Thermal regimes, but not mean temperatures, drive patterns of rapid

climate adaptation at a continent-scale: evidence from the introduced

European earwig across North America

Running title: Climate adaptation in earwigs

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ABSTRACT

The recent development of human societies has led to major, rapid and often inexorable

changes in the environment of most animal species. Over the last decades, a growing number

of studies formulated predictions on the modalities of animal adaptation to climate change,

questioning how and at what speed animals should adapt to such changes, discussing the levels

of risks imposed by changes in the mean and/or variance of temperatures on animal

performance, and exploring the underlying roles of phenotypic plasticity and genetic

inheritance in this adaption. These fundamental predictions, however, remain poorly tested

using field data. Here, we tested these predictions using a unique continental-scale data set in

the European earwig *Forficula auricularia* L, a univoltine insect introduced in North America

one century ago. We conducted a common garden experiment, in which we measured 13 life-

history traits in 4158 field-sampled earwigs originating from 19 populations across North

America. Our results first demonstrate that in less than 100 generations, this species modified

10 of the 13 measured life-history traits in response to the encountered thermal regimes, defined

as a variation of temperatures between seasons or months (here winter-summer and autumn-

spring temperatures). We found, however, no response to the overall mean monthly

temperatures of the invaded locations. Our use of a common garden setup also suggests that

the observed changes in earwigs' life-history traits first emerged as a plastic response to the

thermal constraints of the different localities, then diverged between populations through

canalization, and ultimately became inherited traits. Overall, these findings provide continent-

scale support to the claims that adaptation to thermal changes occurs quickly, even in insects

with long life cycles, and emphasize the importance of thermal regimes over mean population

temperatures in climate adaptation.

Keywords: Temperature, Adaptation, Reproductive strategy, Climate change, Invasion,

Dermaptera

INTRODUCTION

The Earth's climate has changed several times throughout history, leading to gradual

modifications of both its natural environments and multiple traits of its resident organisms,

such as their reproduction, survival, abundance and geographic range (Parmesan 2006, Merilä

and Hendry 2014). The dramatic acceleration of climate change observed over the last decades,

however, challenges the ability of resident organisms to track these changes and adapt their life

histories accordingly (Meehl and Tebaldi 2004, Parmesan 2006, Williams et al. 2007), raising

major issues in terms of biodiversity, conservation, ecosystems functioning and animal

evolution (Bellard et al. 2012, Travis et al. 2013).

Over the last decades, modelling and theoretical approaches have been developed to

better understand the nature and extent of animals' response to such a climate change

(Parmesan 2006). These studies formulated key predictions on how and at what speed animals

should adapt to such changes, on the respective importance of an increase in the overall mean

temperature and/or seasonality of a population on animal performance, as well as on the

underlying roles of phenotypic plasticity and genetic inheritance in adaption (Nylin and

Gotthard 1998, Kingsolver et al. 2013, Paaijmans et al. 2013, Gilbert et al. 2014, Merilä and

Hendry 2014, Levis and Pfennig 2016, Williams et al. 2017, Corl et al. 2018, Fox et al. 2019,

Rohner et al. 2019). For instance, these studies suggest that a rapid adaptation to climate change

should be facilitated in organisms with fast development and short life-cycles, as found in many

arthropods, whereas it should be more difficult in organisms exhibiting slow development and

long life-cycle, as found in many vertebrates. Species should also be less sensitive to changes

in seasonality compared to changes in overall mean temperatures when there are endotherms

and/or when their entire life-cycle occur within a single season, whereas the opposite pattern

is expected when they are ectotherms and/or have a life-cycle encompassing several seasons.

Finally, phenotypic plasticity is often considered a keystone of rapid adaptation to

environmental changes, whereas fixed and inherited patterns of adaptation are often thought to

secondarily derive from the canalization of ancestral plastic variation.

Although central in our current understanding of animal's responses to climate change,

these fundamental predictions remain poorly tested in the field (Janion-Scheepers et al. 2017,

Blanckenhorn et al. 2018). This is probably because such field data are difficult to collect, as

it typically requires measuring variation in life-history traits across multiple natural

populations, over several years, and under different levels of climate change. However, a

powerful alternative consists in using field data of introduced species that quickly invaded large

geographic areas exhibiting a broad diversity of thermal constraints (Huey et al. 2000, Bellard

et al. 2016). In this study, we present and analyze such a unique field data set in one of these

species, the European earwig *Forficula auricularia* Linnaeus (Dermaptera: Forficulidae), after

its introduction in North America. This insect exhibits a broad native range extending across

