

# Single-neuron representations of spatial memory targets in humans

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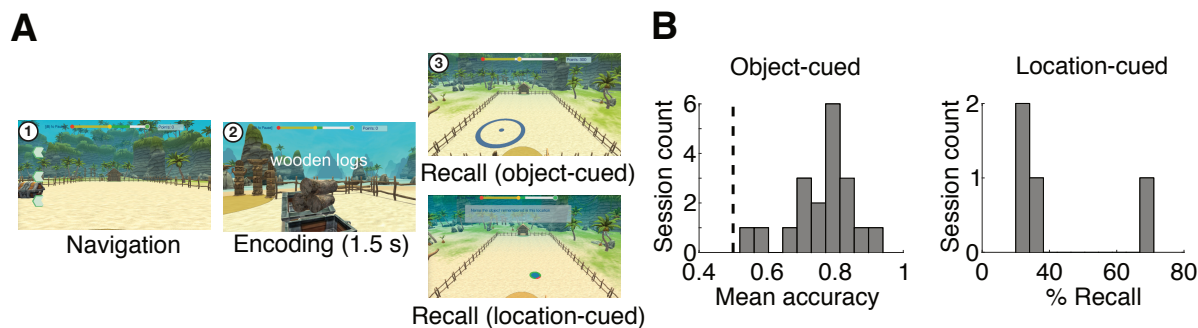
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## 1 Summary

2 The hippocampus and surrounding medial-temporal-lobe (MTL) structures are critical for both memory  
3 and spatial navigation (Scoville and Milner, 1957; Morris et al., 1982). Much research has focused  
4 on neurons that activate according to an animal's own spatial properties, such as "place cells" that  
5 represent the current location (O'Keefe and Dostrovsky, 1971), or "head-direction cells" that code  
6 for the current heading (Taube et al., 1990). In addition to representing the current spatial setting,  
7 these same MTL structures are important for other behaviors such as memory, which can involve  
8 remote locations among other contextual information (Schiller et al., 2015; Epstein et al., 2017).  
9 However, the human cellular representations that underlie our ability to form memories that involve  
10 remote locations are unclear. We recorded single-neuron activity from neurosurgical patients playing  
11 Treasure Hunt (TH), a virtual-reality object–location memory task. We found that the activity of  
12 many MTL neurons was significantly modulated by the position of the to-be-remembered object. In  
13 addition, we observed neurons whose firing rates during navigation were explained by the subject's  
14 heading direction, and others that predicted subsequent memory performance. By showing evidence  
15 for neurons encoding remote locations that are the targets of memory encoding, our results suggest  
16 that the human MTL represents to-be-remembered locations in service of memory formation.

## 17 Results

18 Humans have rich navigation and spatial memory skills, including the ability to not only accurately  
19 navigate through complex environments but also to imagine and remember events that occur at remote  
20 locations (Spiers and Maguire, 2007; Miller et al., 2013; Ekstrom, 2015; Vass et al., 2016). We  
21 hypothesized that the types of neural activity in the human MTL that represent one's current location  
22 during navigation are also involved in representing relevant remote locations during spatial cognition.  
23 To examine this issue directly, we asked neurosurgical patients with microelectrodes implanted in their  
24 MTL to play a virtual-reality spatial-memory task, and we examined how their neural responses related  
25 to their simultaneous movement and memory. The fifteen participants in our study played Treasure  
26 Hunt (TH), a video-game–like task that measures subjects' ability to remember the spatial locations  
27 where various objects are hidden (Miller et al., 2018). In each trial of the task, subjects explored a  
28 virtual beach and traveled to a series of treasure chests. Upon reaching each chest, it opened and an  
29 object appeared. The subject's goal was to encode the spatial location corresponding to the position



**Figure 1: Treasure Hunt (TH) task performance.** A. Screen images from the Navigation, Encoding, and Recall phases of TH. The two images for the recall phase correspond to different versions of the task (see Methods). B. Histogram showing mean performance on the recall phase of TH in each of the two task versions.

30 of each item. To characterize the neural basis of spatial memory, we analyzed the neural recordings  
31 to identify neurons that represented memory target locations, as well as the subject's current virtual  
32 location, heading direction, and subsequent memory performance.

33 **Behavior.** Before characterizing neural signatures of spatial memory encoding, we first examined  
34 subjects' behavior in TH (Figure 1a). In each trial subjects navigated to chests ("Navigation"), and  
35 upon reaching each chest viewed an object whose location they tried to encode ("Encoding"). Finally,  
36 in the "Recall" phase subjects were asked to recall the object locations. The subjects in our study  
37 performed one of two task versions, which differed in terms of the Recall phase. In the "object-cued"  
38 version, they viewed the image of a cued object and indicated its location by moving an on-screen  
39 cursor accordingly. In the "location-cued" version of the task they viewed a probed location and  
40 verbally responded by speaking the name of the corresponding object into a microphone.

41 We scored behavioral performance during the recall phase as a measure of subjects' object–location  
42 memory. Subjects showed good performance in both versions of the task (Figure 1b), indicating that  
43 they were successfully able to orient and remember locations in our virtual reality environment. Mean  
44 accuracy on all object-cued sessions was above chance (chance=0.5). The mean performance on  
45 location-cued sessions (43%) was consistent with levels seen in other paired-associate memory tasks  
46 that required verbal responses, such as in Greenberg et al. (2015) where subjects exhibited 40% recall  
47 rates.

