

1 **TITLE: Model systems for large scale ecological research: *parva sub ingenti***

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3 **SHORT TITLE: Model systems for landscapes**

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16 **Abstract**

17 Large-scale ecological research (i.e., macroecology, biogeography, and landscape ecology) is
18 limited by the inability to have robust experimental replication due to scale and spatial changes
19 in ecological patterns. Model systems may offer one solution to this challenge. We propose that
20 considering smaller patterns in the context of larger ones (here, patches of lichen thalli on the
21 trunks of trees) as model systems for large-scale research, can provide sufficient replication.
22 Appropriate model systems will facilitate experimentation to elucidate links between spatial
23 ecological patterns and processes. To function as replicate landscape units, patterns of patches
24 should not differ significantly between trees across a sampling area of interest. We compared a
25 previously-demonstrated model system of patches of lichen on balsam fir (*Abies balsamea*) tree
26 trunks within a single small lichen-rich forest stand on the Avalon Peninsula in Newfoundland,
27 Canada to a set of more widely dispersed trees of two species (balsam fir and yellow birch,
28 *Betula alleghaniensis*) to assess if this model system could be useful across broader spatial
29 extents. We found that lichen composition generally followed consistent patterns between north
30 and south sides of the tree, as well as along an elevational gradient up the trunk at both the more
31 constrained, and at the more extensive, sampling extents. However, the reliability of the trees as
32 model landscapes varied by tree species and with the suite of lichens included. Considering
33 lichens on trees as a model landscape system can allow sufficient replication in experimentation
34 to address questions about spatial ecological patterns and thus provide a useful model system for
35 research in landscape ecology and biogeography.

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39 1. Introduction

40 When asked to describe what landscape ecologists study, the reply is often “what you can see
41 from an airplane window”, namely, landforms, land cover, and land use [1]. Understanding links
42 between spatial patterns in these three elements of the landscape, and the ecological processes
43 therein, has been the focus of landscape ecology. Scientific research in landscape ecology, in
44 turn, has informed real-world management problems in forestry (e.g., [2]), wildlife management
45 (e.g., [3]), and land-use planning (e.g., [4]). However, after decades of research, landscape
46 ecologists lack comprehensive theories to explain the patterns observed, and the variation
47 between regions. The spatial extents at which landscape ecology research is often conducted
48 make it difficult (if not impossible) to carry out manipulative experiments [5, 6] and to
49 sufficiently replicate experimental units to enable robust hypothesis testing [5, 7]. This, in turn,
50 makes it difficult to elucidate mechanisms that link patterns and processes. Hargrove and
51 Pickering [7] referred to pseudoreplication as the *sine qua non* for regional ecology, meaning
52 that without allowing for pseudoreplication in studies, hypothesis testing (and hence, reliable
53 information to inform management-decisions) within large-scale ecology research is not
54 possible. However, pseudoreplication is an anathema to many researchers [8, 9], and makes it
55 difficult to carry out hypothesis testing and draw meaningful inference from statistical analysis.
56 Thus, the suggestion that pseudoreplication may be unavoidable can be a difficult one to accept.

57 Here, by experimental replicate units we refer to repeated sampling units, not full-scale
58 replication of experiments [10], which has also been termed reproducibility [11]. We propose
59 that considering smaller patterns in the context of larger patterns (*parva sub ingenti* – the small
60 under the huge), that is treating microlandscapes as model systems, can provide a solution to the
61 problem of adequate replication in large-scale ecological research. Model systems (e.g.,

62 *Drosophila melanogaster* or *Escherischi coli*) are common in biomedical and molecular biology
63 research as their faster processes and small size make them amendable for manipulative
64 experiments, and their biological traits make them good analogs for other, larger and less easy-
65 to-manipulate systems. In ecology, Vitousek [12] suggested the use of natural model systems,
66 such as islands or lakes, to test ecological hypotheses that might apply to other, less well-
67 delineated or more-difficult-to-manipulate systems.

68 Microlandscapes have been proposed (but not yet widely adopted) as model systems for
69 landscape ecologists to examine spatial patterns and processes with replicate units [5, 13, 14].
70 Wiens and Milne [5] proposed constructing experimental microlandscapes to assess how
71 variation in patch configuration affected beetle movement. Bowker et al. [13] suggested using
72 biocrusts (soil crusts comprised of fungi, algae, bryophytes, lichens and cyanobacteria) as a
73 model system for research in community, ecosystem, and landscape ecology. Wiersma and
74 McMullin [14] demonstrated that arboreal lichens growing on the trunk of trees could be
75 considered as analogues to patches of landcover on a landscape. Both of the systems proposed
76 previously [13, 14] differ from the system described by Wiens and Milne [5] in that they are
77 naturally occurring as opposed to artificially constructed. The advantage of Wiersma and
78 McMullin's [14] model system over Bowker et al.'s [13] is that the boundaries of the proposed
79 microlandscapes (tree trunks) can be very clearly defined and delineated *in situ*. In fact, others
80 [15] have posited that trees could be considered as "islands". A disadvantage is that it is
81 infeasible to transport tree trunks to the lab for controlled experiments as can be done with soil
82 crusts [13]. Nonetheless, having microlandscapes that can be continually monitored within a
83 natural setting has the potential to advance the science of landscape ecology. Below, we discuss
84 the standards for replicate experimental units and replicate landscapes in more detail, before

85 introducing the sampling and testing carried out to evaluate whether the model system described
86 for a smaller area [14] has traction across a broader region.

