

Capturing site-specific heterogeneity with large-scale N-glycoproteome analysis

Nicholas M. Riley,^{1,2} Alexander S. Hebert,¹ Michael S. Westphall,¹ and Joshua J. Coon^{1-4*}

¹Genome Center of Wisconsin, Departments of ²Chemistry and ³Biomolecular Chemistry, University of Wisconsin-Madison, Madison, WI, 53706, USA

⁴Morgridge Institute for Research, Madison, Wisconsin, USA

*to whom correspondence should be addressed: jcoon@chem.wisc.edu

ABSTRACT

Protein glycosylation is a highly important, yet a poorly understood protein post-translational modification. Thousands of possible glycan structures and compositions create potential for tremendous site heterogeneity and analytical challenge. A lack of suitable analytical methods for large-scale analyses of intact glycopeptides has ultimately limited our abilities to both address the degree of heterogeneity across the glycoproteome and to understand how it contributes biologically to complex systems. Here we show that N-glycoproteome site-specific microheterogeneity can be captured via large-scale glycopeptide profiling with methods enabled by activated ion electron transfer dissociation (AI-ETD), ultimately characterizing 1,545 N-glycosites (>5,600 unique N-glycopeptides) from mouse brain tissue. Moreover, we have used this large-scale glycoproteomic data to develop several new visualizations that will prove useful for analyzing intact glycopeptides in future studies. Our data reveal that N-glycosylation profiles can differ between subcellular regions and structural domains and that N-glycosite heterogeneity manifests in several different forms, including dramatic differences in glycosites on the same protein.

MAIN TEXT

As insights into the role of glycosylation in health and disease continue to emerge, new technologies are required to improve the depth and breadth of glycoproteome analysis.^{1–4} Especially needed are methods for intact glycopeptide analysis – an approach that preserves biological context of the modification and enables understanding of proteome wide glycan heterogeneity.^{5,6} The recent investment in glycoscience technology development made by the National Institutes of Health Common Fund program evince this importance and need.⁷ Mass spectrometry (MS)-based methods are the premier approach to glycoproteome characterization. The bulk of our glycoproteome knowledge comes from methods that enzymatically cleave the glycan from the peptide and then sequence each molecular class separately. This approach has revealed a stunning diversity of hundreds of unique N-linked glycan structures that can decorate proteins. The specific residues that carried the modification can likewise be identified; however, the information of which glycan structures go on which sites is lost. For this reason, one cannot determine whether particular sites or classes of proteins have preference for one structure or another nor ultimately what the functional roles of the various glycans are. It is known that glycan heterogeneity can affect structure and function, such as binding specificities in immunoglobulins,^{8,9} but the functional effects of heterogeneity across the glycoproteome remain largely uncharacterized.

To gain a better understanding of glycan heterogeneity at a given site one can analyze intact glycopeptides. Similar to many other post-translational modifications (PTMs), glycopeptides require enrichment prior to analysis because of low stoichiometry and suppressed ionization efficiency compared to unmodified peptides.¹⁰ Other methodological factors, e.g., MS acquisition speeds, online and offline chromatographic separations, choice of protease, also influence PTM characterization and must be considered in glycopeptide analysis.^{2,5,6,10} Unlike the majority of other PTMs, however, the glycan can be as large or larger than the peptide itself. Beyond that, it has been challenging to find tandem mass spectrometry dissociation methods that can cleave both the peptide backbone and glycosidic bonds to offer successful peptide and glycan composition determination. As such, multiple dissociation strategies (mainly electron-driven dissociation and collision-based methods) are increasingly used to access both glycan and peptide information from intact glycopeptides.^{11,12,21–26,13–20} Even with these methods, large-scale analysis of intact glycopeptides remains largely limited to fewer than ~1,000 unique glycosite identifications from any one system or tissue (approximately an order of magnitude behind other

PTMs),^{27–32} and few studies assess heterogeneity across the glycoproteome with site-specific resolution.

Activated-ion electron transfer dissociation (AI-ETD) has rapidly developed as a highly effective tandem MS approach for proteomic applications.^{33–41} The method uses concurrent infrared (IR) photoactivation to improve dissociation efficiencies and increase product ion generation in electron transfer dissociation (ETD) reactions.^{11,42} We report here that the combination of simultaneous vibrational activation from IR photon bombardment and electron-driven dissociation via ETD is particularly well-suited for intact glycopeptide fragmentation. Using an AI-ETD-enabled method for large-scale glycoproteomic analysis, we characterize 5,662 unique N-glycopeptides mapping to 1,545 unique N-glycosites on 771 glycoproteins and use this dataset to explore profiles of heterogeneity present at multiples levels of proteomic information, from glycosites to subcellular regions.

RESULTS

AI-ETD Performance for Intact Glycopeptides

AI-ETD provides information about both peptide and glycan components of intact N-glycopeptides by concomitantly capitalizing on two complementary modes of fragmentation in a single MS/MS event (**Figure 1a**). The combination of vibrational activation and electron-driven dissociation is concurrent in both space and time with AI-ETD, which also reduces overhead time in MS/MS scans compared to other supplemental activation techniques (e.g., ETcAD and EThcD). This enables slightly more scans per unit time and, ultimately, more identifications (**Supplemental Figure 1**), although other supplemental activation methods can still be quite valuable. Indeed, EThcD has proven suitable for glycoproteome characterization in a number of recent studies,^{18,19,26} and future studies will likely focus on more systematic comparisons of multiple supplemental activation strategies that include AI-ETD. AI-ETD generates extensive fragmentation along the peptide backbone, including mainly c- and z[•]-type products with some y-type fragments (100% sequence coverage in this example of the glycopeptide TN*SSFIQGFDVHKEDCDR, where N* is the glycosite modified with a high mannose HexNAc(2)Hex(9) glycan). Importantly, product ions from peptide backbone cleavage largely retain the entire intact glycan, as is seen here in a series of doubly charged c-type fragments. Minimal b-type product generation indicates that the majority of peptide backbone fragmentation comes from electron-driven dissociation via ETD rather than vibrational activation, matching

observations from non-modified peptides and proteins.^{35,37} That said, vibrational activation from infrared photons does impart enough energy to dissociate more labile glycosidic bonds, producing extensive series of Y-ion fragments (i.e., ions that have lost a portion of the non-reducing end of the glycan but retain the intact peptide sequence) that provide details about glycan composition. Furthermore, the infrared photoactivation of AI-ETD also generates complementary B-type fragments and other oxonium ions to indicate the presence of various sugar moieties. Thus, the vibrational and electron-driven dissociation modes together provide information rich spectra for high quality glycopeptide identifications.

We leveraged AI-ETD for glycoproteomic data collection by triggering scans based on the presence of oxonium ions in HCD spectra (HCD-pd-AI-ETD),^{43,44} which allowed straightforward comparisons of AI-ETD and HCD spectra. AI-ETD produced more peptide backbone fragments and more Y-type fragments (mainly glycan fragments from the charge reduced precursors) than HCD, while HCD produced more oxonium ions (**Supplemental Figure 2**). **Supplemental Figure 3** displays the percent of AI-ETD and HCD identifications that contain a number of common glycopeptide Y-ion fragments and oxonium ions. Only small fraction of spectra from both AI-ETD and HCD contained the Y1-ion (i.e., the intact peptide plus one HexNAc) that carries the same charge of the precursor, while the Y1-ion with one less charge than the precursor was observed in 59.2% and 69.2% of AI-ETD and HCD spectra, respectively. Some database search strategies for intact glycopeptides utilize the presence of Y1-ions in HCD spectra,⁴⁵ and this data shows that AI-ETD may be a reasonable candidate for such an approach in future work. Also, AI-ETD more often produced larger Y-type fragments, including the intact peptide with two HexNAc moieties and the intact peptide with the full HexNAc(2)Hex(3) common N-linked glycan core, and these fragments could also be used to improve glycopeptide searching strategies with AI-ETD spectra. Both AI-ETD and HCD produced at least one of these Y-ions in approximately 72% identified spectra.

All HCD spectra contained the HexNAc oxonium ion (m/z 204.0867), which was also present in 99.97% of AI-ETD spectra (all but four spectra). Conversely, effectively no AI-ETD spectra (0.25%) contained the Hex oxonium ion at m/z 163.06, yet it was observed in 97.27% of HCD spectra. Similar to the HexNAc oxonium ion, the m/z 366.14 oxonium ion (HexNAcHex) was present in nearly all HCD and AI-ETD spectra, but three common larger oxonium ions were more often observed in HCD spectra (**Supplemental Figure 3**). That said, oxonium ions from sialylated glycans were the exception to this trend, which is discussed further below.

Others have reported the ability to distinguish glycan isomers using ratios of oxonium ion intensities in higher energy collisional dissociation (HCD) spectra, namely to distinguish the presence of N-acetylglucosamine (GlcNAc, present in both N- and O-linked glycans) and N-acetylgalactosamine (GalNAc, only in O-linked glycans).^{46–48} In a second dataset, we extended the low mass range of AI-ETD spectra to 115 Th and calculated the GlcNAc/GalNAc ratio for AI-ETD and HCD spectra of intact glycopeptides (**Supplemental Figure 4a**). No GalNAc residues are expected to be present in this data set because of the focus on N-glycopeptides, so ratios for each dissociation method should only indicate the presence of GlcNAc. As noted by Nilsson and co-workers, a GlcNAc/GalNAc ratio below 1 indicates the presence of GalNAc, while a ratio above 2 is significant for the presence of GlcNAc.^{46,47} Nearly the entire distribution (99.9%) of calculated GlcNAc/GalNAc ratios for AI-ETD spectra is greater than 2 (median of 6.52), providing a strong indication for the sole presence of GlcNAc as the primary isomer for all HexNAc residues. HCD spectra also provide ratios with a median value greater than 2 (median of 3.41), but 13% of HCD spectra provide a ratio below 2 despite the collision energy being within the previously investigated range.

We also examined oxonium ions (m/z 292.1027 and m/z 274.0921) from the sialic acid residue N-acetylneuraminic acid (Neu5Ac). Both AI-ETD and HCD generated the m/z 274.0921 ion with high frequency for spectra from sialylated glycans (96% and 95%, respectively), and the m/z 292 was also present in both, although slightly less frequently (87% and 93%, respectively). We also observed these ions to some degree in both AI-ETD and HCD spectra assigned to glycopeptides without a Neu5Ac moiety. This false indication of a Neu5Ac moiety can be controlled for by calculating a ratio of intensity of the m/z 274.0921 ion to the HexNAc oxonium (m/z 204.0867). Setting a threshold of > 0.1 for this Neu5Ac/HexNAc oxonium ion ratio eliminated 97% and 99% of AI-ETD and HCD spectra, respectively, that were assigned an identification without a Neu5Ac residue while retaining 83% and 88% of AI-ETD and HCD spectra that were assigned an identification with a sialylated glycan. Such a calculation could be considered in future glycopeptide-centric search algorithms that are capable of handling both AI-ETD and HCD spectra. Remarkably, Pap et al. observed that EThcD fragmentation preserves larger sialylated oxonium ions than HCD for O-linked glycans (namely m/z 657.2349, HexNAcHexNeuAc),⁴⁹ and we observed a similar trend with AI-ETD for sialylated N-glycans (**Supplemental Figure 4b**). The m/z 657.2349 ion was present in 87% of AI-ETD spectra from identifications containing a Neu5Ac residue, but only 44% of the analogous HCD spectra.

We also calculated Ln/Nn ratios for AI-ETD and HCD spectra to investigate the presence of isomeric glycoforms of Neu5Ac with either α 2,3 and α 2,6 linkages (**Supplemental Figure 4c**).⁵⁰ Both AI-ETD and HCD generate a wide range of Ln/Nn ratios, but distributions within the low values (from 0-3) of Ln/Nn ratios in spectra from both dissociation methods are the most interesting. AI-ETD ratios show a distribution with a median close the previously reported value for α 2,3 linkages but lack a distinct distribution for the higher values that would indicate α 2,6 linkages. HCD has two distinct distributions but they are much closer to each other than previously reported, and the lower distribution has a median with a greater value than expected. Even with these observations, it is difficult to comment on the accuracy of these calculations without predefined glycopeptide standards with known linkage information. Furthermore, others have used the presence of specific oxonium and neutral loss ions to discriminate between structure isomers (see Wu et al. for an example⁴⁵), and observation of both ion types in AI-ETD spectra indicates that AI-ETD could prove useful toward this goal. The ability of AI-ETD to distinguish glycan isomers needs to be further investigated and validated with dedicated future studies, but these data indicate that AI-ETD may be as valuable as HCD for generating oxonium ion distributions to distinguish GlcNAc and GalNAc isomers of HexNAc residues and that the method may also be able to provide insight on NeuAc linkage information.

Large-scale Glycoproteome Characterization Enabled by AI-ETD

Given the AI-ETD method is fast and easily automated, we reasoned the technique could provide analysis of the glycoproteome at a large-scale. To test this hypothesis we extracted proteins from mouse brain lysate, digested them with trypsin, enriched for glycosylated peptides, and performed high-throughput LC-MS/MS analysis using AI-ETD scans triggered by the presence of oxonium ions in HCD scans. In total, we identified 5,662 unique N-glycopeptides (24,099 glycopeptide spectral matches) mapping to 1,545 unique N-glycosites on 771 glycoproteins with 117 different glycan compositions, which were included in a database compiled from literature on previous mouse and rat brain glycosylation studies.^{27,51,52} These data are the result of several steps of post-Byonic search filtering, which were performed because caveats still exist in automated glycopeptide identification – as evidenced by the current HUPO glycoproteomics initiative (<https://hupo.org/HPP-News/6272119>). Note, we do not offer any fundamentally new approach to address such challenges here, but rather we present AI-ETD data for large-scale glycoproteomics using the tools that are currently available in the field. See the results for discussion of the six post-Byonic search filtering steps we performed. Following post-search filtering, no decoy

peptides remained in the dataset. All the data reported here comprise tryptic N-glycopeptides carrying only one glycan modification and have a Delta Mod Score that indicates the correct glycosite has been identified within the confidence range suggested by Byonic.

With this extensive dataset in hand we next characterized several Figures of Merit. First, we examined the percentage of cleaved bonds observed relative to total possible backbones bonds (for both peptide and glycan backbones, **Figure 1b**). Here we achieve 89% median peptide backbone sequence coverage and 78% median glycan sequence coverage with AI-ETD, which significantly outperforms HCD (**Supplemental Figure 5a**). **Figure 1c** presents the average distribution of explainable signal amongst fragment ion types in AI-ETD spectra. On average AI-ETD produces relatively equal proportions of signal in Y-type and peptide backbone fragments (41% and 45%, respectively), compared to HCD which has less signal in peptide backbone fragments and more in Y-type and oxonium ions (**Supplemental Figure 5b**). This is congruent observations presented in **Supplemental Figure 2** (discussed above). Approximately 29% and 46% of total ion current could be explained on average in AI-ETD and HCD spectra, respectively, but we only considered the following fragment ion classes: 1) unmodified peptide backbone fragments (*i.e.*, b/y/c/z-type), 2) peptide backbone fragments with intact glycan still attached, 3) peptide backbone fragments with only a HexNAc moiety still attached, 4) Y-type ions (intact peptide plus glycan fragments), and 5) oxonium ions/glycan B-type ions. It is possible that photoactivation generated some degree of glycan fragmentation on peptide backbone fragments, which could provide more explainable signal in AI-ETD spectra, and this will be the subject of future work. Even so, 71% of AI-ETD spectra (compared to less than 4% of HCD spectra) contained fragments with the intact glycan species retained on peptide backbone fragments. This percentage would likely increase (especially for AI-ETD) by extending the *m/z* range of MS/MS scans above 2,000 Th. Note, intact glycopeptides have considerably larger precursor *m/z* distributions (**Supplemental Figure 6**), as compared to unmodified peptides, and these low-charge density precursors ($z \leq 3$) can be a challenge to dissociate. Even so, AI-ETD provided robust fragmentation that often facilitated identification of these challenging glycopeptides (**Figure 1d**). AI-ETD spectra provided evidence for 4,680 unique N-glycopeptides (83%) and 1,361 (88%) of the glycosites reported in this study, with the remaining identifications/glycosites supported only by HCD spectral evidence. In all, these data represent the one of the most in-depth N-glycoproteome characterization to date to rely on intact glycopeptide identifications (**Figure 1e**). Importantly, this method is amenable to *in vivo* sources, as demonstrated here, for applicability to practically any mammalian system (workflow in **Supplemental Figure 7**).

The ability to profile glycosites with intact glycan modifications at this scale provides new opportunities to investigate system-wide glycosylation patterns. Despite differences in enrichment strategies and fragmentation methods, the overlap in identified glycosites is relatively high between this study and two other recent intact glycopeptide studies in mouse brain (**Figure 2a**) and de-glycoproteomic datasets (**Supplemental Figure 8**). **Supplemental Figure 9** compares one example of overlapping glycosites between this study and the Liu et al. dataset,³⁰ demonstrating that similar glycosites and glycan heterogeneity were identified on integrin alpha-1 in mouse brain, for example, and AI-ETD methods further add to the number of glycosites and glycosite-glycan combinations observed.

Figure 2b demonstrates that approximately 69% of the glycosites identified in this study are annotated as glycosites in the UniProt database. Of the 1,065 UniProt-annotated glycosites, the majority of them were assigned via ‘sequence analysis,’ which indicates a prediction based on presence of the N-X-S/T sequon rather than by experimental observation. In total, we provide new experimental evidence for over 850 UniProt-predicted glycosites, in addition to identifying nearly 200 previously observed glycosites. Expected N-X-S and N-X-T sequons were observed in our identified glycosites (**Figure 2c**), with approximately 59% of the glycosites having the N-X-T sequon. **Figure 2d** displays the percentage of glycosites containing high mannose glycans, fucosylated glycans, or sialylated glycans, which resembles previous studies (although Liu et al. observed significantly higher proportions of fucosylated glycopeptides).^{27,30} Note, this calculation did not consider glycosites exclusively, so one site can count toward multiple types if multiple glycans were identified at that site. Gene ontology (GO) enrichments of functional category terms from identified glycoproteins are available in **Supplemental Figure 10**, which shows expected enriched terms such as glycoprotein, several membrane terms, and extracellular/secreted protein related terms. See the Discussion for more about differences in glycosylation profiles between this study and other published datasets, where we also discuss the implications of our lectin enrichment strategy compared to other strategies.

Visualizing Glycoproteome Heterogeneity

Intact glycopeptide analysis uniquely enables characterization of site-specific microheterogeneity, and our large-scale data set can provide an initial glimpse at this fascinating facet of glycosylation. Trinidad et al. explored heterogeneity to some degree but ultimately provided a limited overview from a global perspective.²⁷ Others have explored several facets discussed herein to some

degree, including subcellular glycosylation profiles and glycosylation based on glycosite accessibility/structural motifs.^{30,53,54} Even so, we sought to approach these questions from a systems level using our large pool of intact glycopeptide identifications, and we developed several new ways to visualize such data. **Figure 2e** captures the prevalence of both singly- or multiply-glycosylated proteins (right) and the degree of glycan microheterogeneity for each of the 1,545 characterized glycosites (left). More than half of the 771 identified glycoproteins were observed with only one glycosite, but nearly 60% of glycosites have more than one glycan that modify them. A network diagram in **Figure 2f** maps which glycans (outer nodes) were observed on identified glycoproteins (inner column, organized by number of glycosites). Several discernable patterns appear, perhaps most notably the prevalence of high mannose glycosylation. The network diagram also indicates that the majority of fucosylated, paucimannose, and sialylated glycans occur on proteins with multiple glycosylation sites, and it indicates which glycans contribute more to heterogeneity. **Supplemental Figure 11** provides a larger version of this network diagram with glycan identities in **Supplemental Table 1**.

