

1 **Accelerating surveillance and research of antimicrobial resistance –**
2 **an online repository for sharing of antimicrobial susceptibility data**
3 **associated with whole genome sequences.**

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21

22 **Abstract**

23 Antimicrobial resistance (AMR) is an emerging threat to modern medicine. Improved diagnostics and
24 surveillance of resistant bacteria require the development of next generation analysis tools and
25 collaboration between international partners. Here, we present the “AMR data hub”, an online
26 infrastructure for storage and sharing of structured phenotypic AMR data linked to bacterial genome
27 sequences.

28 Leveraging infrastructure built by the European COMPARE Consortium and structured around the
29 European Nucleotide Archive (ENA), the AMR data hub already provides an extensive data collection
30 for some 500 isolates with linked genome and AMR data. Representing these data in standardized
31 formats, we provide tools for the validation and submission of new data and services supporting
32 search, browse and retrieval.

33 The current collection was created through a collaboration by several partners from the European
34 COMPARE Consortium, demonstrating the capacities and utility of the AMR data hub and its
35 associated tools. We anticipate growth of content and offer the hub as a basis for future research
36 into methods to explore and predict AMR.

37 Introduction

38 Antimicrobials are widely regarded as one of the major advances in modern medicine ¹. The global
39 emergence, however, of antimicrobial resistance (AMR) threatens the very core of modern medicine
40 with the potential to turn the global population back in time to the pre-antibiotic era in which simple
41 surgical procedures and common infections could have deadly consequences ².

42 The decreased costs of next generation sequencing (NGS) combined with the progress made in big
43 data analysis such as machine learning (ML) represent innovative opportunities to tackle the AMR
44 crisis³. Many bacterial phenotypic traits, including AMR, can be directly linked to the presence of
45 genomic determinants such as genes, Single Nucleotide Polymorphism's (SNPs) or transcription
46 promoters which can be identified using functional genomics approaches on large databases of
47 genomes. Recent studies have used computational approaches such as ML to predict antimicrobial
48 susceptibility from genomic data or to discover previously unidentified antibiotic resistance
49 determinants⁴⁻⁶. Today, the major limitation for such approaches is not the lack of advanced
50 computational methods or hardware resources but the lack of large enough well curated, annotated
51 data sets where phenotypic AMR data and genomes are linked.

52 Academic research initiatives and public health organisations could benefit from the implementation
53 of online repositories capable of storing large amounts of genome sequence and antimicrobial
54 susceptibility testing (AST) data. ⁷ For example, the PATRIC database (<https://patricbrc.org/> ⁶) has
55 been used for development of AMR prediction algorithms, but is a closed architecture using a project
56 specific data entry template. The NCBI is offering a similar service, linking antibiograms with genomes
57 deposited in the Sequence Repository Archive (SRA), but all data have to be made public immediately
58 (<https://www.ncbi.nlm.nih.gov/pathogens/isolates#/search/>).

59 Different stakeholders in the AMR field may have different requirements regarding the accessibility
60 of AST data. Making optimal use of the opportunities described above requires enabling the global
61 sharing of data, but some institutions are reluctant to immediately make their AST data publicly
62 accessible for privacy, legal or other reasons ⁸. National public health institutes would be encouraged
63 to create supra-national networks using a standard format to share data for AST result analysis, while
64 academics would find a solution to share post-publication data, encouraging reproducibility and
65 cross-validation experiments, if such a database structure would become available. Thus, we have
66 identified a clear need for a database structure that can support public AST data as well as those data
67 that are to be shared privately for a period of time until publication.

68 The Horizon 2020 funded EU consortium COMPARE aims at bringing NGS to public health and clinical
69 practice (<http://www.compare-europe.eu/>). European experts in AMR working within the COMPARE
70 consortium, including the European Bioinformatics Institute (EMBL-EBI), which is part of the
71 International Nucleotide Sequence Database Collaboration (INSDC; <http://www.insdc.org/>), have
72 deployed the "data hub" system to allow sharing of isolate NGS and linked phenotypic AMR data.
73 The data hub system allows data providers, such as public health and clinical laboratories, food
74 safety agencies and veterinary institutes, to share and download genomic and related data sets. Data
75 can either be kept private (pre-publication) or released as open-access at the discretion of the data
76 providers (Amid *et al.*, 2019, *in preparation*). Novel software was developed for use in the AMR data
77 hub that validates the conformity of submitted datasets. The system supports both qualitative and
78 quantitative AST data such as those resulting from disk diffusion and micro-broth dilution tests. The

79 AMR data hub “Notebook” reporting system has been configured to support rapid data mining of
80 content.

81 Here, we present the AMR data hub, which permits sharing of large amounts of information that
82 could be used for ML and other data analysis approaches, eventually resulting in accurate and
83 quantitative, hence clinically relevant, predictions of AMR phenotypes based on NGS data.

