Title: Parasite dispersal influences introgression rate

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Abstract: Organisms vary in their dispersal abilities, and these differences can have important biological consequences, such as impacting the likelihood of hybridization events. However, the factors shaping the frequency of hybridization are still poorly understood, and therefore how dispersal ability affects the opportunities for hybridization is still unknown. Here, using the ecological replicate system of dove wing and body lice (Insecta: Phthiraptera), we show that species with higher dispersal abilities exhibited increased genomic signatures of introgression. Specifically, we found a higher proportion of introgressed genomic reads and more reticulated phylogenetic networks in wing lice, the louse group with higher dispersal abilities. Our results illustrate how differences in dispersal ability can drive differences in the extent of introgression through hybridization. The results from this study represent an important step for understanding the factors driving hybridization. We expect our approach will stimulate future studies on the ecological factors shaping hybridization to further understand this important process.

One Sentence Summary: Parasite species with higher dispersal abilities show increased levels of introgression.

Keywords: feather lice, host-symbiont, hybridization, lice, parasites, phoresis, symbionts

Introduction

Dispersal is the permanent movement of organisms away from their place of origin. It is a fundamental process in biology with significant implications at multiple scales of organization (Barton, 1992; Clobert et al., 2001; Matthysen, 2012; Nathan, 2001), including the reproduction of individuals, the composition of populations and communities, and the geographical distribution of species (Clobert et al., 2012, 2001).

Organisms differ in their dispersal abilities, and these differences have an impact on their biology, such as on the distributional range of a species or gene flow between populations (Bohonak, 1999). For example, organisms with lower dispersal abilities tend to have smaller distributional ranges and populations that are genetically more structured (Bohonak, 1999; Dawson et al., 2014; DiBlasi et al., 2018).

Dispersal ability might also affect the opportunities for hybridization between species because the rates at which individuals encounter different species are likely to be higher in organisms with higher dispersal capabilities. Indeed, recent evidence supports this prediction by demonstrating that range expansion is associated with the extent of introgression (Currat et al., 2008; Nussberger et al., 2018). Similarly, dispersal differences explain more than 30% of the variation in the width of hybrid zones across animals (McEntee et al., 2018). However, overall the factors influencing hybridization events are poorly known (Arnold, 2015; Randler, 2006; Taylor and Larson, 2019), and, in particular, the influence of dispersal ability on the rate of hybridization remains understudied.

Comparisons of the effect of dispersal on hybridization should ideally hold constant most factors other than dispersal. The ecological replicate system of wing and body lice (Insecta: Phthiraptera) of pigeons and doves (Aves: Columbidae) has proven to be an ideal system for comparing the impact of dispersal differences on other aspects of biology, such as population structure and codivergence (Clayton et al., 2015; Clayton and Johnson, 2003; DiBlasi et al., 2018; Johnson and Clayton, 2004; Sweet and Johnson, 2018). Specifically, this is an excellent system in which to assess the effect of differences in dispersal capabilities on levels of introgression because both of these two lineages of feather lice: 1) co-occur across the diversity of pigeons and doves, 2) present highly comparable temporal patterns of diversification, 3) have the same basic life history and diet (Clayton et al., 2015; Sweet and Johnson, 2018, 2015), but 4) they significantly differ in their dispersal ability (Bartlow et al., 2016; Harbison et al., 2009, 2008). Both wing and body lice disperse vertically between parents and offspring in the nest. However, wing lice can also attach to and hitchhike on hippoboscid flies to disperse "phoretically" between host individuals or host species (Bartlow et al., 2016; Harbison et al., 2009, 2008). Indeed, this hitch-hiking dispersal mechanism profoundly influences their degree of population structure and cophylogenetic history (Clayton and Johnson, 2003; DiBlasi et al., 2018; Sweet et al., 2017b; Sweet and Johnson, 2018). In addition, wing lice have a higher rate of host-switching (Clayton et al., 2015; Clayton and Johnson, 2003; Sweet et al., 2017b) (i.e., successful colonization of new host species) and of straggling (Whiteman et al., 2004) (i.e., dispersal to new host species without reproduction on that new host).