To further investigate site-specific microheterogeneity, we calculated how many times glycan pairs co-occurred at the same site, as shown in the glycan co-occurrence heat map in **Figure 2g** (larger version in **Supplemental Figure 12**). This data shows glycan pair combinations, i.e., glycans that appeared together at the same glycosite, and the darker color indicates more incidences of co-occurrence. High mannose glycans appear to co-occur together with high frequency, and they also co-occur with several groups of complex/hybrid, fucosylated, and sialylated glycans. Furthermore, numerous other co-occurrence patterns are observed, including co-occurrence of certain complex/hybrid and fucosylated glycans, different fucosylated glycans, and some specific fucosylated and sialylated glycans. We generated glycan co-occurrence networks to display the frequency of co-occurrence of specific glycans with all other glycans across glycosites. **Figure 3** shows an example of a co-occurrence network for a biantennary sialylated complex glycan and **Supplemental Figure 13** displays how co-occurrence networks facilitate visualization for co-occurrence for both a specific glycan, HexNAc(2)Hex(9), and an entire class of glycans, such as all high mannose glycans. Glycan identities are given in **Supplemental Table 3**. Arc plots in **Supplemental Figures 14-17** visualize glycan microheterogeneity delineated by the number of glycans per glycosite, showing increases in co-occurrence complexity as the number of glycans per site rises. We note that calculating mass differences between co-occurring glycans is straight-forward and can provide some information about glycan microheterogeneity (**Supplemental Figure 18**), but the limited resolution in

information about glycan differences and lack of dimensionality in this analysis inspired us to generate the other analyses and visualization discussed herein.

Glycoproteins with a high degree of glycan heterogeneity are readily observed by plotting the number of unique glycans versus total number of glycosites for a protein (**Figure 4**). Several interesting cases where the number of glycans is significantly higher than the total number of glycosites are highlighted. The distribution of glycan types is provided, showing that the types of glycans that contribute to this heterogeneity can vary based on the glycoprotein. Investigations into specific glycoprotein examples also indicate that glycan microheterogeneity can manifest in several different forms (**Figure 5** and **Supplemental Figures 19-21**). Some proteins can have several glycosites but relatively little glycan heterogeneity overall (e.g., protein sidekick-2, **Figure 5a**), while others can have one glycosite with a multitude of glycans modifying it (e.g., SPARC, **Figure 5b**). Or, in the case of sodium/potassium ATPase $\beta 2$ subunit (Atp1b2), some glycosites on a protein can show notably little heterogeneity while others have 15-20 different glycans modifying them (**Figure 5c**). Interestingly, Atp1b2 glycosites with lower glycan heterogeneity (N96, N156, N193, N197, and N250) are on one face of the protein while the sites with relatively high (>10 glycans) heterogeneity (N118, N153, and N238) are on the opposite face, where the protein interacts with alpha subunits.⁵⁵ Moreover, sites N118 and N238 have been shown to be important in mammalian systems for folding and localization of the Na/K-ATPase complex to the plasma membrane,⁵⁵ where it creates concentration gradients important for a variety of cell physiological functions. The tilt of the transmembrane helix of the $\beta 2$ subunit, which is close to N118, N153, and N238 glycosites, mediates functional differences in the Na/K-ATPase complex,⁵⁶ suggesting that various glycans at these sites also have the potential to alter function via conformational changes. The $\beta 1$ subunit of Na/K-ATPase is also glycosylated (all three known sites are also characterized in this dataset), but the importance of specific glycosites is less pronounced in $\beta 1$ versus $\beta 2$ subunits,^{57,58} highlighting the differential roles glycosylation heterogeneity can play even within isoforms of the same complex.

Next we examined glycosylation profiles of glycoproteins in different cellular components (CC) (**Figure 6**). Each of the sixteen different subcellular groups in **Figure 6a** has edges connecting to glycan nodes that are arranged in a circle based on glycan type. A striking feature of this analysis is the increased level of glycan diversity at glycosites in the plasma membrane, other membranes, and extracellular proteins, where glycosites have noticeably more sialylated glycans. Other interesting trends arise, such as the presence of a relatively high occurrence of mannose-6-

phosphate (M6P) in lysosomal proteins. Note, this is expected because of the role of M6P in trafficking proteins to the lysosome for degradation, and this data serves as an internal control to support our approach of analyzing glycosylation profiles. Some trends match those reported by Medzihradszky et al. for cellular compartment in their glycoproteomic comparison of mouse brain and liver glycosites,⁵³ including high mannose glycans in secreted and ER glycoproteins.

To compare glycosylation profiles we calculated a Euclidean distance between each subcellular group (**Figure 6b**), where darker color indicates more similar (i.e., closer) glycosylation profiles. The most similar subcellular groups were plasma membrane/membrane, and also synapse/plasma membrane/membrane groups. Other closely related groups included vesicle glycoproteins and other cell surface-related subcellular locations (groups 1-7), while lysosomal glycoproteins were most closely related vesicles but not many other groups. Among other patterns, secreted proteins were most similar to Golgi and ER groups, and ER glycoproteins had the shortest Euclidean distance to Golgi glycoproteins. That said, the Golgi was related to more groups beyond just ER glycoproteins, and the “none listed” group had the most similarity to Golgi glycoproteins. This is perhaps unsurprising, as the Golgi is central to most glycosylation processing for proteins trafficked to the cell surface while ER glycosylation pathways are often followed by further processing in the Golgi.

One shortcoming of this approach to analyzing subcellular localization is the presence of several GO CC terms for a single UniProt entry. **Figure 6c** displays how many proteins mapped to a given number of subcellular groups based on their GO CC terms, and **Supplemental Figure 22** shows the proportion of proteins in each subcellular group that mapped to “other groups”. A more robust analysis would require subcellular fractionation during sample preparation and/or the use of proximity labeling strategies to investigate the glycoproteomes of each cellular component individually. These strategies present a challenge because of low amounts of starting material for subsequent steps, but coupling intact glycopeptide characterization to these subcellular location methods will be a worthwhile endeavor in future experiments to gain a more refined understanding of glycoproteome organization. Note, Thaysen-Andersen and co-workers have performed such subcellular fractionation analyses with some success using a combination of glycomic and proteomic approaches.⁵⁴

Finally, we conducted analysis of protein domains and their characteristic glycosylation profiles (**Figure 7**). Glycosites were mapped to protein domains to which they belong using information

available in UniProt. In total 745 of the 1,545 glycosites could be mapped to domains. The top bar graph (dark blue) shows the number of glycosites mapping to a given domain type, with the heat map above it (orange) showing the percent of a given domain that was seen as glycosylated relative to the total number of domains present in the mouse proteome. The grey bar graph compares glycan heterogeneity ratios for domains compared to the ratio for all 1,545 glycosites. The ratio is number of glycan-glycosite combinations (i.e., a glycosite site with three different glycans would count as a three) compared to the number of glycosites. Thus, a higher ratio indicates a larger amount of glycan heterogeneity for glycosites in that domain. The heat map on the bottom indicates differences in glycan types observed at sites within a domain type compared to the distribution of glycan types seen in all 1,545 sites. Here, a difference of zero shows that the proportion of glycosites containing a given glycan type is equivalent to the overall proportion observed for all glycosites, whereas positive or negative values indicate that a higher or lower proportional contribution, respectively, of a given glycan type at the glycosites mapped to that domain. Note, only domains with 7 or greater glycosites are delineated here, with all other domains grouped into the “other domains” category, which shows little difference in glycan type expression from the total number of glycosites.

Of the 745 sites that could be mapped to a domain, 197 of them existed within an immunoglobulin (Ig)-like domain, where glycosites had a slightly higher proportion of fucosylated and sialylated glycans relative to all glycosites. Peptidase, EGF-like, and Ig-like domains tended to have glycosites that contained more diverse glycan types, while glycosites observed in Sushi, CUB, Laminin, Cadherin, and Sema domains had lower glycan heterogeneity and contained a high proportion of high mannose glycans. Glycosites in fibronectin and EGF-domains harbored proportionally high amounts of fucosylated glycans whereas the majority of other domain types did not. Interestingly, glycosites in peptidase domains have a relatively higher contribution from M6P, which correlates with the known lysosomal targeting role of that glycan. Lee et al. suggested previously that differences in glycosylation profiles can be explained by differential solvent accessibility of glycosites (which they link to differences in subcellular glycosylation profiles).⁵⁴ This presents an intriguing future avenue to explore for domain-specific glycotypes, although the integration of proteomic and glycomic data (as Lee et al. performed) with intact glycopeptide analysis (as is provided here) is likely needed for such an investigation.

For the majority of domains, only a fraction of the total known number of domains across the entire mouse proteome were detected to contain glycosites in this study (~6% or less of any given

domain), except for Sema domains where 25 glycosites were observed from only 99 known Sema domains. The Sema domain exists in a large family of secreted and transmembrane proteins called semaphorins which can function in axon guidance, and the majority of glycosites in this dataset that are in Sema domains contained high mannose glycans. That said, investigating glycosites in various protein domains in this study is limited by the information available in UniProt, but it represents an intriguing perspective for future large-scale glycoproteomic studies.

DISCUSSION

In all, the AI-ETD method presented here is a straightforward approach to improve glycopeptide fragmentation by combining the strengths of electron-driven dissociation and vibrational activation to access information about both peptide and glycan moieties simultaneously. AI-ETD enabled the most in-depth glycoproteome profiling of a single tissue to date and this strategy is amenable to practically any biological system. Ultimately, this study demonstrates that >1,500 N-glycosites can be characterized via intact glycopeptide analysis from a single tissue, marking a new era of robust glycoproteome characterization that will enable much-needed large-scale studies to investigate the role of glycosylation in various biological systems.

Further studies will be needed to explore the utility of AI-ETD for glycopeptides with more than one glycosite, such as those encountered in middle-down and top-down glycoproteomic experiments.²⁴ Assigning correct glycan modifications for multiply glycosylated peptides poses significant challenges, so we excluded all glycopeptide identifications that harbored more than one glycan in this dataset to ensure higher quality identifications. The middle-down approach can add considerable information to glycoforms and co-occurring glycans, but middle-down methods typically use specifically developed proteolytic and chromatographic methods. Electron-driven dissociation methods have been valuable in middle-down glycoproteomic methods,²⁴ so it is reasonable to suggest that AI-ETD may prove useful in characterizing multiply glycosylated peptides and proteins as well.

Another caveat of any glycoproteomic experiment is that there is not a “universal” or “ideal” glycopeptide enrichment method.²¹ This is markedly different from other PTM-centric proteomic methodologies. Lectin-based methods tend to have high enrichment yields (high percentage of glycopeptides compared to remaining non-modified peptide background), but lectins have glycan specificities that make them better suited for certain glycopeptides/glycan classes than others.⁵⁹

Hydrophilic interaction liquid chromatography (HILIC) and electrostatic repulsion hydrophilic interaction chromatography (ERLIC) have also been successfully explored as glycopeptide enrichment methods.^{12,21,30} ERLIC-based methods show the most promise for applicability to a broad range of glycan classes, but they can have a high background of non-modified peptides present post-enrichment (likely because of charged moieties on peptides that cause their retention on ERLIC material). We relied on Concanavalin A (ConA) lectin for enrichment in this study, meaning there are some limitations in the range of glycan classes observed. ConA binds oligomannose-type N-glycans with high affinity (which includes hybrid-type N-glycans), but is also known to bind complex-type N-glycans, albeit it with lower affinity.⁵⁹ Thus, there is a bias toward oligomannose-type glycans to consider in this dataset. Even with this, however, we do characterize a diverse pool of N-glycans and provide evidence of varying degrees of heterogeneity at the glycosite, glycoprotein, and subcellular location levels across the glycoproteome as discussed above. Furthermore, we also see many similar trends to other studies that used different enrichment methods. A prevalence of high mannose structures was seen in early glycomics studies of rodent brain⁵¹ and has been noted in glycoproteomic studies of rodent brain tissue by Trinidad et al. and Medzihradszky et al. with a lectin-based approaches^{27,53} and Liu et al. with zwitterionic-HILIC methods.³⁰ Woo et al. also noticed a significant degree of oligomannose glycopeptides even with their chemical-tag-based enrichment (although in human cell lines instead of rodent brain tissue), which enriches glycopeptides based on clickable metabolically-incorporated sugars.²⁰ This makes our observations of a high degree of oligomannose glycopeptides, which is likely due to the use of ConA for enrichment in part, still in congruence with observations using several other enrichment strategies. Current and future experiments in our group are exploring combinations of lectin-based approaches with HILIC and ERLIC methods to observe an even broader scope of the glycoproteome.

Profiling the glycoproteome at this depth also requires new ways to interpret complex data that comes from intact glycopeptide analysis. While others have commented on similar trends in smaller-scale datasets, e.g., glycosylation differences in cellular compartments or the observation of varying degrees of heterogeneity on the same protein, we can now comment on trends across more than a thousand glycosites with the data presented here. We present several new ways to analyze and visualize large-scale glycoproteomic data, providing a new perspective into the site-specific microheterogeneity of protein N-glycosylation at a systems level. We also show that glycosylation profiles differ based on subcellular localization and protein domain types and that heterogeneity can present itself in many different forms that can even differ between glycosites

on the same protein. This work underscores the value of intact glycopeptide analysis to capture this complexity and provides an avenue forward to continue advancing our understanding of protein glycosylation.

METHODS

Sample preparation and glycopeptide enrichment. A whole mouse brain was homogenized in lysis buffer (8M Urea, 50 mM tris) using a probe sonicator, and protein concentration was determined using a BCA Protein Assay Kit (Thermo Pierce, Rockford, IL). Brain tissue was harvested from C57BL/6J adult female mice after euthanasia and immediately frozen in liquid nitrogen. Mice were housed at the University of Wisconsin-Madison, and all experiments were performed in accordance with the National Institute of Health Guide for the Care and Use of Laboratory Animals and approved by the Animal Care and Use Committee at the University of Wisconsin-Madison. Tryptic digestion was performed similarly as described elsewhere.⁶⁰ Briefly, 4 mg of mouse brain lysate was brought to 90% methanol by volume, and proteins were precipitated by centrifugation for 5 minutes at 12,000 G. The supernatant was discarded, and the resultant protein pellet was dissolved in 8 M urea, 10 mM tris(2-carboxyethyl)phosphine (TCEP), 40 mM chloroacetamide (CAA) and 100 mM tris pH 8. The sample was diluted with 100 mM tris to a final urea concentration of 1.5 M urea and digested with trypsin (Promega, Madison, WI) overnight at room temperature (1:50, enzyme/protein). Peptides were desalted using Strata X columns (Phenomenex Strata-X Polymeric Reversed Phase). Following desalting, peptides were resuspended in 10 mM HEPES, 150 mM NaCl (wash buffer) and were split into 4 equal mass equivalents. Each aliquot of tryptic peptide mixture was enriched for glycopeptides using 150 μ L of agarose-bound concanavalin A (ConA) (Vector Laboratories, Burlingame, CA). The ConA solution was added to the SigmaPrep spin column (Sigma-Aldrich, St. Louis, MO, USA) and washed with 500 μ L of wash buffer three times. The agarose-bound ConA was then suspended in 500 μ L wash buffer and transferred to the mixture of tryptic peptides, and rotated at ambient temperature for 2 hours. The sample was then washed five times with 500 μ L wash buffer and glycopeptides were eluted with two 500 μ L washes of elution buffer (0.2% TFA in water). Eluted glycopeptide enriched material from all four equivalents were desalted and combined. For deglycan analyses, 20% of the total mixture was incubated at 37 °C with PNGaseF and fractionated into 16 high pH reversed phase fractions using a 1260 Infinity II HPLC (Agilent Technologies, Santa Clara, CA) with a 4.6 x 150 mm XBridge C18 column and a 30 minute gradient (mobile phase A: 10 mM ammonium formate pH 10, mobile phase B: 80% ACN, 10 mM ammonium

formate pH 10). The remaining 80% of the glyco-enriched sample was fractionated into 12 fractions

LC-MS/MS. All samples were injected onto and separated over an in-house fabricated 75/360 μm I.D/O.D. bare fused silica capillary with an integrated nanoelectrospray tip and packed 30 cm with 1.7 μm , 130 Å, BEH C18 particles (Waters). The mobile phases (A: 0.2% formic acid and B: 80% acetonitrile/0.2% formic acid) were driven and controlled by a Dionex Ultimate 3000 RPLC nano system. For intact glycopeptide analyses, two microliters of glycopeptides (10% of total sample) were injected onto the column and gradient elution was performed at 325 nL/min, B was increased to 4% over 6 min, followed by an increase to 53% at 75 min, a ramp to 99% B at 76 min, and a wash at 99% B for 3 min. The column was then re-equilibrated at 0% B for 10 min, for a total analysis of 90 minutes. All MS data was collected on a quadrupole-Orbitrap-linear ion trap hybrid MS system (Orbitrap Fusion Lumos, Thermo Fisher Scientific) modified to perform AI-ETD as previously described.³⁵ Precursors were ionized using a nanoelectrospray source held at +2 kV compared to ground and the inlet capillary temperature was held at 275 °C. Survey scans of peptide precursors were collected from 350-1800 Th with an AGC target of 400,000, a maximum injection time of 50 ms, and a resolution of 120,000 at 200 m/z. Monoisotopic precursor selection was enabled for peptide isotopic distributions, precursors of $z = 2-8$ were selected for data-dependent MS/MS scans for 3 seconds of cycle time, and dynamic exclusion was set to 60 seconds with a ± 10 ppm window set around the precursor. Priority was given for higher precursor charge states and lower precursor m/z values, and an isolation window of 2 Th was used to select precursor ions with the quadrupole. MS/MS scans were collected in a higher-energy collision dissociation-product dependent-activated ion electron transfer dissociation manner (HCD-pd-AI-ETD),^{43,44} where MS/MS scans were collected using HCD at 28 normalized collision energy (nce) with an AGC target of 50,000 and a maximum injection time of 60 ms, and product ions were mass analyzed in the Orbitrap with a resolution of 30,000 at 200 m/z. If oxonium ions 204.0867, 138.0545, or 366.1396 were present in the top 20 fragment ions, an AI-ETD scan was triggered for that precursor ion. For the triggered AI-ETD scans, calibrated charge dependent ETD parameters were enabled to determine ETD reagent ion AGC and ETD reaction times,⁶¹ a laser power of 18 W was used, the laser was on for the duration of the ETD reaction only (adding no time to the standard ETD scan cycle), and product ions were analyzed in the Orbitrap with a resolution of 30,000 at 200 m/z. Each fraction was analyzed in triplicate for AI-ETD data. For oxonium ion analysis for glycan isomer differentiation, the scan range was set from 115-2000 Th for AI-ETD, and only one replicate was collected for each fraction. For ETD and EThcD analyses

for comparisons, all settings were the same, with EThcD having a supplemental activation of 25 nce. De-glycopeptides were analyzed using the same chromatography conditions, but the MS acquisition differed. Precursors were ionized using a nanoelectrospray source held at +2.2 kV compared to ground and the inlet capillary temperature was held at 275 °C. Survey scans of peptide precursors were collected from 300-1350 Th with an AGC target of 500,000, a maximum injection time of 50 ms, and a resolution of 240,000 at 200 m/z. Monoisotopic precursor selection was enable for peptide isotopic distributions, precursors of $z = 2-5$ were selected for data-dependent MS/MS scans for 1 second of cycle time, an isolation window of 2 Th was used to select precursor ions with the quadrupole, and dynamic exclusion was set to 20 seconds with a ± 10 ppm window set around the precursor. Precursors were fragmented using HCD at 30 nce and MS/MS scans were performed in the ion trap using the “rapid” scan rate over a 200-1400 Th range. The AGC target was 30,000 and the maximum injection time was 22 ms.