48 **Neurons responsive to the current memory target.** We hypothesized that we would observe  
49 "memory target cells," which we defined as neurons whose activity was modulated by the location of  
50 the current, to-be-remembered chest as subjects drove to it during navigation. We were motivated  
51 by previous work in animal models showing MTL cells that represent salient remote locations (Rolls  
52 and O'Mara, 1995; Wilming et al., 2018; Wirth et al., 2017; Omer et al., 2018; Danjo et al., 2018;  
53 Gauthier and Tank, 2018), as well as related evidence from recordings of human theta oscillations (Lee  
54 et al., 2018) and fMRI (Brown et al., 2016). Therefore, we examined how neuronal firing rates during  
55 navigation varied according to the location of the current target chest. We identified single-neuron  
56 action potentials (Fried et al., 1999), and isolated a total of 131 putative MTL neurons across 23  
57 task sessions. Forty-five of these neurons were in the hippocampal formation (HF) and 86 were in the  
58 parahippocampal gyrus (PHG).

59 To identify these "memory target cells," we analyzed each cell's spiking activity as a function  
60 of the location of the upcoming chest, in addition to the subject's current position (Fig. 2a). We  
61 generated firing rate heatmaps both based on the location of the upcoming memory target, as well  
62 as the subject's own position. We identified neurons whose firing rates were significantly modulated  
63 using permutation tests.

64 We labeled neurons as memory-target cells if their firing rate significantly varied as function of  
65 the chest location. Figure 2b illustrates the activity of one example memory-target cell from the left  
66 entorhinal cortex (EC) of Patient 9. This neuron increased its firing rate when the subject navigated  
67 to chests that were located in the "south-central" part of the environment ( $p < 0.001$ ). Critically,  
68 while this cell's firing rate was modulated by the location of the upcoming object, it did not vary  
69 significantly according to the subject's own position (Figure 2b-right,  $p = 0.437$ ). Thus, this neuron's  
70 representation of the current navigational target constitutes a novel coding pattern that is distinct  
71 from the activity of conventional place cells, which generally represent an animal's own location.  
72 Figure 2c shows a second example of this phenomenon from a cell in Patient 12's right EC, which  
73 significantly increased its firing rate when the subject had memory targets in the "east" section of the

74 environment ( $p = 0.004$ ), and did not show a firing-rate modulation according to the subject's own  
75 position ( $p = 0.49$ ).

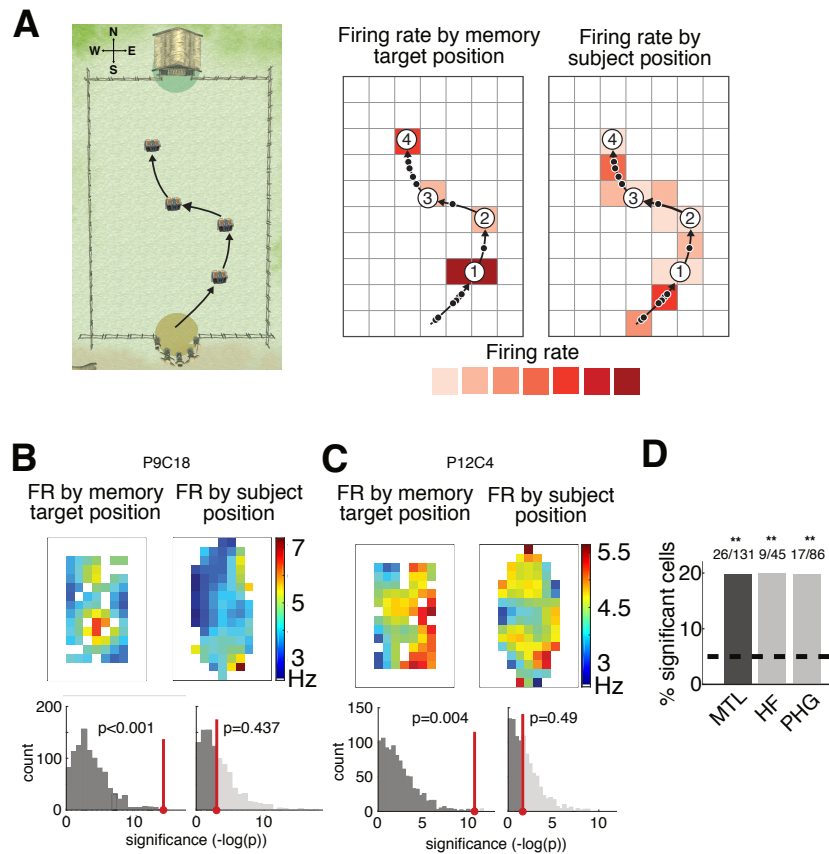
76 Across the population, 20% of MTL cells (26 of 131) had firing rates that were significantly  
77 modulated by memory target location, which is significantly greater than the 5% expected by chance  
78 ( $p < 0.01$ , binomial test). The number of these memory-target cells that we observed was significantly  
79 above chance both when measured separately for the HF and for the PHG regions (binomial tests  
80  $p$ 's  $< 0.01$ ). None of these cells' firing rates were modulated by the subject's virtual position. Thus,  
81 these results indicate that a substantial number of neurons throughout the human MTL specifically  
82 represent remote locations in a task when they are important behaviorally.

