

# **Influence of Organic Matrix and Cations on Bio- Methane Yield with Anthracite Methanogenic Consortium**

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## **Abstract**

Organic compounds fermentation of coal has been used to generate secondary biogenic gas and enhance gas reservoirs in coalbed. To enhance the bio-degradation process, culture nutrition plays an important role in remediating the nutritional deficiency of the coal seam. The influence of bio-methane yield with organic inputs and cations concentrations was examined. Research of organic matrix influence revealed that the traditional organic material except yeast extract should forbid, and the input of yeast extract should limit at 1.00g/L also. Further, the study demonstrated that the ion concentration of

19 sodium, potassium, magnesium, calcium and ammonia nitrogen also influenced methane  
20 and carbon dioxide yields. And the optimize concentrations for  $\text{Ca}^{2+}$ ,  $\text{K}^+$ ,  $\text{Na}^+$ ,  $\text{Mg}^{2+}$  were  
21 5.1, 1.7, 23 and 1.3 mmol/L. The  $\text{Mg}^{2+}$  was particularly sensitive in inhibiting  $\text{CH}_4$   
22 metabolism processes largely for gas-coal methanogenic consortium.

23 **Keywords:** Anaerobic bacteria; Culture media; Organic matrix; Coal; Bio-methane.

## 24 1. INTRODUCTION

25 The demand for gas has seen a tremendous increase in the last 10 years [1]. In  
26 particular, coal-bed methane enhance with biotechnology represents an ideal method to  
27 increase methane productivity from oil deposits and coal seams using underground  
28 anaerobic digestion processing [2,3]. With the addition of adequate nutrient supplements,  
29 methanogenic consortium has the potential to degrade organic compounds in coal into  
30 methane. In this way, gas reserves and productivity could be enhanced along with  
31 secondary biogenic gas generation [4–6].

32 Previous studies involved with the methanogenic microbial traits and fermentation  
33 characteristics for bio-methane generation have focused on the processes of bio-  
34 degradation by microbial communities in coal seam [2,7,8] Anion research identified that  
35 methane biosynthesis process could active where coalbed waters exhibit relatively low  
36 salinities ( $<2 \text{ mol/L Cl}^-$ ) and low  $\text{SO}_4^{2-}$  concentrations ( $<10 \text{ mmol/L}$ ) [9]. Nutrients  
37 supply by meteoric water plays an important role in methanogenic microbial metabolism

38 [10,11]. However, nutritional deficiency exists in most coal seam. Most organic  
39 compounds in coal cannot be readily used as nutrients for methanogenic microbes. In  
40 addition to the inherent elements present in coal beds, supplemental nutrients are  
41 important to remediate the nutritional deficiency and promote microbial growth [12,13].  
42 If concentrations of supplemental nutrients are excessive, the methanogenic consortium  
43 will be adversely influenced [14]. The goal of this report is to identify the optimal  
44 nutrient dose for microbial community culture. Methane and carbon dioxide yield rate  
45 were the indicators used to assess the results of experiments.

## 46 **2. Materials and methods**

### 47 **2.1 Materials**

48 The coal and coalbed water samples used in this study were collected from Laohutai  
49 Fm 103<sup>#</sup> coalbed located in Shenyang province of China (GPS coordinates 41.830256,  
50 123.957639). The depth of the Fm 103<sup>#</sup> gas-coal sample was 550 meters and the  
51 thickness of the coal seam was 6-18 meters. Laohutai Mine, in the east part of Fushun  
52 near Shenyang, Liaoning province, opened in 1901. The mine has 55.6 million tons of  
53 recoverable coal reserves until 2004[15]. The samples were collected by the State Key  
54 Laboratory of Coal Resources and Safety Mining. The field studies did not involve  
55 endangered or protected species, and only small sample quantities were used. And no  
56 specific permissions were required for these locations sample collection and research.

