

1 **Evolutionary rates are correlated between cockroach symbiont**  
2 **and mitochondrial genomes**

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20 evolution, substitution rate, cockroach.

## 21 **Abstract**

22 Bacterial endosymbionts evolve under strong host-driven selection. Factors influencing host  
23 evolution might affect symbionts in similar ways, potentially leading to correlations between  
24 the molecular evolutionary rates of hosts and symbionts. Although there is evidence of rate  
25 correlations between mitochondrial and nuclear genes, similar investigations of hosts and  
26 symbionts are lacking. Here we demonstrate a correlation in molecular rates between the  
27 genomes of an endosymbiont (*Blattabacterium cunenoti*) and the mitochondrial genomes of  
28 their hosts (cockroaches). We used partial genome data for multiple strains of *B. cunenoti* to  
29 compare phylogenetic relationships and evolutionary rates for 55 cockroach/symbiont  
30 pairs. The phylogenies inferred for *B. cunenoti* and the mitochondrial genomes of their hosts  
31 were largely congruent, as expected from their identical maternal and cytoplasmic mode of  
32 inheritance. We found a strong correlation between evolutionary rates of the two genomes,  
33 based on comparisons of root-to-tip distances and on estimates of individual branch rates. Our  
34 results underscore the profound effects that long-term symbiosis can have on the biology of  
35 each symbiotic partner.

## 36 1. Introduction

37 Rates of molecular evolution are governed by a multitude of factors and vary significantly  
38 among species [1,2]. In the case of symbiotic organisms, such rates could be influenced not  
39 only by factors associated with their own biology, but also those of their symbiotic partner.  
40 This is particularly the case for strictly vertically transmitted, obligate intracellular symbionts  
41 (hereafter ‘symbionts’), which have a highly intimate relationship with their host [3]. For  
42 example, a small host effective population size will potentially lead to increased fixation of  
43 slightly deleterious mutations within both host and symbiont genomes, owing to the reduced  
44 efficacy of selection.

45 When the phylogenies of host and symbiont taxa are compared, simultaneous changes  
46 in evolutionary rate between host-symbiont pairs might be evident in their branch lengths.  
47 Some studies have found a correlation in evolutionary rates between nuclear and  
48 mitochondrial genes in sharks [4], herons [5], and turtles [6], suggesting that host biology  
49 affects substitution rates in nuclear and cytoplasmic genomes in similar ways. In insects,  
50 nuclear genes that interact directly with mitochondrial proteins and mitochondrial data have  
51 shown rate correlation [7].

52 There has not yet been any study of rate correlations between hosts and bacterial  
53 symbionts. Evidence for correlated levels of synonymous substitutions was found in a study  
54 of one nuclear gene and two mitochondrial genes from *Camponotus* ants and three genes from  
55 their *Blochmannia* symbionts [8]. However, the study did not determine whether this  
56 correlation was driven by rates of evolution, time since divergence, or both. Numbers of  
57 substitutions tend to be low for closely related pairs of hosts and their corresponding  
58 symbionts, and high for more divergent pairs, leading to a correlation with time that does not  
59 necessarily reflect correlation in evolutionary rates.

60 *Blattabacterium cuenoti* (hereafter *Blattabacterium*) is an intracellular bacterial  
61 symbiont that has been in an obligatory intracellular and mutualistic relationship with  
62 cockroaches for over 200 million years [9,10]. These bacteria are transovarially transmitted  
63 from the mother to the progeny. The genomes of 21 *Blattabacterium* strains sequenced to date  
64 are highly reduced compared with those of their free-living ancestors, ranging in size from  
65 590 to 645 kb [11,12]. They contain genes encoding enzymes for DNA replication and repair,  
66 with some exceptions (*holA*, *holB*, and *mutH*) [12–14]. The extent to which host nuclear  
67 proteins are involved in the cell biology of *Blattabacterium*, and particularly DNA  
68 replication, is not well understood.

