**EvolvingSTEM: A microbial evolution-in-action curriculum that enhances learning**

**of evolutionary biology and increases interest in STEM**

Vaughn S. Cooper[1,2,3,5], Taylor M. Warren[3], Abigail M. Matela[1,2], Michael Handwork[4],

Shani Scarponi[4]

1: Department of Microbiology and Molecular Genetics, and 2: Center for Evolutionary

Biology and Medicine, University of Pittsburgh, School of Medicine, Pittsburgh, PA USA

3: Department of Molecular, Cellular, and Biomedical Sciences, University of New

Hampshire, Durham, NH USA

4: Winnacunnet High School, Hampton, NH USA

5: Corresponding author: vaughn.cooper@pitt.edu

**Abstract**

Evolution is a central, unifying theory for all of life science, yet the subject is

poorly represented in most secondary-school biology courses, especially in the United

States. One challenge to learning evolution is that it is taught as a conceptual,

retrospective subject with few tangible outcomes for students. These typical passive

learning strategies lead to student disengagement with the material and

misunderstanding of evolutionary concepts. To promote greater investment and

comprehension, we developed EvolvingSTEM, an inquiry-based laboratory curriculum

that demonstrates concepts of natural selection, heredity, and ecological diversity

through experimental evolution of a benign bacterium. Students transfer populations of

*Pseudomonas fluorescens* growing on plastic beads, which selects for biofilm formation

and mutants with new, conspicuous phenotypes. We introduced our curriculum to four

introductory high school biology classes alongside their standard curriculum materials

and found that students who learned evolution through EvolvingSTEM scored

significantly better on a common assessment targeted to Next Generation Science

Standards than students taught only the standard curriculum. This latter group

subsequently achieved similar scores once they too completed our curriculum. Our work

demonstrates that inquiry-based, hands-on experiences with evolving bacterial

populations can greatly enhance student learning of evolutionary concepts.

## Introduction

**Introduction**

Understanding evolutionary processes is fundamental to all areas of life science

because evolution serves as a conceptual framework to organize other life science

topics, such as organismal diversity and ecological interactions. Furthermore, some of

the most significant threats to human health are evolutionary phenomena; therefore,

knowledge of evolutionary processes has a direct impact on public health and medicine

(Wells et al. 2017). For example, antimicrobial resistance and cancer are caused by the

rapid evolution of microbes and our own cells, respectively (Karatan and Watnick 2009;

Greaves and Maley 2012; Berendonk et al. 2015; Makohon-Moore and Iacobuzio-

Donahue 2016; Alizon and Méthot 2018). In addition, ongoing revolutions in

biotechnology and personalized medicine, such as gene-editing (i.e., CRISPR), can

only be understood in the context of the evolutionary concept of descent from a shared

ancestral lineage (Makarova et al. 2015; Knott and Doudna 2018). A strong knowledge

base of evolution is therefore invaluable for a literate society to understand scientific

and medical advances and for a prepared workforce to excel in jobs in science,

technology, and engineering. The value of evolutionary biology knowledge is highlighted

by its inclusion as a core concept for STEM education practices (National Research

Council 2012; NGSS Lead States 2013; NSTA 2013).

Although the importance of evolutionary biology is well-established,

misconceptions of its basic principles remain prevalent among students, the general

public, and even the teachers who are providing instruction (Cunningham and Wescott

2009; Gregory 2009; Sickel and Friedrichsen 2013; Yates and Marek 2014; Glaze and

Goldston 2015). While many coexisting factors likely contribute to poor understanding

(Smith 2010a; 2010b; Pobiner 2016), one potential reason that evolutionary concepts

are misunderstood is that typical curricula use passive learning strategies, where

instruction relies on lectures and textbook readings. Current evolution curriculum design

runs counter to evidence that student-centered, active learning strategies are the most

effective method for science teaching and have been shown to improve student

understanding of evolutionary concepts (Nehm and Reilly 2007; Nelson 2008; Freeman

et al. 2014; Romine et al. 2017). Courses that provide students with authentic research

experiences are especially effective at increasing student engagement and promoting a

deeper understanding of evolution (Jordan et al. 2014; Ratcliff et al. 2014; Broder et al.

2018).

75           There is therefore a critical need for engaging and informative evolutionary

biology curricula that provide young students the opportunity to explore the concept of

changing frequencies of inherited traits just as they attempt to quantify gravity in physics

or acid-base reactions in chemistry. To meet this need, we developed EvolvingSTEM, a

curriculum that provides hands-on, inquiry-based learning of evolution, microbiology,

ecology, and heredity with an experiment that employs real scientific research practices.

