Title: Removal of a subset of non-essential genes fully attenuates a highly virulent *Mycoplasma* strain

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23 **ABSTRACT:**

24 Mycoplasmas are the smallest free-living organisms and cause a number of economically
25 important diseases affecting humans, animals, insects and plants. Here, we demonstrate that
26 highly virulent *Mycoplasma mycoides* subspecies *capri* (*Mmc*) can be fully attenuated *via*
27 targeted deletion of non-essential genes encoding, among others, potential virulence traits. Five
28 genomic regions, representing approximately ten percent of the original *Mmc* genome, were
29 successively deleted using *Saccharomyces cerevisiae* as an engineering platform. Specifically,
30 a total of 68 genes out of the 432 genes verified to be individually nonessential in the JCVI-
31 Syn3.0 minimal cell, were excised from the genome. *In vitro* characterization showed that this
32 mutant was similar to its parental strain in terms of its doubling time, even though ten percent
33 of the genome content were removed. A novel *in vivo* challenge model in goats revealed that
34 the wild-type parental strain caused marked necrotizing inflammation at the site of inoculation,
35 septicemia and all animals reaching endpoint criteria within seven days after experimental
36 infection. This is in contrast to the mutant strain, which caused no clinical signs nor
37 pathomorphological lesions. These results highlight, for the first time, the rational design,
38 construction and complete attenuation of a *Mycoplasma* strain via synthetic genomics tools.
39 Trait addition using the yeast-based genome engineering platform and subsequent *in vitro* or *in*
40 *vivo* trials employing the *Mycoplasma* chassis will allow us to dissect the role of individual
41 candidate *Mycoplasma* virulence factors and lead the way for the development of an attenuated
42 designer vaccine.

43 **IMPORTANCE**

44 Members of the *Mycoplasma mycoides* cluster cause important animal plaques in Africa and
45 Asia, which impact animal welfare, provision of food and the life of thousands of small-scale
46 farmers. We applied synthetic biology tools to *Mycoplasma mycoides* in order to design and
47 create a fully attenuated *Mycoplasma* strain that was subsequently confirmed *in vivo* using a
48 novel caprine infection model. This is the first time that a *Mycoplasma* mutant developed by
49 applying synthetic biology tools has been tested *in vivo* to pin point candidate virulence traits.
50 The mutant strain is similar to "apathogenic *E. coli* K12" strains that boosted the research on
51 host-pathogen interactions for the genus *Escherichia* and other bacterial genera.

52 **INTRODUCTION**

53 Bacteria belonging to the genus *Mycoplasma* are wall-less bacteria that cause massive
54 economic losses in the livestock sector (chickens, ruminants and pigs) and are responsible for
55 human pneumonia and sexually transmitted diseases (STDs). Currently, there is an absence of
56 commercial vaccines against infections with the human pathogens *Mycoplasma pneumoniae*
57 and *Mycoplasma genitalium* (1). In contrast, many livestock vaccines are commercialized,
58 which rely either on adjuvanted killed bacteria or on attenuated strains obtained after successive
59 rounds of sub-culturing or chemical mutagenesis (2). Due to these empirical approaches, the
60 exact mechanism triggering the attenuation is unknown for many of the previously developed
61 live attenuated *Mycoplasma* vaccines. Strikingly, these vaccines are far from being optimal
62 since they often display short durations of immunity and limited efficacy (3-5). A better
63 understanding of pathogenicity and the identification of virulence traits would foster next
64 generation vaccines.

65 For many years, the lack of genetic tools has limited our basic understanding of *Mycoplasma*
66 pathogenicity. Due to their regressive evolution by gene loss , mycoplasmas appear to lack
67 many of the common bacterial effectors and toxins used to interact with their hosts or to escape
68 the hosts' immune systems (6, 7). Lipoproteins have been proposed to be involved in both
69 aspects by using their cytoadherent properties and allowing antigenic variability through phase
70 or sequence variation (8). Other candidate virulence traits, such as the *Mycoplasma* Ig binding
71 protein-*Mycoplasma* Ig protease (MIB-MIP) system (9) and the hydrogen peroxide production
72 system (10) have been suggested, but not confirmed *in vivo*.

73 The availability of a genome engineering platform that allows directed and precise mutagenesis
74 for *Mycoplasma mycoides* is undoubtedly a new starting point towards better understanding of
75 host-pathogen interactions. In this work, we engineered a *Mmc* strain by deleting approximately
76 one tenth of the genome, including candidate virulence traits. The resulting mutant retains

77 almost wild-type like growth characteristics and was attenuated both *in vitro* and *in vivo*. The
78 construction of this fully attenuated and safe laboratory *Mycoplasma* strain paves the way for
79 research into host-pathogen interactions and is a good starting point to revisit the actual role of
80 suggested virulence determinants in *Mycoplasma*.

81 **RESULTS**

82 **Generation of the mutant strain GM12::YCpMmyc1.1-*Δ68***

83 To demonstrate attenuation of *Mmc* by rational design, five genomic regions were targeted in
84 this study. The precise localizations in the *Mmc* GM12 genome are shown in Figure 1. The first
85 two target deletion regions contained genes encoding the glycerol-dependent hydrogen
86 peroxide metabolic pathway and its suggested ABC transporter encoded by the *gtsABCD*
87 operon (11). This pathway has been suggested to be a main virulence mechanism for *M.*
88 *mycoides* (11), but *in vivo* confirmation is still missing and in *M. gallisepticum* the pathway
89 does not seem to be linked to virulence (12). Thus, the genes *glpF*, *glpK* and *glpO*
90 (MMCAP2_0217-0219; 2,984-bp region; D1) and the *gts* gene region that includes the gene
91 *lppB* (MMCAP2_0456-0459; 4,950-bp region; D2) were deleted in the *Mmc* genome by the
92 yeast-based engineering method (13). As previously mentioned, lipoproteins were another
93 target of interest since they likely trigger not only host-pathogen interactions but also,
94 overwhelming immune reactions that result in inflammation (14). The lipoproteins encoded in
95 the D3 (MMCAP2_0014-0016; 4,677-bp region; D3) as well as the D5 region (*lppQ,*
96 MMCAP2_0889-0904; 24,906-bp region, D5) were also excised employing again the yeast-
97 based engineering method. Finally, we deleted a large genomic region that contained the
98 *Mycoplasma*-specific F1-likeX0 ATPase (15), the MIB-MIP system (9) that has been shown *in*
99 *vitro* to modulate the action of immunoglobulins through specific degradation, an integrative
100 and conjugative element (ICE) (16) and several genes encoding other lipoproteins. The ICE
101 was targeted in an effort to reduce mobile elements from the *Mmc* genome. In this case, about
102 70 kbp (MMCAP2_0550-0591; 69,220-bp region; D4) were targeted and deleted from the *Mmc*
103 genome using the yeast-based engineering method in one stretch.