To compare differences in the extent of introgression between wing and body lice, we used whole-genome data from 71 louse individuals belonging to five taxa of wing lice (*Columbicola*) and seven taxa of body lice (*Physconelloides*) occurring across the same host species and have highly comparable patterns of diversification (Sweet and Johnson, 2018, 2015). We predicted that wing lice, which have higher dispersal abilities and thus higher odds of

encountering individuals of a different louse species on the same host, should show more
extensive evidence of introgression (Fig. 1).

Materials and Methods

<u>Data</u>
We studied whole genome data from 71 louse individuals belonging to five and seven taxa of
Columbicola and *Physconelloides*, respectively (Supplementary Table S2). Data were available
from previous studies (Boyd, et al. 2017; Sweet, Boyd, et al. 2017; Sweet and Johnson 2018) and
represent all described New World ground-dove wing and body louse species, most host species
in this group, including samples across multiple biogeographic areas within species (Sweet and
Johnson, 2018) (Supplementary Table S2). Illumina genome sequence data pre-processing
included several steps (Sweet and Johnson, 2018). First, we discarded duplicate read pairs using
the fastqSplitDups script (https://github.com/McIntyre-Lab/mcscriptand
https://github.com/McIntyre-Lab/mclib). We then eliminated the Illumina sequencing adapters
with Fastx_clipper v0.014 from the FASTX-Toolkit (http://hannonlab.cshl.edu/fastx_toolkit).
Also, we removed the first 5 nt from the 5' ends of reads using Fastx_trimmer v0.014 and
trimmed bases from the 3' ends of reads until reaching a base with a phred score ≥28 using
Fastq_quality_trimmer v0.014. Finally, we removed any reads less than 75 nt and analyzed the
cleaned libraries with Fastqc v0.11.5 to check for additional errors. We assembled nuclear loci in
aTRAM following previous studies (Allen et al., 2015; Boyd BM, et al., 2017; Sweet and
Johnson, 2018). In particular, we mapped modest coverage (25-60X), multiplexed genomic data
to reference loci from a closely related taxon. For our reference set of nuclear loci for wing lice,
we used 1,039 exons of *Columbicola drowni* generated in a previous study (Boyd BM, et al.,
2017) (raw data: SRR3161922). This data set was assembled de novo (Allen et al., 2015) using
orthologous protein-coding genes from the human body louse genome (*Pediculus humanus*
humanus (Kirkness et al., 2010)) as a set of target sequences. We mapped our newly generated
Columbicola reads and the reads obtained from GenBank to the *C. drowni* references using
Bowtie2 (Langmead and Salzberg, 2012). For body lice, we obtained nuclear data using the same
pipeline and software parameters, except that we used 1,095 loci from *P. emersoni* as the
reference for mapping. To generate the input ultrametric gene trees for Phylonet v3.6.8 (Than et
al., 2008; Wen et al., 2018; Yu and Nakhleh, 2015), we first aligned each nuclear locus in
MAFFT (Katoh, 2002)(--auto) and removed columns with only ambiguous sequences ("N").
Then, we estimated gene trees in RAxML v8.1.3 (Stamatakis, 2006) with a GTR + Γ substitution
model for each gene alignment. Finally, we made trees ultrametric using the nnls method in the
force.ultrametric function within the "phytools" R package (Revell, 2011).