Europe, Asia and northern Africa (Lamb and Wellington 1975) from which it has been

introduced to Australia, New Zealand, East Africa, East Indies, Tasmania and America (Frank

1918, Guillet et al. 2000, Quarrell et al. 2018, Hill et al. 2019). Its presence in North America

was first reported on the Pacific coast in Seattle (WA) in 1907 (Fulton 1924), and then on the

Atlantic coast in Newport (RI) in 1911 (Jones 1917) and in Vancouver (BC) in 1919 (Treherne

1923). From these introductory foci, *F. auricularia* first spread along the coasts to cover areas

ranging from British Columbia to California and from Newfoundland to South Carolina, and

then reached the interior of the continent in both United States of America (Crumb et al. 1941)

and Canada (Wilson 1971, Cantrall 1972, Tourneur 2017). Given that this species produces

only one generation per year (Tourneur and Gingras 1992, Meunier et al. 2012), these historical

records reveal that its successful colonization of North America and thus its adaptation to a

broad diversity of thermal environments occurred in less than 100 generations.

Because the univoltine life cycle of the European earwig lasts up to 2 years and

encompasses all seasons and temperatures (Lamb 1976, Meunier et al. 2012), it has long been

thought that annual mean temperatures and/or temperature seasonality could be major

constraints in the success of *F. auricularia* invasions (Vancassel 1984, Hill et al. 2019).

However, it remains unclear whether this species can mitigate these thermal constraints, and

whether it does so by adapting its life cycle and life-history traits (Ratz et al. 2016, Tourneur

2018). The life cycle of the European earwig generally starts with the emergence of new adults

in late spring to early July, with variation among populations. These adults form groups of up

to several hundred individuals, in which both males and females typically mate with several

partners (Weiß et al. 2014, Sandrin et al. 2015, Tourneur 2017). Females then burrow in the

ground from mid fall to early winter and build a nest where they lay their first clutch of eggs.

After egg laying, females stop their foraging activity and provide extensive forms of egg care

until hatching (Gingras and Tourneur 2001, Boos et al. 2014, Koch and Meunier 2014, Thesing

et al. 2015, Diehl and Meunier 2018, Körner et al. 2018). The eggs of this first clutch hatch in

spring and mothers remain with their newly hatched larvae for several weeks, during which

mothers provide larvae with multiple forms of care (Gingras and Tourneur 2001, Kölliker et

al. 2015, Kramer et al. 2015) and larvae exhibit forms of sibling cooperation (Falk et al. 2014,

Kramer et al. 2015, Kramer and Meunier 2016, Körner et al. 2016). A few weeks later, the

family unit is naturally disrupted. While larvae continue their development to adults in new

social groups, some females produce a second clutch of eggs (i.e. iteroparous as compared to

semelparous females), which will also receive pre- and post-hatching care and will hatch in

late spring (Lamb and Wellington 1975, Meunier et al. 2012, Ratz et al. 2016). All females

generally die during the following summer (Albouy and Caussanel 1990).

In this study, we used a common garden experiment to explore how *F. auricularia*

responded to the different thermal environments encountered during their North American

invasion over the last century, i.e. in less than 100 generations. In particular, we 1) tested

whether and how individuals altered their life history traits in response to the thermal

constraints of the invaded locations, 2) identified to which thermal constraints they adapted to

and 3) investigated the role of phenotypic plasticity in this adaptation. From 1988 to 1995, we

field-sampled individuals originating from 19 populations located from the East to the West

coasts, maintained them under standard laboratory conditions and measured the properties of

the 1st and 2nd clutches produced by each female in terms of egg laying date, egg number, egg

development time and number of newly hatched larvae. We also recorded the reproductive

strategy of the females (iteroparity versus semelparity), their reproductive outcome (total

number of eggs and larvae produced over lifetime), as well as the experimental survival

duration of the field-sampled males and females. To identify which thermal constraints the

tested earwigs adapted to, we tested whether our measurements could be explained by the

results of a principal component analyses (PCA) of the mean monthly temperatures of each

population. This process characterizes patterns of variation among populations' temperatures

without *a priori* definitions of their associations, i.e. without predetermining the focus on

overall mean temperatures and/or specific thermal regimes (defined as variation of

temperatures between seasons or months). If *F. auricularia* individuals adapted their life-cycle

and life-history traits to the mean temperatures and/or thermal regimes of the population in

which they have been sampled (and if this adaptation is determined by their genetic background

and/or early life experience), we predict these traits to covary with the overall mean

temperatures and/or variation in seasonal temperatures of their population (i.e. all sampled

populations should show different performance in the common garden). Conversely, if earwig

life-history traits are independent of the thermal environment of the population in which they

have been sampled (i.e. no adaptation) and/or are plastic to their current thermal environment,

we predict no apparent association between the traits measured in our field-sampled individuals

and the thermal regimes of their populations (i.e. all sampled populations should show similar

performance in the common garden).