104

Fig. 1

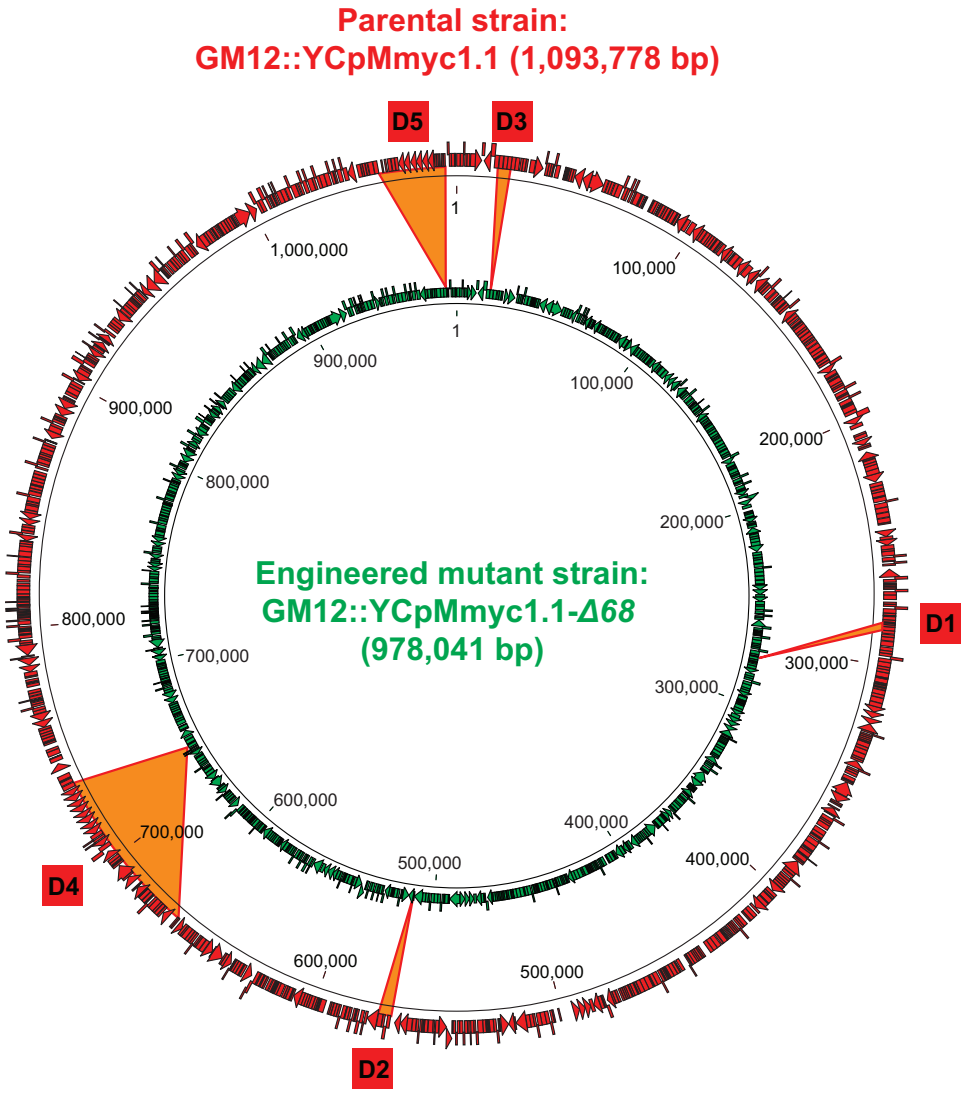

Figure 1: Design of deletion mutant *Mmc* strain. (A) Schematic displaying the genomes of the isogenic parental strain GM12::YCpMmyc1.1 and its derivative, the deletion mutant GM12::YCpMmyc1.1-*Δ68*.

105

106 After each cycle of deletions, the modified *Mmc* genome was isolated from yeast cells and
107 transplanted back into *M. capricolum* subsp. *capricolum* (*Mcc*) recipient cells to confirm the
108 viability of each mutant *Mmc* strain. Overall, the final mutant strain, named *Mmc*
109 GM12::YCpMmyc1.1-*Δ68,* was generated in five sequential deletion cycles (Figure 1). The
110 gene knock-outs were verified by amplifying across each deleted region (Figure S1). Genomic
111 DNA from the GM12::YCpMmyc1.1-*Δ68* was isolated and analyzed by sequencing to confirm

112 the deletions (Figure1B). The genome sequence of GM12::YCpMmyc1.1-*Δ68* was deposited
113 at the ENA database under the accession number LS483503.

114 **The mutant strain GM12::YCpMmyc1.1-*Δ68* is viable and unaffected in its morphology**
115 **or growth in axenic medium**

116 The colonies of *Mmc* GM12::YCpMmyc1.1-*Δ68* were of similar size to those of
117 GM12::YCpMmyc1.1 and GM12. Cell morphology of the GM12, the isogenic parental strain
118 GM12::YCpMmyc1.1 and GM12::YCpMmyc1.1-*Δ68* strains was evaluated using scanning
119 electron microscopy (Figure 2A). All strains tested were globular in shape and lacked any
120 special morphological features. The diameter of the microorganisms was in the range of 500
121 nm, as expected for a *Mycoplasma* cell. The mutant GM12::YCpMmyc1.1-*Δ68* grew with a
122 doubling time somewhat similar to that of the parental strains GM12 and GM12::YCpMmyc1.1
123 (Figure 2B). Together, these results strongly suggest that the deletion of approximately 100 kbp
124 of genomic content from the *Mmc* genome did not adversely affect structural integrity or *in*
125 *vitro* growth of the mutant GM12::YCpMmyc1.1-*Δ68*.

126 **Inability of the mutant strain GM12::YCpMmyc1.1-*Δ68* to produce hydrogen peroxide**
127 **in the presence of glycerol *in vitro***

128 This pathway was completely deleted in the construction of the mutant strain
129 GM12::YCpMmyc1.1-*Δ68*. Therefore to phenotypically confirm the deletion, we measured and
130 compared hydrogen peroxide production levels between the control GM12,
131 GM12::YCpMmyc1.1 and GM12::YCpMmyc1.1-*Δ68 in vitro*. In the presence of the glycerol
132 substrate, the mutant GM12::YCpMmyc1.1-*Δ68* shows a significant decrease in hydrogen
133 peroxide production when compared to its parental strains (Figure 2C). Indeed, while GM12
134 and GM12::YCpMmyc1.1 produced >0.3μM of H_2O_2, the mutant strain produced very low
135 amounts of H_2O_2 (0.01μM), at least 30-fold lower under these conditions.

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Fig. 2

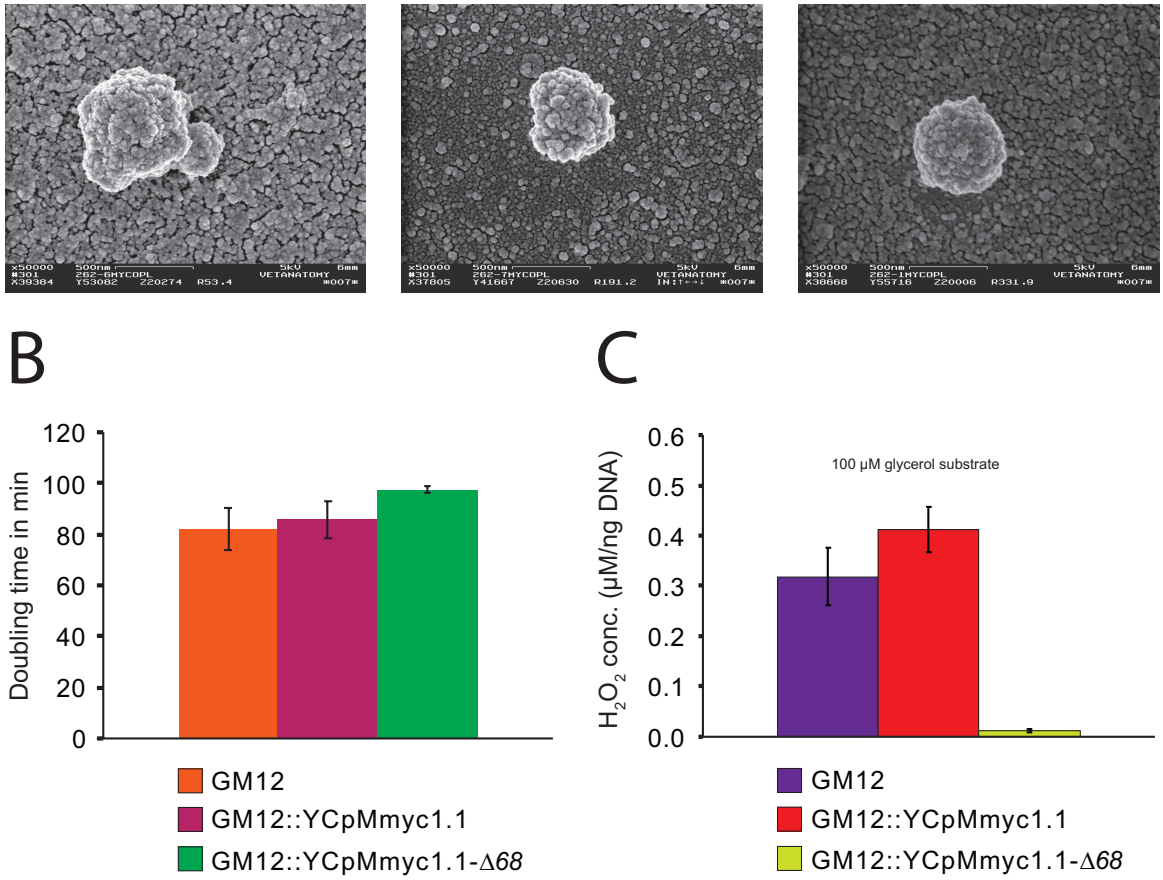

Figure 2: *In vitro* characteristics of the parental strains GM12, GM12::YCpMmyc1.1 and its deletion mutant GM12::YCpMmyc1.1-Δ*68* with respect to their morphology revealed by scanning electron microscopy (A), their doubling time (B), their production of hydrogen peroxide in the presence of glycerol (C), the bars display the standard deviation in B and C

138

139 **Inability of the mutant strain GM12::YCpMmyc1.1-*Δ68* to degrade immunoglobulin *in***

140 ***vitro***

141 Another potential virulence trait encoded by mycoplasmas is the MIB-MIP system, which may

142 play a role in immune evasion by cleavage of immunoglobulins (Figure 3A) (9). Incubation of

143 caprine IgG with GM12, GM12::YCpMmyc1.1 and GM12::YCpMmyc1.1-*Δ68* showed a clear

144 difference in the strains' abilities to degrade IgG (Figure 3B). The two bands at 25 kDa and

145 50kDa corresponds to the IgG light and heavy chains. The mutant strain GM12::YCpMmyc1.1-

146 *Δ68* exhibited no degradation of IgG, as noted by the lack of the 44 kDa band (Figure 3B, black
147 asterisk). This band, clearly visible in the other two strains, is indicative of proteolytic cleavage
148 of the IgG heavy chain. Another pattern of degradation, with a band at a size of about 30 kDa,
149 is visible in the three strains. It was previously reported that this IgG cleavage is not specific or
150 directly linked to the MIB-MIP system (9).

