

1 **Maternal RhD heterozygous genotype is associated with**
2 **male biased secondary sex ratio**

3 Šárka Kaňková^{a*}, Jaroslav Flegr^a, Jan Toman^a, Pavel Calda^b

4

5 ^aDepartment of Philosophy and History of Science, Faculty of Science, Charles University,

6 Prague, CZ-128 44 Prague 2, Czech Republic

7 ^bDepartment of Obstetrics and Gynaecology, General University Hospital and First Faculty of

8 Medicine, Charles University, Prague, CZ-128 08 Prague 2, Czech Republic

9

10 *Corresponding author:

11 Tel.: +420 221951821, fax: +420 224919704

12 E-mail: kankova.sarka@gmail.com

13 Division of Biology, Faculty of Science, Charles University, Prague, Viničná 7, 128 44,

14 Czech Republic

15

16 **Declarations of interest**

17 None.

18

19 **Acknowledgment**

20 This work was supported by Czech Science Foundation (grant No. 18-13692S); Charles

21 University (Research Centre program No. 204056); and the Ministry of Health of the Czech

22 Republic (grant RVO-VFN64165). The funding sources were not involved in study design, in

23 the collection, analysis and interpretation of data, in the writing of the report and in the

24 decision to submit the article for publication.

25

26 **Abstract**

27 The results of previous studies overwhelmingly suggest that RhD positive heterozygotes
28 express better health status than Rh positive homozygous, especially in RhD negative
29 subjects. This also applies to pregnant women. According to the Trivers-Willard hypothesis,
30 women in better physical condition should have a male-skewed sex ratio. The aim of the
31 present study was to test the hypothesis that RhD positive heterozygous mothers give birth to
32 more sons than daughters. In the present cross-sectional study, we analysed data from 5,655
33 women who have given birth in the General University Hospital in Prague, Czech Republic
34 between 2008-2012. Clinical records comprised maternal weight before pregnancy, number of
35 previous deliveries, sex of the newborn, maternal RhD phenotype, and RhD phenotype of the
36 newborn. Secondary sex ratio was significantly higher ($P=0.028$) in RhD positive mothers
37 who had RhD negative newborns, i.e. in heterozygotes ($SR=1.23$), than in RhD positive
38 mothers who had RhD positive newborns, i.e. in a mixed population of heterozygotes and
39 homozygotes ($SR=1.00$), especially in primiparous women ($P=0.013$; $SR=1.37$ and 0.99
40 resp.). In line with the Trivers-Willard effect, RhD maternal heterozygous genotype is
41 associated with male biased secondary sex ratio. The results supported the hypothesis that
42 RhD polymorphism may be maintained due to heterozygote health advantages.

43

44 **Key words**

45 sex ratio; RhD polymorphism; Trivers-Willard hypothesis; frequency-dependent selection;
46 heterozygote advantage

47

48 **1. Introduction**

49 The RhD protein, a product of the *RHD* gene, is a major component of the Rh blood group
50 system. It carries the strongest blood group immunogen, which is the D antigen. Considering
51 its role, the molecular structure of the RhD protein suggests it is a part of an ion pump present
52 in the red blood cell membrane. The whole complex probably serves for the transport of NH₃
53 or CO₂ molecules across the erythrocyte cell membrane (Flegel, 2011; Kustu and Inwood,
54 2006). However, its physiological role is still unclear. Several possible options have been
55 discussed, e.g. in Flegr et al. (2015).

56 Almost 85 % of Europeans express an RhD positive phenotype – the RhD protein is
57 present in their erythrocyte cell membranes. However, the RhD antigen is absent in a
58 considerable portion of the European population (RhD negative subjects) due to the *RHD*
59 deletion (Wagner and Flegel, 2000). RhD polymorphism is comparably high in numerous
60 other human populations (Golassa et al., 2017; Mourant, 1976), but its existence is an
61 evolutionary enigma. Theoretically, populations should be RhD monomorphic. This is
62 because of a strong selection against RhD positive children born to RhD negative mothers due
63 to hemolytic disease of the newborn (Bowman, 1997; Filbey et al., 1995). Before the
64 introduction of prophylactic treatment, this disorder, which can cause serious illness, brain
65 damage, or even death of the fetus or newborn of multiparous women, was one of the leading
66 causes of newborn mortality in highly RhD polymorphic populations. Therefore, the
67 representatives of the minor phenotype had lower fitness before the advent of modern
68 medicine. This includes either RhD negative women in a mostly RhD positive population or
69 RhD positive men in a mostly RhD negative population.

70 Erythrocytes of RhD- and RhD+ homozygotes differ in the molecular complexes
71 present on their cell membranes, and most likely in their biological activities as well (Le Van
72 Kim et al., 2006; Wagner and Flegel, 2000). Moreover, a difference in the erythrocytes of

73 RhD positive homozygotes and heterozygotes was also observed. About 33,560 D antigen
74 sites were detected on the surface of RhD positive homozygous erythrocytes, whereas the
75 surface of RhD heterozygous erythrocytes contain about 17,720 D antigen sites (McGann et
76 al., 2012). It is thus possible that the vulnerability of RhD positive homozygotes, RhD
77 positive heterozygotes and, especially, RhD negative homozygotes to various irregular
78 conditions, including various diseases, may differ dramatically. RhD heterozygotes may link
79 the advantages of both RhD negative and positive homozygotes and the RhD polymorphism
80 in human populations thus could be maintained by selection in favour of heterozygotes
81 (Feldman et al., 1969; Fincham, 1972).