MATERIAL AND METHODS

Earwig sampling and laboratory rearing

All *F. auricularia* individuals were collected over 7 years among 19 natural populations located

across North America (Figure 1, Table 1). These individuals were mostly collected as adults

using wooden traps (Tourneur 2018) between July and August, and were immediately setup in

glass containers (Mason Jars Company, Erie, Pennsylvania, United States of America) in

groups of 20 to 30 individuals. These containers received two sheets of creased toilet paper as

resting places for earwigs, and were then transported to our laboratory in Montreal, Canada.

Upon their arrival, containers were deposited in a shelf covered by a shelter and maintained

under the natural outdoor conditions of Montreal. During their transport and outdoor

maintenance, containers received an *ad libitum* amount of carrots and pollen as a food source

for earwigs, and were supplied with water by means of a cotton pad regularly soaked in water.

This setup allowed earwigs to continue their development, while performing non-controlled

mating and expressing social interactions (Weiß et al. 2014, Sandrin et al. 2015, Kohlmeier et

al. 2016, Körner et al. 2018).

One to two months later (between the 7th and the 19th day of October of each year), we

used 4158 of these field-sampled individuals to set up 2079 mating pairs (from 17 to 356 pairs

per population, see Table 1), in which we subsequently measured 13 life-history traits (see

below). These pairs were set up in Petri dishes (diameter 10 cm) lined with a thin layer of moist

sand, and in which food was changed and substrate humidified once a week. Each Petri dish

was then transferred in a climate chamber and then maintained at 10 ± 1 °C, a temperature

close to the overall median temperature of the 19 sampled populations (i.e. 9.5°C, see Table

S1). Food was removed at egg laying to mimic the natural end of earwigs' foraging activity

(Kölliker 2007). At egg hatching, we discarded all newly emerged larvae from the experiments

to trigger a novel ovarian cycle in the mothers and allow their production of a subsequent clutch

(Vancassel and Foraste 1980, Meunier et al. 2012). We then maintained the pairs under the

rearing conditions described above until our experiment ended, i.e. either one year after the

beginning of our laboratory setup or at the death of the adult males and females. Overall, 3927

of the 4158 (94.4%) tested individuals died within the year following the beginning of our

experiments, a value in line with previous data on *F. auricularia* lifespan (Albouy and

Caussanel 1990). Note that recent studies revealed that North American *F. auricularia* L.

encompasses two genetic subspecies with no apparent mixing of their populations (Wirth et al.

1998, Quarrell et al. 2018, Tourneur 2018). Although these subspecies were not considered in

our analyses (our data were collected before the publication of these genetic analyses), the

continuous distribution (unimodal data) of the life history traits measured across populations

(Figures 2 to 4) suggests an absence of species-specific values regarding these measurements.

The potential co-occurrence of the two subspecies in our data set is thus unlikely to bias our

study and its main conclusions.

Measurements of the life-history traits

For each mating pair, we measured 13 life-history traits encompassing the properties of the

resulting 1st and 2nd clutches (when present), the reproductive strategy and reproductive

outcomes of each female, as well as the experimental survival duration of both field-sampled

males and females. These properties were obtained by recording the date of egg production,

counting the number of eggs produced, calculating the duration of egg development until

hatching (in days) and finally counting the number of larvae at egg hatching in both 1st and 2nd

clutches (when present). The reproductive strategies and reproductive outcomes of females

were obtained by recording whether they were semelparous or iteroparous (i.e. produced one

or two clutches in their lifetime, respectively), and by counting the total number of eggs and

larvae produced per female during their lifetime. Finally, we measured the experimental

survival duration of adult's by counting the number of days each male and female survived

after October 1st of the year of field sampling. Although our measurement of survival duration

does not necessarily reflect adults' longevity, as individuals could have different age at field

sampling (see discussion), it nevertheless provides important insights into the period at which

males and females of each population die during the year. Note that 8.1% and 5.4% females

from Santa Cruz and Asheville, respectively, produced a third clutch. This third clutch was not

considered in the present study, as our experiment ended before their hatching.