84

85 **The AMR data hub**

86 **The data hub system**

87 The data hub system has been built as a broad infrastructure for the sharing and analysis of pathogen
88 NGS data and related data types. Here is presented a brief outline of the system while a full
89 description is provided in Amid *et al.*, (2019, in preparation). The data hubs are provided upon the
90 foundation of the European Nucleotide Archive (ENA; <https://www.ebi.ac.uk/ena>), an open
91 repository for sequence and related data⁹. The concept was developed and introduced as a model
92 for rapid sharing of data and analysis outputs in public and pre-publication confidential status within
93 the COMPARE consortium. Data hubs are restricted (by login and password) to the members of the
94 project authorized to access the data. Pre-publication confidential data sharing between partners has
95 been considered only in projects where immediate release of data/metadata was not possible, i.e.
96 sensitive content has been awaiting publication, but partners needed access to confidential data for
97 analysis. Ultimately, all data archived in data hubs are released into the public domain (the standard
98 ENA database) after a period defined by data owners. Data and metadata reported by data providers
99 are submitted to the hubs through systematic processes supported by a number of tools.
100 Subsequently, structured and accessioned data/metadata are available for sharing between data
101 consumers who have received consent from data providers. Data appropriate for a given
102 computational analysis are selected and fed autonomously through cloud-based analysis workflows
103 of which the outputs – “derived” data products – are fed back into the system.

104

105 **The antibiogram**

106 In order to represent AST data, we have defined a new data type, the “antibiogram”, for use within
107 the AMR data hub and, more broadly, within ENA. This new data type leverages the extensible
108 “analysis object” system, with the addition of a new class specifically for the storage of phenotypic
109 AMR data, designated “AMR_ANTIBIOGRAM”. Antibiograms are treated as data objects within the
110 system and are supported in data submission and access services. As a new data type, building this
111 support has required the development of open software that is distributed publicly
112 (<https://github.com/EBI-COMMUNITY/compare-amr>) and used internally at EMBL-EBI for the
113 validation and submission of incoming AST data.

114 We have aimed with the antibiogram for a format that is flexible and complete. Minimum
115 requirements include, for each combination of isolate/ antibiotic provided, INSDC Sample accession
116 (SAM*); species; antibiotic name; antibiotic susceptibility testing standard; breakpoint version;
117 antibiotic susceptibility test method; measurement; measurement units; measurement sign;
118 susceptibility phenotype; and test platform. Any combination of bacterial species and antimicrobial is
119 supported. Reported antimicrobial susceptibility can be measured by microbroth dilution or zone

120 diameter, all major testing platforms (Sensititre, VITEK and Phoenix) and standards (EUCAST or CLSI)
121 are accepted. More uncommon methods can be added by using the free-text format of these
122 sections. To help data providers, a detailed protocol explaining the preparation of the metadata form
123 is presented on the GitHub repository of the project ([https://github.com/EBI-COMMUNITY/compare-](https://github.com/EBI-COMMUNITY/compare-amr)
124 [amr](https://github.com/EBI-COMMUNITY/compare-amr)) along with tools for batch creation of antibiograms from excel files. An interactive web page
125 allowing the manual creation of antibiograms is in preparation. It will provide an easy alternative for
126 data providers registering a limited number of samples. Finally, a tutorial explaining the steps
127 required to retrieve data (genomes and antibiograms) from the datahub is available on the GitHub
128 repository

129 Each antibiogram is linked to a bacterial genome within ENA. The association is asserted by linking
130 the analysis object, i.e. the antibiogram, with the corresponding study, example:
131 <https://www.ebi.ac.uk/ena/data/view/PRJEB14981>. As with other data deposited in the ENA,
132 antibiograms can be kept confidential for a provider-defined period but must ultimately be released
133 into public view. Antibigrams can be queried and retrieved through the AMR data hub (while
134 confidential) as well as (when made public) through other alternatives, such as the Pathogen data
135 portal (<https://www.ebi.ac.uk/ena/pathogens/home>¹), a Discovery Application Programming
136 Interface (API - <https://www.ebi.ac.uk/ena/portal/api/>), the ENA browser
137 (<https://www.ebi.ac.uk/ena>) and services providing high-volume data access such as the ENA File
138 Downloader (<https://github.com/enasequence/ena-ftp-downloader/>) for public data. Using the
139 Pathogen data portal or the API, various filters can be used to refine the query such as bacterial
140 species, country of origin, host, and more.