82 The correlation of particular RhD genotypes with specific health conditions has been
83 supported by several studies (see, e.g. Flegr et al., 2008, 2009, 2010, 2012, 2013; Kankova et
84 al., 2010; Novotna et al., 2008). One interesting theme is that RhD negativity and
85 heterozygosis often affect the individual's health in opposite directions. Recently, two
86 independent studies showed that RhD phenotype strongly affects the incidence and
87 prevalence of many disorders. RhD negative subjects reported having more frequent allergic,
88 digestive, heart, haematological, immunity, mental health, and neurological problems, as well
89 as a higher incidence of some infectious diseases. This was corroborated by the reported
90 frequency of their visits to medical specialists, usage of prescribed drugs, headaches, and
91 general tiredness (Flegr et al., 2015). The results showed a complex picture. Certain
92 significantly elevated medical problems were specific to RhD negative individuals, others to
93 RhD positive persons, most of them in a sex-specific way. However, taken together, the RhD
94 negative subjects had more serious health problems than the RhD positive subjects in all six
95 variables, significantly differing according to RhD of 22 variables analysed. The difference in
96 RhD phenotype was obviously caused by the underlying RhD genotype, but RhD positive
97 homozygotes and heterozygotes were not separated in this study. Some of the results indicate

98 that RhD negative phenotype may confer increased immunity to infections of viral origin. In
99 the light of these results and considering the long-term persistence of RhD polymorphism,
100 however, it seems more probable that it could be RhD positive heterozygotes who are
101 selectively advantageous in human populations. RhD polymorphism thus could be sustained
102 in populations by negative frequency-dependent selection, namely by its specific form – the
103 selection in favour of heterozygotes (heterozygous advantage).

104 A recent ecological regression study performed on a set of 65 countries for which the
105 RhD genotype frequencies data were available, showed the strongest evidence yet for the
106 heterozygote advantage hypothesis (Flegr, 2016). The results showed that both the
107 frequencies of RhD negative homozygotes and RhD positive heterozygotes (whose frequency
108 was calculated from data on the frequency of homozygotes using Hardy-Weinberg equation)
109 correlated with specific disease burdens in particular countries. In general, the burdens (both
110 in the sense of Disability Adjusted Life Year, see WHO, 2008, and disease mortality rates)
111 were higher in RhD negative homozygotes. Moreover, the observed correlations mostly lead
112 in opposite directions in RhD negative homozygotes and RhD positive heterozygotes. The
113 general pattern showed that the countries with a high frequency of Rhesus negative
114 homozygotes had a lower burden associated with congenital anomalies and neuropsychiatric
115 conditions. Additionally, such countries had a higher burden of cardiovascular diseases and,
116 especially, of malignant neoplasm. This strongly supports the hypothesis that RhD
117 polymorphism could be sustained in populations by selection in favour of heterozygotes at the
118 expense of alternative hypotheses that consider its current distribution to be a consequence of
119 founder effects, genetic disequilibrium, and/or gene flow attenuated by viability selection,
120 partial reproductive incompatibility, and reproductive compensation (see, e.g. Feldman et al.,
121 1969; Nei et al., 1981).

122 Several studies directly supporting RhD heterozygous advantage have also been
123 published. Flegr et al. (2008) and Novotna et al. (2008) showed that healthy RhD negative
124 homozygotes exhibit faster reactions than RhD positive subjects. This reaction time, however,
125 strikingly changes when subjects were infected by *Toxoplasma gondii*, a common parasite
126 whose chronic prevalence in various countries ranges between ten and ninety percent.
127 *Toxoplasma* positive subjects showed prolonged reaction times. This impairment was most
128 severe in RhD negative homozygotes, whereas RhD positive individuals, and especially RhD
129 positive heterozygotes, were considerably protected from this effect. In consequence, the
130 performance of *Toxoplasma* positive subjects was best in RhD positive heterozygotes
131 followed by RhD positive homozygotes and RhD negative homozygotes. The same data also
132 showed that RhD positivity (most probably RhD heterozygosity) modulates certain effects of
133 smoking, fatigue, and ageing (Flegr et al., 2012). For example, the positive effect of age on
134 performance and intelligence was stronger in RhD positive subjects, whereas the effect of
135 smoking on the number of viral and bacterial diseases was about three times higher in RhD
136 negative subjects. In a different study, *Toxoplasma* positive RhD negative subjects expressed
137 lower performance in weight-holding and hand-grip tests in comparison with both RhD
138 negative and positive *Toxoplasma* negative subjects (Flegr et al., 2018). It is clear that faster
139 reaction times or greater stamina conferred a selective advantage in the past, and that they still
140 do in the present. It was, for example, demonstrated that the probability of being involved in a
141 traffic accident is elevated more than twice (in the case of chronic infection) or even five
142 times (in the case of recent infection) in *Toxoplasma* positive RhD negative homozygotes
143 (Flegr et al., 2009). This could lead to the spread of the RhD negative allele in a similar
144 manner to the recessive *HBB* allele for sickle cell disease. *HBB* recessive homozygotes have
145 severely impaired viability. In a heterozygous condition, the *HBB* allele does not impair
146 viability in most situations but confers an increased resistance to malaria. The *HBB* allele thus

147 spread in tropical areas with *Plasmodium falciparum* by the means of frequency-dependent
148 selection, or its specific form – heterozygous advantage – until a disproportionate fraction of
149 nonviable recessive homozygotes are born and its frequency in a population is stabilized
150 (Allison, 1954).