Data analysis. A focused protein database was created from the de-glycopeptide data by searching the data with Byonic.⁶² A mass tolerance of ± 10 ppm was used for precursors, monoisotopic mass tolerance was set to ± 0.4 Da for product ions, and HCD fragmentation type was selected. Oxidation of methionine and deamidation of asparagine were specified as variable modifications, while carbamidomethylation of cysteine was a set as a fixed modification. Trypsin specificity allowing for ≤ 3 missed cleavages was used and spectra was searched against a UniProt mouse (*mus musculus*) database (canonical and isoforms) downloaded on May 12, 2016, which was concatenated with a reversed sequence version of the forward database. Results were filtered at a 1% protein FDR, and a focused database was made of proteins that were identified with at least one peptide that had both a deamidated asparagine modification and the N-glycosylation sequon (N-X-S/T, where X is any residue but proline) present (a total of 3,574 proteins). Intact glycopeptide data were also searched with Byonic by converting .raw files to .mgf files using MSConvert.⁶³ The focused protein database described above and a glycan database of 182 mammalian N-glycans compiled from literature sources were used.^{27,51,52,64} N-glycosylation was set as a variable modification, and each glycan was only allowed to be used once per identified peptide (common1 setting). Oxidation of methionine was set as a common variable modification (common2) and conversion of glutamine and glutamate residues to pyroglutamate were set as rare variable modifications (rare1). A total of three common and one rare modification were allowed per identification, and carbamidomethylation of cysteine was a set as a fixed modification. Trypsin specificity was used with three missed cleavages allowed, and the mass tolerance settings were ± 10 ppm and ± 20 ppm for precursor and product ions, respectively. The

fragmentation type was set to EThcD (no AI-ETD option exists). Results were filtered at a 1% protein FDR as set in the Byonic parameters, and data was further processed using in-house scripts written in C#, some of which used the C# Mass Spectrometry Library (CSMSL, <https://github.com/-dbaileychess/CSMSL>). Our post-processing steps included: manual filtering to 1% false discovery rate (FDR) at the peptide spectral match level using the 2D-FDR score⁶² (Byonic typically retains identifications that are above the 1% FDR cutoff set in the Byonic software but pass protein FDR, especially for glycopeptides,³⁰ which necessitates this step); removing identifications that had a Byonic Score below 150 (as suggested by Lee et al.⁶⁵); setting a threshold for peptide length at 5 residues or greater; and retaining glyco PSMs that had $|\log\text{Prob}|$ value above 1 (which is the absolute value of the log base 10 of the protein p-value). This allowed for an estimated 0.33% FDR at the glycopeptide spectral match level (i.e., specifically counting the number of target and decoy hits that are glycopeptides, *not* including non-modified sequences). AI-ETD and HCD spectra had estimated FDRs of 0.07% and 0.59%, respectively. Furthermore, we removed glycopeptide identifications that contained more than one glycosite (because of known issues with properly assigning modifications in multiply glycosylated peptides²⁴). A further filtering step was added that only allowed for identifications with a Delta Mod Score of 10 or greater, which removed all decoy hits, and this pool of filtered identifications comprises the reported identifications in the manuscript. Note, Byonic only considers glycopeptide identifications for peptides with the N-X-S/T sequon. It is important to note that glycopeptide identification remains challenging with automated methods due to difficulties associated with calculating accurate FDRs.²³ Using the six steps of filtering here was our attempt to best control false discovery rates in large-scale glycopeptide analyses, although this is still an area of development in the field. Our 0.33% estimated FDR prior to our final DeltaMod score filtering, which left no decoy hits in the final dataset used, indicates a promising level of FDR mediation. That said, the data presented here are still subject to the challenges of glycopeptide FDR estimation. **Supplemental Figure 3** provides a distribution of identified glycopeptide masses in addition to the masses of the peptide and glycan moieties separately. Note, structural information about glycans cannot yet be reliably discerned from AI-ETD spectra (although this is under investigation, as noted in the Results section), so we report only compositional information about glycans here (as is standard in high-throughput intact glycopeptide analyses). Overlap in glycosites from this study and two deglycoproteomic experiments were performed from Zielinska et al. and Fang et al. studies.^{66,67} Gene ontology enrichment analysis was performed using DAVID⁶⁸ with a mouse brain proteome as the background.⁶⁹ The protein-glycan network, glycan co-occurrence networks, and glycosylation profiles for subcellular groups were created in R 3.2.2

using the igraph library,⁷⁰ and the arcplot were created with arcdiagram library (Sanchez, G. arcdiagram: Plot pretty arc diagrams. package ver. 0.1.11., <http://www.gastonsanchez.com>, 2014). When grouping glycan types, any glycan with a NeuAc moiety was categorized as sialylated, meaning some glycans in the sialylated group are also fucosylated. Thus the fucosylated glycan type group contains any glycan that contains a fucose moiety and also is not sialylated. Furthermore, this means that glycans in the complex/hybrid glycan class are neither fucosylated nor sialylated. For grouping subcellular locations from GO cellular component terms, GO terms were collapsed into a smaller number of subcellular locations displayed in **Figure 6** and **Supplemental Figure 22** using the following: plasma membrane was assigned only if the GO term specifically matched “plasma membrane”; other membrane includes any GO term with the word membrane that did not include the plasma membrane; neural includes GO terms that contain “axon”, “neuro”, or “myelin”; Golgi includes terms that contain “Golgi” and do not contain “endoplasmic” while “ER” contains any term that contains “endoplasmic”; cell surface includes any GO term that includes “surface”; and “other cellular component” includes any GO term that did not map to the other 11 subcellular groups used. Euclidean distances were calculated between subcellular groups by considering each group as a 117-dimension vector (each glycan is a direction) with the number of occurrences as the magnitude. All domain information was also retrieved from the UniProt database. When mapping glycosites to protein domains, domains were collapsed to the broadest common term possible, e.g., “Ig-like”, “Ig-like C1-type”, “Ig-like C2-type, and “Ig-like V-type” were all categorized as “Ig-like”. “Differences in percent composition” for glycan classes can be considered as “mean normalized” comparisons and were calculated by: 1) calculating a percentage of glycosites with a given glycan type compared to the total number of glycan type-glycosite combinations for all 1,545 sites (this is considered the mean value for each glycan type), and 2) comparing the ratios for each glycan type for a given domain to the mean value for that glycan type. Thus, summing all the differences across six glycan types for a given domain equals zero. Hierarchical clustering to provide the domain ordering was done using R 3.2.2, where each domain type was treated as a six-dimension vector with the difference in percent composition as magnitudes. Glycan heterogeneity ratios were calculated by dividing the total number of glycan type-glycosite combinations by the number of glycosites. The “percent domains with glycosite” values were calculated by summing the total number of known domains of a given type in the entire mouse proteome (from UniProt) and calculating a percentage based on the number of domains observed in this dataset to have a glycosite.

DATA AVAILABILITY

Raw data files (.RAW files) are available at online at the Chorus Project (chorusproject.org), Project ID: 1441. Supplemental data files including identified glycopeptides, glycoproteins, glycosites, glycans, glycoPSMS, and proteins included in the glycoproteomics focused protein database – as well as Supplemental Information, Tables, and Figures – are available in the online version of the paper. Freely available Byonic results files containing all identified and assigned spectra are available at the following link: <https://figshare.com/s/23abd7250324fbc81115>. Note, these results files will contain identifications that were filtered out of the final dataset presented in this manuscript (using the post-Byonic filtering steps indicated above). Only spectral matches indicated in the provided glycoPSMs supplemental file were included in the dataset presented here. We recommend setting the “Max number of peaks per 100 m/z” to >50 (under “Annotation Options”) to ensure annotation of all fragments in more complex spectra.

CODE AVAILABILITY

The C# Mass Spectrometry Library (CSMSL) is available at <https://github.com/-dbaileychess/CSMSL>.

ACKNOWLEDGEMENTS

We thank J. Saba, M. Bern, K. Schauer, and K. Overmyer for helpful discussions and N. Kwiecien and D. Bailey for assistance in data analysis. This work was supported by US National Institutes of Health (NIH) grant R35 GM118110 and P41 GM108538 to J.J.C. N.M.R. gratefully acknowledges support from an NIH Predoctoral to Postdoctoral Transition Award (F99 CA212454).

DISCLOSURE OF INTERESTS

J.J.C. is an inventor of electron transfer dissociation and is a consultant for Thermo Fisher Scientific.

AUTHOR CONTRIBUTIONS

N.M.R and A.S.H. designed and performed research. N.M.R. analyzed data. J.J.C. and M.S.W. supervised all steps of the project. N.M.R. and J.J.C. wrote the paper. Current affiliation for N.M.R. is Department of Chemistry, Stanford University, Stanford, CA 94305

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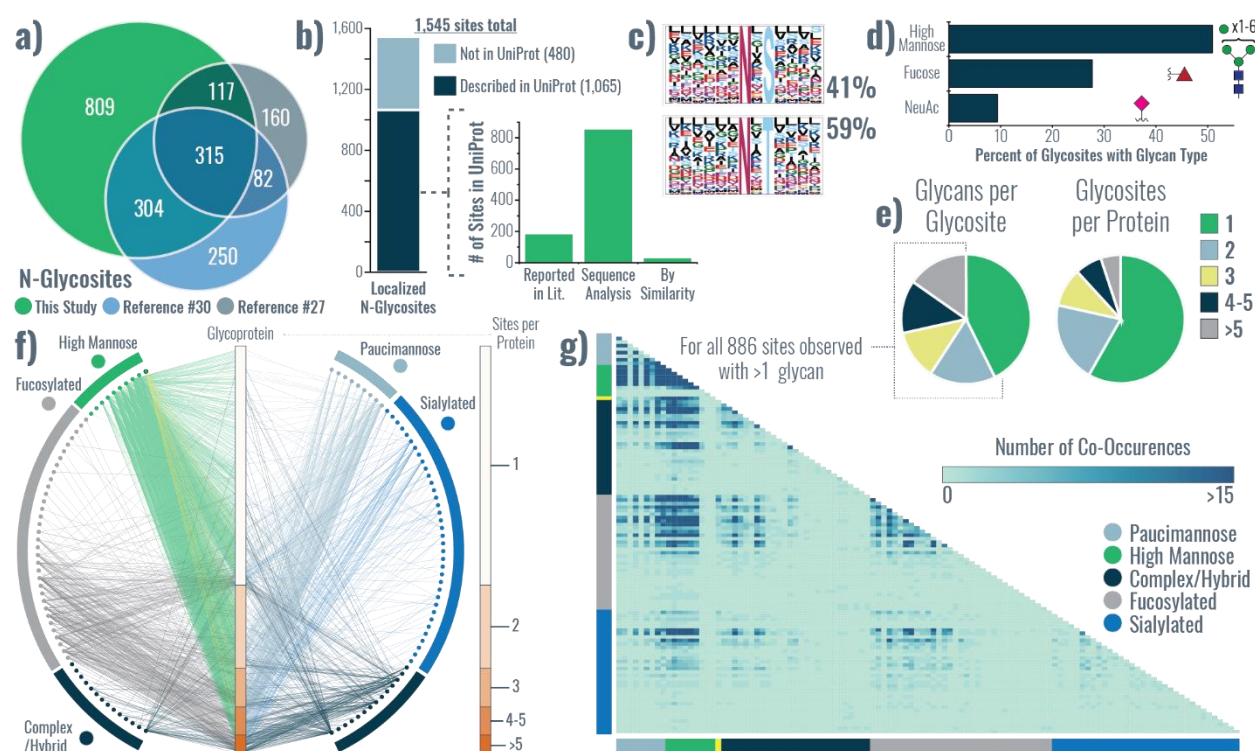


Figure 2. Characteristics of glycosites identified with AI-ETD. **a)** Overlap of mouse brain N-glycosites identified in this study with those from the Liu et al. Nat. Comm. 2017 (ref. #30) and Trinidad et al. MCP 2013 (ref. #27) studies. **b)** Approximately 69% of identified glycosites are described as known glycosites in the UniProt database, and the majority of them have that description based on sequence analysis (i.e., prediction of glycosite based on the presence of the N-X-S/T sequon). **c)** Sequence motifs for N-glycosites having either the N-X-S or N-X-T sequon and their relative percentage in the unique glycosites identified. **d)** Percentage of total glycosites that had glycans of high mannose type or that contained a fucose or NeuAc residue. **e)** Distribution of the number of different glycans seen at a given glycosite, i.e., the degree of glycan microheterogeneity (left), and the number of glycosites per glycoprotein identified (right). **f)** A glycoprotein-glycan network maps which glycans (outer circle, 117 total) modify which proteins (inner bar, 771 total). Glycoproteins are sorted by number of glycosites (scale to the right). Glycans are organized by classification, and edges are colored by the glycan node from which they originate, except for mannose-6-phosphate which has yellow edges. See supplemental information for grouping details. **g)** A heat map represents the number of times glycan pairs appeared together at the same glycosite, indicating which glycans contribute most to microheterogeneity of the >880 sites that had more than one glycan modifying them.

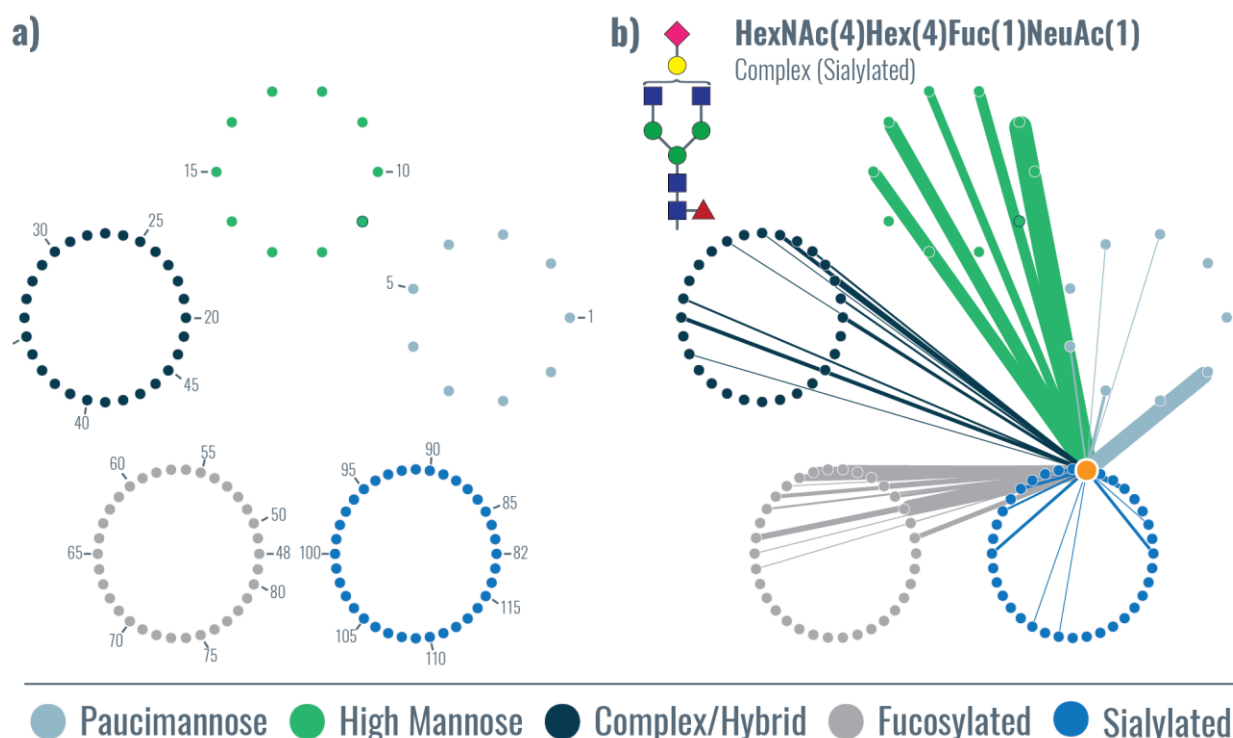


Figure 3. Glycan co-occurrence networks. a) The organization of the glycan co-occurrence network is given, where glycans are sorted into larger circles based on glycan type, each node is one of the 117 glycans identified, and the numbers indicate glycan identities given in **Supplemental Table 3**. Glycan 19 (green with dark blue border) indicates mannose-6-phosphate. b) The glycan co-occurrence network shows all the glycans that co-occurred with HexNAc(4)Hex(4)Fuc(1)NeuAc(1) (highlighted as an orange node, i.e., the source node), with the relative number of occurrences indicated by edge thickness. Edge color indicates the target node.

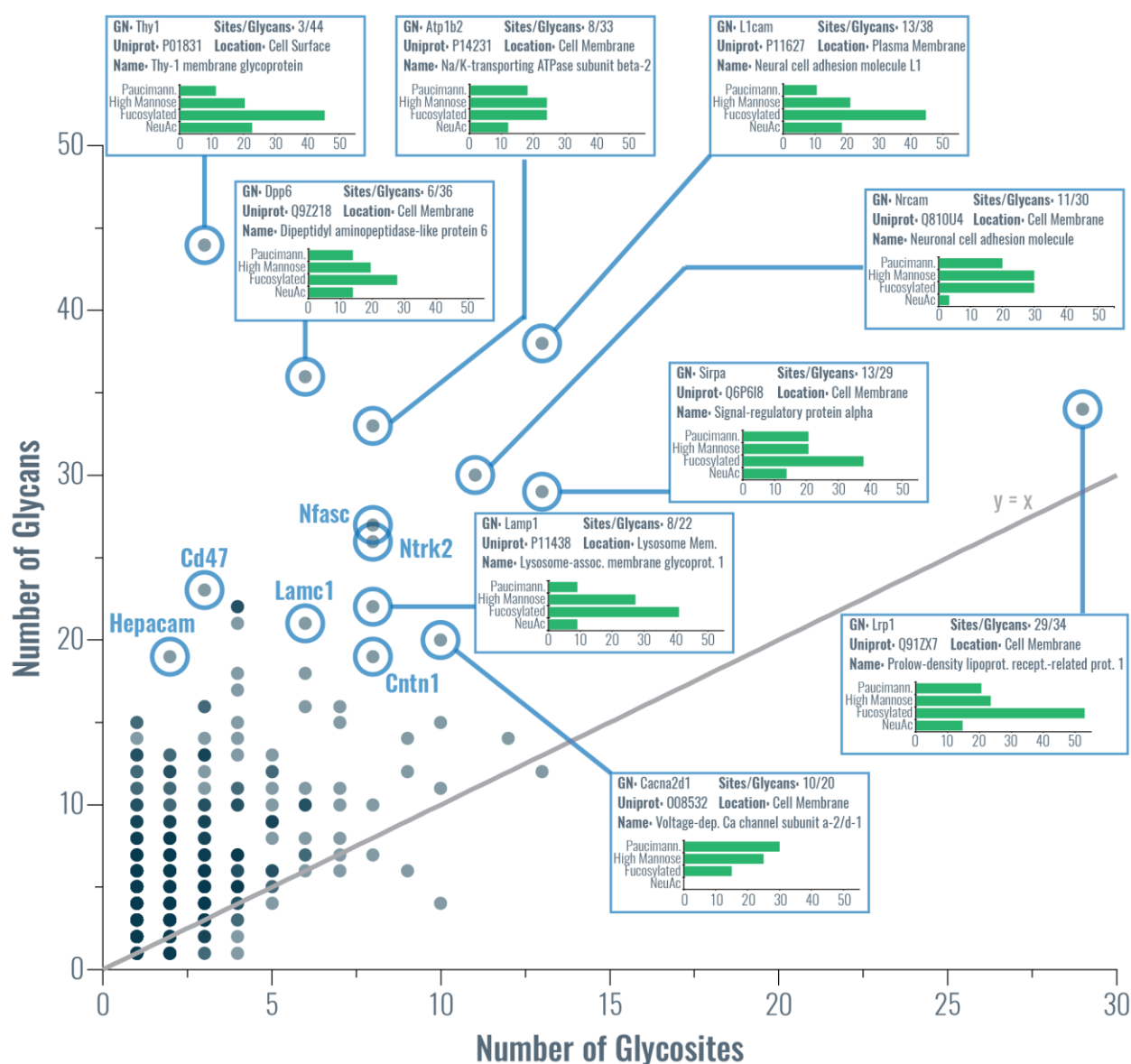


Figure 4. Glycan heterogeneity by glycoprotein. A scatter plot showing the number of glycans identified per glycoprotein vs. the number of glycosites identified for that protein summarizes a degree of glycosylation heterogeneity at the protein level. An $y=x$ line is shown in gray to provide a eye guide for proteins that had a particularly high number of glycans relative to the number of glycosites identified, some of which are highlighted. Boxes for highlighted proteins display gene name (GN), UniProt accession number, number of glycosites/glycans identified, the cellular location assigned to this protein, and a common name for the protein. Additionally, they provide a bar chart that displays the percentage of the total number of identified glycans that can be classified as paucimannose, high mannose, fucosylated, or sialylated (NeuAc). Note, if a paucimannose or sialylated glycan was fucosylated, it was also counted as fucosylated for this calculation. Gene names of other interesting proteins with high a glycan-to-glycosite ratio are also provided.