<u>Quantifying introgression</u>
We used two different approaches to quantify differences in the extent of introgression between
the two louse genera. We employed methods suitable to both detect introgression between
species and individuals from the same species (i.e., we did not employ methods aimed to detect
differences at the population level, e.g., TreeMix; Pickrell and Pritchard 2012). First, we used
sppIDer (Langdon et al., 2018) to quantify the genomic contributions of different louse species in
an individual louse genome. We built our reference for each genus using all the nuclear loci from
a single individual per species. For the reference, we selected those individuals for which we
assembled the highest number of genes for each genus. Finally, we estimated the extent of
introgression as the sum of the mean coverages of reads mapped from all the species excluding

the focal louse species, divided by the mean coverage of the focal louse species. Second, we
quantified introgression at the species level, while accounting for ILS (i.e., reticulations in this
method can be attributed to hybridization events), using a maximum pseudo-likelihood
framework with PhyloNet 3.6.1 (Than et al., 2008; Wen et al., 2018; Yu and Nakhleh, 2015). We
trimmed the unrooted gene trees to the same individuals used as reference taxa in sppIDer, and
performed eleven independent analyses with a differing maximum number of reticulation nodes
(i.e., from zero to ten). We conducted ten runs per analysis. We then selected the optimal
network for each genus based on AIC values.

Analyses
We compared the sppIDer results using generalized linear models (GLMs). We used a Gaussian
distribution of errors and an identity link function. We performed one GLM for each simulation
iteration using the glm function of the "stats" R package (R Core Team, 2013). The extent of
introgression for each louse genus was the dependent variable, the genus identity was the
independent variable, and we accounted for the introgression differences between louse species
including louse identity as a fixed factor. We confirmed assumptions underlying GLMs by
testing the normality of regression residuals for normality against a Q-Q plot. We also
considered the possibility that some of the reads mapping to other species were technical
contaminations, i.e., due to index-swapping (Carlsen et al., 2012; Esling et al., 2015; Schnell et
al., 2015; Sinha et al., 2017). Previous studies have found that the misassignment of reads
generally ranges from 1 to 9% (Carlsen et al., 2012; Esling et al., 2015; Schnell et al., 2015;
Sinha et al., 2017). Thus, to account for possible contaminants, we wrote a simulation in R that
randomly subtracted 9% (Carlsen et al. 2012; Esling et al. 2015; Schnell et al. 2015; Sinha et al.
2017) from the mean coverage value of a particular sample (i.e., we subtracted a random
proportion of the mean coverage value for each sample until reaching 9 %). We ran 100
iterations of the simulation and ran a GLM for each iteration (Table S1). Finally, we used the χ^2
test to compare the number of species in pairwise comparisons of each genus with the number of
reticulations found in each optimal phylogenetic network.

Results

We used two different approaches to quantify the overall (i.e., ancient plus recent) differences in
introgression between louse genera. First, in individual louse genomes, we quantified the
genomic contributions from different closely related louse species of the same genus (Langdon et
al., 2018). Second, we quantified introgression at the species level, while accounting for
incomplete lineage sorting (ILS), by inferring phylogenetic networks using a maximum pseudo-
likelihood framework (Than et al., 2008; Wen et al., 2018; Yu and Nakhleh, 2015).
Both approaches revealed highly concordant results: higher levels of introgression among
species of wing lice compared to body lice. In particular, using a read-mapping based method,
the genomic signature of introgression was significantly higher in wing louse species than in
body louse species (GLM with the mean values of the simulations; F = 21.0705, df = 69, P =
2.367 x 10^{-5}; Fig. 2, Supplementary Table S1, Figs. S1-S12). Contrary to this effect, one body
louse individual (included in the GLMs) exhibited the highest level of introgression (Fig. 2,
Figs. S1-S12). However, the other individual from the same taxon, inhabiting the same host
species and collected in the same geographic region, did not show these elevated levels of
introgression (Supplementary Table S2, Figs. S1-S12).
Secondly, in a phylogenetic network framework, the optimal networks of wing lice were
more reticulated than those of body lice even though the number of taxa included in the networks

was lower (seven reticulations in *Columbicola* vs. four in *Physconelloides*, Fig. 3). Accordingly,
the number of reticulations given the number of potential combinations was significantly higher
(χ^2= 3.8132; df=1; *P*= 0.03). Also, the specific lineages involved in the reticulations were
generally congruent with signatures of introgression from the read-mapping based approach (Fig.
S1-S12).