57           The coal samples were obtained from a newly exposed coal seam in the heading face  
58 by the channel sampling method following ASTM D 4596-86 with nitrogen protection.  
59 Two sample points were selected from the Fm 103<sup>#</sup> coal seam. These samples were  
60 sealed in gas-tight steel canisters and set with gas-tight valves (manufactured by J&D  
61 Technologies) which enabled inert gas to flush the canister and protected with nitrogen  
62 immediately. The coal to be used for the microbe community source was crushed into  
63 small pieces (10-15 mm in diameter) at an aseptic bench (manufactured by JingXue  
64 Technologies). Nitrogen protection was used to maintain the coal in a continuous  
65 anaerobic environment. The sample was sealed in a sterilized gas desorption canister  
66 (manufactured by J&D Technologies) to desorb the absorbed gas until no gas desorbed in  
67 atmospheric pressure at 25 °C[16].

68           The formation of water samples from the Fm 103<sup>#</sup> coalbed were collected in the  
69 same coal-bed in sterile glass bottles (manufactured by Fisher Scientific) which were  
70 filled to overflow to prevent oxygen ingress. Coalbed water ( $\leq$  24 hours after collection)  
71 was autoclaved at 121 °C for 45 minutes and sealed in sterile glass bottles. Argon was  
72 infused to seal the top space of the bottles. The coalbed water samples were stored at 4 °C  
73 for no more than 3 days.

## 74 **2.2 Culture**

75           Five different nutrition culture media were tested in these experiments: MAC-1,

76 MAC-2, MAC-3, MAC-4, MAC-5 [4,17] (MAC is the abbreviation of “the influence test  
77 of organic matrix and cations”). The final concentrations of the compounds (g/L) were as  
78 list in table 1, and 0.1 mL/L of resazurin was added as an oxygen indicator. Yeast  
79 extract was produced by Fisher BioReagents, and other chemicals were supplied by  
80 Acros Organics.

81 **Table 1.** The MAC1-MAC5 nutrition culture media concentrations (g/L)

Culture Media	Sodium Acetate	Glucose	Beef Extract	Yeast Extract	Common Concentrations
MAC-1	2.00	3.00	3.00	2.00	
MAC-2	2.00	1.00	1.50	1.00	NH <sub>4</sub> Cl, 1.00; MgCl•7H <sub>2</sub> O, 1.00;
MAC-3	0.00	1.00	1.50	1.00	KH <sub>2</sub> PO <sub>4</sub> , 0.40; KCl, 0.50;
MAC-4	0.00	0.00	0.00	1.00	NaHCO <sub>3</sub> , 1.00; L-Cysteine Hydrochloride Monohydrate, 0.45
MAC-5	0.00	0.00	0.00	0.00	

82 The distilled water was autoclaved at 121 °C for 45 minutes with dissolved oxygen  
83 removed. Nutrition medium was prepared in a 500 mL flask with distilled water. The  
84 medium was mixed using a magnetic stirrer for 2 hours at 60 °C at an aseptic bench and  
85 then combined with an equal volume of coalbed water (volume ratio 1:1) for another hour  
86 at room temperature. Nitrogen protection was used to maintain nutrition in a continuous  
87 anaerobic environment throughout the entire experiment. The final pH was maintained at  
88 6.0 for all nutrition cultures. Control samples of 100 ml MAC-1, MAC-2, MAC-3, MAC-

89 4, and MAC-5 were sealed in separate sterile glass bottles and stored at -40 °C. Argon  
90 was infused to seal the top space of bottles.

91 Anaerobic conditions were ensured in flasks using a gas-replacement method. This  
92 gas replacing process was monitored in real time with a carbon dioxide monitor system  
93 (manufactured by E2V). For each experiment 50.00±1 g of the coal sample and 500.00  
94 mL of medium were used. Nitrogen was used to seal the upper space of the flask at the  
95 beginning of the experiment. The flasks were placed in an incubation shaker at 35 °C and  
96 were agitated at 80 rpm to maximize the coal-liquid mass transfer rates. Twelve parallel  
97 tests were designed for each nutrition group which were cultured for 40 days under  
98 identical conditions.

99 The control experiments were performed without coal sample, which named MAC-  
100 1\*, MAC-2\*, MAC-3\*, MAC-4\*, MAC-5\*, to identify whether exogenous organic has  
101 the possibility to supply extra carbon to enhance the bio-methane yield. For each control  
102 experiment of each nutrition group 100 mL of cultured medium and 400 mL new medium  
103 was used. And cultured for 40 days in the same condition as above.