87

88 **1.1 Standards for replicate experimental units and replicate landscapes**

89 To have true replication in any study, replicates should be independent but as closely similar as
90 possible except for the factor of interest (the treatment). In laboratory experiments, this is
91 achieved through replicate model organisms which are bred explicitly to be virtual clones of each
92 other. Outside the controlled lab environment, observational experiments are designed to collect
93 data from individual organisms or sites which share certain traits (e.g., age, sex, soil pH, sun
94 exposure) but differ in a single trait of interest (e.g., diet, disturbance history). Much of the lack
95 of perfect replication can be dealt with in various ways at the analysis stage. Statistical tests can
96 treat individuals as random effects to separate variation due to individual replicates from
97 variation due to the experimental treatment (but see critiques of this approach in [16]). Although
98 proper replication is emphasized in classical statistical tests, others [17] have suggested
99 alternatives for experiments in large-scale systems, including replicated controls, or to conduct
100 unreplicated experiments and acknowledge and accommodate the lack of replication through
101 Bayesian statistics or through analysis of spatial and temporal variation. However, both Hurlbert
102 [8] and Oksanen [17] acknowledge that replication is still necessary for inductive approaches to
103 experimentation.

104 To determine how landscapes might be replicated, and to properly evaluate the proposed
105 model system of lichen-covered tree trunks, it is important to understand how the term
106 “landscape” is defined and understood. In their foundational paper, Forman and Godron [18]
107 describe landscapes as being comprised of interacting stands or patches, which are repeated

108 across kilometers-wide extents in similar form. The “patch” has been described as the
109 fundamental unit of landscapes and is defined as a relatively homogenous area that differs from
110 its surroundings [1, 18]. The spatial pattern of patches can be quantified using a wide range of
111 landscape metrics [19] and is assumed to be driven by abiotic and biotic factors and processes
112 operating at multiple spatial and temporal scales [18]. For example, climate dictates the range of
113 plant species that can occur, while topography might influence the plant cover within a specific
114 patch as a function of microclimate.

115 To meet the standards of replicate units described above, replicate landscapes have to
116 have similar spatial configuration of patches (i.e., landscape pattern) and thus similar landscape
117 structure, but yet be spatially independent to avoid problems of spatial autocorrelation and
118 pseudoreplication. In natural systems, we can expect that no two landscapes will be identical as
119 might be the case in artificial ecosystems (e.g., [20]), but we do know that there are predictable
120 patterns. For example, von Humboldt’s early biogeographical thinking (which influenced the
121 development of the field of landscape ecology in Europe) on the geography of plants described
122 predictable patterns of vegetation as a function of climate that were similar across continents
123 [21]. Similarly, Merriam’s “life zones” concept describes patterns of similar plant communities
124 as a function of elevation in mountainous regions [22]. While the exact patterns of vegetation
125 patches along an elevational gradient will differ between mountain ranges in different continents,
126 patterns will be similar along two slopes within a single mountain chain. Similarly, there are
127 predictable differences in vegetation patterns on north-facing vs. south-facing slopes, which are
128 generally consistent across different hillsides. A landscape study along different slopes within a
129 single mountain chain might appear to constitute robust replicates for experimentation, but the
130 spatial extent of these landscapes provides logistical challenges while also introducing

131 confounding factors that might be correlated with altitude or location [17]. Thus, we propose
132 replicating within model systems of tree trunks to more efficiently increase sample size, while
133 making studies more amenable to manipulation.

134 Wiersma and McMullin [14] showed that tree trunks growing in a single, small
135 homogeneous stand (< 1 ha is area) had similar “patch” patterns of lichen distribution along both
136 the gradient of the trunk and between the north and south sides of the trunk. Given the
137 hierarchical structure of ecological systems, we posit that microlandscapes sampled across a
138 wider spatial extent than a single stand will still adequately function as replicate experimental
139 units. Here, we test whether the micolandscape patterns documented earlier [14] within a single
140 stand hold within the wider ecoregion in which their original study was located.