Discussion

Taken together, evidence from wing and body louse genomes suggests that differences in
dispersal ability drive differences in the extent of introgression in this system of ecological
replicate parasites. This work is among the first studies of introgression in a host-symbiont
system (Detwiler and Criscione, 2010). Notably, recent studies have found that straggling and
host-switching are relatively common processes in host-symbiont systems (Bourguignon et al.,
2018; De Vienne et al., 2013; Doña et al., 2019; Nylin et al., 2018). Our study suggests that in a
straggling/host-switching scenario, hybridization can provide further variation with important
eco-evolutionary consequences (Barton, 2018). Indeed, we may have found a potential recent
hybridization event (i.e., the *Physconelloides* individual showing the highest level of
introgression), though this requires further study to rule out methodological issues (e.g., wet-lab
contamination). Overall, the results from this study represent a significant step towards
understanding the factors driving hybridization, because most previous studies focus on the
presence/absence of hybridization and the evolutionary consequences of hybridization events
(Arnold, 2015; Folk et al., 2018; Taylor and Larson, 2019). Further research is needed to
understand the factors shaping the frequency of hybridization and to ascertain their influence
across different scales (e.g., from ancient to recent hybridization events).

Data availability: All data needed to evaluate the conclusions in the paper are present in the
paper and/or the Supplementary Materials. Additional data related to this paper may be requested
from the authors.

Supplementary Materials:

197 Supplementary data are available at Figshare (DOI: 10.6084/m9.figshare.9176204).

Acknowledgments:

200 This study was supported by the US National Science Foundation (DEB-1239788 and DEB-
201 1342604 to K.P.J).

References:

Allen JM, Huang DI, Cronk QC, Johnson KP. 2015. aTRAM - automated target restricted assembly method: a fast
method for assembling loci across divergent taxa from next-generation sequencing data. *BMC
Bioinformatics* **16**. doi:10.1186/s12859-015-0515-2
Arnold ML. 2015. Divergence with genetic exchange. Oxford, UK: OUP Oxford.
Bartlow AW, Villa SM, Thompson MW, Bush SE. 2016. Walk or ride? Phoretic behaviour of amblyceran and
ischnoceran lice. *Int J Parasitol* **46**:221–227.
Barton NH. 2018. The consequences of an introgression event. *Mol Ecol* **27**:4973–4975.
Barton NH. 1992. The genetic consequences of dispersal. In: Animal Dispersal. Netherlands: Springer.

Bohonak AJ. 1999. Dispersal Gene Flow, and Population Structure. *Q Rev Biol* **74**:21–45.

Bourguignon T, Lo N, Dietrich C, Šobotník J, Sidek S, Roisin Y, Brune A, Evans TA. 2018. Rampant host
switching shaped the termite gut microbiome. *Curr Biol* **28**:649–654.

Boyd BM, Allen JM, Nguyen N, Sweet AD, Warnow T, Shapiro MD, et al. 2017. Phylogenomics using Target-
restricted Assembly Resolves Intra-generic Relationships of Parasitic Lice (Phthiraptera: Columbicola).
Syst Biol **66**:896–911.

Carlsen T, Aas AB, Lindner D, Vrålstad T, Schumacher T, Kauserud H. 2012. Dont make a mista(g)ke: is tag
switching an overlooked source of error in amplicon pyrosequencing studies? *Fungal Ecol* **5**:747–749.

Clayton DH, Bush SE, Johnson KP. 2015. Coevolution of life on hosts: integrating ecology and history. Chicago,
IL: University of Chicago Press.

Clayton DH, Johnson KP. 2003. Linking coevolutionary history to ecological process: doves and lice. *Evolution*
57:2335–2341.

Clobert J, Baguette M, Benton TG, Bullock JM, editors. 2012. Dispersal Ecology and Evolution. Oxford University
Press. doi:10.1093/acprof:oso/9780199608898.001.0001

Clobert J, Danchin E, Dhondt AA, D NJ. 2001. Dispersal. Oxford Univ. Press.

