Thermal regimes, but not mean temperatures, drive patterns of rapid climate adaptation at a continent-scale: evidence from the introduced European earwig across North America

Running title: Climate adaptation in earwigs

Jean-Claude Tourneur[1], Joël Meunier[2*]

[1] Département des Sciences biologiques, Université du Québec à Montréal 141. Avenue du Président-Kennedy, Montréal, Québec, H2X 1Y4, Canada.

Email: jc.tourneur@nb.sympatico.ca

[2] Institut de Recherche sur la Biologie de l'Insecte (IRBI), UMR 7261, CNRS, University of Tours, Tours, France.

Email: joel.meunier@univ-tours.fr | ORCID: 0000-0001-6893-2064

*Corresponding author.

ABSTRACT

The recent development of human societies has led to major, rapid and often inexorable changes in the environment of most animal species. Over the last decades, a growing number of studies formulated predictions on the modalities of animal adaptation to climate change, questioning how and at what speed animals should adapt to such changes, discussing the levels of risks imposed by changes in the mean and/or variance of temperatures on animal performance, and exploring the underlying roles of phenotypic plasticity and genetic inheritance. These fundamental predictions, however, remain poorly tested using field data. Here, we tested these predictions using a unique continental-scale data set in the European earwig *Forficula auricularia* L, a univoltine insect introduced in North America one century ago. We conducted a common garden experiment, in which we measured 13 life-history traits in 4158 field-sampled earwigs originating from 19 populations across North America. Our results first demonstrate that in less than 100 generations, this species modified 10 of the 13 measured life-history traits in response to the encountered thermal regimes, defined as a variation of temperatures between seasons or months (here winter-summer and autumn-spring temperatures). We found, however, no response to the overall mean monthly temperatures of the invaded locations. Furthermore, our use of a common garden setup reveals that the observed changes in earwigs' life-history traits are not mere plastic responses to their current environment, but are either due to their genetic background and/or to the environmental conditions they experienced during early life development. Overall, these findings provide continent-scale support to the claims that adaptation to thermal changes occurs quickly, even in insects with long life cycles, and emphasize the importance of thermal regimes over mean population temperatures in climate adaptation.

Keywords: Temperature, Adaptation, Reproductive strategy, Climate change, Invasion, Dermaptera

INTRODUCTION

The dramatic acceleration of climate change observed over the last decades challenges the ability of resident organisms to track these changes and adapt their life histories accordingly (Meehl and Tebaldi 2004, Parmesan 2006, Williams et al. 2007). Over the last decades, modelling and theoretical approaches have been developed to better understand the nature and extent of animals' response to such a climate change (Parmesan 2006). These studies formulated key predictions on how and at what speed animals should adapt to such changes, on the respective importance of an increase in the overall mean temperature and/or seasonality of a population on animal performance, as well as on the underlying roles of phenotypic plasticity and genetic inheritance in adaption (Nylin and Gotthard 1998, Kingsolver et al. 2013, Paaijmans et al. 2013, Gilbert et al. 2014, Merilä and Hendry 2014, Levis and Pfennig 2016, Williams et al. 2017, Corl et al. 2018, Fox et al. 2019, Rohner et al. 2019). For instance, these studies suggest that a rapid adaptation to climate change should be facilitated in organisms with fast development and short life-cycles, as found in many arthropods, whereas it should be more difficult in organisms exhibiting slow development and long life-cycle, as found in many vertebrates. Species should also be less sensitive to changes in seasonality compared to changes in overall mean temperatures when they are endotherms and/or when their entire life-cycle occur within a single season, whereas the opposite pattern is expected when they are ectotherms and/or have a life-cycle encompassing several seasons. Finally, phenotypic plasticity is often considered a keystone of rapid adaptation to environmental changes, whereas fixed and inherited patterns of adaptation are often thought to secondarily derive from the canalization of ancestral plastic variation (Chevin et al. 2010).

Although central in our current understanding of animal's responses to climate change, these fundamental predictions remain poorly tested in the field (Janion-Scheepers et

al. 2017, Blanckenhorn et al. 2018). This is probably because such field data are difficult to collect, as it typically requires measuring variation in life-history traits across multiple natural populations, over several years, and under different kind of climates. However, a powerful alternative consists in using field data of introduced species that quickly invaded large geographic areas exhibiting a broad diversity of thermal constraints (Huey et al. 2000, Bellard et al. 2016). In this study, we present and analyze such a unique field data set in one of these species, the European earwig *Forficula auricularia* Linnaeus (Dermaptera: Forficulidae), after its introduction in North America. This insect exhibits a broad native range extending across Europe, Asia and northern Africa (Lamb and Wellington 1975) from which it has been introduced to Australia, New Zealand, East Africa, East Indies, Tasmania and North America (Frank 1918, Guillet et al. 2000, Quarrell et al. 2018, Hill et al. 2019). Its presence in North America was first reported on the Pacific coast in Seattle (WA) in 1907 (Fulton 1924), and then on the Atlantic coast in Newport (RI) in 1911 (Jones 1917) and in Vancouver (BC) in 1919 (Treherne 1923). From these introductory foci, *F. auricularia* first spread along the coasts to cover areas ranging from British Columbia to California and from Newfoundland to South Carolina, and then reached the interior of the continent in both United States of America (Crumb et al. 1941) and Canada (Wilson 1971, Cantrall 1972, Tourneur 2017). Given that this species produces only one generation per year (Tourneur and Gingras 1992, Meunier et al. 2012), these historical records reveal that its successful colonization of North America and thus its adaptation to a broad diversity of thermal environments occurred in less than 100 generations.

Because the univoltine life cycle of the European earwig lasts up to 2 years and encompasses all seasons and temperatures (Lamb 1976, Meunier et al. 2012), it has long been thought that annual mean temperatures and/or temperature seasonality could be major constraints in the success of *F. auricularia* invasions (Vancassel 1984, Hill et al. 2019).