141

142 **Visualisation tools**

143 To visualize the contents of the AMR data hub, a Notebook was configured (Figure 1) that has several
144 options for comparison of a defined set of parameters from the database, such as the distribution of
145 the minimal inhibitory concentration (MIC) of different antimicrobials as a function of the country of
146 origin, or the comparison of MIC distributions between different antimicrobials. As such, this
147 functionality can be used for surveillance purposes, providing a rapid overview of MIC distribution for
148 a specific collection of isolates and how this compares to isolates from the same host or from
149 different geographical regions.

150 The Notebook is integrated into the Pathogen portal and can be accessed from
151 <https://www.ebi.ac.uk/ena/pathogens/home> under the “Explore” tab. Access to pre-publication data
152 in the AMR data hub requires authentication via login and password with authorization to the
153 corresponding project.

		resistance_phenotype											scientific_name
Count		measurement_units	measurement										
antibiotic_name		country	mg/L										Totals
antibiotic_name	country	0.015	0.25	2.0	8.0	0.03	0.5	1.0	4.0	0.12	0.06	Totals	
ciprofloxacin	Denmark	121	6			12					1	140	
	Italy	13										13	
	Netherlands		2					1	1			4	
	USA	49	11			3	2	2	6	2	1	76	
	United Kingdom	92	3		1	4						100	
	Viet Nam		36	51			16	8				111	
	nan		6		24							30	
Totals		275	64	51	25	19	18	11	7	2	2	474	

154

155 Figure 1: Visualization of the AST data deposited in the database on 15/10/2018. Data were filtered
 156 for: ciprofloxacin (antibiotic_name); *E. coli* (scientific_name) and mg/L (measurement_units).