141

142 **2. Methods**

143 *Study area* - The stand studied by Wiersma and McMullin [14] is in the Avalon Forest Ecoregion
144 on the island of Newfoundland, Canada. The Avalon Forest is the smallest (500 km²) ecoregion
145 in the province of Newfoundland and Labrador, characterized by high humidity and
146 precipitation, cool summers, and mild winters [23]. The forested areas are on rolling hills called
147 ribbed moraines, landscape features created by glaciers [24]. Interspersed between the moraines
148 are open, sphagnum-dominated wetlands [23]. Forests are dominated by balsam fir (*Abies*
149 *balsamea*) with black spruce (*Picea mariana*) in wet areas and occasional yellow birch (*Betula*
150 *alleghaniensis*) stands on north facing slopes [23]. In this study, we test whether the consistent
151 microlandscape patterns previously observed on 24 balsam fir trees in a single stand [14] are
152 consistent across a broader region on the same species, and whether such patterns are also found
153 on a less-common deciduous tree, yellow birch.

154

155 **2.1. Experimental Design**

156 We expanded our study area from the previous study [14]. In this new study, our spatial extent
157 encompassed the entirety of the Avalon Forest Ecoregion (Fig. 1). Sites were all similar to those
158 in Wiersma and McMullin [14] in that they were balsam-fir dominated stands, mostly occurring
159 on moraines. We visited 21 sites across the region (Fig. 1) and at each site selected two trees, one
160 balsam fir and one yellow birch that were similar in diameter and within 25 m of each other. On
161 each tree we sampled the north and south sides of the trunk using a 10 cm x 50 cm “lichen
162 ladder”, divided into five 10 x 10 cm “blocks”, placed from 1.1 m to 1.6 m up the trunk (Fig. 2).
163 This is similar to the 10 cm by 1 m “transect” that Wiersma and McMullin [14] placed on each
164 side of the trunk (theirs was positioned from 0.9-1.9 m along the bole). For the purposes of
165 comparing the pattern within the single stand and across the wider ecoregion, we only used data
166 from the 10 cm blocks in Wiersma and McMullin’s [14] data that matched those of the expanded
167 survey (i.e., we used the data from Wiersma and McMullin [14] only from the 10 cm blocks
168 between 1.1 m and 1.6 m up the tree trunk). In the present study, we identified and counted all
169 lichen species within each 10 cm block. Species that could not be identified in the field were
170 collected for identification using standard processes, including microscopy, chemical spot tests
171 [25], and thin-layer chromatography [26]. In addition, we inventoried both macro- (those with a
172 more three-dimensional growth form, and growing on the substrate, which includes foliose and
173 fruticose growth forms) and micro-lichens (those growing within the substrate, i.e., crustose
174 growth forms); this is in contrast to Wiersma and McMullin [14], who limited their study to
175 field-identifiable macro-lichens. Thus, we analyzed the new data with macro-lichens only to
176 compare to the previous study; we also repeated the analysis for more dispersed trees using data

177 on both macro- and micro-lichens to assess whether the landscape patterns observed by Wiersma
178 and McMullin [14] held when examining a broader suite of lichen species and when looking
179 across a larger sampling area.

180

181 **2.2. Statistical analysis**

182 We used a perMANOVA analysis [27] to assess whether the pattern of lichen patches along the
183 trunk was consistent across all trees when stratifying for aspect, and whether the patterns
184 between the north and south sides were consistent when stratifying by height up the trunk.
185 Wiersma and McMullin [14] found significant patterns in both cases along a 1 m transect along
186 the trunk across 24 trees. We re-tested their data using a 50 cm transect that covered the same
187 distance up the trunk as in this expanded study. We separately analysed the data from the 21
188 balsam fir and the 21 yellow birch in the expanded study area and did not combine the data from
189 the 21 balsam fir in the wider ecoregion with the data from the 24 balsam fir in the single stand
190 because of differences in geographic sampling intensity. We carried out all statistical analysis
191 using R (version 1.0.136 [28]) with the package vegan [29].

192

193 **3. Results**

194 The 24 balsam fir trees in the single stand (that were previously analyzed for lichen diversity
195 along a 1 m “microtransect” along the north and south sides) showed a significantly consistent
196 lichen patch pattern between the north and south sides when we re-analyzed only a 50 cm portion
197 of the trunk, when stratifying by tree (perMANOVA $R^2 = 0.00552$, $p = 0.05$) but not when we
198 controlled for position along the trunk (perMANOVA $R^2 = 0.00552$, $p = 0.223$). There was also
199 a significant pattern for position along the trunk when controlling for the tree (perMANOVA R^2

200 = 0.001727, $p = 0.024$) but not for position along the trunk when we stratified for aspect
201 (perMANOVA $R^2 = 0.01727$, $p = 0.412$).