However, it remains unclear whether this species can mitigate these thermal constraints, and whether it does so by adapting its life cycle and life-history traits (Ratz et al. 2016, Tourneur 2018). The life cycle of the European earwig generally starts with the emergence of new adults in late spring to early July, with variation among populations. These adults form groups of up to several hundred individuals, in which both males and females typically mate with several partners (Weiß et al. 2014, Sandrin et al. 2015, Tourneur 2017). Females then burrow in the ground from mid fall to early winter and build a nest where they lay their first clutch of eggs. After egg laying, females stop their foraging activity and provide extensive forms of egg care until hatching (Gingras and Tourneur 2001, Boos et al. 2014, Koch and Meunier 2014, Thesing et al. 2015, Diehl and Meunier 2018, Körner et al. 2018). The eggs of this first clutch hatch in spring and mothers remain with their newly hatched larvae for several weeks, during which mothers provide larvae with multiple forms of care (Gingras and Tourneur 2001, Kölliker et al. 2015, Kramer et al. 2015) and larvae exhibit forms of sibling cooperation (Falk et al. 2014, Kramer et al. 2015, Kramer and Meunier 2016, Körner et al. 2016). A few weeks later, the family unit is naturally disrupted. While larvae continue their development to adults in new social groups, some females produce a second clutch of eggs (i.e. iteroparous as compared to semelparous females), which will also receive pre- and post-hatching care and will hatch in late spring (Lamb and Wellington 1975, Meunier et al. 2012, Ratz et al. 2016). All females generally die during the following summer (Albouy and Caussanel 1990).

In this study, we used a common garden experiment to explore how *F. auricularia* responded to the different thermal environments encountered during their North American invasion over the last century, i.e. in less than 100 generations. In particular, we 1) tested whether and how individuals altered their life history traits in response to the thermal constraints of the invaded locations, 2) identified the thermal constraints to which they

111 adapted and 3) investigated the role of phenotypic plasticity in this adaptation. From 1988 to
112 1995, we field-sampled individuals originating from 19 populations located from the East to
113 the West coasts, maintained them under standard laboratory conditions and measured the
114 properties of the 1st and 2nd clutches produced by each female in terms of egg laying date, egg
115 number, egg development time and number of newly hatched larvae. We also recorded the
116 reproductive strategy of the females (iteroparity versus semelparity), their reproductive
117 outcome (total number of eggs and larvae produced over lifetime), as well as the
118 experimental survival duration of the field-sampled males and females. To identify which
119 thermal constraints the tested earwigs adapted to, we tested whether our measurements could
120 be explained by the results of a principal component analyses (PCA) of the mean monthly
121 temperatures of each population. This process characterizes patterns of variation among
122 populations' temperatures without *a priori* definitions of their associations, i.e. without
123 predetermining the focus on overall mean temperatures and/or specific thermal regimes
124 (defined as variation of temperatures between seasons or months). If *F. auricularia*
125 individuals adapted their life-cycle and life-history traits to the mean temperatures and/or
126 thermal regimes of the population in which they have been sampled (and if this adaptation is
127 determined by their genetic background and/or early life experience), we predict these traits
128 to covary with the overall mean temperatures and/or variation in seasonal temperatures of
129 their population (i.e. all sampled populations should show different performance in the
130 common garden). Conversely, if earwig life-history traits are independent of the thermal
131 environment of the population in which they have been sampled (i.e. no adaptation) and/or
132 are plastic to their current thermal environment, we predict no apparent association between
133 the traits measured in our field-sampled individuals and the thermal regimes of their
134 populations (i.e. all sampled populations should show similar performance in the common
135 garden).

MATERIAL AND METHODS

Earwig sampling and laboratory rearing

All *F. auricularia* individuals were collected over 7 years among 19 natural populations located across North America (Figure 1, Table 1). These individuals were mostly collected as adults using wooden traps (Tourneur 2018) between July and August, and were immediately setup in glass containers (Mason Jars Company, Erie, Pennsylvania, United States of America) in groups of 20 to 30 individuals. These containers received two sheets of creased toilet paper as resting places for earwigs, and were then transported to the laboratory in Montreal, Canada. Upon their arrival, containers were deposited in a shelf covered by a shelter and maintained under the natural outdoor conditions of Montreal. During their transport and outdoor maintenance, containers received an *ad libitum* amount of carrots and pollen as a food source for earwigs, and were supplied with water by means of a cotton pad regularly soaked in water. This setup allowed earwigs to perform non-controlled mating and to live in groups – just like they do under natural conditions (Weiß et al. 2014, Sandrin et al. 2015, Kohlmeier et al. 2016, Körner et al. 2018).

One to two months later (between the 7th and the 19th day of October of each year), we used 4158 of these field-sampled individuals to set up 2079 mating pairs (from 17 to 356 pairs per population, see Table 1), in which we subsequently measured 13 life-history traits (see below). These pairs were set up in Petri dishes (diameter 10 cm) lined with a thin layer of moist sand, and in which food was changed and substrate humidified once a week. Each Petri dish was then transferred in a climate chamber and then maintained at 10 ± 1 °C, a temperature close to the overall median temperature of the 19 sampled populations (i.e. 9.5°C, see Table S1). Food was removed at egg laying to mimic the natural end of earwigs'

158 foraging activity (Kölliker 2007). At egg hatching, we discarded all newly emerged larvae
159 from the experiments to trigger a novel ovarian cycle in the mothers and allow their
160 production of a subsequent clutch (Vancassel and Foraste 1980, Meunier et al. 2012). We
161 then maintained the pairs under the rearing conditions described above until our experiment
162 ended, i.e. either one year after the beginning of our laboratory setup or at the death of the
163 adult males and females. Overall, 3927 of the 4158 (94.4%) tested individuals died within the
164 year following the beginning of our experiments, a value in line with previous data on *F.*
165 *auricularia* lifespan (Albouy and Caussanel 1990). Note that recent studies revealed that
166 North American *F. auricularia* encompasses two genetic subspecies with no apparent mixing
167 of their populations (Wirth et al. 1998, Quarrell et al. 2018, Tourneur 2018). Although these
168 subspecies were not considered in our analyses (our data were collected before the
169 publication of these genetic analyses), the continuous distribution (unimodal data) of the life
170 history traits measured across populations (Figures 2 to 4) suggests an absence of subspecies-
171 specific values regarding these measurements. The potential co-occurrence of the two
172 subspecies in our data set is thus unlikely to bias our study and its main conclusions.