157

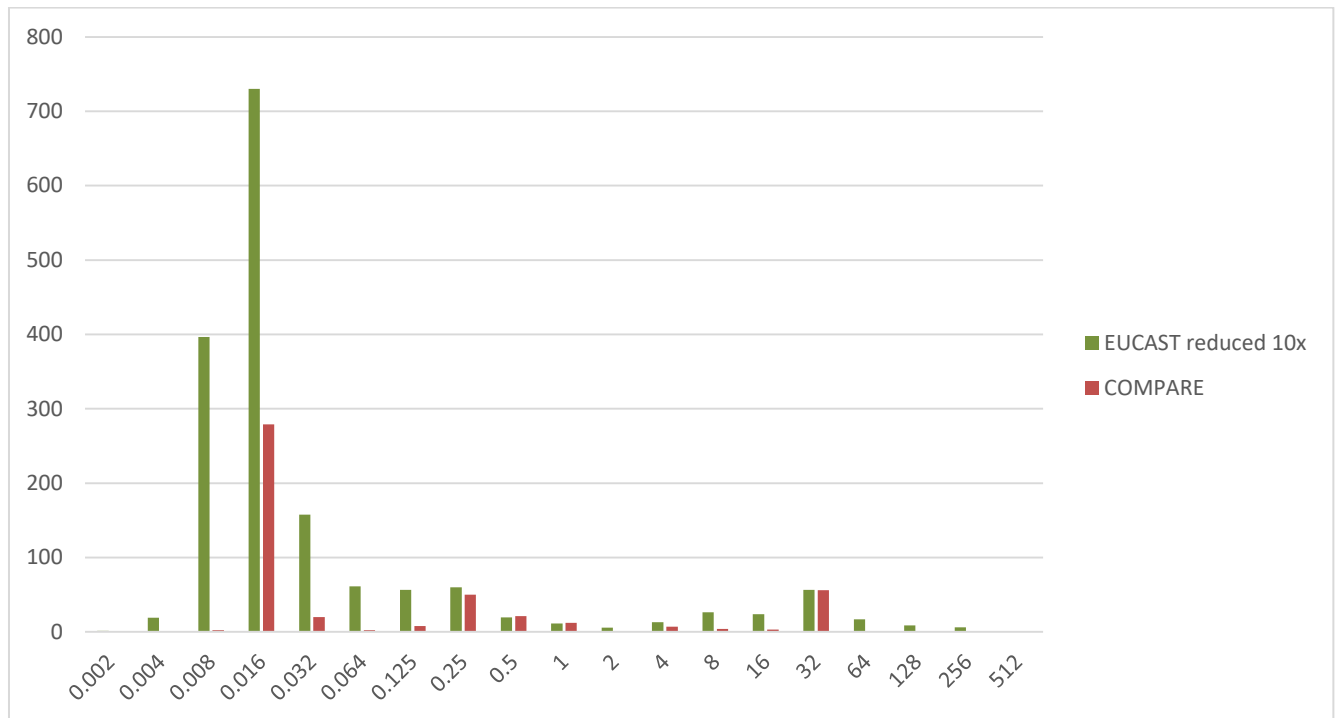
158 Current content

159 Statistics

160 As of 15-10-2018 the AMR data hub contains 577 *Escherichia coli* genomes with attached AST data,
 161 1,842 *Salmonella enterica* and 16 *Enterococcus faecium* originating from 9 different countries. Data
 162 on susceptibility against 55 different antibiotics (or combinations) have been entered in the database
 163 so far. As an example, 577 *E. coli* isolates originating from seven different countries (Bangladesh;
 164 Denmark; Italy; Netherlands; UK; USA and Vietnam) have been tested for ciprofloxacin susceptibility,
 165 474 by various dilution-based methods and 103 by disk diffusion.

166 A total of 470 *E. coli* antibiograms were submitted directly by COMPARE partners while 107 were
 167 imported from the US CDC database (n = 31) or previous publications (n = 76)^{10,11}.

