Expanding the watch list for potential Ebola virus antibody escape mutations

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Abstract

The 2014 outbreak of Ebola virus (EBOV) in Western Africa is the largest recorded filovirus disease outbreak and lead to the death of over 11,000 people. This deadly virus still poses a grave epidemic threat as evidenced by the current (since May 2018) EBOV outbreak in the Democratic Republic of the Congo which has already claimed the lives of over 250 people. One important strategy for combating EBOV epidemics is to anticipate how the evolution of EBOV might undermine treatment since the development of vaccines and antibody therapies are typically based on a single strain (often the 1976 Mayinga) of the EBOV envelope glycoprotein (GP). In a previous study we initiated a watch list of potential antibody escape mutations of EBOV by modeling interactions between EBOV GP and the monoclonal antibody KZ52. This watch list was generated using molecular modeling to estimate the effect of every possible mutation of GP. The final watch list containing 34 mutations were predicted to disrupt GP-KZ52 binding but not to disrupt the ability of GP to fold and to form trimers. In this study, we expand our watch list by including three more monoclonal antibodies with distinct epitopes on GP, namely Antibody 100 (Ab100), Antibody 114 (Ab114) and 13F6-1-2. Our updated watch list contains 127 mutations, three of which have been seen in humans or are experimentally associated with reduced efficacy of antibody treatment. We believe mutations on this broad watch list require attention since they may be a signal of an evolutionary response from EBOV to treatment that could diminish the effectiveness of interventions.

Introduction

Ebola virus (EBOV) has five known species: Zaire EBOV, Sudan EBOV, Ta Forest EBOV, Bundibugyo EBOV, and Reston EBOV and can cause disease with up to 90% case fatality rates.[1, 2] The outbreak of Zaire EBOV infection in Western Africa between 2014 to 2016 is the largest recorded filovirus disease outbreak with death typically occurring 5-7 days after the appearance of symptoms.[3] This outbreak receded in 2016, but EBOV still constitutes an epidemic threat as evidenced by the current (since May 2018) Zaire EBOV outbreak in the Democratic Republic of the Congo which has already claimed lives of 271 people and more than 458 positive cases (WHO report, 4th December 2018).[4]

The 2014–2016 EBOV outbreak in West Africa led to clinical development of therapeutics and vaccines.[5, 6] The success of ZMapp, a cocktail of three chimeric monoclonal antibodies (mAbs) derived from immunized mice, in nonhuman primates (NHP) demonstrated the potential of mAb therapies against EBOV infection, and ZMapp is currently undergoing human trials.[7-9] Subsequently, several other mAbs were designed and demonstrated protection against EBOV infection and some of them are currently being used to manage the current EBOV outbreak. (https://www.nih.gov/news-events/news-releases/nih-begins-testing-ebola-treatment-early-stage-trial)[10, 11] To prepare for future outbreaks it is critical to anticipate and monitor EBOV evolution since it could lead to antibody escape mutants that could compromise treatment efforts. Sequencing studies have revealed that there is a significant genetic variation in EBOV. Most of the variation is found in the EBOV glycoprotein (GP) that is the target for the all mAbs currently under development and being used in the management of the current EBOV outbreak.[12-14]

To address the potential threat of EBOV evolution outpacing antibody treatment efforts we

previously initiated a watch list of potential antibody escape mutants for the EBOV GP.[15] We

focused on the KZ52 mAb as it had a 3-D experimental structure bound to EBOV GP available.

In this previous study, we combined use of FoldX software with molecular dynamics simulations

to estimate folding and binding stabilities. We placed 34 mutations on the watch list by

considering every possible EBOV GP mutation and choosing those that disrupt binding between

GP and KZ52 but do not disrupt the ability of GP to fold and bind to form a complex. One of

these 34 mutations (*N550K*) was seen in humans in a previous outbreak.

