov/Internet/FSE_PLANTMATERIALS/publications/idpmcsr13140.pdf
1595 f. [S1, S2, QC]
- 1596 Tisdale, E. W. 1961. "Intraspecific Variation in *Festuca*." *Carnegie Institution of Washington*
1597 *Yearbook* 60:388–91. [S1]
- 1598 Toole, Vivian K. 1941. "Factors Affecting the Germination of Various Dropseed Grasses
1599 (*Sporobolus* Spp.)." *Journal of Agricultural Research* 62:691–715. [S1, S2]
- 1600 USDA NRCS Bridger Plant Materials Center. 1996. Notice of Release of 'Rimrock' Indian
1601 Ricegrass. USDA NRCS Bridger Plant Materials Center, Montana Agricultural Experiment
1602 Station, Wyoming Agricultural Experiment Station, USDA Agricultural Research Station.
1603 [S1]
- 1604 USDA NRCS Los Lunas Plant Materials Center. 1973. Supporting Data For the Release of Arriba
1605 Western Wheatgrass. New Mexico State University's Los Lunas Agricultural Science Center,
1606 Colorado State University, New Mexico Department of Transportation, and the USDA Natural
1607 Resources Conservation Service Los Lunas Plant Materials Center. [S1, QC]
- 1608 Waldron, B L, K B Jensen, A J Palazzo, T J Cary, J G Robins, M D Peel, D G Ogle, and L St John.
1609 2011. "'Recovery', a New Western Wheatgrass Cultivar with Improved Seedling
1610 Establishment on Rangelands." *Journal of Plant Registrations* 5 (3):367–73.

- 1611 <https://doi.org/Doi10.3198/Jpr2010.09.0527crc>. [S1]
- 1612 Wan, Changgui, Ronald E. Sosebee, and Bobby L. McMichael. 1998. "Water Relations and Root
1613 Growth of Two Populations of *Gutierrezia Sarothrae*." *Environmental and Experimental*
1614 *Botany* 39 (1):11–20. [https://doi.org/10.1016/S0098-8472\(97\)00021-X](https://doi.org/10.1016/S0098-8472(97)00021-X). [S1]
- 1615 Wan, Changgui, Ronald E. Sosebee, and Bobby L. McMichael. 1995. "Water Acquisition and
1616 Rooting Characteristics in Northern and Southern Populations of *Gutierrezia Sarothrae*."
1617 *Environmental and Experimental Botany* 35 (1):1–7. [https://doi.org/10.1016/0098-
1618 8472\(94\)00038-7](https://doi.org/10.1016/0098-8472(94)00038-7). [S1, S2]
- 1619 Wan, Changgui, Ronald E. Sosebee, and Bobby L. McMichael. 1996. "Lateral Root Development
1620 and Hydraulic Conductance in Four Populations of *Gutierrezia Sarothrae*." *Environmental and*
1621 *Experimental Botany* 36 (2):157–65. [https://doi.org/10.1016/0098-8472\(96\)01008-8](https://doi.org/10.1016/0098-8472(96)01008-8). [S1]
- 1622 Ward, Richard T. 1969. "Ecotypic Variation in *Deschampsia Caespitosa* (L.) Beauv. from
1623 Colorado." *Ecology* 50 (3):519. <https://doi.org/10.2307/1933914>. [S1, S2, QC]
- 1624 Waser, Nickolas M., and Mary V. Price. 1985. "Reciprocal Transplant Experiments with
1625 *Delphinium Nelsonii* (Ranunculaceae): Evidence for Local Adaptation." *American Journal of*
1626 *Botany*, 1726–32. [S1, S2, S3]
- 1627 Wood, M. Karl, Robert W. Knight, and James A. Young. 1976. "Spiny Hopsage Germination."
1628 *Journal of Range Management*, 53–56. [S1]
- 1629 Workman, John P, and Neil E West. 1967. "Germination of *Eurotia Lanata* in Relation to
1630 Temperature and Salinity." *Ecology* 48 (4):659–61. [S1]
- 1631 Workman, John P, and Neil E West. 1969. "Ecotypic Variation of *Eurotia Lanata* Populations in
1632 Utah." *Botanical Gazette* 130 (1):26–35. <https://doi.org/10.2307/2473599>. [S1, S2]
- 1633 Young, J A, C D Clements, and T Jones. 2003. "Germination of Seeds of Big and Bottlebrush
1634 Squirreltail." *Journal of Range Management* 56 (3):277–81. <https://doi.org/10.2307/4003819>.
1635 [S1]
- 1636 Young, JA, RA Evans, and DE Palmquist. 1989. "Big Sagebrush (*Artemisia Tridentata*) Seed
1637 Production." *Weed Science* 37 (1):47–53. <http://www.jstor.org/stable/10.2307/4044754>. [S1,
1638 S3, QC]
- 1639 Young, James A., and Raymond A. Evans. 1989. "Reciprocal Common Garden Studies of the
1640 Germination of Seeds of Big Sagebrush (*Artemisia Tridentata*)." *Weed Science* 37 (3):319–
1641 325. <https://doi.org/none>. [S1, QC]
- 1642 Young, James A, Raymond A Evans, and B L Kay. 1984. "Persistence and Colonizing Ability of
1643 Rabbitbrush Collections in a Common Garden." *Journal of Range Management* 37 (4):373–77.
1644 <https://doi.org/10.2307/3898715>. [S1, QC]
- 1645 Zhang, Huarong, Laura E. DeWald, Thomas E. Kolb, and Dan E. Koepke. 2011. "Genetic Variation
1646 in Ecophysiological and Survival Responses to Drought in Two Grasses: *Koeleria Macrantha*
1647 and *Elymus Elymoides*." *Western North American Naturalist* 71 (1):25–32.
1648 <https://doi.org/10.3398/064.071.0104>. [S1, S2]
- 1649

1650 **Appendix 3. Additional Results**

1651 *Additional results of literature summary*

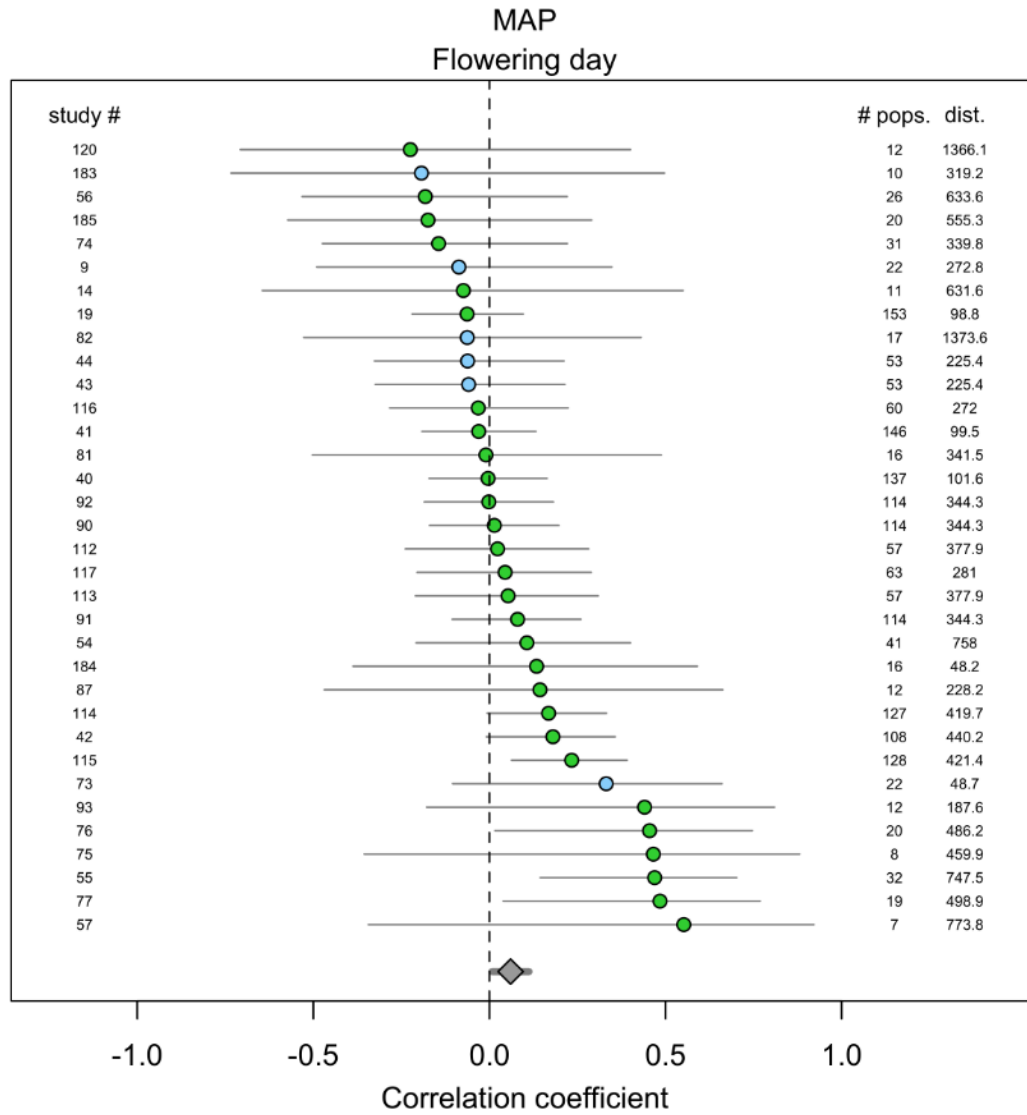
1652 Dicots accounted for 23.6% of the taxa and 42.8% of the experiments in the final pool of
1653 reviewed literature. Regional taxa accounted for 47.2% of the taxa and 48.9% of experiments,
1654 widespread taxa accounted for 26.0% of taxa and 36.7% of experiments, narrow taxa accounted
1655 for 24.4% of taxa and 13.5% of experiments, and endemic taxa accounted for 2.4% of taxa and
1656 0.9% of experiments. Perennials accounted for 46.3% of taxa and 32.1% of experiments, long-
1657 lived perennials accounted for 32.5% of taxa and 39.4% of experiments, short-lived perennials
1658 accounted for 14.6% of taxa and 25.1% of experiments, annuals accounted for 5.7% of taxa and
1659 3.1% of experiments, and biennials accounted for 0.8% of taxa and 0.3% of experiments.
1660 Primarily outcrossing plants accounted for 71.4% of taxa and 72.2% of experiments, primarily
1661 selfing plants accounted for 11.1% of taxa and 14.1% of experiments, and plants with mixed
1662 mating accounted for 17.5% of taxa and 13.8% of experiments.