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Fig. 3

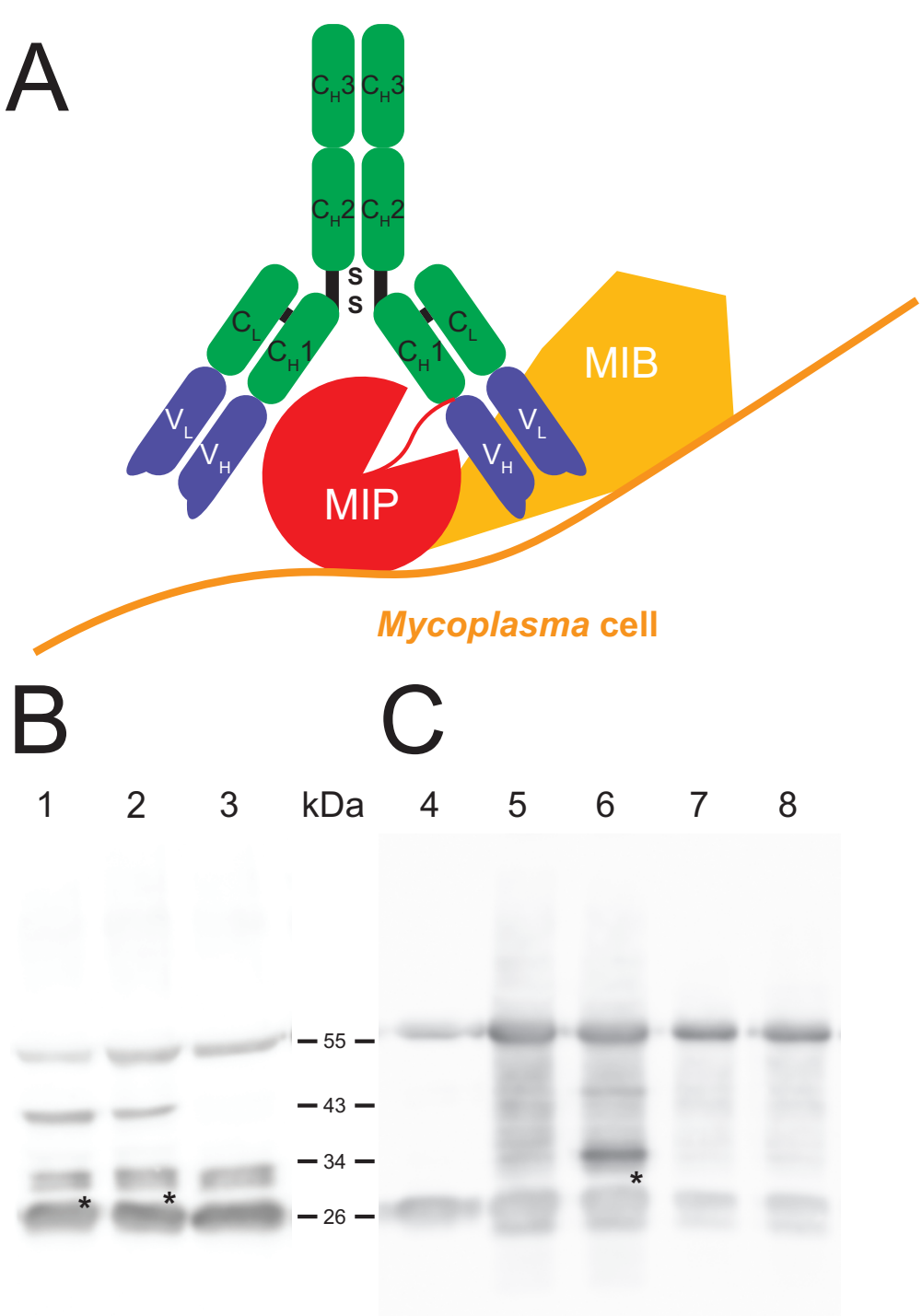

Figure 3: *In vitro* and *in vivo* ability of the *Mmc* MIB-MIP system to degrade caprine immunoglobulin: (A) cartoon displaying the suggested action of the MIB-MIP system, (B) *in vitro* ability of the parental strains GM12, GM12::YCpMmyc1.1 and their deletion mutant GM12::YCpMmyc1.1-Δ*68* to degrade caprine immunoglobulin, 1-GM12 + 5 µg caprine IgG, 2-GM12::YCpMmyc1.1 + 5 µg caprine IgG, 3-GM12::YCpMmyc1.1-Δ*68* + 5 µg caprine IgG; the black asterisk marks the 44 kDa cleaved fragment (C) *In vivo* detection of degradation of the *Mmc* MIB-MIP system using serum samples from the animal CK51 that succumbed from disease and had septicemia, 4-5 µg caprine IgG, 5-serum from goat CK51: -1 dpi (GM12 group), 6-serum from goat CK51: 4 dpi (GM12 group), 7-serum from goat CK45: -1 dpi (GM12::YCpMmyc1.1-Δ*68* group), 8-serum from goat CK45: 4 dpi (GM12::YCpMmyc1.1-Δ*68* group)

153 **The mutant strain GM12::YCpMmyc1.1-*Δ68* is fully attenuated *in vivo.***

154 We next tested whether GM12::YCpMmyc1.1-*Δ68* was able to cause disease in its native host.
155 Sixteen male outbred goats (*Capra aegagrus hircus*) were used in this animal infection trial.
156 The animals were separated into two groups of equal numbers. After the infection, no
157 immediate clinical signs of disease were observed. Two animals in the GM12::YCpMmyc1.1-
158 *Δ68* group had to be removed from the experiment, because of acquired wounds unrelated to
159 the infectious agent. Animal euthanasia was planned 28 days post infection (dpi). However, all
160 eight animals inoculated with the GM12 strain developed severe clinical signs, with pyrexia
161 starting 2-3 dpi (Figure 4B). Their body temperature continued to increase, up to 41-41.5°C,
162 during the following days. The animals stopped feeding, were apathic and showed signs of pain.
163 According to the endpoint criteria stated in the animal experiment protocol, they had to be
164 euthanized between 5-6 dpi (Figure 4A, red line). In sharp contrast, animals inoculated with the
165 GM12::YCpMmyc1.1-*Δ68* mutant strain did not develop any clinical signs of disease and were
166 all monitored until the end of the trial (Figure 4A). Their body temperatures fluctuated between
167 38°C and 39°C, with a few isolated cases where animals showed temperatures above 39°C but,
168 never for more than 2 consecutive days (Figure 4B). The animals appeared to remain healthy
169 and gained weight during the experiment (Figure 4C). Their heartbeat and respiratory rates,
170 between 80-110 beats/min and 20-30 breaths/min, respectively, remained constant over the
171 course of experimentation.
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Fig. 4