202 The perMANOVA results for the 21 more spatially dispersed balsam fir also showed a
203 significant pattern for macrolichens between the north and south sides of the tree, when
204 stratifying by tree (perMANOVA $R^2 = 0.00988$, $p = 0.009$) but not when controlling for position
205 along the trunk (perMANOVA $R^2 = 0.0098$, $p = 0.081$). Unlike for the trees in the single stand,
206 there was no significant pattern for position along the trunk when controlling for the tree
207 (perMANOVA $R^2 = 0.00476$, $p = 0.157$) nor for position along the trunk when stratified for
208 aspect (perMANOVA $R^2 = 0.00476$, $p = 0.444$). When examining a different tree species, yellow
209 birch, there was no significantly consistent pattern of lichen between the north and south sides of
210 the trunk, either when stratifying by tree (perMANOVA $R^2 = 0.00147$, $p = 0.772$) or by position
211 along the trunk (perMANOVA $R^2 = 0.00147$, $p = 0.85$). Nor was there any significant pattern
212 along the trunk of the yellow birch when stratifying by tree (perMANOVA $R^2 = 0.00143$, $p =$
213 0.721) or when controlling for aspect (perMANOVA $R^2 = 0.00143$, $p = 0.811$). Overall patterns
214 for macro-lichens are summarized in Table 1.

215 When we looked at both macro- and micro-lichens, the patterns were different. Balsam
216 fir did not show any significant pattern (aspect stratified by tree perMANOVA $R^2 = 0.00497$ $p =$
217 0.051 ; aspect stratified by position along trunk perMANOVA $R^2 = 0.00497$, $p = 0.439$; position
218 along trunk stratified by tree perMANOVA $R^2 = 0.00245$, $p = 0.471$; position along trunk
219 stratified by aspect perMANOVA $R^2 = 0.00245$, $p = 0.843$). In contrast, there was a significant
220 pattern for aspect for yellow birch, both when stratifying by tree (perMANOVA $R^2 = 0.01528$, p
221 $= 0.001$) and by position along the trunk (perMANOVA $R^2 = 0.01528$, $p = 0.002$). However,
222 there was not a significant pattern for location up the trunk for yellow birch, neither when

223 stratifying by tree (perMANOVA $R^2 = 0.00373$, $p = 0.297$) nor by aspect (perMANOVA $R^2 =$
224 0.00373 , $p = 0.647$). Table 2 summarizes the overall patterns for macro- and micro-lichens
225 combined.

226

227 **4. Discussion**

228 The consistent lichen patterns of macrolichens along the trunks of balsam fir which had
229 previously been observed within a single stand [14] showed some similarities when we looked at
230 21 balsam fir trees scattered across a wider region. The lichen pattern along the north vs. south
231 sides of trees held between the single stand and the wider region. The pattern along the gradient
232 of the tree trunk that was observed in the initial study did not hold in the new study. However, in
233 this more spatially dispersed sampling, we only looked at lichen patterns along a 50 cm
234 microtransect along the tree bole. Thus, it is possible that balsam fir across a wider region exhibit
235 consistent patterns, but only when examining at least a 1 m “microtransect”. Yellow birch, on the
236 other hand did not show any of the landscape patterns posited by Wiersma and McMullin [14]
237 for macro-lichens. However, when micro-lichens were included, yellow birch did show
238 significant patterns by aspect and thus might be considered potential replicate landscapes when
239 the full suite of lichen species is included.

240 The comparison of the microlandscape pattern in the 24 balsam fir in the single stand
241 (from [14]) to the 21 more widely dispersed trees sampled here, suggests that the idea of ‘trees as
242 replicate landscapes’ has support across the Avalon Forest Ecoregion. The lichen pattern on the
243 more widely dispersed trees was consistent when examining lichen patterns on north- vs. south-
244 facing sides of the trunks. Although we did not observe the same strong pattern of a gradient
245 along the trunk, we believe that this might be due to the shorter microtransect in this new study
246 (50 cm vs. 1 m). The consistency of pattern with aspect in the more widely dispersed balsam fir

247 is especially notable given that the canopy cover and size of the more dispersed trees was quite
248 different (more open canopy and larger trees) in the 21 dispersed trees compared to the 24 trees
249 in the single stand (Table 3).

250 Although this study shows support for the concept of treating lichens on tree trunks as
251 analogous to patches on a larger landscape [14] for balsam fir, it also illustrates that the proposed
252 model system may not hold true for all tree species, nor for all lichens. There was no consistent
253 pattern for yellow birch when we included only macro-lichens, and when assessed macro- and
254 micro-lichens together, there was a consistent pattern for yellow birch but the patterns on balsam
255 fir disappeared. This might be due to the fact that the lichen community on the two trees differed
256 [30]; yellow birch had higher overall lichen diversity (mean of 11.9 ± 2.86 species on yellow
257 birch vs. 9.86 ± 3.26 on balsam fir [30]). There were nine lichen species found on yellow birch
258 that were not on the balsam fir in either study site; of these, six species were micro-lichens. Thus,
259 the possibility to treat trees as landscapes is only supported partially by this study. Assessments
260 of whether trees are replicate units need to consider variation between host species tree and the
261 extent of lichen sampling taxonomically. Micro-lichens require a much higher degree of
262 specialization to recognize in the field and identify than macro-lichens. We only saw consistent
263 landscape patterns on yellow birch when we included micro-lichens. Thus, in this ecosystem at
264 least, researchers wishing to use yellow birch as replicates need to be mindful of the need for
265 specialized lichen expertise. In other systems, some pre-sampling to assess whether certain trees
266 are substrates for a wider array of specialized species may be useful before assessing the
267 potential of any one tree to function as an experimental microlandscape.