1663

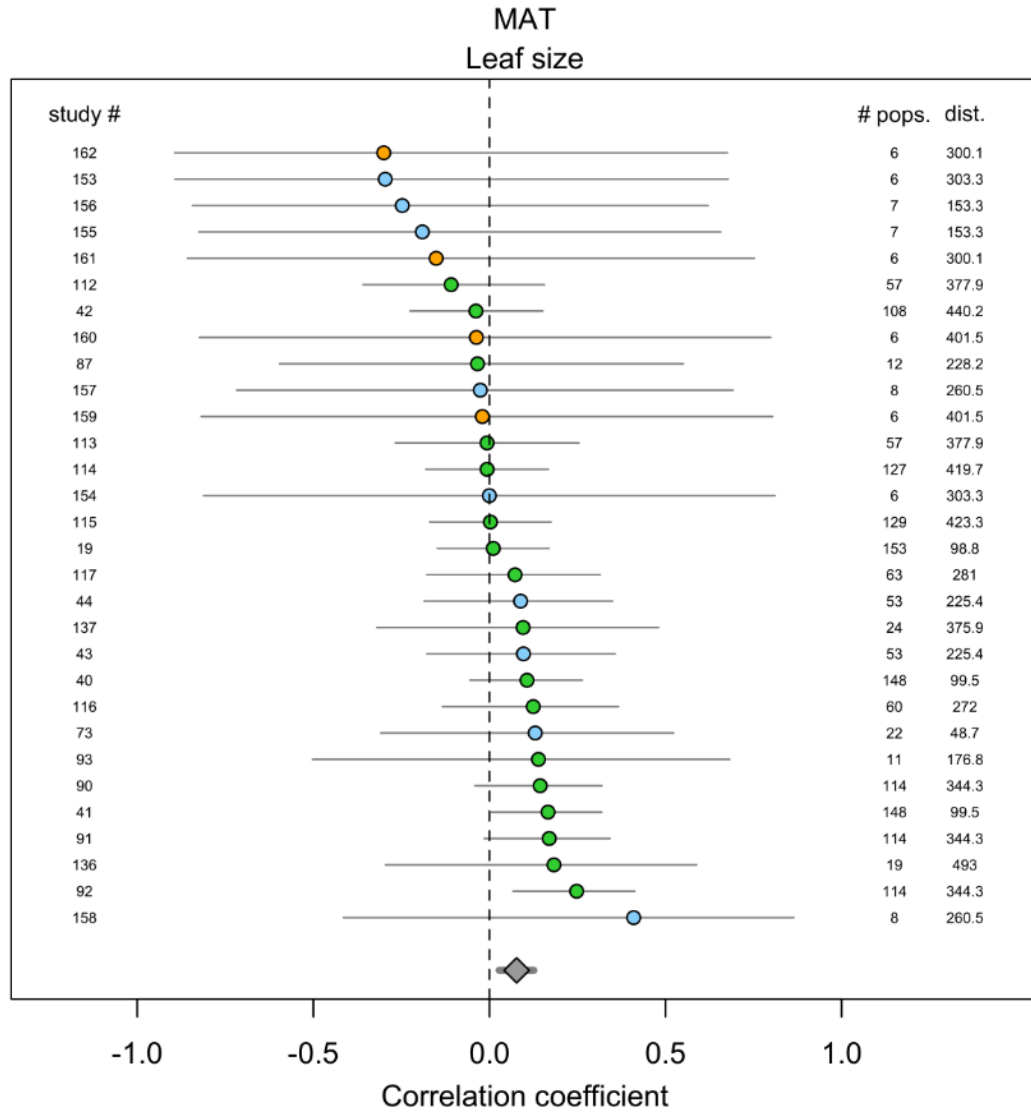
1664 *Additional results for quantitative comparison of trait-by-environment associations*

1665 Appendix 3 Figures 1-16. For each trait/environment correlation (16 combinations), the
1666 results of correlation coefficients (with 95% confidence intervals) for the pairwise comparisons,
1667 for each population in each experiment, between the difference in phenotypic trait and
1668 environmental characteristic at the collection location, while controlling for geographic distance
1669 among populations (see methods). MAT = Mean Annual Temperature, MAP = Mean Annual
1670 Precipitation. Study number identifies the particular experiment in the quantitative comparison
1671 dataset in the electronic supplementary material, “# pops.” is the number of populations included
1672 in each study, and “dist.” is the average pairwise distance between populations (in km). Grasses

