Analysis of genetic networks regulating refractive eye development in collaborative cross progenitor strain mice reveals new genes and pathways underlying human myopia

Short title: Refractive eye development is regulated by evolutionary conserved genetic networks

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Abstract

Background: Population studies suggest that genetic factors play an important role in refractive error development; however, the precise role of genetic background and the composition of the signaling pathways underlying refractive eye development remain poorly understood.

Methods: Here, we analyzed normal refractive development and susceptibility to form-deprivation myopia in the eight progenitor mouse strains of the Collaborative Cross (CC). We used RNA-seq to analyze gene expression in the retinae of these mice and reconstruct genetic networks and signaling pathways underlying refractive eye development. We also utilized genome-wide gene-based association analysis to identify mouse genes and pathways associated with myopia in humans.

Results: Genetic background strongly influenced both baseline refractive development and susceptibility to environmentally-induced myopia. Baseline refractive errors ranged from -21.2 diopters (D) in 129S1/svlmj mice to +22.0 D in CAST/EiJ mice and represented a continuous distribution typical of a quantitative genetic trait. The extent of induced form-deprivation myopia ranged from -5.6 D in NZO/HILtJ mice to -20.0 D in CAST/EiJ mice and also followed a continuous distribution. Whole-genome (RNA-seq) gene expression profiling in retinae from CC progenitor strains identified genes whose expression level correlated with either baseline refractive error or susceptibility to myopia. Expression levels of 2,302 genes correlated with the baseline refractive state of the eye, whereas 1,917 genes correlated with susceptibility to induced myopia. Genome-wide gene-based association analysis in the CREAM and UK Biobank human

cohorts revealed that 985 of the above genes were associated with myopia in humans, including 847 genes which were implicated in the development of human myopia for the first time. Although the gene sets controlling baseline refractive development and those regulating susceptibility to myopia overlapped, these two processes appeared to be controlled by largely distinct sets of genes. **Conclusions:** Comparison with data for other animal models of myopia revealed that the genes identified in this study comprise a well-defined set of retinal signaling pathways, which are highly conserved across different vertebrate species. These results identify major signaling pathways involved in refractive eye development and provide attractive targets for the development of anti-myopia drugs.

Keywords: myopia; refractive eye development; genetic networks; signaling pathways; genetic variation; RNA-seq; gene-based genome-wide association analysis; evolutionary conservation of pathways

Background

Myopia is the most common ocular disorder worldwide [1]. The prevalence of myopia in the U.S. has increased from 25% to ~48% in the last 40 years [2-4]. The worldwide prevalence of myopia is predicted to increase from the current 25% to 50% in the next three decades [5], while the prevalence already exceeds 80% in several parts of Asia [6, 7]. Myopia often leads to serious blinding complications such as myopic maculopathy, retinal floaters, chorioretinal atrophy, retinoschisis, retinal tears, retinal detachment, and myopic macular degeneration [8-24]. It also represents a major risk factor for a number of other serious ocular pathologies such as cataract and glaucoma [9, 10, 25-27]. Because of the increasing prevalence, myopia is rapidly becoming

one of the leading causes of vision loss in several parts of the world, and World Health Organization designated myopia as one of five priority health conditions [1, 8-10, 28].

Development of myopia is controlled by both environmental and genetic factors [29-32]. Although environmental factors, such as reading and nearwork, play a very important role in the development of myopia [33-36], genetic studies suggest that the impact of environmental factors on refractive development is determined by genetic variation in "myopia-susceptibility genes" [37]. The role of genetic background in refractive eye development is also supported by animal studies, which revealed that the extent of myopia experimentally induced in animal models is strongly influenced by genetic background [38-41]. Analysis of the size of ocular components in different strains of mice suggested a significant role of genetic background in the regulation of refractive eye development [42, 43]. Wong and Brown [44] also described significant differences in visual detection, pattern discrimination and visual acuity among different strains of mice. The contribution of genetic factors to myopia has been estimated to be as high as 70-80% [45-50], and human genetic mapping studies have identified over 270 chromosomal loci linked to myopia [51-54]. These loci implicate genes involved in multiple cellular and biological processes related to extracellular matrix organization, eye morphogenesis, retinal signaling, and visual perception [53, 54]. Gene expression profiling studies also showed that development of myopia is accompanied by changes in gene expression in the retina, choroid, and sclera [39, 55-62]. Moreover, these studies suggested that similar biological processes underlie refractive development in animal models and humans [63].

Although these studies revealed important roles of genetic variation and gene expression in the development of refractive errors, the relationship between genetic background, gene expression and development of refractive errors remains unexplored. Here, we systematically

analyzed the role of genetic background in the regulation of retinal gene expression and signaling pathways underlying refractive eye development in eight inbred strains of mice and their association with human myopia using genome-wide gene expression profiling (RNA-seq) and gene-based genome-wide association analysis in the CREAM and UK Biobank human cohorts. We found that both baseline refractive state of the eye and susceptibility to myopia are inherited as quantitative traits, demonstrating strong dependence on genetic background. Furthermore, genetic background strongly influenced expression of genes in the retina and modulated a well-defined set of signaling pathways highly conserved in chickens, mice, monkeys, and humans. Our data suggest that refractive eye development is regulated by hundreds to thousands of genes across ocular tissues and point to high evolutionary conservation of signaling pathways underlying refractive development across vertebrate species.

Methods

Ethics statement

Mice were obtained from the Jackson Laboratory (Bar Harbor, ME) and were maintained as an in-house breeding colony. All procedures adhered to the Association for Research in Vision and Ophthalmology (ARVO) statement on the use of animals in ophthalmic and vision research and were approved by the Columbia University Institutional Animal Care and Use Committee. Animals were anesthetized via intraperitoneal injection of ketamine (90 mg/kg) and xylazine (10 mg/kg) and were euthanized using CO_2 followed by cervical dislocation.

All human studies were approved by the relevant institutional review boards and/or medical ethics committees and conducted according to the Declaration of Helsinki. All CREAM

participants provided written informed consent. The UK Biobank received ethical approval from the National Health Service National Research Ethics Service (reference 11/NW/0382).

Analysis of refractive state of the eyes in mice

To examine the effect of genetic background on the refractive state of the eye in mice, we analyzed baseline refractive errors in the eight strains of mice comprising Collaborative Cross. The refractive state of both left and right eyes was determined on alert animals at P40 using an automated eccentric infrared photorefractor as previously described [64, 65]. The animal to be refracted was immobilized using a restraining platform, and each eye was refracted along the optical axis in dim room light (< 1 lux), 20-30 min. after instilling 1% tropicamide ophthalmic solution (Alcon Laboratories) to ensure mydriasis and cycloplegia. Five independent measurement series (~300-600 measurements each) were taken for each eye. The measurements were automatically acquired by the photorefractor every 16 msec. Each successful measurement series (i.e., Purkinje image in the center of the pupil and stable refractive error for at least 5 sec.) was marked by a green LED flash, which was registered by the photorefractor software. Sixty individual measurements from each series, immediately preceding the green LED flash, were combined, and a total of 300 measurements (60 measurements x 5 series = 300 measurements) were collected for each eye. Data for the left and right eyes were combine (600 measurements total) to calculate mean refractive error and standard deviation for each animal.

Analysis of form-deprivation myopia in mice

To examine the effect of genetic background on susceptibility to environmentally induced myopia in mice, we analyzed the extent of myopia induced by the diffuser-imposed retinal image

degradation (visual form deprivation) in the eight strains of mice comprising Collaborative Cross. Visual input was degraded in one of the eyes by applying plastic diffusers, and refractive development of the treated eye was compared to that of the contralateral eye, which was not treated with a diffuser, as previously described [66, 67]. Diffusers represented low-pass optical filters, which degraded the image projected onto the retina by removing high spatial frequency details. Frosted hemispherical plastic diffusers were hand-made from zero power rigid contact lenses made from OP3 plastic (diameter = 7.0 mm, base curve = 7.0 mm; Lens.com). Lenses were frosted using a fine sandpaper and inserted into a 3D-printed plastic frames (Proto Labs). On the first day of the experiment (P24), animals were anesthetized via intraperitoneal injection of ketamine and xylazine, and frames with diffusers were attached to the skin surrounding the right eye with six stitches using size 5-0 ETHILON™ microsurgical sutures (Ethicon) and reinforced with Vetbond™ glue (3M Animal Care Products) (the left eye served as a control). Toenails were covered with adhesive tape to prevent mice from removing the diffusers. Animals recovered on a warming pad and were then housed under low-intensity constant light in transparent plastic cages for the duration of the experiment as previously described [66, 67]. Following 21 days of visual form deprivation (from P24 through P45), diffusers were removed and refractive status of both treated and control eyes was assessed using an automated eccentric infrared photorefractor as previously described [64, 65]. The interocular difference in refraction between the treated and contralateral control eye served as an indication of the extent of induced myopia.

RNA extraction and RNA-seq

Animals were euthanized following an IACUC-approved protocol. Eyes were enucleated, the retinae were dissected from the enucleated eyes and the choroid/RPE removed. The retinae were washed in RNAlater (Thermo Fisher Scientific) for 5 min., frozen in liquid nitrogen, and stored at -80°C until processed for this study. To isolate RNA, tissue samples were homogenized at 4°C in a lysis buffer using Bead Ruptor 24 tissue homogenizer (Omni). Total RNA was extracted from each tissue sample using miRNAeasy mini kit (QIAGEN) following the manufacturer's protocol. The integrity of RNA was confirmed by analyzing 260/280 nm ratios ($Ratio_{260/280}$ = 2.11-2.13) on a Nanodrop (Thermo Scientific) and the RNA Integrity Number (RIN = 9.0-10.0) using Agilent Bioanalyzer. Illumina sequencing libraries were constructed from 1 µg of total RNA using the TruSeq Stranded Total RNA LT kit with the Ribo-Zero Gold ribosomal RNA depletion module (Illumina). Each library contained a specific index (barcode) and were pooled at equal concentrations using the randomized complete block (RCB) experimental design before sequencing on Illumina HiSeq 2500 sequencing system. The number of libraries per multiplexed sample was adjusted to ensure sequencing depth of ~70 million reads per library (paired-end, 2 x 100 nucleotides). The actual sequencing depth was 76,773,554 ± 7,832,271 with read quality score 34.5 ± 0.4.