Extraction of mean temperatures and thermal regimes of each population

We extracted the mean monthly temperature of the 19 studied populations using their GPS

coordinates (Table 1) and the Worldclim database v2.0 (http://www.worldclim.org/) with a

spatial resolution of 30 seconds. The mean temperatures provided by the Worldclim database

are calculated over 30 years, from 1970 to 2000. To reduce dimensionality of co-varying

temperatures in our data set while characterizing potential thermal regimes of each population

without *a priori* definitions of their composition, we then conducted a Principal Component

Analysis (PCA) on the set of 12 mean monthly temperatures per population (Table S1). This

analysis provided us with 12 orthogonal principal components (PCs), out of which we retained

the first three PCs (total variance explained = 98.6%, Table 2). The first components (PC1)

was positively loaded by almost all monthly temperatures, therefore positively reflecting the

overall mean temperature of a population. The second component (PC2) revealed variation in

seasonality between February on one hand, and June, July, and August on the other hand. In

particular, high values of PC2 reflected populations with cold February (winter) and warm

summer, whereas small values of PC2 reflected populations with warm February (winter) and

cold summer. Finally, the third component (PC3) captured variation in seasonality between

October and November on one hand, and April and May on the other hand. High values of PC3

therefore characterized populations with cold autumn and warm spring, whereas small values

of PC3 reflected populations with warm autumn and cold spring.

Statistical analyses

To test whether *F. auricularia* adapt their life-cycle and life-history traits to North American

temperatures, we conducted a series of 12 linear models (LM in R) and one generalized linear

model (GLM in R) – see Table 3. In the 12 LMs, the three selected PCs and their interactions

were entered as explanatory variables (PC1, PC2 and PC3), whereas the response variable was

either egg laying date, egg number, egg development time and larvae number for the 1st or 2nd

clutches (a total of 8 LMs), the total number of eggs or larvae produced, or the survival duration

of males or females. Note that both egg laying date and adult's survival duration were

calculated using October 1st as day 0. In the GLM, the response variable was the ratio of

iteroparous females per population, which was entered using the command *cbind* in R (to

weight each ratio by the sample size of its population) and fitted to a binomial error distribution

corrected for overdispersion. In all our statistical models, the response variables were the mean

values of each measured trait per population. They were also checked for homoscedasticity and

normality of residuals, as well as simplified stepwise by removing all non-significant

interaction terms (all $P > 0.05$). To correct for inflated Type-I errors due to multiple testing

(and provide an experiment-wide Type I error rate of 5%), all *P*-values were adjusted using

False Discovery Rate (FDR) correction (Benjamini and Hochberg 1995). All analyses were

conducted using the software R v3.5.1 loaded with the packages *raster*, *FactoMineR*, *rsq* and

rcompanion.

RESULTS

The 19 studied populations greatly varied in their mean temperatures and thermal regimes

(Table S1), as well as in the mean values of the 13 traits measured in their sampled individuals

(Figures 2 to 4; Tables S2 to S4). Mean monthly temperatures overall ranged from 22.9°C (July

in Saluda) to -10.1°C (January in Montreal), while thermal amplitudes over a year ranged from

30.7°C (Montreal) to 7.9°C (Santa Cruz). For the traits measured in the 1st clutches, the mean

dates of egg production ranged from 47.8 to 132.6 days after the 1st of October, the mean

number of eggs per clutch from 23.2 to 66.0, the mean egg development time from 42.2 to 71.4

days and the mean number of larvae per clutch from 11.6 to 44.8. For the 2nd clutches, the mean

dates of egg production ranged from 142.0 to 248.2 days after the 1st of October, the mean

number of eggs from 14.0 to 38.4, the mean egg development time from 10.0 to 63.7 days and

the mean number of larvae from 0 to 17.7. Finally, the total number of eggs produced ranged

from 28.1 to 83.4, the total number of larvae produced from 13.0 to 46.3, the proportion of

iteroparous females from 0 to 70.8%, the survival duration of males from 82.0 to 299.8 days

after the 1st of October and the survival duration of females from 146.0 to 322.5 days after the

1st of October.

Of the 13 measured traits, 10 varied together with the thermal regimes of the population

of origin (Table 3). Five of these 10 traits were exclusively associated with PC2 (February-

summer temperatures), two traits were exclusively associated with PC3 (autumn-spring

temperatures), and three traits were associated with both PC2 and PC3. By contrast, no traits

were associated with PC1 (overall mean temperatures). The associations with PC2 revealed

that populations with cold February and warm summers (high PC2 values) had females that

produced their 1st clutch of eggs earlier and these eggs had longer development time compared

to populations exhibiting warm February and cold summers (low PC2 values, Figure 2).