151 However, phenotypic correlates of RhD phenotype may be more complex. A study
152 performed on 502 soldiers surprisingly showed that *Toxoplasma* positive RhD positive
153 subjects express lower, while *Toxoplasma* positive RhD negative subjects express higher,
154 verbal and nonverbal intelligence than their *Toxoplasma* negative peers (Flegr et al., 2013).
155 This may be the consequence of psychological differences between *Toxoplasma* positive and
156 *Toxoplasma* negative subjects (a lower total N-70 score of potentially pathognomic factors,
157 anxiety, depression, phobia, hysteria, vegetative lability, hypochondria, psychasteny, and
158 neuroticism, but see Flegr et al., 2010, for somewhat contradictory results) that were observed
159 in this study and were more prominent among RhD negative subjects. However, these
160 psychological differences may result from the increased tendency of *Toxoplasma* positive
161 military personnel to mask any negative properties. Moreover, there were signs of higher
162 verbal and nonverbal intelligence even in *Toxoplasma* negative RhD negative subjects, which
163 points to a possible direct effect of RhD phenotype. Regardless, the protective role of RhD
164 positive phenotype against the effects of toxoplasmosis was documented even in this study.
165 The strongest effect of Rh phenotype was reported in a study on the influence of *Toxoplasma*
166 on weight gain in pregnancy. In the study of Kankova et al. (2010), RhD negative women
167 with latent toxoplasmosis gained nearly two times more weight in 16th week of pregnancy (N
168 = 27) than RhD negative *Toxoplasma* negative women (N = 139) or the RhD positive women
169 of any infection status (N = 813). The difference of about 1600 g remained approximately
170 constant until delivery.

171 The secondary sex ratio (sex ratio at birth) in humans is around 1.06 in most
172 populations (Davis et al., 1998). Within the population, the sex ratio may be influenced by
173 many factors, such as paternal hormones (James, 1996, 2010, 2015; James and Grech, 2018),
174 immunosuppression (James, 1996), and several important pathologies (e.g. hepatitis, James,
175 2010, or toxoplasmosis, Kankova et al., 2007). As the Rh phenotype modulate the effects of
176 many detrimental factors on human performance and physiology, we decided to examine its
177 potential effects on sex ratio at birth. The specific aim of the present study was to analyse the
178 association between the sex of newborns and RhD phenotype of both the newborns and
179 mothers. Our working hypothesis was to expect a higher secondary sex ratio in RhD
180 heterozygous women. The generally accepted Trivers-Willard hypothesis (Trivers and
181 Willard, 1973) suggests that females, including women in “good condition”, e.g. women with
182 a good health status, tend to give birth to more sons than daughters. The reason is that
183 mothers in good condition may invest disproportionately more time, energy, and resources
184 into their sons, which may, in turn, reach a better condition and leave more offspring in
185 polygynous (or serially monogamous) species where the biological fitness of males, but not so
186 much females, strongly depends on their health status. According to the studies, heterozygous
187 RhD positive mothers probably express better health status than Rh positive homozygous
188 mothers and especially than the RhD negative mothers. This can result in a higher sex ratio in
189 RhD positive than in RhD negative mothers and an even higher sex ratio in RhD positive
190 mothers who give birth to RhD negative children, i.e., in RhD heterozygous mothers. We
191 expect this effect to be more prominent in primiparous women on the basis of already
192 published results (Christiansen et al., 2004; Maraz et al., 1973; Nielsen et al., 2008).
193

194 **2. Material and Methods**

195 ***2.1. Subjects***

196 The study was designed as a cross-sectional study. The main data set covered women who have
197 given birth in the General University Hospital in Prague, Czech Republic between 2008-2012.
198 Clinical records comprised maternal weight before pregnancy, number of previous deliveries
199 (primiparae/multiparae), sex of the newborn, maternal RhD phenotype (positive/negative), and
200 RhD phenotype of the newborn (positive/negative). The women that gave birth to twins were
201 excluded from the analyses. During the whole study, we worked with an anonymized data set.

202 ***2.2. Statistics***

203 The program Statistica 10.0 was used for all statistical testing. The association between the
204 sex of the newborn and the RhD phenotype of the newborn was analysed using the Chi-
205 Square test, initially for all women, and then separately for RhD positive and RhD negative
206 women. Furthermore, the analysis was conducted separately for both primiparous and
207 multiparous women. The effects of maternal weight as a continuous predictor and the RhD
208 phenotype of newborn (positive/negative) as a categorical predictor of newborn sex, were
209 evaluated by the generalized linear model (GLZ) separately for both RhD positive and RhD
210 negative women. Binomial distribution and logit link function, as recommended by Wilson
211 and Hardy (2002), were used for the construction of the model. For some women, the variable
212 “maternal weight before pregnancy” was not available, and therefore the number of women
213 varied between analyses. Sex ratio (SR) in this article is expressed as the ratio of male to
214 female.

215 ***2.3. Ethical approval***

216 The project was approved by IRB Faculty of Science, Charles University (No. 2018/19) (Etická
217 komise pro práci s lidmi a lidským materiálem Přírodovědecké fakulty Univerzity Karlovy).