268 There are several implications for research based on the consistent landscape patterns
269 across more widely dispersed trees, as we have shown here for macro-lichens on balsam fir and

270 for macro- and micro-lichens on yellow birch. For one thing, this means that there are replicate
271 experimental units across a broader spatial extent against which different hypotheses can be
272 tested than would be possible with replicate trees in a single stand (e.g., [31]). The wider extent
273 of replicate “landscapes” allows for natural-experiments to see how landscapes respond to meso-
274 scale conditions such as distance to different types of habitat (e.g., open bogs), or broader
275 gradients in elevation and climate. It also allows for manipulative experiments at larger extents
276 that might mimic real-world processes, for example, looking at responses to different disturbance
277 levels. We suggest that future research focuses on testing whether the concept of replicate
278 microlandscapes holds for lichens along tree trunks in different forest types, beyond the boreal.
279 Such patterns would allow for more extensive and expansive experimentation.

280

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293 **References**

- 294 1. Forman RTT. Land Mosaics: the ecology of landscapes and regions. Cambridge: Cambridge
295 University Press; 1995.
- 296 2. Gustafson EJ, Lytle DE, Swaty R., Loehle C. Simulating the cumulative effects of multiple
297 forest management strategies on landscape measures of forest sustainability. *Landscape*
298 *Ecology*. 2007; 22: 141–156. <https://doi.org/10.1007/s10980-006-9017-y>
- 299 3. Kie JG, Ager AA, Bowyer RT. Landscape-level movements of North American elk (*Cervus*
300 *elaphus*): effects of habitat patch structure and topography. *Landscape Ecology*. 2005; 20:
301 289–300. <https://doi.org/10.1007/s10980-005-3165-3>
- 302 4. Theobald DM, Hobbs NT, Bearly T, Zack JA, Shenk T, Riebsame WE. Incorporating
303 biological information in local land-use decision making: designing a system for
304 conservation planning. *Landscape Ecology*. 2000; 15: 35–45.
305 <https://doi.org/10.1023/a:1008165311026>
- 306 5. Wiens JA, Milne BT. Scaling of 'landscapes' in landscape ecology, or, landscape ecology
307 from a beetle's perspective. *Landscape Ecology*. 1989; 3: 87–96.
308 <https://doi.org/10.1007/bf00131172>
- 309 6. Ims RA. The role of experiments in landscape ecology. In: Wiens JA, Moss MR (eds.),
310 *Issues and perspectives in landscape ecology*. Cambridge: Cambridge University Press;
311 2005.
- 312 7. Hargrove WW, Pickering J. Pseudoreplication: a *sine qua non* for regional ecology.
313 *Landscape Ecology*. 1992; 6: 251–258. <https://doi.org/10.1007/BF00129703>
- 314 8. Hurlbert SH. Pseudoreplication and the design of ecological field experiments. *Ecological*
315 *Monographs*. 1984; 54: 187–211. <https://doi.org/10.2307/1942661>