69 We recently performed a study of cockroach evolution and biogeography using  
70 mitochondrial genomes [9]. During this process, we obtained partial genomic information for  
71 several *Blattabacterium* strains. These data provide the opportunity to test for correlation of  
72 molecular evolutionary rates between *Blattabacterium* and host-cockroach mitochondrial  
73 DNA.

74 Here we infer phylogenetic trees for 55 *Blattabacterium* strains on the basis of 104  
75 genes, and compare branch lengths and rates of evolution for host-symbiont pairs across the  
76 phylogeny. We find evidence of markedly increased rates of evolution in some  
77 *Blattabacterium* lineages, which are matched by increased rates of evolution in mitochondrial  
78 DNA of host lineages.

79

## 80 **2. Materials and methods**

81 A list of samples and collection data for each cockroach examined is provided in table S1  
82 (electronic supplementary material (ESM)). We used assembled data obtained from a  
83 previous study, in which we used cockroach mitochondrial genomes to build phylogenetic  
84 trees [9]. We searched each assembly against previously published *Blattabacterium* genomes  
85 using blastn [15] to identify *Blattabacterium* contigs. Each contig was annotated using Prokka  
86 v1.12 [16]. We determined orthology among 104 genes of 55 *Blattabacterium* strains and  
87 seven Flavobacteriales outgroups with OMA v1.0.6 [17]. Further details are available in the  
88 ESM.

89 The 104 orthologous *Blattabacterium* genes were aligned individually using MAFFT  
90 v7.300b [18] and concatenated. The mitochondrial genome dataset included all protein coding  
91 genes from each taxon plus 12S rRNA, 16S rRNA, and the 22 tRNA genes. Third codon sites  
92 were removed from each dataset on the basis of saturation tests using Xia's method  
93 implemented in DAMBE 6 [19, 20] (see ESM). Trees were inferred using maximum  
94 likelihood in RAxML v8.2 [21]. We examined congruence between host and symbiont  
95 topologies using the distance-based ParaFit [22]. Further details of phylogenetic analyses and  
96 congruence testing are provided in the ESM.

97 The evolutionary timescale of hosts and symbionts, as well as their evolutionary rates,  
98 were inferred using BEAST v 1.8.4 [23], using a fixed topology from the RAxML analysis  
99 (figure 1). We calibrated the molecular clock using minimum age constraints based on four  
100 fossils (table S2, ESM). A soft maximum bound of 311 Ma was set for the root node,

101 representing the oldest known cockroach-like fossil [24]. To allow evolutionary rates to be  
102 estimated in a single framework, a random subset of 12 *Blattabacterium* protein-coding genes  
103 (from the larger set of 104) plus the full mitochondrial data set were concatenated and  
104 partitioned into host and symbiont subsets. These analyses were carried out a total of three  
105 times, with a novel subset of 12 randomly selected *Blattabacterium* genes for each replicate.  
106 The inferred branch rates were then compared using Pearson correlation analysis using  
107 ggpubr [25] in R [26].

108       Root-to-tip distances from the RAxML analysis for each host and symbiont pair were  
109 calculated using the R packages ape [27], phylobase [28], and adephylo [29]. The use of root-  
110 to-tip distances removes the confounding effects of time, because all lineages leading to  
111 terminal taxa have experienced the same amount of time since evolving from their common  
112 ancestor. We used Seq-Gen v1.3.4 [30] to simulate the evolution of sequences from both host  
113 and symbiont along the *Blattabacterium* tree topology inferred using RAxML, with  
114 evolutionary parameters obtained from our separate RAxML analyses of the original data.  
115 Tree lengths for host and symbiont were rescaled according to their relative rates, but the  
116 relative branch rates were maintained between the two trees (see ESM).

117

### 118 **3. Results**

119 In all analyses, there was strong support for the monophyly of each cockroach family with the  
120 exception of Ectobiidae (figure 1). The topologies inferred from the host and symbiont data  
121 sets were significantly congruent ( $p=0.001$ ). Although there were some apparent  
122 disagreements between the two trees (for example, the position of Corydiidae), support at  
123 these nodes was generally low for both trees.