The aim of this study is to expand our watch list of potential antibody escape mutants for EBOV

GP. After we published our previous watch list, three more 3-D structures of mAbs interacting

with different epitopes on EBOV GP (necessary for our molecular modeling approach) were

published in the Protein Data Bank (PDB). In this study, we expand our watch list to include

possible antibody escape mutations from three antibodies with distinct epitopes: Antibody 100

(Ab100), Antibody 114 (Ab114) and 13F6-1-2. Our watch list now contains 127 mutations, three

of which have been seen in humans or are experimentally associated with reduced efficacy of

antibody treatment.

Methods

For a mutation to be placed on a watch list for EBOV it must: (1) disrupt binding to a protective

antibody, and (2) leave the viral proteins functional. It is thus necessary to determine how amino

acid mutations alter stabilities ($\Delta\Delta G$ values) for GP folding, forming a trimer and binding to the

antibody. In our previous work, we have obtained $\Delta\Delta G$ values of GP folding, GP trimer

formation and GP binding to KZ52 antibody using our molecular dynamics (MD) plus FoldX

approach.[15] In this study we calculate $\Delta\Delta G$ values of binding for Ab100, Ab114 and 13F6-1-2

antigen-antibody complexes using the same modeling approach.

Structure Preparation

EBOV GP – Ab100 and Ab114 complexes

Structures of Ab100 and Ab114 bound to EBOV GP were obtained from the Protein Data Bank

(PDB) accession number 5FHC.[16] The EBOV GP amino acid sequence was based on the 1976

Mayinga strain, just like our previous study. The PDB file 5hfc.pdb contains coordinates of both

Ab100 and Ab114 bound to EBOV GP (GP1 & GP2) monomer at different sites. This file was

first modified to remove all but one copy each of GP1, GP2, antibody light chain and antibody

heavy chain of Ab100 and Ab114 (one third of the GP-Ab100/Ab114 trimeric complex), and

then was split into two files where one file had the GP – Ab100 complex and the other had the

GP – Ab114 complex. The MODELLER software[17] was then used to build the missing

residues. This included residues 190 – 213 that are predicted to be intrinsically disordered; the

resulting complexes had no secondary structure content in this region. The full EBOV trimer

protein complex was then created using the symexp command in PyMOL (see Figs 1A and 2A)

and contained three copies each of GP1 (residues 33 - 278), GP2 (residues 502 – 599), heavy and

light chains of the antibody.

Fig 1. (A) Structure of EBOV GP trimer in complex with the Ab100 antibody. GP1 is gray, GP2

is red and the heavy and the light chains of Ab100 are brown. (B) & (C) Mutations in the GP

complex with $\Delta\Delta G_{\text{Bind}} > 2$ kcal/mol (i.e., above black dashed line) are considered disruptive and

are highlighted using green stick representation. (D) $\Delta\Delta G_{\text{Bind}}$ values (gray circles) for all 19

possible mutations at each site of GP1 (33 – 278) and GP2 (502 – 599). Different colors and

counts in the legend indicate locations and number of mutations with $\Delta\Delta G_{\mathrm{Bind}}$ > 2 kcal/mol on

the GP complex.

Fig 2. (A) Structure of EBOV GP trimer in complex with the Ab114 antibody. GP1 is gray, GP2

is yellow and the heavy and the light chains of Ab114 are orange. (B) Mutations in the GP

complex with $\Delta\Delta G_{\mathrm{Bind}}$ > 2 kcal/mol (i.e., above black dashed line) are considered disruptive and

are highlighted using green stick representation. (D) $\Delta\Delta G_{\mathrm{Bind}}$ values (gray circles) for all 19

possible mutations at each site of GP1 (33 – 278) and GP2 (502 – 599). Different colors and

counts in the legend indicate locations and number of mutations with $\Delta\Delta G_{\mathrm{Bind}}$ > 2 kcal/mol on

the GP complex.