173 **Measurements of the life-history traits**

174 For each mating pair, we measured 13 life-history traits encompassing the properties of the
175 resulting 1st and 2nd clutches (when present), the reproductive strategy and reproductive
176 outcomes of each female, as well as the experimental survival duration of both field-sampled
177 males and females. These properties were obtained by recording the date of egg production,
178 counting the number of eggs produced, calculating the duration of egg development until
179 hatching (in days) and finally counting the number of larvae at egg hatching in both 1st and
180 2nd clutches (when present). The reproductive strategies and reproductive outcomes of
181 females were obtained by recording whether they were semelparous or iteroparous (i.e.
182 produced one or two clutches in their lifetime, respectively), and by counting the total

number of eggs and larvae produced per female during their lifetime. Finally, we measured the experimental survival duration of adults by counting the number of days each male and female survived after October 1st of the year of field sampling. Although our measurement of survival duration does not necessarily reflect adults longevity, as individuals could have different ages at field sampling (see discussion), it nevertheless provides important insights into the period at which males and females of each population die during the year. Note that 8.1% and 5.4% females from Santa Cruz and Asheville, respectively, produced a third clutch. This third clutch was not considered in the present study, as our experiment ended before their hatching.

Extraction of mean temperatures and thermal regimes of each population

We extracted the mean monthly temperature of the 19 studied populations using their GPS coordinates (Table 1) and the Worldclim database v2.0 (http://www.worldclim.org/) with a spatial resolution of 30 seconds. The mean temperatures provided by the Worldclim database are calculated over 30 years, from 1970 to 2000. To reduce dimensionality of co-varying temperatures in our data set while characterizing potential thermal regimes of each population without *a priori* definitions of their composition, we then conducted a Principal Component Analysis (PCA) on the set of 12 mean monthly temperatures per population (Table S1). This analysis provided us with 12 orthogonal principal components (PCs), out of which we retained the first three PCs (total variance explained = 98.6%, Table 2). The first component (PC1) was positively loaded by almost all monthly temperatures, therefore positively reflecting the overall mean temperature of a population. The second component (PC2) revealed variation in seasonality between February on one hand, and June, July, and August on the other hand. In particular, high values of PC2 reflected populations with cold February (winter) and warm summer, whereas small values of PC2 reflected populations with warm February (winter) and cold summer. Finally, the third component (PC3) captured variation in

seasonality between October and November on one hand, and April and May on the other hand. High values of PC3 therefore characterized populations with cold autumn and warm spring, whereas small values of PC3 characterized populations with warm autumn and cold spring.

Statistical analyses

To test whether *F. auricularia* adapt their life-cycle and life-history traits to North American temperatures, we conducted a series of 12 linear models (LM in R) and one generalized linear model (GLM in R) – see Table 3. In the 12 LMs, the three selected PCs and their interactions were entered as explanatory variables (PC1, PC2 and PC3), whereas the response variable was either egg laying date, egg number, egg development time and larvae number for the 1st or 2nd clutches (for a total of 8 LMs), the total number of eggs or larvae produced, or the survival duration of males or females. Note that both egg laying date and adult survival duration were calculated using October 1st as day 0. In the GLM, the response variable was the ratio of iteroparous females per population, which was entered using the command *cbind* in R (to weight each ratio by the sample size of its population) and fitted to a binomial error distribution corrected for overdispersion. In all our statistical models, the response variables were the mean values of each measured trait per population. They were also checked for homoscedasticity and normality of residuals, as well as simplified stepwise by removing all non-significant interaction terms (all $P > 0.05$). To correct for inflated Type-I errors due to multiple testing (and provide an experiment-wide Type I error rate of 5%), all *P*-values were adjusted using False Discovery Rate (FDR) correction (Benjamini and Hochberg 1995). All analyses were conducted using the software R v3.5.1 loaded with the packages *raster*, *FactoMineR*, *rsq* and *rcompanion*.

RESULTS

The 19 studied populations greatly varied in their mean temperatures and thermal regimes (Table S1), as well as in the mean values of the 13 traits measured in their sampled individuals (Figures 2 to 4; Tables S2 to S4). Mean monthly temperatures overall ranged from 22.9°C (July in Saluda) to -10.1°C (January in Montreal), while thermal amplitudes over a year ranged from 30.7°C (Montreal) to 7.9°C (Santa Cruz). For the traits measured in the 1st clutches, the mean dates of egg production ranged from 47.8 to 132.6 days after the 1st of October, the mean number of eggs per clutch from 23.2 to 66.0, the mean egg development time from 42.2 to 71.4 days and the mean number of larvae per clutch from 11.6 to 44.8. For the 2nd clutches, the mean dates of egg production ranged from 142.0 to 248.2 days after the 1st of October, the mean number of eggs from 14.0 to 38.4, the mean egg development time from 10.0 to 63.7 days and the mean number of larvae from 0 to 17.7. Finally, the total number of eggs produced ranged from 28.1 to 83.4, the total number of larvae produced from 13.0 to 46.3, the proportion of iteroparous females from 0 to 70.8%, and the survival duration of males and females from 82.0 to 299.8 days and from 146.0 to 322.5 days after the 1st of October, respectively.

Of the 13 measured traits, 10 varied together with the thermal regimes of the population of origin (Table 3). Five of these 10 traits were exclusively associated with PC2 (February-summer temperatures), two traits were exclusively associated with PC3 (autumn-spring temperatures), and three traits were associated with both PC2 and PC3. By contrast, no traits were associated with PC1 (overall mean temperatures). The associations with PC2 revealed that populations with cold February and warm summers (high PC2 values) had females that produced their 1st clutch of eggs earlier and these eggs had longer development time compared to populations exhibiting warm February and cold summers (low PC2 values, Figure 2). Similarly, females from the former populations were less likely to produce a

255 second clutch (i.e. to be iteroparous, Figure 3) and when they did so, their 2nd clutch eggs
256 were less numerous (Figure 3) and showed longer development time (Figure 3). Moreover,
257 females and males from populations with cold February and warm summers lived less long
258 compared to adults from warm February and cold summers (Figure 4). On the other hand, the
259 effects of PC3 reveal that populations exhibiting cold autumns and warm springs (high PC3
260 values) had females that produced their 1st clutch of eggs later in the season and these eggs
261 were less numerous compared to females from populations with warm autumns and cold
262 springs (low PC3 values, Figure 2). Females from the former populations also had 2nd clutch
263 eggs that exhibited a shorter developmental time (Figure 3), they produced an overall lower
264 number of eggs (Figure 4) and had males with a longer survival duration (Figure 4). By
265 contrast, PC1, PC2 and PC3 did not shape the number of 1st clutch larvae, as well as their
266 total number and the dates of 2nd clutch egg laying (Figures 2, 3 and 4; Table 3).