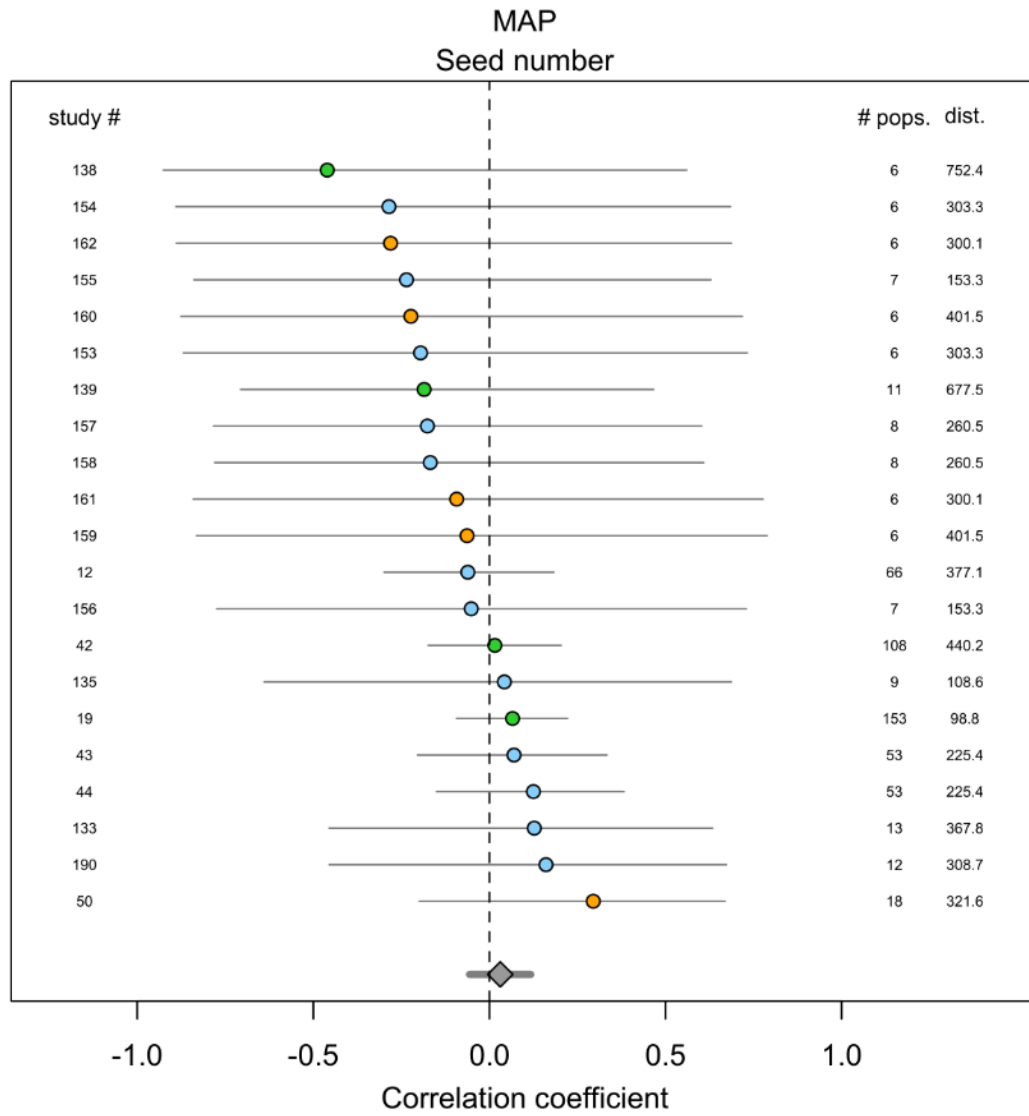
1673 are shown in green, shrubs in blue, and forbs in orange. The overall effect size and confidence
1674 intervals, across all studies, is shown in gray at the bottom of each figure. See main text, Table 1,
1675 for descriptions of phenotypic traits.