83 Given the large literature on place cells (Muller, 1996), we also characterized neurons whose firing  
84 rates were modulated by the subject's current position. Figure 3 shows three example cells that  
85 showed significant spatial modulation according to the subject's current virtual location. We found  
86 that during navigation some cells individually exhibited significant place coding, but at the population  
87 level the total number of observed place-like cells was not above chance. Across the population 8 of  
88 131 (6%) of MTL cells were significantly modulated by subject position (Fig. 3d) ( $p = 0.21$ , binomial  
89 test).

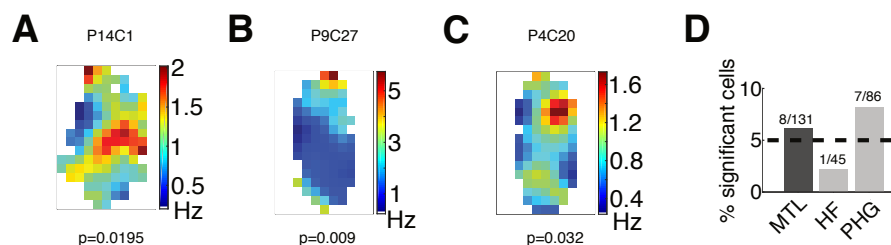
90 **Neurons responsive to heading direction.** In rodents there is evidence for neurons whose firing  
91 rates are modulated by the direction of the animal's head during movement (Taube et al., 1990;  
92 Robertson et al., 1999). These head-direction cells were first discovered in the dorsal presubiculum  
93 and are also commonly found in the anterodorsal thalamus, but have also been found in areas of the  
94 MTL such as the entorhinal cortex (Sargolini et al., 2006). These cells have not previously been found  
95 in humans, but the representation of heading direction could play a role in forming memories. As such,  
96 we tested for the existence of "heading-direction" cells in our dataset, which we defined as neurons  
97 that varied their firing rate according to the direction that subjects moved in the virtual environment,  
98 grouped into four 90° bins, which we refer to as north, south, east, and west. Figure 4a–d illustrates  
99 the activity of four significant heading-direction cells. As these examples illustrate, the heading that  
100 elicited peak firing activity varied across individual heading-direction cells. In addition, some cells  
101 showed increased firing rates at multiple distinct headings (Figure 4c–d), similar to "bidirectional  
102 cells" recently observed in rodents (Jacob et al., 2017). We did not find that any particular preferred  
103 angle was dominant across the population of heading-direction cells (Figure 4e).

104 Overall, 12% of MTL cells (16 of 131) were heading-direction cells, showing significant changes  
105 in firing rate according to the virtual heading, which is more than expected by chance ( $p < 0.0003$ ,  
106 binomial test). There were significant proportions of heading-direction cells in both the HF and PHG  
107 ( $p = 0.007$  and  $p = 0.004$ , respectively; Figure 2f). No heading-direction cells showed firing rates that  
108 varied with the subject's current position; two heading-direction cells showed effects of memory target  
109 position. Because the population of cells modulated by heading direction is largely nonoverlapping  
110 from those that were significantly modulated by memory target position, it suggests that our finding  
111 of memory-target cells is not explained by direction-related modulations.

112 **Neurons modulated by subsequent memory performance.** We next tested whether each cell's  
113 firing rate during navigation significantly varied as a function of whether or not memory targets were  
114 subsequently remembered (Miller et al., 2018). Figure 5a–b shows two example "memory cells" whose  
115 firing rates during navigation significantly varied according to whether or not the subject subsequently  
116 recalled the correct location of the item in the current chest. As illustrated by these examples, individual  
117 cells showed either significant increases or decreases in their firing rate according to subsequent memory  
118 performance.



**Figure 2: Neural activity related to memory target position.** *A. Analysis framework for binning navigation period neuronal data by subject position and memory target location, shown for an example trial. Left, overhead view of TH environment with example paths to 4 chests (only one chest is visible at a given time). Middle, example path spikes binned by memory target location to calculate firing rate during navigation based on the chest location. Right, same spikes binned by subject position to calculate firing rate on the path. B. Top-left, firing rate map of navigation activity binned by memory target position for a neuron in the left EC from Patient 9. Black line indicates the perimeter of the traversable virtual environment. Bottom-left, histogram of p-values from ANOVA (see Methods) assessing memory target location modulation of firing rate for the observed data (red) versus shuffled data (gray). This cell's activity is significantly modulated by the memory target position (permutation-corrected ANOVA,  $p < 0.001$ ). Top-right, firing rate map for current location. Bottom-right, histogram of p-values from ANOVA assessing current location modulation of firing rate. Neuron is not significantly modulated by subject position ( $p = 0.437$ ). C. Same as B but for another example neuron in the right EC from Patient 12. Neuron is significantly modulated by memory target position ( $p = 0.004$ ) and not subject position ( $p = 0.49$ ). D. Percentage of significant memory-target cells by region. Shown for all MTL neurons and also split into HF and PHG. \*\* indicates  $p < 0.01$  for binomial test.*



**Figure 3: Neural activity related to the subject's current location.** A. Firing rate map of navigation activity binned by subject location for a right parahippocampal cortex neuron from Patient 14. This cell's activity is significantly modulated by the subject's virtual position (permutation-corrected ANOVA,  $p = 0.0195$ ). B. Example cell from left EC in Patient 9 ( $p = 0.009$ ). C. Example neuron from Patient 4 in right EC. Neuron is significantly modulated by subject position ( $p = 0.032$ ). D) Percentage of significant spatially-tuned cells by region. \*\* indicates  $p < 0.01$  for binomial test.