EvolvingSTEM allows students to visualize evolutionary adaptations arising in real time

by growing populations of the harmless bacterium *Pseudomonas fluorescens* under

conditions that select for the formation of a biofilm. A biofilm is a surface dwelling

community of microbes encased in a protective coating of self-produced polymers;

biofilms are the dominant form of microbial life (Costerton et al. 1987). They are also

structured, heterogeneous environments that include varied ecological niches (Karatan

and Watnick 2009). Bacteria with advantageous mutations colonize these niches, and

their adaptations cause visible differences in colony morphology from the ancestral

genotype (Rainey and Travisano 1998; Flynn et al. 2016). This evolution-in-action

occurs within days, requires little specialized equipment, and can be offered in any

classroom laboratory that can support sterile technique. Our curriculum is intended to

replace standard, passive learning curricula to meet competencies for natural selection

and evolution described in the Next-Generation Science Standards (HS-LS4, (NGSS

Lead States 2013)). We hypothesized that students who learn evolutionary concepts

with our curriculum would have significant increases in content knowledge relative to

students that were provided only the standard curriculum.

**Results**

***Developing and refining an amenable protocol for teaching bacterial evolution to***

***high school students***

The idea to teach evolutionary concepts to high school students with a bacterial

evolution experiment grew from our research on identifying the causes of rapidly

evolving mutant colony morphologies of the opportunistic pathogens *Burkholderia*

*cenocepacia* and *Pseudomonas aeruginosa* (Poltak and Cooper 2011; Flynn et al.

2016). These species are particularly threatening to persons with cystic fibrosis, where

they cause chronic airway infections by forming biofilms (Starkey et al. 2009; Ashish et

al. 2013). Biofilm-associated infections are inherently more resistant to host immunity

and antimicrobials because secreted adhesive polymers are protective and the cells

within grow more slowly (Harrison et al. 2005). Eventually, some bacteria disperse from

the colony, either as individuals or clusters, to inhabit new surfaces and resume the

biofilm lifecycle (Poltak and Cooper 2011; Martin et al. 2016).

In order to study the dynamics of bacterial evolution *in vitro*, we developed a

simple method to model the biofilm lifecycle of surface attachment, biofilm formation,

dispersal, and recolonization (Figure 1, (Poltak and Cooper 2011; Traverse et al. 2013;

O'Rourke et al. 2015; Flynn et al. 2016; Turner et al. 2018). In short, we culture bacteria

for 24 hours in test tubes containing growth media and a polystyrene bead. A subset of

the bacteria colonize the bead and form a biofilm. We then transfer only the biofilm-

covered bead to a new tube with a fresh bead. We repeat this process daily to select for

bacterial mutants that are best adapted to aspects of the entire biofilm lifecycle.

Conveniently, we found that biofilm adapted mutants also display altered colony

morphologies when grown on agar plates, making them conspicuous to students.

In collaboration with science teachers at Winnacunnet High School (Hampton,

NH), we modified our research laboratory protocol to accommodate implementation in a

high school classroom. We selected the plant probiotic bacterium, *Pseudomonas*

*fluorescens* SBW25*,* as our study subject because it had several qualities that made it a

good candidate for use in a high school classroom: (1) it is benign, and thus safe for

students with no microbiology experience, (2) it had previously been suggested as a

good candidate for use in educational settings (Green et al. 2011; Spiers 2014), and (3)

it is the subject of a large body of research on its capacity for rapid and conspicuous

adaptive evolution in biofilm-related conditions (Rainey and Travisano 1998; Spiers

2005). Adaptive *P. fluorescens* mutants are often characterized by rugose or rosette-like

colony morphologies resulting from greater production of polysaccharides for

attachment (Rainey et al. 2000). We found that experimental evolution of *P. fluorescens*

SBW25 in the biofilm lifecycle model selected for a high frequency of adaptive mutants

with novel colony morphologies in less than two weeks.

Fig. 1. Biofilm lifecycle model. Bacteria are grown in test tubes with plastic beads on which biofilm forms. Daily bead transfers select for bacterial attachment, assembly, dispersal, and reattachment. Figure adapted from (Turner et al. 2018).

To accelerate this process and ensure that our experiment could be performed

within the timeframe of a high school biology lesson, we conducted a series of trials in

different media to determine conditions that resulted in predictable, rapid adaptations.