Currat M, Ruedi M, Petit RJ, Excoffier L. 2008. The hidden side of invasions: massive introgression by local genes.
Evolution **62**:1908–1920.

Dawson MN, Hays CG, Grosberg RK, Raimondi PT. 2014. Dispersal potential and population genetic structure in
the marine intertidal of the eastern North Pacific. *Ecol Monogr* **84**:435–456.

De Vienne DM, Refrégier G, López-Villavicencio M, Tellier A, Hood ME, Giraud T. 2013. Cospeciation vs host-
shift speciation: methods for testing, evidence from natural associations and relation to coevolution. *New*
Phytol **198**:347–385.

Detwiler JT, Criscione CD. 2010. An infectious topic in reticulate evolution: introgression and hybridization in
animal parasites. *Genes (Basel)* **1**:102–23.

DiBlasi E, Johnson KP, Stringham SA, Hansen AN, Beach AB, Clayton DH. 2018. Phoretic dispersal influences
parasite population genetic structure. *Mol Ecol* **27**:2770–2779.

Doña J, Serrano D, Mironov S, Montesinos-Navarro A, Jovani R. 2019. Unexpected bird–feather mite associations
revealed by DNA metabarcoding uncovers a dynamic ecoevolutionary scenario. *Mol Ecol* **28**:379–390.

Esling P, Lejzerowicz F, Pawlowski J. 2015. Accurate multiplexing and filtering for high-throughput amplicon-
sequencing. *Nucleic Acids Res* **43**:2513–24.

Folk RA, Soltis PS, Soltis DE, Guralnick R. 2018. New prospects in the detection and comparative analysis of
hybridization in the tree of life. *Am J Bot* **105**:364–375.

Harbison CW, Bush SE, Malenke JR, Clayton DH. 2008. Comparative transmission dynamics of competing parasite
species. *Ecology* **89**:3186–3194.

Harbison CW, Jacobsen MV, Clayton DH. 2009. A hitchhiker's guide to parasite transmission: The phoretic
behaviour of feather lice. *Int J Parasitol* **39**:569–575.

Johnson KP, Clayton DH. 2004. Untangling coevolutionary history. *Syst Biol* **53**:92–4.

Katoh K. 2002. MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform.
Nucleic Acids Res **30**:3059–3066.

Kirkness EF, Haas BJ, Sun W, Braig HR, Perotti MA, Clark JM. 2010. Genome sequences of the human body louse
and its primary endosymbiont provide insights into the permanent parasitic lifestyle. *Proc Natl Acad Sci U*
S A **107**:12168–73.

Langdon QK, Peris D, Kyle B, Hittinger CT. 2018. sppIDer: A Species Identification Tool to Investigate Hybrid
Genomes with High-Throughput Sequencing. *Mol Biol Evol* **35**:2835–2849.

Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. *Nat Methods* **9**:357–9.

Matthysen E. 2012. Multicausality of dispersal: a review. In: Dispersal Ecology and Evolution. Oxford University
Press.

McEntee JP, Burleigh JG, Singhal S. 2018. Dispersal predicts hybrid zone widths across animal diversity:
Implications for species borders under incomplete reproductive isolation. *bioRxiv*. doi:10.1101/472506

Nathan R. 2001. The challenges of studying dispersal. *Trends Ecol Evol* **16**:481–483.

Nussberger B, Currat M, Quilodran CS, Ponta N, Keller LF. 2018. Range expansion as an explanation for
introgression in European wildcats. *Biol Conserv* **218**:49–56.

Nylin S, Agosta S, Bensch S, Boeger WA, Braga MP, Brooks DR, Forister ML, Hambäck PA, Hoberg EP, Nyman
T. 2018. Embracing colonizations: a new paradigm for species association dynamics. *Trends Ecol Evol*
33:4–14.