119 Twelve MTL neurons (9%) showed significant changes in firing rate in relation to subsequent  
120 memory performance, which is greater than the 5% expected by chance ( $p = 0.014$ , binomial test).  
121 This effect was most prominent in the PHG ( $p = 0.011$ , binomial test; Fig. 5d). Of the twelve cells  
122 that showed significant memory-related firing-rate changes, seven demonstrated significant memory-  
123 related firing rate increases while five showed significant decreases (see Figure 5C). Of the 12 memory  
124 cells, zero were place-like cells, 3 were memory-target cells, and 2 were heading-direction cells.

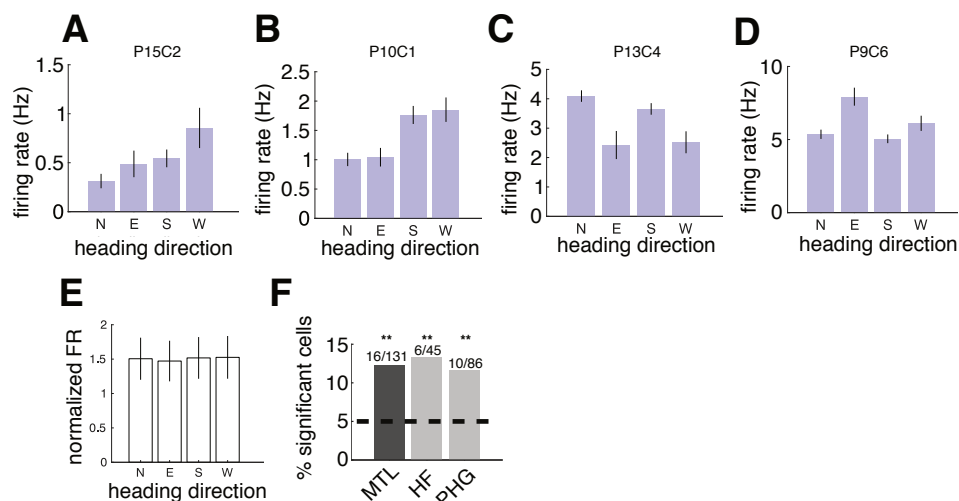
## 125 Discussion

126 In this study, we found that the firing rates of subjects' MTL cells were modulated by the locations  
127 of to-be-remembered targets, heading direction, and subsequent memory performance, but not by  
128 self location. By showing that single-neuron activity in the human MTL can represent remote spatial  
129 information, these results help explain how contextual information, such as relevant remote locations,  
130 may be used to support memory formation in humans.

131 Several other studies have described MTL activity in animals with spatial responses that are related  
132 to our findings in humans. The existence of hippocampal "spatial view" cells in non-human primates  
133 provides one possible explanation for the memory-target cells we describe (Rolls and O'Mara, 1995;  
134 Wirth et al., 2017). Those studies characterized how MTL neurons code for remote spatial locations,  
135 and our study extends this area of research to relate to memory encoding. Furthermore, there is recent  
136 evidence of entorhinal grid cells coding for viewed locations in 2D (Wilming et al., 2018), as well as  
137 view-related sharp-wave-ripple responses in the primate hippocampus (Leonard and Hoffman, 2017).  
138 Additionally, "social place cells," whose activity reflect allocentric representations of conspecifics, are  
139 another example of coding for salient locations albeit not for memory targets (Omer et al., 2018;  
140 Danjo et al., 2018). Finally, the cells we describe may be related to the view cells found by Ekstrom  
141 et al. (2003). However, whereas Ekstrom et al.'s view cells represent visually distinctive buildings,  
142 the memory-target cells we describe represent locations that are denoted by identical treasure chests.  
143 Thus, our study adds to the growing evidence that MTL neurons can be modulated as a function of  
144 salient remote spatial locations.

145 In addition to our primary finding of neurons that activate for remote target locations, we also  
146 observed other types of navigationally relevant neural responses. The first of these are heading-  
147 direction cells, in which we provide some of the first evidence of this cell type in humans. Head-



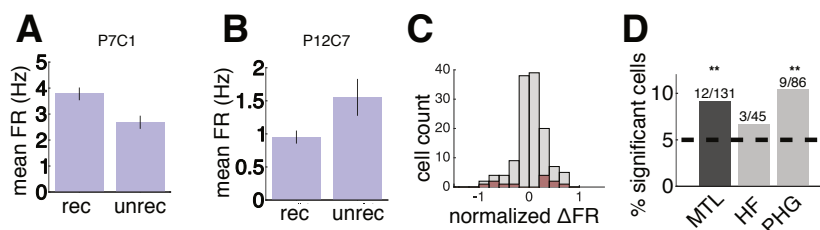


**Figure 4: Neural activity related to the subject's heading.** Firing rate by heading direction (split by N=north, E=east, S=south, W=west) for example cells significantly modulated by heading direction. Error bars are SEM of each group. A) Example cell from Patient 15, in the left hippocampus, significantly modulated by heading direction ( $p = 0.026$ ). B) Example cell from Patient 10, in the left parahippocampal cortex, significantly modulated by heading direction ( $p = 0.005$ ). C-D) Two more significant heading-direction cells in the left hippocampus and left EC respectively (from Patient 13;  $p = 0.0315$ , from Patient 9;  $p = 0.0015$ ). E) Average of z-scored firing rates for each cell split by heading direction, across all significant heading-direction cells. F) Percentage of significant heading-direction cells by region. \*\* indicates  $p < 0.01$  for binomial test.