DISCUSSION

267 Shedding light on how species successfully adapt to a broad set of environmental constraints
268 is of major importance to improve our general understanding of the mechanisms underlying
269 animal adaptations to climate change. In this study, we demonstrate that the successful
270 invasion of the European earwig across North America came with multiple changes in their
271 life-history traits in response to the thermal regimes (sets of winter-summer and autumn-
272 spring temperatures), but not to the overall mean temperature of the invaded areas. In
273 particular, our data from 19 populations revealed that females changed their timing of first
274 reproduction, their reproductive strategy and investment into egg production when facing
275 different thermal regimes, while experimental survival duration of males and females varied
276 accordingly. By contrast, we found no association between thermal regimes and both the
277 timing of second reproduction and the total number of larvae produced per female.

We first showed that females produced their first clutch of eggs earlier when they came from populations facing warm summers and/or warm autumns (PC2 and PC3, respectively), and were less likely to produce a second clutch in populations with cold February. A plastic response to warm temperatures on egg laying date could be expected in nature: adult earwigs typically develop and mate during summer and autumn, so that warm temperatures during these seasons could accelerate their reproductive physiology (as shown in other insect species, Singh et al., 2018) and thus accelerate egg laying (Tourneur 2018). Similarly, cold Februaries might slow down the development of 1st clutch eggs and thus extend the corresponding period of egg care. This, in turn, might inhibit females' physiological transformation to produce a second clutch (Vancassel 1984, Gingras and Tourneur 2001, Tourneur 2018, Körner et al. 2018). However, our results were obtained under common garden conditions, which reveals that the observed effects of thermal regime on egg laying dates are not a plastic response to their current environment, but are either due to the environment experienced during their early life development (i.e. before field sampling), or due to an inherited basis that possibly emerged through canalization (Nylin and Gotthard 1998, Van Buskrik and Steiner 2009). It has been proposed that traits tightly linked to fitness are more strongly canalized due to past stabilizing selection (Falconer 1990). Our findings may therefore suggest that the observed changes in the timing of first reproduction and females' reproductive strategy may have first emerged as a plastic response to the thermal constraints of the different localities, then diverged between populations through canalization and ultimately become inherited traits – all this in a maximum of 100 generations. Further experiments with naïve individuals remain, however, required to rule out an effect of early life experience.

Our data also reveal that thermal regimes are associated with lifetime egg production, but not with lifetime larvae production. In particular, the total number of eggs produced per

303 female decreased with decreasing autumn temperatures, whereas this association vanished
304 with larvae number. This apparent discrepancy suggests that females from populations with
305 the warmest autumns lost a larger number of eggs during egg development. A first
306 explanation could be that these females produced eggs of lower quality and/or were less
307 efficient in egg care, a process that is essential to ensure egg development until hatching in
308 earwigs (Boos et al. 2014, Van Meyel et al. 2019). Whereas both effects should be tested in
309 future studies, previous results may suggest that the second effect is unlikely, as maternal
310 investment in post-hatching care is not population-specific, at least in Europe (Ratz et al.
311 2016). Another explanation is that females consumed a larger part of their clutch in
312 populations with the warmest compared to the coldest autumns. Filial egg consumption is a
313 common phenomenon in insects (Elgar and Crespi 1992) and it has been recently reported in
314 several Dermapteran species, such as the species studied here (Koch and Meunier 2014, Van
315 Meyel et al. 2019) and the maritime earwig *Anisolabis maritima* Bonelli (Miller and Zink
316 2012). In the European earwig, this phenomenon has been proposed to reflect an adaptive
317 strategy to limit female weight loss during the period of egg care (i.e. when they stop all other
318 foraging activities) and by doing so, to reallocate resources into post-hatching care and/or
319 into a 2nd oogenesis cycle (Koch and Meunier 2014, Tourneur 2018). Given that females lay
320 eggs earlier in populations with the warmest autumns, this increased egg consumption could
321 be an adaptive strategy to limit the cost of tending newly hatched offspring earlier in the
322 season (middle of winter) when food sources are scarce or absent. If this hypothesis holds
323 true, it would suggest that filial egg cannibalism could be a strategy that *F. auricularia*
324 females have evolved to better cope with warmer autumns.

325 In addition to the above findings, our results show that the survival duration of both
326 males and females was associated with the thermal regime of the population of origin. In
327 particular, females' and males' survival duration decreased together with warm summers (and

328 cold Februaries), while male's survival duration also decreased with warm autumns (and cold
329 springs). The first results may be a by-product of the effect of temperature on their date of
330 egg laying and/or egg hatching. In particular, we showed that females from populations
331 facing warm summers are the first to lay their eggs. Individuals from these populations might
332 thus have been the oldest at the date of our field sampling, therefore leading to the shortest
333 survival duration in our subsequent experiment. Surprisingly, there was a sex-specific effect
334 of spring (and autumn) temperatures on adult survival duration: males lived up to two times
335 longer in populations with warm compared to cold springs (as well as cold compared to warm
336 autumns), whereas this effect was absent in females. This finding may reflect sex-specific
337 sensitivity to high temperatures in terms of, for instance, physiology or expression of costly
338 behaviors. Whereas some physiological traits are known to be sex-specific in this species
339 (Kohlmeier et al. 2016, Vogelweith et al. 2017), further studies should explore the effects of
340 temperature on the observed differences. Notwithstanding its underlying mechanisms, the
341 long survival duration of males in warm spring populations opens scope for these males to
342 mate with females of the subsequent generation, as well as for a possible involvement of
343 fathers into larva care – a phenomenon reported in other insect species (Smiseth 2014). These
344 two processes remain unknown in the European earwig, but they could be of central
345 importance in their successful adaptation to climate change.