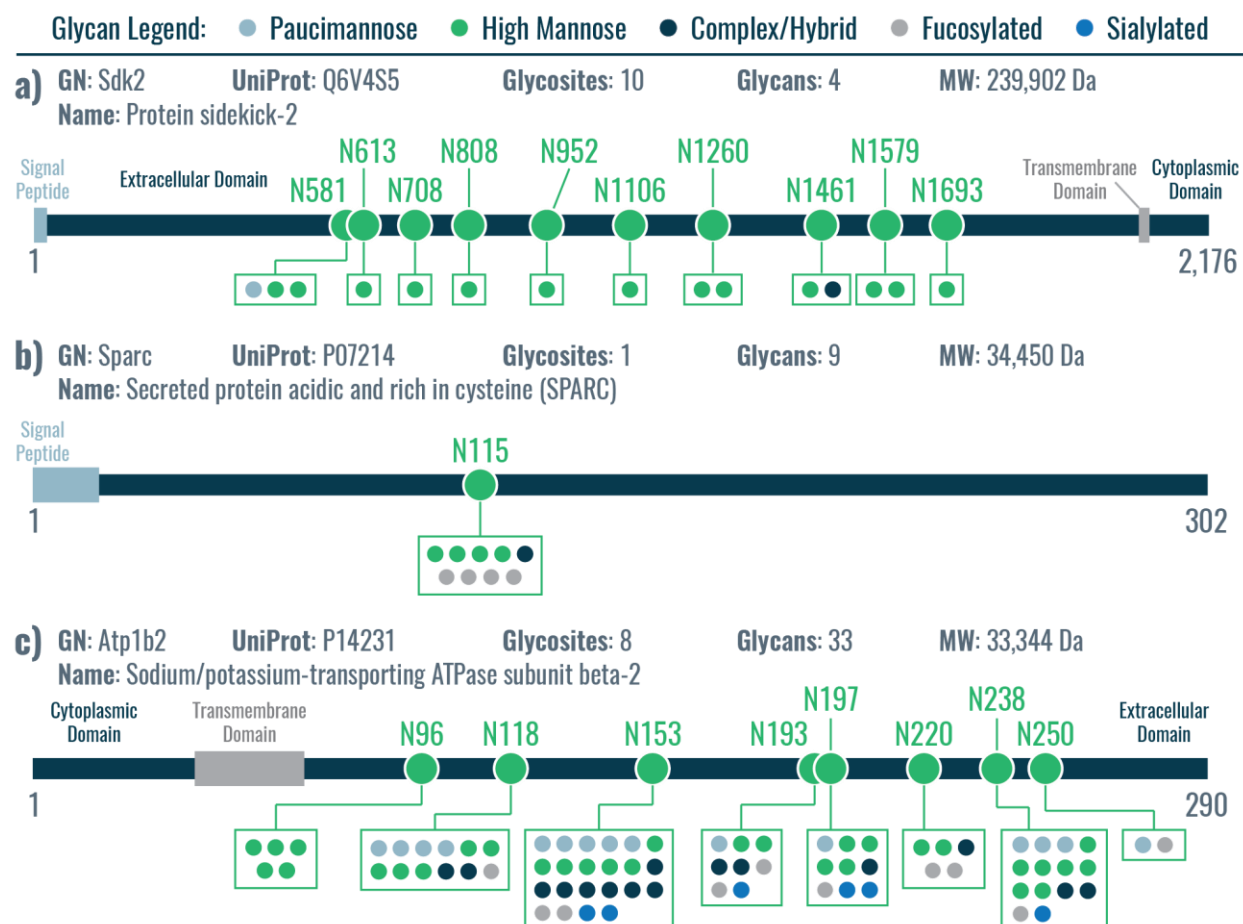


Figure 5. Glycan microheterogeneity can manifest in several different forms. Glycosites can have small or large degrees of glycan heterogeneity, and this level of glycan diversity can even differ for glycosites on the same protein. Here three examples of different modes of heterogeneity are provided: **a)** several glycosites on one protein with limited glycan heterogeneity (Protein sidekick-2), **b)** a protein with one glycosite that has some degree of heterogeneity (SPARC), and **c)** several glycosites on one protein that show either little or significant glycan heterogeneity (Na/K-transporting ATPase subunit beta-2).

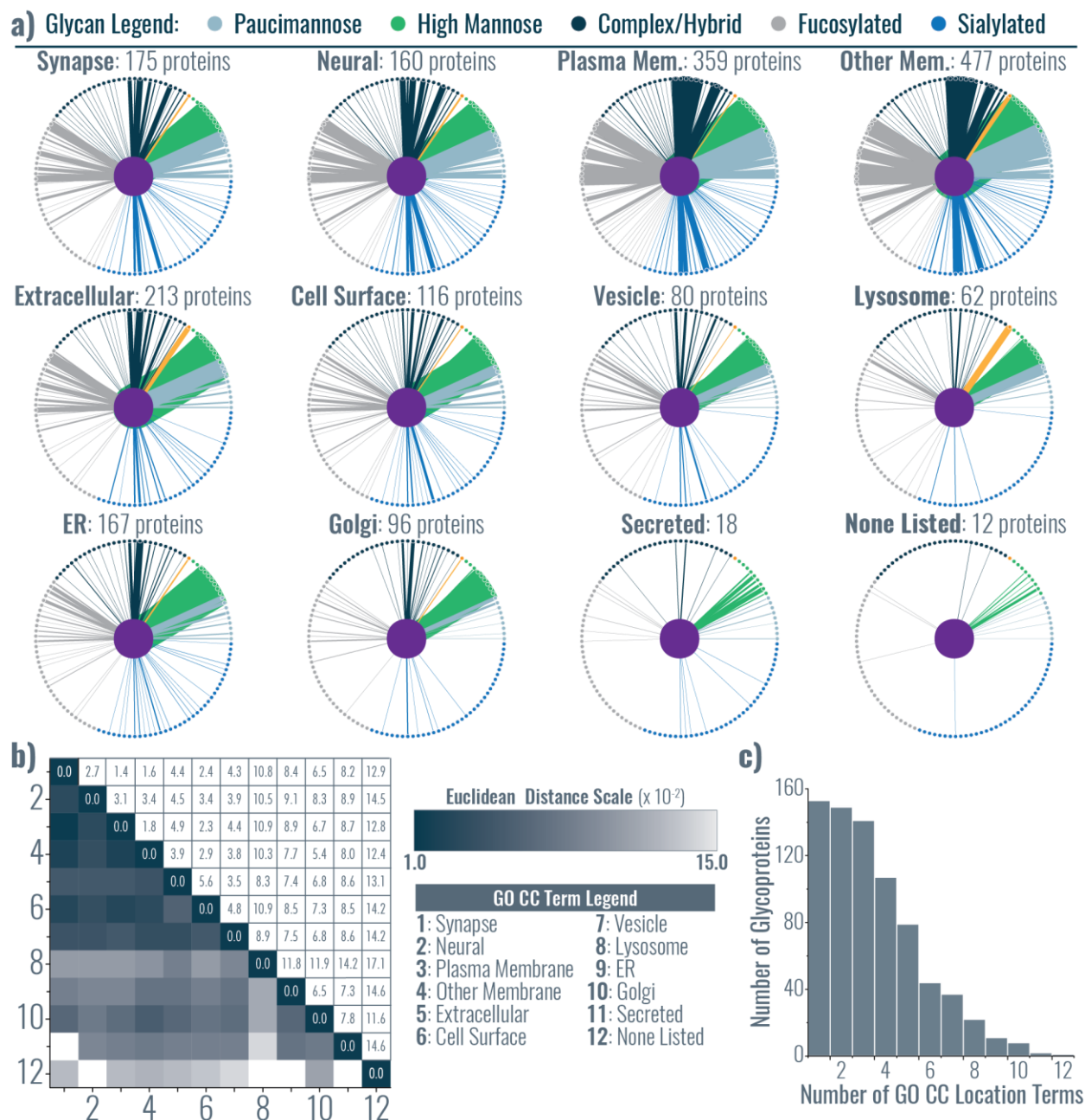


Figure 6. Delineating glycosylation profiles by subcellular cellular locations. a) Glycosylation profiles for glycoproteins from 12 subcellular locations (derived from GO cellular component terms) are shown, with colors indicating glycan type and line thickness indicating frequency. Orange denotes mannose-6-phosphate. **b)** Euclidean distances were calculated between each of the 12 subcellular localizations to indicate similarity in their glycosylation types (darker indicates a higher degree of similarity). **c)** Number of GO cellular component terms associated with identified glycoproteins.

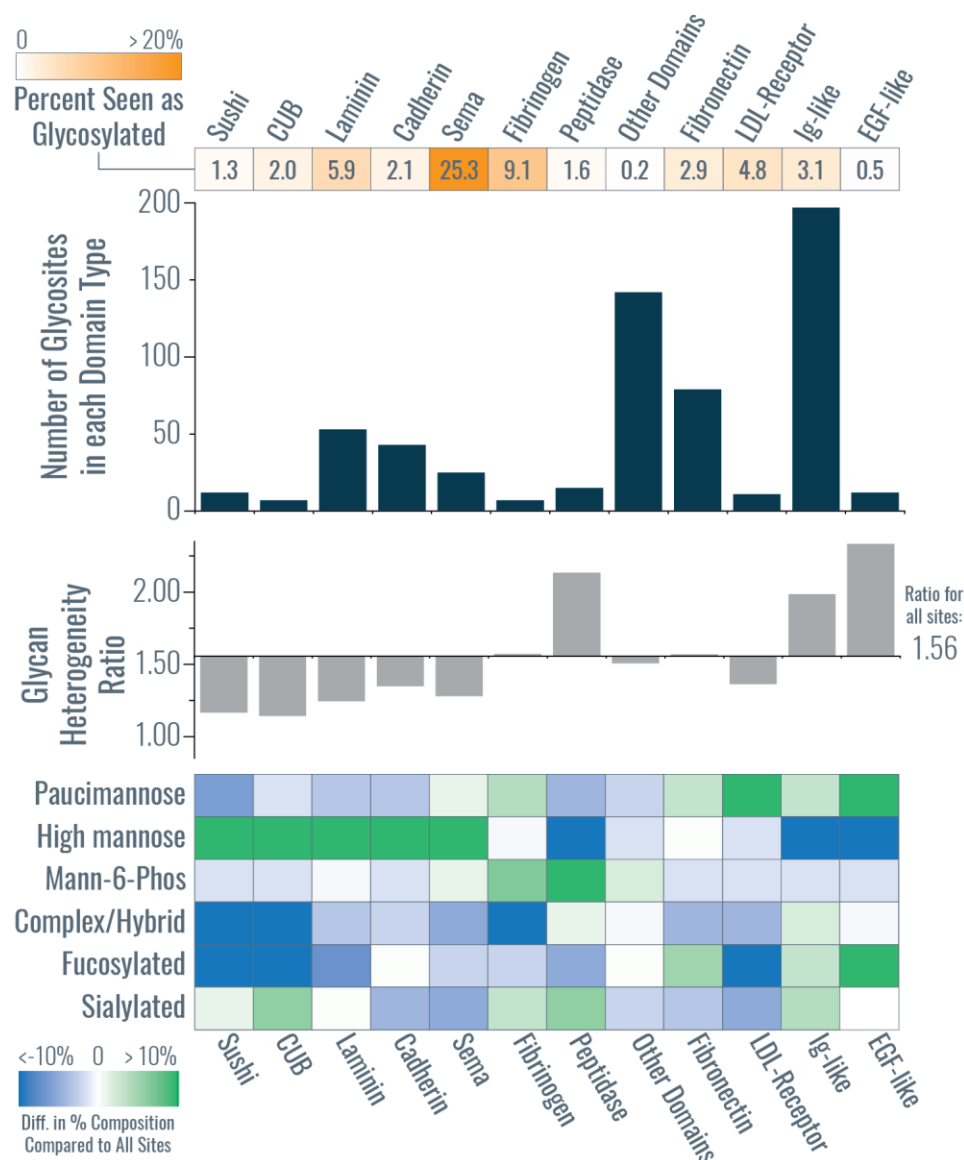


Figure 7. Mapping glycosites to protein domains. The number of glycosites mapping to a given domain and the percent of a given domain observed as glycosylated are provided in the dark blue bar chart and the orange heat map above it, respectively. The grey bar graph compares glycan heterogeneity ratios for domains compared to the ratio for all 1,545 glycosites (with an average ratio of 1.56 for all sites), and the heat map at the bottom indicates differences in glycan types observed at sites within a domain type compared to the distribution of glycan types seen in all 1,545 sites.

SUPPLEMENTAL MATERIAL

Capturing site-specific heterogeneity with large-scale N-glycoproteome analysis

Nicholas M. Riley,^{1,2} Alexander S. Hebert,¹ Michael S. Westphall,¹ Joshua J. Coon^{1-4*}

¹Genome Center of Wisconsin, Departments of ²Chemistry and ³Biomolecular Chemistry, University of Wisconsin-Madison, Madison, WI, 53706, USA

⁴Morgridge Institute for Research, Madison, Wisconsin, USA

*to whom correspondence should be addressed: jcoon@chem.wisc.edu

Supplemental Files (available for download)

Supplemental File 1. Glycopeptides.xlsx

Supplemental File 2. Glycoproteins.xlsx

Supplemental File 3. Glycosites.xlsx

Supplemental File 4. Glycans.xlsx

Supplemental File 5. GlycoPSMs.xlsx

Supplemental File 6. GlycoFocusedProteinDB.fasta

Raw data available on Chorus (Project ID 1441) (<https://chorusproject.org>). Freely available Byonic results files containing all identified and assigned spectra are available at the following link: <https://figshare.com/s/23abd7250324fbc81115>.

Supplemental Tables and Figures

Supplemental Table 1. Glycan identities for node labels from Supplemental Figure 11.

Supplemental Table 2. Glycan identities for the co-occurrence heat map from Figure 2g Supplemental Figure 12.

Supplemental Table 3. Glycan identities for node labels from Figure 3 and Supplemental Figure 13.

Supplemental Figure 1. Comparing ETD, ETD with supplemental activation using higher energy collisional dissociation (ETHcD) and AI-ETD.

Supplemental Figure 2. Distributions of fragment ions generated by AI-ETD (green) and HCD (dark blue).

Supplemental Figure 3. Percent of spectra containing specific oxonium and Y-type ions.

Supplemental Figure 4. Using oxonium ions to differentiate glycan isomers.

Supplemental Figure 5. HCD performance characteristics.

Supplemental Figure 6. Mass and m/z distributions of glycoproteomics data.

Supplemental Figure 7. Workflow for in-depth profiling of the glycoproteome via intact glycopeptides with AI-ETD.

Supplemental Figure 8. Comparison of glycosites to large-scale deglycoproteomic experiments.

Supplemental Figure 9. N-glycosites characterized in integrin alpha-1 (GN: Itga1, UniProt: Q3V3R4) in mouse brain.

Supplemental Figure 10. Gene ontology functional categories enriched terms for identified glycoproteins.

Supplemental Figure 11. A larger version of the glycoprotein-glycan map in Figure 2f.

Supplemental Figure 12. A larger version of the co-occurrence heat map in Figure 2g.

Supplemental Figure 13. Glycan co-occurrence networks for a specific glycan and a class of glycans.

Supplemental Figure 14. Arc plot representing glycan microheterogeneity for glycosites with two glycans per site.

Supplemental Figure 15. Arc plot representing glycan microheterogeneity for glycosites with three glycans per site.

Supplemental Figure 16. Arc plot representing glycan microheterogeneity for glycosites with four glycans per site.

Supplemental Figure 17. Arc plot representing glycan microheterogeneity for glycosites with five glycans per site.

Supplemental Figure 18. Mass differences between glycans that occupy the same glycosite.

Supplemental Figure 19. Protein sidekick-2: relatively high number of glycosites with relatively low glycan microheterogeneity.

Supplemental Figure 20. Secreted protein acidic and rich in cysteine (SPARC): only one glycosite with moderate glycan microheterogeneity.

Supplemental Figure 21. Sodium/potassium transporting-ATPase β 2 subunit (Atp1b2): glycosites with varying degrees of glycan microheterogeneity.

Supplemental Figure 22. Heat map of the number of subcellular groups (derived from GO cellular component terms) for identified glycoproteins.

Supplemental Table 1. Glycan identities for node labels from Supplemental Figure 11.

Node	Glycan
1	HexNAc(1)
2	HexNAc(2)
3	HexNAc(2)Fuc(1)
4	HexNAc(2)Hex(1)
5	HexNAc(2)Hex(1)Fuc(1)
6	HexNAc(2)Hex(2)
7	HexNAc(2)Hex(2)Fuc(1)
8	HexNAc(2)Hex(3)
9	HexNAc(2)Hex(3)Fuc(1)
10	HexNAc(3)Hex(4)Fuc(1)NeuAc(1)
11	HexNAc(3)Hex(4)Fuc(2)NeuAc(1)
12	HexNAc(3)Hex(4)NeuAc(1)
13	HexNAc(3)Hex(5)Fuc(1)NeuAc(1)
14	HexNAc(3)Hex(5)NeuAc(1)
15	HexNAc(3)Hex(6)Fuc(1)NeuAc(1)
16	HexNAc(3)Hex(6)NeuAc(1)
17	HexNAc(4)Hex(3)NeuAc(1)
18	HexNAc(4)Hex(4)Fuc(1)NeuAc(1)
19	HexNAc(4)Hex(4)NeuAc(1)
20	HexNAc(4)Hex(5)Fuc(1)NeuAc(1)
21	HexNAc(4)Hex(5)Fuc(1)NeuAc(2)
22	HexNAc(4)Hex(5)Fuc(2)NeuAc(1)
23	HexNAc(4)Hex(5)Fuc(3)NeuAc(1)
24	HexNAc(4)Hex(5)NeuAc(1)
25	HexNAc(4)Hex(6)Fuc(1)NeuAc(1)
26	HexNAc(4)Hex(6)NeuAc(1)
27	HexNAc(4)Hex(7)NeuAc(1)
28	HexNAc(5)Hex(3)Fuc(1)NeuAc(1)
29	HexNAc(5)Hex(4)Fuc(1)NeuAc(1)
30	HexNAc(5)Hex(4)NeuAc(1)
31	HexNAc(5)Hex(4)NeuAc(2)
32	HexNAc(5)Hex(5)Fuc(1)NeuAc(2)
33	HexNAc(5)Hex(5)Fuc(2)NeuAc(1)
34	HexNAc(5)Hex(6)NeuAc(2)
35	HexNAc(6)Hex(3)Fuc(1)NeuAc(1)
36	HexNAc(6)Hex(3)Fuc(1)NeuAc(2)
37	HexNAc(6)Hex(5)Fuc(1)NeuAc(2)
38	HexNAc(6)Hex(5)Fuc(2)NeuAc(1)
39	HexNAc(6)Hex(6)Fuc(2)NeuAc(1)

Node	Glycan
40	HexNAc(6)Hex(6)NeuAc(1)
41	HexNAc(6)Hex(7)Fuc(3)NeuAc(1)
42	HexNAc(6)Hex(7)NeuAc(3)
43	HexNAc(6)Hex(7)NeuAc(4)
44	HexNAc(6)Hex(9)Fuc(1)NeuAc(2)
45	HexNAc(7)Hex(8)Fuc(1)NeuAc(1)
46	HexNAc(3)Hex(3)
47	HexNAc(3)Hex(4)
48	HexNAc(3)Hex(5)
49	HexNAc(3)Hex(6)
50	HexNAc(4)Hex(3)
51	HexNAc(4)Hex(4)
52	HexNAc(4)Hex(5)
53	HexNAc(4)Hex(6)
54	HexNAc(4)Hex(7)
55	HexNAc(5)Hex(3)
56	HexNAc(5)Hex(4)
57	HexNAc(5)Hex(6)
58	HexNAc(5)Hex(8)
59	HexNAc(6)Hex(3)
60	HexNAc(6)Hex(4)
61	HexNAc(6)Hex(5)
62	HexNAc(6)Hex(7)
63	HexNAc(6)Hex(9)
64	HexNAc(7)Hex(3)
65	HexNAc(7)Hex(4)
66	HexNAc(7)Hex(6)
67	HexNAc(8)Hex(3)
68	HexNAc(8)Hex(4)
69	HexNAc(8)Hex(5)
70	HexNAc(8)Hex(8)
71	HexNAc(9)Hex(10)
72	HexNAc(9)Hex(3)
73	HexNAc(9)Hex(6)
74	HexNAc(2)Hex(4)Fuc(1)
75	HexNAc(2)Hex(5)Fuc(1)
76	HexNAc(2)Hex(6)Fuc(1)
77	HexNAc(3)Hex(3)Fuc(1)
78	HexNAc(3)Hex(4)Fuc(1)

Node	Glycan
79	HexNAc(3)Hex(4)Fuc(2)
80	HexNAc(3)Hex(5)Fuc(1)
81	HexNAc(3)Hex(6)Fuc(1)
82	HexNAc(4)Hex(3)Fuc(1)
83	HexNAc(4)Hex(4)Fuc(1)
84	HexNAc(4)Hex(4)Fuc(2)
85	HexNAc(4)Hex(5)Fuc(1)
86	HexNAc(4)Hex(5)Fuc(2)
87	HexNAc(4)Hex(6)Fuc(1)
88	HexNAc(4)Hex(6)Fuc(2)
89	HexNAc(4)Hex(7)Fuc(1)
90	HexNAc(5)Hex(3)Fuc(1)
91	HexNAc(5)Hex(3)Fuc(2)
92	HexNAc(5)Hex(4)Fuc(1)
93	HexNAc(5)Hex(4)Fuc(2)
94	HexNAc(5)Hex(5)Fuc(1)
95	HexNAc(5)Hex(6)Fuc(4)
96	HexNAc(5)Hex(8)Fuc(1)
97	HexNAc(6)Hex(4)Fuc(1)
98	HexNAc(6)Hex(4)Fuc(2)

Node	Glycan
99	HexNAc(6)Hex(6)Fuc(2)
100	HexNAc(6)Hex(7)Fuc(3)
101	HexNAc(7)Hex(3)Fuc(1)
102	HexNAc(7)Hex(7)Fuc(1)
103	HexNAc(8)Hex(5)Fuc(1)
104	HexNAc(8)Hex(9)Fuc(1)
105	HexNAc(9)Hex(3)Fuc(1)
106	HexNAc(9)Hex(6)Fuc(1)
107	HexNAc(9)Hex(9)Fuc(1)
108	HexNAc(2)Hex(12)
109	HexNAc(2)Hex(11)
110	HexNAc(2)Hex(10)
111	HexNAc(2)Hex(9)
112	HexNAc(2)Hex(8)
113	HexNAc(2)Hex(7)
114	HexNAc(2)Hex(6)
115	HexNAc(2)Hex(5)
116	HexNAc(2)Hex(4)
117	HexNAc(2)Hex(6)Phospho(1)

Supplemental Table 2. Glycan identities for the co-occurrence heat map from **Figure 2g**
Supplemental Figure 12.