We found that growth in King's B medium (KB) generated multiple, heritable colony

phenotypes within seven days. In the interest of accelerating the evolutionary dynamics,

we repeated the experiment in KB medium with various glycerol concentrations. We

found that an increase from 1.5% to 2.0% glycerol selected for novel colony

morphologies at detectable frequencies in four days. We named this modified media

recipe "Queen's B" (QB) and used this recipe thereafter. Media recipes are available as

a supplemental file (Supplemental File 1).

| **Tube appearance** | **Colony appearances on agar plates** | **Darwin's finch analogy** |
|---|---|---|

Fig. 2. Adaptation to biofilm selection can occur within days and produce conspicuous phenotypic differences. Populations were founded with equal ratios of Lac+ (blue) and Lac- (white) ancestral genotypes that do not differ in morphology. After 5-7 days, new colony morphologies evolve and represent different biofilm-associated ecological strategies, as different beak shapes of Darwin's finches represent distinct feeding strategies (Rainey and Travisano 1998, Poltak and Cooper 2011).

Students can use our modified protocol to guide an inquiry-based experiment

that allows them to visualize evolution in their bacterial populations in only six class

periods (Fig 2). For example, on Monday, students inoculate glass test tubes containing

QB media and a polystyrene bead with a clone of *P. fluorescens* SBW25, and then

perform bead transfers for the following three days (Tuesday-Thursday). During the

process of bead transfer, students can identify effects of natural selection by observing

increased biofilm production on the walls of their test tubes. In addition, at the beginning

and end of the week, students sample their populations by growing individual bacterial

colonies on agar plates. Students can make observations of mutant colonies on the

Monday of the following week and compare these colonies to those of the ancestral

population that were plated earlier in the week. Students can be given additional

curriculum materials, such as homework and pretests, to prepare them for each step in

the laboratory protocol and provide opportunities for them to link the heritable, adaptive

evolutionary change they observe in their experiment to the evolutionary processes that

produced this dynamic. Through EvolvingSTEM, students can acquire the knowledge to

meet Next Generation Science Standards for Natural Selection and Evolution (Box 1;

(NGSS Lead States 2013)). Curriculum materials are available as supplemental files

(Supplemental File 2-3).

***Learning outcomes***

The exact outcome of any individual experiment is unknown because the biofilm

selection acts on randomly occurring mutations in the bacterial populations that were

founded from a single clone. In fact, this variability among these independent "replays"

of evolution is realistic and demonstrates effects of chance and contingency on

evolution (Blount et al. 2018). Nonetheless, student groups propagate multiple

populations in different culture tubes under identical experimental conditions, and this

replication means they are very likely to see mutants with novel morphologies in at least

one experimental population. In addition, students compare their experimental

populations to a control population that does not contain the bead and therefore is not

under selection for increased biofilm production. Students can examine the phenotypes

found in each population over time, compare their findings to those of other classmates,

and develop their own explanations for their observations. This allows students to apply

the comparative method of evolutionary biology and begin the process of scientific

inquiry. Students are encouraged to consider why their replicate populations vary and

propose reasons for that variation, ranging from experimental error, to peculiarities of

the bead transfers, to genuine evolutionary randomness.

**Box 1.** Next Generation Science Standards (NGSS) Targeted by EvolvingSTEM. NGSS (2013) are based on *A Framework for K-12 Science Education: Practices, Crosscutting Concepts, and Core Ideas* (National Research Council 2012) and designed through a collaboration between 26 states, the National Research Council, the National Science Teachers Association, the American Association for the Advancement of Science, and Achieve, Inc.

EvolvingSTEM provides students with the knowledge to meet the following NGSS HS-LS4 standards:
1. Communicate scientific information that common ancestry and biological evolution are supported by multiple lines of empirical evidence.
2. Construct an explanation based on evidence that the process of evolution primarily results from four factors: (1) the potential for a species to increase in number, (2) the heritable genetic variation of individuals in a species due to mutation and sexual reproduction, (3) competition for limited resources, and (4) the proliferation of those organisms that are better able to survive and reproduce in the environment.
3. Apply concepts of statistics and probability to support explanations that organisms with an advantageous heritable trait tend to increase in proportion to organisms lacking this trait.
4. Construct an explanation based on evidence for how natural selection leads to adaptation of populations.
5. Evaluate the evidence supporting claims that changes in environmental conditions may result in (1) increases in the number of individuals of some species, (2) the emergence of new species over time, and (3) the extinction of other species.