346 All our results are based on a common garden experiment, a method that is often
347 considered a powerful tool to disentangle the roles of phenotypic plasticity and genetic
348 background on adaptation (Franks et al. 2014, Stoks et al. 2014, Blanckenhorn et al. 2018).
349 Individuals reared under a common environment are typically expected to exhibit
350 homogenized life-history traits if adaptation is the outcome of phenotypic plasticity, whereas
351 they should exhibit population-specific traits otherwise. Our results are in line with the latter
352 process for the great majority of the measured life-history traits (10 out of 13), therefore

353 suggesting that the observed associations between thermal regimes and life-history traits do
354 not stem from a plastic response to their current environment. Nevertheless, common garden
355 experiments often have some limits: they do not prevent maternal and grand maternal effects,
356 they cannot preclude the possibility of genotype-by-environment interactions on the
357 measured life-history traits, and they are poorly efficient at shedding light on the multiple
358 facets of plasticity (e.g. some traits can be partially plastic, the plastic responses can vary in
359 intensity and slope, and plasticity may become apparent only after certain thresholds) (Franks
360 et al. 2014, Merilä and Hendry 2014, Stoks et al. 2014, Bodensteiner et al. 2019). These
361 limits can be particularly important here, as maternal effects and harsh environments shape
362 the nature and outcomes of several family interactions in earwigs (Meunier and Kölliker
363 2012a, 2012b, Thesing et al. 2015, Raveh et al. 2016, Kramer et al. 2017). Concluding on the
364 absence or limited role of plasticity in earwigs' adaptation to North American' thermal
365 regimes would therefore need further empirical works exploring its multiple facets under
366 several common garden conditions (Bodensteiner et al. 2019), and if present, demonstrating
367 the adaptive value of this apparent plasticity.

368 To conclude, our results demonstrate that the spread of the European earwigs across
369 North America came with important changes in their life-history traits and life cycle, and that
370 these changes emerged in a maximum of 100 generations. Whereas we show that some of
371 these changes are by-products of novel thermal constraints (timing of first reproduction and
372 female iteroparity), we reveal that others are likely to reflect adaptive strategies to cope with
373 different autumn temperatures (egg production and the possibility of egg cannibalism).
374 Overall, these findings emphasize that adaptation of an insect with a relatively long life-cycle
375 does not necessarily operate in response to the overall mean temperatures of the invaded
376 environments, but to their thermal regimes – i.e. to seasonality and/or mean temperature at a
377 specific time of their life-cycle. Whether the reported adaptations are the product of

population-differences in energetic/metabolic constraints experienced by adults during their early development (Wong and Kölliker 2014, English et al. 2016), and/or the product of an inherited genetic basis that varies with thermal regimes (Levis and Pfennig 2016; Corl et al. 2018; Fox et al. 2019), as well as whether these adaptations are similar across its worldwide distribution (Frank 1918, Guillet et al. 2000, Huey et al. 2000, Quarrell et al. 2018, Hill et al. 2019) will be investigated in future studies. On a more general level, our findings emphasize that studying invasive species can provide unique data sets to empirically and comprehensively test general predictions on animals' responses to climate change (Gilbert et al. 2014, Merilä and Hendry 2014, Levis and Pfennig 2016, Hulme 2017, Fox et al. 2019, Rohner et al. 2019), and therefore call for their open access to the entire research community - a timely task to which the present study contributes.

AUTHOR CONTRIBUTION STATEMENT

JCT designed the experiment, conducted field samplings, and run the experiments. JM analysed the data and wrote the first version of the manuscript. The final manuscript was commented and corrected by all authors.

DATA AVAILABILITY

The complete data set and R script are archived in the open data repository Zenodo (https://doi.org/10.5281/zenodo.2652192).

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CONFLICT OF INTEREST

The authors of this preprint declare that they have no financial conflict of interest with the content of this article. J Meunier is one of the PCI Evol Biol recommenders.

REFERENCES

Table 1 – Details of the 19 sampled populations. *The table shows the name and location of each population, their GPS coordinates (Latitude, Longitude), samplings years, total number of mating pair setup across years (N. pairs), and thermal regimes (defined as PC1, PC2 and PC3).*

Populations	*Country*	*State (USA)/Province (CDN)*	*Latitude*	*Longitude*	*Samplings*	*N. pairs*	*PC1*	*PC2*	*PC3*
Asheville	*USA*	*North Carolina*	*35.612*	*-82.566*	*1994-95*	*80*	*4.60*	*0.73*	*0.74*
Charlestown	*USA*	*Rhode Island*	*41.383*	*-71.642*	*1990*	*42*	*1.37*	*0.73*	*-0.84*
Deschutes	*USA*	*Oregon*	*44.157*	*-121.256*	*1990*	*17*	*-1.54*	*-2.32*	*-0.27*
Enderby	*CDN*	*British Columbia*	*50.551*	*-119.14*	*1989-90*	*121*	*-1.68*	*-0.02*	*1.14*
Ennis lake	*USA*	*Montana*	*45.447*	*-111.695*	*1990*	*36*	*-2.86*	*-0.36*	*0.02*
Kimberley	*CDN*	*British Columbia*	*49.635*	*-115.998*	*1990*	*94*	*-5.27*	*-0.95*	*0.73*
Kingston	*USA*	*Rhode Island*	*41.486*	*-71.531*	*1991*	*137*	*1.00*	*0.76*	*-0.75*
Montreal	*CDN*	*Quebec*	*45.542*	*-73.893*	*1988,1990-95*	*356*	*-2.78*	*2.32*	*-0.07*
Pointe Pelée	*CDN*	*Ontario*	*41.963*	*-82.518*	*1992*	*47*	*1.25*	*2.31*	*-0.37*
Revelstoke	*CDN*	*British Columbia*	*50.998*	*-118.196*	*1989-90*	*100*	*-2.69*	*-0.11*	*0.95*
Rocky knob	*USA*	*Virginia*	*36.832*	*-80.345*	*1993-94*	*304*	*1.44*	*-0.07*	*0.48*
Saluda	*USA*	*North Carolina*	*35.198*	*-82.353*	*1993-95*	*117*	*5.03*	*0.69*	*0.77*
*Santa Cruz**	*USA*	*California*	*36.926*	*-121.845*	*1991*	*130*	*5.04*	*-4.50*	*-0.57*
Selkirk	*CDN*	*Ontario*	*42.834*	*-79.932*	*1992-94*	*233*	*-0.69*	*1.35*	*-0.67*
Selinsgrove	*USA*	*Pennsylvania*	*40.832*	*-76.872*	*1993-94*	*134*	*1.76*	*1.83*	*0.27*
Truro	*CDN*	*Nova Scotia*	*45.372*	*-63.264*	*1988*	*39*	*-3.46*	*-0.13*	*-1.42*
Vancouver	*CDN*	*British Columbia*	*49.252*	*-123.24*	*1989,1991*	*84*	*0.23*	*-2.89*	*-0.05*
Waterrock knob	*USA*	*North Carolina*	*35.464*	*-83.138*	*1991-94*	*167*	*-1.88*	*-1.69*	*0.22*
Wheatley	*CDN*	*Ontario*	*42.094*	*-82.445*	*1992*	*52*	*1.13*	*2.31*	*-0.31*