Pickrell JK, Pritchard JK. 2012. Inference of Population Splits and Mixtures from Genome-Wide Allele Frequency

268   Data. *PLoS Genetics* **8**.
269   R Core Team. 2013. R: A language and environment for statistical computing.
270   Randler C. 2006. Behavioural and ecological correlates of natural hybridization in birds. *Ibis* **148**:459–467.
271   Revell L. 2011. phytools: an R package for phylogenetic comparative biology (and other things). *Methods Ecol Evol*
272   **3**:217–223.
273   Schnell IB, Bohmann K, Gilbert MT. 2015. Tag jumps illuminated–reducing sequence-to-sample misidentifications
274   in metabarcoding studies. *Mol Ecol Resour* **15**:1289–303.
275   Sinha R, Stanley G, Gulati GS, Ezran C, Travaglini KJ, Wei E. 2017. Index Switching Causes Spreading-Of-Signal
276   Among. *bioRxiv*. doi:10.1101/125724
277   Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa
278   and mixed models. *Bioinformatics* **22**:2688–2690.
279   Sweet AD, Boyd BM, Allen JM, Villa SM, Valim MP, Rivera-Parra JL. 2017a. Integrating phylogenomic and
280   population genomic patterns in avian lice provides a more complete picture of parasite evolution. *Evolution*
281   **72**:95–112.
282   Sweet AD, Chesser RT, Johnson KP. 2017b. Comparative cophylogenetics of Australian phabine pigeons and doves
283   (Aves: Columbidae) and their feather lice (Insecta: Phthiraptera). *Int J Parasitol* **47**:347–356.
284   Sweet AD, Johnson KP. 2018. The role of parasite dispersal in shaping a host–parasite system at multiple
285   evolutionary scales. *Mol Ecol* **27**:5104–5119. doi:10.1111/mec.14937
286   Sweet AD, Johnson KP. 2015. Patterns of diversification in small New World ground doves are consistent with
287   major geologic events. *Auk* **132**:300–312. doi:10.1642/AUK-14-193.1
288   Taylor SA, Larson EL. 2019. Insights from genomes into the evolutionary importance and prevalence of
289   hybridization in nature. *Nat Ecol Evol* **3**:170.
290   Than C, Ruths D, Nakhleh L. 2008. PhyloNet: a software package for analyzing and reconstructing reticulate
291   evolutionary relationships. *BMC Bioinformatics* **9**:322.
292   Wen D, Yu Y, Zhu J, Nakhleh L. 2018. Inferring Phylogenetic Networks Using PhyloNet. *Syst Biol* **67**:735–740.
293   Whiteman NK, Santiago-Alarcon D, Johnson KP, Parker PG. 2004. Differences in straggling rates between two
294   genera of dove lice (Insecta: Phthiraptera) reinforce population genetic and cophylogenetic patterns. *Int J*
295   *Parasitol* **34**:1113–1119.
296   Yu Y, Nakhleh L. 2015. A maximum pseudo-likelihood approach for phylogenetic networks. *BMC Genomics*
297   **16**:Suppl 10: S10.

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Fig. 1. Diagram depicting the ecological replicate system and the hypothesis of this study. Wing lice (*Columbicola*) have higher dispersal abilities than body lice (*Physconelloides*), and thus higher odds of encountering individuals of a different louse species on the same host. Thus, wing lice are predicted to show higher levels of introgression compared to body lice.

306 **Fig. 2.** Boxplot showing the differences in levels of introgression between wing (green) and body
307 (orange) lice. Level of introgression represents the sum of the mean coverage of reads mapped
308 from all the species excluding the focal louse species, divided by the mean coverage of the focal
309 louse species (see Methods). Black dots represent individual samples (horizontally jittered).

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312 **Fig. 3.** Optimal phylogenetic networks of feather lice genera. Orange branches depict
313 reticulations. From left to right, *Columbicola* (seven reticulations) and *Physconelloides* (four
314 reticulations) networks (See Methods).