124       A molecular-clock analysis of the *Blattabacterium* data set indicated that the basal  
125 divergence occurred 314 Ma (95% credibility interval 219–420 Ma; figure S1, electronic  
126 supplementary material), giving rise to a one clade containing strains infecting Corydiidae,  
127 termites, Cryptocercidae, Blattidae, Anaplectidae, Tryonicidae, and Lamproblattidae, and a  
128 second clade containing strains infecting Ectobiidae and Blaberidae. We repeated the analysis  
129 using the mitochondrial data set and found that divergence times were markedly younger (184  
130 Ma, 95% CI 160–212 Ma; figure S2, electronic supplementary material). In an analysis of the  
131 combined data set, divergence times were approximately midway between those from the

132 separate analyses (216 Ma, 95% CI 159–278 Ma; figure S3, electronic supplementary  
133 material).

134 The inferred substitution rates along each pair of equivalent host and symbiont branches  
135 were found to be highly correlated ( $R=0.88$ ,  $p<2.2\times 10^{-16}$ ; figure 2a). Almost identical results  
136 were found in replicate analyses involving different sets of *Blattabacterium* genes (data not  
137 shown). The highest rates of evolution in the host and symbiont data sets (on the basis of  
138 branch lengths; figure 1) were in members of an ectobiid clade containing *Allacta*,  
139 *Amazonina*, *Balta*, *Chorisoserrata*, and *Euphyllodromia*, and a separate clade containing the  
140 two anaplectids *Anaplecta omei* and *Anaplecta calosoma*. After excluding these taxa,  $R$  was  
141 reduced to 0.47 but remained highly significant ( $p=1.5\times 10^{-6}$ ). As expected, analyses of  
142 simulated host and symbiont data yielded highly correlated estimates of branch rates (figure  
143 2b).

144 Regression analysis of root-to-tip distances for host-symbiont pairs also indicated that  
145 these two variables were correlated ( $R=0.7$ ; figure 2c). However, the sharing of branches  
146 between taxa in the estimation of root-to-tip distances renders the data in this plot  
147 phylogenetically non-independent and precludes statistical analysis.

148

#### 149 **4. Discussion**

150 Our study provides evidence for a correlation in molecular evolutionary rates between  
151 *Blattabacterium* and host mitochondrial genomes, based on two different approaches (branch-  
152 rate comparisons and analysis of root-to-tip distances). To our knowledge, this is the first  
153 demonstration of such a correlation in a host-symbiont relationship. Previous studies found a  
154 correlation in evolutionary rates between mitochondrial and nuclear genes [5–7], and this  
155 relationship appears especially pronounced for nuclear genes encoding proteins that are  
156 associated with mitochondria [7]. The pattern that we found was consistent across multiple  
157 subsets of *Blattabacterium* genes, indicating that it represents a genome-wide phenomenon  
158 for this symbiont.

159 Similar forces acting on the underlying mutation rates of both host and symbiont  
160 genomes could translate into a relationship between their rates of substitution. This could  
161 potentially occur if symbiont DNA replication depends on the host's DNA replication and  
162 repair machinery [31]. Because the genome of *Blattabacterium* is known to possess an almost  
163 complete suite of replication and repair enzymes [31], the scope for host enzymes to

164 significantly influence symbiont mutation rate appears to be limited. A better understanding  
165 of the level of integration of host-encoded proteins in the metabolism of *Blattabacterium* is  
166 required to explore this issue further.

167 Short host generation times could potentially lead to elevated evolutionary rates in host  
168 [32] and symbiont, assuming that increased rates of symbiont replication are associated with  
169 host reproduction, as is found in *Blochmannia* symbionts of ants [33]. Variations in metabolic  
170 rate and effective population size between host taxa could also explain the rate correlations  
171 that we observed here. Unfortunately, with the exception of a few pest and other species,  
172 generation time, metabolic rates, and effective population sizes are poorly understood in  
173 cockroaches. This precludes an examination of their influence on evolutionary rates in host  
174 and symbiont.