** This population was called San Francisco in (Tourneur 2018).*

408 ***Table 2 – Loadings of the four first principal components (PCs) reflecting combinations of***

409 ***the 12 mean monthly temperatures across populations.*** *The traits having significant*

410 *loadings on each PC are in bold.*

	PC1	PC2	PC3	PC4
Jan	**0.800**	-0.589	-0.066	0.083
Feb	0.716	**-0.668**	0.131	0.139
Mar	**0.844**	-0.486	0.216	0.048
Apr	**0.949**	-0.140	**0.267**	-0.082
May	**0.890**	0.321	**0.286**	**-0.145**
Jun	0.731	**0.665**	0.123	-0.060
Jul	0.547	**0.823**	-0.006	**0.143**
Aug	0.641	**0.746**	-0.013	**0.175**
Sep	**0.905**	0.380	-0.175	-0.019
Oct	**0.951**	0.019	**-0.292**	-0.064
Nov	**0.931**	-0.174	**-0.296**	-0.112
Dec	**0.872**	-0.469	-0.113	0.041
Eigenvalues	8.153	3.224	0.453	0.130
Variance explained (%)	**67.9**	26.9	3.8	1.1
Cumulative variance explained (%)	67.9	94.8	98.6	99.7

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Table 3 –Results of the statistical models on the 13 measured life-history traits. *PC1 positively reflects the overall mean temperature of a population. High values of PC2 reflect populations with cold February (winter) and warm summer, and vice-versa. High values of PC3 reflect populations with warm spring and cold autumn, and vice-versa. P-values significant after FDR correction (adj-P) are in bold. Note that FDR correction transforms each P-value in function of its rank of statistical significance in the data set, which can lead to similar corrected p-values. Model estimates (estim).*

	PC1				PC2				PC3			
	estim.	*SE*	*P*	*adj-P*	*estim.*	*SE*	*P*	*adj-P*	*estim.*	*SE*	*P*	*adj-P*
First clutch												
Egg laying date	*-1.55*	*1.47*	*0.307*	*0.665*	*-8.92*	*2.34*	*0.002*	***0.011***	*20.37*	*6.23*	*0.005*	***0.014***
Egg number	*-0.32*	*0.82*	*0.705*	*0.896*	*1.87*	*1.30*	*0.171*	*0.234*	*-11.77*	*3.47*	*0.004*	***0.014***
Egg development time	*-0.31*	*0.53*	*0.575*	*0.896*	*2.16*	*0.85*	*0.022*	***0.041***	*-4.57*	*2.26*	*0.061*	*0.132*
Larvae number	*-1.28*	*0.72*	*0.098*	*0.425*	*1.53*	*1.08*	*0.180*	*0.234*	*-3.85*	*3.39*	*0.275*	*0.357*
Second clutch												
Egg laying date	*0.79*	*2.01*	*0.700*	*0.896*	*-3.28*	*3.26*	*0.331*	*0.391*	*13.13*	*8.46*	*0.143*	*0.233*
Egg number	*0.08*	*0.44*	*0.855*	*0.896*	*-1.97*	*0.72*	*0.016*	***0.041***	*0.27*	*1.86*	*0.887*	*0.887*
Egg development time	*1.92*	*0.92*	*0.059*	*0.381*	*3.93*	*1.48*	*0.021*	***0.041***	*-19.02*	*4.04*	*0.001*	***0.005***
Larvae number	*-0.05*	*0.38*	*0.896*	*0.896*	*-1.70*	*0.59*	*0.012*	***0.041***	*2.96*	*1.78*	*0.121*	*0.224*
General												
Total egg number	*0.17*	*1.01*	*0.866*	*0.896*	*-0.15*	*1.60*	*0.929*	*0.929*	*-13.93*	*4.26*	*0.005*	***0.014***
Total larvae number	*-1.04*	*0.77*	*0.198*	*0.642*	*0.60*	*1.16*	*0.615*	*0.666*	*-3.45*	*3.61*	*0.355*	*0.419*
Ratio of iteroparous females	*0.01*	*0.09*	*0.896*	*0.896*	*-0.35*	*0.13*	*0.022*	***0.041***	*-0.48*	*0.40*	*0.254*	*0.357*
Male longevity	*-10.26*	*3.63*	*0.013*	*0.165*	*-23.93*	*5.77*	*0.001*	***0.011***	*65.49*	*15.39*	*0.001*	***0.005***
Female longevity	*-4.30*	*3.74*	*0.268*	*0.665*	*-14.80*	*5.94*	*0.025*	***0.041***	*13.63*	*15.86*	*0.404*	*0.437*

418 ***Figure 1 – Map showing the 19 sampled populations across Canada (CND) and United***

419 ***States of America (USA)***. ** This population was called San Francisco in Tourneur (2018).*