### 104 **2.3 The cation orthogonal analysis**

105 The cation orthogonal analysis was based on consumption of cation elements in bio-  
106 degradation processes, according to the L<sub>16</sub>(4<sup>5</sup>) orthogonal table which includes Na<sup>+</sup>, K<sup>+</sup>,  
107 Ca<sup>2+</sup>, Mg<sup>2+</sup>. The lowest ion level was based on the MAC-4 medium while the highest ion

108 level was based upon that of the East China Sea ion concentration. A quantity of  $50.00 \pm 1$   
109 g of Fm 103<sup>#</sup> gas-coal sample was used in each experiment. The upper space of the flasks  
110 was sealed with nitrogen. The flasks were placed on an incubation shaker at 35 °C and  
111 agitated at 80 rpm to maximize coal-liquid mass transfer rates. Cation orthogonal  
112 experiments were of 40 days of culture.

#### 113 **2.4 Gas analysis**

114 Gas samples were obtained with a 50- $\mu$ L gas syringe. Methane and carbon dioxide  
115 analyses were performed using an Agilent 7890A gas chromatograph (manufactured by  
116 Agilent). The nitrogen (carrier gas) flow rate was set at 1.00 mL/min. The injection port  
117 was maintained at 150°C with the oven temperature set at 25°C and the Thermal  
118 Conductivity Detector (TCD) at 200°C. Retention times for methane were 3.76 minutes  
119 and 5.0 minutes for carbon dioxide. Calibration standards consisted of 40% methane,  
120 20% carbon dioxide, 10% hydrogen and 30% nitrogen, which were injected at  
121 atmospheric pressure to generate the calibration plot.

#### 122 **2.5 Nutrient Metabolism Analysis**

123 Nutrient ion concentrations were analyzed using an HC-800 ionization analyzer  
124 (manufactured by Histrong Technologies). The main indicators included fluoride,  
125 chloride, nitrate and nitrite nitrogen, phosphate, sulfate, carbonate, bicarbonate, ammonia  
126 nitrogen, sodium, potassium, magnesium, calcium, pH, water hardness and total

127 alkalinity.

## 128 **3. Results**

### 129 **3.1 The nutrient abundance influence for bio-degradation of coal**

130 Five different organic compound nutrient concentrations designated as MAC-1,  
131 MAC-2, MAC-3, MAC-4, and MAC-5 were assessed in this study. In these medium,  
132 Sodium Acetate is a collective medium, as carbon resource, for *methanogenic* normally  
133 [18]. Glucose is the medium for *hydrogen-producing acetogens*. Beef Extract is a mixture  
134 of peptides and amino acids, nucleotide fractions, organic acids, minerals, and some  
135 vitamins. It is often used to supply carbon and nitrogen sources. Yeast extract contains a  
136 mixture of amino acids, peptides, water soluble vitamins, and carbohydrates. And it is  
137 often used in culture media [19]. The rank order of organic concentrations were MAC-1 >  
138 MAC-2 > MAC-3 > MAC-4 > MAC-5.

139 Coalbed methanogenic groups are comprised of a variety of microbial types. With  
140 regard to *methanogens*, only a limited number of simple carbon compounds such as CO<sub>2</sub>  
141 or acetate can serve as substrates. For the conversion of complex organic compounds to  
142 methane, fermentative and *acetogenic bacteria* are required. Thus they group an  
143 interactive methanogenic consortium [20]. In the process of gas-coal bio-degradation, the  
144 capacity for fermentative bacteria to hydrolyze and ferment the organic compounds of  
145 coal plays an important role, and CO<sub>2</sub> is the main compound of gas productions. Beef



146 extract in medium supplied the extra carbon and nitrogen for bacteria. For the *acetogenic*  
147 *bacteria* fermentation process, the long chain fatty acids and sugar degrade to form  
148 acetate, CO<sub>2</sub> and H<sub>2</sub> [18]. Glucose is the medium to enhance the *acetogenic bacteria*  
149 fermentation. And *methanogens* yield CH<sub>4</sub> with CO<sub>2</sub> and H<sub>2</sub> or acetate. Sodium Acetate  
150 was introduced to raise the acetate supply for *methanogens* [21].