Order#	Glycan
1	HexNAc(1)
2	HexNAc(2)
3	HexNAc(2)Fuc(1)
4	HexNAc(2)Hex(1)
5	HexNAc(2)Hex(1)Fuc(1)
6	HexNAc(2)Hex(2)
7	HexNAc(2)Hex(2)Fuc(1)
8	HexNAc(2)Hex(3)
9	HexNAc(2)Hex(3)Fuc(1)
10	HexNAc(2)Hex(4)
11	HexNAc(2)Hex(5)
12	HexNAc(2)Hex(6)
13	HexNAc(2)Hex(7)
14	HexNAc(2)Hex(8)
15	HexNAc(2)Hex(9)
16	HexNAc(2)Hex(10)
17	HexNAc(2)Hex(11)
18	HexNAc(2)Hex(12)
19	HexNAc(2)Hex(6)Phospho(1)
20	HexNAc(3)Hex(3)
21	HexNAc(3)Hex(4)
22	HexNAc(3)Hex(5)
23	HexNAc(3)Hex(6)
24	HexNAc(4)Hex(3)
25	HexNAc(4)Hex(4)
26	HexNAc(4)Hex(5)
27	HexNAc(4)Hex(6)
28	HexNAc(4)Hex(7)
29	HexNAc(5)Hex(3)
30	HexNAc(5)Hex(6)
31	HexNAc(5)Hex(8)
32	HexNAc(6)Hex(3)
33	HexNAc(6)Hex(4)
34	HexNAc(6)Hex(5)
35	HexNAc(6)Hex(7)
36	HexNAc(6)Hex(9)
37	HexNAc(7)Hex(3)
38	HexNAc(7)Hex(4)

Order#	Glycan
39	HexNAc(7)Hex(6)
40	HexNAc(8)Hex(3)
41	HexNAc(8)Hex(4)
42	HexNAc(8)Hex(5)
43	HexNAc(8)Hex(8)
44	HexNAc(9)Hex(10)
45	HexNAc(9)Hex(3)
46	HexNAc(9)Hex(6)
47	HexNAc(2)Hex(4)Fuc(1)
48	HexNAc(2)Hex(5)Fuc(1)
49	HexNAc(2)Hex(6)Fuc(1)
50	HexNAc(3)Hex(3)Fuc(1)
51	HexNAc(3)Hex(4)Fuc(1)
52	HexNAc(3)Hex(4)Fuc(2)
53	HexNAc(3)Hex(5)Fuc(1)
54	HexNAc(3)Hex(6)Fuc(1)
55	HexNAc(4)Hex(3)Fuc(1)
56	HexNAc(4)Hex(4)Fuc(1)
57	HexNAc(4)Hex(4)Fuc(2)
58	HexNAc(4)Hex(5)Fuc(1)
59	HexNAc(4)Hex(5)Fuc(2)
60	HexNAc(4)Hex(6)Fuc(1)
61	HexNAc(4)Hex(6)Fuc(2)
62	HexNAc(4)Hex(7)Fuc(1)
63	HexNAc(5)Hex(3)Fuc(1)
64	HexNAc(5)Hex(3)Fuc(2)
65	HexNAc(5)Hex(4)Fuc(1)
66	HexNAc(5)Hex(4)Fuc(2)
67	HexNAc(5)Hex(5)Fuc(1)
68	HexNAc(5)Hex(6)Fuc(4)
69	HexNAc(5)Hex(8)Fuc(1)
70	HexNAc(6)Hex(4)Fuc(1)
71	HexNAc(6)Hex(4)Fuc(2)
72	HexNAc(6)Hex(6)Fuc(2)
73	HexNAc(6)Hex(7)Fuc(3)
74	HexNAc(7)Hex(3)Fuc(1)
75	HexNAc(7)Hex(7)Fuc(1)
76	HexNAc(8)Hex(9)Fuc(1)

Order#	Glycan
77	HexNAc(9)Hex(3)Fuc(1)
78	HexNAc(9)Hex(6)Fuc(1)
79	HexNAc(9)Hex(9)Fuc(1)
80	HexNAc(3)Hex(4)Fuc(1)NeuAc(1)
81	HexNAc(3)Hex(4)Fuc(2)NeuAc(1)
82	HexNAc(3)Hex(4)NeuAc(1)
83	HexNAc(3)Hex(5)Fuc(1)NeuAc(1)
84	HexNAc(3)Hex(5)NeuAc(1)
85	HexNAc(3)Hex(6)Fuc(1)NeuAc(1)
86	HexNAc(3)Hex(6)NeuAc(1)
87	HexNAc(4)Hex(3)NeuAc(1)
88	HexNAc(4)Hex(4)Fuc(1)NeuAc(1)
89	HexNAc(4)Hex(4)NeuAc(1)
90	HexNAc(4)Hex(5)Fuc(1)NeuAc(1)
91	HexNAc(4)Hex(5)Fuc(1)NeuAc(2)
92	HexNAc(4)Hex(5)Fuc(2)NeuAc(1)
93	HexNAc(4)Hex(5)Fuc(3)NeuAc(1)
94	HexNAc(4)Hex(5)NeuAc(1)
95	HexNAc(4)Hex(6)Fuc(1)NeuAc(1)
96	HexNAc(4)Hex(6)NeuAc(1)
97	HexNAc(4)Hex(7)NeuAc(1)

Order#	Glycan
98	HexNAc(5)Hex(3)Fuc(1)NeuAc(1)
99	HexNAc(5)Hex(4)Fuc(1)NeuAc(1)
100	HexNAc(5)Hex(4)NeuAc(1)
101	HexNAc(5)Hex(4)NeuAc(2)
102	HexNAc(5)Hex(5)Fuc(1)NeuAc(2)
103	HexNAc(5)Hex(5)Fuc(2)NeuAc(1)
104	HexNAc(5)Hex(6)NeuAc(2)
105	HexNAc(6)Hex(3)Fuc(1)NeuAc(1)
106	HexNAc(6)Hex(3)Fuc(1)NeuAc(2)
107	HexNAc(6)Hex(5)Fuc(1)NeuAc(2)
108	HexNAc(6)Hex(5)Fuc(2)NeuAc(1)
109	HexNAc(6)Hex(6)Fuc(2)NeuAc(1)
110	HexNAc(6)Hex(7)Fuc(3)NeuAc(1)
111	HexNAc(6)Hex(7)NeuAc(3)
112	HexNAc(6)Hex(7)NeuAc(4)
113	HexNAc(6)Hex(9)Fuc(1)NeuAc(2)
114	HexNAc(7)Hex(8)Fuc(1)NeuAc(1)

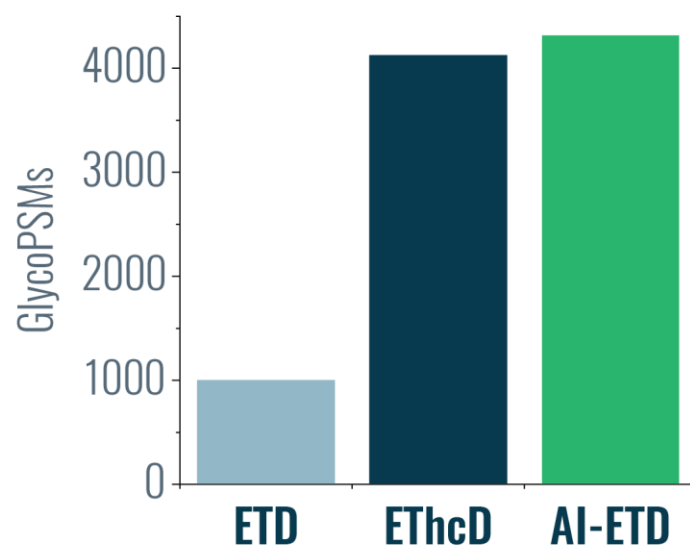
Supplemental Table 3. Glycan identities for node labels from Figure 3 and Supplemental Figure 13.

Node	Glycan
1	HexNAc(1)
2	HexNAc(2)
3	HexNAc(2)Fuc(1)
4	HexNAc(2)Hex(1)
5	HexNAc(2)Hex(1)Fuc(1)
6	HexNAc(2)Hex(2)
7	HexNAc(2)Hex(2)Fuc(1)
8	HexNAc(2)Hex(3)
9	HexNAc(2)Hex(3)Fuc(1)
10	HexNAc(2)Hex(4)
11	HexNAc(2)Hex(5)
12	HexNAc(2)Hex(6)
13	HexNAc(2)Hex(7)
14	HexNAc(2)Hex(8)
15	HexNAc(2)Hex(9)
16	HexNAc(2)Hex(10)
17	HexNAc(2)Hex(11)
18	HexNAc(2)Hex(12)
19	HexNAc(2)Hex(6)Phospho(1)
20	HexNAc(3)Hex(3)
21	HexNAc(3)Hex(4)
22	HexNAc(3)Hex(5)
23	HexNAc(3)Hex(6)
24	HexNAc(4)Hex(3)
25	HexNAc(4)Hex(4)
26	HexNAc(4)Hex(5)
27	HexNAc(4)Hex(6)
28	HexNAc(4)Hex(7)
29	HexNAc(5)Hex(3)
30	HexNAc(5)Hex(4)
31	HexNAc(5)Hex(6)
32	HexNAc(5)Hex(8)
33	HexNAc(6)Hex(3)
34	HexNAc(6)Hex(4)
35	HexNAc(6)Hex(5)
36	HexNAc(6)Hex(7)
37	HexNAc(6)Hex(9)
38	HexNAc(7)Hex(3)

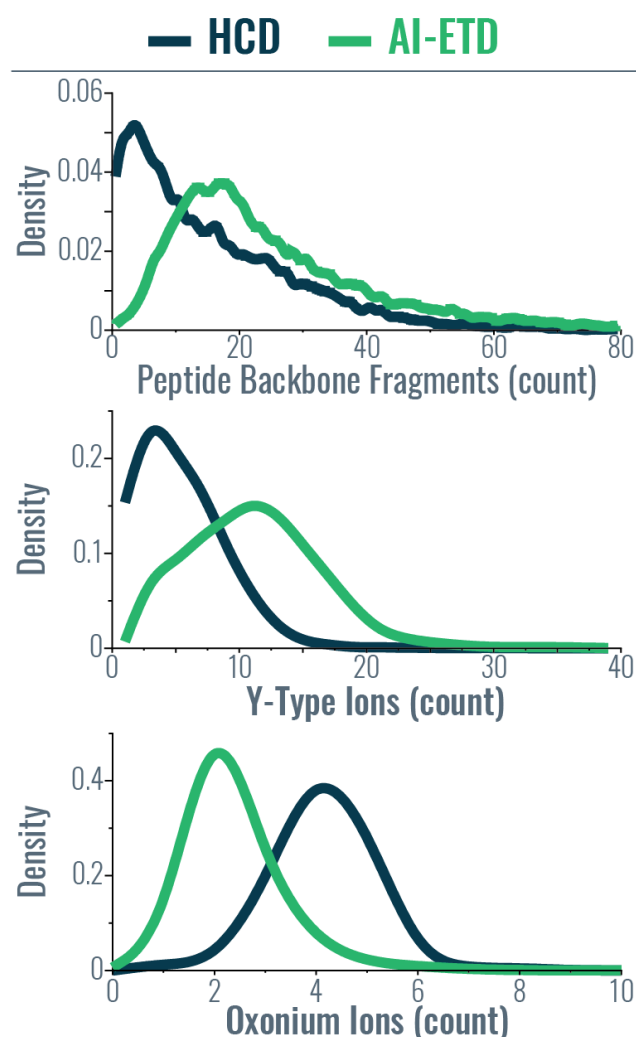
Node	Glycan
39	HexNAc(7)Hex(4)
40	HexNAc(7)Hex(6)
41	HexNAc(8)Hex(3)
42	HexNAc(8)Hex(4)
43	HexNAc(8)Hex(5)
44	HexNAc(8)Hex(8)
45	HexNAc(9)Hex(10)
46	HexNAc(9)Hex(3)
47	HexNAc(9)Hex(6)
48	HexNAc(2)Hex(4)Fuc(1)
49	HexNAc(2)Hex(5)Fuc(1)
50	HexNAc(2)Hex(6)Fuc(1)
51	HexNAc(3)Hex(3)Fuc(1)
52	HexNAc(3)Hex(4)Fuc(1)
53	HexNAc(3)Hex(4)Fuc(2)
54	HexNAc(3)Hex(5)Fuc(1)
55	HexNAc(3)Hex(6)Fuc(1)
56	HexNAc(4)Hex(3)Fuc(1)
57	HexNAc(4)Hex(4)Fuc(1)
58	HexNAc(4)Hex(4)Fuc(2)
59	HexNAc(4)Hex(5)Fuc(1)
60	HexNAc(4)Hex(5)Fuc(2)
61	HexNAc(4)Hex(6)Fuc(1)
62	HexNAc(4)Hex(6)Fuc(2)
63	HexNAc(4)Hex(7)Fuc(1)
64	HexNAc(5)Hex(3)Fuc(1)
65	HexNAc(5)Hex(3)Fuc(2)
66	HexNAc(5)Hex(4)Fuc(1)
67	HexNAc(5)Hex(4)Fuc(2)
68	HexNAc(5)Hex(5)Fuc(1)
69	HexNAc(5)Hex(6)Fuc(4)
70	HexNAc(5)Hex(8)Fuc(1)
71	HexNAc(6)Hex(4)Fuc(1)
72	HexNAc(6)Hex(4)Fuc(2)
73	HexNAc(6)Hex(6)Fuc(2)
74	HexNAc(6)Hex(7)Fuc(3)
75	HexNAc(7)Hex(3)Fuc(1)
76	HexNAc(7)Hex(7)Fuc(1)

Node	Glycan
77	HexNAc(8)Hex(5)Fuc(1)
78	HexNAc(8)Hex(9)Fuc(1)
79	HexNAc(9)Hex(3)Fuc(1)
80	HexNAc(9)Hex(6)Fuc(1)
81	HexNAc(9)Hex(9)Fuc(1)
82	HexNAc(3)Hex(4)Fuc(1)NeuAc(1)
83	HexNAc(3)Hex(4)Fuc(2)NeuAc(1)
84	HexNAc(3)Hex(4)NeuAc(1)
85	HexNAc(3)Hex(5)Fuc(1)NeuAc(1)
86	HexNAc(3)Hex(5)NeuAc(1)
87	HexNAc(3)Hex(6)Fuc(1)NeuAc(1)
88	HexNAc(3)Hex(6)NeuAc(1)
89	HexNAc(4)Hex(3)NeuAc(1)
90	HexNAc(4)Hex(4)Fuc(1)NeuAc(1)
91	HexNAc(4)Hex(4)NeuAc(1)
92	HexNAc(4)Hex(5)Fuc(1)NeuAc(1)
93	HexNAc(4)Hex(5)Fuc(1)NeuAc(2)
94	HexNAc(4)Hex(5)Fuc(2)NeuAc(1)
95	HexNAc(4)Hex(5)Fuc(3)NeuAc(1)
96	HexNAc(4)Hex(5)NeuAc(1)
97	HexNAc(4)Hex(6)Fuc(1)NeuAc(1)

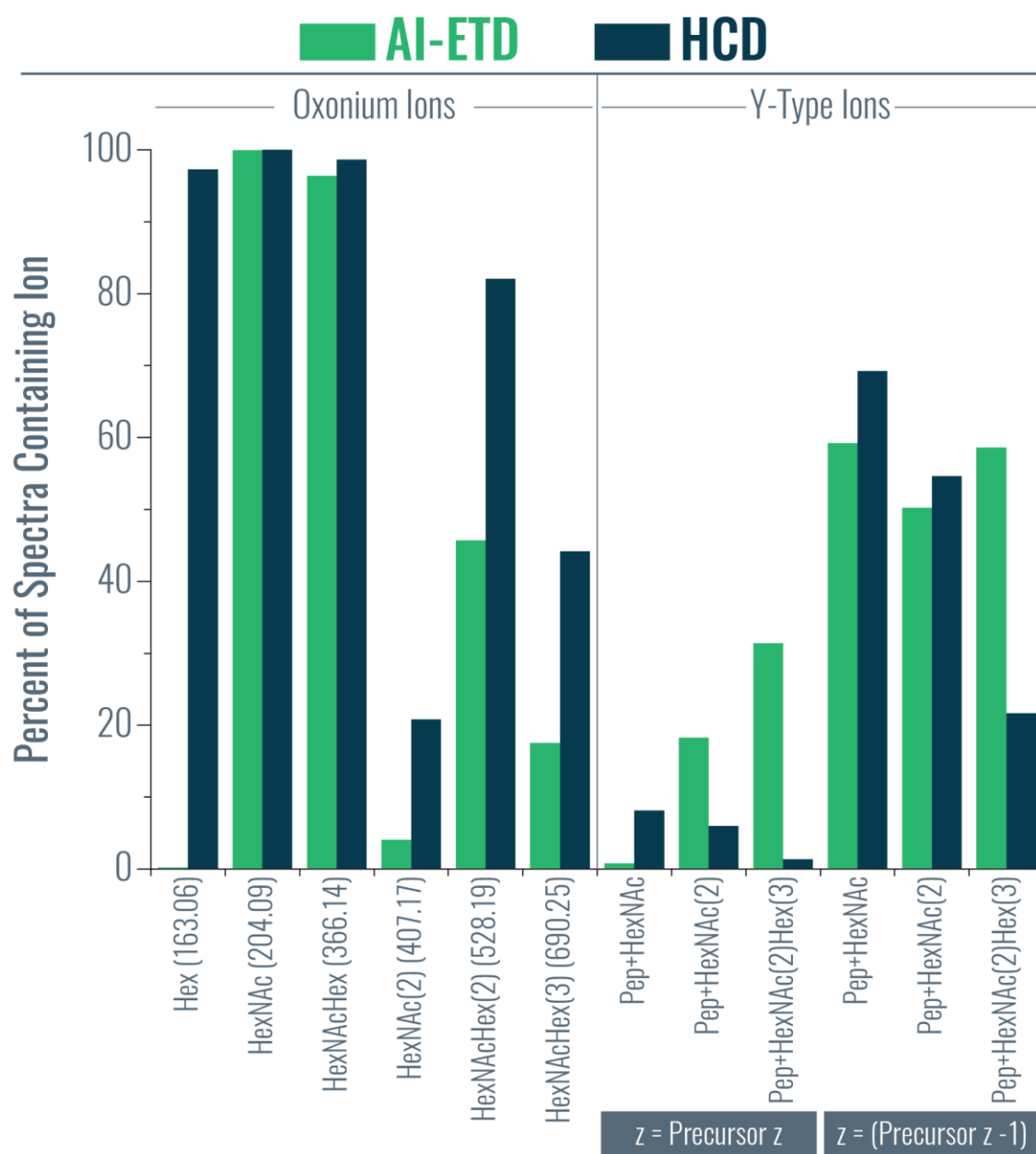
Node	Glycan
98	HexNAc(4)Hex(6)NeuAc(1)
99	HexNAc(4)Hex(7)NeuAc(1)
100	HexNAc(5)Hex(3)Fuc(1)NeuAc(1)
101	HexNAc(5)Hex(4)Fuc(1)NeuAc(1)
102	HexNAc(5)Hex(4)NeuAc(1)
103	HexNAc(5)Hex(4)NeuAc(2)
104	HexNAc(5)Hex(5)Fuc(1)NeuAc(2)
105	HexNAc(5)Hex(5)Fuc(2)NeuAc(1)
106	HexNAc(5)Hex(6)NeuAc(2)
107	HexNAc(6)Hex(3)Fuc(1)NeuAc(1)
108	HexNAc(6)Hex(3)Fuc(1)NeuAc(2)
109	HexNAc(6)Hex(5)Fuc(1)NeuAc(2)
110	HexNAc(6)Hex(5)Fuc(2)NeuAc(1)
111	HexNAc(6)Hex(6)Fuc(2)NeuAc(1)
112	HexNAc(6)Hex(6)NeuAc(1)
113	HexNAc(6)Hex(7)Fuc(3)NeuAc(1)
114	HexNAc(6)Hex(7)NeuAc(3)
115	HexNAc(6)Hex(7)NeuAc(4)
116	HexNAc(6)Hex(9)Fuc(1)NeuAc(2)
117	HexNAc(7)Hex(8)Fuc(1)NeuAc(1)



