Horizontal transmission of the entomopathogenic fungal isolate

INRS-242 of *Beauveria bassiana* in emerald ash borer,

***Agrilus planipennis* Fairmaire**

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Abstract

Emerald ash borer (EAB), *Agrilus planipennis* Fairmaire, is an invasive and destructive beetle that causes extensive damage to ash trees in North America. The entomopathogenic fungus *Beauveria bassiana* is considered as an effective biological control agent for EAB adult populations. Using an autodissemination device with a fungal isolate of *B. bassiana*, our research aims to investigate the possibility of horizontal transmission of the fungal disease from infected to uninfected EAB adults during mating. Results show that the efficiency of fungal transmission is significantly related to the sex of EAB carrying the fungal pathogen. EAB males are the promising vector to transmit mycosis to their partners during mating. Results strengthen the potential of the fungal autodissemination device as a powerful biological strategy to control EAB populations.

Keywords: Emerald ash borer, Horizontal transmission, Autodissemination device, *Beauveria bassiana*

Introduction

Emerald ash borer (EAB), *Agrilus planipennis* Fairmaire, is an extremely invasive insect that is responsible for killing hundreds of million ash trees, *Fraxinus* spp., in urban and forest ecosystems of North America. EAB was first detected in 2002 in Detroit, Michigan (USA) and in Windsor, Ontario (Canada) after its introduction from northeastern Asia (Haack et al. 2002). To date, EAB is found in 35 American states and five Canadian provinces (Emerald ash borer information network). After hatching, EAB larvae tunnel through the bark and begin feeding the phloem and the cambium (Cappaert et al. 2005). The abundance of larval galleries disrupts the flow of nutrients and water within the tree and may cause the tree death in 3-4 years (Siegert et al. 2009).

Various strategies have currently developed to suppress EAB populations. These approaches include the use of systemic insecticides (Herms et al. 2014), parasitoids (Duan et al. 2012) and fungal control agents (Lyons et al. 2012). However, management of EAB is still challenging because of the beetle's cryptic behaviors. A fungal autodissemination device developed by Dr. Guertin and Dr. Lavallée has shown its potential in controlling EAB (Lyons et al. 2012). Results shown that most EAB adults who contacted the autodissemination chamber died after five days. This study builds upon previous research and aims to test the hypothesis that infected adults released from the autodissemination device can horizontally transmit the fungal pathogen to uninfected partners during mating.

Materials and Methods

Emerald ash borer Collection

During summer of 2017, six 12-funnel green Lindgren traps (Synergy Semiochemicals, Burnaby, BC, Canada) were suspended in the upper third of the canopy of ash trees located in Laval, QC, Canada (Long. 45.541309; Lat. -73.718103). EAB adults captured in the collecting jars were harvested daily. In the laboratory, male and female EAB were separated using morphological characters (Rodriguez-Saon et al. 2007). Insects were then reared in cages and fed with fresh ash leaves until their use in experiments.

Fungal production

The isolate INRS-242 of *Beauveria bassiana* (also known as INRS-CFL), which is an effective entomopathogen of *A. planipennis* adults (Lyons et al. 2012), was used in this study. This isolate is an endogenous fungus recovered by Lavallée and Guertin from the pine shoot beetle, *Tomicus piniperda* L., near Cookeshire, QC, Canada. The fungal isolate is stored in 70% of glycerol at -80°C in the Fungal bank of the INRS-Armand-Frappier Santé and Biotechnologie, Laval, QC, Canada.

Autodissemination devices containing fungal coated-pouch were used in this experiment to inoculate EAB males and females. The production of the pouches was achieved as described by Srei (2017). Briefly, the INRS-242 isolate was first grown in Yeast Extract Peptone Dextrose broth (YPD, Alpha Bioscience Inc., Baltimore, MD, USA), in a rotary shaker (150 rpm) for four days at 25°C. The pouches (14.5 x 11 cm; fiberglass mosquito net) containing sterile pearled barley were then inoculated with the fungal conidial suspension before incubation for ten days in the growth chamber under controlled conditions (25°C, 70% of humidity and in the darkness).