As an example, for HS-LS4-2, students will learn:
- Random mutation results in genetic variation between members of a population.
- Genetic variation can result in trait variation that leads to performance differences among individuals.
- Competition for limited resources results in differential survival. Individuals with more favorable phenotypes are more likely to survive and reproduce, thus passing traits to subsequent generations.
- Evolutionary fitness is measured by reproductive success.
- An adaptation is a heritable genetic variant manifested as a trait that provides an advantage to an individual in a particular environment.
- In addition to natural selection, chance and random events can influence the evolutionary process, especially for small populations.

In addition, students will be skilled at:
- Developing experimental investigations that can be used to test specific hypotheses.
- Evaluating evidence to qualitatively and quantitatively investigate the role of natural selection in evolution.
- Constructing evidence-based explanations that the process of evolution is a consequence of the interaction of four factors: (1) the potential for population size to increase, (2) genetic variation, (3) competition for resources, and (4) proliferation of individuals better able to survive and reproduce in a particular environment.
- Applying basic mathematics to calculate the fitness advantages of selected mutants and/or to compare differences in levels of biofilm production.
- Developing generalizations of the results obtained and/or the experimental design and applying them to new problems, including the design of new experiments and interpreting results in the context of natural and infectious bacterial biofilms.

The speed of adaptation in biofilm models results from strong selection for more

adherent mutants that bind not only the provided surface (e.g. polystyrene), but also

other attached bacteria or secreted substances. Consequently, selection often favors

the evolution of diverse, conspicuous phenotypes within each tube and not just a single,

more adherent type. This result not only simulates the process of adaptive radiation

often illustrated using Darwin's finches in textbooks (Figure 2), but also reproduces the

selection for traits associated with adherence that often occurs during biofilm-associated

infections (Traverse et al. 2013; Cooper et al. 2014; O'Rourke et al. 2015; Gloag et al.

2018). The "wrinkly" colony morphologies that evolve in our model are genetically and

functionally identical to those commonly isolated from infections of the related species

*Pseudomonas aeruginosa* in the airways of cystic fibrosis patients and in chronic skin

wounds (Gloag et al. 2018(Starkey et al. 2009)). Students can therefore connect their

classroom experiments to recent findings at the interface of evolutionary biology and

medicine to see how basic biological research impacts their everyday lives.

Furthermore, making connections from classroom activities to real-world examples can

increase students' evolutionary understanding and engagement (Beardsley et al. 2011;

Infanti and Wiles 2014).

**Assessment of student learning**

We used a delayed intervention approach to assess learning in 4 classes of 9th

grade biology honors students at Winnacunnet High School, a suburban public high

school in New England. Group 1 included classroom A, taught by MH, and classroom B,

taught by SS. This group used an earlier version of our EvolvingSTEM curriculum that

did not use a control population alongside their standard curriculum materials, which

included textbook readings, lectures, and an educational video. Group 2 included

classrooms C and D, both taught by SS. This group first received the standard

curriculum with additional lecture materials, followed by EvolvingSTEM (Table 1).

Students conducted the experiments and analyses for our curriculum in groups of three

or four individuals, requiring collaborative teamwork.

A knowledge assessment was used to determine whether students achieved an

increased understanding of evolutionary concepts. The test consisted of multiple choice

and free response questions to address student learning across multiple categories of

Bloom's Taxonomy of cognitive domains, which evaluates student knowledge gains at

hierarchical levels of complexity (Zoller 1993; Crowe et al. 2008). Test questions were

devised to directly assess whether students met NGSS (2013) performance

expectations HS-LS4-1,2,3, and 5. We developed a grading rubric for the free response

questions based on templates suggested by Wiggins and McTighe (2005) that required

answers with accurate information, specific vocabulary, and a well-structured defense

that incorporated outside examples (Wiggins and McTighe 2005). Our assessment and

grading rubric are available as supplemental files (Supplemental File 4).

| Group | Class – Teacher | Number of Students per Class | Total Number of Students per Group |
|---|---|---|---|
| 1 | A – Teacher MH | 19 | 41 |
| | B – Teacher SS | 22 | |
| 2 | C – Teacher SS | 18 | 37 |
| | D – Teacher SS | 19 | |

282
283    **Table 1: Composition of Study Groups.**
284

Pretests were given to both groups prior to the start of classroom evolution

activities. Group 1 students were given a posttest after completing the EvolvingSTEM

curriculum. Group 2 students were given a midtest after completing the standard

curriculum, and then a posttest after completing EvolvingSTEM. We found no significant

difference between the average pretest score of Group 1 and Group 2 students (13.17