148 direction cells have been described extensively in rodents and are most frequently found in areas such  
 149 as the postsubiculum, retrosplenial cortex, anterodorsal thalamus (Taube et al., 1990; Taube, 1998;  
 150 Robertson et al., 1999). However, they have also been found in the hippocampus and entorhinal cortex  
 151 (Leutgeb et al., 2000; Sargolini et al., 2006). In addition to the cells we found that respond most to  
 152 movement in a single direction, we also found evidence of cells with bidirectional responses, which are  
 153 similar to patterns reported recently in rodents (Jacob et al., 2017).

154 Finally, we observed neurons that varied their firing rates during navigation according to subsequent  
 155 memory performance. Human fMRI studies consistently report greater hippocampal BOLD activity  
 156 for subsequent memory (Schacter and Wagner, 1999; Chua et al., 2007; Suthana et al., 2009).  
 157 However, few studies have found that single-neuron firing rates relate to episodic memory in the  
 158 human hippocampus and surrounding MTL. For example, Rutishauser et al. (2010) found that mean  
 159 firing rates during learning did not differ as a function of subsequent memory performance. Here we  
 160 found cells whose firing rates significantly increased for successful memory encoding and others whose  
 161 firing significantly decreased. This variability could explain why previous studies of population neural  
 162 signals did not find a consistent firing-rate response relating to subsequent memory, because the mixed  
 163 patterns from individual neurons may cancel at the population level. In accordance with our results,  
 164 Wixted et al. (2014) report evidence of a sparse and distributed code of episodic memory in human  
 165 hippocampus, which is consistent with our finding that individual neurons showed both increases and  
 166 decreases in firing rates during successful memory encoding.

167 In light of the literature on place cells in rodents (O'Keefe and Dostrovsky, 1971), monkeys (Wirth  
 168 et al., 2017; Gulli et al., 2018), and humans (Ekstrom et al., 2003), it might be considered surprising  
 169 that we did not observe a large proportion of neurons with activity that varied as a function of the  
 170 subject's own location. As we describe below, this pattern may have stemmed from the behavioral  
 171 demands of our task, which differed compared to the paradigms used previously to study human



**Figure 5: Neural activity related to subsequent memory.** Firing rate by subsequent memory performance for two example cells significantly modulated by subsequent recall. Error bars are SEM of the trial means in each condition. A. Example neuron from Patient 7 in the left PRC, significantly modulated by subsequent memory ( $p = 0.017$ ). B. Example neuron from Patient 12 in the right EC,  $p = 0.044$ . C. Histogram of cell count by normalized firing rate between conditions  $((rec-unrec)/rec)$ . Red indicates the significant cells. D. Percentage of significant memory-related cells by region. \*\* indicates  $p < 0.01$  for binomial test.

172 single-neuron electrophysiology in navigation. We hypothesize that the nature of the MTL's spatial  
173 coding varies with task demands. In our experiment, on each trial subjects remembered the positions  
174 of objects that had been placed at locations in an open environment that were subsequently unmarked.  
175 Therefore, this task allowed us to examine neural activity used to represent new spatial locations for  
176 memory formation. On each trial in our task, new, previously unmarked locations become salient  
177 and relevant for memory encoding. This may have led to greater attentional focus to those upcoming  
178 locations during navigation instead of the subject's current location. However, in other tasks where the  
179 environmental structure was fairly static, subjects were likely to focus more on their current location,  
180 which would have been more likely to elicit place-cell activity (e.g. Ekstrom et al., 2003; Jacobs et al.,  
181 2010, 2013; Miller et al., 2013, 2015; Watrous et al., 2018; Qasim et al., 2018).

182 Our primary result is that single neurons in the human MTL code for remote locations that are  
183 relevant for memory. We also find coding of heading direction and subsequent memory. A remaining  
184 open question is whether the neuronal representation of memory targets relates specifically to where  
185 subjects are viewing, and future work utilizing eye tracking will be able to address this directly. By  
186 showing human MTL neuronal activity related to the location of upcoming memory targets rather  
187 than a subject's own location, our findings indicate that the human hippocampal system changes the  
188 nature of its representation to support task demands. This opens new directions for future research  
189 on what causes these representational schemes to change and how the brain links between different  
190 types of representations for individual memories.

## 191 Methods

### 192 Experimental Task

193 The subjects in our study were neurosurgical patients, who performed one of two versions of the  
194 Treasure Hunt (TH) task (referred to as 'object-cued' and 'location-cued' in Figure 1a). TH is a 3D  
195 virtual spatial-memory game developed in Unity3D. Subjects played TH on a bedside laptop computer  
196 and controlled their movement with a handheld joystick. In each trial of the task subjects explored  
197 a virtual beach (100 × 70 virtual units) to reach treasure chests that revealed hidden objects, with  
198 the goal of encoding the location of each encountered item. For a more detailed description of the  
199 object-cued task, see Miller et al. (2018). Briefly, the main components of the task are the Navigation,  
200 Encoding, and Retrieval phases. The focus of this paper is on the Navigation period.