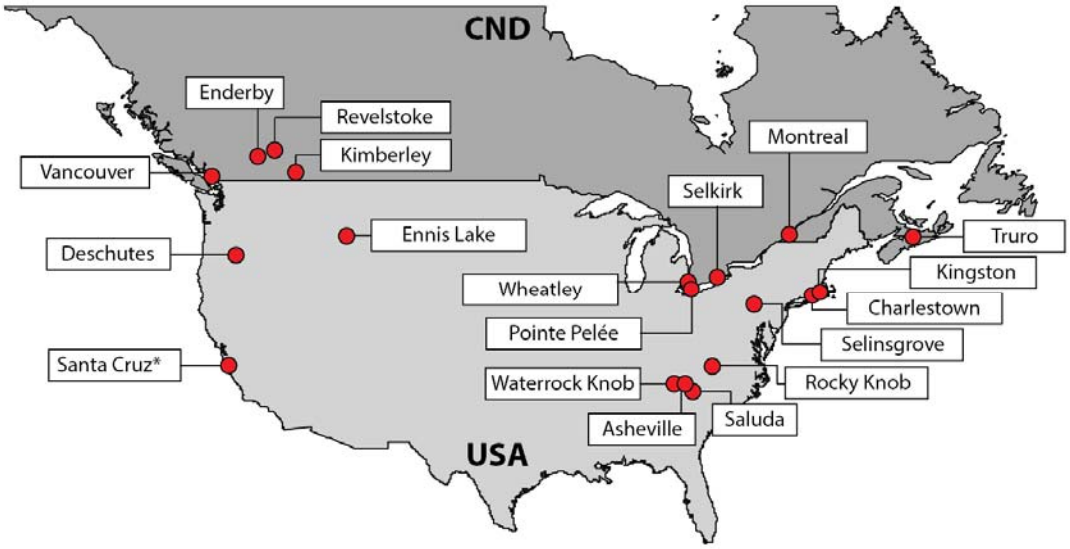

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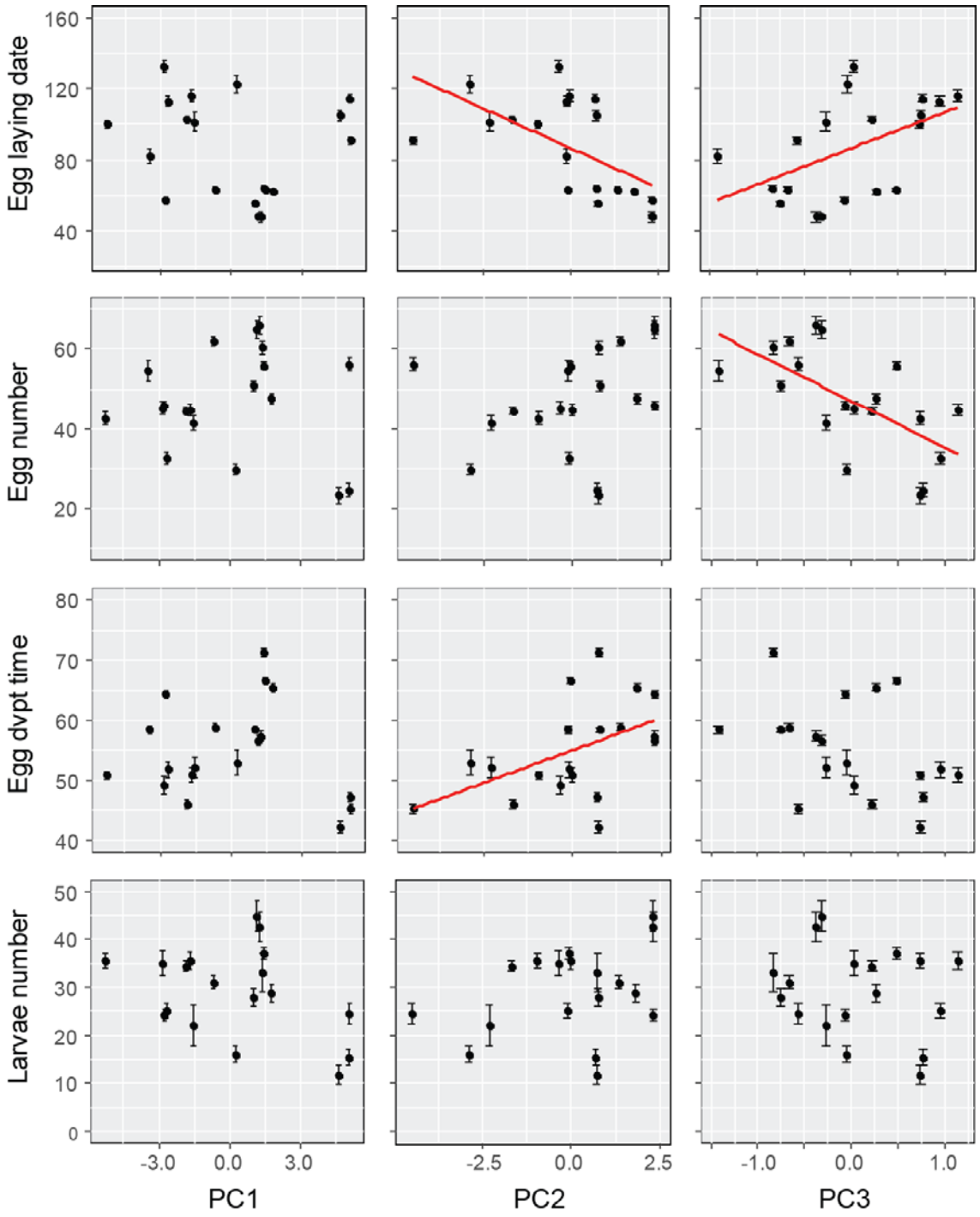
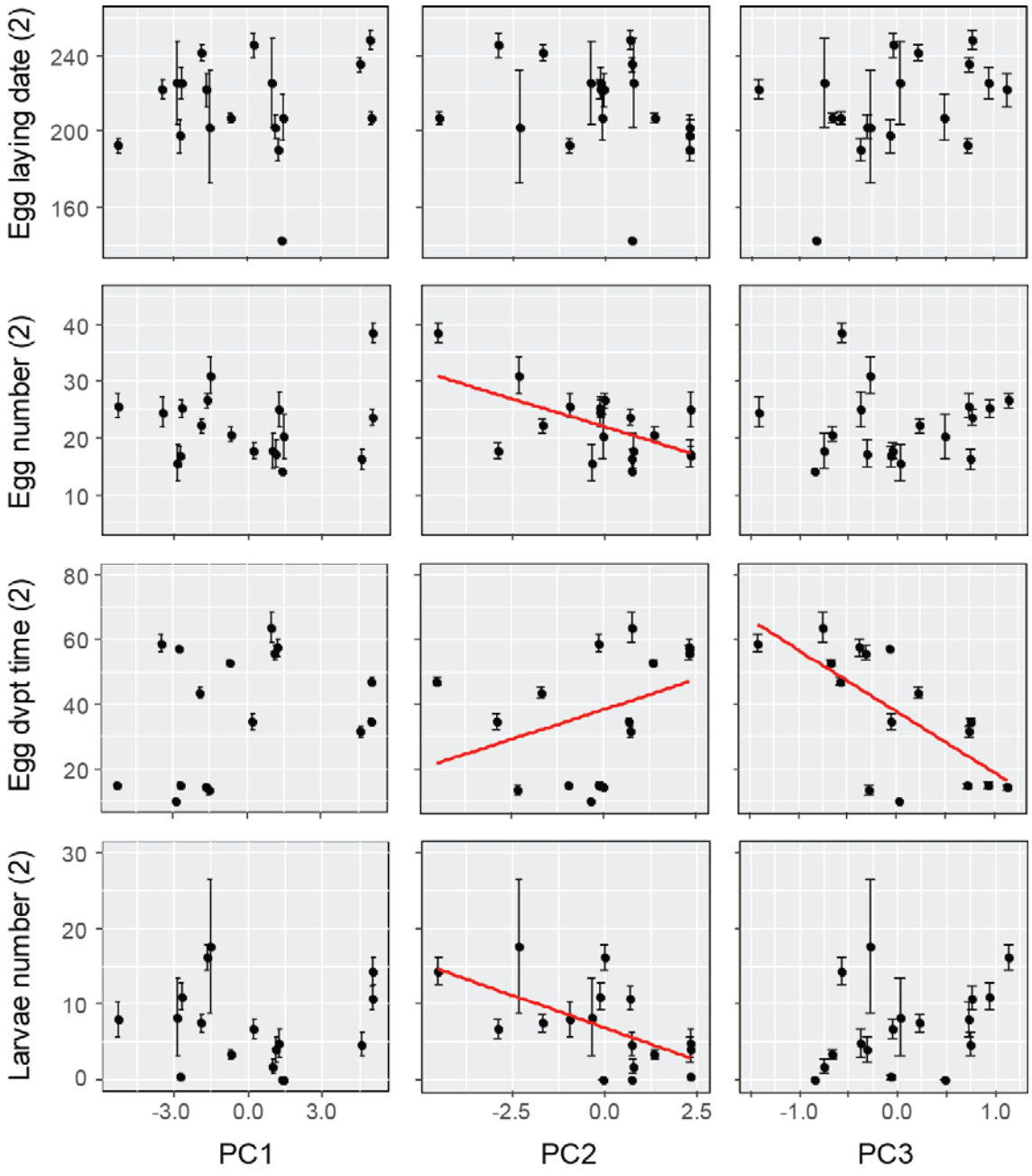
Figure 2 *– Associations between the thermal regimes (PC1, PC2, PC3) of the 19 populations of origin and 1st clutch parameters. The red lines represent correlations that are significant after FDR correction. Mean values ± SE. Egg laying date was calculated using October 1st as a reference (i.e. as day 0).*