218 3. Results

219 The total data set contained the records of 5,655 women. The relationships between the sex of
220 newborns and the RhD phenotype of the newborns analysed using the Chi-Square test showed
221 only a non-significantly male biased proportion of the newborns in the group of RhD negative
222 newborns (N= 5,655, P=0.088, $\chi^2=2.91$). Among 1,356 RhD negative newborns, 717 were
223 boys (SR=1.12), while among 4,299 RhD positive newborns, only 2,151 were boys
224 (SR=1.01). However, the situation differed in the RhD positive and RhD negative
225 subpopulations of women. In women with the RhD positive phenotype, who included Rh
226 positive homozygotes and heterozygotes, the relationships between the sex of newborn and
227 the RhD phenotype of the newborn were significant (N=3,406, P=0.028, $\chi^2=4.84$). Among
228 422 RhD negative newborns, the offspring of the RhD positive heterozygote mothers, 235
229 were boys (SR=1.26), while among 2,984 RhD positive newborns, the offspring of either
230 RhD positive heterozygote or RhD positive homozygote mothers, 1493 were boys (SR=0.99).
231 In women with the RhD negative phenotype, the relationships between the sex of the
232 newborns and the RhD phenotype of the newborns did not exist (N=2,229, P=0.660, $\chi^2=0.19$).
233 Among 929 RhD negative newborns, 479 were boys (SR=1.06), while among 1,300 RhD
234 positive newborns, 658 were boys (SR=1.02).
235 The same analyses were conducted separately for both primiparous and multiparous women.
236 In primiparous women, the relationship between the sex of the newborn and the RhD
237 phenotype of the newborn analysed using the Chi-Square test showed a significantly male
238 biased proportion of the newborns in the group of RhD negative newborns (N= 3,418,
239 P=0.013, $\chi^2=6.21$). Among 844 RhD negative newborns, 466 were boys (SR=1.23) and
240 among 2,574 RhD positive newborns, 1,294 were boys (SR=1.01). In women with the RhD
241 positive phenotype, the relationship between the sex of the newborn and the RhD phenotype
242 of the newborn was also significant (N=2,047, P=0.013, $\chi^2=6.18$). Among 258 RhD negative

243 newborns, 150 were boys (SR=1.37) and among 1,789 RhD positive newborns, 892 were
244 boys (SR=0.99). In primiparous women with RhD negative phenotype, the relationship
245 between the sex of the newborns and the RhD phenotype of the newborns did not exist
246 (N=1,360; P=0.335; $\chi^2=0.93$). Among 583 RhD negative newborns, 314 were boys (SR=1.17)
247 and among 777 RhD positive newborns, 398 were boys (SR=1.02). In multiparous women,
248 both RhD negative (P=0.559) and RhD positive (P=0.690), no significant results were
249 observed.

250 In the next part of the study, we analysed the influence of the RhD phenotype of newborns
251 (positive/negative) on the sex of newborns using a generalized linear model (GLZ). At first,
252 this test was conducted only for RhD positive women, while the variables “maternal weight”
253 and “maternal age” were used as the continuous covariates. However, the maternal weight
254 (P=0.998) and the maternal age (P=0.484) did not make a significant contribution to the
255 equation. Finally, the reduced statistical model was conducted without them. The results of
256 GLZ (P values) were similar to the results of the Chi-Square test mentioned above. Again, no
257 significant effect of the RhD phenotype of newborns on the sex of newborns was observed in
258 the group of Rh negative women (data not shown).

259

260 **4. Discussion**

261 Our results showed that the sex ratio at birth was significantly higher (male skewed) in RhD
262 positive mothers who had RhD negative newborns (SR=1.23) than in RhD positive mothers
263 who had RhD positive newborns (SR=1.00). This effect was stronger in primiparous women
264 (SR=1.37 and 0.99 resp.). In our study, we had only data regarding RhD phenotypes. It can be
265 deduced, however, that the RhD negative mothers were undoubtedly homozygotes (dd) while
266 the RhD positive mothers could have either RhD positive heterozygous genotype (Dd) or
267 RhD positive homozygous genotype (DD). This uncertainty applies to the group of RhD

268 positive mothers who had RhD positive newborns. On the other hand, the RhD positive
269 mothers who had RhD negative newborns necessarily had RhD positive heterozygous
270 genotype. Therefore, the main observed result, i.e., higher secondary sex ratio in RhD
271 negative newborns of RhD positive mothers, confirms our *a priori* hypothesis that RhD
272 maternal heterozygous genotype is associated with male biased secondary sex ratio.

273 Recent results support the hypothesis that the Rhesus factor polymorphism is
274 maintained in human populations due to a higher resistance or tolerance of heterozygotes to
275 specific diseases (see Introduction). The hypothesis was repeatedly supported by empirical
276 data showing that RhD positivity, and especially RhD heterozygosity, protects people against
277 certain negative effects of toxoplasmosis (Flegr et al., 2008, 2018; Kankova et al., 2010;
278 Novotna et al., 2008). In line with the Trivers-Willard effect (Trivers and Willard, 1973), our
279 working hypothesis was to expect RhD heterozygous mothers, i.e., mothers with supposedly
280 higher health status, to exhibit a higher secondary sex ratio. The results of our study therefore
281 supported our working hypothesis and further supported the hypothesis that RhD
282 polymorphism may be maintained due to heterozygote health advantages.