151 If consortia in a good balance, most CO<sub>2</sub> and H<sub>2</sub> will be used to form CH<sub>4</sub>. And  
152 the concentration of CH<sub>4</sub> should be high, meanwhile, the concentration of CO<sub>2</sub> and H<sub>2</sub>  
153 should keep in low. So the yield gas concentration with microbial fermentation process  
154 could reflect the bacteria balance conditions. The presence of sufficient organic nutrients  
155 in the medium could promote fermentative and *acetogenic bacteria* flourish and improve  
156 methanogen nutrient generation. However, if excessive amounts of organic nutrients are  
157 contained within the medium, which like beef extract and glucose, the high propagation  
158 of fermentative or *acetogenic bacteria* would break the microbial balance, and carbon  
159 dioxide yield rate could be enhanced, meanwhile, methanogenesis could be inhibited.  
160 This phenomenon showed in MAC-1, MAC-2 and MAC-3 culture experiments (Figure  
161 1).

162 The results confirmed that MAC-4 was the most effective medium for enhancing the  
163 bio-methane generation rate. 1 g/L yeast extract provided the best concentration of  
164 organics amino acids, peptides, and water-soluble vitamins to optimize the microbial  
165 fermentation. The maximal methane concentrations of the MAC-4 culture group achieved

166 23.62% (Figure 1). It was 4 times higher than that of the MAC-2, MAC-3, MAC-5  
167 groups on average. The MAC-4\* control experiment and MAC-5 group verified that the  
168 anthracite is important to provide carbon for bio-methane yield (Figure 1).

169 **Figure 1 here**

170 The high carbon dioxide and low methane concentration data obtained from MAC-1  
171 and MAC-2 media demonstrate that hydrolytic bacterial had enhanced with an abundance  
172 of organic substrate supplement, however, methane biosynthesis tended to inhibit in  
173 culture even with additional sodium acetate. In MAC-1 and MAC-2 experiments, the  
174 *acetogenic bacteria* inhibited also with sodium acetate influence in the medium.

175 The MAC-3 medium, which follows the same medium concentration of MAC-2  
176 except sodium acetate, enhanced *acetogenic bacteria*, and inhibited the *methanogens*.  
177 After 40 days of culture, the average H<sub>2</sub> volume concentration for the MAC-3 and MAC-  
178 3\* group achieved 67.2% and 59.15% as indicated by gas analysis. This amount was 33-  
179 40 times greater than that of all other experimental groups. The exogenous organic  
180 material played an important role in carbon supply.

181 However, organic nutrition plays an important to supply the vitamin and other  
182 microelements besides organic nutrients. Without the vitamin and microelements besides  
183 organic nutrients supply, the consortia would grow at a slower rate, such as that observed  
184 for MAC-5.

### 185 **3.2 The nutritional metabolism analysis of methanogenic consortium**

186 Changes in MAC-4 ion concentrations were analyzed in the initial and final media.  
187 The data from this analysis revealed that the concentrations of sodium, potassium,  
188 ammonia nitrogen and magnesium were consumed microbially. In particular, 85.78% of  
189 the sodium was utilized with microbial fermentation. The medium pH changed from 6.65  
190 to 7.32 over this period. In contrast, the concentrations of sulfate and bicarbonate  
191 increased 1.5-7 fold (Table 2). These data indicated that sodium, nitrogen, potassium, and  
192 magnesium are the key elements for methanogenic consortium fermentation. However,  
193 the sulfate should be maintained at a low level in the medium.