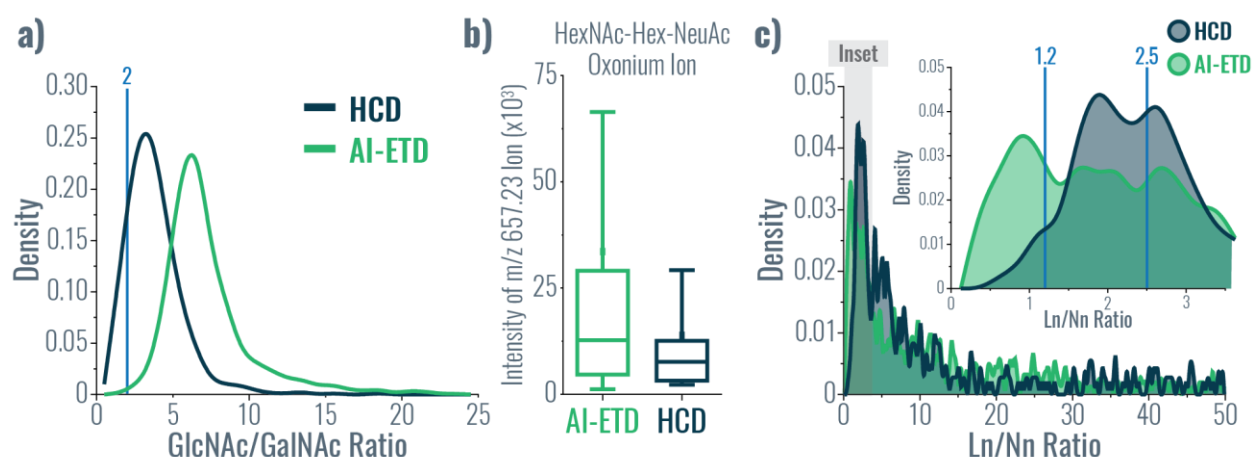
Supplemental Figure 1. Comparing ETD, ETD with supplemental activation using higher energy collisional dissociation (EThcD) and AI-ETD. Supplemental activation methods like EThcD and AI-ETD provide substantially more intact glycopeptide identifications than standard ETD, and AI-ETD provides a small boost in identifications over EThcD.



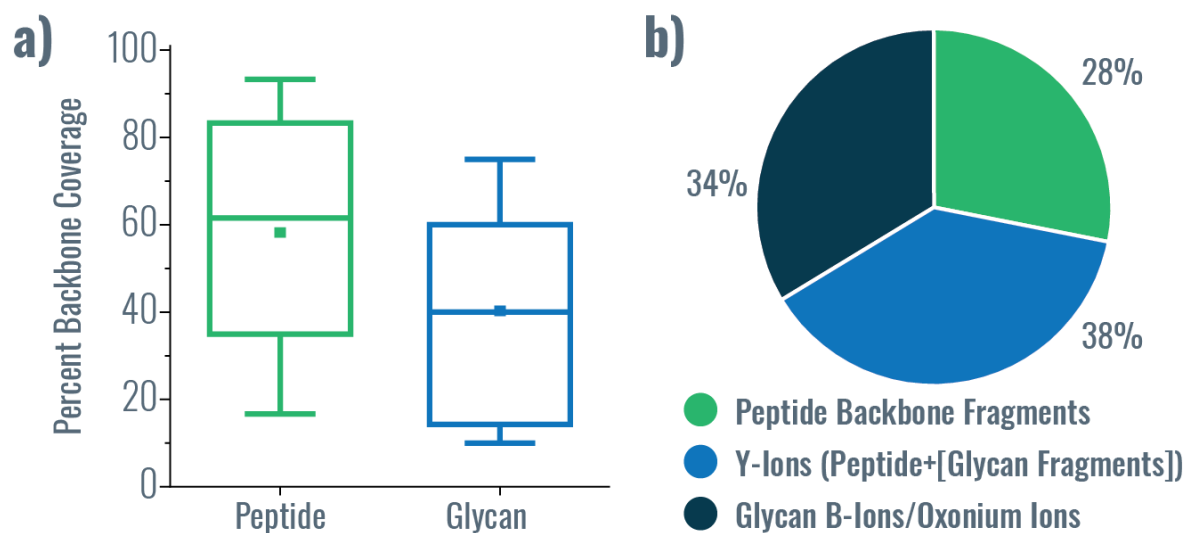
Supplemental Figure 2. Distributions of fragment ions generated by AI-ETD (green) and HCD (dark blue). Density plots show the number of peptide backbone fragments (top), Y-type ions (middle), and oxonium ions (bottom) produced by each fragmentation method.



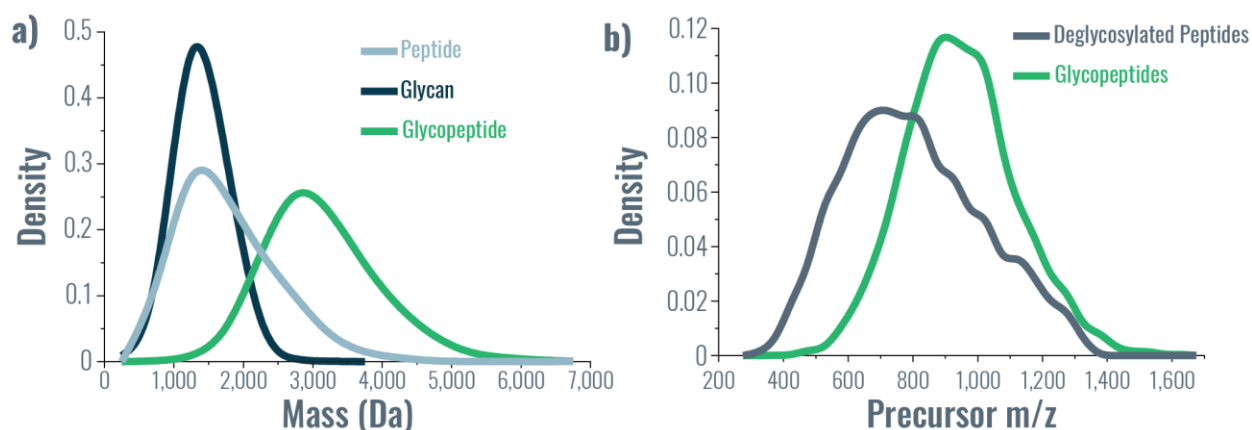
Supplemental Figure 3. Percent of spectra containing specific oxonium and Y-type ions. The bar graph provides the percent of glycoPSMs from AI-ETD (green) and HCD (dark blue) spectra containing specific oxonium and Y-type ion. For the Y-type ions, the left three ions have the same charge as the precursor ion, while the right three ions are for those Y-type ions that had one fewer charge than the precursor ion. This is indicated by the grey boxes below the x-axis, where z = charge.



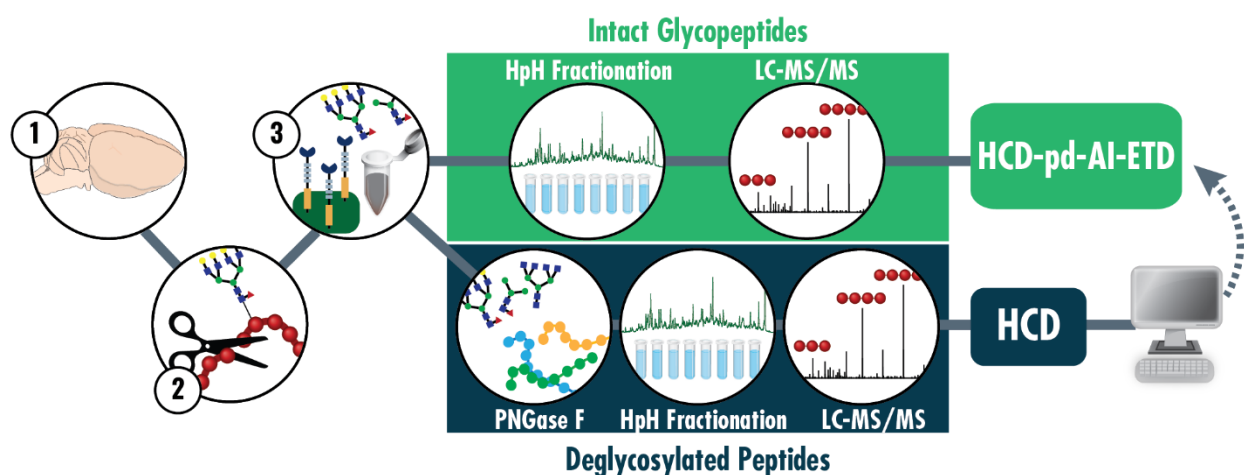
Supplemental Figure 4. Using oxonium ions to differentiate glycan isomers. **a)** GlcNAc/GalNAc ratios, as defined by references #41 and #42, are calculated based on oxonium ion intensities to aid in defining HexNAc isomers as either GalNAc or GlcNAc. Ratios less than one indicate the presence of GalNAc residues, while ratios greater than two indicate the presence of GlcNAc residues (both cutoffs defined by vertical lines on the graph). The distributions of GlcNAc/GalNAc ratios calculated from oxonium ions from HCD and AI-ETD spectra are shown in dark blue and green, respectively. **b)** The box plot shows the distribution of intensities measured for the HexNAcHexNeuAc oxonium ion (m/z 657.2349) in AI-ETD and HCD glycoPSMs that contained a Neu5Ac residue. **c)** The Ln/Nn ratio, as defined in reference #49, provides insight in the presence of either α 2,3 and α 2,6 linked NeuAc residues. The total distributions of Ln/Nn ratios are given for HCD (dark blue) and AI-ETD (green) spectra, with the inset showing a zoom on ratios less than 3.5 (the inset region is shown in grey on the larger graph). Vertical lines show approximate cutoffs for determining α 2,3 (less than 1.2) versus α 2,6 (greater than 2.5) linked NeuAc residues based on uncorrected Ln/Nn ratios provided in reference #44.



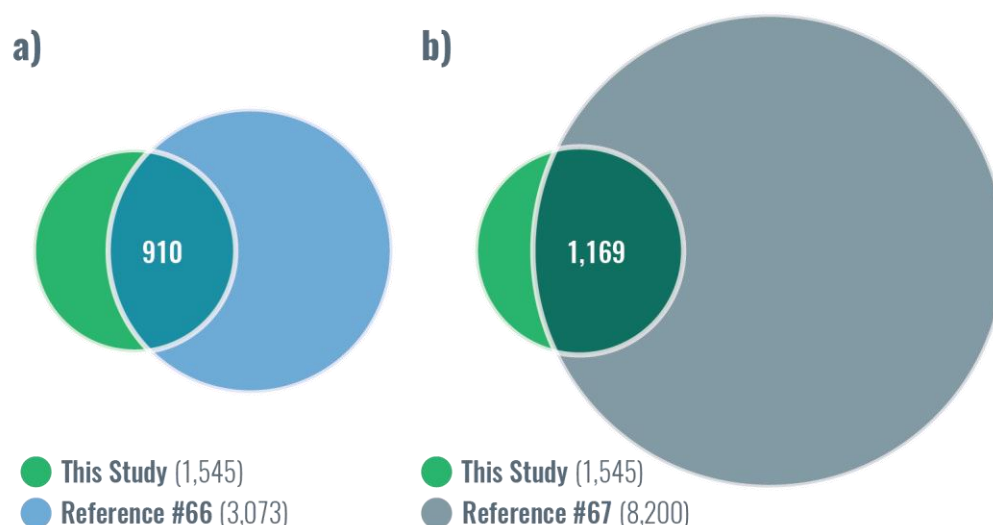
Supplemental Figure 5. HCD performance characteristics. **a)** Distribution of percent peptide backbone coverage and glycan coverage seen in HCD spectra. **b)** Average percent of explained ion current in product ions in HCD spectra from peptide backbone cleavage fragments, Y-ions (i.e., intact peptide sequence with fragments of the glycan moiety), and B-ions/oxonium ions.



Supplemental Figure 6. Mass and m/z distributions of glycoproteomics data. **a)** Density plots show the mass distributions of intact glycopeptides (green), peptide sequences from glycopeptides without the glycan mass considered (light blue), and glycans from glycopeptides with the peptide mass considered (dark blue). **b)** Density plots provide the precursor m/z distribution from deglycosylated peptides, i.e., from the PNGaseF treated samples (grey) and the intact glycopeptides (green), highlighting the shift to significantly higher precursor m/z values in experiments analyzing intact glycopeptides.



Supplemental Figure 7. Workflow for in-depth profiling of the glycoproteome via intact glycopeptides with AI-ETD. Mouse brains are (1) lysed, (2) digested with trypsin, and (3) enriched using a lectin-spin column format. A portion of enriched glycopeptides are deglycosylated with PNGaseF, fractionated using high pH (HpH) reversed phase separations, and analyzed to generate a glycoprotein database to search intact glycopeptide data. The majority of the enriched glycopeptides are kept as intact species, fractionated with HpH separations, and analyzed with LC-MS/MS using a higher-energy collision dissociation-product dependent-activated ion electron transfer dissociation (HCD-pd-AI-ETD) acquisition method.

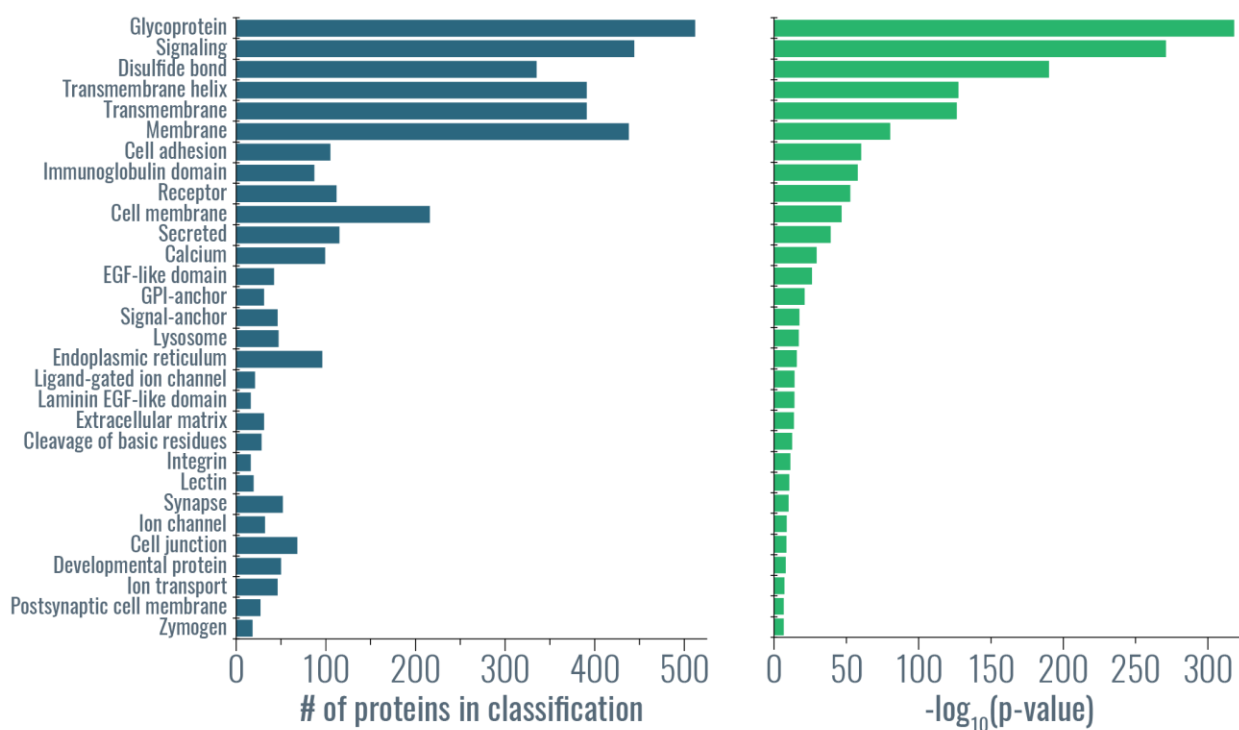


Supplemental Figure 8. Comparison of glycosites to large-scale deglycoproteomic experiments. Zielinska et al. Cell 2010 (reference #66) **(a)** and Fang et al. Oncotarget 2016 (reference #67) **(b)** both used PNGaseF to removed N-glycans and do large-scale sequencing of the deglycoproteome to capture sites of N-glycosylation. Venn diagrams show the overlap in N-glycosites characterized in our intact glycopeptide data and their deglycoproteomic data. Note, Zielinska et al. used trypsin and GluC as proteases and lectins to enrich, while Fang et al. used seven proteases and four different enrichment strategies.