(26%) vs. 12.5 (25%) out of 50 points total; $t=0.60$, $p=$n.s.), indicating that all students

began with a similar knowledge base (Fig. 3). Quantitative analyses of student

knowledge gains revealed that students who completed EvolvingSTEM (Group 1)

showed significant improvement on their average posttest scores, with an average gain

of 19.16 points, thereby increasing their overall score by 38% between the pre- and

posttest ($t=16.61$, $p<0.0001$). Students provided the standard curriculum (Group 2) also

showed significant improvement on their average midtest score, which increased by

10.14 points (t= 9.72, p<0.0001), resulting in an overall increase of 21% between pre-

and midtest. Although both student groups showed improvement, Group 1 achieved

significantly higher average test scores after completing EvolvingSTEM than Group 2

did after completing the standard curriculum (t=5.87, p<0.0001). Students who learned

evolution with EvolvingSTEM therefore achieved significantly greater gains in

comprehension of evolution than students who learned it from the standard curriculum.

Fig. 3. Boxplot of student assessment scores. The EvolvingSTEM curriculum produces significantly greater gains in comprehension of NGSS topic HS-LS-4 than the standard curriculum (Group 1 Post vs Group 2 Mid, t=5.87, p<0.0001). After experiencing our curriculum, Group 2 students subsequently achieved equivalent scores to Group 1 students (Group 1 Post vs Group 2 Post, t=0.14, ns). Mean values are indicated with diamonds.

Once students in Group 2 were exposed to EvolvingSTEM, their average

posttest scores had an overall increase of 20% in comparison to their midtest scores,

reaching knowledge gains made by Group 1 students (Fig. 3). Knowledge gains by both

Groups were overwhelmingly attributable to increased scores on the free-response

section of the assessment. Average free-response scores from pretests to posttests

increased by 18.09 points (48%) for Group 1 students and 20.59 points (54%) for Group

2 students. In comparison, average multiple-choice scores increased by 1.07 points for

Group 1 students and decreased by 0.54 points for Group 2 students. These results

may indicate that EvolvingSTEM has a greater impact on improving students' higher-

order cognitive skills, such as applying knowledge to an unknown problem and

performing data analysis. There was no significant difference between Group 1 and 2

posttest scores ($t=0.14$, $p=$n.s.), even though Group 2 students were provided more

detailed verbal instruction and took one additional assessment. This result speaks to the

power of EvolvingSTEM to increase student knowledge and suggests that our

curriculum can serve to replace, rather than supplement, the standard evolution

curriculum.

**Discussion**

We developed an inquiry-based, microbiology curriculum to improve the

engagement of high school biology students with topics central to evolutionary biology

and their subsequent understanding of related NGSS concepts. We observed high

levels of engagement when students participated in our curriculum. Students

recognized that success with the microbiology experiments required preparation, so

they almost invariably completed their homework assignments. Teachers indicated that

students who rarely participate in class-based discussions emerged as enthusiastic

group leaders while performing the EvolvingSTEM experiment. Informal post-surveys of

student attitudes towards the curriculum were overwhelmingly positive. Students

indicated that they were enthusiastic about the bacterial model, enjoyed coming to class

to work on the experiment, and felt that our curriculum was better at teaching them than

the standard lecture-style class. The group format for the experiments and analyses

encouraged the students to collaborate and support one another throughout the

program. Students tended to hold one another accountable, but also demonstrated

cohesion when groups compared their replicate populations, demonstrating both

friendly competition and pride and ownership in their results. Further, many students

expressed that they felt like "real scientists" using equipment like pipettes, vortexes, and

the incubator. They shared a greater sense of what science was actually like and asked

more questions about microbiology and evolution research and other scientific careers.

Crucially, teachers found EvolvingSTEM to be effective at demonstrating

evolution in action, thereby increasing student understanding of natural selection,

mutation, and the effects of chance, and increasing student interest and engagement

with biology. Student assessments also demonstrated the substantial benefit of our

curriculum to student learning, and consequently, our curriculum replaced the standard

WHS evolution curriculum in subsequent years. The sustainability of the EvolvingSTEM

curriculum has been greatly facilitated by the involvement of returning students who

demonstrated particular interest in the program and who served as *de facto* teaching

assistants through an Extended Learning Opportunity program. (More information about

this program will be the subject of a future report.) This teaching experience was made

possible by engaging first-year students in laboratory research, which allowed them to

help teach new students for up to three subsequent years prior to graduating.