Post-sequencing RNA-seq data validation and analysis

The FASTQ raw data files generated by the Illumina sequencing system were imported into Partek Flow software package (version 7.0.18.1210, Partek), libraries were separated based on their barcodes, adapters were trimmed and remaining sequences were subjected to pre-alignment quality control using Partek Flow pre-alignment QA/QC module. After the assessment of various quality metrics, bases with the quality score < 34 were removed (≤ 5 bases) from each end.

Sequencing reads were then mapped to the mouse reference genome Genome Reference Consortium Mouse Build 38 (GRCm38/mm10, NCBI) using the STAR aligner (version 2.5.2b) resulting in $95.0 \pm 0.4\%$ mapped reads per library, which covered $35.4 \pm 1.0\%$ of the genome. Aligned reads were quantified to transcriptome using Partek E/M annotation model and the NCBI's RefSeq Transcripts 80 annotation file to determine read counts per gene/genomic region. The generated read counts were normalized by the total read count and subjected to the analysis of variance (ANOVA) to detect genes whose expression correlates with either refractive error or susceptibility to myopia. Differentially expressed transcripts were identified using a P-value threshold of 0.05 adjusted for genome-wide statistical significance using Storey's q-value algorithm [68]. To identify sets of genes with coordinate expression, differentially expressed transcripts were clustered using Partek Flow hierarchical clustering module using average linkage for the cluster distance metric and Euclidean distance metric to determine the distance between data points. Each RNA-seq sample was analyzed as a biological replicate, thus, resulting in three biological replicates per strain.

Gene ontology analysis and identification of canonical signaling pathways

To identify biological functions (gene ontology categories), which were significantly affected by the genes whose expression correlated with either baseline refractive errors or susceptibility to myopia, we used the database for annotation, visualization and integrated discovery (DAVID) version 6.8 [69] and GOplot R package [70]. DAVID uses a powerful gene-enrichment algorithm and DAVID Gene Concept database to identify biological functions (gene ontology categories) affected by differential genes, while GOplot integrates gene ontology information with gene expression information and predicts the effect of gene expression changes on

biological processes. DAVID uses a modified Fisher's exact test (EASE score) with a P-value threshold of 0.05 to estimate statistical significance of enrichment for specific gene ontology categories. IPA Pathways Activity Analysis module (QIAGEN) was used to identify canonical pathways effected by the genes involved in baseline refractive eye development or regulating susceptibility to myopia, and predict the effect of gene expression changes in different strains on specific pathways. The activation z-score was employed in the IPA Pathways Activity Analysis module to predict activation or suppression of the canonical pathways. The z-score algorithm is designed to reduce the chance that random data will generate significant predictions. The z-score provides an estimate of statistical quantity of change for each pathway found to be statistically significantly affected by the changes in gene expression. The significance values for the canonical pathways were calculated by the right-tailed Fisher's exact test. The significance indicates the probability of association of molecules from a dataset with the canonical pathway by random chance alone. Pathways Activity Analysis module determines if canonical pathways, including functional end-points, are activated or suppressed based on the gene expression data in a dataset. Once statistically significant canonical pathways were identified, we subjected the datasets to the Core Functional Analysis in IPA to compare the pathways and identify key similarities and differences in the canonical pathways underlying baseline refractive development and susceptibility to myopia.

Identification of candidate genes for human myopia within known myopia QTLs

To identify candidate genes for myopia in the QTLs previously found to be linked to human myopia, we compared the genes that we found to be involved in refractive eye development in mice with a list of genes located within human myopia QTLs. We first compiled a list of all

SNPs or markers exhibiting statistically significant association with myopia in the human linkage or GWAS studies. LDlink's LDmatrix tool (National Cancer Institute) was used to identify SNPs in linkage disequilibrium and identify overlapping chromosomal loci. We then used UCSC Table Browser to extract all genes located within critical chromosomal regions identified by the human linkage studies or within 200 kb (±200 kb) of the SNPs found by GWAS. The list of genes located within human QTLs was compared with the list of genes that we found to be associated with either baseline refractive errors or susceptibility to myopia in mice using Partek Genomics Suite (Partek). The statistical significance of the overlaps was estimated using probabilities associated with the hypergeometric distribution using Bioconductor software package GeneOverlap version 1.14.0 and associated functions.

Identification of genes associated with refractive error in the human population using gene-based genome-wide association analysis

To identify genes associated with the development of refractive errors in humans among the genes whose expression correlated with refractive eye development in mice, human homologs of candidate mouse genes were examined for association with refractive error in the international genome-wide association study (GWAS) of refractive error carried out by the Consortium for Refractive Error and Myopia (CREAM) [54] and the UK Biobank Eye and Vision consortium sample [71] using the Multi-marker Analysis of GenoMic Annotation (MAGMA) [72]. Human homologs of candidate mouse genes were obtained from Ensembl BioMart and were mapped according to gene definitions in the NCBI Entrez Gene database. Genes were defined according to NCBI build 37 (hg19/GRCh37) coordinates with 200kb flanking regions appended to the transcription start/stop sites. LD patterns were estimated by MAGMA using the 1000 Genomes

Phase 1, version 3 European ancestry reference panel. As summary statistics were used as input, MAGMA gene-based analysis was performed using the default "snp-wise=mean" model.

The CREAM sample included 148,485 individuals of European ancestry from 28 cohorts and 11,935 individuals of Asian ancestry from eight studies. All participants included in this analysis from CREAM were 25 years of age or older.

UK Biobank is a large prospective study following the health and wellbeing of approximately 500,000 UK residents aged between 40 and 69 years-old at the baseline recruitment visit (during the period 2006–2010). 130,521 participants had non-cycloplegic autorefraction performed for at least one eye using the Tomey RC 5000 autorefractor-keratometer (Tomey Corp., Nagoya, Japan), with up to ten measurements taken for each eye. After the exclusion of unreliable readings, 130,459 participants had measures for refractive astigmatism and spherical equivalent refractive error.

Participants with conditions that might alter refraction, such as cataract surgery, laser refractive procedures, retinal detachment surgery, keratoconus, or ocular or systemic syndromes were excluded from the analyses. Refractive error was represented by measurements of refraction and analyzed as spherical equivalent (SphE□=□spherical refractive error +1/2-cylinder refractive error).

Results

Genetic background modulates baseline refractive eye development and susceptibility to myopia in mice

Genetic background was shown to influence the size of ocular components in mice. To investigate whether genetic differences between mice would have an impact on the baseline

refractive state of the eye and susceptibility to myopia, we analyzed baseline refractive development and susceptibility to form-deprivation myopia in eight inbred strains of mice, which served as founder strains for the Collaborative Cross (CC) [73], i.e., 129S1/svlmj, A/J, C57BL/6J, CAST/EiJ, NOD/ShiLtJ, NZO/HlLtJ, PWK/PhJ, and WSB/EiJ mice.

We first measured baseline refractive errors in all eight strains at P40, i.e., when baseline refractions reach a plateau in mice [64]. We found that C57BL/6J mice were emmetropic on average (+0.3 ± 0.9 D). CAST/EiJ, NZO/HlLtJ, PWK/PhJ, and WSB/EiJ mice exhibited various degrees of hyperopia ranging from +10.6±2.2 D to +22±4.0 D, whereas A/J, NOD/ShiLtJ, and 129S1/svlmj mice developed various degrees of myopia ranging from -3.5±3.6 D to -21.2±3.9 D (Additional file 1: Table S1, Fig. 1A). The differences between the strains were statistically significant as revealed by ANOVA ($F_{(7, 145)} = 429.76$, $P < 0.00001$). More importantly, the distribution of refractive errors in the CC mice was continuous, suggesting that refractive state of the eye in mice is inherited as a quantitative trait.

We then analyzed susceptibility to myopia in the same strains of mice by evaluating the extent of induced form-deprivation myopia (Additional file 2: Table S2, Fig. 1B). Susceptibility to form-deprivation myopia was estimated by applying a diffuser to one eye at P24 and comparing the extent of induced myopia in the form-deprived eye versus the contralateral control eye after 21 days of treatment. We found large differences in susceptibility to induced myopia between the strains (Additional file 2: Table S2, Fig 1B), which ranged from -5.5 ± 2.1 D in NZO/HlLtJ mice to -18.7 ± 3.1 D in CAST/EiJ mice. Other strains occupied intermediate positions between NZO/HlLtJ and CAST/EiJ mice and differences between the strains in the extent of induced myopia were statistically significant as revealed by ANOVA ($F_{(7, 48)} = 9.8$, $P < 0.00001$). The distribution of induced refractive errors was continuous, similar to baseline

refractive errors, suggesting that susceptibility to myopia was also inherited as a quantitative trait in mice. Spearman's rank-order correlation analysis showed that there was no statistically significant correlation between baseline refractive errors and the extent of induced myopia (r_s = -0.60, P = 0.12). Collectively, these data suggest that genetic background plays important role in baseline refractive development and susceptibility to environmentally induced myopia in mice and that both baseline refractive error and susceptibility to myopia are inherited as quantitative traits.