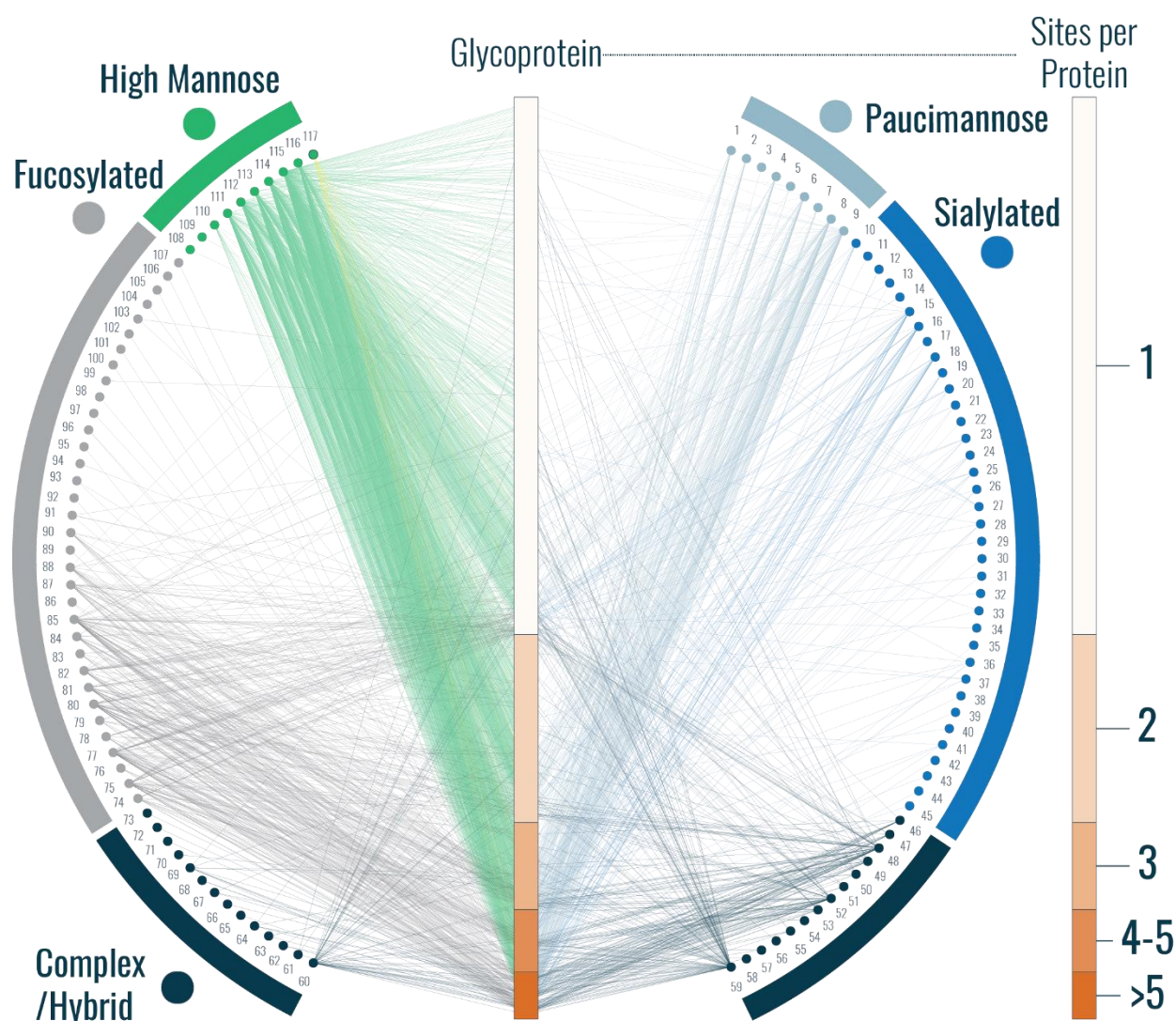
Itga1	N418	N459	N779	N839	N882	N1102	N1113
				★ ▲		★ ▲	★
						★	
		★			★ ▲	★ ▲	
		★ ▲			★ ▲	★	
		★ ▲			★ ▲	★	
	▲						
			★		★		

This Study
 Liu et al. Nat. Comm. 2017 (ref. #30)

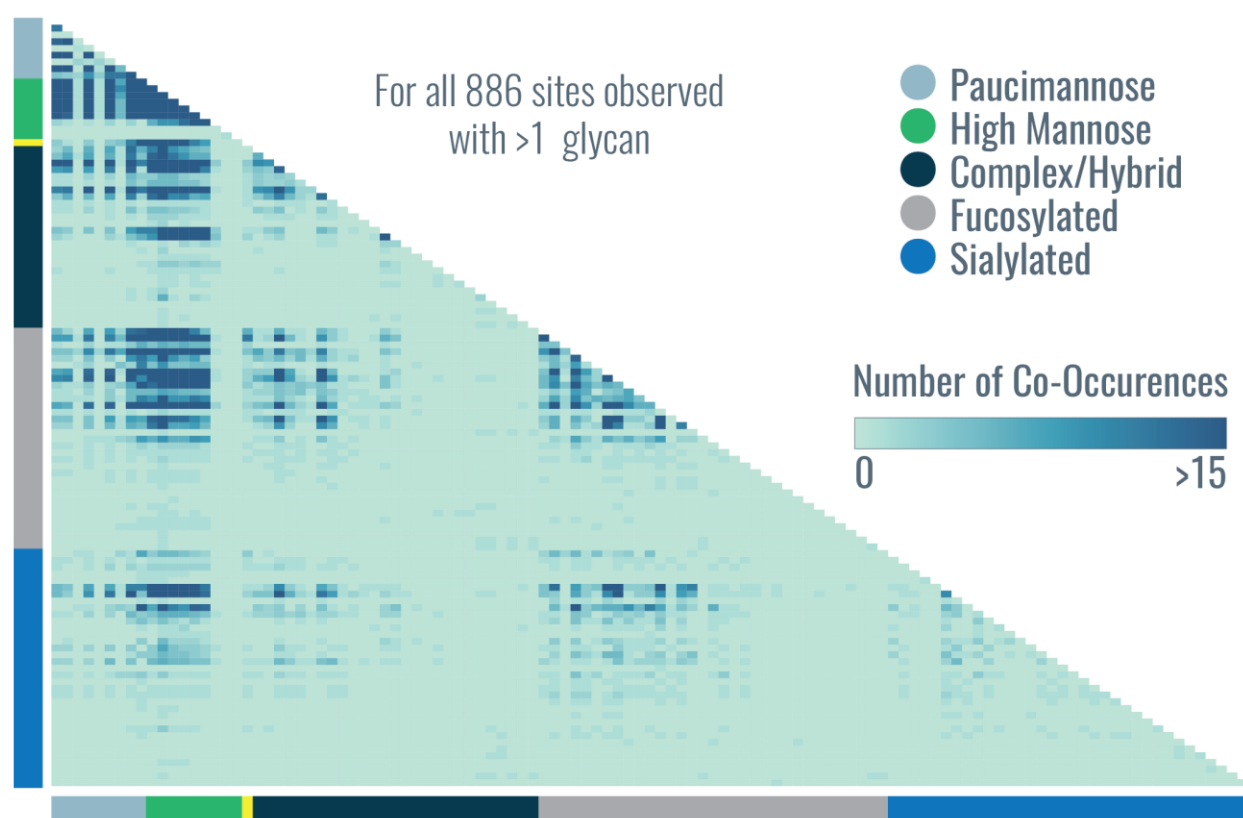
Supplemental Figure 9. N-glycosites characterized in integrin alpha-1 (GN: Itga1, UniProt: Q3V3R4) in mouse brain. Identified N-glycosites are listed in the top row of the table and glycans are in the leftmost column. Dark blue stars indicate glycosite/glycan combinations identified in this study and light blue triangles show identifications from Liu et al. Nat. Comm. 2017 (ref. #30, from the table in Figure 8 of that manuscript). Eight of the nine combinations from the Liu et al. study are seen in this dataset, in addition to seven novel ones (including two novel glycosites).



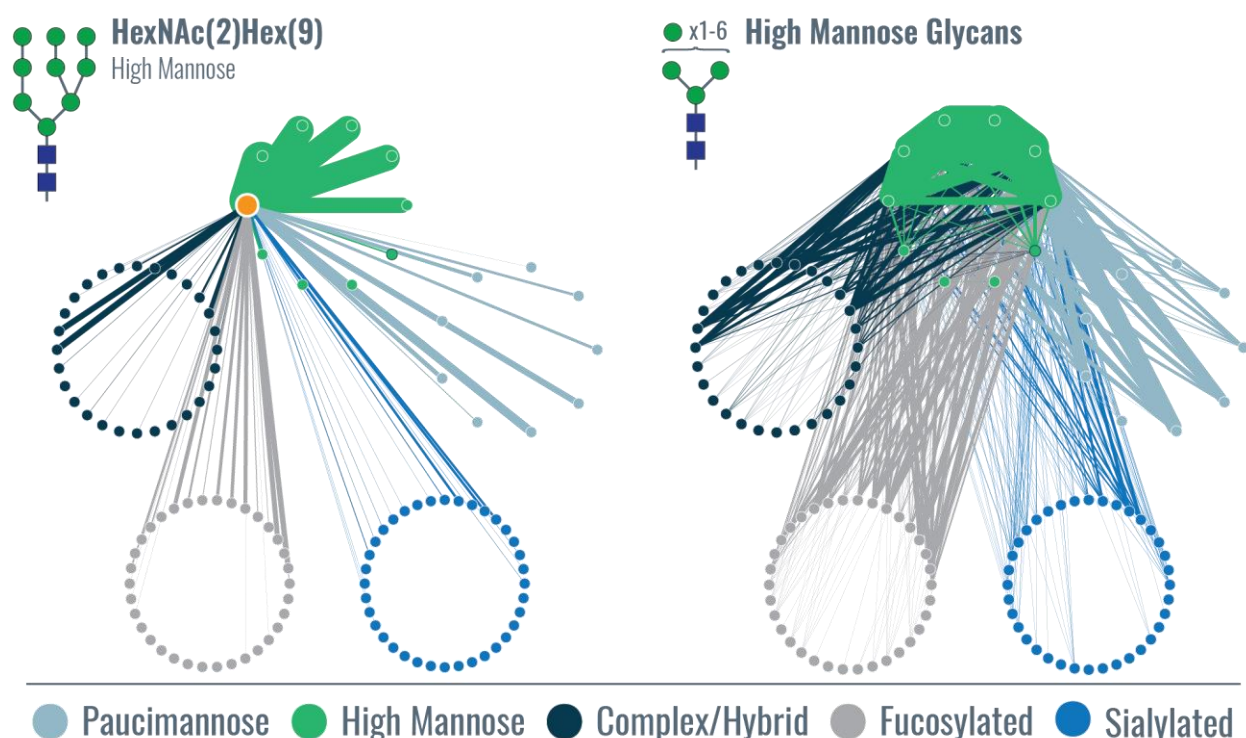
Supplemental Figure 10. Gene ontology functional categories enriched terms for identified glycoproteins. Gene ontology categories for the most enriched terms showing the number of proteins (dark blue) and the significance of the enrichment (green) for each category.



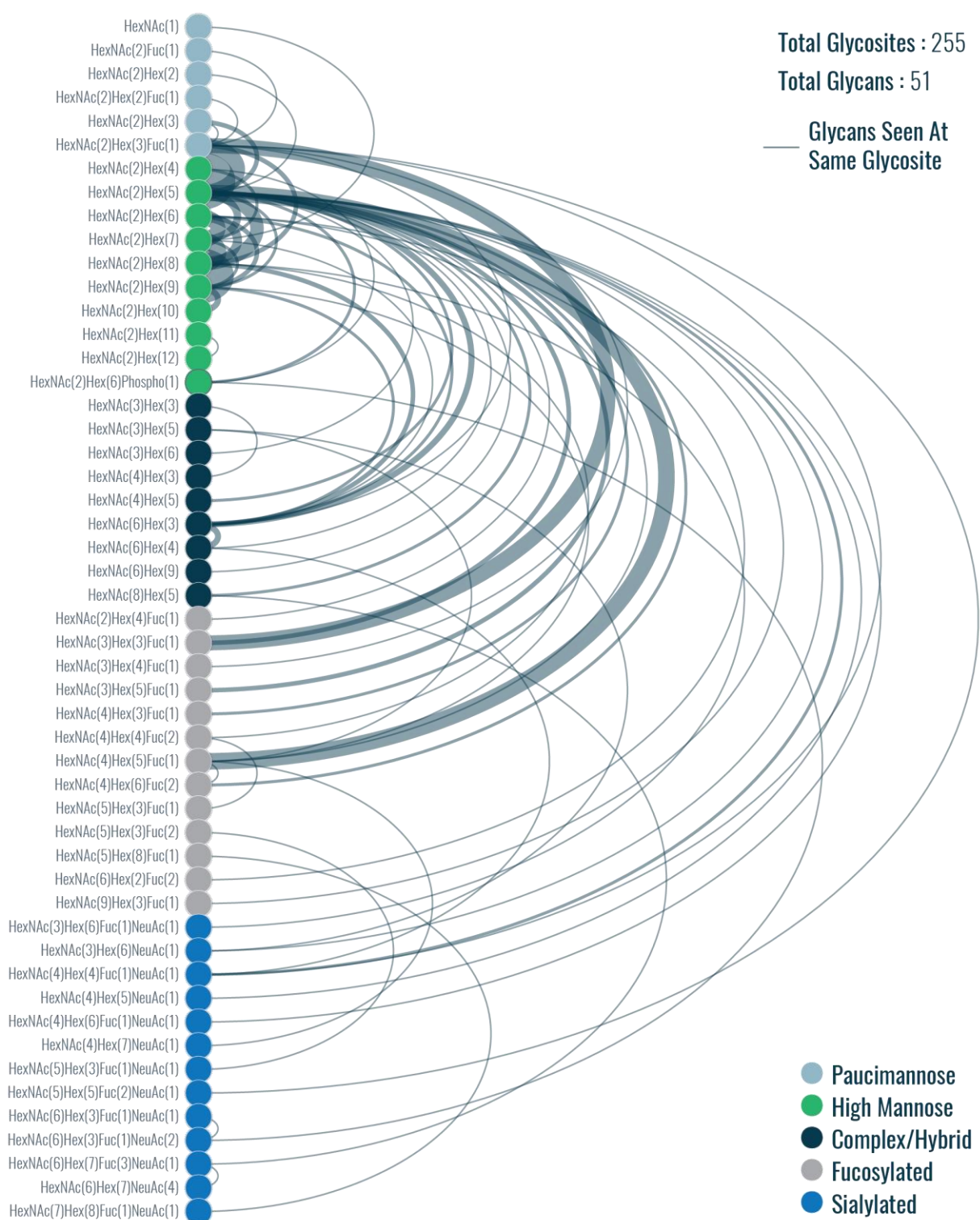
Supplemental Figure 11. A larger version of the glycoprotein-glycan map in Figure 2f. A glycoprotein-glycan network maps which glycans (outer circle, 117 total) modify which proteins (inner bar, 771 total). Glycoproteins are sorted by number of glycosites (scale to the right). Glycans are organized by classification, and edges are colored by the glycan node from which they originate, except for mannose-6-phosphate which has yellow edges. Glycan identities corresponding to node labels are given **Supplemental Table 1**. Note, paucimannose and sialylated glycans that contained fucose moieties are classified as paucimannose or sialylated instead of as fucosylated.



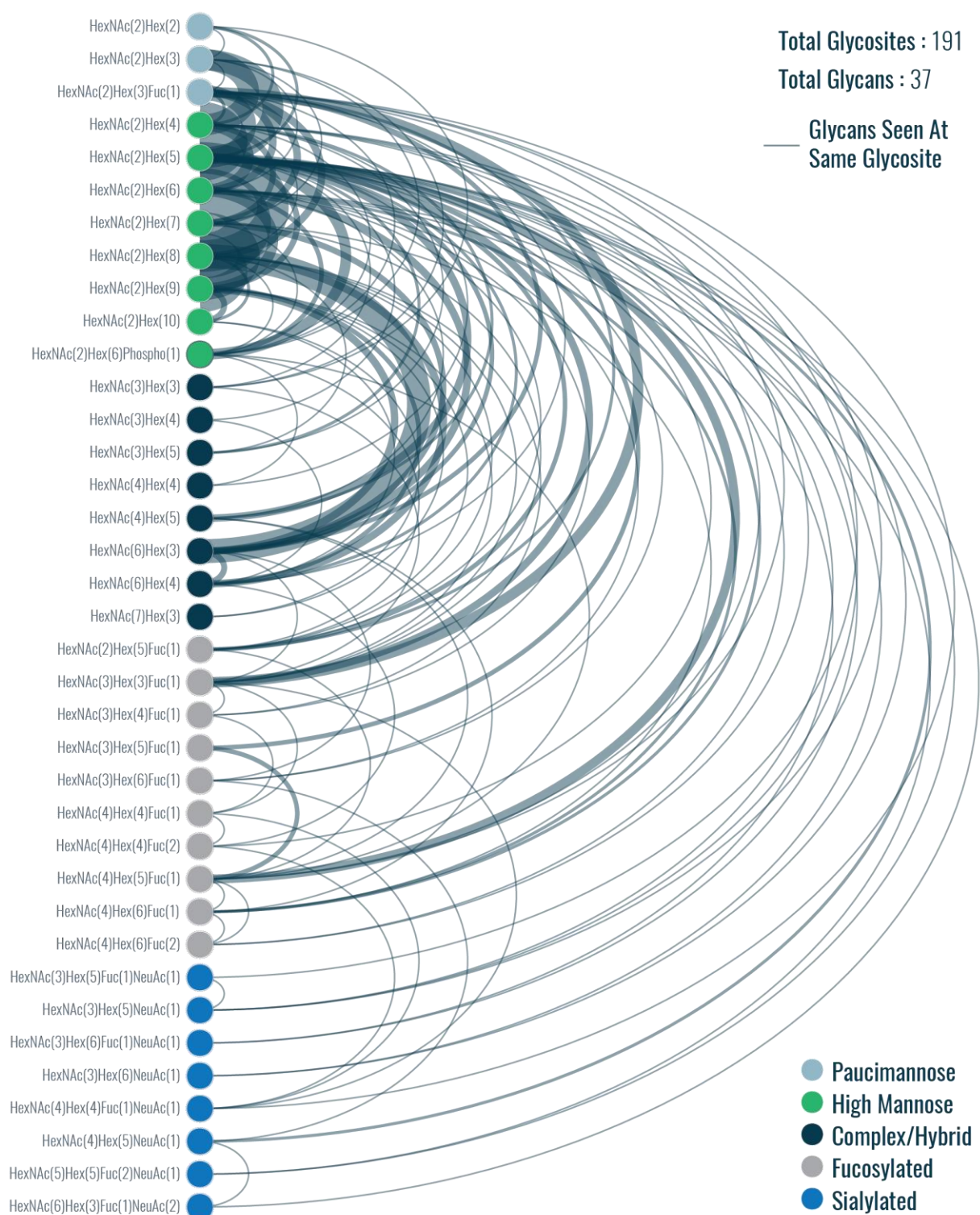
Supplemental Figure 12. A larger version of the co-occurrence heat map in Figure 2g. A heat map represents the number of times glycan pairs appeared together at the same glycosite, indicating which glycans contribute most to microheterogeneity of the >880 sites that had more than one glycan modifying them. Glycans are grouped together by type, as indicated in the key at the top right. Glycan identities are provided by order number in **Supplemental Table 2**.



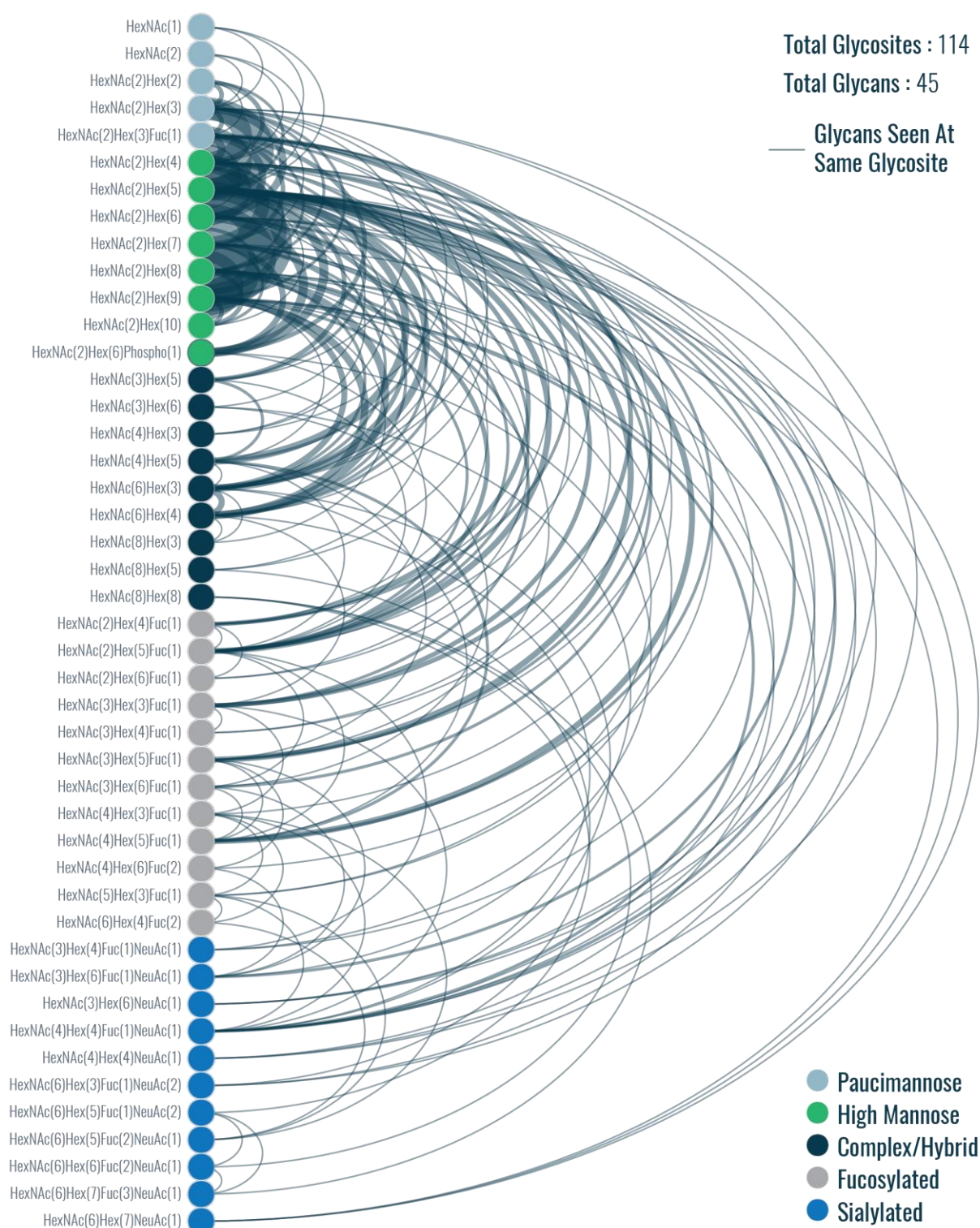
Supplemental Figure 13. Glycan co-occurrence networks for a specific glycan and a class of glycans. Glycan co-occurrence networks can show (left) all the glycans that co-occurred with the specific high mannose glycan HexNAc(2)Hex(9) (highlighted as an orange node, i.e., the source node), with the relative number of occurrences indicated by edge thickness. Edge color indicates the target node. They can also be used to visualize co-occurrence for more than one glycan or entire classes of glycans, in this case shown for all high mannose glycans. Interesting structure arises in the all high mannose glycan co-occurrence network. For example, co-occurrence of certain sialylated glycans (glycans #87-90) with several high mannose glycans is observed while others, e.g., glycans #109-113, occur with no high mannose glycans. Other such trends can be delineated amongst the glycan types that co-occur with high mannose glycans. The organization of the glycan co-occurrence network is given in **Figure 3a**, where glycans are sorted into larger circles based on glycan type, each node is one of the 117 glycans identified, and the numbers indicate glycans identities given in **Supplemental Table 3**.