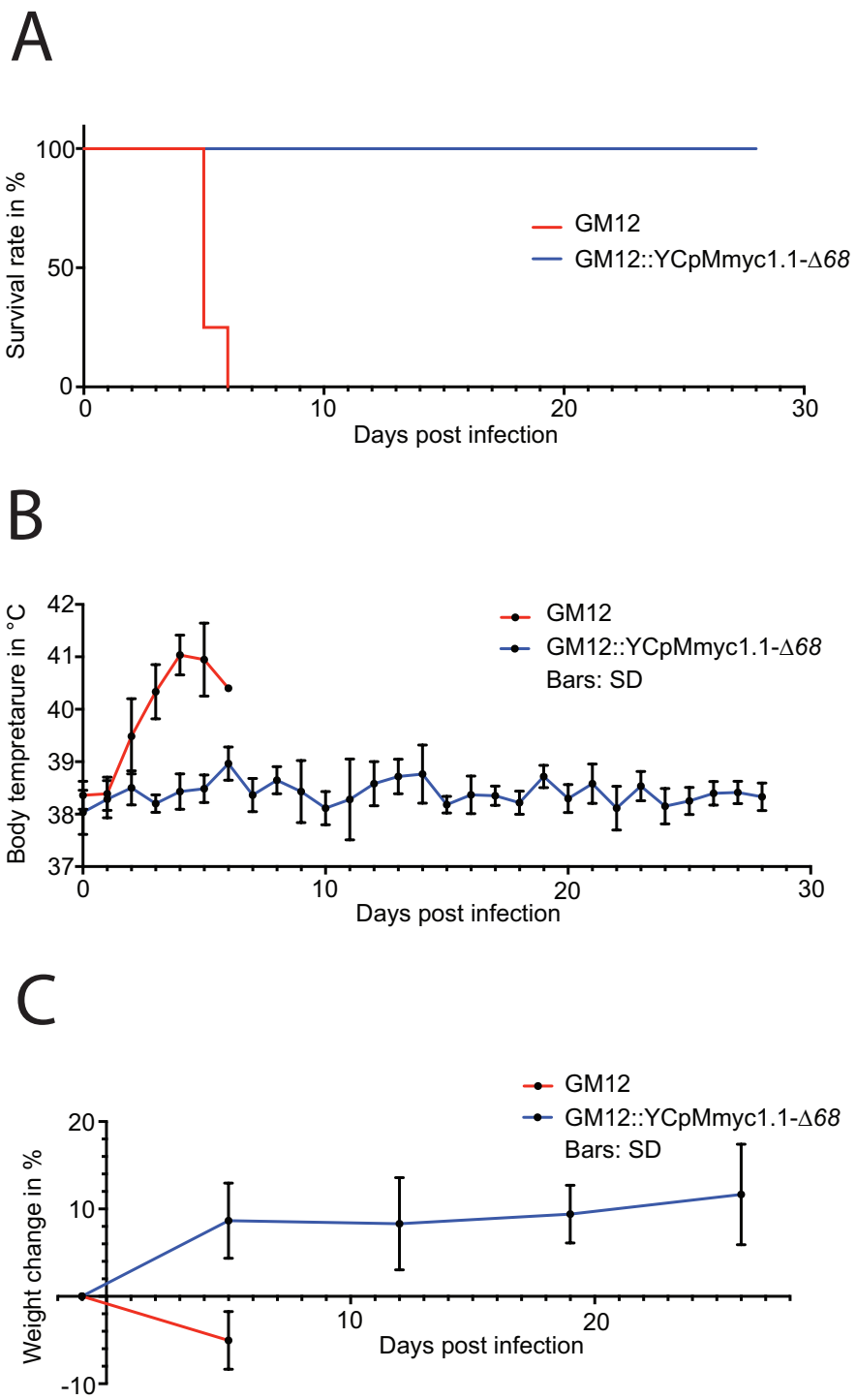

Figure 4: Comparison of the clinical parameters monitored during the in vivo challenge trial between the animals that received the GM12 and its derivative GM12::YCpMmyc1.1-Δ*68*. (A) Kaplan Meier survival curve based on animals reaching endpoint criteria, (B) Average body temperatures during experimental infection. Values were generated using daily rectal temperatures from the two groups. (C) Average weight gain during experimental infection. Values were generated using interval measures from the two groups. The standard deviations are displayed as bars in B and C.

179

180 In all animals infected with the GM12 strain, the main pathological lesions were similar, with
181 a severe and extensive inflammation of the soft tissues of the neck around the site of inoculation.
182 Additional macroscopic findings were severe pulmonary edema and congestion. Histologically,
183 there was extensive coagulation necrosis of the connective tissue (Figure 5B, thick arrows) and
184 musculature surrounding the trachea, in the vicinity of the inoculation site (Figure 5B and D,
185 diamonds). A marked infiltration of mainly degenerate neutrophilic granulocytes (Figure 5B
186 and D, asterisks) was always found associated with the necrosis. The necrotizing process
187 extended to the trachea, the subcutis and skin. In addition, all animals showed multifocal acute
188 necrosis with infiltration of neutrophilic granulocytes in liver and kidney and for three animals,
189 in the lung. These lesions were indicative of an acute septicemia. Among all animals infected
190 with the mutant strain GM12::YCpMmyc1.1-*Δ68*, and upon euthanasia 28-29 dpi, no
191 pathological lesions were found around the inoculation site. The soft tissue around the
192 inoculation site was within normal limits. Additionally, neither inflammation nor necrosis
193 associated with the epithelium, the submucosa or any cartilage tissues was histologically
194 observed (Figure 5, panels A and C).
195 *Mmc* GM12 was re-isolated from the blood of all animals experimentally infected with the
196 wild-type parental strain. Bacteremia was characterized by 10^6 up to 10^9 CCU.ml^{-1} of blood, as
197 measured by serial dilutions (Table 1). Blood samples collected from goats infected with
198 GM12::YCpMmyc1.1-Δ*68* did not reveal any bacteremia.

199

Fig. 5

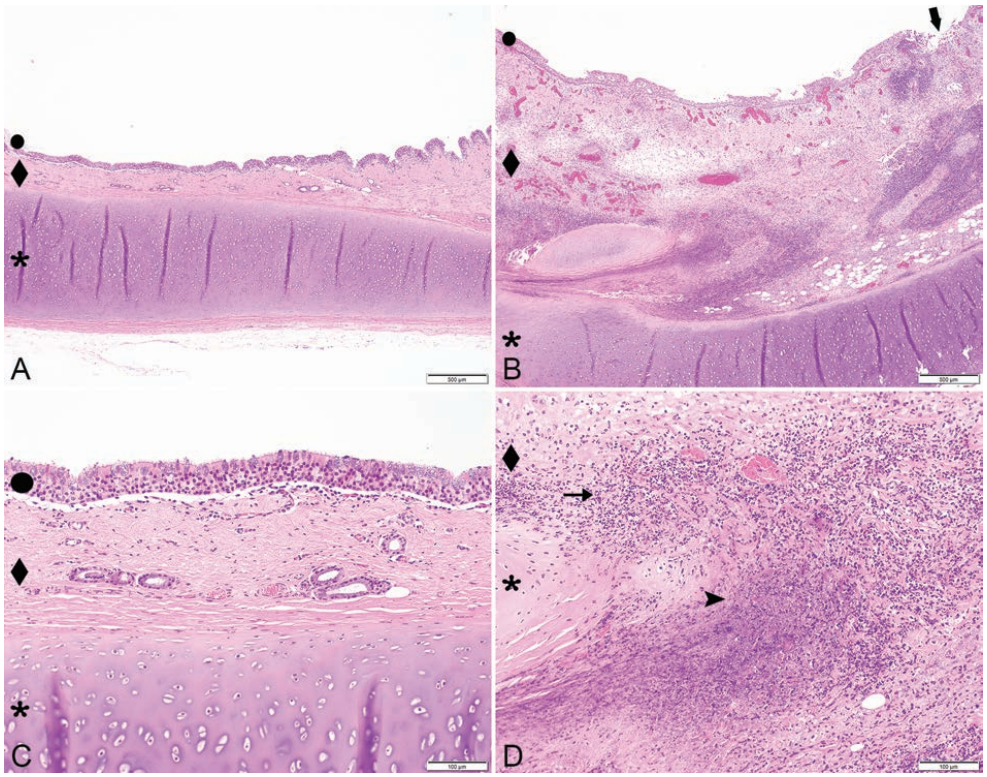

Figure 5: Composite figure displaying representative histological results from tracheal tissue at the inoculation site. Tissues were stained with hematoxylin and eosin. Low (40x) and high (200x) magnification of trachea of a goat inoculated with *Mmc* GM12::YCpMmyc1.1-Δ*68* (panels A and C) depicting unaffected epithelium (circle), submucosa (diamond) and cartilage (asterics). or of a goat inoculated with *Mmc* GM12 (panels B and D), depicting ulceration of the epithelium (thick arrow, panel B), massive extension of the submucosa due to extensive areas of necrosis (diamond, panels B and D) and infiltration with large numbers of degenerate neutrophilic granulocytes (asterisk, panels B and D). Size standards are displayed in each picture.

200

201 **The MIB-MIP system in *Mmc* GM12 is functional *in vivo***

202 The MIB-MIP system was shown to be active *in vitro*, and to be present in large amounts at the
203 cell surface (17) during infection (18). Two animal sera were selected from the *in vivo* infection
204 trial: CK51 from the GM12 group and CK45 from the GM12::YCpMmyc1.1-*Δ68* group and
205 tested for cleavage of IgG. The pre-infection sera from both animals demonstrated no
206 proteolytic cleavage of IgG when compared to the IgG control (Figure 3C). Conversely, the
207 post-infection serum of CK51, which had succumbed to disease and had a high titer of bacteria
208 in the venous blood, clearly exhibited the typical band at 44 kDa, consistent with the size of a

209 cleaved IgG heavy chain by the MIB-MIP system. As expected, no band at 44 kDa was seen in
210 the post-infection serum of CK45. This clearly demonstrates, for the first time, that the MIB-
211 MIP system is functional within the caprine host and that its proteolytic activity is triggered
212 during an *Mmc* infection.

213 **DISCUSSION**

214 The first aim of this work was to fully attenuate a highly pathogenic strain of
215 *Mycoplasma mycoides* following a rational deletion design. The second aim was to verify this
216 attenuation *in vivo* using the native host, since no rodent animal models for highly virulent *M.*
217 *mycoides* exist (3). Many candidate virulence factors of *Mycoplasma mycoides* have been
218 suggested but, all except capsular polysaccharide (19) have never been confirmed according to
219 Falkow's postulates *in vivo* (20).