EBOV mucin-like domain peptide – 13F6-1-2 antibody complex
Crystal structure of the 13F6-1-2 Fab fragment bound to its EBOV GP mucin-like domain (GP

MLD) peptide epitope (11 amino acids, VEQHHRRTDND) was downloaded from the PDB

using the 2QHR accession number.[18] Unlike other epitopes used in our previous and current

study, this structure was based on the 1976 Eckron strain. However, the alignment of the 11-

residue long peptide was 100% identical to the 1976 Mayinga strain. The PDB file 2qhr.pdb was

edited to remove everything except GP MLD peptide (residues 404 – 414) and heavy and light

chains of 13F6-1-2 antibody. (see Fig 3A) WHAT IF web server (https://swift.cmbi.umcn.nl)

was then used to add the missing atoms to the complex structure.

Fig 3. (A) Structure of EBOV GP MLD peptide bound to the 13F6-1-2 antibody. GP MLD

peptide is in gray tube representation, and the heavy and the light chains of 13F6-1-2 are black.

Mutations in the GP MLD peptide with $\Delta\Delta G_{\mathrm{Bind}}$ > 2 kcal/mol (i.e., above black dashed line) are

considered disruptive and are highlighted using green stick representation. (B) $\Delta\Delta G_{Bind}$ values

(gray circles) for all 19 possible mutations at each of the 11 sites of GP MLD peptide (404 –

414). Different colors and counts in the legend indicate locations and number of mutations with

$\Delta\Delta G_{Bind}$ >2 kcal/mol on the GP complex.

Molecular Dynamics Simulations
All three EBOV – antibody complexes were subjected to atomistic MD simulations using the

protocol reported in our previous study. Briefly, the software package GROMACS 5.0.7[19] was

used for all MD simulations with the Charmm22* forcefield.[20] GP MLD peptide – 13F6-1-2

simulations were 100 ns, and the GP – Ab100 and Ab114 production simulations were run for a

shorter 50 ns due to the large number of atoms in the final simulation box. During the production

simulation snapshots were saved every 1 ns resulting in 50 snapshots for each of the GP – Ab100

and GP – Ab114 systems, and 100 snapshots for the peptide – 13F6-1-2 complex.

FoldX
Snapshots of all three complexes were analyzed using FoldX software.[21, 22] As with our

previous study, we initially minimize each snapshot six times in succession using the RepairPDB

command to obtain convergence of the potential energy. BuildModel command was then used to

generate all possible 19 single mutations in EBOV GP and GP MLD peptide at each amino acid

site. Finally, the binding stability of the protein complex due to each mutation was estimated

using AnalyseComplex command. For each mutation, we then estimated $\Delta\Delta G_{Bind}$ by averaging

the FoldX results across all individual snapshot estimates.

To calculate $\Delta\Delta G_{Bind}$ values for all possible 19 mutations at amino acid site of EBOV GP and GP

MLD peptide, we carried out 980,400 FoldX calculations (344 GP residues $\times$ 19 possible

mutations at each site × 50 MD snapshots × 3 copies of GP-Antibody in a complex) for each GP

– Ab100 and GP – Ab114 complexes and 20,900 calculations (11-residue MLD peptide × 19

possible mutations × 100 MD snapshots) for GP MLD peptide – 13F6-1-2 antibody complex.

Averaging estimates across all individual snapshots ultimately resulted in 6,441 $\Delta\Delta G_{Bind}$ values

each for Ab100 and Ab114 antibody complexes and 209 $\Delta\Delta G_{Bind}$ values for 13F6-1-2 antibody

complex. (see SI Files)

166 **Results and discussion**

We have expanded the watch list generated by us in a previous study by including antibody

escape mutations against three additional antibodies interacting with EBOV GP. These watch list

mutations are those predicted to both disrupt GP - antibody binding and yet allow GP to fold and

form trimers.