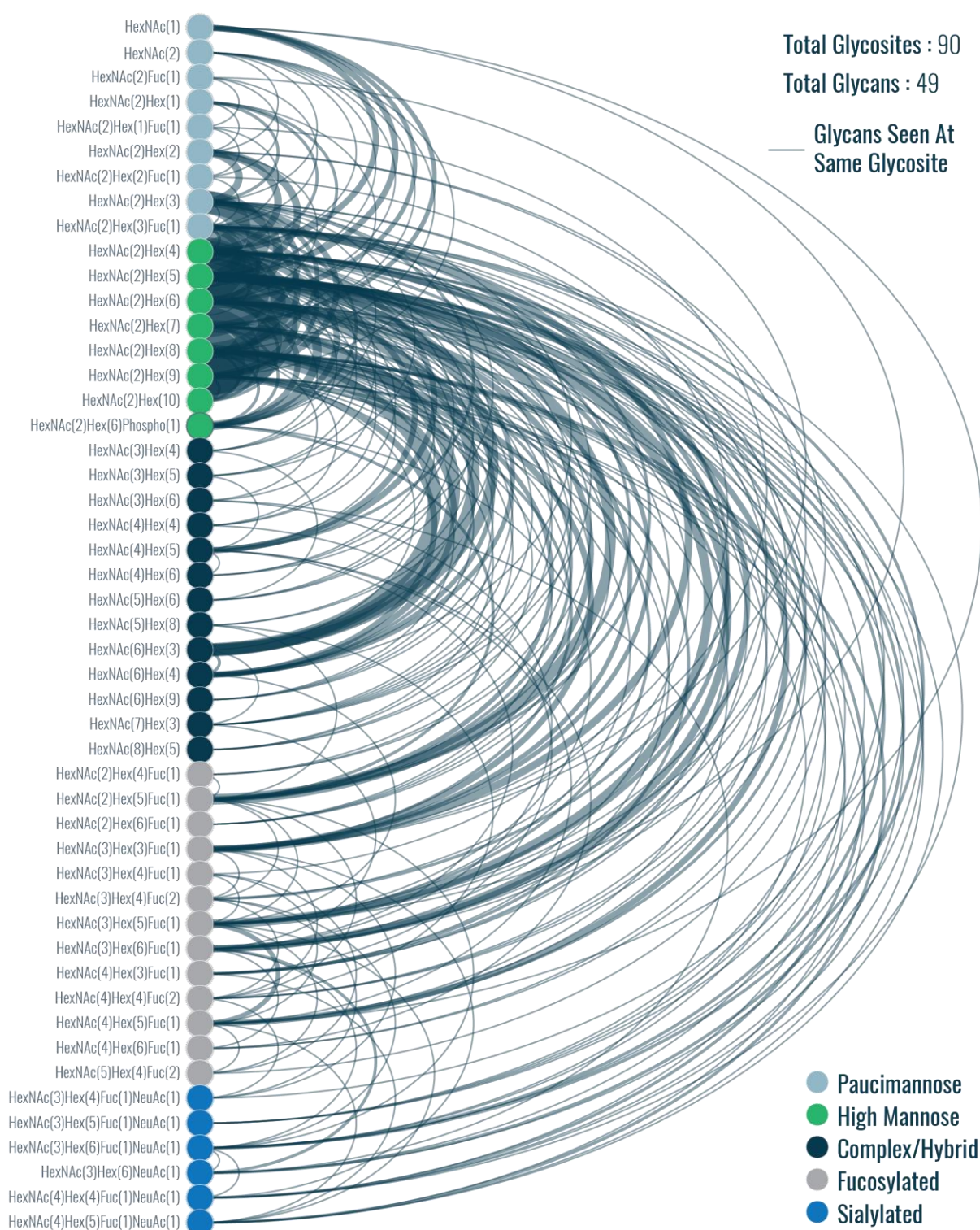
Supplemental Figure 14. Arc plot representing glycan microheterogeneity for glycosites with two glycans per site. Nodes are glycans (colors indicate glycan classification), and edges connect glycans that were seen at the same glycosite (weight indicates number of co-occurrences). Note, paucimannose and sialylated glycans that contain fucose moieties are classified as paucimannose or sialylated instead of as fucosylated.



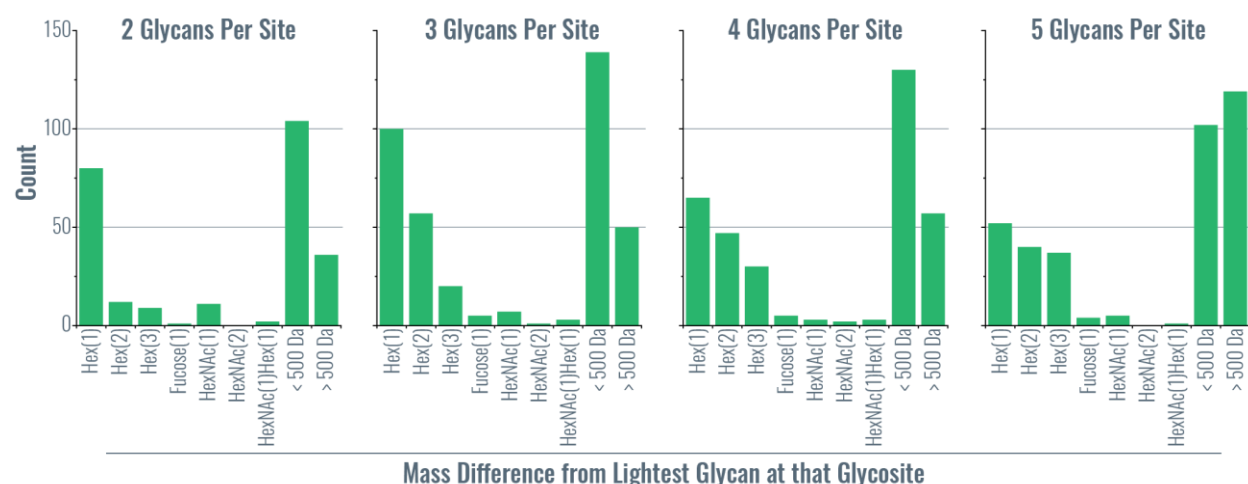
Supplemental Figure 15. Arc plot representing glycan microheterogeneity for glycosites with three glycans per site. Nodes are glycans (colors indicate glycan classification), and edges connect glycans that were seen at the same glycosite (weight indicates number of co-occurrences). Note, paucimannose and sialylated glycans that contain fucose moieties are classified as paucimannose or sialylated instead of as fucosylated.



Supplemental Figure 16. Arc plot representing glycan microheterogeneity for glycosites with four glycans per site. Nodes are glycans (colors indicate glycan classification), edges connect glycans that were seen at the same glycosite (weight indicates number of co-occurrences). Note, paucimannose and sialylated glycans that contain fucose moieties are classified as paucimannose or sialylated instead of as fucosylated.



Supplemental Figure 17. Arc plot representing glycan microheterogeneity for glycosites with five glycans per site. Nodes are glycans (colors indicate glycan classification), and edges connect glycosites that were seen at the same glycan (weight indicates number of co-occurrences). Note, paucimannose and sialylated glycans that contain fucose moieties are classified as paucimannose or sialylated instead of as fucosylated.



Supplemental Figure 18. Mass differences between glycans that occupy the same glycosite. A distribution of mass differences relative to the lightest glycan identified are plotted for all glycans seen at a given glycosite for glycosites with two, three, four, and five distinct glycans. Common mass differences for glycan are shown on the x-axis. For example, if both HexNAc(2)Hex(8) and HexNAc(2)Hex(9) were identified at a glycosite that had two distinct glycans, that would contribute to a count of one to the Hex(1), i.e., one mannose residue, mass difference in the leftmost plot. Observing HexNAc(6)Hex(3) and HexNAc(6)Hex(4), i.e., a difference of one galactose residue, at the same glycosite would contribute to the same Hex(1) count. This provides a crude summary of glycan microheterogeneity at sites with multiple glycans, but discerning all patterns is difficult, especially for >2 glycans per site, because mass difference are all relative to the lightest glycan. For example, a glycan with three glycans HexNAc(2)Hex(8), HexNAc(6)Hex(3), and HexNAc(2)Hex(9) would give mass differences of ~2 Da and ~162 Da (i.e., Hex(1)). This shows a clear difference in a pair of glycans differing by one hexose residue, but the ~2 Da mass difference does not give clear insight into the relationship between the two high mannose glycans [HexNAc(2)Hex(8), HexNAc(2)Hex(9)] and the complex glycan [HexNAc(6)Hex(3)].

GN• Sdk2 UniProt• Q6V4S5

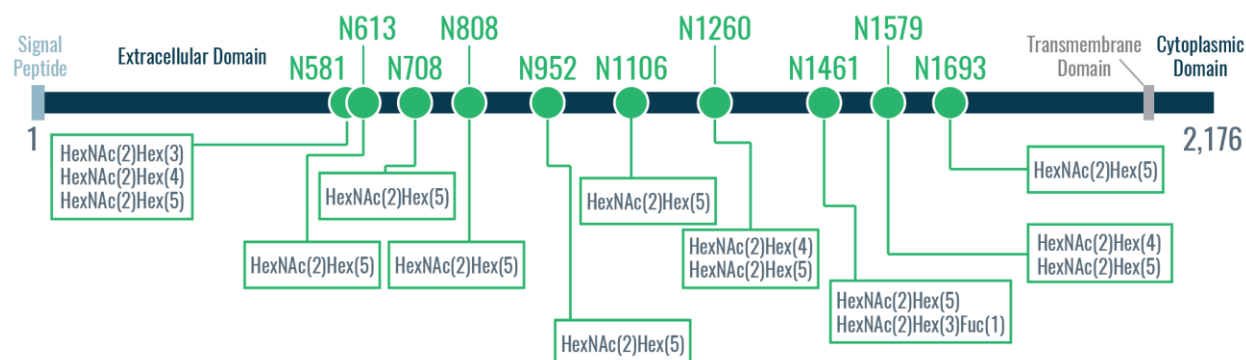
Name• Protein sidekick-2

Keywords/Cellular Component• cell junction, cell membrane, membrane, synapse

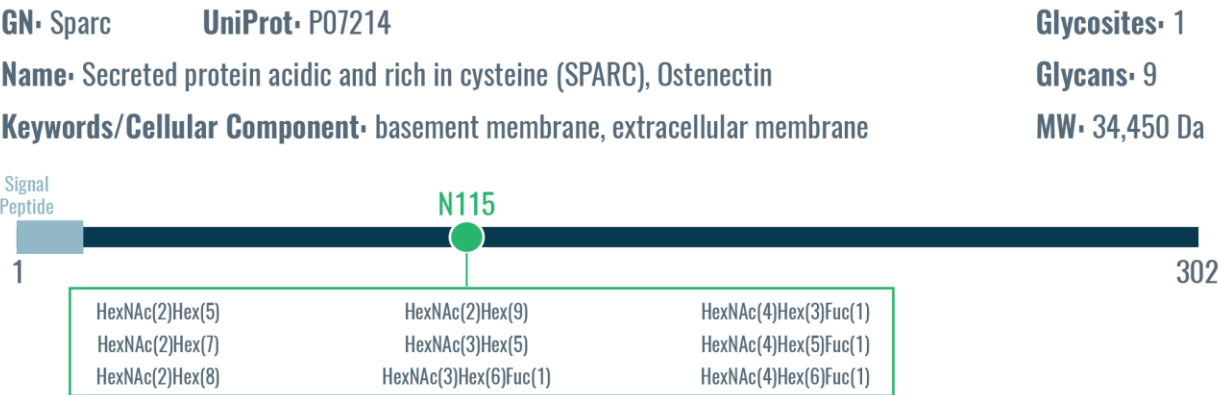
Glycosites• 10

Glycans• 4

MW• 239,902 Da



Supplemental Figure 19. Protein sidekick-2: relatively high number of glycosites with relatively low glycan microheterogeneity. Sdk2 is a somewhat large single-pass membrane glycoprotein found at the cell membrane that functions in cell adhesion, synapse assembly, and formation of neural circuits that detect motion. Even with ten glycosites identified, only four different glycans were characterized, which are smaller high mannose and fucosylated paucimannose core structure. Six of the ten sites had only one glycan, which was HexNAc(2)Hex(5) in all six cases.



Supplemental Figure 20. Secreted protein acidic and rich in cysteine (SPARC): only one glycosite with moderate glycan microheterogeneity. Sparc is an extracellular membrane protein with only one known glycosite but nine different glycans were observed at that site. The degree of glycan heterogeneity is moderate, with high mannose, complex, and fucosylated glycans.

GN· Atp1b2

UniProt· P14231

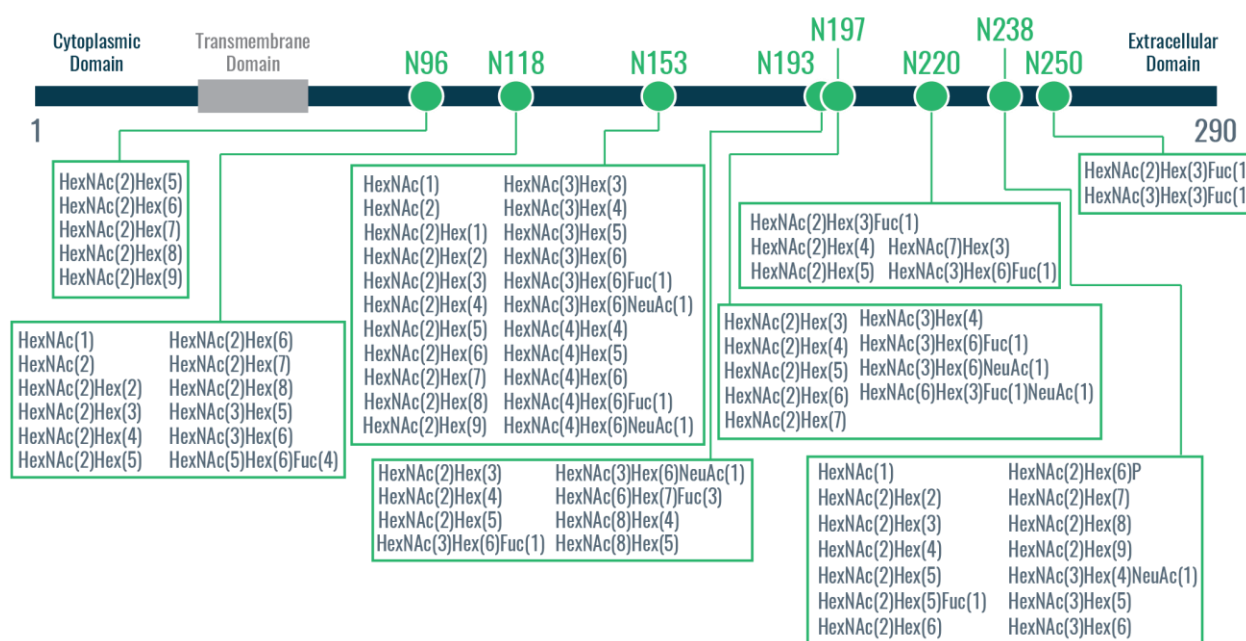
Glycosites· 8

Name· Sodium/potassium-transporting ATPase subunit beta-2

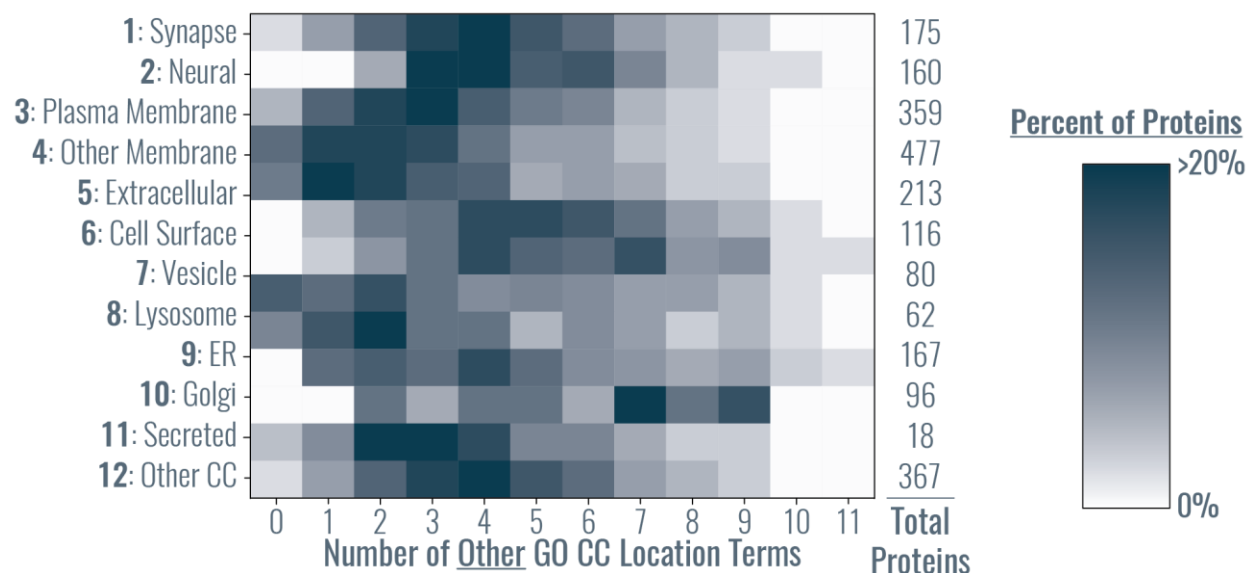
Glycans· 33

Keywords/Cellular Component· plasma membrane

MW· 33,344 Da



Supplemental Figure 21. Sodium/potassium transporting-ATPase $\beta 2$ subunit (Atp1b2): glycosites with varying degrees of glycan microheterogeneity. Five of the eight glycosites display relatively low glycan microheterogeneity (<10 glycans), but N118, N153, and N238 show a higher degree of microheterogeneity with 12, 22, and 14 localized glycans, respectively. Overall, this presents an interesting case where some glycosites display relatively little microheterogeneity while other glycosites on the same protein have a high degree of microheterogeneity.



Supplemental Figure 22. Heat map of the number of subcellular groups (derived from GO cellular component terms) for identified glycoproteins. The y-axis shows the twelve subcellular groups (determined by GO cellular component terms associated with UniProt identification) and the x-axis is the number of GO “other” cellular component terms associated with a UniProt entry. For example, if a protein could be assigned to only one or two subcellular groups, the number of “other” location terms would be zero or one, respectively. The color of the heat map indicates the percentage of total protein identifications from that subcellular group that had a given number of other GO cellular component terms. This graph shows that a significant proportion of proteins could be classified as more than one subcellular group (i.e., location).