220 First, we relied on previous knowledge to select five genomic regions that encode candidate
221 virulence traits. These regions, distributed around the *Mmc* genome, consisted of 68 genes. The
222 first two regions (D1 and D2) encode enzymes and putative glycerol transporters involved in
223 the production of hydrogen peroxide using the glycerol-dependent metabolism of mycoplasmas
224 (10). The region D3 encodes major antigens (LppA/P72) in *Mycoplasma mycoides* (21) that
225 induced T cell responses early in infection (22). The region D4 includes an integrative
226 conjugative element (ICE), the MIB-MIP system (9) and the F1-likeX0 ATPase (15). The last
227 region (D5) encodes six lipoproteins including the lipoprotein Q (LppQ), which has been
228 suspected to be involved in exacerbating immune responses and other virulence determinants
229 (23). Interestingly, among these 68 genes deleted in our mutant, 67 were defined as non-
230 essential in the JCVI-Syn3.0 minimal cell, a study in which 432 genes were classified as non-
231 essential to sustain the life of a minimal *Mycoplasma* cell (24). Only one gene encoding the
232 glycerol phosphate kinase (GlpK) (MMCAP2_0218, region D1), which was retained in the
233 minimal JCVI-Syn3.0 cell but, classified as a "quasi-essential" gene. This means that the
234 function encoded by the glycerol kinase, i.e. the transfer of a phosphate group on the glycerol
235 molecule to produce glycerol-3-phosphate, could be compensated for by another gene which
236 encodes a similar function in the full-length genome. The compensatory gene for *glpK* is
237 probably absent in the minimal JCVI-Syn3.0 cell, but present in our mutant

238 GM12::YCpMmyc1.1-*Δ68*, allowing the deletion of *glpK* without any defect in growth.
239 Therefore, our results are consistent with the quasi-essentiality of *glpK.* Altogether we retained
240 69 out of the 87 lipoproteins (D1-0, D2-1, D3-3, D4-8, D5-6) present in GM12, while the
241 minimal cell retained only 15 lipoproteins (24).
242 It was paramount for us that the mutant strain GM12::YCpMmyc1.1-*Δ68* maintains a doubling
243 time similar to its parental strain, since we wanted to create a 'K12-like' strain that can be
244 further used as a cellular platform to introduce antigens or stretches of DNA. It was interesting
245 for us to observe that, despite complete removal of the glycerol pathway, which maybe
246 important for cell metabolism, there was no significant impact on *in vitro* growth. It is known
247 that there is a trade-off between genome size and growth rate. The drastic deletions in the
248 genome of JCVI-syn3.0 strain led to a substantial increase in the generation time, from ~60 min
249 to ~180 min (24). Recently, we also have shown that the deletion of a gene encoding an enzyme
250 important for synthesis of carbohydrates can subsequently lead to an increase in the generation
251 time (25). However, in this work, we significantly reduced the genome of GM12 by more than
252 100 kbp (i.e. 106,737 bp) without seeing any significant difference in the growth rate of
253 GM12::YCpMmyc1.1-Δ*68* in comparison to the wild-type strain. Still, this reduction represents
254 ~10% of the initial genome size confirming that, in addition to the size of the deletions, the
255 nature of the genes deleted is also very likely to influence the generation time of mycoplasmas.
256 The main goal of this work was to construct a fully attenuated *Mmc* strain, that is safe to handle
257 in the laboratory. The first confirmation of the attenuation of the GM12::YCpMmyc1.1-Δ*68*
258 strain was obtained *in vitro.* The production of hydrogen peroxide was almost completely
259 abolished in the mutant strain, confirming the participation of the *glpFKO* and/or *gtsABCD*
260 pathways in the metabolism of glycerol. In addition, the loss of the IgG specific cleavage band
261 at 44 kDa in GM12::YCpMmyc1.1-Δ*68* confirmed the role of the MIB-MIP system in the
262 degradation of the host immunoglobulins.

263 To confirm the mutant strain's attenuation *in vivo*, we developed an animal challenge model
264 using Kenyan goats, outbred animals derived from different herds. The use of such animals
265 increased variability to get a better idea of reproducibility and significance of the results (26).
266 Animals infected with the GM12 strain developed specific symptoms (fever, heavy breathing,
267 septicemia, etc.) and were all euthanized by 6 dpi. Strikingly, none of the goats infected with
268 GM12::YCpMmyc1.1-Δ*68* developed such symptoms and were healthy for the entire course of
269 the experimentation. These results exceeded our expectations and confirmed the complete
270 abolishment of pathogenicty of the mutant strain GM12::YCpMmyc1.1-Δ*68*. In addition, the
271 massive septicemia associated with very high titers of *Mycoplasma* observed in animals
272 infected with the GM12 strain prompted us to investigate whether the MIB-MIP system would
273 leave signatures of its action on immunoglobulins (Ig) in the serum of an animal (CK51) that
274 had a titer of 10^9 CFU/ml. Specific IgG cleavage, characteristic of the MIB-MIP system (9),
275 was observed. No such cleavage was observed in the serum of animals infected with the mutant
276 strain. This work shows, for the first time, that the MIB-MIP system of *Mycoplasma mycoides*
277 is functional *in vivo*. *Mycoplasmas* have been viewed as stealth pathogenic organisms because
278 they lack most of the immune activators or PAMPs found in other bacteria (27). Indeed, the
279 lack of a cell wall or the capacity to produce either LPS or flagellins likely contribute to the
280 chronicity of infection. The only PAMP that has been described for several *Mycoplasma* species
281 is the surface lipoproteins, abundant components of their membrane (8). In the present study,
282 we suggest another mechanism that could contribute to activating the immune system. Indeed,
283 Ig cleaved by several bacteria, including those generated by *Mycoplasma hyorhinis*, have been
284 described as ligands of the innate immune receptor LILRA2 (28). Once bound to this receptor,
285 it triggers the activation of the innate immune system. It is also possible that this cleavage is in
286 line with the 'nutritional virulence' of the parasite (29). The exact significance of the Ig cleavage
287 regarding *Mycoplasma* infection of mucosal surfaces remains to be studied.

288 Interestingly, we observed severe inflammation around the site of injection in the animals that
289 received GM12 whereas animals that were injected with the strain GM12::YCpMmyc1.1-*Δ68*
290 developed no such pathomorphological lesions. Overwhelming immune reactions at the site of
291 vaccination have been reported from immunizations against contagious bovine
292 pleuropneumonia using live *M. mycoides* subsp. *mycoides* based vaccines such as T1/44, which
293 is the closest relative of *Mmc* (30). Therefore, it is likely that any one or several of the deleted
294 genes encode proteins that drive this overwhelming immune reaction in the GM12.
295 To conclude, we confirmed, *in vitro* and *in vivo*, our ability to design a fully attenuated strain
296 *via* the precise reduction of ~10 % of the *Mmc* genome. However, we cannot currently pinpoint
297 the weight of each deletion on the observed attenuation. The total clearance of the pathogen
298 and the absence of a compelling humoral immune response, even at the inoculation point, is
299 surprising and supports the total abolishment of pathogenicity. Now it is necessary to test more
300 defined mutants such as a *glpOKF* mutant strain to get clarity about its real role in
301 pathogenicity.
302 In addition, the design of next generation vaccines for, but not restricted to, *Mycoplasma*
303 diseases will benefit from this study since a chassis that is fully attenuated and able to
304 accommodate antigens for vaccine delivery that can be constructed based on our deletion
305 mutant is now within reach. To induce a proper immune response via such a chassis, we have
306 the option to add genes that appropriately stimulate an inflammatory immune response or
307 alternatively, we can construct different chassis that direct responses towards Th1 or Th2 using
308 TLR agonists. We consider a genetically modified *Mycoplasma* less problematic than other
309 potential chassis since the survival time of *Mycoplasma* in general in the environment is very
310 short. In addition, the unconventional codon usage (where UGA encodes tryptophan) and high
311 AT content of *Mycoplasma* minimizes the risk of spread of genes to other bacteria. Additional
312 experiments are necessary to decipher the role of individual virulence traits to understand these

313 minimal bacterial pathogens better and to develop next generation rationale vaccines.
314 Regardless, this study provides an attractive blueprint towards these goals, especially for those
315 that are needed in low and middle-income countries.

316 **MATERIALS AND METHODS**

317 ***Mycoplasma* strains**

318 The *Mycoplasma mycoides* subsp. *capri* outbreak strain GM12 was used as positive control in
319 the *in vivo* experiment (31). A modified *Mycoplasma capricolum* subsp. *capricolum* strain CK
320 was used as recipient strain in genome transplantation protocols (13).