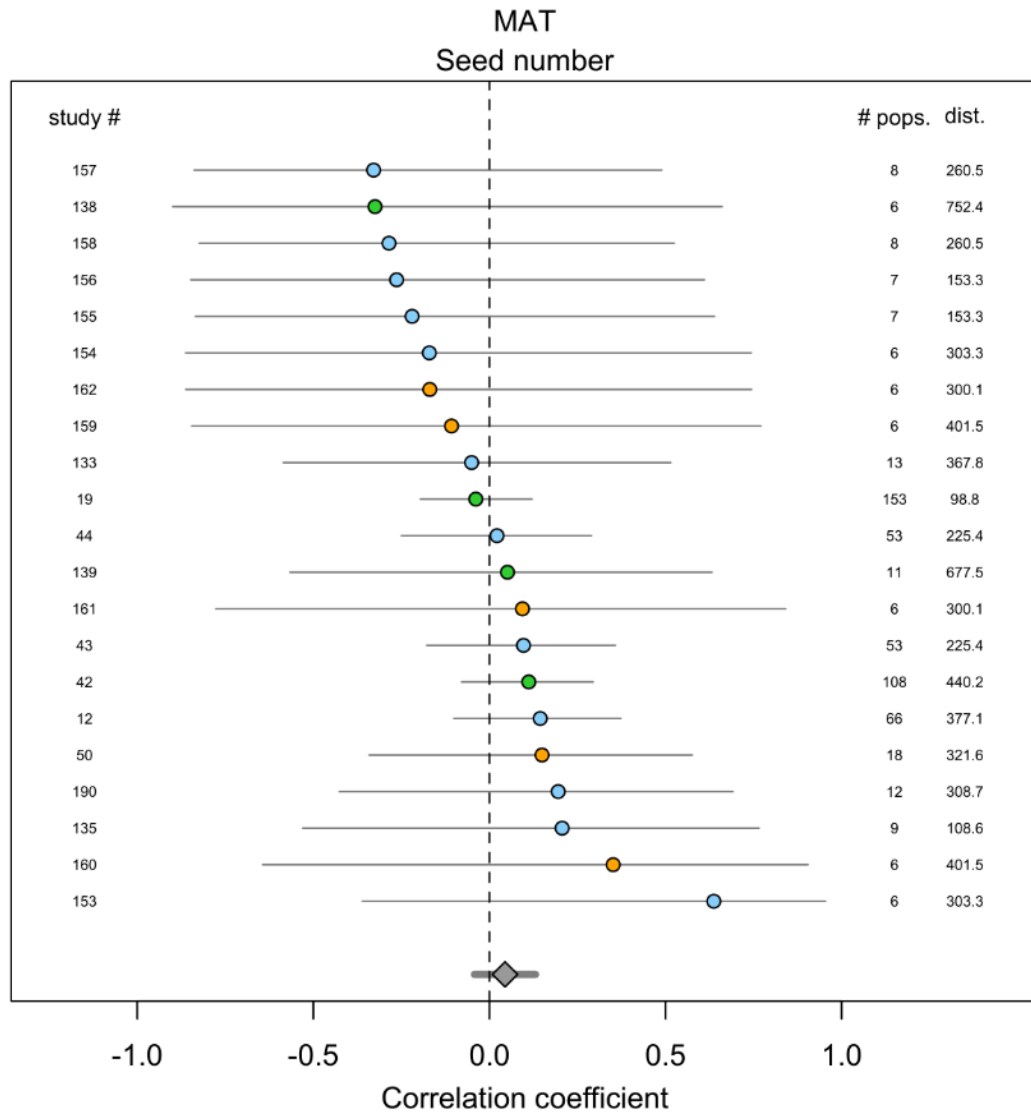
1676



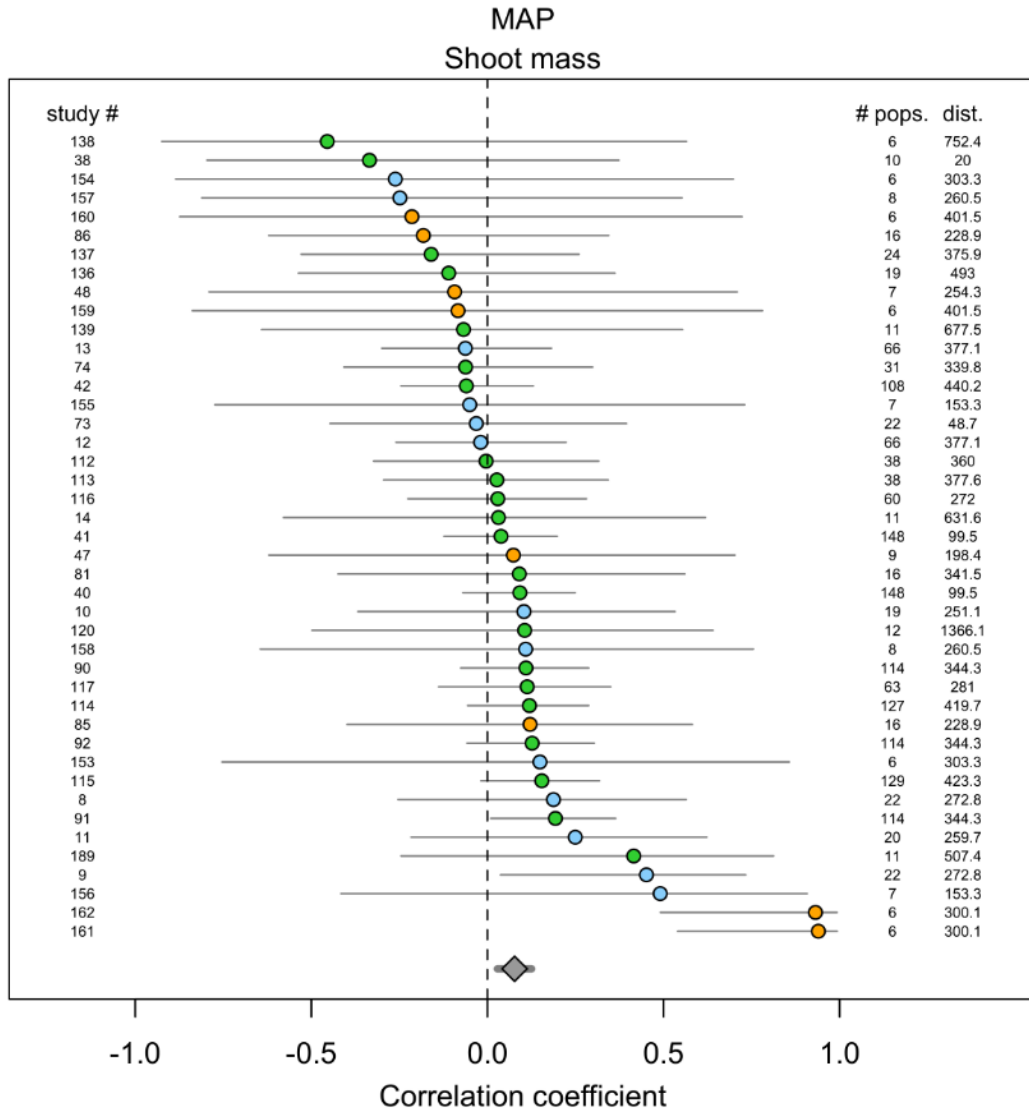
1677



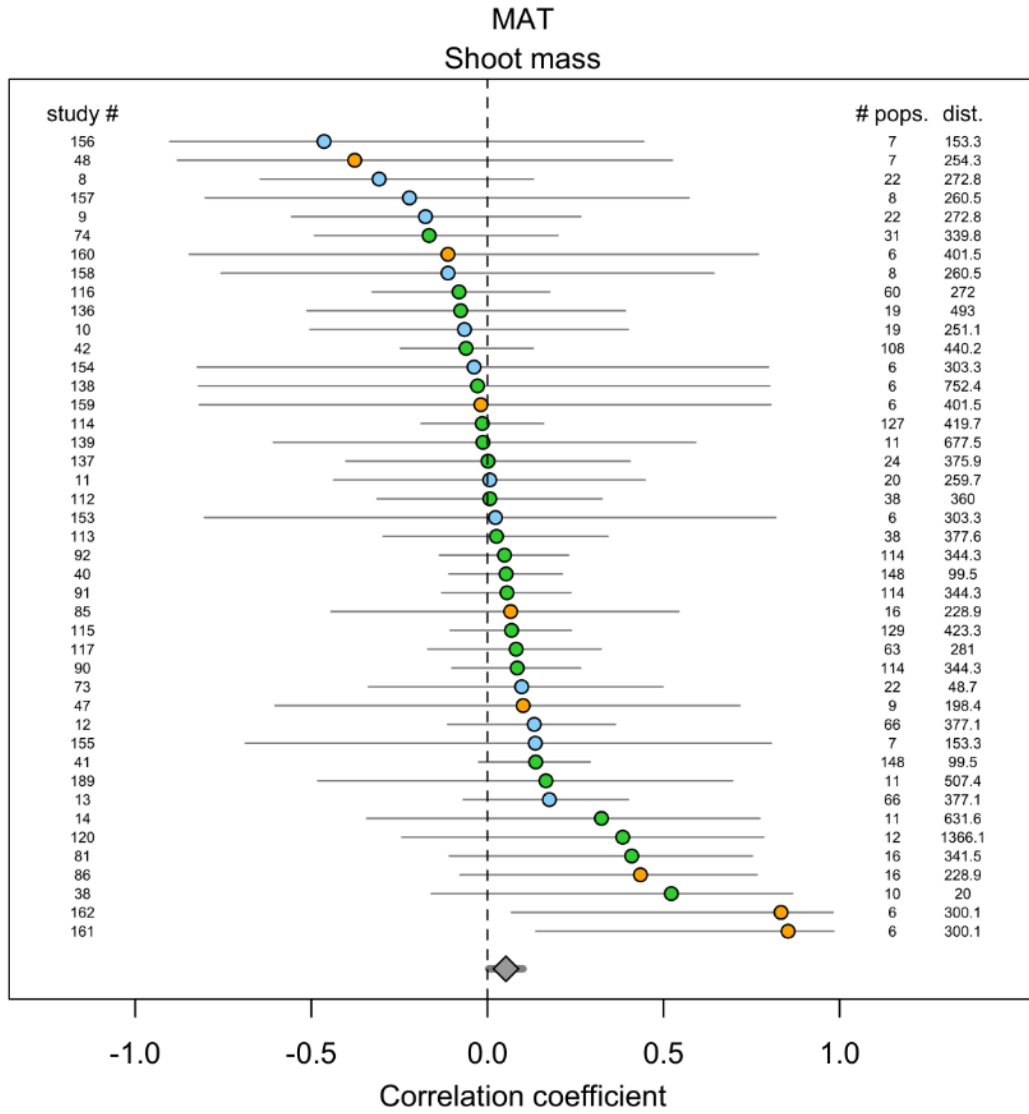