Large number of genes are involved in regulation of baseline refractive error in mice via multiple retinal biological processes and signaling pathways

To identify retinal genes influencing baseline refractive eye development in mice, we used RNA-seq to analyze gene expression in the retina of eight CC strains at P28 (an age when refractive development is progressing towards its stable plateau) (Fig 2). We found that expression of 2,302 retinal genes strongly correlated with the baseline refractive state (Additional file 3: Table S3, Fig 2A). Genes were organized in two distinct clusters. Expression of 793 genes comprising the first cluster was positively correlated with hyperopia, i.e., expression was increased in the strains with positive refractive errors and decreased in the strains with negative refractive errors. Conversely, expression of 1,509 genes comprising the second cluster was positively correlated with myopia, i.e., expression of these genes was increased in the mouse strains with negative refractive errors and decreased in the strains with positive refractive errors. We observed a clear transition from the "hyperopic" gene expression pattern in CAST/EiJ and PWK/PhJ mice with highly hyperopic refractive errors to the "myopic" gene expression pattern in 129S1/svlmj mice with highly myopic refractive errors. Other strains occupied intermediate positions between these

two extremes and exhibited transitional patterns of gene expression, which correlated with average baseline refractive errors in these strains.

Gene ontology analysis revealed that the 2,302 genes whose expression correlated with baseline refractive error were associated with 116 biological processes, 77 cellular components, and 69 molecular functions in the retina (Additional file 4: Table S4). Biological processes involved in the regulation of baseline refractive development ranged from regulation of neurogenesis and neuron migration to regulation of DNA methylation, visual perception, and synaptic vesicle endocytosis. Figure 2B shows the top 15 biological processes associated with these genes, including regulation of protein kinase B, regulation of transcription and translation, covalent chromatin modification, insulin receptor signaling, dendrite morphogenesis, and response to oxidative stress, among others. Genes underlying baseline refractive development were also associated with multiple canonical signaling pathways in the retina (Additional file 5: Table S5, Fig 2C). Negative refractive errors were associated with activation of mTOR, EIF2, AMPK, β-adrenergic, and dopamine-DARPP32 feedback signaling pathways and suppression of HIPPO and RhoGDI signaling pathways, among others. Taken together, these data suggest that refractive eye development is regulated by a large number of genes and pathways. In summary, the development of hyperopic and myopic refractive errors was associated with specific patterns of gene expression, and the activation or suppression of many retinal signaling pathways.

Large number of genes are involved in regulation of susceptibility to myopia in mice via multiple retinal biological processes and signaling pathways

We found that genetic background influences susceptibility to experimentally induced myopia in mice and that susceptibility to myopia appears to be controlled as a quantitative trait by multiple

genes (Fig 1B). To identify genes underlying inter-strain differences in susceptibility to myopia in mice, we analyzed gene expression in the retina of the eight CC strains upon induction of form-deprivation myopia at the whole-genome level, using RNA-seq (Fig 3). We found that expression of 1,917 genes strongly correlated with the extent of form-deprivation myopia in different strains of mice (Additional file 6: Table S6, Fig 3A). Similar to what we found with baseline refractive errors, these genes were organized in two clusters. Expression of 643 genes comprising the first cluster was positively correlated with the susceptibility to myopia, whereas expression of 1,274 genes in the second cluster was negatively correlated with the susceptibility to myopia. We found that there was a transition from a "high susceptibility" gene expression pattern in CAST/EiJ mice, which developed -18.7 ± 3.1 D of myopia, to a "low susceptibility" gene expression pattern in NZO/HlLtJ mice, in which only -5.5 ± 2.1 D of myopia was induced by visual form deprivation. Other strains exhibited transitional gene expression patterns, which correlated with the extent of induced myopia.

Gene ontology analysis suggested that 55 biological processes, 61 cellular components, and 41 molecular functions were associated with the 1,917 genes correlated with susceptibility to myopia (Additional file 7: Table S7). Figure 3B shows the top 15 biological processes involved in the regulation of susceptibility to myopia, including regulation of signal transduction, cell-cell adhesion, transcription, translation, protein transport, and lysosome organization, among others. Analysis of the canonical pathways influenced by the genes correlated with susceptibility to myopia revealed that increased susceptibility to myopia was associated with suppression of mTOR signaling, EIF2 signaling, protein kinase A signaling, D-myo-inositol-5-phosphate metabolism, cholesterol and choline biosynthesis, as well as with activation of amyloid processing, HIPPO signaling, PTEN signaling, and PPARα/RXRα signaling pathways

(Additional file 8: Table S8, Fig 3C). Collectively, these data implicate an elaborate retinal genetic network and multiple signaling pathways in the regulation of susceptibility to myopia in mice.

Baseline refractive eye development and susceptibility to myopia in mice are regulated via overlapping but largely distinct retinal genetic networks

To estimate the relative contribution of genes whose expression level correlated with baseline refractive error versus susceptibility to myopia, we analyzed the overlap between the two gene sets (Fig 4). We found that 714 genes were correlated with both baseline refractive development and susceptibility to myopia (Additional file 9: Table S9, Fig 4A). Gene ontology analysis revealed that these 714 genes were associated with 24 biological processes, 24 cellular components, and 14 molecular functions (Additional file 10: Table S10). Figure 4B shows the top 15 biological processes for the overlapping genes. Interestingly, the majority of these biological processes were suppressed in animals with high susceptibility to myopia (lower panel) and were activated in animals with negative baseline refractive errors (upper panel). A similar trend was observed when we compared canonical signaling pathways affected by the overlapping genes (Additional file 11: Table S11, Fig 4C). Signaling pathways, which were activated in mice with highly negative baseline refractive errors, were suppressed in mice with high susceptibility to myopia, and vice versa. The top pathways associated with both baseline refractive development and susceptibility to myopia were EIF2 signaling, protein kinase A signaling, regulation of eIF4 and p70S6K signaling, mTOR pathway, HIPPO pathway, and axonal guidance signaling. Figure 5 shows a summary of all signaling pathways correlated with baseline refractive eye development and susceptibility to myopia. In addition to the pathways listed

above, a number of other pathways were also implicated in both baseline refractive development and susceptibility to myopia, including GP6 signaling pathway, melatonin signaling, RhoGDI signaling, PTEN signaling, opioid signaling pathway, PPARα/RXRα activation, PI3K/AKT signaling, estrogen receptor signaling, and tight junction signaling. Conversely, synaptic long-term potentiation, α-adrenergic and β-adrenergic signaling, androgen and aldosterone signaling, ephrin receptor signaling, relaxin signaling, dopamine-DARPP32 feedback signaling, dopamine receptor signaling, eNOS and nNOS signaling, somatostatin receptor 2 signaling, neurotrophin/TRK signaling, protein ubiquitination pathway, gap junction signaling, phototransduction pathway, and several other pathways were associated with baseline refractive development but not susceptibility to myopia. The amyloid processing, IGF-1 signaling, DNA methylation and transcriptional repression signaling, epithelial adherens junction signaling, iron homeostasis signaling pathway, RAR activation, RAN signaling, and several other pathways were associated with susceptibility to myopia but not baseline refractive error. Thus, these data suggest that baseline refractive development and susceptibility to myopia are regulated by genetic networks with considerable overlap; however, the two genetic networks have substantial unique components, which may independently regulate either baseline refractive development or susceptibility to environmentally induced myopia.

Many genes regulating baseline refractive eye development or susceptibility to myopia in mice are localized within chromosomal loci linked to human myopia

Genes comprising genetic networks underlying important developmental and physiological processes often harbor mutations causing human diseases. Therefore, to identify genes associated with susceptibility to myopia in humans, we analyzed the overlap between the genes that we

found to be associated with either baseline refractive development or susceptibility to myopia in mice and genes located within human quantitative trait loci (QTLs) linked to myopia (Additional file 12: Table S12, Fig 6, 7 and 8). We found that 90 genes whose expression correlated with baseline refractive errors, 51 genes whose expression correlated with susceptibility to myopia, and 23 genes whose expression correlated with both baseline refractive errors and susceptibility to myopia in mice were localized within human QTLs linked to myopia (Additional file 13: Table S13, Additional file 14: Table S14, Additional file 15: Table S15, Fig 6). GeneOverlap analysis (Fig 6) revealed that the overlaps for the genes involved in baseline refractive development and genes involved in the regulation of susceptibility to myopia were highly significant (OR = 2.75, $P = 3.3 \times 10^{-18}$; OR = 2.03, $P = 1.2 \times 10^{-07}$; respectively). The overlap between the genes located within human QTLs and genes associated with both baseline refractive development and susceptibility to myopia in mice was weaker (OR = 1.62, $P = 0.022$); however, overall, these data suggest a functional association between genes associated with refractive eye development in mice and genes causing myopia in humans. GeneOverlap analysis revealed that a total of 164 mouse genes were located within 109 human QTLs, producing 1.5 candidate genes per QTL (Additional file 16: Table S16, Fig 7 and 8).