194 **Table 2.** The MAC-4 medium ion concentration changes with methanogenic consortium  
195 bio-degradation.

	pH	Fluoride	Chloride	Nitrate Nitrogen	Nitrite Nitrogen	Phosphate	Sulfate	Carbonate
Initial	6.65	3.70	81.58	-	-	44.67	195.80	-
final	7.32	5.18	95.90	-	-	38.60	478.40	-
	Bicarbonate	Sodium	Ammonia Nitrogen	Potassium	Magnesium	Calcium	Water Hardness	Total Aalkalinity
Initial	40.66	811.90	193.60	159.00	38.89	21.06	212.56	40.66
final	328.77	115.40	39.23	35.30	14.30	22.76	115.62	328.77

### 196 **3.3 The cations orthogonal analysis**

197 From the analysis of nutrient element concentrations, it is clear that sodium,  
198 ammonia, nitrogen, potassium, and magnesium play important roles in the methanogenic

199 microbial metabolism. The orthogonal analysis experiment was designed to identify the  
 200 influence of cations in bio-methane yield. The orthogonal module introduced 4 cations,  
 201 ( $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ ) as tested with 4 different concentrations. The lowest ion  
 202 concentration was set by MAC-4 and the highest reference level for cation analysis was  
 203 that of the East China Sea ion concentration. The  $L_{16}(4^5)$  orthogonal table was used as an  
 204 orthogonal analysis module (Table 3).

205 **Table 3.** The cation orthogonal analysis table for  $\text{CH}_4$  and  $\text{CO}_2$ . The results represent  
 206 volume percent units for both gases. The table shows the analysis of  $\text{CH}_4$  and  $\text{CO}_2$   
 207 ranges.

Serial Number	Factors (mmol/L)				Results	
	Na	Mg	Ca	K	$\text{CH}_4$	$\text{CO}_2$
1	23	1.3	0.5	10.7	6.65	4.71
2	23	6.7	1.7	1.7	3.92	4.27
3	23	13.0	3.4	3.4	0.92	4.03
4	23	20.0	5.1	5.1	0.46	3.23
5	130	1.3	1.7	3.4	6.59	4.84
6	130	6.7	0.5	5.1	3.85	3.56
7	130	13.0	5.1	10.7	0.99	3.31
8	130	20.0	3.4	1.7	0.38	2.94
9	260	1.3	3.4	5.1	6.49	4.47
10	260	6.7	5.1	3.4	3.76	4.73
11	260	13.0	0.5	1.7	0.57	2.47
12	260	20.0	1.7	10.7	0.34	2.84
13	390	1.3	5.1	1.7	7.01	4.86
14	390	6.7	3.4	10.7	0.30	3.90
15	390	13.0	1.7	5.1	0.47	2.28
16	390	20.0	0.5	3.4	0.30	2.67
$\text{CH}_4$	$k_{1j}$	2.99	6.69	2.84	2.97	
	$k_{2j}$	2.95	2.96	2.83	2.89	
	$k_{3j}$	2.79	0.74	2.02	2.82	

	k <sub>4j</sub>	2.02	0.37	3.06	2.07
	R <sub>j</sub>	0.97	6.32	1.03	0.90
	k <sub>1j</sub>	4.06	4.72	3.35	3.64
	k <sub>2j</sub>	3.66	4.12	3.56	4.07
CO <sub>2</sub>	k <sub>3j</sub>	3.63	3.02	3.84	3.39
	k <sub>4j</sub>	3.43	2.92	4.03	3.69
	R <sub>j</sub>	0.63	1.80	0.68	0.68

208 Range analysis was used to indicate the affected order among factors. Range R<sub>j</sub> was  
 209 calculated with module 1.

$$R_j = \text{Max}_{k_{jm}} - \text{Min}_{k_{jm}} \quad (1)$$

210

211 Where  $k_{jm}$  is the average result of the  $m$  level  $j$  factor (module 2), ( $j, m = 1, 2, 3, 4$ ).

$$k_{jm} = \sum_{i=1}^4 y_{jmi} / 4 \quad (2)$$

212

213 Where  $y_{jmi}$  is the result of the  $m$  level  $j$  factor number  $i$  data, ( $j, m, i = 1, 2, 3, 4$ ).

214 Range analysis confirmed that for methanogenic activity, the best ion concentrations  
 215 for enhancement CH<sub>4</sub> yield were Na<sub>1</sub>, Mg<sub>1</sub>, Ca<sub>4</sub>, and K<sub>1</sub>, with the rank order of element  
 216 effectiveness being Mg > Ca > Na > K. The most effective ion concentration for control  
 217 CO<sub>2</sub> generation were Na<sub>4</sub>, Mg<sub>4</sub>, Ca<sub>1</sub>, and K<sub>3</sub>, with its ran order being Mg > Ca = K > Na.