283 As expected on the basis of already published data (Maraz et al., 1973), the results of
284 our analyses were not significant in multiparous women. This could have been caused by a
285 broader spectrum of factors that influence the secondary sex ratio in multiparous women in
286 comparison with primiparous women. These factors are, for example, sex (Christiansen et al.,
287 2004; Gualtieri et al., 1984; Renkonen et al., 1962) and RhD (Renkonen and Seppala, 1962)
288 of previous siblings, the existence or absence of previous miscarriages (Christiansen et al.,
289 2004; Nielsen et al., 2008), and immunosuppression (James, 1996), especially the reaction
290 against y-antigens (Nielsen et al., 2008, 2009). Due to these sources of latent variability, the
291 effect of RhD phenotype (resp. genotype) on sex ratio at birth cannot be detected in the
292 multiparous women, even if it existed. It must be emphasized, however, that the broader

293 spectrum of these confounding factors cannot be the only reason for the difference in the
294 effect of RhD on primiparous and multiparous women. Confounding factors may increase the
295 variability of the focal variable (the probability of a birth of a son in this study), and by this,
296 they can increase the P value of tests. However, they cannot influence the size of the observed
297 effect. For this reason, our results suggest that effects sizes in primiparous mothers are much
298 higher than in multiparous women, the difference in the effect of RhD on primiparous and
299 multiparous women cannot be explained solely by the aforementioned confounding factors.

300 Only RhD phenotype, not RhD genotype, of newborns and mothers is examined and
301 recorded in the current clinical praxis. The genotype of RhD negative mothers is dd and the
302 genotype of one third of RhD positive heterozygote mothers (Dd) can deduced from the RhD
303 phenotype of their children. Therefore, we can compute SRB (sex ratio at birth) in these two
304 subpopulations, but not in the subpopulation of RhD positive women with RhD positive
305 children, which represent a mixture of RhD positive homozygotes and heterozygotes. The
306 secondary sex ratio of RhD positive newborns of RhD positive mothers was 0.99. We can
307 expect that the secondary sex ratio in RhD positive newborns of RhD positive mothers with
308 RhD heterozygous genotype (part of RhD positive newborns of RhD positive mothers) is
309 comparable with the secondary sex ratio ($SR = 1.37$) of RhD negative newborns of RhD
310 positive mothers (with RhD heterozygous genotype). This would then suggest (in the case that
311 the Hardy-Weinberg equilibrium applies and the frequency of RhD negative homozygotes is
312 about 16 % in Czech population, see Flegr, 2016) that the secondary sex ratio in newborns of
313 RhD positive mothers with a RhD positive homozygous genotype is female biased ($SR =$
314 0.74) to make the total sex ratio 0.99 for the mixture of RhD positive heterozygotes ($SR =$
315 1.37) and RhD positive homozygotes.

316 It is also worth mentioning that the observed frequency of the RhD negative allele
317 (about 40 % of the standing polymorphism) conspicuously well approaches the proportion of

318 the recessive allele that ensures the production of the relatively highest proportion of
319 heterozygotes (i.e., the most fit individuals; in this case 48 %) and dominant homozygotes
320 (i.e., the individuals with mediocre fitness; 36 %) at the expense of recessive homozygotes
321 (i.e., the least fit individuals; 16 %). The highest possible stable proportion of the most fit
322 heterozygotes in the population is 50 %. However, both dominant homozygotes and the least
323 fit recessive homozygotes would constitute 25 % of the population in this scenario. Gradual
324 change in the proportion of both alleles in favour of dominant one would be expected in this
325 case. The observed deflection in the proportion of RhD alleles thus further supports the
326 hypothesis of the RhD heterozygote advantage and the relatively lowest fitness value of the
327 recessive RhD allele.

328 Possible alternative scenarios consistent with the observed data suggest it is possible
329 that the observed phenomenon of biased sex ratio in RhD negative children of RhD positive
330 mothers is associated with the child's RhD genotype. It cannot be excluded that RhD negative
331 embryos are rejected by maternal body less often, which would point to the ability of the RhD
332 allele or some closely linked allele to manipulate the mother's body or a general preference
333 and selective advantage of RhD negative homozygotes. However, this hypothesis does not
334 seem very probable because the sex ratio of RhD negative mothers was not significantly
335 biased.

336 Another option is based on the observation that adverse effects of RhD negativity on
337 health probably manifest at an older age (Flegr et al., 2015). This is corroborated by our
338 results (unpublished data), which show that better reaction times in RhD negative individuals
339 are specific to younger age. This points to a possible conditional selective advantage of RhD
340 negative homozygotes. Under this scenario, RhD negativity can be beneficial at a younger
341 age, or at least exhibit its adverse effects on health or psychomotor performance exclusively
342 or more prominently at an older age. Negative effects that manifest only at an older age could

343 hide away into the “selection shadow” (Fisher, 1930; Medawar, 1946). It was supported that
344 mutations that negatively affect fitness at an older age are under a much weaker selective
345 pressure than mutations that affect the fitness of young individuals (Gavrilova et al., 1998). It
346 is also possible that the adverse effects of RhD negative phenotype would be individually
347 reflected during ageing and thus masked by an adaptive behaviour (see, e.g. Flegr et al., 2009,
348 2013; Novotna et al., 2008). It is even possible that the RhD negative allele represents one of
349 the alleles with antagonistic effects on fitness dependent on age (see, e.g. Pedersen, 1995). Its
350 presence may easily result in a positive feedback loop of selection on fast life strategy and
351 maximum performance at a young age (Nettle, 2010; Promislow and Harvey, 1990). In such a
352 situation, it might be more advantageous to have more RhD negative sons than RhD negative
353 daughters for mothers with RhD positive heterozygous genotypes. Such sons would have a
354 better chance of reproductive success at a young age. In contrast, at least mediocre longevity
355 would be always advantageous for daughters. This presents a special case of the generalized
356 Trivers-Willard hypothesis (Kanazawa, 2005). Note, however, that this option does not
357 exclude advantages of preferentially producing sons by RhD heterozygous mothers in good
358 condition postulated by the classical Trivers-Willard hypothesis (Trivers and Willard, 1973).
359 The benefit of producing RhD negative sons because of their young age advantage therefore
360 might be masked in RhD negative mothers by their own advantage in producing daughters,
361 which results in the pattern we observed in our study.