We next analyzed biological processes linked to the genes associated with refractive development in mice and localized within human myopia QTLs (Additional file 17: Table S17, Additional file 18: Table S18, Fig 9). Surprisingly, we found that although several biological processes, such as cell growth and proliferation, circadian regulation of gene expression, and regulation of neuron differentiation, were implicated in both baseline refractive development and regulation of susceptibility to myopia, many biological processes underlying baseline refractive development and susceptibility to myopia appeared to be different. Our data on the genes

associated with baseline refractive development in mice and localized in human QTLs implicate several unique processes, including camera-type eye development, regulation of ion/calcium transport, regulation of membrane polarization during action potential and long-term synaptic potentiation, nitric oxide signaling, ephrin receptor signaling, glucocorticoid receptor signaling, regulation of glutamate secretion, and regulation of JAK-STAT and MAPK signaling cascades (Additional file 17: Table S17, Fig 9A). Conversely, the genes associated with susceptibility to myopia in mice and localized in human QTLs highlighted several different processes, including regulation of cell shape and cell migration, dorsal/ventral pattern formation, vasculature morphogenesis, photoreceptor cell function and development, cellular response to DNA damage, and small-GTPase-mediated signal transduction (Additional file 18: Table S18, Fig 9B). Collectively, these data suggest that there is a significant functional overlap between genes we found to be correlated with refractive eye development in mice and genes causing myopia in humans. Moreover, our data suggest that development of refractive errors in humans is associated with genes that, in mice, regulate both baseline refractive development and susceptibility to refractive changes induced by the visual environment.

Gene-based genome-wide association analysis of mouse genes identifies novel gene candidates for human myopia

To build on the significant overlap observed above between genes associated with refractive eye development in mice and human myopia QTLs, we examined whether the mouse genes whose expression level correlated with refractive error or myopia susceptibility were enriched for genetic variants associated with refractive error in humans. Human genes enriched for variants associated with refractive error were identified with MAGMA (Multi-marker Analysis of

GenoMic Annotation), using single-marker summary statistics from a genome-wide association study (GWAS) for refractive error and age-of-onset-of-spectacle-wear reported by the CREAM Consortium (N=160,420 participants; [54]) or a GWAS for refractive error by the UK Biobank Eye & Vision Consortium (N= 88,005 participants; [71]) as input. Flanking regions of 200 kb upstream and downstream of transcription start and stop sites were included, in order to capture regulatory variants influencing the expression of nearby genes.

Of the 2,302 genes associated with baseline refractive development in mice for which there were human homologs, 277 genes in the CREAM dataset were associated with refractive error in humans (*FDR* < 0.05), with 86 genes surviving Bonferroni correction ($P_{Bonferroni}$ < 0.05) (Additional file 19: Table S19). Sixty-seven of these genes (including 43 genes with $P_{Bonferroni}$ < 0.05) were localized within previously identified human QTLs (Fig 7 and 8), while 210 of the 277 genes (including 43 genes with $P_{Bonferroni}$ < 0.05) have not previously been implicated in the development of refractive errors in the human population (Additional file 19: Table S19). When the above analysis was repeated using the (independent) UK Biobank dataset, 560 genes were associated with refractive error (*FDR* < 0.05), including 156 genes with genome-wide significance ($P_{Bonferroni}$ < 0.05) (Additional file 20: Table S20). 488 of the 560 genes (including 104 genes with $P_{Bonferroni}$ < 0.05) have not previously been implicated in the development of refractive errors in humans (Additional file 20: Table S20). Importantly, 190 genes associated with baseline refractive development in mice were replicated in both the CREAM and UK Biobank cohorts, including 69 genes which achieved genome-wide significance ($P_{Bonferroni}$ < 0.05) (Additional file 23: Table S23).

Of the 1,917 genes associated with the regulation of susceptibility to myopia in mice, gene-based analysis using the CREAM dataset identified 223 genes associated with refractive

error in humans (*FDR* < 0.05), including 72 genes at genome-wide significance ($P_{Bonferroni}$ < 0.05) (Additional file 21: Table S21). Forty-six of these genes (including 33 genes with $P_{Bonferroni}$ < 0.05) were localized within previously identified human QTLs (Fig 7 and 8), whereas 177 of the 223 genes (including 39 genes with $P_{Bonferroni}$ < 0.05) have not previously been implicated in the development of refractive errors in humans (Additional file 21: Table S21). MAGMA analysis of the 1,917 genes using the UK Biobank dataset revealed that 465 genes were associated with refractive error (*FDR* < 0.05), including 119 genes that withstood Bonferroni correction ($P_{Bonferroni}$ < 0.05) (Additional file 22: Table S22). Forty-nine of these genes (including 35 genes with $P_{Bonferroni}$ < 0.05) were localize within known human myopia QTLs (Fig 7 and 8), whereas 416 of the 465 genes (including 84 genes with $P_{Bonferroni}$ < 0.05) were not previously known to be associated with the development of refractive errors in the human population (Additional file 22: Table S22). One hundred and fifty-two genes involved in the regulation of susceptibility to myopia in mice were found to be linked to the development of refractive errors in both CREAM and UK Biobank cohorts, including 53 genes which reached genome-wide significance ($P_{Bonferroni}$ < 0.05) in both samples (Additional file 24: Table S24).

Analysis of the biological processes associated with the above genes linked to either baseline refractive development or regulation of susceptibility to myopia in humans (Additional file 25: Table S25, Additional file 26: Table S26, Fig 10A and 11A) revealed processes primarily related to calcium-mediated cell adhesion, synapse assembly, synaptic transmission, protein translation, small GTPase-mediated signal transduction, and GABA receptor signaling. Several canonical signaling pathways were also identified, including EIF2 and mTOR signaling, eIF4 and p70S6K signaling, epithelial adherence junction signaling, sumoylation pathway, and

regulation of cellular mechanics by calpain protease, among others (Additional file 27: Table S27, Additional file 28: Table S28, Fig 10B and 11B).

However, there were also substantial differences between the two sets of human genes identified using either the mouse baseline refractive error or the mouse myopia susceptibility gene sets. The mouse baseline refractive error-derived human gene set was associated with nervous system development, post-embryonic camera-type eye development, retinal cone development, neuron migration, dendrite morphogenesis, regulation of glutamate metabolism, extracellular matrix organization, and beta-amyloid formation (Additional file 25: Table S25, Fig 10A). This gene set was associated with several canonical pathways distinct from those identified using the mouse susceptibility to myopia-derived human gene set. These pathways included integrin signaling, semaphorin signaling in neurons, glucocorticoid receptor signaling, phospholipase C signaling, synaptic long term potentiation, ephrin receptor signaling, nNOS signaling in neurons, and estrogen receptor signaling (Additional file 27: Table S27, Additional file 28: Table S28, Fig 10B and 11B).

The mouse susceptibility to myopia-derived human gene set, on the other hand, was associated with biological processes related to developmental growth, neuron fate commitment, regulation of mesenchymal cell proliferation, potassium ion transmembrane transport, protein transport, response to estradiol and cholesterol, as well as cellular response to hypoxia (Additional file 26: Table S26, Fig 11A). Analysis of canonical pathways suggested that pathways associated with TGF-β signaling, PPARα/RXRα activation, PTEN signaling, regulation of the epithelial-mesenchymal transition, STAT3 signaling, regulation of stem cell pluripotency, VEGF and IGF-1 signaling, NRF2-mediated oxidative stress response, and

PI3K/AKT signaling, among others, were involved in the regulation of susceptibility to myopia (Additional file 28: Table S28, Fig 11B).

Discussion

Human population studies and studies in animal models strongly suggest that both environmental and genetic factors play important roles in refractive eye development. Numerous linkage and genome-wide association studies in humans identified over 270 chromosomal loci linked to the development of myopia in humans [74]; however, very little is known how genetic variation causing myopia affects gene expression and how changes in gene expression associated with differences in genetic backgrounds affect refractive eye development and susceptibility to myopia.

Our data suggest that differences in genetic backgrounds play a very important role in refractive eye development and regulation of susceptibility to myopia. Moreover, we found that variations in genetic background produce a continuous distribution of refractive errors and susceptibilities to myopia in a mouse population, characteristic of quantitative traits. Genetic variations in different mouse strains also produce unique patterns of gene expression, which strongly correlate with either baseline refractive errors or susceptibility to induced myopia. Surprisingly, we found that the baseline refractive development and susceptibility to myopia are controlled by largely distinct sets of genes, which suggests that signaling pathways that regulate the trajectory of refractive eye development towards emmetropia, myopia, or hyperopia might be different from the pathways that modulate the impact of optical defocus and other environmental factors on refractive development. Nevertheless, expression of 714 genes correlated with both baseline refractive errors and susceptibility to myopia, suggesting that at least some genes

control both the pathways regulating the trajectory of refractive development and the impact of visual input on it.

Genes that influenced either baseline refractive development or susceptibility to myopia affected a long list of biological and molecular functions; however, most noteworthy is the finding that over 29 canonical signaling pathways were involved in both baseline refractive development and regulation of susceptibility to myopia. Interestingly, the majority of these pathways were suppressed in animals with high susceptibility to myopia and activated in animals with negative baseline refractive errors. The exceptions from this rule were sumoylation, RhoGDI signaling, PTEN, and HIPPO signaling pathways, which we found to be activated in animals with high susceptibility to myopia and suppressed in animals with negative baseline refractive errors. The observation that the same pathways were effected in the opposite directions in animals with high susceptibility to myopia and animals with highly negative baseline refractive errors may be explained by the role of optical defocus in the development of myopia. Considering that mice are housed in small cages and the exposure to distant vision is limited, animals with hyperopic refractive errors would be exposed to high levels of hyperopic optical defocus (which was shown to cause myopia) compared to the animals with negative refractive errors; thus explaining why signaling pathways are effected in the same direction in animals with high susceptibility to myopia and animals with hyperopic baseline refractive errors.

Remarkably, we also found that many genes whose expression correlated with either baseline refractive errors or susceptibility to myopia in mice were localized in the known human QTLs linked to myopia. The majority of these genes (90 genes) were exclusively involved in baseline refractive eye development, 51 genes were linked to the regulation of susceptibility to

myopia, and 23 genes affected both baseline refractive development and susceptibility to myopia.