201 Each trial of the object-cued task begins with navigation to a chest (i.e., the Navigation phase)



202 using a joystick. Upon arrival at the chest, the chest opens and either reveals an object or is empty.  
203 The subjects remain facing the open chest for 1.5 s (Encoding phase). In each trial the subjects  
204 navigate to a sequence of 4 chests. Two or three (randomly selected) of the chests contain an object  
205 and the rest are empty. In each session there are a total of 100 full chests and 60 empty chests, over  
206 40 trials. In each trial the chests are placed in random locations such that their locations span the  
207 environment across trials, and they are never located in the outermost positions in the environment.  
208 After all four chests have been reached in each trial, subjects are transported automatically so they  
209 view the environment from an overhead perspective. They then perform a distractor minigame before  
210 entering the Recall phase. In Recall subjects are cued with each of the objects from the trial in a  
211 random sequence and asked to recall their location by placing the cursor in the correct location from  
212 the overhead view.

213 The location-cued task is a variation of the object-cued version with a different Recall phase, in  
214 which the subjects are cued with a location and asked to verbally recall the corresponding object. Each  
215 session of this task version consists of 30 trials, each with 3 or 4 chests, for a total of 105 chests  
216 per session. None of the chests are empty. During the Recall phase subjects are probed with 4 or 5  
217 locations, one of which is a lure location that does not match the location of any of the trial's objects.

## 218 **Intracranial Recordings**

219 Fifteen patients (10 Male, mean age=32 years, minimum age=20 years) with medication-resistant  
220 epilepsy participated in this study. Subjects underwent a surgical procedure in which depth electrodes  
221 were implanted to localize epileptogenic regions at 3 different hospital sites (Columbia University Med-  
222 ical Center, Emory University School of Medicine, Thomas Jefferson University). Electrode placement  
223 was determined solely by the clinical team at each collaborating hospital. Behnke-Fried microelectrodes  
224 with 9 platinum-iridium microwires (40  $\mu\text{m}$ ) extending from the macroelectrode tip were implanted  
225 in the participating subjects following previously reported methods (Fried et al., 1999; Misra et al.,  
226 2014). All patients provided informed consent for both the Behnke-Fried implants and the behavioral  
227 task, under IRB protocols at all three institutions. The microwire data were recorded at 30 kHz using  
228 NeuroPort recording systems (Blackrock Microsystems, Salt Lake City, UT). We conducted automatic  
229 spike detection and sorting in Combinato (Niediek et al., 2016) and followed by manual sorting to  
230 identify putative single neurons following criteria described in Valdez et al. (2013).

231 Subjects participated in a total of 23 TH sessions. Across these sessions we successfully isolated  
232 a total of 131 putative neurons from microelectrodes localized to the medial temporal lobe (MTL).  
233 Forty-five of these neurons were in the HF: hippocampus and subiculum, and 86 were in the PHG:  
234 EC, perirhinal cortex, amygdala, and parahippocampal cortex (see Table 1) .

## 235 **Behavioral performance**

236 We assessed performance on the object-cued task as in earlier work (Miller et al., 2018). For each  
237 object the distance error is defined as the Euclidean distance between the subject's response and  
238 the correct location. Accuracy is defined as 1 minus the percentile rank of the actual distance error  
239 computed relative to the distribution of all possible distance errors that could have been made for the  
240 object's location. Performance on the location-cued task was calculated as the percentage of words  
241 correctly freely recalled.

## 242 **Anatomical localization**

243 We determined the anatomical location of each microwire electrode bundle by co-registering the pre-  
244 surgical T1 and T2 weighted structural MRIs to the post-surgical CT scan. Only subjects with depth  
245 electrodes extending into the MTL were included in this study. MTL subregions were automatically  
246 labeled using a multi-atlas based segmentation technique on the T2-weighted MRI. A neuroradiologist  
247 identified the electrode contacts on the post-surgical CT. Electrode contact coordinates were then  
248 mapped to MRI space and a neuroradiologist manually determined the anatomical locations of the  
249 microwire electrodes based on the co-registered images.

## 250 **Analysis of spatial coding during navigation epochs**

251 We analyzed neural data collected from the navigation periods of each task session. The rectangular  
252 environment was binned into a  $5 \times 7$  grid, and both the current location and current memory target  
253 location were logged for each navigation epoch. For the data from a grid location to be used in the  
254 analysis, the subject must have occupied that location for a minimum of 5 s or 2 s when binning by  
255 subject position or memory target position, respectively. We discretized the behavioral navigation data  
256 into 100-ms epochs, and calculated the subject's current average x- and y-coordinate, current chest  
257 x- and y-coordinate, and speed within each 100-ms bin. We excluded navigation epochs during which  
258 subjects were not moving for more than 500 ms from the analyses. We binned the spike data into the  
259 corresponding 100-ms epochs and calculated the firing rate for each spatial bin. We calculated firing  
260 rate by subject position using the subject's virtual position, and firing rate by memory target position  
261 using each path's corresponding chest location.

262 We identified spatially modulated cells using an ANOVA to predict firing rate with current and  
263 memory target location as predictors. We assessed statistical significance using random circular per-  
264 mutation, as in earlier work (Ekstrom et al., 2003; Miller et al., 2015). This procedure was repeated  
265 1000 times with circularly time-shifted firing rate values, whereby the firing rate of the cell was ran-  
266 domly shifted relative to the behavioral navigation data. If the test statistic calculated on the real data  
267 was in the 95<sup>th</sup> percentile of the test statistics from the shifted data, the parameter was considered  
268 a significant factor in modulating firing rate.

269 To test for modulation of firing rate by heading direction, a separate ANOVA was conducted  
270 with heading quadrant (N,E,S,W) as a factor. The heading direction was determined using the x,y  
271 coordinates of the path and calculating the movement angle. To test for modulation of firing rate by  
272 subsequent memory another separate ANOVA was conducted with memory performance as a factor.