168 Distribution of the MICs



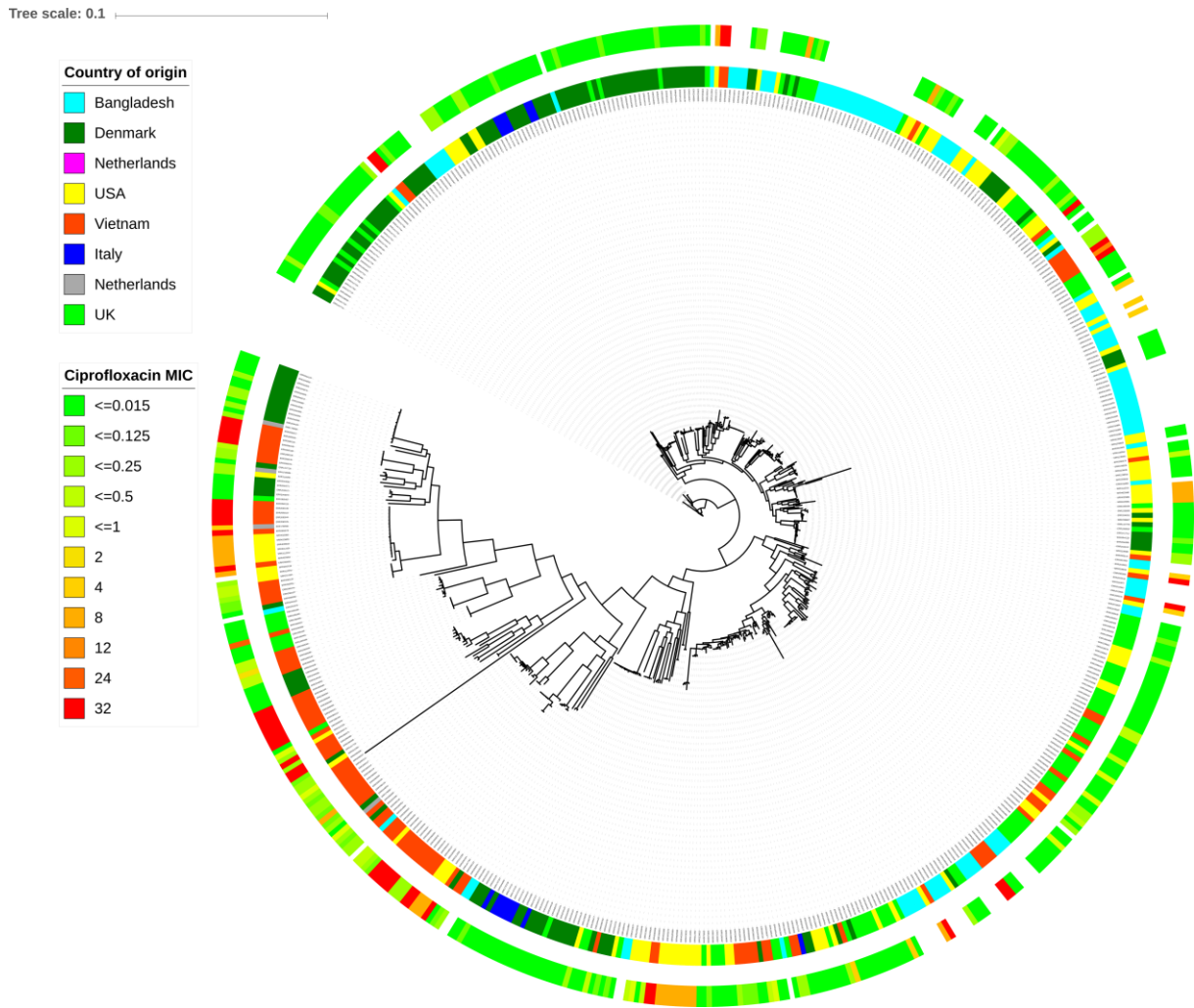
169

170 Figure 2: Distribution of the *E. coli* ciprofloxacin MICs and comparison with ECOFFs
171 (<https://mic.eucast.org/Eucast2/>).

172 As shown in Figure 2, the distribution of the *E. coli* ciprofloxacin MICs, as an example, recorded in the
173 present database follows a similar pattern as the ciprofloxacin MICs distribution reported by EUCAST
174 which is based on more than 20,000 isolates (<https://mic.eucast.org/Eucast2/>).

175 Phylogenetic analysis

176 As an example of the possibilities offered by the AMR data hub, the entire collection of *E. coli*
177 genomes was downloaded and a phylogenetic analysis of the population, in relation to ciprofloxacin
178 MICs, was performed (methods in supplementary materials). As shown in Figure 3, this population is
179 highly diverse and features a large range of ciprofloxacin MICs. Several isolates tend to cluster by
180 country, such as those from Denmark or Vietnam. Additionally, a strong association between
181 resistance and country can be observed. Most isolates from Vietnam show an MIC higher than 2
182 mg/ml. The study during which these isolates were collected focused on the presence of ESBL genes
183 in *E. coli*¹² and it is possible that ciprofloxacin resistance was co-selected for, as was previously
184 suggested¹³. Additionally, 24 out of 30 isolates retrieved from the CDC antimicrobial resistance
185 isolate bank (<https://www.cdc.gov/drugresistance/resistance-bank/currently-available.html>)
186 exhibited an MIC of 8 mg/ml. This unusually high proportion of resistant isolates can be explained by
187 the purpose of the CDC database, which is to provide a panel of well characterized resistant bacteria
188 for testing of diagnostic devices and new antibiotic agents. Conversely, isolates from Denmark were
189 collected as part of a routine surveillance effort from the veterinary institute and appear to all be
190 susceptible to ciprofloxacin.



191

192 Figure 3: Maximum likelihood tree based on the alignment of concatenated core genes of 572 *E. coli*
193 genomes (5 genomes failed quality control or assembly). Country of origin (inner circle) and
194 ciprofloxacin minimum inhibitory concentration (outer circle) are indicated for each isolate. Zone
195 diameters values for antibiotic susceptibility were not included in this analysis (isolates from
196 Bangladesh).

197

198 Discussion

199 We have built the AMR data hub with aim to provide a system for public health, food and veterinary
200 institutes, clinical laboratories and researchers to share their genomic and related AST data. It can be
201 used for standardized open-access data sharing, for example for published data, thus creating an
202 ever-growing source of AST metadata available to researchers worldwide. The large volume of data
203 made available will make it easier to use advanced statistical methods such as machine-learning to
204 predict AMR phenotypes from genomic data and discover new AMR determinants.

205 It has been recently underlined that application of the Nagoya Protocol, which regulates material and
206 data sharing, to genetic information might threaten the timely sharing of data in times of public
207 health emergencies¹⁴. In allowing the organisation and sharing of linked genomics and AST data, the
208 AMR data hub promotes openness and accessibility for these important data types while at the same

209 time meeting the privacy concerns for pre-publication data. Considering the exponential rise in the
210 number of bacterial genomes available, and the threat to modern medicine represented by the rise
211 of AMR, the establishment of the AMR data hub represents a timely effort to improve collaboration
212 in this field.