The EBOV GP is a class I fusion protein consisting of disulfide-linked subunits, GP1 and GP2,

that bind to form a chalice-shaped trimer. (see Figs 1A & 2A) Ab100 interacts at the base of the

GP trimer (see Fig 1A). This interaction is similar to that of KZ52, the prototypic neutralizing

antibody used in our previous study. However, Ab100 contacts GP1 and GP2 of a monomer and

the disordered (residue 190 -213) loop of the neighboring monomer (see Fig 1A) in contrast to

KZ52 which interacts with GP2 of a single monomer. The epitope for Ab114 spans both the

glycan cap and the inner chalice of GP, (see Fig 2A) where it remains bound after proteolytic

cleavage of the glycan cap and prevents interaction of cleaved GP to its host receptor.[16] 13F6-

1-2 is a monoclonal antibody that binds to an 11-residue peptide located in the heavily

glycosylated mucin-like domain (MLD) of the EBOV GP.[18] (see Fig 3A)

Fig 4 is a graphical representation of our expanded watch list of EBOV mutations which includes

KZ52, Ab100, Ab114 and 13F6-1-2 antibody complexes. The maximum functional stability

change (i.e., folding and trimer formation) for all mutations is plotted against the corresponding

change in the antibody binding affinity. The 127 mutations in the lower right quadrant are those

that belong on the watch list since they are classified as both functional and disruptive to

antibody binding. There were 21, 33, 39 and 34 mutations respectively in Ab100, Ab114, 13F6-

1-2 and KZ52 antibody complexes. The specific mutations on the watch list are given in the

Table 1 and shows that they are concentrated at just six residues in KZ52 and at five residues in

each of the other three antibody complexes. (see Figs 1B & 1C, Fig 2B and Fig 3A) All of these

mutations lie at the interface as one might intuitively expect from the structure, however, most

mutations at the interface are not predicted to disrupt antibody binding.

Fig 4. Maximum of folding stability, dimer binding stability (interaction of GP1 and GP2) or

trimer binding stability (interaction of a GP1-GP2 dimer with other dimers), $\Delta\Delta G_{Max}$, as a

function of $\Delta\Delta G_{Bind}$ for all antibody complexes. $\Delta\Delta G_{Max}$ values are considered to be zero for the

intrinsically disordered 11-residue MLD peptide. Symbols in the inset legend indicate the

corresponding antibody. Watch list mutations are shown as colored symbols and are predicted to

disrupt binding to any one of the four antibodies (KZ52, Ab100, Ab114 and 13F6-1-2) but not to

disrupt GP folding and trimer formation. Consistent with our previous study, mutations with

$\Delta\Delta G_{Bind} > 2$ kcal/mol are considered disruptive to antibody binding and those with $-3 < \Delta\Delta G_{Max}$

< 3 kcal/mol are considered functional. The number of watch list mutations associated with each

antibody is shown in the legend.

Table 1. Watch list mutations

Systems	Amino acid site	Predicted Escape Mutations
EBOV GP – KZ52	N506	W, Y
	P513	H, W
	N550	Q, K, P, F, H, I, E, R, W, V, Y, M
	P552	S, Q, K, T, F, A, H, G, R, W, V, Y
	G553	M
	G557	F, H, R, W, Y
EBOV GP – Ab100	P34	W, Y
	D47	R
	P198	F, W, Y, D, H
	E564	K, P, S, T, Y, A, R, G
	A568	F, W, Y, R, H
EBOV GP – Ab114	P116	F, W, Y, R, D, G, H
	G118	L, K, M, F, S, W, Y, R, N, C, Q, E, H
	T144	F, P, W, Y, D, E, H
	T223	F, W, Y, R, H
	L233	W
GP MLD peptide – 13F6-1-2	Q406	R, D, G, H, F, S, T, W, Y
	H407	P
	R409	A, N, D, C, Q, E, G, H, I, L, K, M, F, P, S, T, W, Y, V
	R410	G, W
	R411	R, Q, G, H, K, F, W, Y

Table 1. Watch list mutations are shown for all four antibodies (KZ52, Ab100, Ab114 and 13F6-1-2) at a predicted amino acid sites on EBOV GP.

If any of the mutations in Table 1 were to appear in a real population, it would indicate a possible increased risk of escaping the normal immune response or render treatments with these antibodies ineffective. Mutation *N550K* against KZ52 has already appeared in humans and thought to have been infected by gorillas in Central Africa between 2001 and 2003. This mutation is present in all sequenced isolates from that outbreak.