428 ***Figure 3*** *– Associations between the thermal regimes (PC1, PC2, PC3) of the 19 populations*
429 *of origin and 2nd clutch parameters (when produced). The red lines represent correlations*
430 *that are significant after FDR correction. Mean values ± SE. Egg laying date was calculated*
431 *using October 1st as a reference (i.e. as day 0).*



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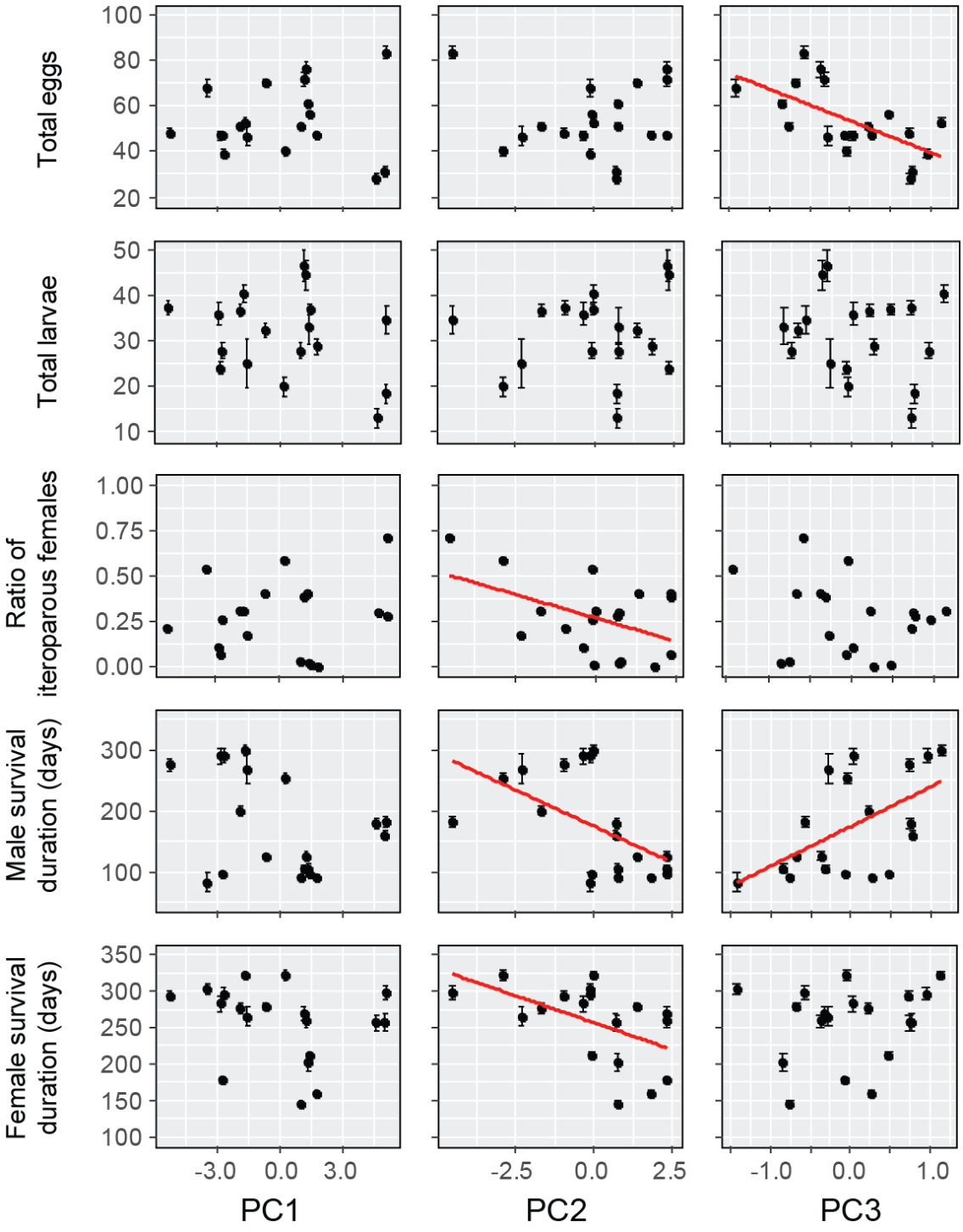
434 ***Figure 4*** *– Associations between the thermal regimes (PC1, PC2, PC3) of the 19 populations*
435 *of origin and females' reproductive strategies and outcomes, as well as adult's survival*
436 *duration. The red lines represent correlations that are significant after FDR correction.*
437 *Mean values ± SE.*



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