We found that EvolvingSTEM provided students with significant learning benefits

in comparison to standard curricula. After completing our curriculum, students achieved

significantly higher scores on an knowledge assessment of evolution than students who

had followed the standard curriculum. After completing our curriculum, students who

were originally provided only the standard curriculum were able to further increase their

assessment scores to meet the gains made by students who were taught evolution only

with EvolvingSTEM. Our results demonstrate the power of microbial evolution

experiments to effectively teach concepts in population genetics and evolution while

also providing valuable experience in microbiology. Furthermore, EvolvingSTEM can

serve as an instructional foundation of other life science topics. For example, further

investigations by students could identify the genetic mutations (using inexpensive

whole-genome sequencing, i.e. (Cooper 2018)) that underlie the adaptive mutant

phenotypes, supporting a greater understanding of inheritance and trait variation (NGSS

HS-LS3). Previous research in our lab indicates that many commonly identified

mutations are found in the *wsp* (wrinkly spreader phenotype) gene cluster (Cooper et al.

2014; Gloag et al. 2018), which coordinates bacterial surface recognition with increased

biofilm production (Hickman et al. 2005). Students are likely to identify *wsp* mutants in

their classroom experiments and can therefore connect how changes in DNA can result

in changes in protein structure and intracellular signaling that lead to increased biofilm

production and changes to colony morphology, supporting a greater understanding of

DNA, protein structure, and cellular function (NGSS HS-LS1). Furthermore, the bacterial

adaptations are in response to environmental changes that provide new niches,

supporting a greater understanding of interdependent relationships in ecosystems

(NGSS HS-LS2). Classroom experiments that build upon the core evolution study can

therefore span much of the NGSS-recommended introductory biology curriculum and

also be readily adapted to cover more advanced topics for Advanced Placement

Biology.

**Summary**

EvolvingSTEM is an engaging, inquiry-based curriculum that provides students

with a hands-on approach to visualize evolutionary change occur in real time. It also can

be delivered at a low cost per student (<$5 in consumables) and is therefore potentially

suitable for broad distribution. As evidence, it is beinng offered in 10 public and private

high schools in 2019. Our curriculum provides students with the tools to understand

evolutionary concepts and apply their knowledge to other areas of life science and

medicine. For example, students can make a direct link between the adaptive

phenotypes they see in the classroom for increased biofilm production and the nearly

identical phenotypes seen in clinically relevant biofilm-associated bacterial infections. In

addition, students are provided an introduction to microbiological techniques that have

important applications for biotechnology. A particularly powerful aspect of our curriculum

is its positive effect on teacher and student engagement. Teachers and students

embark on the research experiment together, which provides a collaborative classroom

environment where both have the opportunity for greater understanding and discovery.

EvolvingSTEM has exceptional ability to improve scientific literacy and the promise of

promoting broad acceptance of evolution as a central, unifying theory for life science.

**Acknowledgements**

We thank Stephen Hale from the University of New Hampshire for assistance in

designing the assessment and Caroline Turner for critical feedback. This research was

supported by NSF CAREER DEB-0845851, NASA CAN-7NNA15BB04A, and

discretionary funds from the University of Pittsburgh, School of Medicine to VSC.

## References

Alizon S, Méthot P-O. Reconciling Pasteur and Darwin to control infectious diseases. Read A, editor. Plos Biol. 2018 Jan 18;16(1):e2003815–12.

Ashish A, Paterson S, Mowat E, Fothergill JL, Walshaw MJ, Winstanley C. Extensive diversification is a common feature of Pseudomonas aeruginosa populations during respiratory infections in cystic fibrosis. Journal of Cystic Fibrosis. European Cystic Fibrosis Society; 2013 Dec 1;12(6):790–3.

Beardsley PM, Stuhlsatz MAM, Kruse RA, Eckstrand IA, Gordon SD, Odenwald WF. Evolution and Medicine: An Inquiry-Based High School Curriculum Supplement. Evolution: Education and Outreach. 2nd ed. Springer; 2011 Oct 8;4(4):603–12.

Berendonk TU, Manaia CM, Merlin C, Fatta-Kassinos D, Cytryn E, Walsh F, et al. Tackling antibiotic resistance: the environmental framework. Nature Publishing Group. Nature Publishing Group; 2015 Mar 30;13(5):310–7.

Blount ZD, Lenski RE, Losos JB. Contingency and determinism in evolution: Replaying life's tape. Science. 2018 Nov 8;362(6415):eaam5979–12.

Broder ED, Angeloni LM, Simmons S, Warren S, Knudson KD, Ghalambor CK. Authentic Science with Live Organisms Can Improve Evolution Education. The American Biology Teacher. 2018 Jan 29;80(2):116–23.

Cooper VS. Experimental Evolution as a High-Throughput Screen for Genetic Adaptations. Gales AC, editor. mSphere. 2018 Jun 27;3(3):45–7.