218 Notable differences exist regarding the influence of CH<sub>4</sub> and CO<sub>2</sub> on yield rates as a  
 219 function of cation concentrations. The CH<sub>4</sub> biosynthesis is substantially more sensitive to  
 220 element content. Maximal Mg<sup>2+</sup> concentrations to enhance methanogen metabolism  
 221 processes were 1.3 mmol/L, while maximal cation concentrations for Ca<sup>2+</sup>, K<sup>+</sup>, Na<sup>+</sup> were  
 222 5.1, 1.7 and 23 mmol/L, respectively.

223 Variance analysis has been used to analyze the Factor Significance (F) in this  
 224 system. F was calculated with module 3.

$$225 \quad F_j = \frac{4 \sum_{i=1}^4 K_{ij}^2 - (\sum_{i=1}^{16} x_i)^2}{16 \sum_{i=1}^{16} x_i^2 - (\sum_{i=1}^{16} x_i)^2} \times 5 \quad (3)$$

$$226 \quad K_{ij} = \sum_{i=1}^4 y_{jmi} \quad (4)$$

227 Where  $x_i$  is the experiment results for every factor and level.

228 **Table 4.** The variance analysis table for 4 factors with CH<sub>4</sub> and CO<sub>2</sub> data.

	Factor	Square of Deviance	Degree of Freedom	F Value	Significant
CH <sub>4</sub>	Na <sup>+</sup>	2.465	3	0.892	
	Mg <sup>2+</sup>	100.905	3	36.533	*
	Ca <sup>2+</sup>	2.486	3	0.900	
	K <sup>+</sup>	2.080	3	0.753	
	Error	2.762	3		
CO <sub>2</sub>	Na <sup>+</sup>	0.842	3	8.592	
	Mg <sup>2+</sup>	9.120	3	93.061	**
	Ca <sup>2+</sup>	1.079	3	11.010	*
	K <sup>+</sup>	0.954	3	9.735	
	Error	0.098	3		

229 Based upon the F calculation,  $Mg^{2+}$  was the significant cation factor for  $CH_4$  and  
230  $CO_2$  (Table 4). This ion is particularly sensitive in inhibiting  $CH_4$  metabolism processes  
231 largely for gas-coal methanogenic consortium.

## 232 **4. Discussion**

233 Microbial bio-degradation of some organic compounds of coal represents a current  
234 technology available to enhance coal-bed methane. Microbial cooperation via anaerobic  
235 digestion processes results in the implementation of secondary biogenic methane  
236 generation and gas reservoirs enhancement in mining. Based on the experiments  
237 performed in this report, the following conclusions can be garnered: (1) The organic  
238 nutrient dose should be adjusted to methanogenic consortium requirements. Excessive or  
239 deficient organic nutrient support doses adversely affecting biomethane yield; (2) MAC-4  
240 was the most effective medium in enhancing anaerobic digestion in Fushun gas-coal  
241 seam; (3) The sodium, nitrogen, potassium, and magnesium are the key elements for  
242 methanogenic consortium fermentation; (4) The cation rank order of influence Fushun  
243 methanogenic consortium metabolism was  $Mg > Ca > Na > K$ , and maximal cation  
244 concentrations of  $Mg^{2+}$ ,  $Ca^{2+}$ ,  $K^+$ ,  $Na^+$  were 1.3, 5.1, 1.7, and 23 mmol/L, respectively;  
245 (5)  $Mg^{2+}$  was a particularly sensitive factor, which could inhibit methanogenic bacteria.

## 246 **5. Conclusion**

### 247 **5.1 The nutrient concentration for gas-coal methanogenic consortium culture**

248           The biomethane generation from coal involves a complex interaction between  
249 environmental factors and biotic communities. Hydrolytic fermentative bacteria,  
250 syntrophic *acetogenic bacteria*, methanogenic bacteria, and many other bacteria comprise  
251 the biotic community. The environment includes not only the physical factors but also the  
252 coal, coalbed water, coal-bed gas and other complex formations which we call the  
253 environment.