1678



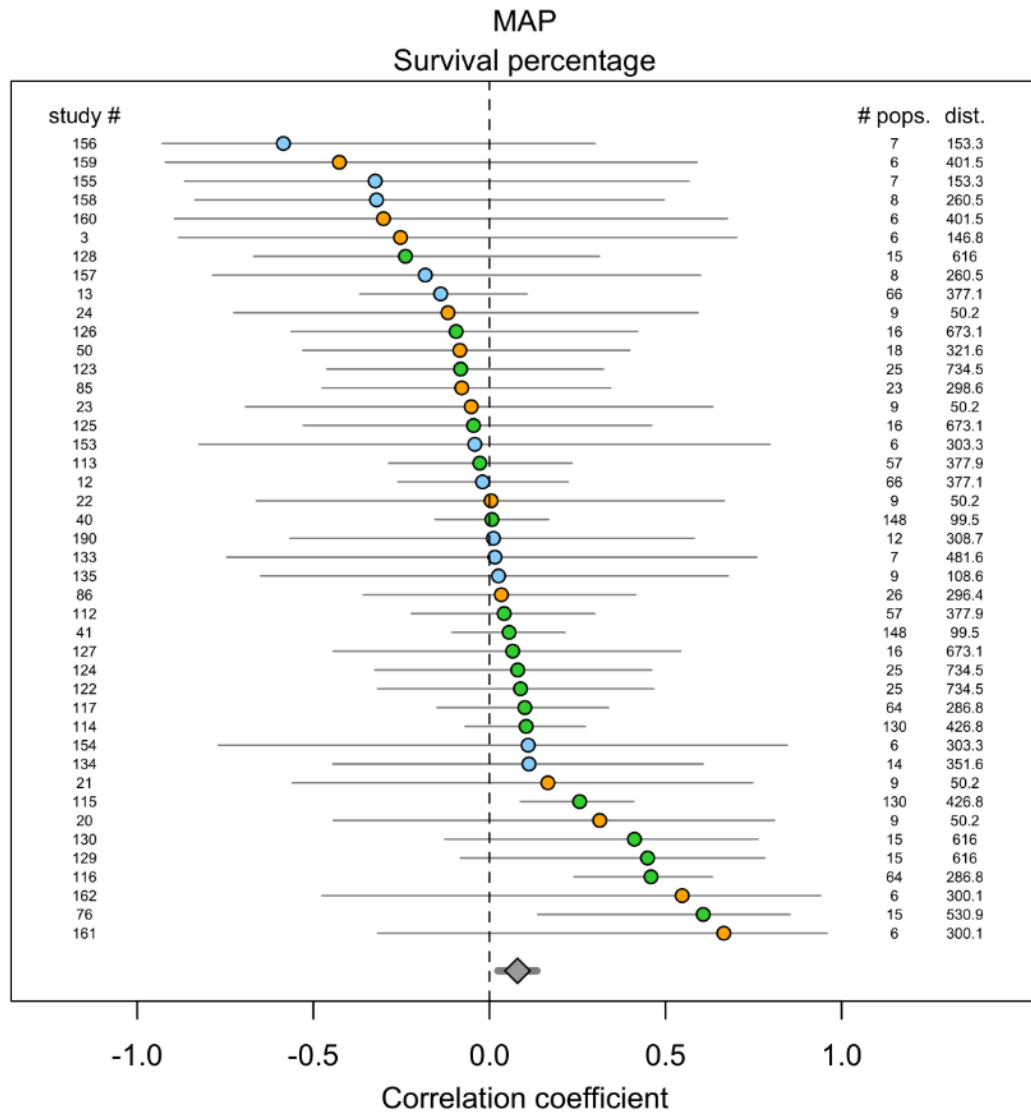
1679



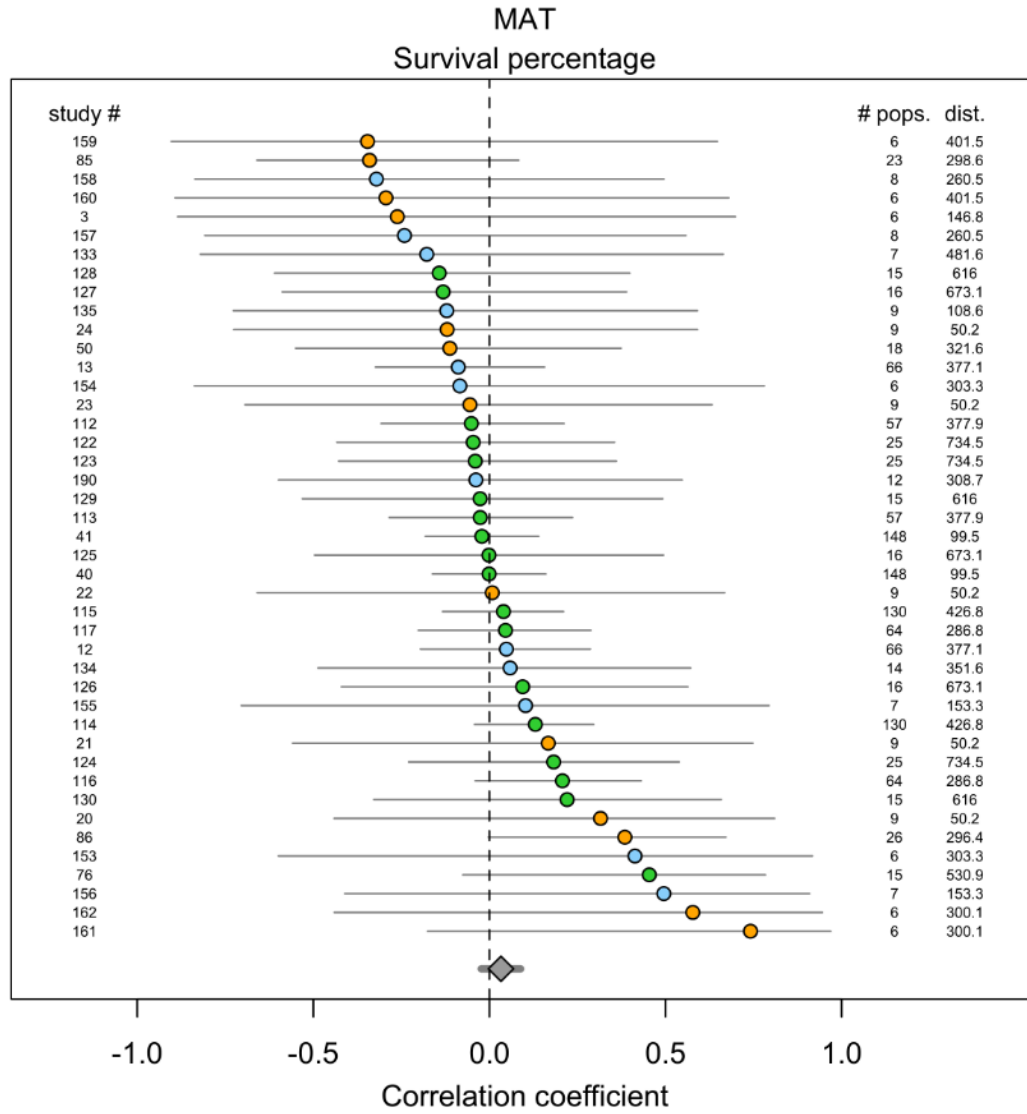
1680