Experimental design

A randomized block design with three replicates (blocks) was used to assess the transmission of the fungal disease from infected to uninfected EAB adults during mating behavior. Three treatments were randomly assigned to each block, with 20 EAB pairs per treatment. The first treatment represents the control pair, in which one uninfected male and one uninfected female were introduced in an arena (Petri Dishes 100 x 150 mm, Fisher Scientific, Ontario, Canada). For the second treatment, only the EAB male was inoculated with the fungus (IM+UF), whereas in the third treatment, just the EAB female was contaminated with the fungus (IF+UM). Three sterilized pouches without fungal isolate were used for the control treatments. For each infection treatment of each block, a fungal-coated pouch was employed once to inoculate EAB adults (Srei et al. 2017). Briefly, after walked on the fungal-coated pouch for 1 min, each EAB male or female was transferred into the arena using sterilized tweezers. For all treatments, each EAB female was glued to the Petri dish surface using Evo-Stik (Evode Industry Ltd., Newtown, Ireland). This approach avoids the possibility of indirect horizontal transfer resulting from insect movement inside the arena. After being placed in the Petri dishes, the insects were observed for 18 hours. Two parameters were measured during the assay: presence of mating and fungal transmission. The fungal transmission was recorded as occurring when conidia were present on the ventral side of uninfected males, or on the dorsal part of uninfected females. In addition to these parameters, the subsequent mortality of EAB was also reported. After 18 hours, EAB males and females from each arena were individually transferred to a new Petri dish in which a fresh ash leaf was provided as food substrate. All Petri dishes were then incubated in the growth chamber at 25°C with 50% of relative humidity and 16:8 h photoperiod (light:dark). Mycosis was confirmed by the appearance of white muscardine on EAB cadavers. Both mortality of EAB adults and the presence of the muscardine were recorded daily for 14 days.

 Statistical analysis

All statistical analyses were performed using the software R, version 3.4.3. The proportion

of mated pairs and horizontal transmission efficacy of INRS-242 isolate of *B. bassiana* among

EAB adults were assessed using an analysis of variance (ANOVA) to show significant

differences between treatments. Means associated with different treatments were then coated

using Duncan's multiple comparisons. Student's *t*-test was used to compare the fungal infection

percentage of mated pairs and unmated pairs. For all statistical tests, the level of rejection was

set at α = 0.05.

Results

A significant difference in the percentage of EAB adults that mated was observed between

treatments (ANOVA, df = 2; F value = 6.75; *p* = 0.0291) (Table 1). More precisely, no difference

was recorded between the control group and IM+UF treatment. However, the percentage of

insects that mated was significantly lower in IF+UM treatment. In both infection treatments

(IM+UF and IF+UM), mycosis transmission was of 100% for mated beetles. On the other hand,

no mycosis was observed in mated EAB pairs of control groups. As expected, the percentage

of infected EAB was significantly higher in mated pairs than unmated pairs, with 66.7% and

4.4% respectively *(t*-test = 3.6766; df = 16; *p* = 0.002). All infected EAB adults died within six

days following the fungal contamination, and white muscardine was recorded on all

contaminated individuals.

Discussion

Horizontal transmission by direct contact between fungal-infected and uninfected insects is

a mechanism of natural dispersion of conidia that regulates insect population size (Steinkraus

2006). This fungal transmission could occur from the time of initial contact during mating (Kreutz

et al. 2004), as has previously observed in insect species including the eastern larch beetle,

Dendroctonus simplex LeConte   (Srei   2017),   the   European   spruce   bark   beetle,

Ips typographus L. (Kreutz et al. 2004), the Mexican fruit fly, *Anastrepha ludens* Loew (Toledo

et al. 2007), and the stem borer, *Busseola fusca* Fuller (Maniania et al. 2011). Under laboratory

conditions, our results have demonstrated for the first time that an infected EAB adult can

transmit *B. bassiana* isolate INRS-242 to an uninfected mate by contact during mating.

Furthermore, our results shown that no fungal transmission was recorded on insects having no

contact, which provides evidence that the confinement did not mediate mycosis transfer. For all

EAB mated pairs, the average duration of mating was about 42 min (data not shown). This

observation corroborates to that of Pureswaran and Poland (2009). Following the mortality of

EAB adults and the appearance of muscardine, it is possible to confirm that the fungal dose

transmitted from infected to uninfected EAB adults during mating is strong enough to cause the

death of insects.

The efficiency of fungal transmission depends on the sex of the insect that carries the

pathogen. Transmission of entomopathogenic fungus *Metarhizium anisopliae* (Metschn.)

Sorokin within the populations of the Mediterranean fruit fly, *Ceratitis capitata* Wiedemann, was

higher when males were carriers of the pathogen (Quesada-Moraga et al. 2008). In *D. simplex*,

horizontal transmission of *B. bassiana* isolate INRS-242 was greatest when females were

inoculated with the fungus (Srei 2017). Our results show that male-to-female transmission of

the fungal isolate within EAB adults was higher than from female-to-male. One distinct

characteristic of EAB males is their pubescence. This may facilitate the adhesion of conidia and

could explain the difference observed in fungal transmission between infected males and

infected females. Moreover, a preliminary experiment suggests that some EAB males can mate

with at least two different females (data not shown). This observation suggests that increasing

the proportion of contaminated males can be a valuable strategy for increasing the mortality

rate of the adult EAB population. Adding another mortality factor to other biotic and abiotic

factors could contribute to slow or to keep the EAB population at a lower level and may

eventually reduce oviposition on ash trees.

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References

Cappaert, D., D. G. McCullough, T. M. Poland, and N. W. Siegert. 2005. Emerald ash borer in North America: a research and regulatory challenge. Am. Entomol. 51: 152-165.