175 *Blattabacterium* is a vertically transmitted, obligate intracellular mutualistic symbiont,  
176 whose phylogeny is expected to mirror that of its hosts. This is especially the case for  
177 phylogenies inferred from mitochondrial DNA, since mitochondria are linked with  
178 *Blattabacterium* through vertical transfer to offspring through the egg cytoplasm. As has been  
179 found in previous studies [34–36], we observed a high level of agreement between the  
180 topologies inferred from cockroach mitochondrial genomes and from the 104-gene  
181 *Blattabacterium* data set. Owing to long periods of co-evolution and co-cladogenesis between  
182 cockroaches and *Blattabacterium* [9,34], potential movement of strains between hosts (for  
183 example, via parasitoids) is not expected to result in the establishment of new symbioses,  
184 especially between hosts that diverged millions of years ago.

185 In conclusion, our results highlight the profound effects that long-term symbiosis can  
186 have on the biology of each symbiotic partner. The rate of evolution is a fundamental  
187 characteristic of any species, and our study shows that it can become closely linked between  
188 organisms as a result of symbiosis. Further studies are required to determine whether the  
189 correlation that we have found here also applies to the nuclear genome of the host. Future  
190 investigations of generation time, metabolic rate, and effective population sizes in  
191 cockroaches and *Blattabacterium* will allow testing of their potential influence on  
192 evolutionary rates.

193

194 **Author contributions.** D.A.A., T.B. generated sequence data; Z.Q. collected and provided  
195 specimens; D.A.A., T.B, N.L., and S.Y.W.H. performed data analysis and interpreted results;  
196 N.L., D.A.A., T.B. and S.Y.W.H. designed the study; N.L. and D.A.A. wrote the manuscript,

197 with contributions from T.B. and S.Y.W.H. All authors approved the final version of the  
198 manuscript and agree to be accountable for all aspects of the work.

199

200 **Ethics.** This article does not present research with ethical considerations.

201 **Data accessibility.** Sequence data have been uploaded to GenBank (accession  
202 numbers XXXXXX–XXXXXX (TBA) and alignments have been uploaded to DRYAD: DOI:  
203 <https://doi.org/10.5061/dryad.ft6240m>.

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210 **Competing interests.** There are no competing interests.

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213

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302 between a wood-feeding cockroach and its bacterial endosymbiont, across a  
303 topographically complex landscape. *J. Biogeogr.* **44**, 1500–1511.
- 304