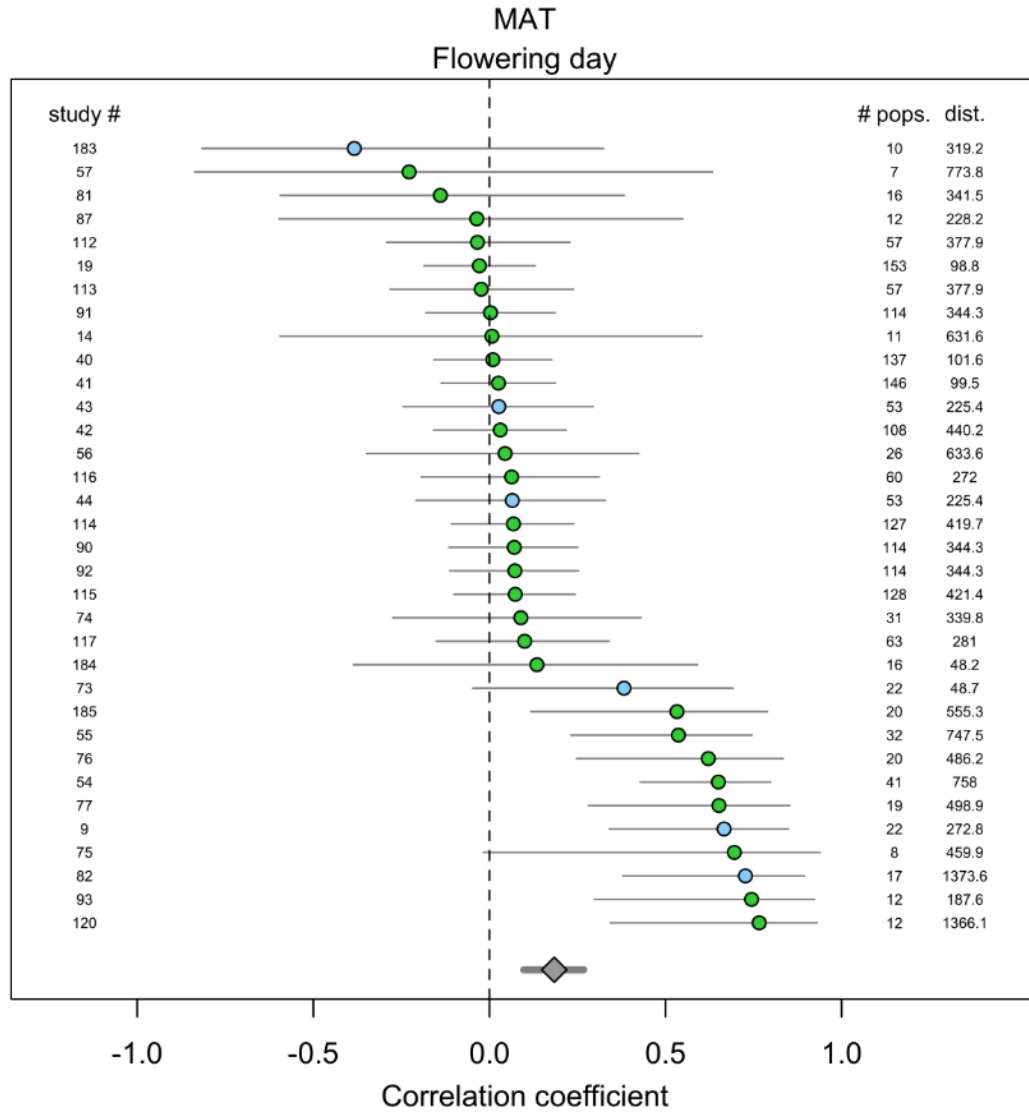
1681



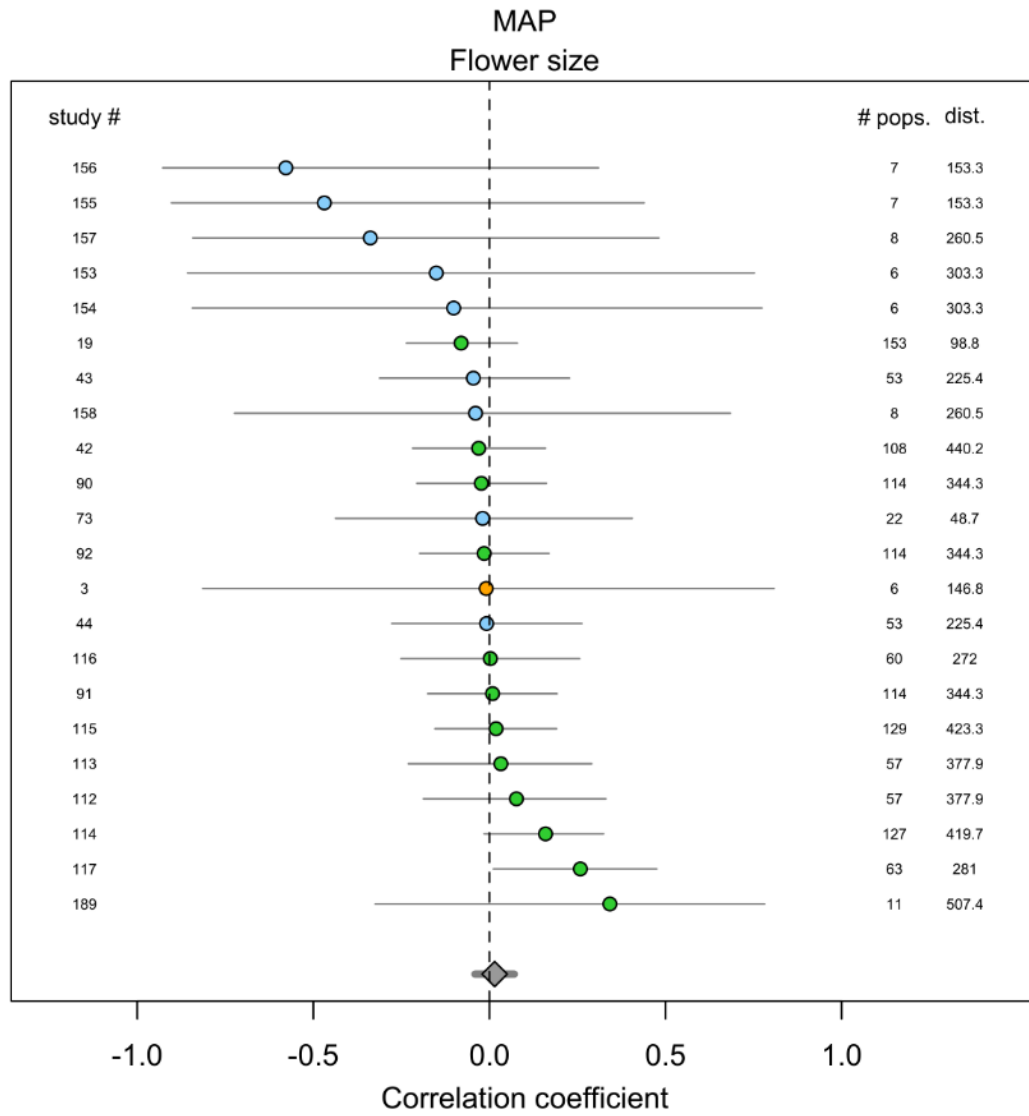
1682



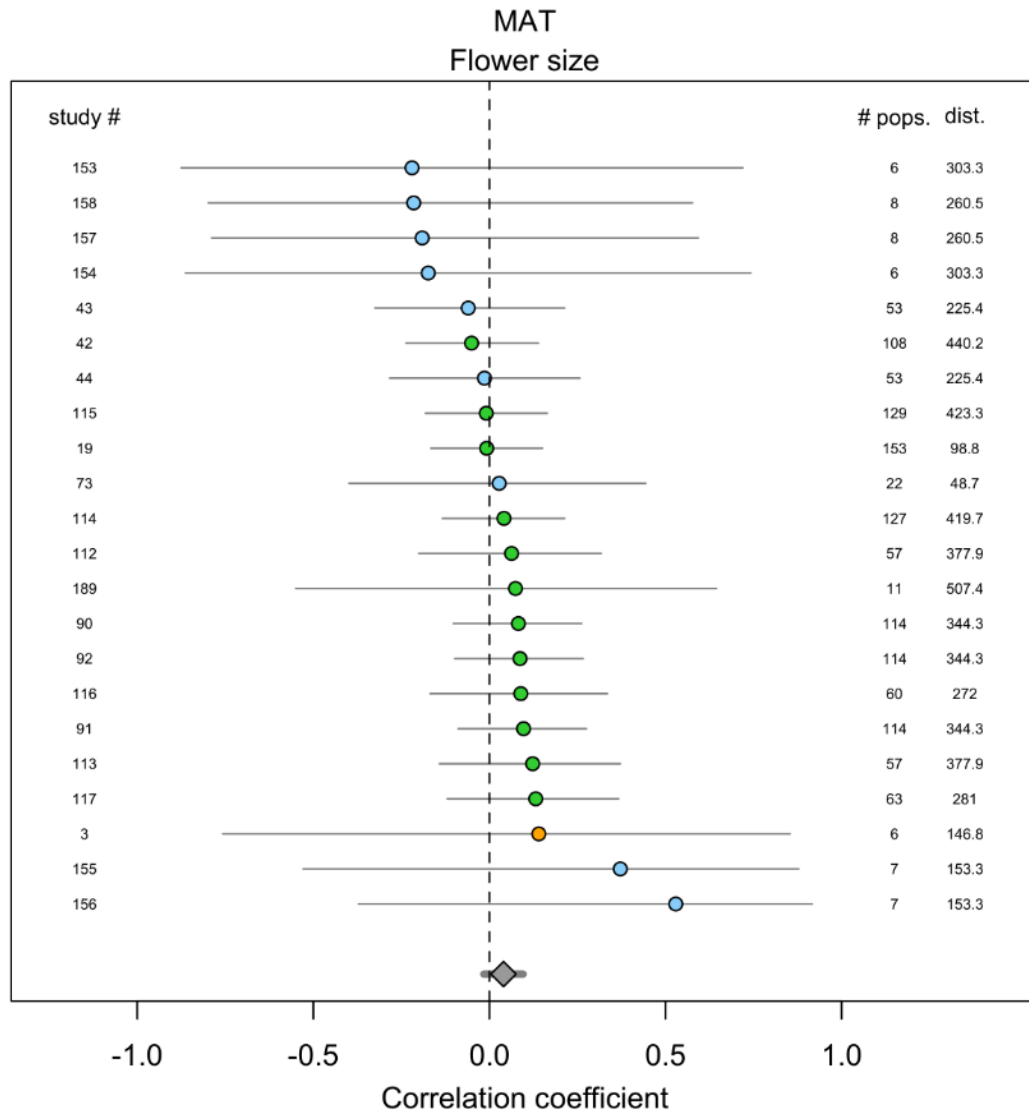
1683