Duan, J. J., L. S. Bauer, K. J. Abell, and R. van Driesche. 2012. Population responses of hymenopteran parasitoids to the emerald ash borer (Coleoptera: Buprestidae) in recently invaded areas in north central United States. BioControl 57: 199-209.

Emerald ash borer information network. http://www.emeraldashborer.info/. January 23rd, 2019.

Haack, R. A., E. Jendak, L. Houping, K. R. Marchant, T. R. Petrice, T. M. Poland, and H. Ye. 2002. The emerald ash borer: a new exotic pest in North America. Newsletter of the Michigan Entomological Society 47: 1-5.

Herms, D. A., D. G. McCullough, D. R. Smitley, C. S. Sadof, and W. Cranshaw. 2014. Insecticide options for protecting ash trees from emerald ash borer,   Second ed. USDA-NIFA North Central IPM Center.

Kreutz, J., G. Zimmermann, and O. Vaupel. 2004. Horizontal transmission of the entomopathogenic fungus *Beauveria bassiana* among the spruce bark beetle, *Ips typographus* (Col., Scolytidae) in the laboratory and under field conditions. Biocontrol Sci. Technol. 14: 837-848.

Lyons, D. B., R. Lavallée, G. Kyei-Poku, K. Van Frankenhuyzen, S. Johny, C. Guertin, J. A. Francese, G. C. Jones, and M. Blais. 2012. Towards the development of an autocontamination trap system to manage populations of emrald ash borer (Coleoptera: Buprestidae) with the native entomopathogenic fungus, *Beauveria bassiana*. J. Econ. Entomol. 105: 1929-1939.

Maniania, N. K., E. Ouna, P. Ahuya, B. Frérot, A.-E. Félix, B. L. Ru, and P. A. Calatayud. 2011. Dissemination of entomopathogenic fungi using *Busseola fusca* male as vector. Biol. Control 58: 374-378.

Pureswaran, D. S., and T. M. Poland. 2009. The role of olfactory cues in short-range mate finding by the emerald ash borer, *Agrilus planipennis* Fairmaire (Coleoptera: Buprestidae). J. Insect Behav. 22: 205-216.

Quesada-Moraga, E., I. Martin-Carballo, I. Garrido-Jurado, and C. Santiago-Álvarez. 2008. Horizontal transmission of *Metarhizium anisopliae* among laboratory populations of *Ceratitis capitata* (Wiedemann) (Diptera: Tephritidae). Biol. Control 47: 115-124.

Rodriguez-Saon, C. R., J. R. Miller, T. M. Poland, K. T. M., G. W. Otis, T. Turk, and D. L. Ward. 2007. Behaviors of adult *Agrilus planipennis* (Coleoptera: Buprestidae). The Great Lakes Entomologist 40: 1-16.

Siegert, N. W., D. G. McCullough., A. M. Liebhold, and F. W. Telewski. 2009. Reconstruction of the establishment and spread of the emerald ash borer: dendrochronological analysis, pp. 70. In K. McManus and K. Gottschalk (eds.), 19th Annual Proceedings of the USDA Interagency Research Forum on Invasive Species. General Technical Report NRS-P-36.

Srei, N. 2017. Impact de la transmission horizontale d'un champignon entomopathogène chez
le dendroctone du mélèze, *Dendroctonus simplex* LeConte (Coleoptera: Scolytinae).
Philosophiæ doctor Thesis, INRS-Institut Armand-Frappier.

Srei, N., R. Lavallée, and C. Guertin. 2017. Susceptibility of *Dendroctonus simplex* to
Hypocreales fungi: towards the development of a biological control strategy. J. Appl.
Entomol. 141: 487-495.

Steinkraus, D. C. 2006. Factors affecting transmission of fungal pathogens of aphids. J.
Invertebr. Pathol. 92: 125–131.

Toledo, J., S. E. Campos, S. Flores, P. Liedo, J. F. Barrera, A. Villaseñor, and P. Montoya.
2007. Horizontal transmission of *Beauveria bassiana* in *Anastrepha ludens* (Diptera:
Tephritidae) under laboratory and field cage conditions. J. Econ. Entomol. 100: 291-297.

Table 1. Percentage of mated pairs per treatment and the proportion of horizontal transmission
of *B. bassiana* isolate INRS-242 recorded in adults of *A. planipennis* mated pairs. Control:
male and female EAB were not exposed to the fungal isolate; IM+UF: fungal transmission
between infected male and uninfected female EAB; and IF+UM: fungal transmission between
infected female and uninfected male EAB. For each column, distinct letters indicate a
significant difference (Duncan, n = 60, $p < 0.05$).

Treatments	Number of mated pairs (mean ± SD)	Fungal transmission between mated pairs (mean ± SD)
Control	73.3 ± 5.8 [a]	0.0 ± 0.0 [a]
IM+UF	73.3 ± 2.9 [a]	100.0 ± 0.0 [b]
IF+UM	58.3 ± 7.6 [b]	100.0 ± 0.0 [b]

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