Cooper VS, Staples RK, Traverse CC, Ellis CN. Parallel evolution of small colony variants in Burkholderia cenocepacia biofilms. Genomics. The Authors; 2014 Dec 1;104(Part A):447–52.

Costerton JW, Cheng KJ, Geesey GG, Ladd TI, Nickel JC, Dasgupta M, et al. Bacterial biofilms in nature and disease. Annual Review of Microbiology, Vol 64, 2010. 1987;41:435–64.

Crowe A, Dirks C, Wenderoth MP. Biology in Bloom: Implementing Bloom's Taxonomy to Enhance Student Learning in Biology. Sundberg M, editor. LSE. 2008 Dec;7(4):368–81.

Cunningham DL, Wescott DJ. Still More "Fancy" and "Myth" than "Fact" in Students' Conceptions of Evolution. Evolution: Education and Outreach. Springer; 2009 Sep 1;2(3):505–17.

Flynn KM, Dowell G, Johnson TM, Koestler BJ, Waters CM, Cooper VS. Evolution of Ecological Diversity in Biofilms of Pseudomonas aeruginosa by Altered Cyclic Diguanylate Signaling. O'Toole GA, editor. J Bacteriol. 2016 Sep 9;198(19):2608–

18.

Freeman S, Eddy SL, McDonough M, Smith MK, Okoroafor N, Jordt H, et al. Active
learning increases student performance in science, engineering, and mathematics.
Proc Natl Acad Sci USA. 2014 Jun 10;111(23):8410–5.

Glaze AL, Goldston MJ. US science teaching and learning of evolution: A critical review
of the literature 2000–2014. Sci Ed. 2015;99(3):500–18.

Gloag ES, Marshall C, Snyder D, Lewin GR, Harris JS, Chaney SB, et al. The
Pseudomonas aeruginosa Wsp pathway undergoes positive evolutionary selection
during chronic infection. bioRxiv. 2018.

Greaves M, Maley CC. Clonal evolution in cancer. Nature. 2012 Jan 19;481(7381):306–
13.

Green JH, Koza A, Moshynets O, Pajor R, Ritchie MR, Spiers AJ. Evolution in a test
tube: rise of the Wrinkly Spreaders. Journal of Biological Education. 2011 Jan
20;45(1):54–9.

Gregory TR. Understanding Natural Selection: Essential Concepts and Common
Misconceptions. Evolution: Education and Outreach. 2nd ed. 2009 Apr 9;2(2):156–
75.

Harrison JJ, Turner RJ, Ceri H. Persister cells, the biofilm matrix and tolerance to metal
cations in biofilm and planktonic Pseudomonas aeruginosa. Environ Microbiol. 2005
Jul;7(7):981–94.

Hickman JW, Tifrea, Harwood. A chemosensory system that regulates biofilm formation
through modulation of cyclic diguanylate levels. PNAS. 2005.

Infanti LM, Wiles JR. " Evo in the News:" Understanding Evolution and Students'
Attitudes toward the Relevance of Evolutionary Biology. Bioscene. 2014.

Jordan TC, Burnett SH, Carson S, Caruso SM, Clase K, DeJong RJ, et al. A Broadly
Implementable Research Course in Phage Discovery and Genomics for First-Year
Undergraduate Students. MBio. 2014;5(1):493–8.

Karatan E, Watnick P. Signals, Regulatory Networks, and Materials That Build and
Break Bacterial Biofilms. Microbiol Mol Biol Rev. 2009 Jun 1;73(2):310–47.

Knott GJ, Doudna JA. CRISPR-Cas guides the future of genetic engineering. Science.
2018.

Makarova KS, Wolf YI, Alkhnbashi OS, Costa F, Shah SA, Saunders SJ, et al. An
updated evolutionary classification of CRISPR–Cas systems. Nature Publishing
Group. Nature Publishing Group; 2015 Sep 28;13(11):722–36.

Makohon-Moore A, Iacobuzio-Donahue CA. Pancreatic cancer biology and genetics
from an evolutionary perspective. Nature Publishing Group. Nature Publishing
Group; 2016 Jul 22;16(9):553–65.

Martin M, Hölscher T, Dragoš A, Cooper VS, Kovács ÁT. Laboratory Evolution of
Microbial Interactions in Bacterial Biofilms. O'Toole GA, editor. J Bacteriol. 2016 Sep
9;198(19):2564–71.

National Research Council. A framework for K-12 science education: Practices,
crosscutting concepts, and core ideas. Washington, DC: The National Academies
Press; 2012.