362 The important limitation of our study is that we knew only maternal and child RhD
363 phenotypes and not genotypes. We were not able to count maternal RhD genotypes for all
364 data, but only for the subpopulation of 2,651 women (RhD negative women and RhD positive
365 women with RhD negative newborns). It will be necessary to genotype the RhD positive
366 mothers in future studies to confirm or reject our predictions that the SR of RhD positive
367 heterozygotes will be male skewed and the SR of RhD positive homozygotes will be female

368 skewed. The second limitation was a relatively small sample size of sub-analyses evaluated
369 across the different RhD phenotype categories for both primiparous and multiparous women.
370 However, the small number of subjects may lead only to a false negative, not a false positive
371 result of a study. Another, more general, problem was the absence of information on many
372 factors that could have influenced sex ratio at birth (e.g. maternal socio-economic status,
373 health, sex of previous child). Existence of known and unknown confounding factors may
374 result in the failure of a statistical test to detect the existent effect, not in the detection of non-
375 existing effect (Flegr and Horacek, 2017). However, researchers should focus on these factors
376 in future studies on the role of RhD polymorphism in the origin of skewed human sex ratios.

377 ***4.1. Conclusions***

378 In our relatively large data set, the maternal RhD heterozygosity was associated with a male
379 biased secondary sex ratio. The most parsimonious explanation of the observed pattern
380 suggests the preference of male offspring by mothers in good condition according to the
381 Trivers-Willard hypothesis. This hypothesis proposes that mothers who exhibit a good
382 physiological condition and/or social status give birth to a higher proportion of sons, while
383 those in a worse situation, e.g. those with worse health, give birth to more daughters. If the
384 explanation is correct, then our data provides new indirect support for the heterozygous
385 advantage hypothesis of sustaining RhD polymorphism in human populations. Moreover, the
386 possibility that the RhD homozygotes, both RhD negative and RhD positive, have worse
387 health than RhD heterozygotes could have serious clinical implications and should be
388 examined in detail in future studies.

389

390 **5. Data Availability**

391 The data associated with this research are available at

392 [https://figshare.com/articles/Maternal_RhD_heterozygous_genotype_is_associated_with_mal](https://figshare.com/articles/Maternal_RhD_heterozygous_genotype_is_associated_with_male_biased_secondary_sex_ratio/7687544)
393 [e_biased_secondary_sex_ratio/7687544](https://figshare.com/articles/Maternal_RhD_heterozygous_genotype_is_associated_with_male_biased_secondary_sex_ratio/7687544)

394

395 **6. References**

396 Allison, A. (1954). The distribution of the sickle-cell trait in East Africa and elsewhere, and
397 its apparent relationship to the incidence of subtertian malaria. Transactions of the Royal
398 Society of Tropical Medicine and Hygiene, 48, 312-318. [https://doi.org/10.1016/0035-](https://doi.org/10.1016/0035-9203(54)90101-7)
399 [9203\(54\)90101-7](https://doi.org/10.1016/0035-9203(54)90101-7).

400 Bowman, J. (1997). The management of hemolytic disease in the fetus and newborn.

401 Seminars in perinatology, 21, 39-44. [https://doi.org/10.1016/S0146-0005\(97\)80018-3](https://doi.org/10.1016/S0146-0005(97)80018-3).

402 Christiansen, O., Pedersen, B., Nielsen, H., & Andersen, A. (2004). Impact of the sex of first
403 child on the prognosis in secondary recurrent miscarriage. Human Reproduction, 19, 2946-
404 2951. <https://doi.org/10.1093/humrep/deh516>.

405 Davis, D., Gottlieb, M., & Stampnitzky, J. (1998). Reduced ratio of male to female births in
406 several industrial countries - A sentinel health indicator? The Journal of the American
407 Medical Association, 279, 1018-1023. <https://doi.org/10.1001/jama.279.13.1018>.

408 Feldman, M., Nanholtz, M., & Bodmer, W. (1969). Evolution of Rh polymorphism - a model
409 for interaction of incompatibility reproductive compensation and heterozygote advantage.
410 American Journal of Human Genetics, 21, 171-193.

411 Filbey, D., Hanson, U., & Wesstrom, G. (1995). The prevalence of red-cell antibodies in
412 pregnancy correlated to the outcome of the newborn - a 12 year study in central Sweden. Acta