321 **Yeast strain and media**

322 The yeast *Saccharomyces cerevisiae*, strain VL6-48N (MATαhis3-Δ200 trp1-Δ1 KlURA3-Δ1
323 lys2 ade2-101 met14) containing the 1.08 Mb genome of *Mycoplasma mycoides* subsp. *capri*
324 (*Mmc*) strain GM12 with an integrated yeast centromeric plasmid (YCp) (13) was used for
325 construction of the mutants. Yeast cells were grown and maintained in either synthetic minimal
326 medium containing dextrose (SD, Takara Bio) (13), or in standard rich medium containing
327 glucose (YPD, Takara Bio) or galactose (YPG, Takara Bio) (32). SD medium was
328 supplemented with 5-fluoroorotic acid (5-FOA), for KlURA3 counter-selection (13, 33).

329 **Preparation of mutagenesis cassettes**

330 Sixty eight genes that encode candidate virulence traits were seamlessly deleted from the
331 genome of *Mmc* GM12::YCpMmyc1.1 in five consecutive cycles (D1, D3, D4 and D5) using
332 Tandem Repeat coupled with Endonuclease Cleavage [TREC] as described (32, 34) or a
333 variation of TREC involving the Cre-lox system for the D2 deletion, see below. Primer
334 sequences to target and confirm the insertion of the mutagenesis cassette into each target site
335 and to verify seamless deletion of the targeted genes are shown in Table S 1.

336 The *gts* gene cluster (D2) was targeted and deleted in the *Mmc* genome in the back-ground of
337 the *glpFKO* deletion strain by employing a derivative of the *Mmc* synthetic cell JCVI-syn1.0
338 (35). Primers RC0905 and RC0906 (Table S1) were used to amplify the mutagenesis cassette
339 from the synthetic cell derivative and was targeted to the *gts* region. Specific primers were used
340 to confirm correct insertion at the target site by amplifying the junctions between the

341 GM12::YCpMmyc1.1 genome and the inserted cassette. Galactose induction resulted in the
342 Cre-mediated deletion of the *gts* region, leaving 13 bp of the 5' end of the *gtsA* region, the 34
343 bp *loxP* site, and 27 bp of the 3' end of the *lppB* gene. Specific primers were used to verify the
344 knock-out.

345 **Transformation and PCR analysis**

346 Transformation of the CORE3 cassette was performed by the lithium acetate method as
347 described previously (36). Transformed yeast were plated on appropriate selection media [SD
348 medium minus His (Teknova, CA) or SD medium (minus His and minus Ura)] and incubated
349 at 30°C for 48 hours. Yeast colonies were patched on appropriate selective media and total
350 DNA was isolated for PCR screening (37). The correct insertion of the mutagenesis cassette
351 was verified by PCR amplification using upstream and downstream specific primers (Integrated
352 DNA Technologies, Coralville, IA, USA) (Table S1).

353 **Transplantation**

354 The modified GM12::YCpMmyc1.1 genomes (D1 to D5) were transplanted into *Mycoplasma*
355 *capricolum* subsp. *capricolum* (*Mcc*) recipient cells with polyethylene glycol and selected for
356 tetracycline resistance as described previously (13, 37). The resulting mutant strains were
357 subjected to multiplex PCRs and pulsed-field gel electrophoresis as described elsewhere (38)
358 to confirm integrity of the genome.

359 **Confirmation of the mutants using next generation sequencing and mapping assembly**

360 Total DNA of the strains GM12, GM12::YCpMmyc1.1 and GM12::YCpMmyc1.1-Δ*68* was
361 isolated as described before (39). DNA was sheared using sonication and subjected to Illumina
362 sequencing using a MiSeq machine by University of California Santa Cruz, CA (USA). Reads
363 were mapped to the designed genome sequences based on the parental strains GM12 and
364 GM12::YCpMmyc1.1 and GM12::YCpMmyc1.1-Δ*68*. The raw reads (300bp PE) were QC
365 with FastQC (https://www.bioinformatics.babraham.ac.uk/projects/fastqc/). The corrected

366 reads were mapped onto the reference genome WT-YCP.fa with bwa mem (40) and converted
367 to sorted bam with samtools (41). The bam files were analyzed for deletions using Delly2 (42)
368 and Sprites (43), and the predictions validated visually using IGV (44). The list of strains and
369 their deleted regions is summarized in Table 2.

370 **Scanning electron microscopy of *Mycoplasma***

371 Unless stated otherwise, chemicals were obtained from Merck (Schaffhausen, Switzerland).
372 Mycoplasmas were washed with distilled water (dH_2O) and fixed with 4% para-formaldehyde
373 (Life Technologies, Thermo Fisher, Zug, Switzerland; Cat. No. 28906) in dH_2O for five days
374 at 4°C. Thereafter, samples of 40 µl of cell suspension were centrifuged onto gold-sputtered
375 poly-L-lysine coated coverslips (high molecular poly-L-Lysine hydrobromide) at 125 *rcf* for 4
376 min. Coverslips were washed 1x with PBS and 2x with 0.1% bovine serum albumin in PBS
377 (BSA/PBS). Free aldehydes were blocked with 0.05M glycine in 0.1% BSA-c/PBS (Aurion,
378 ANAWA Trading, Wangen, Switzerland) for 15 min. at room temperature. After 3x washes
379 with 0.1% BSA/PBS, cells were fixed with 2.5% glutaraldehyde (Merck 104239) in 0.1M
380 cacodylate buffer (dimethylarsinic acid sodium salt trihydrate), washed 3x with dH_2O and
381 postfixed with 1% OsO_4 (Polysciences, Warrington, PA, USA) in 0.1M cacodylate buffer for
382 15 min. at room temperature. Another 5x washes with dH_2O were followed by dehydration in
383 an ascending ethanol series. Samples were then transferred to hexamethyldisilazane (Merck
384 814051) for 10 min, air-dried, mounted onto aluminum stubs with carbon conductive adhesive
385 tabs (Ted Pella Inc., Redding, CA, USA) and coated with approximately 25 nm of gold in an
386 SCD004 (Leica Microsystems, Heerbrugg, Switzerland). Secondary electron micrographs and
387 corresponding backscattered images were obtained with a fully digital field emission scanning
388 electron microscope DSM 982 Gemini (Zeiss, Oberkochen, Germany) at an accelerating
389 voltage of 5 kV, a working distance of 6–8 mm and primary magnifications ranging from
390 30,000 to 50,000×.

391 **Growth assay**

392 Overnight cultures of *Mycoplasma* strains were grown at 37°C in SP4 medium containing
393 streptomycin (GM12) or tetracycline (GM12::YCpMmyc1.1, GM12::YCpMmyc1.1-*Δ68*) for
394 about 16 hours. Doubling times of the Mycoplasma strains were then determined as described
395 elsewhere (24), except that time interval samples were collected and processed at 0, 1, 2, 3, 4,
396 5, 6, 7, 9, 12, 15, and 24 h.

397 ***In vitro* hydrogen peroxide assay**

398 Overnight cultures of *Mycoplasma* strains were grown as described above. When the pH of
399 overnight cultures reached 6.0 - 6.5, they were inoculated into fresh SP4 medium at 1:200
400 dilution and incubated at 37 °C for different time intervals of 0, 5, 7, and 24 h. At each time
401 interval, an aliquot of culture was taken for DNA extraction (24) and another aliquot was taken
402 to determine hydrogen peroxide levels.

403 To determine hydrogen peroxide levels, the aliquots were spun at 14,000 rpm for 10 minutes at
404 4°C. The pellets were washed with 1 ml of cold PBS, pH 7.5 to remove traces of media, then
405 resuspended in 400 µl of cold PBS and stored at 4°C. Hydrogen peroxide levels were
406 determined using the Amplex Red Hydrogen Peroxide Assay Kit (Life Technologies, NY)
407 according to the manufacturer's instructions. Briefly, 50 µl of diluted samples (1:5 in PBS) was
408 aliquoted onto 96-well plates and warmed to 37°C for 1 h prior to starting the assay. 100 µM
409 final concentration of glycerol (Sigma Aldrich, MO) or GPC (Sigma Aldrich, MO) was then
410 added to the diluted sample and incubated at 37°C for 1 h. 50 µl of the Amplex Red reagent
411 was added to the samples, incubated at room temperature in the dark for 30 minutes and
412 fluorescence was measured using a spectrophotometer (SpectraMax M5, Molecular Devices,
413 CA). Three technical replicates were performed for each sample and normalized to their
414 respective DNA concentrations.