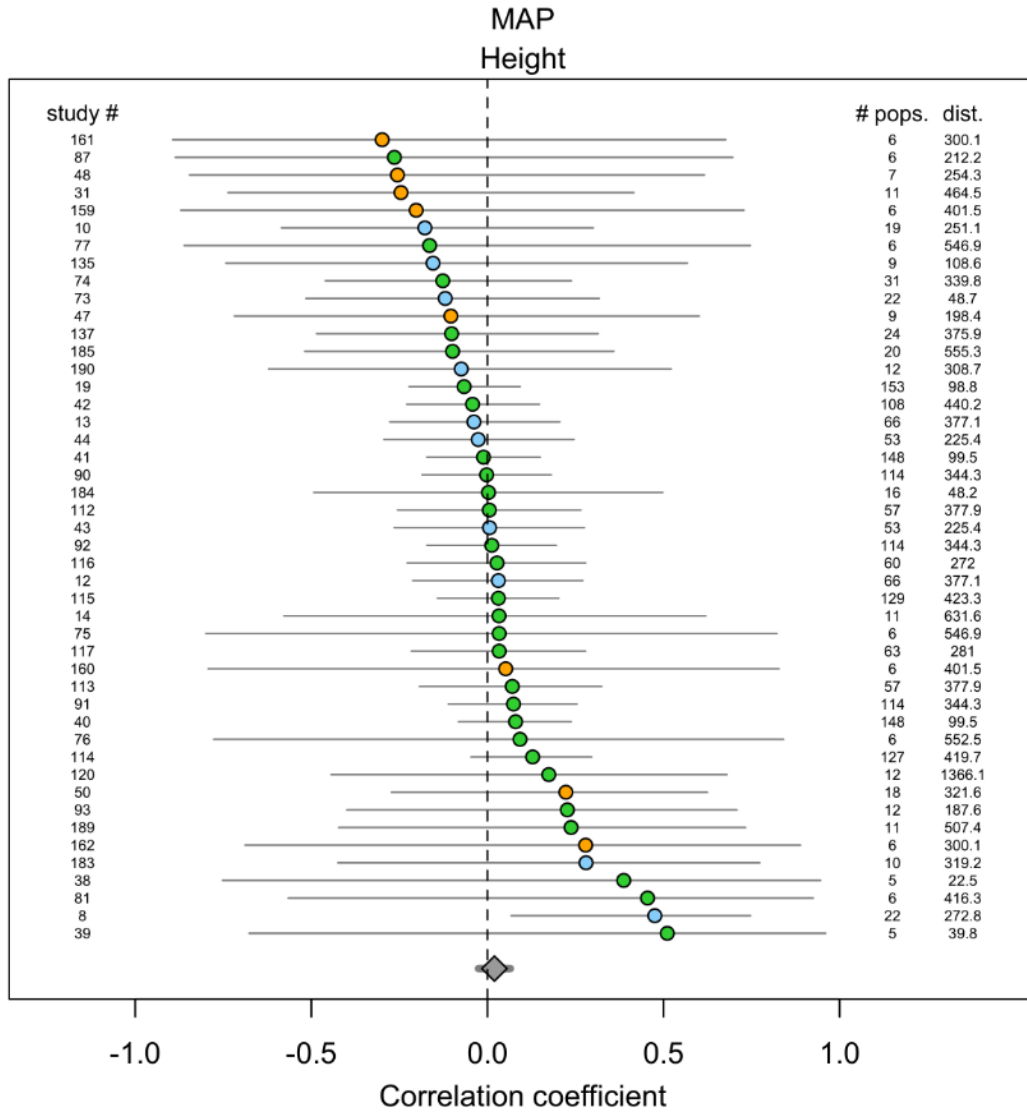
1684



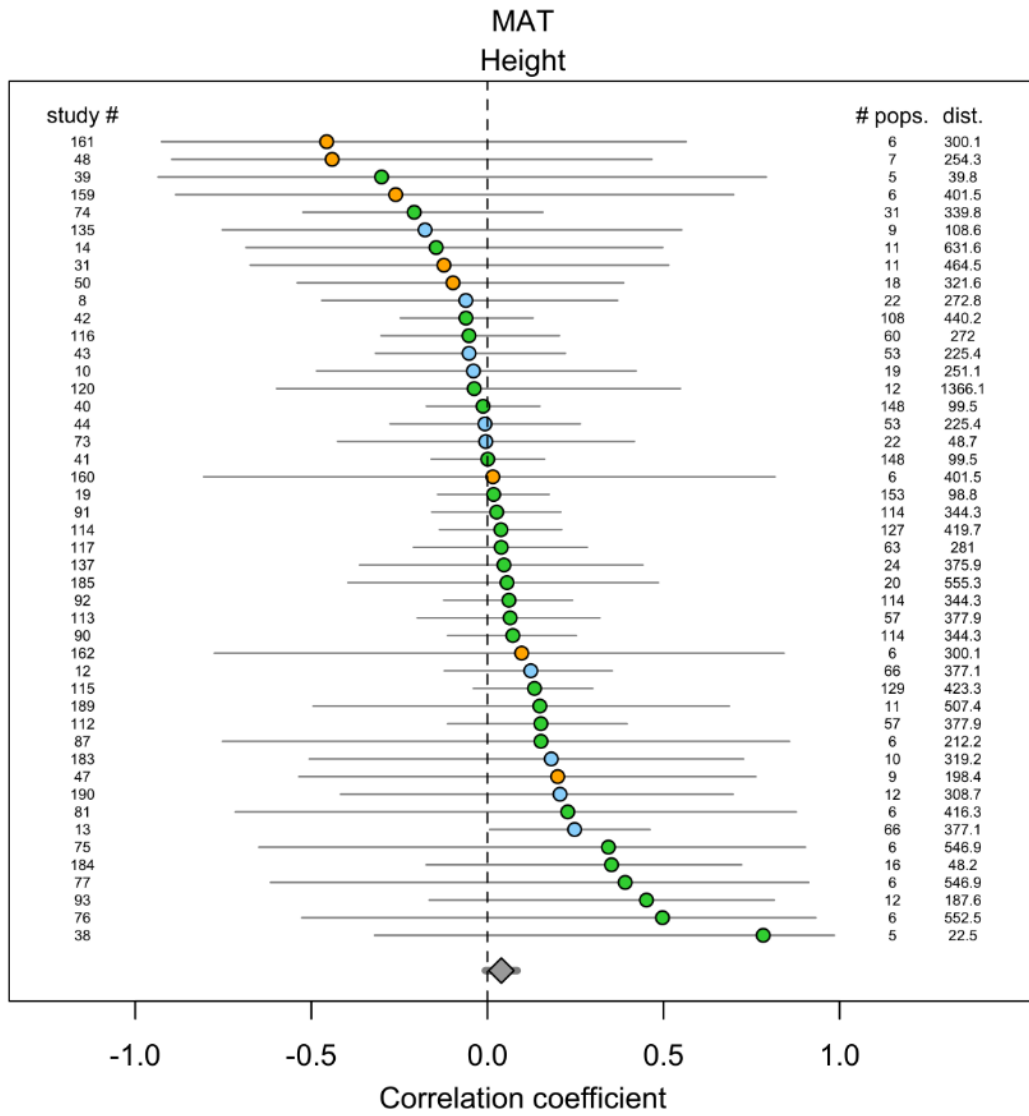
1685



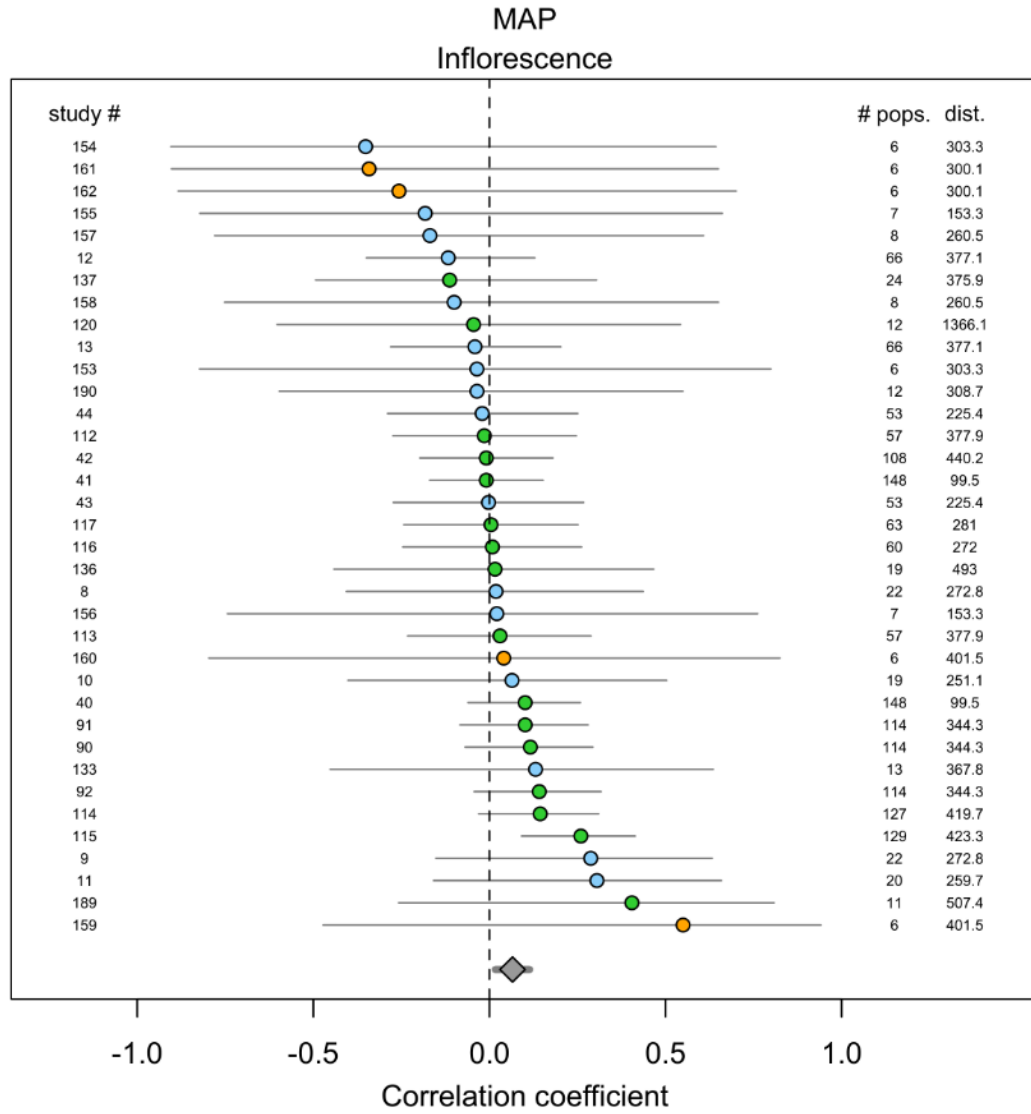