- 413 *Obstetricia et Gynecologica Scandinavica*, 74, 687-692.
- 414 <https://doi.org/10.3109/00016349509021175>.
- 415 Fincham, J. R. (1972). Heterozygous advantage as a likely general basis for enzyme
- 416 polymorphisms. *Heredity*, 28, 387-391. <https://doi.org/10.1038/hdy.1972.49>.
- 417 Fisher, R. A. (1930). *The genetical theory of natural selection*. Oxford: Clarendon press.
- 418 Flegel, W. (2011). Molecular genetics and clinical applications for RH. *Transfusion and*
- 419 *Apheresis Science*, 44, 81-91. <https://doi.org/10.1016/j.transci.2010.12.013>.
- 420 Flegr, J. (2016). Heterozygote Advantage Probably Maintains Rhesus Factor Blood Group
- 421 Polymorphism: Ecological Regression Study. *Plos One*, 11, 1-12.
- 422 <https://doi.org/10.1371/journal.pone.0147955>.
- 423 Flegr, J., Novotna, M., Lindova, J., & Havlicek, J. (2008). Neurophysiological effect of the
- 424 Rh factor. Protective role of the RhD molecule against *Toxoplasma*-induced impairment of
- 425 reaction times in women. *Neuro Endocrinology Letters*, 29, 475-481.
- 426 Flegr, J., Klose, J., Novotna, M., Berenreitterova, M., & Havlicek, J. (2009). Increased
- 427 incidence of traffic accidents in *Toxoplasma*-infected military drivers and protective effect
- 428 RhD molecule revealed by a large-scale prospective cohort study. *BMC Infectious Diseases*,
- 429 9, 1-7. <https://doi.org/10.1186/1471-2334-9-72>.
- 430 Flegr, J., Novotná, M., Fialová, A., Kolbeková, P., & Gasová, Z. (2010). The influence of
- 431 RhD phenotype on toxoplasmosis- and age-associated changes in personality profile of blood
- 432 donors. *Folia Parasitologica*, 57, 143-150. <https://doi.org/10.14411/fp.2010.018>.

- 433 Flegr, J., Geryk, J., Volný, J., Klose, J., & Cernochová, D. (2012). Rhesus factor modulation
434 of effects of smoking and age on psychomotor performance, intelligence, personality profile,
435 and health in Czech soldiers. *Plos One*, 7, 1-10. <https://doi.org/10.1371/journal.pone.0049478>.
- 436 Flegr, J., Preiss, M., & Klose, J. (2013). Toxoplasmosis-Associated Difference in Intelligence
437 and Personality in Men Depends on Their Rhesus Blood Group but Not ABO Blood Group.
438 *Plos One*, 8, 1-8. <https://doi.org/10.1371/journal.pone.0061272>.
- 439 Flegr, J., Hoffmann, R., & Dammann, M. (2015). Worse Health Status and Higher Incidence
440 of Health Disorders in Rhesus Negative Subjects. *Plos One*, 10, 1-14.
441 <https://doi.org/10.1371/journal.pone.0141362>.
- 442 Flegr, J., & Horacek, J. (2017). Toxoplasma-infected subjects report an Obsessive-
443 Compulsive Disorder diagnosis more often and score higher in Obsessive-Compulsive
444 Inventory. *European Psychiatry*, 40, 82-87. <https://doi.org/10.1016/j.eurpsy.2016.09.001>.
- 445 Flegr, J., Sebankova, B., Priplatova, L., Chvatalova, V., & Kankova, S. (2018). Lower
446 performance of Toxoplasma-infected, Rh-negative subjects in the weight holding and hand-
447 grip tests. *Plos One*, 13, 1-12. <https://doi.org/10.1371/journal.pone.0200346>.
- 448 Gavrilova, N., Gavrilov, L., Evdokushkina, G., Semyonova, V., Gavrilova, A., Evdokushkina,
449 N., Kushnareva, Y., Kroutko, V., & Andreyev, A. (1998). Evolution, mutations, and human
450 longevity: European royal and noble families. *Human Biology*, 70, 799-804.
- 451 Golassa, L., Tsegaye, A., Erko, B., & Mamo, H. (2017). High rhesus (Rh(D)) negative
452 frequency and ethnic-group based ABO blood group distribution in Ethiopia. *BMC Research*
453 *Notes*, 10, 1-5. <https://doi.org/10.1186/s13104-017-2644-3>.

454 Gualtieri, C., Hicks, R., & Mayo, J. (1984). Influence of sex ratio antecedent siblings on the
455 human sex-ratio. *Life Sciences*, 34, 1791-1794. [https://doi.org/10.1016/0024-3205\(84\)90670-](https://doi.org/10.1016/0024-3205(84)90670-2)
456 2.

457 James, W. (1996). Evidence that mammalian sex ratios at birth are partially controlled by
458 parental hormone levels at the time of conception. *Journal of Theoretical Biology*, 180, 271-
459 286. <https://doi.org/10.1006/jtbi.1996.0102>.

460 James, W. (2010). Behavioural and biological determinants of human sex ratio at birth.
461 *Journal of Biosocial Science*, 42, 587-599. <https://doi.org/10.1017/S002193201000012X>.

462 James, W. (2015). Hypothesis: High levels of maternal adrenal androgens are a major cause
463 of miscarriage and other forms of reproductive suboptimality. *Journal of Theoretical Biology*,
464 364, 316-320. <https://doi.org/10.1016/j.jtbi.2014.09.027>.

465 James, W., & Grech, V. (2018). Offspring sex ratio: Coital rates and other potential causal
466 mechanisms. *Early Human Development*, 116, 24-27.
467 <https://doi.org/10.1016/j.earlhumdev.2017.10.006>.

468 Kanazawa, S. (2005). Big and tall parents have more sons: Further generalizations of the
469 Trivers-Willard hypothesis. *Journal of Theoretical Biology*, 235, 583-590.
470 <https://doi.org/10.1016/j.jtbi.2005.02.010>.