- 316 9. Hurlbert SH. On misinterpretation of pseudoreplication and related matters: a reply to
317 Oksanen. *Oikos*. 2004; 104: 591–597. <https://doi.org/10.1111/j.0030-1299.2004.12752.x>
- 318 10. Eberhardt LL, Thomas JM. Designing environmental field studies. *Ecological Monographs*.
319 1991; 61: 53–73. <https://www.jstor.org/stable/1942999>
- 320 11. Borregaard MK, Hart EM. 2016. Towards a more reproducible ecology. *Ecography*. 2016;
321 39: 349–353. <https://doi.org/10.1111/ecog.02493>
- 322 12. Vitousek PM. Ocean islands as model systems for ecological studies. *J Biogeogr*. 2002; 29:
323 573–582. <https://doi.org/10.1046/j.1365-2699.2002.00707.x>
- 324 13. Bowker MA, Maestre FT, Eldridge D, Belnap J, Castillo-Monroy A, Escolar C. Biological
325 soil crusts (biocrusts) as a model system in community, landscape and ecosystem ecology.
326 *Biodiversity and Conservation*. 2014; 23: 1619–1637. [https://doi.org/10.1007/s10531-014-](https://doi.org/10.1007/s10531-014-0658-x)
327 [0658-x](https://doi.org/10.1007/s10531-014-0658-x)
- 328 14. Wiersma YF, McMullin RT. Is it common to be rare on the landscape? A test using a novel
329 model system. *Landscape Ecology*. 2018; 33: 183–195. [https://doi.org/10.1007/s10980-017-](https://doi.org/10.1007/s10980-017-0599-3)
330 [0599-3](https://doi.org/10.1007/s10980-017-0599-3)
- 331 15. Patiño J, Gómez-Rodríguez C, Pupo-Correia A, Sequeira M, Vanderpoorten A. Trees as
332 habitat islands: temporal diversity in alpha and beta diversity in epiphytic laurel forest
333 bryophyte communities. *J Biogeogr*. 2018; 45:1727–1738. <https://doi.org/10.1111/jbi.13359>
- 334 16. Underwood AJ. *Experiments in Ecology. Their logical design and interpretation using*
335 *analysis of variance*. Cambridge: Cambridge University Press; 1997.
- 336 17. Oksanen J. Logic of experiments in ecology: is pseudoreplication a pseudoissue? *Oikos*.
337 2001; 94: 27–38. <https://doi.org/10.1034/j.1600-0706.2001.11311.x>
- 338 18. Forman RTT, Godron M. *Landscape Ecology*. New York: John Wiley; 1986.

- 339 19. Li X, Wu J. Use and misuse of landscape indices. *Landscape Ecology*. 2004; 19: 389–399.
340 <https://doi.org/10.1023/b:land.0000030441.15628.d6>
- 341 20. Elköf, JS, Alsterberg C, Havenhand JN, Sundbäck K, Wood HL, Gamfeldt L. Experimental
342 climate change weakens the insurance effect of biodiversity. *Ecology Letters*. 2012; 15:
343 864–872. <https://doi.org/10.1111/j.1461-0248.2012.01810.x>
- 344 21. von Humboldt, A. 1805. Excerpt from “Essay on the geography of plants”. Translations by
345 Kern F, Janvier P. from *Essai sur la Géographie des Plantes*. Paris: Leevrault, Schoell et Cie,
346 1805. As published in: Lomolino MV, Sax DF, Brown JH, editors. *Foundations of*
347 *biogeography, classic papers with commentaries*. Chicago: The University of Chicago Press;
348 2004. pp. 49-57.
- 349 22. Merriam CH. Excerpts from “Results of a biological survey of the San Francisco Mountain
350 region and the desert of the Little Colorado, Arizona”. *North American Fauna* no. 3.
351 Washington: U.S. Department of Agriculture; 1890. As published in: Lomolino MV, Sax
352 DF, Brown JH, editors. *Foundations of biogeography, classic papers with commentaries*.
353 Chicago: The University of Chicago Press; 2004. pp. 202-233.
- 354 23. South G. *Biogeography and ecology of the Island of Newfoundland*, 1st ed. The Hague: Dr.
355 W. Junk Publishers; 1983.
- 356 24. Hättestrand C, Kleman J. Ribbed moraine formation. *Quaternary Science Reviews*. 1999;
357 18(1): 43–61. [https://doi.org/10.1016/S0277-3791\(97\)00094-2](https://doi.org/10.1016/S0277-3791(97)00094-2)
- 358 25. Brodo IM, Sharnoff SD, Sharnoff S. *Lichens of North America*. New Haven: Yale
359 University Press; 2001.

- 360 26. Culberson CF, Kristinsson H. A standardized method for the identification of lichen
361 products. *Journal of Chromatography*. 1970; 46: 85–93. [https://doi.org/10.1016-s0021-](https://doi.org/10.1016/s0021-9673(00)83967-9)
362 [9673\(00\)83967-9](https://doi.org/10.1016/s0021-9673(00)83967-9)
- 363 27. Anderson MJ. A new method for non-parametric multivariate analysis of variance. *Austral*
364 *Ecology*. 2001. 26: 32–46. <https://doi.org/10.1111/j.1442-9993.2001.01070.pp.x>
- 365 28. R Core Team. R: A language and environment for statistical computing; 2016. R Foundation
366 for Statistical Computing, Vienna, Austria. Available from: <https://www.R-project.org/>
- 367 29. Oksanen J, Blanchet GF, Kindt R, Legendre P, McGlinn D, Minchin PR, et al. *vegan*:
368 *Community Ecology Package*; 2016. R package version 2.3-4. Available from:
369 <https://CRAN.R-project.org/package=vegan>
- 370 30. Wigle RD. Arboreal lichen community structure and diversity on yellow birch (*Betula*
371 *alleghaniensis*) and balsam fir (*Abies balsamea*) in the Avalon Forest Ecoregion in
372 Newfoundland, Canada. MSc Thesis, Memorial University of Newfoundland; 2018.
- 373 31. Bokhorst S, Asplund J, Kardol P, Wardle DA. Lichen physiological traits and growth forms
374 affect communities of associated invertebrates. *Ecology*. 2015; 96: 2394–2407.
375 <https://doi.org/10.1890/14-1030.1>

377 **Figure Captions**

378 **Figure 1.** Location of sampling sites in the Avalon Forest Ecoregion on the island of
379 Newfoundland, Canada. Inset map shows the location of the Avalon Forest Ecoregion (black
380 polygon). Black stars on the main map are sampling locations for the 21 sites where we sampled
381 both balsam fir and yellow birch. The location labelled “Halls Gullies” designates the stand
382 within which 24 balsam fir were sampled.