Similarly, females from the former populations were less likely to produce a second clutch (i.e.

to be iteroparous, Figure 3) and when they did so, their 2nd clutches eggs were less numerous

(Figure 3) and showed longer development time (Figure 3). Moreover, females and males from

populations with cold February and warm summers lived less long compared to adults from

warm February and cold summers (Figure 4). On the other hand, the effects of PC3 reveal that

populations exhibiting cold autumn and warm spring (high PC3 values) had females that

produced their 1st clutch of eggs later in the season and these eggs were less numerous

compared to females from populations with warm autumn and cold spring (low PC3 values,

Figure 2). Females from the former populations also had 2nd clutch eggs that exhibited a shorter

developmental time (Figure 3), they produced an overall lower number of eggs (Figure 4) and

had males with a longer survival duration (Figure 4). By contrast, PC1, PC2 and PC3 did not

shape the number of 1st clutch larvae, as well as their total number and the dates of 2nd clutch

egg laying (Figures 2, 3 and 4; Table 3).

DISCUSSION

Shedding light on how species successfully adapt to a broad set of environmental constraints

is of major importance to improve our general understanding of the mechanisms underlying

animal adaptations to climate change. In this study, we demonstrate that the successful invasion

of the European earwig across North America came with multiple changes in their life-history

traits in response to the thermal regimes (sets of winter-summer and autumn-spring

temperatures), but not to the overall mean temperature of the invaded areas. In particular, our

data from 19 populations revealed that females changed their timing of first reproduction, their

reproductive strategy and investment into egg production when facing different thermal

regimes, while both males and females experimental survival duration varied accordingly. By

contrast, we found no association between thermal regimes and both the timing of second

reproduction and the total number of larvae produced per female.

277   We showed that females produced their first clutch of eggs earlier when they came from
278 populations facing warm summers and/or warm autumns (PC2 and PC3, respectively), and
279 were less likely to produce a second clutch in populations with cold February. A plastic
280 response to warm temperatures on egg laying date would not be surprising: adult earwigs
281 typically develop and mate during summer and autumn, so that warm temperatures during these
282 seasons could accelerate their reproductive physiology (as shown in other insect species, Singh
283 et al., 2018) and thus accelerate egg laying (Tourneur 2018). Similarly, cold Februaries might
284 slow down the development of 1st clutch eggs and thus extend the corresponding period of egg
285 care. This, in turn, might inhibit females' physiological transformation to produce a second
286 clutch (Vancassel 1984, Tourneur 2018). Such an apparent plasticity is in line with previous
287 results showing that cold winters increase the duration of egg development time in *F.*
288 *auricularia* (Gingras and Tourneur 2001, Körner et al. 2018). It is also in line with other results
289 demonstrating that tending eggs inhibits vitellogenic activity and the production of juvenile
290 hormone in the earwig *Labidura riparia* Pallas, two parameters required to activate ovary
291 development and allow egg production (Vancassel et al. 1984). However, our results were
292 obtained under common garden conditions, revealing that the observed effects of thermal
293 regime on egg laying dates are not the outcome of plasticity, but are either due to canalization
294 (i.e. the same phenotype results regardless of environmental variation) or to the effect of early
295 life environments faced before field sampling (Nylin and Gotthard 1998). It has been suggested
296 that traits tightly linked to fitness are more strongly canalized due to past stabilizing selection
297 (Falconer 1990). Our findings therefore suggest that the observed changes in the timing of first
298 reproduction and females' reproductive strategy may have first emerged as a plastic response
299 to the thermal constraints of the different localities, then diverged between populations through
300 canalization to ultimately become inherited traits – all this in a maximum of 100 generations.

Our data also reveals that thermal regimes are associated with lifetime egg production,

but not with lifetime larvae production. In particular, the total number of eggs produced per

female decreased with decreasing autumn temperatures, whereas this association vanished with

larvae number. This apparent discrepancy suggests that females from populations with the

warmest autumns lost a larger number of eggs during egg development. A first explanation

could be that these females produced eggs of lower quality and/or were less efficient in egg

care, a process that is essential to ensure egg development until hatching in earwigs (Boos et

al. 2014, Van Meyel et al. 2019). Whereas both effects should be tested in future studies,

previous results may suggest that the second effect is unlikely, as maternal investment in post-

hatching care is not population-specific, at least in Europe (Ratz et al. 2016). Another

explanation is that females consumed a larger part of their clutch in populations with the

warmest compared to the coldest autumns. Filial egg consumption is a common phenomenon

in insects (Elgar and Crespi 1992) and it has been recently reported in several Dermapteran

species, such as the European earwig *F. auricularia* (Koch and Meunier 2014, Van Meyel et

al. 2019) and the maritime earwig *Anisolabis maritima* Bonelli (Miller and Zink 2012). In the

European earwig, this phenomenon has been proposed to reflect an adaptive strategy to limit

female weight loss during the period of egg care (i.e. when they stop all other foraging

activities) and by doing so, to reallocate resources into post-hatching care and/or into a 2nd

oogenesis cycle (Koch and Meunier 2014, Tourneur 2018). Given that females lay eggs earlier

in populations with the warmest autumns, this increased egg consumption could be an adaptive

strategy to limit the cost of tending newly hatched offspring earlier in the season (middle of

winter) when food sources are scarce or absent. If this hypothesis holds true, it would suggest

that filial egg cannibalism could be a strategy that *F. auricularia* females have evolved to better

cope with warmer autumns.