273 For each cell-type category, we tested the proportion of significant cells against the null hypothesis  
274 that the proportion was not significantly different from chance using a binomial test with  $\alpha =$   
275 0.05. We smoothed the firing rate maps for visualization purposes by binning into a  $11 \times 16$  grid and  
276 applying a Gaussian filter with a 1.1 bin standard deviation. Any grid with at least 100 ms spent in it  
277 was included in these plots, and all other locations were plotted as white.

## 278 **Control for epileptic regions**

279 The units from three subjects in our study were localized to what was clinically determined to be the  
280 seizure onset zone. To control for a possible confound caused by epileptic activity, we re-calculated  
281 the proportions excluding the 15 units from these patients (Patients 2, 10, 12). Our main findings  
282 remained consistent after this exclusion:  $23/116 = 20\%$  of the units were classified as memory-target  
283 cells,  $6/116 = 5\%$  as place-like cells,  $14/116 = 12\%$  as heading-direction cells, and  $9/116 = 8\%$  as  
284 memory-related cells.

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## References

- 289  
290 T. I. Brown, V. A. Carr, K. F. LaRocque, S. E. Favila, A. M. Gordon, B. Bowles, J. N. Bailenson,  
291 and A. D. Wagner. *Science*, 352(6291):1323–1326, 2016.
- 292 E. F. Chua, D. L. Schacter, E. Rand-Giovannetti, and R. A. Sperling. *Hippocampus*, 17(11):1071–  
293 1080, 2007.
- 294 T. Danjo, T. Toyozumi, and S. Fujisawa. *Science*, 359(6372):213–218, 2018.
- 295 A. D. Ekstrom. *Hippocampus*, 25(6):731–735, 2015.
- 296 A. D. Ekstrom, M. J. Kahana, J. B. Caplan, T. A. Fields, E. A. Isham, E. L. Newman, and I. Fried.  
297 *Nature*, 425:184–187, 2003.
- 298 R. A. Epstein, E. Z. Patai, J. B. Julian, and H. J. Spiers. *Nature neuroscience*, 20(11):1504, 2017.
- 299 I. Fried, C. Wilson, N. Maidment, J. J. Engel, E. Behnke, T. Fields, K. MacDonald, J. Morrow, and  
300 L. Ackerson. *Journal of Neurosurgery*, 91:697–705, 1999.
- 301 J. L. Gauthier and D. W. Tank. *Neuron*, 99(1):179–193.e7, Jul 2018. doi: 10.1016/j.neuron.2018.  
302 06.008.
- 303 J. A. Greenberg, J. F. Burke, R. Haque, M. J. Kahana, and K. A. Zaghloul. *NeuroImage*, 114:257–  
304 263, Jul 2015. ISSN 1095-9572 (Electronic); 1053-8119 (Linking). doi: 10.1016/j.neuroimage.  
305 2015.03.077.
- 306 R. A. Gulli, L. Duong, B. W. Corrigan, G. Doucet, S. Williams, S. Fusi, and J. C. Martinez-Trujillo.  
307 *bioRxiv*, page 295774, 2018.
- 308 P.-Y. Jacob, G. Casali, L. Spiesser, H. Page, D. Overington, and K. Jeffery. *Nature neuroscience*, 20  
309 (2):173, 2017.
- 310 J. Jacobs, M. J. Kahana, A. D. Ekstrom, M. V. Mollison, and I. Fried. *Proceedings of the National*  
311 *Academy of Sciences*, 107(14):6487–6482, 2010.
- 312 J. Jacobs, C. T. Weidemann, J. F. Miller, A. Solway, J. F. Burke, X. Wei, N. Suthana, M. R. Sperling,  
313 A. D. Sharan, I. Fried, and M. J. Kahana. *Nature Neuroscience*, 16(9):1188–1190, 2013. doi:  
314 10.1038/nn.3466.
- 315 S. A. Lee, J. F. Miller, A. J. Watrous, M. Sperling, A. Sharan, G. A. Worrell, B. M. Berry, B. C. Jobst,  
316 K. A. Davis, R. E. Gross, B. Lega, S. Sheth, S. R. Das, J. M. Stein, G. Richard, D. S. Rizzuto,  
317 and J. Jacobs. *Journal of Neuroscience*, page 218040, 2018.
- 318 T. K. Leonard and K. L. Hoffman. *Current Biology*, 27(2):257–262, 2017.
- 319 S. Leutgeb, K. Ragozzino, and S. Mizumori. *Neuroscience*, 100(1):11–19, 2000.
- 320 J. F. Miller, M. Neufang, A. Solway, A. Brandt, M. Trippel, I. Mader, S. Hefft, M. Merkow, S. M.  
321 Polyn, J. Jacobs, M. J. Kahana, and A. Schulze-Bonhage. *Science*, 342(6162):1111–1114, 2013.  
322 doi: 10.1126/science.1244056.
- 323 J. F. Miller, I. Fried, N. Suthana, and J. Jacobs. *Current Biology*, 2015.