213 The design of a standard data submission format benefited from the expertise of the COMPARE
214 consortium, a group of international experts in bacterial genomics and AMR surveillance and
215 research. It is designed to be as exhaustive, and at the same time as flexible as possible to ensure
216 easy sharing of AST data. The database is hosted at the EMBL-EBI, ensuring its connection to the
217 world's largest online repository of bacterial genomes.

218 As members of the INSDC, EMBL-EBI and NCBI are part of a joined effort for standardization and
219 sharing of genomic data. The NCBI can also host antibiograms in a similar format to that presented
220 here, and efforts are currently ongoing to allow automated synchronization of content from both
221 sides. This will greatly increase the flexibility and the reach of the AMR data hub. While NCBI data
222 must be made public immediately upon deposition, EBI allows for pre-publication data to be kept
223 private for a provider-defined period. Participating institutions can thus choose whether they want
224 their data to be open-access immediately or whether they prefer sharing it with selected members of
225 a consortium before public release.

226 The view is that the AMR data hub will soon become an essential resource for functional genomic
227 studies of AMR. By encouraging data providers from different fields and geographical origin to share
228 their data, this collection can greatly improve our ability to answer questions related to the current
229 AMR crisis.

230

231

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264 **References**

- 265 1. Aminov, R. I. A Brief History of the Antibiotic Era: Lessons Learned and Challenges for the
266 Future. *Front. Microbiol.* **1**, 134 (2010).
- 267 2. WHO. *Antimicrobial resistance: global report on surveillance*. *Who* (2014).
- 268 3. van Dijk, E. L., Auger, H., Jaszczyszyn, Y. & Thermes, C. Ten years of next-generation
269 sequencing technology. *Trends Genet.* **30**, 418–426 (2014).
- 270 4. Kavas, E. S. *et al.* Machine learning and structural analysis of Mycobacterium tuberculosis
271 pan-genome identifies genetic signatures of antibiotic resistance. *Nat. Commun.* **9**, 4306
272 (2018).
- 273 5. Niehaus, K. E., Walker, T. M., Crook, D. W., Peto, T. E. A. & Clifton, D. A. Machine learning for
274 the prediction of antibacterial susceptibility in Mycobacterium tuberculosis. *2014 IEEE-EMBS*
275 *Int. Conf. Biomed. Heal. Informatics, BHI 2014* 618–621 (2014).
- 276 6. Davis, J. J. *et al.* Antimicrobial Resistance Prediction in PATRIC and RAST. *Nat. Publ. Gr.* 1–12
277 (2016).
- 278 7. Otto, M. Next-generation sequencing to monitor the spread of antimicrobial resistance.
279 *Genome Med.* **9**, 68 (2017).
- 280 8. van Panhuis, W. G. *et al.* A systematic review of barriers to data sharing in public health. *BMC*
281 *Public Health* **14**, 1144 (2014).
- 282 9. Harrison, P. W. *et al.* The European Nucleotide Archive in 2018. *Nucleic Acids Res.* **47**, D84–
283 D88 (2019).
- 284 10. Tyson, G. H. *et al.* WGS accurately predicts antimicrobial resistance in Escherichia coli. *J.*
285 *Antimicrob. Chemother.* **70**, 2763–2769 (2015).
- 286 11. Tyson, G. H. *et al.* Establishing genotypic cutoff values to measure antimicrobial resistance in
287 Salmonella. *Antimicrob. Agents Chemother.* **61**, (2017).
- 288 12. Nguyen, V. T. *et al.* Prevalence and risk factors for carriage of antimicrobial-resistant
289 Escherichia coli on household and small-scale chicken farms in the Mekong Delta of Vietnam.
290 *J. Antimicrob. Chemother.* **70**, 2144–2152 (2015).
- 291 13. Wiener, E. S., Heil, E. L., Hynicka, L. M. & Johnson, J. K. Are Fluoroquinolones Appropriate for
292 the Treatment of Extended-Spectrum β -Lactamase-Producing Gram-Negative Bacilli? *J.*
293 *Pharm. Technol.* **32**, 16–21 (2015).
- 294 14. dos S. Ribeiro, C., Koopmans, M. P. & Haringhuizen, G. B. Threats to timely sharing of
295 pathogen sequence data. *Science (80-)*. **362**, 404 LP-406 (2018).

296