254           The medium injected into coal seam enriching nutrients of the coal seam. The series  
255 of experiments of this report confirm that nutrient media require a strict concentration of  
256 control to be effective, especially for those involving organic materials. If the organic  
257 compounds are too rich, like that modeled with our formulations of MAC-1, MAC-2, and  
258 MAC-3, the excessive levels of nutrients will adversely affect the microbial community  
259 structure. High rates growth of hydrolytic fermentative bacteria or *acetogenic bacteria*  
260 could inhibit methane biosynthesis. In contrast, if lack organic nutrition, such as that  
261 modeled in MAC-5, the potential for enhancing methane biosynthesis is low and the yield  
262 proceeds at a slow rate. Concentrations of nutrition required should differ as a function  
263 of: (1) the methane biosynthesis type, such as carbon dioxide reduction or acetate  
264 fermentation; (2) coal maturity grade, such as gas-coal, flame coal, bituminous coal or  
265 lignite. The exact organic nutrition required for different coal ranks needs to be identified  
266 to maximize bio-methane yield rates.

## 267 **5.2 Ion concentration for bio-methane yield enhancement**



268           Organic biological degradation to methane is a microbial cooperation process.  
269   Fermentative bacteria initially hydrolyze complex organic compounds to acetate, longer  
270   chained fatty acids, carbon dioxide, hydrogen,  $\text{NH}_4^+$ , and  $\text{HS}^-$ . Syntrophic hydrogen-  
271   producing (proton-reducing) *acetogenic bacteria* reduce intermediary metabolites to  
272   acetate, carbon dioxide, and hydrogen. Hydrogen-utilizing *acetogenic bacteria*  
273   demethoxylate low molecular weight ligneous compounds and ferment some  
274   hydroxylated aromatic compounds. Carbon dioxide reduction methanogenic bacteria are  
275   dependent on hydrogen, produced by other bacteria, to reduce carbon dioxide or  
276   bicarbonate to methane. And acetate fermentation methanogen yields methane via acetate  
277   bio-degradation.

278           Different nutrients are required for different microbial activity in the community.  
279   Results from the cation orthogonal experiments revealed that cation concentrations  
280   critically influence the metabolism process of the microbial. Except for the examination  
281   of sodium, potassium, magnesium, calcium which analyzed in the cations orthogonal  
282   analysis, the nitrogen, yeast extract, salinity and pH could have the potential to influence  
283   the biomethane synthesis. More researches to identify the effects of ions upon the coal  
284   seam biotic community are needed to reveal the microbial activity in gas-coal beds.

## 285   **6. Competing interests**

286           The authors declare that they have no competing financial interests.

## 287 **7. Acknowledgments**

288 The authors' acknowledge the contributions of the following companies for allowing  
289 access to coal samples and other information used in this paper: Laohutai Mining. We  
290 thank, Ke Li, Yumei Jia's assistance in this work.

291 This work was supported by the State Key Laboratory of Coal Resources and Safe  
292 Mining Subject (grant number SKLCRSM17KFA08 and SKLCRSM19X012), the  
293 Fundamental Research Funds for Central Universities (grant number 2014QNB41).

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## 357 **Figure Legends**

358 **Figure 1. Changes in methane and carbon dioxide production as a function of media**  
359 **nutrition and time in culture.** Figure A is the methane concentration changes with  
360 culture days. And figure B is the control experiment without coal sample supply in  
361 experiment. Figure C is the carbon dioxide concentration changes with culture days. And  
362 figure D is the control experiment without coal sample supply in experiment. Methane is  
363 the main factor to identify the *methanogens* activity. And carbon dioxide is an important  
364 factor to identify the fermentative and *acetogenic bacteria* activity. The methane yield  
365 rate would high, only if the fermentative and *acetogenic bacteria* activity in a limited  
366 condition. Organic material could enhance the fermentative and *acetogenic bacteria*.  
367 However, if the activity of fermentative and *acetogenic bacteria* is too high, it would  
368 inhibit the *methanogens*. The microbial group will in a good balance when achieved the  
369 highest methane yield and the best ratio of methane and carbon dioxide. Thus the MAC-4  
370 medium culture group fit for the requirements.