Gene-based genome-wide association analysis of the genes we found in mice against CREAM and UK Biobank human samples revealed that 647 genes whose expression correlated with baseline refractive errors in mice were also associated with refractive errors in humans, including 173 genes that withstood Bonferroni correction. Using gene-based analysis, we also found that 536 genes whose expression correlated with susceptibility to myopia were associated with refractive errors in humans, including 138 genes which exhibited genome-wide significance ($P_{Bonferroni} < 0.05$). One hundred and ninety-eight of these genes were involved in both baseline refractive development and regulation of susceptibility to myopia, including 34 genes which withstood Bonferroni correction. Although many of these genes were previously implicated in the development of human myopia, 572 genes (92 genes with $P_{Bonferroni} < 0.05$) whose expression correlated with baseline refractive development and 486 genes (79 genes with $P_{Bonferroni} < 0.05$) whose expression correlated with susceptibility to myopia were linked to human myopia for the first time, including 211 genes (54 genes with $P_{Bonferroni} < 0.05$) whose expression correlated with both baseline refractive development and susceptibility to myopia.

Genes that we found to be involved in refractive error development in mice and humans affect a multitude of biological functions in the retina; however, many biological functions underlying refractive development appear to be highly conserved across species. These include camera-type eye development and post-embryonic eye morphogenesis, ephrin receptor signaling, glucocorticoid receptor signaling, regulation of circadian rhythms and circadian regulation of gene expression, glutamate signaling, regulation of neurogenesis and dendrite morphogenesis, regulation of nitric oxide biosynthesis, regulation of long-term synaptic potentiation, synapse

assembly and chemical synapse transmission, calcium-dependent signaling, regulation of translation, small GTPase mediated signal transduction, photoreceptor function and development, cell-cell adhesion, regulation of beta-amyloid formation, regulation of mesenchymal cell proliferation, cellular response to hypoxia. We also found that many retinal signaling pathways involved in refractive development in mice are also subjected to genetic variation causing myopia in humans. These include EIF2 signaling, regulation of eIF4 and p70S6K signaling, mTOR signaling, integrin signaling, semaphorin signaling in neurons, protein kinase A signaling, clathrin-mediated endocytosis signaling, epithelial adherens junction signaling, synaptic long term potentiation, ephrin receptor signaling, nNOS and eNOS signaling, estrogen receptor signaling, PPARα/RXRα activation, signaling by Rho family GTPases, Wnt/Ca+ pathway, dopamine-DARPP32 feedback signaling, PTEN signaling, IGF-1 signaling, insulin receptor signaling, NF-κB signaling, RAR signaling, amyloid processing, among other pathways.

Interestingly, 27 genes that we found to be associated with refractive error development in mice were also among the genes differentially expressed in the retina of green monkeys with form-deprivation-induced myopia [39], and 233 genes were among the genes differentially expressed in the retina of marmosets exposed to positive and negative optical defocus [75]. There was also a 47-gene overlap with the genes found by Riddell et al. to be differentially expressed in the retina of chicks exposed to optical defocus [76], while 292 genes found in mice overlapped with genes found to be differentially expressed in the retina of chicks with lens-induced myopia by Stone et al. [77]. Many of the genes that we found to be involved in refractive eye development in this study had been previously implicated in various physiological and pathological processes in the retina. For example, mutations in *TTC21B* gene, which we

found to be involved in the regulation of susceptibility to myopia, were shown to cause a syndromic form of retinal dystrophy [78]. Mutations in *EFEMP1* (gene involved in baseline refractive development) were found to be associated with Malattia Leventinese retinal dystrophy characterized by the presence of RPE deposits [79]. Several studies linked a manganese transporter *SLC30A10*, which was found in this study to be involved in baseline refractive development, to parkinsonism, thus implicating this gene in the regulation of dopamine signaling [80, 81]. Ephrin receptor A10 (*EPHA10*), which we found to be involved in the regulation of susceptibility to myopia, was shown to influence cone photoreceptor morphogenesis, implicating signaling at the level of photoreceptors in refractive eye development [82]. A GTP-binding protein GNL2, involved in baseline refractive development, was demonstrated to play important role in retinal neurogenesis [83]. Finally, *AGRN* gene, found by this study to be involved in baseline refractive development, was shown to interact with EGR1 (previously implicated in refractive eye development) and regulate synaptic physiology in the retina [84, 85].

Although many pathways that we found to underlie refractive development in mice and humans in this study are novel, many of them are conserved across vertebrate species (Table 1). Seven out of 47 pathways found to be involved in optical defocus response in chickens by Riddell et al. [62] were also found to be involved in refractive eye development in this study, including PPAR signaling, tight junction signaling, Huntington's disease signaling, cysteine metabolism, actin cytoskeleton signaling, TGF-β signaling, and Glutathione metabolism (Additional file 5: Table S5, Additional file 8: Table S8, Additional file 27: Table S27, Additional file 28: Table S28). We replicated 12 out of 20 canonical pathways identified by Stone et al. [77] in chickens with lens-induced myopia, including glutamate receptor signaling, β-adrenergic signaling, synaptic long term depression, synaptic long term potentiation,

amyotrophic lateral sclerosis signaling, CREB signaling in neurons, relaxin signaling, purine metabolism, CDK5 signaling, protein kinase A signaling, hypoxia signaling, and tight junction signaling (Additional file 5: Table S5, Additional file 8: Table S8, Additional file 27: Table S27, Additional file 28: Table S28). We also found that 35 out of 75 pathways that we recently found to be involved in the development of hyperopia and myopia in marmosets [75] were also among the pathways which we found to be involved in refractive eye development in this study, including Ephrin Receptor Signaling, β-adrenergic Signaling, Protein Kinase A Signaling, Relaxin Signaling, Androgen Signaling, Dopamine-DARPP32 Feedback Signaling, nNOS Signaling, RAN Signaling, HIPPO signaling, Wnt/Ca+ pathway, PTEN Signaling, Synaptic Long Term Potentiation, Gap Junction Signaling, Regulation of eIF4 and p70S6K Signaling, mTOR Signaling, Glucocorticoid Receptor Signaling, α-Adrenergic Signaling, and Epithelial Adherens Junction Signaling, among others (Additional file 5: Table S5, Additional file 8: Table S8, Additional file 27: Table S27, Additional file 28: Table S28). An important role of amyloid signaling pathway, which we found to be involved in the regulation of susceptibility to myopia (Additional file 8: Table S8), is an agreement with our recent finding that a component of the amyloid signaling pathway APLP2 regulates susceptibility to myopia in mice and humans [37]. Our data also suggest that the phototransduction pathway is involved in refractive eye development (Additional file 5: Table S5), in agreement with a recent GWAS study [54] and recent marmoset data [75].

Conclusions

We have identified 2,302 genes which are involved in baseline refractive eye development and 1,917 genes which regulate susceptibility to myopia in mice. Our data suggest that at least 985 of

these genes are subjected to genetic variation in the human population and are involved in refractive error development in humans. Eight hundred forty-seven of these genes were implicated in the development of human myopia for the first time. A large number of common genes and canonical pathways that we found to be involved in the regulation of refractive eye development in mice, chickens and humans suggests strong evolutionary conservation of the signaling pathways underlying refractive error development. It appears that the number of genes involved in the regulation of refractive eye development may be as high as 3,505 in the retina alone, suggesting that refractive eye development maybe regulated by hundreds to thousands of genes across ocular tissues. Interestingly, in spite of significant overlap between genes that control baseline refractive development and genes regulating susceptibility to myopia, these two processes appear to be controlled by largely distinct sets of genes. The genes that we found to be involved in refractive eye development control, however, a well-defined set of retinal signaling pathways that we begin to consistently find to be involved in refractive eye development across different species. This provides a solid framework for future studies and for the development of anti-myopia drugs.

Acknowledgements

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Additional files

Additional file 1: Table S1. Baseline refractive errors in Collaborative Cross mice measured at P40 (diopters).

Additional file 2: Table S2. Form-deprivation myopia in Collaborative Cross mice (deprived eye versus control eye, diopters).

Additional file 3: Table S3. List of genes whose expression correlates with baseline refractive error in Collaborative Cross mice.

Additional file 4: Table S4. Gene ontology categories significantly associated with genes whose expression correlates with baseline refractive error in Collaborative Cross mice (BP, biological process; CC, cellular component; MF, molecular function).

Additional file 5: Table S5. Canonical signaling pathways affected by genes whose expression correlates with baseline refractive error in Collaborative Cross mice.

Additional file 6: Table S6. List of genes whose expression correlates with susceptibility to myopia in Collaborative Cross mice.

Additional file 7: Table S7. Gene ontology categories significantly associated with genes whose expression correlates with susceptibility to myopia in Collaborative Cross mice (BP, biological process; CC, cellular component; MF, molecular function).

Additional file 8: Table S8. Canonical signaling pathways affected by genes whose expression correlates with susceptibility to myopia in Collaborative Cross mice.

Additional file 9: Table S9. List of genes whose expression correlates with both baseline refractive error and susceptibility to myopia in Collaborative Cross mice.

Additional file 10: Table S10. Gene ontology categories significantly associated with genes whose expression correlates with both baseline refractive error and susceptibility to myopia in Collaborative Cross mice (BP, biological process; CC, cellular component; MF, molecular function).

Additional file 11: Table S11. Canonical signaling pathways affected by genes whose expression correlates with both baseline refractive error and susceptibility to myopia in Collaborative Cross mice.

Additional file 12: Table S12. List of human myopia QTLs and candidate genes located within 200 kb of the lead SNP (or within critical region).

Additional file 13: Table S13. List of genes localized within human myopia QTLs and whose expression correlates with baseline refractive error in mice.

Additional file 14: Table S14. List of genes localized within human myopia QTLs and whose expression correlates with susceptibility to myopia in mice.