## 305 **Figure captions**

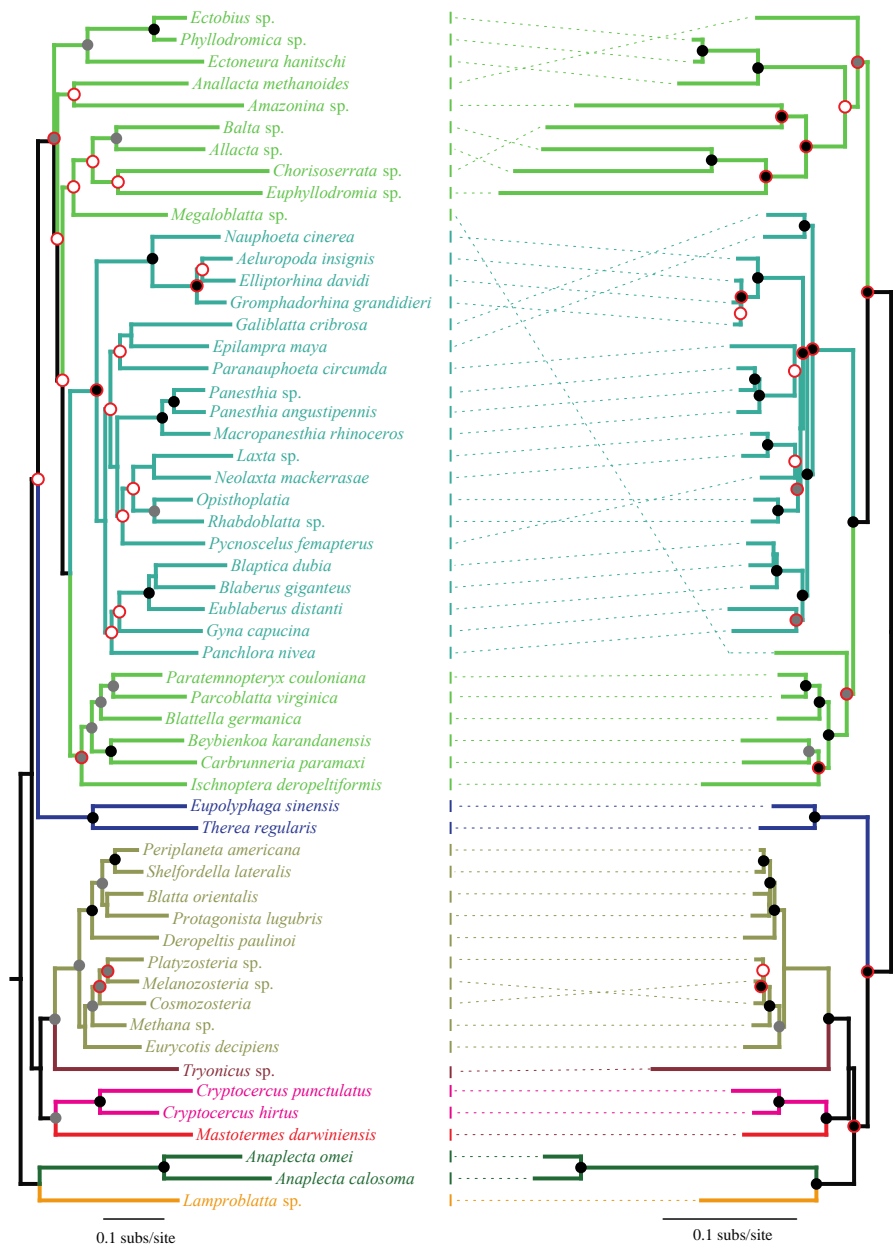
306 **Figure 1.** Congruence between (a) phylogenetic tree of host cockroaches inferred using  
307 maximum likelihood from whole mitochondrial genomes, and (b) phylogenetic tree of  
308 *Blattabacterium* inferred using maximum likelihood from 104 protein-coding genes (3rd  
309 codon sites excluded for both datasets). Circles at nodes indicate bootstrap values (black =  
310 100%, grey = 85–99%). Nodes without circles have bootstrap values <85%. Red outlines on  
311 circles indicate disagreement between the phylogenies, whereas red outlines on white circles  
312 indicate disagreement between the phylogenies and bootstrap values <85%. Colours represent  
313 taxa belonging to different cockroach families: light green = Ectobiidae, teal = Blaberidae,  
314 blue = Corydiidae, olive green = Blattidae, maroon = Tryonicidae, pink = Cryptocercidae, red  
315 = termites, dark green = Anaplectidae, and orange = Lamproblattidae.

316

317 **Figure 2.** Comparison of evolutionary rates of *Blattabacterium* symbionts and their host  
318 cockroaches. (a,b) Correlation between branch rates in the phylogenies of *Blattabacterium*  
319 and cockroaches, obtained from a Bayesian time-calibrated tree inferred from (a) 12  
320 *Blattabacterium* protein-coding genes and whole mitochondrial genomes from cockroaches,  
321 with 3rd codon sites excluded; (b) synthetic sequence data (see ESM). (c,d) Correlation of  
322 root-to-tip distances in phylogenies of *Blattabacterium* and cockroaches, inferred using  
323 maximum-likelihood analysis of (c) 104 *Blattabacterium* protein-coding genes and whole  
324 mitochondrial genomes from cockroaches, with 3rd codon sites excluded; (d) synthetic  
325 sequence data (see ESM). Colours represent data from representatives of different cockroach  
326 families, as described in figure 1. Grey circles represent internal branches.