383

384 **Figure 2.** A lichen ladder, which was used to sample lichen diversity on tree trunks. Each square
385 is 10 x 10 cm; the ladder is 50 cm in length and was placed with the top rung at 1.6 m from the
386 ground.

387

389 **Table 1.** Summary of perMANOVA analysis for trees from two studies within the Avalon Forest
 390 Ecoregion; one for 24 balsam fir in a single stand; and for 21 more widely dispersed sites with
 391 one balsam fir and one yellow birch at each site. Analysis here is for consistency in lichen
 392 patterns on the tree trunks for macro-lichens only along a 50 cm section of the tree trunk on the
 393 north- and south-facing sides.

Variable	Balsam fir in a single stand (<i>n</i> = 24)	Balsam fir dispersed (<i>n</i> = 21)	Yellow birch dispersed (<i>n</i> = 21)
Aspect controlling for tree	significant	significant	non-significant
Aspect controlling for location up tree	non-significant	non-significant	non-significant
Location up tree controlling for tree	significant	non-significant	non-significant
Location up tree controlling for aspect	non-significant	non-significant	non-significant

394

396 **Table 2.** Summary of perMANOVA analysis for 21 widely dispersed sites across the Avalon
 397 Forest Ecoregion with one balsam fir and one yellow birch at each site. Analysis here is for
 398 consistency in lichen patterns along 50 cm of the tree trunks along the north and south-facing
 399 sides, for macro-lichens and mirco-lichens combined.

Variable	Balsam fir dispersed (<i>n</i> = 21)	Yellow birch dispersed (<i>n</i> = 21)
Aspect controlling for tree	non-significant	significant
Aspect controlling for location up tree	non-significant	significant
Location up tree controlling for tree	non-significant	non-significant
Location up tree controlling for aspect	non-significant	non-significant

400

401

403 **Table 3.** Comparison of balsam fir tree measurements. dbh = diameter at breast height (1.3 m).

404 Values are given as mean (+/- standard deviation)

Sample trees	dbh (cm)	Canopy (%)	Height (m)
Dispersed trees (n = 21)	24.5 (5.9)	25 (9.4)	7.9 (1.6)
Trees in a single stand (n = 24)	10.6 (2.7)	82 (12.8)	7.1 (2.2)