415 **Detection of immunoglobulin degradation by *Mycoplasma in vitro* and *in vivo***

416 *In vitro* functionality of the MIP-MIP system was tested using the strains GM12,
417 GM12::YCpMmyc1.1 and GM12::YCpMmyc1.1-Δ*68*. Each strain was grown overnight at
418 37°C in 3 mL modified SP5 medium (containing 5% FBS). 250 µL of each culture was
419 harvested and centrifuged for 10 minutes at 4,000 *g*. The cells were then resuspended in 15 µL
420 modified SP5 medium (containing 5% FBS) and incubated with 5 µg of purified caprine IgG
421 (Sigma) at 37°C for 3 hours. Bacterial CFUs were estimated for each strain by serial dilutions
422 and were $3.2*10^9$ CFUs *Mmc* GM12, $8.5*10^8$ CFUs *Mmc* GM12::YCpMmyc1.1 and $3.5*10^8$
423 CFUs *Mmc* GM12::YCpMmyc1.1-Δ*68*. A sample consisting of 5 µg caprine IgG in dH_2O was
424 included as a control. The incubated samples were mixed with 2x Laemmli Sample Buffer (Bio-
425 Rad,) at a 1:1 ratio, boiled for 10 minutes at 98°C and separated onto a 12% SDS-PAGE gel.
426 They were subsequently transferred onto a 0.2 µm nitrocellulose membrane (Bio-Rad) using a
427 Bio-Rad Trans-Blot® Turbo™ Transfer System (25 volts, 1.0 A, 30 minutes). Next, a Western
428 Blot was performed using PBS supplemented with 0.1% Tween-20 and 2% BSA as a blocking
429 buffer, and mouse anti-goat IgG (H+L) (Jackson Immuno Research, 205-005-108) and goat
430 anti-mouse (Fc) labelled with horseradish peroxidase (Sigma, A0168) as primary and secondary
431 antibodies. The antibodies were diluted in blocking buffer at 1:2000 and 1:70,000, respectively,
432 and incubated with the membrane for 1 hour each. In between the antibody incubations, the
433 membrane was washed once with PBS – 0.1% Tween-20 + 3.2% NaCl and twice with PBS –
434 0.1% Tween-20, for 10 minutes each time. The results were visualized using the Fujifilm LAS-
435 3000 Luminescent Image Analyzer.
436 Serum samples derived from the *in vivo* challenge were diluted in water to achieve a total load
437 of 10-20 µg of protein per sample. Western Blots were performed and analyzed using the same
438 protocol as described above.
439 **Animal experiment setup**

440 All protocols of this study were designed and performed in strict accordance with the Kenyan
441 and US American legislation for animal experimentation and were approved by the institutional
442 animal care and use committee at both institutions (JCVI and ILRI, IACUC reference number
443 2014.08).
444 Sixteen male outbred goats (*Capra aegagrus hircus*), 1-2 years of age and randomly selected
445 in Naivasha, were transferred to the ILRI campus in Nairobi and kept under quarantine for 6
446 months. After arrival at the campus, all animals were dewormed twice using levamisole and
447 treated prophylactically against babesiosis and anaplasmosis using imidocarb. Upon entry to
448 ILRI, the goats were vaccinated against anthrax & blackleg (Blanthax®, Cooper), Foot and
449 Mouth Disease (FOTIVAX®) and Peste des Petits Ruminants (Live attenuated strain Nig.
450 75/1). All animals were tested negative for presence of antibodies against CCPP, using a
451 competitive ELISA (IDEXX). Two weeks before experimental infection, all animals were
452 transferred to the animal biosafety level two (ABSL2) unit. *Mycoplasma* cells were cultivated
453 in PPLO medium supplemented with horse serum (45) to early logarithmic phase, aliquoted
454 and stored at -80°C. Afterwards, we determined the CFU using two aliquots. Just before
455 infection we thawed the vials and adjusted the concentration of *Mycoplasma* to 10^9CFU per
456 mL^{-1} using broth. All 16 goats were infected transtracheally by needle puncture 5-10 cm distal
457 to the larynx. Each animal received 1 mL of *Mmc* GM12 or GM12::YCpMmyc1.1-Δ*68* liquid
458 culture (10^9 colony forming units per animal), followed by 5 mL of phosphate buffered saline
459 (PBS). The animals were allowed to move freely within the ABSL2 unit and had *ad libitum*
460 access to water. They were fed *ad libitum* with hay and received pellets each morning. Three
461 veterinarians monitored the health status of the animals throughout the experiment. Rectal
462 temperature, oxygen blood saturation, heart rate & breathing frequency were measured daily in
463 the morning hours using the GLA M750 thermometer (GLA agricultural electronics, USA),
464 VE-H100B oximeter (Edan, USA), and a stethoscope classic II (Littmann, USA) with a water-

465 resistant wrist watch Seamaster (Omega, Switzerland), respectively. Blood samples for
466 subsequent analysis were taken twice a week by jugular vein puncture. Nasal swabs (Flocked
467 swabs, Copan, Italy) were taken twice a week. Swabs were transferred into cryotubes filled
468 with media and stored at -80°C until further processing. Goats were euthanized when they
469 developed severe disease associated with unwarranted moderate to severe pain. Therefore, they
470 received an intravenous injection of Lethabarb Euthanasia Injection (Virbac, USA) of 200
471 $mg.kg^{-1}$ body weight. Severe disease and pain were determined by a fever of ≥41ºC for >3
472 consecutive days, an oxygen saturation of ≤92% and a lateral recumbency of ≥1 day without
473 the ability to feed or intake water. Goats that were not put down because of ethical reasons were
474 euthanized on 28 dpi.

475 **Pathomorphological and histology analysis**

476 A complete necropsy was performed on all animals. Tissue samples of the neck region around
477 the inoculation site and all internal organs were fixed in 10 % buffered formalin for 72 hours
478 and subsequently routinely processed for paraffin embedding. Tissue sections were cut at 3 µm
479 and stained routinely with hematoxylin and eosin (H&E) and evaluated by a board-certified
480 pathologist.

481 **Microbiology**

482 Venous blood samples, lung samples, carpal joint fluid, and pleural fluid specimens taken at
483 necropsy were used for isolation of *Mmc* as described elsewhere (46) using *Mycoplasma* liquid
484 medium (Mycoplasma Experience, Ltd., United Kingdom). Lung samples and pleural fluid
485 were used for screening of *Pasteurella* and *Mannheimia* spp. using standard methods (47).

486 **Statistical analysis**

487 Exact and normal approximation binomial tests were used to compare the two groups using
488 GenStat 12th Edition (48). P values for differences in parameters were estimated using a 2-
489 sided 2-sample t-test comparing average levels between both groups at 5% level of significance.

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AUTHOR CONTRIBUTIONS

J.J. and S.V. designed research; L.M., N.G-A., S.C., J.J., A.L., P.S., M.S., E.S., V.C.,Y.A., H.P. and F.L. performed research; J.J., F.L., S.C., Y.A., L.F., P.S-P., C.L., A.B. and SV., analyzed data; J.J. and S.V. wrote the paper

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Table 1: Summary of post mortem records of goats

Animal No.	Date of euthanasia	Bacteremia ccu/ml	Inflammation of the neck around the injection site	Pulmonary congestion	Pulmonary oedema	Congested kidneys	Mucoid enteritis & congestion	Pleural fluid in thoracic cavity	Adherent lung to rib cage	Liver abscess
CK032	6 dpi	10^7	X	X	X	X	X	X		
CK034	5 dpi	10^7	X	X	X	X	X			
CK035	29 dpi									
CK040	5 dpi	10^8	X	X	X	X	X			X
CK043	5 dpi	10^8	X	X	X	X	X			
CK045	28 dpi									
CK046	5 dpi	10^8	X	X	X	X	X			
CK047	29 dpi									
CK048	5 dpi	10^9	X	X	X	X	X			
CK049	28 dpi									
CK051	5 dpi	10^9	X	X	X	X	X			
CL001	28 dpi									
CL002	5 dpi	10^8	X	X	X	X	X		X	
CL003	28 dpi									

Animals displayed in bold were infected with GM12::YCpMmyc1.1-*Δ68*, the other animals were infected with wtGM12; dpi - days post infection, ccu – color changing unit

Table 2: Table showing the results of the Illumina sequencing-based mapping assemble of the strains used in this study

Region (size in bp)	**YCP (9,192)**	**D1 (2,984 bp)**	**D2 (4,950 bp)**	**D3 (4,677 bp)**	**D4 (69,220 bp)**	**D5 (24,906 bp)**
GM12	Not present	Present (I)	Present (I)	Present (I)	Present (I)	Present (I)
GM12::YCpMmyc1.1 (position)	Present (25,426-34,618)	Present (283,950-286,934)	Present (569,098-574,048)	Present (18,716-23,393)	Present (665,418-734,638)	Present (1,055,169-1,080,075)
GM12::YCpMmyc1.1-*Δ68*	Present (I)	Deleted (DIS)	Deleted (IS)	Deleted (DIS)	Deleted (DIS)	Deleted (DIS)

Legend: D = Detected by Delley; I = Validated visually by IGV; S = Detected by Sprites

Table S1: Oligonucleotide primers used to construct the mutagenesis cassettes and to confirm the deletions