471 Kankova, S., Sulc, J., Nouzova, K., Fajfrlik, K., Frynta, D., & Flegr, J. (2007). Women
472 infected with parasite *Toxoplasma* have more sons. *Naturwissenschaften*, 94, 122-127.
473 <https://doi.org/10.1007/s00114-006-0166-2>.

- 474 Kankova, S., Sulc, J., & Flegr, J. (2010). Increased pregnancy weight gain in women with
475 latent toxoplasmosis and RhD-positivity protection against this effect. *Parasitology*, 137,
476 1773-1779. <https://doi.org/10.1017/S0031182010000661>.
- 477 Kustu, S., & Inwood, W. (2006). Biological gas channels for NH₃ and CO₂: evidence that Rh
478 (Rhesus) proteins are CO₂ channels. *Transfusion Clinique et Biologique*, 13, 103-110.
479 <https://doi.org/10.1016/j.tracli.2006.03.001>.
- 480 Le Van Kim, C., Colin, Y., & Cartron, J. (2006). Rh proteins: Key structural and functional
481 components of the red cell membrane. *Blood Reviews*, 20, 93-110.
482 <https://doi.org/10.1016/j.blre.2005.04.002>.
- 483 Maraz, A., Impre, G., Keseru, T. L., Szabo E., & Szontaght, F. E. (1973). Sex-ratio, blood-
484 groups and parity. *Folia Biologica*, 19, 174-178.
- 485 McGann P, Despotovic J, Howard T, & Ware R. (2012). A novel laboratory technique
486 demonstrating the influences of RHD zygosity and the RhCcEe phenotype on erythrocyte D
487 antigen expression. *American Journal of Hematology*, 87, 266-271.
488 <https://doi.org/10.1002/ajh.22254>.
- 489 Medawar, P. B. (1946). Old age and natural death. *Modern Quarterly*, 1, 30-56.
- 490 Mourant, A. K, Kopeć A. C., & Domaniewska-Sobczak, K. (1976). *The Distribution of the*
491 *Human Blood Groups and Other Polymorphisms*. London: Oxford University Press.
- 492 Nei, M., Li, W., Tajima, F., & Narain, P. (1981). Polymorphism and evolution of the Rh
493 blood-groups. *Journal of Human Genetics*, 26, 263-278. <https://doi.org/10.1007/BF01876357>.
- 494 Nettle, D. (2010). Dying young and living fast: variation in life history across English
495 neighborhoods. *Behavioral Ecology*, 21, 387-395. <https://doi.org/10.1093/beheco/arp202>.

- 496 Nielsen, H., Andersen, A., Kolte, A., & Christiansen, O. (2008). A firstborn boy is suggestive
497 of a strong prognostic factor in secondary recurrent miscarriage: a confirmatory study.
498 *Fertility and Sterility*, 89, 907-911. <https://doi.org/10.1016/j.fertnstert.2007.04.029>.
- 499 Nielsen, H., Steffensen, R., Varming, K., Van Halteren, A., Spierings, E., Ryder, L., Goulmy,
500 E., & Christiansen, O. (2009). Association of HY-restricting HLA class II alleles with
501 pregnancy outcome in patients with recurrent miscarriage subsequent to a firstborn boy.
502 *Human Molecular Genetics*, 18, 1684-1691. <https://doi.org/10.1093/hmg/ddp077>.
- 503 Novotna, M., Havlicek, J., Smith, A., Kolbekova, P., Skalova, A., Klose, J., Gasova, Z.,
504 Pisacka, M., Sechovska, M., & Flegr, J. (2008). *Toxoplasma* and reaction time: role of
505 toxoplasmosis in the origin, preservation and geographical distribution of Rh blood group
506 polymorphism. *Parasitology*, 135, 1253-1261. <https://doi.org/10.1017/S003118200800485X>.
- 507 Pedersen, B. (1995). An evolutionary-theory of clonal senescence. *Theoretical Population*
508 *Biology*, 47, 292-320. <https://doi.org/10.1006/tpbi.1995.1013>.
- 509 Promislow, D. E. L., & Harvey, P. H. (1990). Living fast and dying young - a comparative-
510 analysis of life-history variation among mammals. *Journal of Zoology*, 220, 417-437.
511 <https://doi.org/10.1111/j.1469-7998.1990.tb04316.x>.
- 512 Renkonen, K., Mäkelä, O., & Lehtovaara, R. (1962). Factors affecting human sex ratio.
513 *Nature*, 194, 308-309. <https://doi.org/10.1038/194308b0>.
- 514 Renkonen, K., & Seppala, M. (1962). Sex of immunizing Rh-positive child. *Annales*
515 *Medicinae Experimentalis et Biologiae Fenniae*, 40, 108-109.
- 516 Trivers, R., & Willard, D. (1973). Natural-selection of parental ability to vary sex ratio of
517 offspring. *Science*, 179, 90-92. <http://dx.doi.org/10.1126/science.179.4068.90>.

518 Wagner, F., & Flegel, W. (2000). RHD gene deletion occurred in the Rhesus box. *Blood*, 95,
519 3662-3668.

520 WHO. (2008). *The Global Burden of Disease: 2004 update*. World Health Organization,
521 Geneva, Switzerland.

522 https://www.who.int/healthinfo/global_burden_disease/2004_report_update/en/ Accessed 5
523 February 2019.

524 Wilson, K., & Hardy, I. (2002). Statistical analysis of sex ratios: an introduction. In I. Hardy
525 (Ed.), *Sex Ratios: Concepts and Research Methods* (pp. 287–311). Cambridge: Cambridge
526 University Press.