405

406

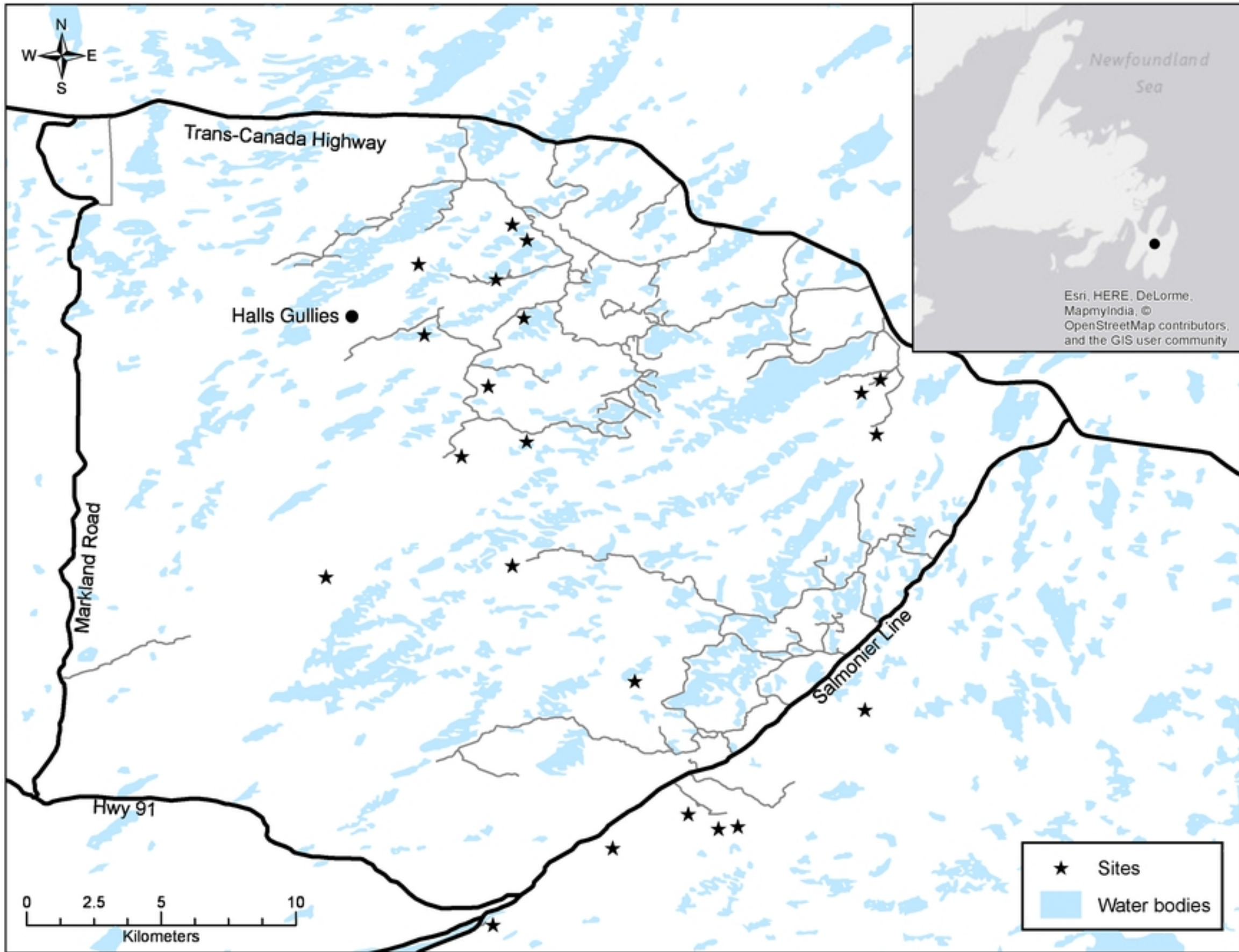


Figure 1

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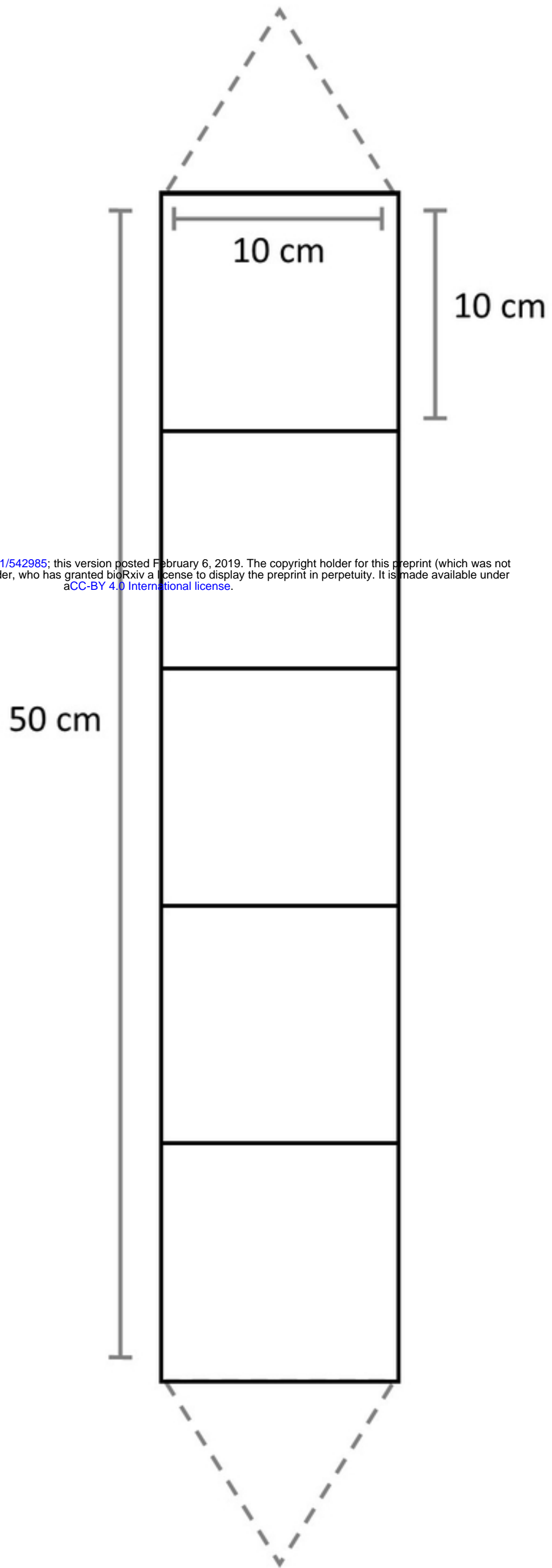


Figure 2