Nehm RH, Reilly L. Biology Majors' Knowledge and Misconceptions of Natural
Selection. BioScience. Oxford University Press; 2007 Mar 1;57(3):263–72.

Nelson CE. Teaching evolution (and all of biology) more effectively: Strategies for
engagement, critical reasoning, and confronting misconceptions. Integrative and
Comparative Biology. 2008 Jun 21;48(2):213–25.

NGSS Lead States. Next generation science standards: For states, by states.
Washington, DC: The National Academies Press; 2013.

NSTA. NSTA Position Statement: The Teaching of Evolution. 2013 Jul 30;:1–7.

O'Rourke D, FitzGerald CE, Traverse CC, Cooper VS. There and back again:
consequences of biofilrn specialization under selection for dispersal. Front Genet.
Frontiers; 2015;6(e225).

Pobiner B. Accepting, understanding, teaching, and learning (human) evolution:
Obstacles and opportunities. Am J Phys Anthropol. 3rd ed. 2016 Jan
25;159(3):232–74.

Poltak SR, Cooper VS. Ecological succession in long-term experimentally evolved
biofilms produces synergistic communities. The ISME Journal. Nature Publishing
Group; 2011;5(3):369–78.

Rainey P, Buckling A, Kassen R, TRAVISANO M. The emergence and maintenance of
diversity: insights from experimental bacterial populations. Trends in Ecology &
Evolution. 2000 Jun;15(6):243–7.

Rainey PB, Travisano M. Adaptive radiation in a heterogeneous environment. Nature.
1998.

Ratcliff WC, Raney A, Westreich S, Cotner S. A novel laboratory activity for teaching
about the evolution of multicellularity. The American Biology Teacher.
2014;76(2):81–7.

Romine WL, Walter EM, Bosse E, Todd AN. Understanding patterns of evolution acceptance-A new implementation of the Measure of Acceptance of the Theory of Evolution (MATE) with Midwestern university students. J Res Sci Teach. 2nd ed. 2017 Jan 12;54(5):642–71.

Sickel AJ, Friedrichsen P. Examining the evolution education literature with a focus on teachers: major findings, goals for teacher preparation, and directions for future research. Evolution: Education and Outreach. 2nd ed. 2013 Jul 5;6(1):1105–15.

Smith MU. Current Status of Research in Teaching and Learning Evolution: I. Philosophical/Epistemological Issues. Sci & Educ. Springer Netherlands; 2010a;19(6-8):523–38.

Smith MU. Current Status of Research in Teaching and Learning Evolution: II. Pedagogical Issues. Sci & Educ. 2nd ed. 2010b;19(6-8):539–71.

Spiers AJ. The Pseudomonas fluorescens SBW25 wrinkly spreader biofilm requires attachment factor, cellulose fibre and LPS interactions to maintain strength and integrity. Microbiology. 2005 Sep 1;151(9):2829–39.

Spiers AJ. Getting Wrinkly Spreaders to demonstrate evolution in schools. Trends in Microbiology. Elsevier Ltd; 2014 Jun 1;22(6):301–3.

Starkey M, Hickman JH, Ma L, Zhang N, De Long S, Hinz A, et al. Pseudomonas aeruginosa rugose small-colony variants have adaptations that likely promote persistence in the cystic fibrosis lung. J Bacteriol. American Society for Microbiology Journals; 2009 Jun;191(11):3492–503.

Traverse CC, Mayo-Smith LM, Poltak SR, Cooper VS. Tangled bank of experimentally evolved Burkholderia biofilms reflects selection during chronic infections. Proc Natl Acad Sci USA. 2013;110(3):E250–9.

Turner, Marshall CW, Cooper VS. Parallel genetic adaptation across environments differing in mode of growth or resource availability. Evolution Letters. Wiley-Blackwell; 2018 Aug 4;2(4):355–67.

Wells J, Nesse RM, Sear R, Johnstone RA, Stearns SC. Evolutionary public health: introducing the concept. The Lancet. 2017.

Wiggins G, McTighe. Understanding by Design (Expanded 2nd Ed.) Alexandria: VA: ASCD. 2005.

Yates TB, Marek EA. Teachers teaching misconceptions: a study of factors contributing to high school biology students' acquisition of biological evolution-related misconceptions. Evolution: Education and Outreach. 2nd ed. 2014 Mar 15;7(1):1–18.

550     Zoller U. Are lecture and learning compatible? Maybe for LOCS: Unlikely for HOCS. J
551        Chem Educ. 1993 Mar;70(3):195–3.

552