1686



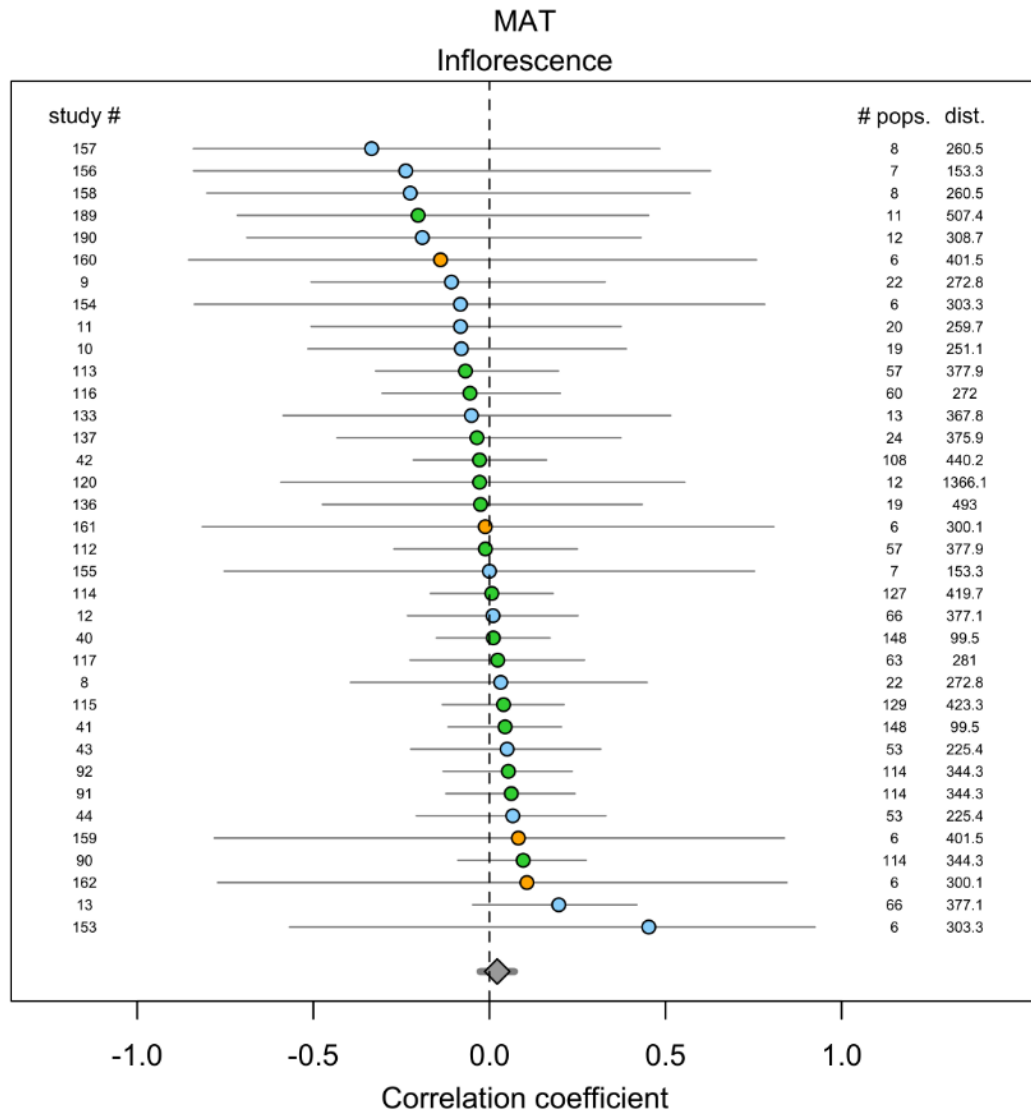
1687



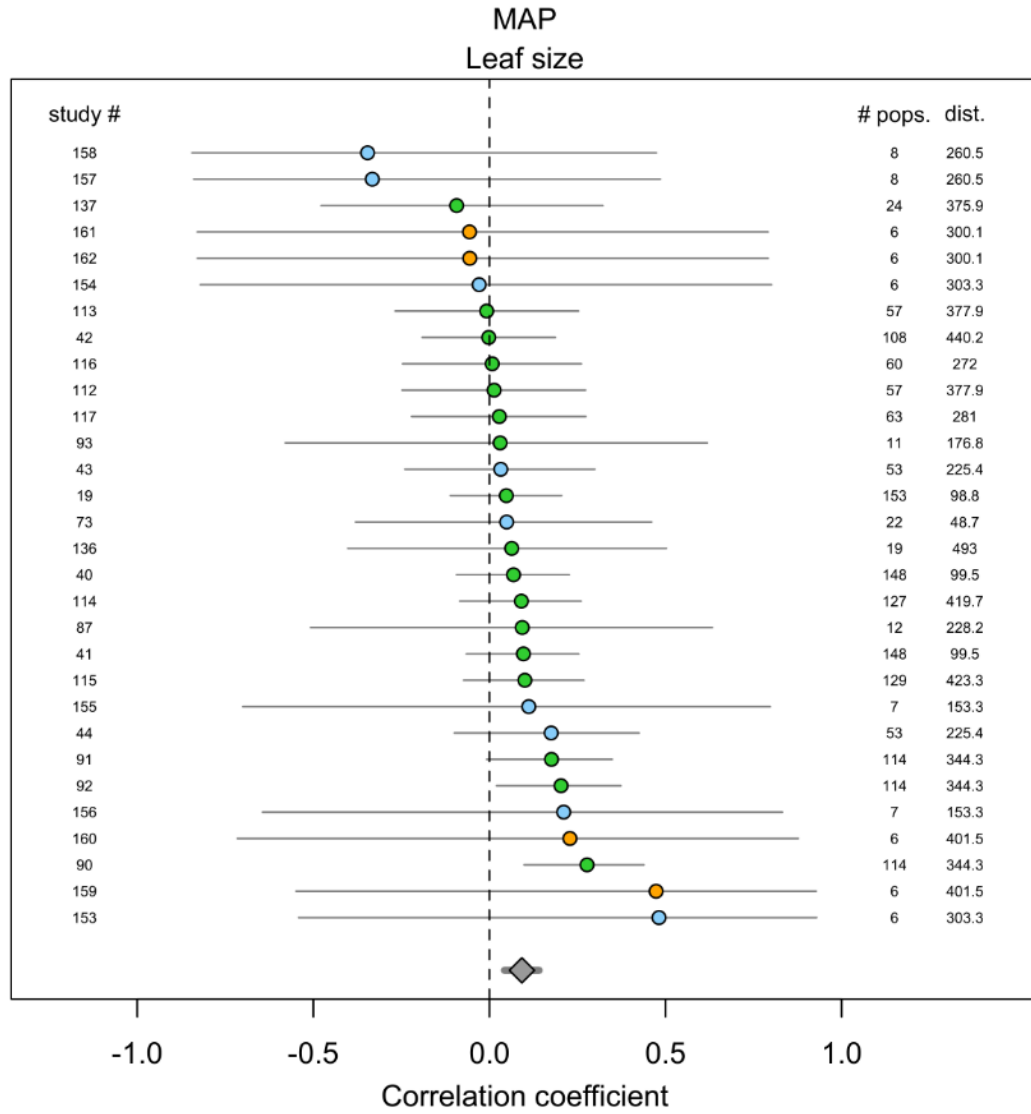
1688



1689



1690



1691