Additional file 15: Table S15. List of genes localized within human myopia QTLs and whose expression correlates with both baseline refractive error and susceptibility to myopia in mice.

Additional file 16: Table S16. List of human myopia QTLs showing overlap with genes exhibiting correlation with either refractive error or susceptibility to myopia in mice.

Additional file 17: Table S17. Gene ontology categories significantly associated with genes localized within human myopia QTLs and whose expression correlates with baseline refractive error in mice (BP, biological process; CC, cellular component; MF, molecular function).

Additional file 18: Table S18. Gene ontology categories significantly associated with genes localized within human myopia QTLs and whose expression correlates with susceptibility to myopia in mice (BP, biological process; CC, cellular component; MF, molecular function).

Additional file 19: Table S19. List of mouse genes displaying significant association with refractive error in CREAM cohort and whose expression correlates with baseline refractive error in mice (Blue identifies $P_{Bonferroni} < 0.05$ in both cohorts).

Additional file 20: Table S20. List of mouse genes displaying significant association with refractive error in UK Biobank cohort and whose expression correlates with baseline refractive error in mice (Blue identifies $P_{Bonferroni} < 0.05$ in both cohorts).

Additional file 21: Table S21. List of mouse genes displaying significant association with refractive error in CREAM cohort and whose expression correlates with susceptibility to myopia in mice (Blue identifies $P_{Bonferroni} < 0.05$ in both cohorts).

Additional file 22: Table S22. List of mouse genes displaying significant association with refractive error in UK Biobank cohort and whose expression correlates with susceptibility to myopia in mice (Blue identifies $P_{Bonferroni} < 0.05$ in both cohorts).

Additional file 23: Table S23. List of mouse genes displaying significant association with refractive error in both UK Biobank and CREAM cohorts, and whose expression correlates with baseline refractive error in mice (Blue identifies $P_{Bonferroni} < 0.05$ in both cohorts).

Additional file 24: Table S24. List of mouse genes displaying significant association with refractive error in both UK Biobank and CREAM cohorts, and whose expression correlates with susceptibility to myopia in mice (Blue identifies $P_{Bonferroni} < 0.05$ in both cohorts).

Additional file 25: Table S25. Biological processes significantly associated with genes linked to refractive error in UK Biobank and CREAM cohorts, and whose expression correlates with baseline refractive error in mice (BP, biological process).

Additional file 26: Table S26. Biological processes significantly associated with genes linked to refractive error in UK Biobank and CREAM cohorts, and whose expression correlates with susceptibility to myopia in mice (BP, biological process).

Additional file 27: Table S27. Canonical signaling pathways significantly associated with genes linked to refractive error in UK Biobank and CREAM cohorts, and whose expression correlates with baseline refractive error in mice.

Additional file 28: Table S28. Canonical signaling pathways significantly associated with genes linked to refractive error in UK Biobank and CREAM cohorts, and whose expression correlates with susceptibility to myopia in mice.

Figure legends

Fig 1. Genetic background modulates refractive eye development and susceptibility to myopia in mice. **a** Refractive error is inherited as a quantitative trait in the founder strains of Collaborative Cross. Baseline refractive errors at P40 range from highly myopic to highly hyperopic depending on the genetic background in different strains. Horizontal red lines show mean refractive errors for each strain, while each dot corresponds to mean refractive errors of individual animals. **b** Susceptibility to form-deprivation myopia is inherited as a quantitative trait in the founder strains of Collaborative Cross. The extent of myopia induced by 21 days of visual form deprivation in different strains ranged from -5.5 ± 2.1 D in NZO/HlLtJ mice to -18.7 ± 3.1 D in CAST/EiJ mice. Horizontal red lines identify means of induced myopia for each strain, while each dot represents a mean interocular difference between deprived eye and control contralateral eye for individual animals.

Fig 2. Baseline refractive eye development in mice is regulated by a large number of genes via multiple retinal signaling pathways. **a** Expression of 2,302 retinal genes correlates with baseline refractive error in mice. Hierarchical clustering results show that genes, whose expression correlates with refractive error in the founder strains of Collaborative Cross, are organized in two clusters, i.e., one (top) cluster exhibiting increased expression in the highly hyperopic mice, and

the second (bottom) cluster showing increased expression in the myopic mice. **b** Top fifteen biological processes affected by the genes whose expression correlates with the baseline refractive errors in the founder strains of Collaborative Cross. Outer circle shows gene ontology IDs for the biological processes; second circle shows up- or down-regulated genes in the myopic mice versus hyperopic mice; inner circle shows activation or suppression of the corresponding biological processes, while the size of the sector corresponds to statistical significance (larger sectors correspond to smaller *P*-values). **c** Top twenty-five canonical pathways affected by the genes whose expression correlates with the baseline refractive errors in the founder strains of Collaborative Cross. Horizontal yellow line indicates $P = 0.05$. Z-score shows activation or suppression of the corresponding pathways.

Fig 3. Susceptibility to myopia in mice is regulated by a large number of genes via multiple retinal signaling pathways. **a** Expression of 1,917 retinal genes correlates with susceptibility to form deprivation myopia in mice. Hierarchical clustering results show that genes, whose expression correlates with susceptibility to myopia in the founder strains of Collaborative Cross, are organized in two clusters, i.e., one (top) cluster exhibiting increased expression in mice with high susceptibility to myopia, and the second (bottom) cluster showing increased expression in mice with low susceptibility to myopia. **b** Top fifteen biological processes affected by the genes whose expression correlates with susceptibility to myopia in the founder strains of Collaborative Cross. Outer circle shows gene ontology IDs for the biological processes; second circle shows up- or down-regulated genes in mice with high susceptibility to myopia versus mice with low susceptibility to myopia; inner circle shows activation or suppression of the corresponding biological processes, while the size of the sector corresponds to statistical significance (larger

sectors correspond to smaller *P*-values). **c** Top twenty-five canonical pathways affected by the genes whose expression correlates with susceptibility to myopia in the founder strains of Collaborative Cross. Horizontal yellow line indicates $P = 0.05$. Z-score shows activation or suppression of the corresponding pathways.

Fig 4. Baseline refractive eye development and susceptibility to myopia in mice are regulated via overlapping pathways. **a** Venn diagram showing substantial overlap between genes underlying baseline refractive eye development and genes regulating susceptibility to myopia in Collaborative Cross progenitor strain mice. **b** Top fifteen biological processes affected by the 714 genes associated with both baseline refractive eye development (top panel) and susceptibility to myopia in mice (bottom panel). Outer circle shows gene ontology IDs for the biological processes; second circle shows up- or down-regulated genes in the myopic mice versus hyperopic mice (top panel), or in mice with high susceptibility to form-deprivation myopia versus mice with low susceptibility to myopia (bottom panel); inner circle shows activation or suppression of the corresponding biological processes, while the size of the sector corresponds to statistical significance (larger sectors correspond to smaller *P*-values). **c** Top twenty-five canonical pathways affected by the 714 genes involved in the regulation of both baseline refractive eye development (top panel) and susceptibility to myopia (bottom panel) in the founder strains of Collaborative Cross. Horizontal yellow line indicates $P = 0.05$. Z-score shows activation or suppression of the corresponding pathways.

Fig 5. Summary of signaling pathways involved in regulation of baseline refractive eye development and susceptibility to myopia. Heatmap showing all statistically significant

canonical pathways affected by the genes involved in baseline refractive development and the genes influencing susceptibility to myopia. Z-score shows activation or suppression of the corresponding pathways. The dots show statistical significance ($P < 0.05$).

Fig 6. Genes localized within human myopia QTLs show functional overlap with genes underlying baseline refractive eye development and susceptibility to myopia in mice. 750 candidate genes localized within 279 human myopia QTLs exhibit statistically significant overlap with both genes involved in baseline refractive eye development and genes regulating susceptibility to myopia in Collaborative Cross progenitor strain mice.

Fig 7. Genes underlying baseline refractive eye development and susceptibility to myopia in mice are localized in human QTLs associated with myopia (part 1). Heatmap depicting genes and odds ratios for the overlaps between 109 human myopia QTLs and genes whose expression correlates with either baseline refractive errors, susceptibility to form deprivation myopia, or both in mice. Colors indicate odds ratios. Bold italic identifies genes found to be associated with refractive error in UK Biobank, CREAM, or both human samples by the gene-based genome-wide association analysis. Red identifies genes exhibiting correlation with both baseline refractive development and susceptibility to myopia in mice.

Fig 8. Genes underlying baseline refractive eye development and susceptibility to myopia in mice are localized in human QTLs associated with myopia (part 2). Heatmap depicting genes and odds ratios for the overlaps between 109 human myopia QTLs and genes whose expression correlates with either baseline refractive errors, susceptibility to form deprivation myopia, or

both in mice. Colors indicate odds ratios. Bold italic identifies genes found to be associated with refractive error in UK Biobank, CREAM, or both human samples by the gene-based genome-wide association analysis. Red identifies genes exhibiting correlation with both baseline refractive development and susceptibility to myopia in mice.

Fig 9. Top biological processes associated with genes linked to refractive eye development in mice and localized in known human myopia QTLs. **a** Chord diagram showing key genes (left semicircle) and top biological processes (right semicircle) associated with genes correlated with baseline refractive development in mice and localized in known human QTLs linked to myopia. **b** Chord diagram showing key genes (left semicircle) and top biological processes (right semicircle) for genes correlated with susceptibility to myopia in mice and localized in known human QTLs linked to myopia. Colored bars underneath gene names show up- or down-regulation of corresponding genes in either myopic mice versus hyperopic mice (A), or mice with high susceptibility to myopia versus mice with low susceptibility to myopia (B).