Primer Name	**Primer sequence (5'→ 3')**	**PCR product and notes**
D1: Deletion using TREC method (modified CORE3 cassette):		
FKO-F1	G**TT GAT TCA GTC TTC ATA ACG TTT TTT ACC TAA AAC ATT TAA TTT TGC ATT** *TAG GGA TAA CAG GGT AA TAC GGA TTA G*	To amplify the 2.4 kb CORE3 mutagenesis cassette with 50 bp homology overhangs at the target site.
FKO-R	**AAT GCA AAA TTA AAT GTT TTA GGT AAA AAA CGT TAT GAA GAC TGA ATC AA***T CGG TAC ATA AAT ATA TGT GAT TCT*	
FKO-F2	CTG AAG ATT TAG CAG CAG TTA ATC CTC AAA TAC CAA CTA ATA GAA CTA CAG **TTG ATT CAG TCT TCA TAA CGT TT**	
FKO-DG-F	CCA GTT GTT CCA CCT AAT G	To amplify 235 bp with RC0421 to confirm the left junction of the glpFKO-CORE3 insertion. To amplify 250 bp with FKO-DG-R to confirm pop-out and seamless deletion of the glpFKO genes.
RC0421	CTT CGG AGG GCT GTC ACC	
FKO-DG-R	TCG CAT TAG AAA CAA GTA GTC	To amplify 283 bp with RC0430 to confirm the right junction of the glpFKO-CORE3 insertion.
RC0430	AAG TGT CAC CAT GAA CGA CA	
D2: Deletion using TREC method (Cre/ URA3 cassette):		
RC0905	CTA ATT AAT ATA AAT GAA AGA ATT AAA	To amplify the 2.4 kb Cre-Ura3 mutagenesis cassette with 50 bp homology overhangs at the target site.
RC0906	TTA AAG AAT TAT TAC TTG GTA TAT TA	
RC0350	CGG CTA AAG CTA AAA TCA AAA GAA C	To amplify 615 bp to confirm the left junction of the gts-CRE/URA3 insertion. To amplify 650 bp with RC0350 and RC0332 to confirm pop-out and seamless deletion of the gts-5kb region.
RC0373	ATA TTT GAG AAG ATG CGG CCA GC	
RC0332	AAG AGC ATA TTA GTA AAA TAT TGT CA	
D3: Deletion using TREC method (CORE3 cassette):		
lppA123-F1	**TAA AAC TGG AAA AAA CTA TGA AAA AAG CAA TTA AAT TAT TAC TAT CTA TT***T AGG GAT AAC AGG GTA ATA CGG ATT A*	To amplify the 2.5 kb CORE3 mutagenesis cassette with 50 bp homology overhangs at the target site.

lppA123-R	**AAT AGA TAG TAA TAA TTT AAT TGC TTT TTT CAT AGT TTT TTC CAG TTT TA***T CGG TAC ATA AAT ATA TGT GAT TCT*	
LppA123-F2	TAG CTA TGT TGT TAT AAT ATT TTC GAA AAA TTT TTT GTA GGT ATA TTA TT**T AAA ACT GGA AAA AAC TAT GAA AAA AG**	
lppA123-DG-F	CTA GAT GTA AAT TAC CAC TAG G	To amplify 360 bp with RC0421 to confirm the left junction of the lppA-CORE3 insertion. To amplify 436 bp with lppA-DG-R to confirm pop-out and seamless deletion of the lppA region.
lppA123-DG-R	GGT TTT GAT GGT GTT TTT GGA G	To amplify 237 bp with RC0430 to confirm the left junction of the lppA-CORE3 insertion.
D4: Deletion using TREC method (CORE3 cassette):		
50CORE-F1	**TTTTAATCCTCCAACCTATTAATATTTTAAATAAGAT AAAACATTGTTGG***TAGGGATAACAGGGTAATACGGATTA G*	To amplify the 2.5 kb CORE3 mutagenesis cassette with 50 bp homology overhangs at the target site.
50CORE-R	CCAACAATGTTTTATCTTATTTAAAATATTAATAGGTTG GAGGATTAAAA*TCGGTACATAAATATATGTGATTCTG*	
50CORE-F2	AGTATAAAACTTTTATCCTAACCGATTTTAAGTTTTATA CAGGAGGAATT**TTTTAATCCTCCAACCTATTAATATTT**	
RC0863	TCAACATATTCTGGTATGTCT	To amplify 425 bp with RC0421 to confirm the left junction of the ICE-CORE3 insertion. To amplify 480 bp with RC0878 to confirm pop-out and seamless deletion of the ICE region.
RC0878	TAATCTAACACAACCGGTAG	To amplify 325 bp with RC0430 to confirm the left junction of the ICE-CORE3 insertion.
D5: Deletion using TREC method (CORE3 cassette):		
TREC37-F1	**TAATTCATTAGTCTCTAATTCTTATTAAAAGATTTAG AGATTTTTTATTT***TAGGGATAACAGGGTAATACGGATTAG*	To amplify the 2.5 kb CORE3 mutagenesis cassette with 50 bp homology overhangs at the target site.
TREC37-R	**AAATAAAAAATCTCTAAATCTTTTAATAAGAATTAGA GACTAATGAATTA***TCGGTACATAAATATATGTGATTCT*	
TREC37-F2	CTAGAAATATATTAATATATCATTAAAAATAAAAATAG CATTTAAGATT**TAATTCATTAGTCTCTAATTCTTATTA A**	

TREC37 -DG-F	TGGTGTTGCTACTGAAATATG	To amplify 250 bp to confirm pop-out and seamless deletion of the D5 region.
TREC37 -DG-R	ACTTAAAAAAGAACGACACCG	

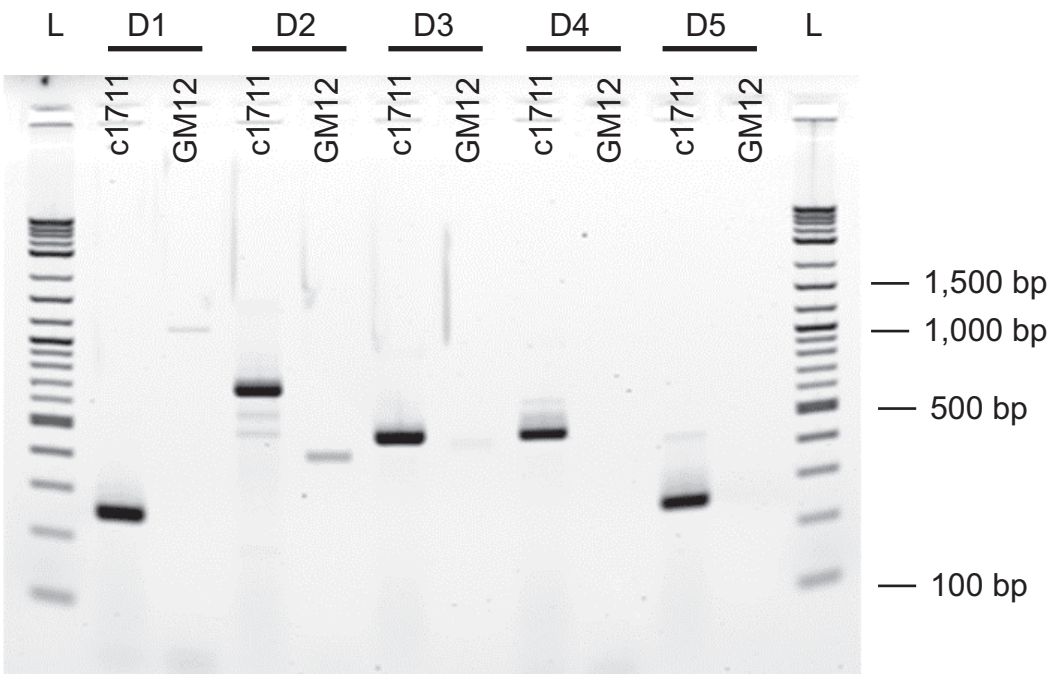

Figure S1: Specific PCRs confirming the deletions in GM12::YCpMmyc1.1-*Δ68*. Genomic DNA from GM12::YCpMmyc1.1-*Δ68* (clone 1711) and its parental strain GM12 was used as template. Specific primers were used to amplify across the deleted regions. DNA from GM12::YCpMmyc1.1-*Δ68* (clone 1711) yielded expected amplicons that are a product covering the flanking regions of the deleted subgenomic fragment. Diagnostic primers FKO-DG-F and FKO-DG-R result in a 250 bp amplicon across the D1 region in clone 1711, primers RC0350 and RC0332 result in a 650 bp amplicon across the D2 region, primers LppA123-DG-F and LppA123-DG-R result in a 436 bp amplicon across the D3 region, primers RC0863 and RC0878 result in a 480 bp amplicon across the D4 region, and primers TREC37-DG-F and TREC37-DG-R result in a 250 bp region across the D5 region. L denotes the DNA ladder.