Furthermore, we found that the survival duration of both males and females were

associated with the thermal regime of the population of origin. In particular, female's and

male's survival duration decreased together with warm summers (and cold Februaries), while

male's survival duration also decreased with warm autumns (and cold springs). The first results

may be a by-product of the effect of temperature on their date of egg laying and/or egg hatching.

In particular, we showed that females from populations facing warm summers are the first to

lay their eggs. Individuals from these populations might thus have been the oldest at the date

of our field sampling, therefore leading to the shortest survival duration in our subsequent

experiment. Surprisingly, there was a sex-specific effect of spring (and autumn) temperatures

on adult's survival duration: males lived up to two times longer in populations with warm

compared to cold springs (as well as cold compared to warm autumns), whereas this effect was

absent in females. This finding may reflect sex-specific sensitivity to high temperatures in

terms of, for instance, physiology or expression of costly behaviors. Whereas some

physiological traits are known to be sex-specific in this species (Kohlmeier et al. 2016,

Vogelweith et al. 2017), further studies should explore the effects of temperature on the

observed differences. Notwithstanding its underlying mechanisms, the long survival duration

of males in warm spring populations opens scope for these males to mate with females of the

subsequent generation, as well as for a possible involvement of fathers into larva care – a

phenomenon reported in other insect species (Smiseth 2014). These two processes remain

unknown in this species, but they could be of central importance in their successful adaptation

to climate change. Hence, follow-up studies should investigate the (re)mating strategies of

earwig males, as well as the expression of paternal care across these populations.

Common garden experiments are powerful tools to disentangle the effects of

phenotypic plasticity and genetic background on adaptation (Franks et al. 2014, Stoks et al.

2014, Blanckenhorn et al. 2018). Individuals reared under a common environment are expected

to exhibit homogenized life-history traits if adaptation is the outcome of phenotypic plasticity, whereas they should exhibit population-specific traits otherwise. Our results are in line with the latter process for the great majority of the measured life-history traits (10 out of 13), therefore suggesting that the observed associations between thermal regimes and life-history traits do not stem from phenotypic plasticity. Further studies are now needed to disentangle whether the observed associations are the outcome of earwigs' genetic background or early-life experience. Our tested individuals were mostly collected as adults in the field, so that we cannot rule out that population-specific constraints occurring during their early developmental stages have had long-lasting effects on their life-history traits (English et al. 2016). In line with this hypothesis, food restriction and poor food quality during juvenile stage has been shown to have negative effects on females' body weight and egg production in *F. auricularia* (Berleur et al. 2001, Wong and Kölliker 2014). Future studies using field-collected eggs and/or F1 generation individuals should help disentangling among these two processes. Common garden experiments also do not preclude the possibility of genotype-by-environment interactions on the measured life-history traits, which can in turn affect the strength and nature of the reported patterns of adaptation (Merilä and Hendry 2014). Because our common garden experiment was set up under a single temperature, future studies may repeat our protocol with multiple temperatures to test the impact of rearing conditions on the strength and nature of the observed associations between life-history traits and populations' thermal regimes in *F. auricularia*.

To conclude, our results demonstrate that the spread of the European earwigs across North America came with important changes in their life-history traits and life cycle, and that these changes emerged in a maximum of 100 generations. Whereas we show that some of these changes are by-products of novel thermal constraints (timing of first reproduction and female iteroparity), we reveal that others are likely to reflect adaptive strategies to cope with different autumn temperatures (egg production and the possibility of egg cannibalism). Overall, these

findings emphasize that adaptation of an insect with a relatively long life-cycle does not

necessarily operate in response to the overall mean temperatures of the invaded environments,

but to their thermal regimes – i.e. to seasonality and/or mean temperature at specific time of

their life-cycle. Whether the reported adaptations are the product of population-differences in

energetic/metabolic constraints experienced by adults during their early development (Wong

and Kölliker 2014, English et al. 2016), and/or the product of an inherited genetic basis that

varies with thermal regimes (Levis and Pfennig 2016; Corl et al. 2018; Fox et al. 2019), as well

as whether these adaptations are similar across its worldwide distribution (Frank 1918, Guillet

et al. 2000, Huey et al. 2000, Quarrell et al. 2018, Hill et al. 2019) will be investigated in future

studies. On a more general level, our findings emphasize that studying invasive species can

provide unique data sets to empirically and comprehensively test general predictions on

animals' responses to climate change (Gilbert et al. 2014, Merilä and Hendry 2014, Levis and