Davidson et al.[23] conducted an alanine-scanning mutagenesis study on GP that can be

qualitatively compared to our work. They individually mutated each residue of the GP protein to

alanine and measured changes in GP-KZ52 binding affinities relative to the unmutated form.

They identified five residues that are critical for KZ52 antibody binding: C511, N550, D552,

G553, and C556. Three of these sites are found on our watch list in Table 1 (N550, D552, and

G553) and 25 of the 34 (74%) watch list mutations are found at these three sites.

In the study of MB-003,[24] a plant-derived monoclonal antibody cocktail composed of c13C6,

13F6-1-2, and c6D8 used effectively in treatment of EBOV virus infection in non-human

primates, was unable to protect two of six animals when initiated one- or two-days post-

infection. Investigation of a mechanism of viral escape in one of the animals shown five

nonsynonymous mutations in the monoclonal antibody target sites. Among these mutations

Q406R and *R409C* were linked to a reduction in 13F6-1-2 antibody binding.[24] Both of these

mutations were correctly identified by our modeling strategy and are present on the watch list.

In spite of our efforts to expand the watch list, it remains incomplete and putative for several

reasons. The list covers only four epitopes for which experimental structures of antibodies

interacting with viral proteins are available. However, there are other epitopes known for EBOV

GP.[10] With more experimental structures it would be possible to further expand the watch list.

The size of the watch list depends on how we define the threshold for antibody binding (Fig 4).

If the threshold is lowered from 2.0 kcal/mol, the watch list will naturally grow in size. The

reasoning behind using a different cutoff for functional as compared to antibody binding is

described in our previous study. The watch list only includes mutations that are individually

predicted to disrupt antibody binding while remaining functional, however, it is possible that

immune escape could arise by the cumulative effect of several changes on antibody binding.

How multiple substitutions interact to produce cumulative effects on stability is not well

understood and is an important consideration for future studies. The watch list has not been

experimentally validated either in terms of mutational effects on GP folding and binding

affinities, nor on the downstream immune system consequences. Our hope is that this work will

encourage such research.

In summary, we have expanded the watch list of potential antibody escape mutations of EBOV

by including three more antibody complexes: EBOV GP – Ab100, GP – Ab114 and GP Mucin-

like domain (GP MLD) peptide – 13F6-1-2. The watch list now contains 127 mutations in 21

sites in EBOV GP. Mutations from the watch list that appear during an outbreak deserve

attention since they may be a signal of an evolutionary response from the virus that could reduce

the efficacy of treatment efforts. Ab114 has been recently approved for the first time to treat

infected individuals during the current EBOV outbreak in Democratic Republic of Congo.

(https://www.nih.gov/news-events/news-releases/nih-begins-testing-ebola-treatment-early-stage-

trial) We hope our watch list will serve as a useful reference for the public health and emerging

infectious disease monitoring agencies. The watch list can still be expanded if more experimental

structures of EBOV – antibody complexes become available. In fact, as we were preparing this

manuscript, the crystal structure of mAb CA45 bound to GP1 and GP2 interface of EBOV GP

were published by Janus et al.[25] Lastly, we believe our in-silico approach could be applied to

determine watch lists for other viruses provided experimental structures are available and for

future design and optimization efforts of antibodies.

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Supporting information

S1 File. S1_Ab100_data.xlsx. Excel spreadsheet with estimated stability effects of all 6,441
mutations against Ab100.

S2 File. S2_Ab114_data.xlsx. Excel spreadsheet with estimated stability effects of all 6,441
mutations against Ab114.

S3 File. S3_13F6-1-2_data.xlsx. Excel spreadsheet with estimated stability effects of all 209
mutations against 13F6-1-2.

S4 File. S4_expanded_watchlist.xlsx. Excel spreadsheet with estimated binding stability effects
of watch list mutations against all the four antibodies (KZ52, Ab100, Ab114, and 13F6-1-2).