- 324 J. F. Miller, A. J. Watrous, M. Tsitsiklis, S. A. Lee, S. A. Sheth, C. A. Schevon, E. H. Smith, M. R.  
325 Sperling, A. Sharan, A. A. Asadi-Pooya, G. A. Worrell, S. Meisenhelter, C. Inman, K. A. Davis,  
326 B. Lega, P. A. Wanda, S. R. Das, J. M. Stein, R. Gorniak, and J. Jacobs. *Nature communications*,  
327 9(1):2423, 2018.
- 328 A. Misra, J. Burke, A. Ramayya, J. Jacobs, M. Sperling, K. Moxon, M. Kahana, J. Evans, and  
329 A. Sharan. *Journal of neural engineering*, 11(2):026013, 2014.
- 330 R. Morris, P. Garrud, J. a. Rawlins, and J. O’Keefe. *Nature*, 297(5868):681, 1982.
- 331 R. Muller. *Neuron*, 17:813–822, 1996.
- 332 J. Niediek, J. Boström, C. E. Elger, and F. Mormann. *PLoS One*, 11(12):e0166598, 2016. doi:  
333 10.1371/journal.pone.0166598.
- 334 J. O’Keefe and J. Dostrovsky. *Brain Research*, 34:171–175, 1971.
- 335 D. B. Omer, S. R. Maimon, L. Las, and N. Ulanovsky. *Science*, 359(6372):218–224, 2018.
- 336 S. Qasim, J. Miller, C. Inman, R. Gross, J. Willie, B. Lega, J. Lin, A. Sharan, C. Wu, M. Sperling,  
337 et al. *bioRxiv*, 2018.
- 338 R. Robertson, E. Rolls, P. Georges-François, and S. Panzeri. *Hippocampus*, 9:206–219, 1999.
- 339 E. Rolls and S. O’Mara. *Hippocampus*, 5(5):409–424, 1995.
- 340 U. Rutishauser, I. Ross, A. Mamelak, and E. Schuman. *Nature*, 464(7290):903–907, 2010.
- 341 F. Sargolini, M. Fyhn, T. Hafting, B. McNaughton, M. Witter, M. Moser, and E. Moser. *Science*,  
342 312(5774):758–762, 2006.
- 343 D. L. Schacter and A. D. Wagner. *Hippocampus*, 9:7–24, 1999.
- 344 D. Schiller, H. Eichenbaum, E. A. Buffalo, L. Davachi, D. J. Foster, S. Leutgeb, and C. Ranganath.  
345 *Journal of Neuroscience*, 35(41):13904–13911, 2015.
- 346 W. B. Scoville and B. Milner. *Journal of Neurology, Neurosurgery, and Psychiatry*, 20:11–21, 1957.
- 347 H. Spiers and E. Maguire. *Neuroscience*, 149(1):7–27, 2007.
- 348 N. Suthana, A. D. Ekstrom, S. Moshirvaziri, B. J. Knowlton, and S. Y. Bookheimer. *Journal of*  
349 *Neuroscience*, 2009.
- 350 J. Taube. *Progress in Neurobiology*, 55(3):225–56, 1998.
- 351 J. Taube, R. Muller, and J. Ranck. *Journal of Neuroscience*, 10(2):420–435, 1990.
- 352 A. B. Valdez, E. N. Hickman, D. M. Treiman, K. A. Smith, and P. N. Steinmetz. *J Neural Eng*, 10  
353 (1):016001, Feb 2013. doi: 10.1088/1741-2560/10/1/016001.
- 354 L. K. Vass, M. S. Copara, M. Seyal, K. Shahlaie, S. T. Farias, P. Y. Shen, and A. D. Ekstrom. *Neuron*,  
355 89(6):1180–1186, 2016.
- 356 A. J. Watrous, J. Miller, S. E. Qasim, I. Fried, and J. Jacobs. *eLife*, 7:e32554, 2018.

- 357 N. Wilming, P. König, S. König, and E. A. Buffalo. *eLife*, 7:e31745, 2018.
- 358 S. Wirth, P. Baraduc, A. Planté, S. Pinède, and J.-R. Duhamel. *PLoS biology*, 15(2):e2001045, 2017.
- 359 J. T. Wixted, L. R. Squire, Y. Jang, M. H. Papesch, S. D. Goldinger, J. R. Kuhn, K. Smith, D. Treiman,  
360 and P. N. Steinmetz. *Proceedings of the National Academy of Sciences*, 111(26):9621–9626, 2014.



Patient number	Age	Gender	# Sessions	Task version	# MTL Behnke-Fried bundles	# MTL units	Electrode locations
1	22.7	M	1	object cued	2	4	R SUB, R HPC
2	52.3	M	2	object cued	6	3	L AMY, L HPC, R EC, R SUB, R HPC
3	41	M	1	object cued	1	2	L HPC
4	22.4	M	3	object cued	1	5	R EC
5	21.5	F	2	object cued	2	7	L HPC, L SUB
6	25.6	F	1	object cued	2	11	R EC
7	29.4	M	2	object cued	2	9	L SUB, L PRC
8	23.4	F	1	object cued	1	1	R HPC
9	21.9	F	3	object cued	2	39	L EC
10	27.6	M	1	object cued	2	4	L SUB, L PHC
11	47	M	1	object cued	1	8	L EC
12	20	M	1	object cued	1	8	R EC
13	21	M	1	location cued	2	12	L HPC, R HPC
14	55	M	1	location cued	2	2	R HPC, R PHC
15	43.5	F	2	location cued	2	16	L HPC, R SUB

**Table 1: Patients and unit information.** Table indicates each patient's demographics and their MTL unit counts. R/L: right/left; HPC: hippocampus, SUB: subiculum, AMY: amygdala, EC: entorhinal cortex, PRC: perirhinal cortex, PHC: parahippocampal cortex.