Pfennig 2016, Hulme 2017, Fox et al. 2019, Rohner et al. 2019), and therefore call for their

open access to the entire research community - a timely task to which the present study

contributes.

AUTHOR CONTRIBUTION STATEMENT

390 JCT designed the experiment, conducted field samplings, and run the experiments. JM analysed

391 the data and wrote the first version of the manuscript. The final manuscript was commented

392 and corrected by all authors.

DATA AVAILABILITY

393 The complete data set and R script are archived in the open data repository

394 Zenodo (https://doi.org/10.5281/zenodo.2652192).

ACKNOWLEDGMENTS

This work is dedicated to the memory of Noelle Tourneur, for her constant support through

JCT career. We thank Maximilian Körner, Franck Dedeine and Sylvain Pincebourde, as well

as Eric Gangloff, Ben Phillips and Fabrice Aubret (PCI Evol Biol referees and recommenders,

respectively) for their comments on a previous version of this manuscript. We also would like

to thank Jean Gingras for his help during laboratory observations and samplings, as well as

Michel Vancassel and Maryvonne Forasté for their help in the late 1980s.

CONFLICT OF INTEREST

401 The authors of this preprint declare that they have no financial conflict of interest with the

402 content of this article. J Meunier is one of the PCI Evol Biol recommenders.

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Table 1 – Details of the 19 sampled populations. The table shows the name and location of

each population, their GPS coordinates (Latitude, Longitude), samplings years, total number

of mating pair setup across years (N. pairs), and thermal regimes (defined as PC1, PC2 and

PC3).

Populations	Country	State (USA)/Province (CDN)	Latitude	Longitude	Samplings	N. pairs	PC1	PC2	PC3
Asheville	USA	North Carolina	35.612	-82.566	1994-95	80	4.60	0.73	0.74
Charlestown	USA	Rhode Island	41.383	-71.642	1990	42	1.37	0.73	-0.84
Deschutes	USA	Oregon	44.157	-121.256	1990	17	-1.54	-2.32	-0.27
Enderby	CDN	British Columbia	50.551	-119.14	1989-90	121	-1.68	-0.02	1.14
Ennis lake	USA	Montana	45.447	-111.695	1990	36	-2.86	-0.36	0.02
Kimberley	CDN	British Columbia	49.635	-115.998	1990	94	-5.27	-0.95	0.73
Kingston	USA	Rhode Island	41.486	-71.531	1991	137	1.00	0.76	-0.75
Montreal	CDN	Quebec	45.542	-73.893	1988,1990-95	356	-2.78	2.32	-0.07
Pointe Pelée	CDN	Ontario	41.963	-82.518	1992	47	1.25	2.31	-0.37
Revelstoke	CDN	British Columbia	50.998	-118.196	1989-90	100	-2.69	-0.11	0.95
Rocky knob	USA	Virginia	36.832	-80.345	1993-94	304	1.44	-0.07	0.48
Saluda	USA	North Carolina	35.198	-82.353	1993-95	117	5.03	0.69	0.77
Santa Cruz*	USA	California	36.926	-121.845	1991	130	5.04	-4.50	-0.57
Selkirk	CDN	Ontario	42.834	-79.932	1992-94	233	-0.69	1.35	-0.67
Selinsgrove	USA	Pennsylvania	40.832	-76.872	1993-94	134	1.76	1.83	0.27
Truro	CDN	Nova Scotia	45.372	-63.264	1988	39	-3.46	-0.13	-1.42
Vancouver	CDN	British Columbia	49.252	-123.24	1989,1991	84	0.23	-2.89	-0.05
Waterrock knob	USA	North Carolina	35.464	-83.138	1991-94	167	-1.88	-1.69	0.22
Wheatley	CDN	Ontario	42.094	-82.445	1992	52	1.13	2.31	-0.31

547   * This population was called San Francisco in (Tourneur 2018).

549 **Table 2 – Loadings of the four first principal components (PCs) reflecting combinations**

550 **of the 12 mean monthly temperatures across populations.** The traits having significant

551 loadings on each PC are in bold.