Fig 10. Summary of key biological processes and pathways affected by genes underlying refractive eye development and found to be linked to myopia in UK Biobank and CREAM human cohorts. **a** Hierarchical clustering diagram showing top 30 biological processes affected by genes underlying baseline refractive eye development in humans. Outer circle shows hierarchical clusters of biological processes (identified by different colors) linked to baseline refractive development; inner circle shows clusters of the corresponding genes up- or down-regulated in myopic mice versus hyperopic mice. **b** Top twenty-five canonical pathways affected

by the genes underlying baseline refractive eye development in humans. Horizontal yellow line indicates $P = 0.05$. Z-score shows activation or suppression of the corresponding pathways.

Fig 11. Summary of key biological processes and pathways affected by genes regulating susceptibility to myopia and found to be linked to myopia in UK Biobank and CREAM human cohorts. **a** Hierarchical clustering diagram showing top 30 biological processes affected by genes regulating susceptibility to myopia in humans. Outer circle shows hierarchical clusters of biological processes (identified by different colors) underlying regulation of susceptibility to myopia; inner circle shows clusters of the corresponding genes up- or down-regulated in mice with high susceptibility to myopia versus mice with low susceptibility to myopia. **b** Top twenty-five canonical pathways affected by the genes regulating susceptibility to myopia in humans. Horizontal yellow line indicates $P = 0.05$. Z-score shows activation or suppression of the corresponding pathways.

Table 1. Evolutionary conservation of retinal signaling pathways involved in refractive eye development.

Chicken	Mouse	Marmoset	Human	Canonical signaling pathways
	●		●	4-hydroxyproline degradation I
●	●		●	Actin cytoskeleton signaling
●	●	●		Aldosterone signaling in epithelial cells
	●		●	Amyloid processing
●	●	●		Amyotrophic lateral sclerosis signaling
●	●	●		Androgen receptor signaling
	●	●		Antiproliferative role of somatostatin receptor 2 signaling
●	●	●	●	Calcium signaling
	●	●		Choline biosynthesis III
	●		●	Chondroitin sulfate biosynthesis
●	●	●	●	Circadian rhythm signaling
	●	●	●	Clathrin-mediated endocytosis signaling
●	●	●		CREB signaling in neurons
●	●	●	●	CXCR4 signaling
	●		●	Diphthamide biosynthesis
	●	●		Dopamine receptor signaling
●	●	●	●	Dopamine-DARPP32 feedback signaling
	●	●	●	EIF2 signaling

	•	•	•	eNOS signaling
•	•	•		Ephrin B signaling
•	•	•	•	Ephrin receptor signaling
•	•	•	•	Epithelial adherens junction signaling
	•	•		ERK/MAPK signaling
•	•	•	•	Estrogen receptor signaling
•	•		•	GABA receptor signaling
•	•	•		Gap junction signaling
•	•	•	•	Glucocorticoid receptor signaling
•	•	•	•	Glutamate degradation/Glutamate receptor signaling
•	•		•	Glutathione biosynthesis
•	•		•	Glutathione redox reactions I
•	•		•	Glutathione-mediated detoxification
	•		•	Glycoaminoglycan-protein linkage region biosynthesis
	•	•	•	Gαq signaling
	•	•		HIPPO signaling
	•	•	•	HMGB1 signaling
•	•	•	•	Huntington's disease signaling
	•	•	•	IGF-1 signaling
•	•		•	ILK signaling
	•	•	•	Insulin receptor signaling
	•	•	•	Integrin signaling
•	•		•	L-cysteine degradation I
•	•	•	•	L-cysteine degradation III
	•	•		Mismatch repair in eukaryotes
	•	•	•	mTOR signaling
	•		•	NF-κB signaling
•	•	•	•	nNOS signaling
	•	•	•	NRF2-mediated oxidative stress response
	•	•	•	Oncostatin M signaling
•	•		•	Phospholipase C signaling
	•	•	•	Phototransduction pathway
•	•	•	•	PI3K/AKT signaling
•	•		•	PPAR signaling
•	•	•	•	PPARα/RXRα activation
	•	•	•	Production of nitric oxide and reactive oxygen species
•	•	•	•	Protein kinase A signaling
•	•	•	•	Protein ubiquitination pathway
	•	•	•	PTEN signaling
•	•			Purine nucleotides de novo biosynthesis II
•	•		•	Purine nucleotides degradation II (aerobic)
	•	•		RAN signaling
	•	•	•	RAR activation
	•	•	•	Regulation of eIF4 and p70S6K signaling
	•	•	•	Regulation of the epithelial-mesenchymal transition pathway
•	•	•		Relaxin signaling
•	•	•	•	RhoGDI signaling
	•		•	Semaphorin signaling in neurons
•	•	•	•	Signaling by Rho family GTPases
	•		•	Sumoylation pathway
•	•			Synaptic long-term depression
•	•	•	•	Synaptic long-term potentiation
•	•		•	TGF-β signaling
•	•			Tight junction signaling
•	•	•		tRNA splicing
	•	•	•	Wnt/Ca+ pathway
	•		•	Xenobiotic metabolism signaling
•	•	•		α-adrenergic signaling
•	•	•		β-adrenergic signaling

A Refractive error genes / Susceptibility to myopia genes
1588 | 714 | 1203

B Refractive error

z-score: decreasing — increasing

Fold change (myopia vs hyperopia)
- downregulated
- upregulated

Susceptibility to myopia

z-score: decreasing — increasing

Fold change (high myopia vs low myopia)
- downregulated
- upregulated

C
- positive z-score
- z-score = 0
- negative score
- no activity pattern available

Refractive error

Susceptibility to myopia

GO:0006412 translation
GO:0007156 homophilic cell adhesion via plasma membrane adhesion molecules
GO:0006596 polyamine biosynthetic process
GO:0031648 protein destabilization
GO:0040008 regulation of growth
GO:0046628 positive regulation of insulin receptor signaling pathway
GO:0098609 cell-cell adhesion
GO:0006810 transport
GO:0008654 phospholipid biosynthetic process
GO:0006635 fatty acid beta-oxidation
GO:0045104 intermediate filament cytoskeleton organization
GO:0006506 GPI anchor biosynthetic process
GO:0035774 positive regulation of insulin secretion in cellular response to glucose stimulus
GO:0070555 response to interleukin-1
GO:0009968 negative regulation of signal transduction

Activation z-score

-3.2   3.7

Left column:

Baseline refraction
Susceptibility to myopia

- GP6 signaling pathway
- Synaptic long-term potentiation
- Protein kinase A signaling
- CXCR4 signaling
- Superpathway of cholesterol biosynthesis
- CREB signaling in neurons
- EIF2 signaling
- Synaptic long-term depression
- Superpathway of inositol phosphate compounds
- Phospholipase C signaling
- β-adrenergic signaling
- NRF2-mediated oxidative stress response
- Signaling by Rho family GTPases
- D-myo-inositol-5-phosphate metabolism
- Melatonin signaling
- Aldosterone signaling in epithelial cells
- Ephrin B signaling
- p70S6K signaling
- ERK/MAPK signaling
- Huntington's disease signaling
- α-adrenergic signaling
- RhoGDI signaling
- IL-1 signaling
- Relaxin signaling
- Amyotrophic lateral sclerosis signaling
- Ephrin receptor signaling
- Sumoylation pathway
- Dopamine-DARPP32 feedback signaling
- 3-phosphoinositide degradation
- Wnt/Ca+ pathway
- HMGB1 signaling
- mTOR signaling
- Gαq signaling
- Amyloid processing
- D-myo-inositol (1,4,5,6)-tetrakisphosphate biosynthesis
- D-myo-inositol (3,4,5,6)-tetrakisphosphate biosynthesis
- Production of nitric oxide and reactive oxygen species
- eNOS signaling
- NF-κB signaling
- PTEN signaling
- Integrin signaling
- D-myo-inositol (1,4,5)-trisphosphate biosynthesis
- ILK signaling

Right column:

Baseline refraction
Susceptibility to myopia

- Insulin receptor signaling
- Regulation of eIF4 and p70S6K signaling
- Nitric oxide signaling
- Opioid signaling pathway
- PPARα/RXRα activation
- Androgen Signaling
- Cell Cycle: G2/M DNA damage checkpoint regulation
- Mitotic roles of polo-like kinase
- tRNA splicing
- AMPK signaling
- IGF-1 signaling
- HIPPO signaling
- Choline biosynthesis III
- Dopamine receptor signaling
- Leucine degradation I
- PI3K/AKT signaling
- Chondroitin sulfate biosynthesis
- Antiproliferative role of somatostatin receptor 2
- nNOS signaling in neurons
- Neurotrophin/TRK signaling
- 4-hydroxyproline degradation I
- Antiproliferative role of TOB in T cell signaling
- Arginine degradation I (arginase pathway)
- Assembly of RNA polymerase II complex
- Axonal guidance signaling
- Clathrin-mediated endocytosis signaling
- Diphthamide biosynthesis
- DNA methylation and transcriptional repression signaling
- Epithelial adherens junction signaling
- Estrogen receptor signaling
- G protein signaling mediated by Tubby
- Gap junction signaling
- Iron homeostasis signaling pathway
- L-cysteine degradation I
- Phosphatidylethanolamine biosynthesis II
- Phototransduction pathway
- Protein ubiquitination pathway
- Purine nucleotides de novo biosynthesis II
- RAN signaling
- RAR activation
- Semaphorin signaling in neurons
- Tight junction signaling
- Xenobiotic metabolism signaling