(a) Cockroach mtDNA

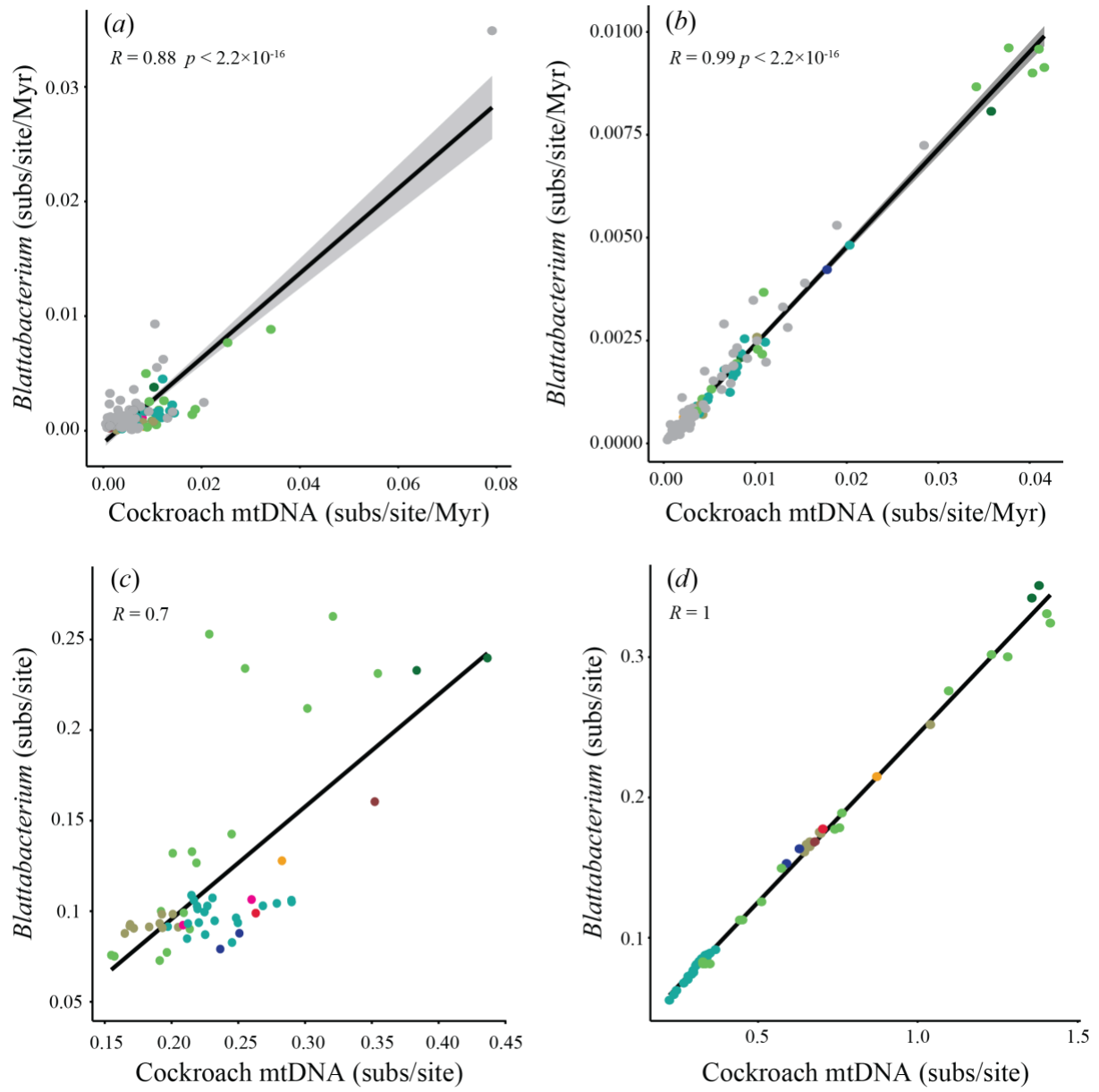
(b) *Blattabacterium*



327

328 Figure 1

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330

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332

333

334 Figure 2