	PC1	PC2	PC3	PC4
Jan	**0.800**	-0.589	-0.066	0.083
Feb	0.716	**-0.668**	0.131	0.139
Mar	**0.844**	-0.486	0.216	0.048
Apr	**0.949**	-0.140	**0.267**	-0.082
May	**0.890**	0.321	**0.286**	**-0.145**
Jun	0.731	**0.665**	0.123	-0.060
Jul	0.547	**0.823**	-0.006	**0.143**
Aug	0.641	**0.746**	-0.013	**0.175**
Sep	**0.905**	0.380	-0.175	-0.019
Oct	**0.951**	0.019	**-0.292**	-0.064
Nov	**0.931**	-0.174	**-0.296**	-0.112
Dec	**0.872**	-0.469	-0.113	0.041
Eigenvalues	8.153	3.224	0.453	0.130
Variance explained (%)	**67.9**	26.9	3.8	1.1
Cumulative variance explained (%)	67.9	94.8	98.6	99.7

553 **Table 3 –Results of the statistical models on the 13 measured life-history traits.** PC1 positively reflects the overall mean temperature of a

554 population. High values of PC2 reflect populations with cold February (winter) and warm summer, and vice-versa. High values of PC3 reflect

555 populations with warm spring and cold autumn, and vice-versa. P-values significant after FDR correction (adj-P) are in bold. Note that FDR

556 correction transforms each P-value in function of its rank of statistical significance in the data set, which can lead to similar corrected p-values.

557 Model estimates (estim).

	PC1				PC2				PC3			
	estim.	SE	P	adj-P	estim.	SE	P	adj-P	estim.	SE	P	adj-P
First clutch												
Egg laying date	-1.55	1.47	0.307	0.665	-8.92	2.34	0.002	**0.011**	20.37	6.23	0.005	**0.014**
Egg number	-0.32	0.82	0.705	0.896	1.87	1.30	0.171	0.234	-11.77	3.47	0.004	**0.014**
Egg development time	-0.31	0.53	0.575	0.896	2.16	0.85	0.022	**0.041**	-4.57	2.26	0.061	0.132
Larvae number	-1.28	0.72	0.098	0.425	1.53	1.08	0.180	0.234	-3.85	3.39	0.275	0.357
Second clutch												
Egg laying date	0.79	2.01	0.700	0.896	-3.28	3.26	0.331	0.391	13.13	8.46	0.143	0.233
Egg number	0.08	0.44	0.855	0.896	-1.97	0.72	0.016	**0.041**	0.27	1.86	0.887	0.887
Egg development time	1.92	0.92	0.059	0.381	3.93	1.48	0.021	**0.041**	-19.02	4.04	0.001	**0.005**
Larvae number	-0.05	0.38	0.896	0.896	-1.70	0.59	0.012	**0.041**	2.96	1.78	0.121	0.224
General												
Total egg number	0.17	1.01	0.866	0.896	-0.15	1.60	0.929	0.929	-13.93	4.26	0.005	**0.014**
Total larvae number	-1.04	0.77	0.198	0.642	0.60	1.16	0.615	0.666	-3.45	3.61	0.355	0.419
Ratio of iteroparous females	0.01	0.09	0.896	0.896	-0.35	0.13	0.022	**0.041**	-0.48	0.40	0.254	0.357
Male longevity	-10.26	3.63	0.013	0.165	-23.93	5.77	0.001	**0.011**	65.49	15.39	0.001	**0.005**
Female longevity	-4.30	3.74	0.268	0.665	-14.80	5.94	0.025	**0.041**	13.63	15.86	0.404	0.437

Figure 1 – Map showing the 19 sampled populations across Canada (CND) and United

States of America (USA). * This population was called San Francisco in Tourneur (2018).

Figure 2 – Associations between the thermal regimes (PC1, PC2, PC3) of the 19 populations

of origin and 1st clutch parameters. The red lines represent correlations that are significant after

FDR correction. Mean values ± SE. Egg laying date was calculated using October 1st as a

reference (i.e. as day 0).

Figure 3 – Associations between the thermal regimes (PC1, PC2, PC3) of the 19 populations

of origin and 2nd clutch parameters (when produced). The red lines represent correlations that

are significant after FDR correction. Mean values ± SE. Egg laying date was calculated using

October 1st as a reference (i.e. as day 0).

Figure 4 – Associations between the thermal regimes (PC1, PC2, PC3) of the 19 populations

of origin and females' reproductive strategies and outcomes, as well as adult's survival

duration. The red lines represent correlations that are significant after FDR correction. Mean

values ± SE.