OR = 1.62, P = 0.022

OR = 2.75, P = 3.3 × 10^{-18}

OR = 2.03, P = 1.2 × 10^{-07}

CC refractive error

CC myopia

1498

691

1152

23

90

51

586

Human QTLs

Odds ratio	Genes correlated with refractive error	Genes correlated with induced myopia	Human myopia QTLs
	AGRN	EPHA10	chr1:973501
	GNL2	ZNF644	chr1:38092723
	ZNF644		chr1:91434335
	RHOC, PPM1J		chr1:113418415
	ZNF281		chr1:200311910
	CR1		chr1:207488004
	CD34		chr1:208062973
	SLC30A10		chr1:219790221
		SH3YL1	chr2:301051
	EFEMP1		chr2:56011517
		TTC21B	chr2:166627558
	PRSS56	PRSS56	chr2:233385396
	KLHDC8B		chr3:49211468
	ACTR8	ACTR8	chr3:53837971
	CEP63		chr3:134086356
	RASA2	ZBTB38	chr3:141093285
	TRMT44		chr4:8461896
	ANTXR2		chr4:80827062
	PRKG2	PRKG2	chr4:81903049
	RASGEF1B	RASGEF1B	chr4:82422327
	NR3C1		chr5:142720770
	DRD1		chr5:174720893
	RREB1, SSR1		chr6:7317483
	ZKSCAN8, ZKSCAN4, PGBD1, ZSCAN12		chr6:28162469
	LMBRD1		chr6:70446345
	PNRC1, GABRR2	SRSF12, PM20D2, GABRR2, UBE2J1, RRAGD	chr6:89918638
	NT5DC1, DSE		chr6:116446576
	ARHGAP18	ARHGAP18	chr6:129820038
	L3MBTL3		chr6:130354855
		SHPRH	chr6:146245589
	AKAP12		chr6:151620414
	CHN2, SCRN1, FKBP14, PLEKHA8, GARS, AQP1	FKBP14, ADCYAP1R1	chr7:27937503_32236643
	CACNA2D1	CACNA2D1	chr7:81400939
		PPP1R9A	chr7:94731338
	CAV1		chr7:116365413
	DPP6		chr7:154406581
	VIPR2		chr7:159093347
	ERI1		chr8:8961990
	MSRA		chr8:9807022
	PPP3CC		chr8:22538384
	EPHX2, CLU	EPHX2	chr8:27466315
		ZMAT4	chr8:40726394
	SFRP1		chr8:41103891
	ST18	ST18	chr8:53362145
	NCOA2	LACTB2	chr8:71420067
	MRPL13		chr8:121622778
	FAM83H	FAM83H	chr8:144811008
	C9ORF66 (DOCK8)	C9ORF66 (DOCK8)	chr9:214501
	FREM1	FREM1	chr9:14822828
	CNTLN		chr9:17639458
	FXN	TJP2, FAM189A2	chr9:71766593
	SEC61B	TGFBR1	chr9:101878622
		RGS3	chr9:116283516
	PROSER2	PROSER2	chr10:11889837
		TFAM	chr10:60304864

refractive error
myopia

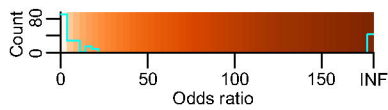| Odds ratio	Genes correlated with refractive error	Genes correlated with myopia	Human myopia QTLs
	ASCC1	DDIT4	chr10:74100279
		KCNMA1	chr10:79061458
	MYOF		chr10:94953258
		MRPL43, SFXN3	chr10:102824349
	MPPED2		chr11:30430331
	RCN1	PAX6	chr11:31612215
	LRRC4C		chr11:40149305
		SNX32, CFL1, FIBP, C11ORF68	chr11:65494260
		AASDHPPT	chr11:105753469
	DSCAML1		chr11:117657679
	KMT2A		chr11:118352372
	ING4, ENO2, C12ORF57, LPCAT3	PIANP	chr12:6954864
	GRIN2B		chr12:14062637
	SLCO1C1		chr12:20720707
		PYROXD1	chr12:21607360
	ETNK1	ETNK1	chr12:22592653
	ARID2	ARID2	chr12:46144855
	CD63, GDF11	ITGA7	chr12:56115778
	PPFIA2	PPFIA2	chr12:81827092
	RMST		chr12:97676235
		TMPO	chr12:98919382
	IGF1	IGF1	chr12:102793569
		AACS	chr12:125588898
		SPATA13	chr13:24432467
		RCBTB1	chr13:50159165
	GGACT		chr13:101175664
	EFNB2		chr13:107164733
		AKAP6	chr14:33303540
	EML5		chr14:89428948
		TRIP11	chr14:92582655
	C14ORF2, RD3L	TDRD9	chr14:104407243
	AQR		chr15:35005886
	JMJD7		chr15:42123687
	FBN1		chr15:48756536
	RPS27L, APH1B	LACTB, APH1B	chr15:63574641
		MEGF11, TIPIN	chr15:66590037
	ANKRD34C, TMED3	CTSH	chr15:79375347
	PLEKHG4		chr16:67318227
	TNFSF12, SAT2, EFNB3		chr17:7434819
	DNAH9	DNAH9	chr17:11407259
	HS3ST3B1		chr17:14138507
		SSH2	chr17:28104992
	TMEM98		chr17:31227593
	MED1	FBXL20	chr17:37576546
	ZNF652		chr17:47263475
	SUPT4A, SEPT4	SEPT4	chr17:56619441
	HGS, SLC25A10	NPLOC4, ARL16, P4HB	chr17:79612397
	MYL12B	MYL12B	chr18:3457606
		SETBP1	chr18:42824449
	BSG		chr19:577974
		RALY	chr20:32610401
	COL6A2		chr21:47377296
	DESI1, LINC00634, SEPT3	DESI1, XRCC6	chr22:42194561
	CACNA1F		chrX:49075678

Count
80
0

0   50   100   150   INF
Odds ratio

A

Fold change (myopia vs hyperopia)

downregulated upregulated

B

positive z-score   z-score = 0   negative score   no activity pattern available

-log(p-value)

EIF2 signaling
Sumoylation pathway
mTOR signaling
Integrin signaling
ILK signaling
Semaphorin signaling in neurons
Regulation of eIF4 and p70S6K signaling
Protein kinase A signaling
Clathrin-mediated endocytosis signaling
Actin nucleation by ARP-WASP complex
Epithelial adherens junction signaling
Glucocorticoid receptor signaling
Phospholipase C signaling
Calcium signaling
Protein ubiquitination pathway
Synaptic long term potentiation
Regulation of actin-based motility by Rho
RhoGDI signaling
Regulation of cellular mechanics by calpain protease
Remodeling of epithelial adherens junctions
Sphingosine-1-phosphate signaling
Ephrin receptor signaling
Adenosine nucleotides degradation II
nNOS signaling in neurons
Estrogen receptor signaling

calcium-dependent cell-cell adhesion via plasma membrane cell adhesion molecules

chemical synaptic transmission

post-embryonic eye morphogenesis

positive regulation of receptor activity

SRP-dependent cotranslational protein targeting to membrane

translational initiation

ATP metabolic process

cellular response to histamine

homophilic cell adhesion via plasma membrane adhesion molecules

cell-cell adhesion

dendrite morphogenesis

negative regulation of epithelial cell migration

positive regulation of dendrite development

positive regulation of beta-amyloid formation

neuron migration

synapse assembly

translation

rRNA processing

camera-type eye development

cell adhesion

nuclear-transcribed mRNA catabolic process, nonsense-mediated decay

cellular response to transforming growth factor beta stimulus

retinal cone cell development

nervous system development

negative regulation of cell migration

glutamate metabolic process

protein oligomerization

extracellular matrix organization

small GTPase mediated signal transduction

plasma membrane to endosome transport

A

Fold change (high myopia vs low myopia)

downregulated — upregulated

B

- positive z-score
- z-score = 0
- negative score
- no activity pattern available

-log(p-value)

EIF2 signaling
TGF-β signaling
mTOR signaling
PPARα/RXRα activation
Aryl hydrocarbon receptor signaling
Glutathione-mediated detoxification
Regulation of eIF4 and p70S6K signaling
PTEN signaling
Regulation of epithelial-mesenchymal transition pathway
Glutathione redox reactions I
STAT3 pathway
Glycoaminoglycan-protein linkage region biosynthesis
Human embryonic stem cell pluripotency
Epithelial adherens junction signaling
Neuregulin signaling
Xenobiotic metabolism signaling
GABA receptor signaling
Sumoylation pathway
VEGF signaling
IGF-1 signaling
Chondroitin sulfate biosynthesis
Mouse embryonic stem cell pluripotency
Regulation of cellular mechanics by calpain protease
NRF2-mediated oxidative stress response
PI3K/AKT signaling

- calcium-dependent cell-cell adhesion via plasma membrane cell adhesion molecules
- homophilic cell adhesion via plasma membrane adhesion molecules
- regulation of nucleic acid-templated transcription
- negative regulation of phosphorylation
- metabolic process
- positive regulation of cellular component movement
- protein transport
- receptor catabolic process
- small GTPase mediated signal transduction
- nitrobenzene metabolic process
- response to estradiol
- response to cholesterol
- glutathione derivative biosynthetic process
- developmental growth
- positive regulation of mesenchymal cell proliferation

- synapse assembly
- translation
- neuron fate commitment
- cellular response to hypoxia
- potassium ion transmembrane transport
- chondroitin sulfate biosynthetic process
- signal transduction by protein phosphorylation
- mitochondrial calcium ion transport
- chemical synaptic transmission
- potassium ion transport
- translational initiation
- nuclear-transcribed mRNA catabolic process, nonsense-mediated decay
- maturation of LSU-rRNA from tricistronic rRNA transcript
- ribosomal large subunit biogenesis
- GPI anchor biosynthetic process